DATTA, MRIDUL, Ph.D. Impact of Tomato Juice on Radiation Side Effects and select Inflammatory Mediators in Prostate Cancer Patients undergoing Intensity Modulated Radiation Therapy. (2011)
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This pilot study assessed tolerance of different volumes of processed tomato juice consumed daily and its impact on serum lycopene, selected serum inflammatory mediator levels and radiation-induced side effects in men with localized prostate cancer undergoing radiation therapy. Participants (n = 17) were randomized into control group or one of three intervention groups (4 oz, 8 oz or 12 oz of processed tomato juice daily). Non-Hispanic Whites comprised 71% of study participants. Tumor staging ranged from $T_{1c-2c}N_0M_0$, with 71% of participant tumors in the $T_{1c}N_0M_0$ stage.

Participants tolerated daily tomato juice supplementation without any adverse gastrointestinal (GI) effects. Serum lycopene decreased in control group participants, while increasing from 0.33±0.11 µg/mL (baseline) to 0.41±0.12 µg/mL (endpoint) in the intervention group. No correlation between serum and dietary lycopene was detected. Control group participants lost weight, while participants in the intervention groups did not. Not surprisingly, participants exhibited systemic inflammation at baseline. Overtime, increased c-reactive protein (CRP) and interleukin-6 (IL-6) was observed in control group, while decreases in serum CRP, IL-6 and prostaglandin E2 (PGE2) levels were observed in intervention groups (p>0.05). No statistically significant within group differences were detected for CRP. Within group differences were statistically

significant for 12 oz group only, when comparing baseline and endpoint with midpoint levels (p = 0.014) for IL-6, and when comparing PGE2 baseline levels with midpoint and endpoint (p = 0.003).

We observed no statistical correlation between inflammatory markers, cancer characteristics and dietary or serum lycopene, or acute side effects of treatment. Lower performance score was observed in intervention group participants. Daily tomato juice intake appeared to offer a GI protective effect during the first three weeks of treatment. Based on the results of this study, daily consumption of processed tomato juice (at least 8-12 oz) may decrease serum levels of CRP, IL-6 and PGE2; lower performance status score; and offer a protective GI effect during radiotherapy for prostate cancer. This information may assist in improving patient tolerance and minimize acute side-effects of radiation therapy in men with localized prostate cancer undergoing intensity modulated radiation therapy.

IMPACT OF TOMATO JUICE ON RADIATION SIDE EFFECTS AND

SELECT INFLAMMATORY MEDIATORS IN PROSTATE

CANCER PATIENTS UNDERGOING INTENSITY

MODULATED RADIATION THERAPY

By

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A Dissertation Submitted to the Faculty of The Graduate School at The University of North Carolina at Greensboro in Partial Fulfillment of the Requirements for the Degree Doctor of Philosophy

Greensboro 2011

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APPROVAL PAGE

This dissertation has been approved by the following committee of the Faculty of the Graduate School at The University of North Carolina at Greensboro.

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CHAPTER I

INTRODUCTION

Chronic inflammation is now recognized as one causative mechanism in carcinogenesis (Colotta, Allavena, Sica, Garlanda, & Mantovani, 2009; Mantovani, Allavena, Sica, & Balkwill, 2008), contributing to about 20% of all cancers (De Marzo et al., 2007). The presence of inflammatory cells and mediators such as cytokines and prostaglandins in tumor tissue is considered a "hallmark" of cancer-related inflammation (Mantovani, et al., 2008). In addition to generating reactive oxygen (ROS) and nitrogen species (RNS), inflammation also activates several pro-inflammatory cytokines (such as tumor necrosis factor-α (TNF- α), interleukin-6 (IL-6), IL-1β) which further increase cancer risk by inducing enzymes that produce more ROS/RNS (Ohshima, Tatemichi, & Sawa, 2003). Additionally, cancer treatments, specifically radiation therapy may also induce inflammation (McBride et al., 2004). Researchers have reported upregulation of proinflammatory cytokines (TNF- α, IL-6, IL-1β) during and after treatment, contributing to fatigue (Bower, 2007) and impacting quality of life.

Prostate cancer is the most commonly diagnosed cancer among men ("Cancer Facts and Figures 2010," 2010), and inflammation has been identified as one causative factor (De Marzo, et al., 2007; Wong, Bray, & Ho, 2009). Cell

and animal studies have demonstrated upregulation of several cytokines and inflammatory enzymes in prostate carcinogenesis (Fujita et al., 2002).

Researchers have also documented upregulation of several inflammatory mediators during radiation therapy for prostate cancer in men (Christensen et al., 2009; Johnke et al., 2009).

Nutritional modulation for chemoprevention is not a new concept. Consumption of phytonutrients such as lycopene, have been associated with a decreased incidence of prostate cancer in epidemiological studies (Giovannucci, 1999, 2002). Several cell (Hwang & Bowen, 2004; Ivanov et al., 2007; Kanagaraj et al., 2007; L. Kim, Rao, & Rao, 2002) and animal studies (Guttenplan et al., 2001; Konijeti et al., 2010) and clinical trials in men at risk for developing prostate cancer (Bunker et al., 2007; Edinger & Koff, 2006; Mohanty, Saxena, Singh, Goyal, & Arora, 2005; Schwarz et al., 2008) have demonstrated a chemopreventive role of lycopene. Researchers have also evaluated the benefits of lycopene in prostate cancer patients prior to initiating treatment (Jatoi et al., 2007; H. Kim et al., 2003; Kucuk et al., 2001) or after failure of treatment (Clark et al., 2006). Clinical trials have demonstrated a decline in serum prostate specific antigen (PSA) level, tumor bulk, and inflammation, and an increase in serum lycopene levels with lycopene supplementation. However, the impact of lycopene supplementation during radiation treatment for prostate cancer and its impact on biomarkers, physical side effects and quality of life remain unknown.

The *long-term goal* of this project was to increase our understanding of lycopene supplementation during radiotherapy for prostate cancer, to minimize side effects, evaluate biomarker responses, and develop specific guidelines for lycopene intake during radiation therapy. The *objectives* of this project were to assess tolerance of an orally ingested food source of lycopene (tomato juice) during radiotherapy for prostate cancer, impact of lycopene supplementation on serum lycopene levels, radiotherapy-related side effects, and select inflammatory markers. The patient population for this study was newly diagnosed patients with non-metastasized prostate cancer who had not received prior treatment and who were scheduled to receive radiotherapy to the prostate gland (treatment volume to include prostate and seminal vesicles). The *central hypothesis* was that tomato juice supplementation during radiotherapy will increase serum lycopene levels and minimize inflammatory response and radiation therapy-related side effects. The dissertation design is presented in Figure 1.1.

The *rationale* for this project was that knowledge of the impact of lycopene supplementation during radiation treatment in prostate cancer patients will provide important insights about a potential method to reduce side effects associated with radiotherapy in prostate cancer patients. Overall this project had the following three specific aims.

Aim 1

To examine the impact of supplementing three different volumes (4 oz, 8 oz, 12 oz) of tomato juice on serum lycopene levels in men with prostate cancer

undergoing radiation therapy. We also quantified dietary lycopene intake using a validated food frequency questionnaire (National Cancer Institute's Diet History Questionnaire (NCI-DHQ)).

Hypothesis: A dose-dependent response will be observed with tomato juice supplementation, with an increase in serum lycopene levels in the intervention groups. To test this hypothesis, we evaluated the effectiveness of three different doses of tomato juice on serum lycopene levels, which were measured using a liquid chromatography-mass spectrometry (LC-MS) analysis at Dr. Wei Jia's laboratory in Kannapolis, NC. We administered the NCI-DHQ once during this study to determine frequency of consumption of lycopene rich foods among the participants. We also used the NCI-common toxicity criteria (CTC) to evaluate if any participants suffered from gastro-intestinal (nausea, vomiting, heartburn) side effects as a result of tomato juice consumption.

Aim 2

Evaluate serum levels of select inflammatory mediators (c-reactive protein (CRP), prostaglandin E2 (PGE2), TNF-α, and IL-6) in men with prostate cancer prior to initiating radiation therapy and then at midpoint and endpoints along with supplementation of three different volumes of tomato juice.

Hypothesis: A dose-dependent decrease in serum inflammatory biomarkers will be observed with tomato juice supplementation.

To test this hypothesis, we evaluated the impact of three different doses of tomato juice on serum CRP, TNF- α , IL-6 and PGE₂ levels. Traditional enzymelinked immunosorbent assay (ELISA) tests were conducted to analyze the serum for the presence of CRP, TNF- α , IL-6 and PGE₂ at three different time points.

Aim 3

Evaluate the time of onset and severity of select side effects (urinary frequency and urgency, proctitis, and diarrhea) of radiation therapy in the control group and three treatment groups. We also wanted to evaluate if the cytokine response was predictive of radiation therapy symptoms.

Hypothesis: A dose-dependent response will be observed with tomato juice supplementation, with delayed onset and decreased severity of side effects, and improvement or maintenance of the patient's performance score. To test this hypothesis, we evaluated the effectiveness of three different doses of tomato juice on performance status (measured using the Eastern Cooperative Oncology Group (ECOG) Scoring), the time of onset and severity of urinary frequency and urgency, proctitis, and diarrhea. The symptom severity was measured using the NCI common terminology criteria for adverse events (CTCAE).

The research population (men with prostate cancer undergoing radiation therapy) and phytonutrient (lycopene/tomato juice) supplementation were based on the identified research gaps (Davis et al., 2005). However, the implication of inflammatory mediator involvement was extrapolated from literature reviews of

patients undergoing radiation therapy to the pelvis and lungs. Researchers have since demonstrated serum/plasma cytokine expression in men with prostate cancer undergoing radiation therapy (Christensen, et al., 2009; Johnke, et al., 2009).

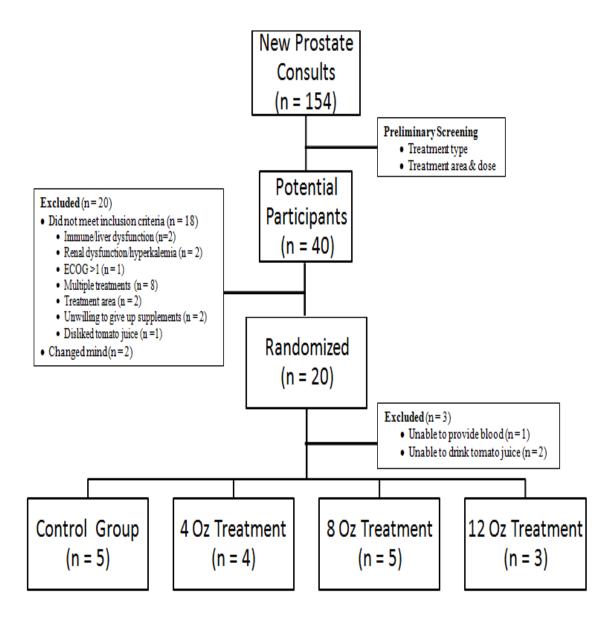


Figure 1.1. Flow diagram representing recruitment and randomization of study participants

References

- Bower, J. E. (2007). Cancer-related fatigue: Links with inflammation in cancer patients and survivors. *Brain, Behavior, and Immunity, 21*(7), 863-871.
- Bunker, C., McDonals, A., Evans, R., de la Rosa, N., Boumosleh, J., & Patrick, A. (2007). A randomized trial of lycopene supplementation in Tobago men with high prostate cancer risk. *Nutrition & Cancer*, *57*(2), 130-137.
- Cancer Facts and Figures 2010. (2010). Retrieved May 14, 2010, from http://www5.cancer.org/docroot/STT/content/STT_1x_Cancer_Facts_Figures_2010.asp
- Christensen, E., Pintilie, M., Evans, K. R., Lenarduzzi, M., Ménard, C., Catton, C.
 N., et al. (2009). Longitudinal Cytokine Expression during IMRT for
 Prostate Cancer and Acute Treatment Toxicity. *Clinical Cancer Research*,
 15(17), 5576-5583.
- Clark, P. E., Hall, M. C., Borden, J. L. S., Miller, A. A., Hu, J. J., Lee, W. R., et al. (2006). Phase I-II prospective dose-escalating trial of lycopene in patients with biochemical relapse of prostate cancer after definitive local therapy.

 Urology, 67(6), 1257-1261.

- Colotta, F., Allavena, P., Sica, A., Garlanda, C., & Mantovani, A. (2009). Cancer-related inflammation, the seventh hallmark of cancer: links to genetic instability. *Carcinogenesis*, *30*(7), 1073-1081.
- Davis, C., Clevidence, B., Swanson, C. A., Ziegler, R. G., Dwyer, J. T., & Milner, J. A. (2005). A Research Agenda for Lycopene/Tomato Supplementation and Cancer Prevention. *Journal of Nutrition*, 135(8), 2074S.
- De Marzo, A. M., Platz, E. A., Sutcliffe, S., Xu, J., Gronberg, H., Drake, C. G., et al. (2007). Inflammation in prostate carcinogenesis. *Nature Reviews, 7*, 256 269.
- Edinger, M. S., & Koff, W. J. (2006). Effect of the consumption of tomato paste on plasma prostate-specific antigen levels in patients with benign prostate hyperplasia. *Brazilian Journal of Medical and Biological Research*, 39, 1115-1119.
- Fujita, H., Koshida, K., Keller, E. T., Takahashi, Y., Yoshimito, T., Namiki, M., et al. (2002). Cyclooxygenase-2 promotes prostate cancer progression. *The Prostate*, *53*(3), 232-240.
- Giovannucci, E. (1999). Tomatoes, tomato-based products, lycopene and cancer: Review of the Epidemiologic literature. *Journal of National Cancer Institute*, 91(4), 317 331.

- Giovannucci, E. (2002). A Review of Epidemiologic Studies of Tomatoes,

 Lycopene, and Prostate Cancer. *Experimental Biology and Medicine*,

 227(10), 852-859.
- Guttenplan, J., Chen, M., Kosinska, W., Thompson, S., Zhao, Z., & Cohen, L. A. (2001). Effects of a lycopene-rich diet on spontaneous and benzo[a]pyrene-induced mutagenesis in prostate, colon and lungs of the lacZ mouse. *Cancer Letters, 164*, 1-6.
- Hwang, E.-S., & Bowen, P. E. (2004). Cell Cycle Arrest and Induction of Apoptosis by Lycopene in LNCaP Human Prostate Cancer Cells. *Journal of Medicinal Food, 7*(3), 284-289.
- Ivanov, N. I., Cowell, S. P., Brown, P., Rennie, P. S., Guns, E. S., & Cox, M. E. (2007). Lycopene differentially induces quiescence and apoptosis in androgen-responsive and -independent prostate cancer cell lines. *Clinical Nutrition*, *26*(2), 252-263.
- Jatoi, A., Burch, P., Hillman, D., Vanyo, J. M., Dakhil, S., Nikcevich, D., et al. (2007). A Tomato-Based, Lycopene-Containing Intervention for Androgen-Independent Prostate Cancer: Results of a Phase II Study from The North Central Cancer Treatment Group. *Urology*, 69(2), 289-294.
- Johnke, R. M., Edwards, J. M., Evans, M. J., Nangami, G. N., Bakken, N. T. G., Kilburn, J. M., et al. (2009). Circulating cytokine levels in prostate cancer

- patients undergoing radiation therapy:influence of neoadjuvant total androgen suppression. *In Vivo*, 23, 827-834.
- Kanagaraj, P., Vijayababu, M. R., Ravisankar, B., Anbalagan, J., Aruldhas, M. M., & Arunakaran, J. (2007). Effect of lycopene on insulin-like growth factor-I, IGF binding protein-3 and IGF type-I receptor in prostate cancer cells. *Journal of Cancer Research and Clinical Oncology*, 133, 351-359.
- Kim, H., Bowen, P., Chen, L., Duncan, C., Ghosh, L., Sharifi, R., et al. (2003).

 Effects of tomato sauce consumption on apoptotic cell death in prostate benign hyperplasia and carcinoma. *Nutrition & Cancer*, *47*(1), 40-47.
- Kim, L., Rao, A. V., & Rao, L. G. (2002). Effect of Lycopene on Prostate LNCaP Cancer Cells in Culture. *Journal of Medicinal Food*, *5*(4), 181-187.
- Konijeti, R., Henning, S., Moro, A., Sheikh, A., Elashoff, D., Shapiro, A., et al. (2010). Chemoprevention of prostate cancer with lycopene in the TRAMP model. *The Prostate*, *70*(14), 1547-1554.
- Kucuk, O., Sarkar, F., Sakr, W., Djuric, Z., Pollak, M., Khachik, F., et al. (2001).
 Phase II randomized clinical trial of lycopene supplementation before radical prostatectomy. *Cancer Epidemiol Biomarkers and Prevention*, 10, 861-868.

- Mantovani, A., Allavena, P., Sica, A., & Balkwill, F. (2008). Cancer-related inflammation. *Nature*, *454*(7203), 436 444.
- McBride, W. H., Chiang, C.-S., Olson, J. L., Wang, C.-C., Hong, J.-H., Pajonk, F., et al. (2004). A Sense of Danger from Radiation. *Radiation Research*, *162*(1), 1-19.
- Mohanty, N. K., Saxena, S., Singh, U. P., Goyal, N. K., & Arora, R. P. (2005).

 Lycopene as a chemopreventive agent in the treatment of high-grade prostate intraepithelial neoplasia. *Urol Oncol*, *23*(6), 383 385.
- Ohshima, H., Tatemichi, M., & Sawa, T. (2003). Chemical basis of inflammation-induced carcinogenesis. *Archives of Biochemistry and Biophysics*, *417*(1), 3 11.
- Schwarz, S., Obermuller-Jevic, U. C., Hellmis, E., Koch, W., Jacobi, G., & Biesalski, H.-K. (2008). Lycopene Inhibits Disease Progression in Patients with Benign Prostate Hyperplasia. *Journal of Nutrition*, *138*(1), 49-53.
- Wong, C. P., Bray, T. M., & Ho, E. (2009). Induction of proinflammatory response in prostate cancer epithelial cells by activated macrophages. *Cancer Letters*, *276*(1), 38 46.

CHAPTER II

REVIEW OF THE LITERATURE

Prostate Cancer

Prostate cancer has the dubious distinction of being the most commonly diagnosed cancer among men in the United States. In 2010, the American Cancer Society projected 217,730 new cases and of these, 6910 new cases were expected in North Carolina. Incidence of prostate cancer is highest among African American men. Prostate cancer is detected by PSA blood test and digital rectal exam, with final confirmation obtained with a prostate biopsy ("Cancer Facts and Figures 2010," 2010). In addition to ethnicity, age and family history (De Marzo et al., 2007), patients with high-grade prostatic intraepithelial neoplasia (HGPIN), genetic variants (such as SNP, CYP3A4, ELAC2, SRD5A2, etc), and elevated biochemical risk factors (e.g., Prostate Specific Antigen (PSA) and/or Insulin like Growth Factor-1) are also considered at a higher risk for developing prostate cancer (Lieberman, 2001). Chronic inflammation has also been implicated in the development of prostate cancer (De Marzo, et al., 2007; Rebbeck et al., 2008).

Inflammation and Prostate Cancer

Inflammation, a natural physiological process, initiated in response to injury as a result of infections (Philip, Rowley, & Schreiber, 2004; Sgambato & Cittadini, 2010) and environmental factors (Grivennikov, Greten, & Karin, 2010), has been implicated in the development of more than 20% of all human cancers, including prostate cancer (Bardia, Platz, Yegnasubramanian, De Marzo, & Nelson, 2009; De Marzo, et al., 2007; Kirschenbaum, Liu, Yao, & Levine, 2001). Epidemiologic, histopathologic and molecular pathological studies have provided evidence to implicate inflammation in prostate carcinogenesis (De Marzo, et al., 2007). Intraprostatic inflammation may be caused by infection, cellular injury, hormonal exposure and/or dietary factors, and direct injury to the prostate epithelium from chronic exposure to these noxious stimuli may lead to proliferative inflammatory atrophy (PIA) or proliferative atrophy (De Marzo, et al., 2007). PIA lesions may transition to PIN, and HGPIN is considered a precursor for prostate carcinogenesis (Wagenlehner et al., 2007). Tissue samples obtained from patients with benign prostate hyperplasia (BPH) have shown the presence of significantly high chronic inflammatory cells and atrophic epithelium indicating the presence of PIA (De Marzo, et al., 2007). A higher risk of developing prostate cancer has been reported in the presence of increased levels of inflammatory cytokines and chemokines in the prostate (Haverkamp, Charbonneau, & Ratliff, 2008). Researchers have demonstrated a decreased risk (21% lower) of developing prostate cancer in current and long-term users of

asprin, further linking inflammation in the development of prostate cancer (Salinas et al., 2010). Tables 2.1 - 2.3 summarize the cell/animal studies and human clinical trials documenting the association of inflammation in prostate pathologies.

Inflammatory Markers:

Key mediators of inflammation that were investigated in this study are briefly reviewed below:

Interleukin-6: Interleukin-6 (IL-6) is a plieotropic pro-inflammatory cytokine produced by leukocytes, fibroblasts and endothelial cells (Lamprecht, Oettl, Schwaberger, Hofmann, & Greilberger, 2007). IL-6 is involved in regulating several key cellular functions such as immune function, acute phase response, inflammation, oncogenesis (Song & Kellum, 2005), proliferation, differentiation, apoptosis and angiogenesis (Culig, Steiner, Bartsch, & Hobisch, 2005). IL-6 is considered a prognostic indicator for prostate cancer development and has been shown to correlate with stages of prostate cancer (Wertz, Siler, & Goralczyk, 2004). While IL-6 is localized to the basal epithelial cells in the normal prostate, in BPH it has been reported to be localized in the stromal and luminal epithelial cells (Royuela et al., 2004). Acting as an autocrine growth factor, IL-6 has been reported to be a key player in the stromal growth in BPH (Kramer, Mitteregger, & Marberger, 2007). Poor outcomes have been reported with elevated IL-6 levels in patients with prostate and other cancers (König, Senge, Allhoff, & König, 2004).

Researchers hypothesize that IL-6 may stimulate the growth of prostate cancer cells and cause progression of the disease through increased IL-6 production (Nakashima et al., 2000). An 18 fold increase in the concentration of IL-6 has been reported in localized prostate cancer compared to normal prostate tissue (Steiner et al., 2004). IL-6 has also been reported to upregulate the expression of COX-2 and PGE2 in a human PIN cell line and has a reciprocal and synergistic effect with COX-2/PGE2 on the growth of human PIN cells (X.-H. Liu et al., 2002). More recently researchers have demonstrated upregulation of IL-6 in men with prostate cancer undergoing radiation therapy (Christensen et al., 2009; Johnke et al., 2009). Diminished IL-6 expression has been reported in prostate tumors with lycopene supplementation (Wertz, et al., 2004). Figure 2.1 illustrates the cascade of events ascribed to IL-6 as discussed by several researchers (Christensen, et al., 2009; Helzlsouer, Erlinger, & Platz, 2006; Kuroda et al., 2007; Nakashima, et al., 2000; Steuber, Helo, & Lilja, 2007; Twillie et al., 1995).

C-Reactive Protein: C-reactive protein (CRP), an acute phase protein (Ford, Liu, Mannino, Giles, & Smith, 2003; Helzlsouer, et al., 2006; Lehrer et al., 2005), is a sensitive, yet non-specific marker of inflammation and is elevated in several diseases and disorders such as obesity, trauma, infection, diabetes and cancer (Walsh, Mahmoud, & Barna, 2003). CRP elevation in cancer patients may signify disease progression or recurrence (Walsh, et al., 2003). Two of the most commonly measured biomarkers of inflammation in prostate cancer are IL-6 and

CRP (Bowen, 2005). CRP levels are inversely associated with circulating levels of lycopene along with other carotenoids, retinoids and antioxidants (Ford, et al., 2003; McMillan et al., 2002).

<u>Tumor Necrosis Factor- α </u>: Tumor necrosis factor- α (TNF- α), a proinflammatory cytokine, is a paracrine and autocrine mediator (Danilko et al., 2007) of systemic inflammatory and immune reactions, and is produced by leukocytes, endothelia and adipocytes (Lamprecht, et al., 2007). Normal serum values are reported to be between 0-8.1 pg/mL (Akmansu, Unsal, Bora, & Elbeg, 2005). Chronic production of TNF may contribute to tumor promotion, leading to tissue remodeling and stromal development, ultimately contributing to tumor growth and metastasis (Wilson & Balkwill, 2002). TNF- α may trigger the inflammatory process by inducing the acute phase response along with increasing prostaglandin, leukotriene and collegenase synthesis (Huang, Ghai, & Ho, 2004). Acting as a macrophage and neutrophil activating factor, TNF- α triggers the synthesis of interleukin cascade, especially IL-1, IL-6, IL-8 and IL-10 (Danilko, et al., 2007). Loss of androgen responsiveness has been linked with tumor cell TNF production in patients with prostate cancer (Balkwill & Mantovani, 2001).

<u>Prostaglandin</u>: Prostaglandins (PGs) are lipid derivatives and regulate several physiological processes such as immune response, clotting, and platelet aggregation (Dubois et al., 1998; Williams, Mann, & DuBois, 1999). Action of cyclooxygenase (COX) enzyme on arachidonic acid leads to the production of

PGs (Keskek et al., 2006; Smyth, Grosser, Wang, Yu, & FitzGerald, 2009). COX-1 and COX-2, the two isoforms of COX are involved in PG synthesis (Keskek, et al., 2006). While COX-1 is the constitutive isoform, COX-2 is the inducible form whose expression is induced by proinflammatory cytokines such as IL-1 and TNF-α (Keskek, et al., 2006). PGs (PGE2, PGI2) generated via COX-2 are reported to be immunosuppressive (Williams, et al., 1999) and a key source of inflammation and inflammation related cancers (Smyth, et al., 2009). COX-2 is over expressed in several malignant conditions and may also be induced by radiation and chemotherapies. Therefore, decreasing expression of COX-2 may be potentially important in the prevention and treatment of these malignant conditions (Dorai & Aggarwal, 2004).

PGE₂ is a key player in the immune suppression associated with inflammation and is secreted by several different cells including tumor cells, macrophages and monocytes (Ben-Baruch, 2006). It is synthesized by cyclooxygenase -2 (COX-2), and together they show great potential as targets for cancer therapy (Ben-Baruch, 2006). COX-2 and PGE2 are over-expressed in both PIN and prostate cancer (Kirschenbaum, et al., 2001). Liu et al (2002) demonstrated in a human PIN cell line that COX-2, PGE₂ and IL-6 have a reciprocal and synergistic effect. PGE2 stimulates the release of soluble IL-6 receptor and activates the STAT-3 signaling (X.-H. Liu, et al., 2002).

Radiation Therapy for Prostate Cancer

In prostate cancer, radiotherapy may be used as an independent therapy or in conjunction with surgery, chemotherapy, or hormone therapy (Samant & Gooi, 2005). Radiotherapy is generally administered in small daily doses (fractions), five days a week over a period of several weeks (Samant & Gooi, 2005; Withers, 1992). The number of treatments may vary based on factors such as organ being treated, patient condition, and size of the tumor (personal communication with radiation oncologist BF on September 12, 2007). Conventional radiation therapy has been available since the 1960's and with the advent of technology, treatment dose has increased while the exposure to the normal tissue in close proximity to the tumor has decreased considerably (Duchesne, 2001). Intensity modulated radiation therapy (IMRT) can conform higher dose volumes as close to the tumor as possible (Duchesne, 2001). Despite this specificity of IMRT, non-cancerous tissues still get irradiated and physiological changes noted in patients are related to this low dose exposure (Okunieff, Chen, Maguire, & Huser, 2008). Thus, patients may develop acute or late intestinal radiation toxicity depending on the radiation dose, fractions and treatment time frame (Hovdenak, Fajardo, & Hauer-Jensen, 2000).

Radiation therapy leads to ionization of intracellular water, producing free radicals (Lewanski & Gullick, 2001; Prasad, Cole, Kumar, & Prasad, 2002), which results in DNA strand breaks and plasma membrane damage ultimately leading to cell death (Lewanski & Gullick, 2001). In addition to DNA damage, apoptosis,

necrosis and inactivation of the cell cycle are additional mechanisms through which radiation may cause cell death (Okunieff, et al., 2008). While total body irradiation can have a significant immunosuppressive effect, loco-regional radiation therapy can also have a suppressive effect on the immune system (McBride et al., 2004). Radiation therapy treatment has also been shown to induce inflammation and up-regulate PGE₂ (Milas & Hanson, 1995; Steinauer et al., 2000).

Acute Side Effects During Pelvic Radiation Therapy: Radiation treatment related adverse effects occurring during or within three months of radiation treatment are considered acute and impact a patient's quality of life (Hauer-Jensen, Wang, Boerma, Fu, & Denham, 2007). Acute toxicity is observed frequently in tissues such as the skin and the gastrointestinal (GI) tract, that experience rapid cell proliferation (Mollà & Panés, 2007). Radiation therapy destroys stem cells which prevents the functional cells lost during normal tissue turnover from being replaced (Mollà & Panés, 2007). In addition to the GI toxicities, genitourinary (GU) toxicities are still the most commonly observed treatment related side effects among patients receiving pelvic radiation, and are dependent on the dose and volume of radiation delivered (Hovdenak, et al., 2000; Teh et al., 2004), along with treatment time (Hovdenak et al., 2003) and its process of dissipation through the tissues (Andreyev, 2007).

Andreyev (2007) proposed that radiotherapy induced damage to the blood vessels may lead to ischemia and fibrosis, thus altering GI function by either

worsening pre-existing GI problems or creating new GI dysfunction depending on the affected site. Other researchers have proposed that acute toxicity in the intestines is secondary to damage to the epithelial cells leading to the breakdown of mucosal barrier and causing mucosal inflammation (Hauer-Jensen, et al., 2007). Radiation-induced bowel damage may be evident in > 75% patients receiving pelvic radiotherapy (Cole, Slater, Sokal, & Hawkey, 1993). Cole et. al (1993) demonstrated a significant increase in eicosanoid inflammatory mediators (such as leukotriene B4, thromboxane B2 and PGE₂), implicating them in radiation-induced bowel inflammatory changes in patients receiving pelvis radiotherapy. Table 2.4 summarizes some of the cell and animal research studies reporting the upregulation of COX-2 in radiation induced bowel injury. Figure 2.2 represents the cascade of events reported during radiation therapy (Ben-Baruch, 2006; Cole, et al., 1993; Dorai & Aggarwal, 2004; Dubois, et al., 1998; Kurzrock, 2001; Larsen et al., 2007; Steinauer, et al., 2000; Steuber, et al., 2007; Wang, Bergh, & Damber, 2005; Williams, et al., 1999).

Lycopene

Lycopene is a 40 carbon, lipid soluble, highly conjugated aliphatic hydrocarbon carotenoid responsible for the red color of some fruits and vegetables (Kun, Lule, & Xiao-Lin, 2006; A V Rao & Agarwal, 1999, 2000; A V Rao & Ali, 2007). Lycopene is not synthesized by animals, only microorganisms and plants (Agarwal & Rao, 2000). The molecular formula for lycopene is C₄₀H₅₆, its molecular weight is 536.85 Daltons (Kun, et al., 2006; A V Rao & Agarwal,

1999; Shi & Maguer, 2000) and its melting point is 172-175°C (Shi & Maguer, 2000). Lycopene resides primarily within cell membranes and is hydrophobic (Clinton, 1998). It contains 11 conjugated and two nonconjugated double bonds (A. Liu et al., 2006) and has a half life of 2-3 days in the serum (Stahl & Sies, 1992), and reaches maximum serum concentration within 15-48 hours of consuming lycopene rich foods (Gustin et al., 2004; Stahl & Sies, 1992). Lycopene is devoid of vitamin A activity due to the absence of the β-ionone ring (Clinton, 1998; Heber, 2004; Shi & Maguer, 2000).

Lycopene exists in a variety of geometric isomers like all-*trans*, mono-*cis*, and poly-*cis* form (Shi & Maguer, 2000). The *trans* configuration is the most common form of lycopene and is the form found in most plants (Kun, et al., 2006), while in the human plasma 50% lycopene exists as *cis* isomers (A V Rao & Agarwal, 2000; Shi & Maguer, 2000). The *all-trans* isomer of lycopene is the most thermodynamically stable form of the commonly identified isomers –5-*cis*, 7-*cis*, 9-*cis*, 11-*cis*, 13-*cis* and 15-*cis* (Shi & Maguer, 2000). The structural differences among some of the geometrical isomers of lycopene are presented in Figure 2.3.

Food sources of lycopene include tomatoes and tomato products, papaya, pink grapefruit, watermelon, and guava (Clinton, 1998; Kun, et al., 2006; A V Rao & Ali, 2007). Tomato and tomato products contribute more than 85% of dietary lycopene consumed in the North American diet (A V Rao & Agarwal, 2000). Tomato variety and the stage of ripening are key determinants of the lycopene

content of tomatoes (Clinton, 1998; Hadley, Miller, Schwartz, & Clinton, 2002). Tomato products may also contain polyphenols like quercetin, naringenin and chlorogenic acid along with small amounts of glucosinolates, tomatine and cyclolycopene (Bowen, 2005). More than 21 carotenoid pigments have been identified in tomatoes (Shi & Maguer, 2000). Some of these include phytoene, phytofluene, ζ-carotene, γ-carotene, β-carotene, neurosporene (Khachik et al., 2002) and lutein (Shi, Kakuda, & Yeung, 2004). The highest concentration of lycopene is found in the liver, adrenal glands, testes and the prostate (Kun, et al., 2006; Wertz, et al., 2004), while the lowest concentrations are noted in the kidney, lung and ovaries (Kun, et al., 2006). This variation in deposition of lycopene may be indicative of specific mechanisms involved in this process (Bramley, 2000). Tissue lycopene concentration in the prostate is reported to be 0-1.8 nM/gm (Clinton, 1998), and mean plasma lycopene concentrations ranging from 50-900 nM have been reported (Bramley, 2000). Concentration of lycopene in tissues, blood and other body secretions (seminal fluid, breast milk) is dependent on several factors such as food preparation practices, composition of the meal, diet composition, lipoprotein metabolism, and factors affecting lipid metabolism (Clinton, 1998).

Lycopene, a potent antioxidant and anti-inflammatory agent (Huang, et al., 2004; Rafi, Yadav, & Reyes, 2007), protects cells against ROS induced damage (Ford, et al., 2003; Neill & Fleshner, 2006; A V Rao & Shen, 2002). In addition to its role as an antioxidant, lycopene can influence several biological processes to

regulate cell cycle progression, gap junction communication and cytokine and growth factor signaling (Wertz, 2009). Some of the other mechanisms of action for lycopene include induction of phase II enzymes, inhibition of insulin like growth factor 1 (IGF-1) signal transduction, apoptosis induction, inhibition of IL-6 expression, inhibition of androgen activation and signaling, (Wertz, 2009) and modulation of the cyclo-oxygenase pathway (De Stefano et al., 2007; Sengupta, Ghosh, Das, Bhattacharjee, & Bhattacharya, 2006).

Recommended Intake of Lycopene: While researchers have recognized the health benefits of lycopene for some time now, it is still not recognized as an essential nutrient (A V Rao & Rao, 2007). No actual Dietary Reference Intake (DRI) levels currently exist for lycopene. Mean per capita intake of lycopene in the US has been reported to be about 8.2 (Matulka, Hood, & Griffiths, 2004) to 10.9 mg/day (McClain & Bausch, 2003). Clinical trials have been conducted using lycopene supplements (both food-based and pills) ranging from 5-120 mg/day, but a consensus is yet to be reached on the optimal daily dose. While Gustin et al., (2004) recommend daily ingestion of lycopene due to its short half life, they do not advocate more than 30 mg lycopene for chronic consumption.

Bioavailability of Lycopene: Lycopene absorption in humans is reported to be only about 10-30% of amount consumed (A V Rao, Ray, & Rao, 2006; Shi, Qu, Kakuda, Yeung, & Jiang, 2005). Lycopene in raw tomatoes is attached to the membranes which weaken during cooking, making lycopene more available from cooked tomato products (A V Rao & Agarwal, 2000; Weisburger, 2002). In

addition to heat, other factors that affect the bioavailability of lycopene include presence of dietary fiber (Kun, et al., 2006), and amount and type of fat (Kun, et al., 2006; Lee, Thurnham, & Chopra, 2000; A V Rao & Ali, 2007; Stahl & Sies, 1992). Other factors such as age, gender, hormonal status, smoking, alcohol consumption, body mass and composition may also affect bioavailability of lycopene (A V Rao, et al., 2006; Stahl & Sies, 1992). In addition, dietary components and drugs affecting lipid metabolism may impact serum lycopene concentration (Hadley, et al., 2002). During digestion lycopene is incorporated into micelles and through passive diffusion is absorbed by the intestinal epithelial cells (Shi, et al., 2005). Chylomicrons carry lycopene into the portal circulation, and after being repackaged, lycopene is released into the blood bound to low density lipoprotein (LDL) (Shi, et al., 2005).

Safety/Toxicity Evaluation of Lycopene: Upon reviewing lycopene supplementation studies, McClain and Bausch (2003) reported no systemic toxicity, genotoxicity, or adverse effects on reproductive parameters. No adverse effects were noted in toxicology studies conducted in rats with intakes up to 3 gm/kg/day (Trumbo, 2005). Safety of food based lycopene supplementation in humans has been established by virtue of its unrestricted use (McClain & Bausch, 2003). While no upper tolerable limit has been established for lycopene consumption, synthetic lycopene, lycopene extracts and crystallized lycopene extracts are generally regarded as safe (GRAS) in foods at levels ranging from 0.5 to 7% (Trumbo, 2005). After reviewing published data on lycopene

supplementation, Shao and Hathcock (2006) recommend 75 mg per day as the upper limit for supplementation in humans. Astley and Elliott (2005) propose that 15 mg of lycopene obtained from supplements is equivalent to the addition of approximately 150 gm of lycopene-rich foods in the diet.

Some side effects of lycopene supplementation have been reported in clinical trials. These include nausea and vomiting (Bunker et al., 2007; Jatoi et al., 2007), diarrhea (Clark et al., 2006; Jatoi, et al., 2007), abdominal distention (Jatoi, et al., 2007), flatulence (Edinger & Koff, 2006; Jatoi, et al., 2007), anorexia (Jatoi, et al., 2007) and dyspepsia (Jatoi, et al., 2007). Edinger and Koff (2006) reported heartburn, skin itching and flatulence among some study participants fed 50 grams tomato paste daily for 10 consecutive weeks. Jatoi et al., (2007) also reported occurrence of severe hypotension, prostatic hemorrhage, diarrhea, and anemia. While Jatoi et al., (2007) raise cautious concern about mild adverse reactions to lycopene supplementation, they do not preclude future clinical trials, but instead stress the importance of patient education about any potential side effects.

Prostate Cancer, Inflammation and Lycopene

Epidemiological studies (Giovannucci, 1999, 2002) have documented an inverse relationship between intake of lycopene rich foods and the development of prostate cancer. Several cell (Hwang & Bowen, 2004; Ivanov et al., 2007; Kanagaraj et al., 2007; L. Kim, Rao, & Rao, 2002) and animal studies (Guttenplan et al., 2001; Konijeti et al., 2010), and clinical trials in men at risk for

developing prostate cancer (Bunker, et al., 2007; Edinger & Koff, 2006; Mohanty, Saxena, Singh, Goyal, & Arora, 2005; Schwarz et al., 2008) have demonstrated the chemopreventive role of lycopene. Researchers have also evaluated the benefits of lycopene in prostate cancer patients prior to initiating treatment (Jatoi, et al., 2007; H. Kim et al., 2003; Kucuk et al., 2001) or after failure of treatment (Clark, et al., 2006). Despite the presence of several carotenoids and polyphenols, the chemoprotective effect of tomatoes on prostate cancer has been attributed to lycopene (Hwang & Bowen, 2004). Researchers have even demonstrated a stronger anti-mutagenic effect of food-based lycopene (tomato puree) compared with pure lycopene. This stronger effect was attributed to the likely synergistic effect of all bioactive compounds found in tomatoes (Polívková, Šmerák, Demová, & Houška, 2010). A summary of some of the key trials/studies are provided in Tables 2.5-2.7.

Despite several clinical trials evaluating the role of lycopene in patients with BPH and prostate cancer, data on the efficacy of lycopene supplementation in prostate cancer patients during radiation therapy are lacking. Based on the existing evidence, while some researchers have advocated for the inclusion of tomato extracts/products in prostate cancer treatment and prevention (Guns & Cowell, 2005), others have proposed urgency in evaluating the effectiveness of lycopene supplementation in prostate cancer patients undergoing radiation and androgen oblation therapies (Clinton, 2005; Davis et al., 2005), in order to formulate evidence-based recommendations for this population.

Lycopene Use During Radiation Therapy: Lycopene and other antioxidants have been proposed to resolve free radical induced oxidative damage produced during radiation therapy (Simone, Simone, Simone, & Simone, 2007). Researchers have demonstrated a beneficial effect of lycopene supplementation during radiation therapy to reduce the associated gastrointestinal (GI) side effects in rats (Andic, Garipagaoglu, Yurdakonar, Tuncel, & Kucuk, 2009). Clinical trials evaluating the impact of lycopene supplementation during radiotherapy are limited. The only documentation in the literature of lycopene supplementation during radiotherapy was found in patients with high grade gliomas undergoing radiotherapy and chemotherapy (Puri et al., 2005). Puri et al., (2005) reported a significant increase in serum lycopene levels, no adverse effects and positive, but not statistically significant (p = 0.1), outcomes in the lycopene (8 mg supplement) treated group compared to the control group. A summary of some of the studies evaluating the effectiveness of lycopene supplementation during radiation therapy are provided in Table 2.8.

Summary

Despite several clinical trials evaluating the role of lycopene in patients with BPH (Edinger & Koff, 2006; Schwarz, et al., 2008) and prostate cancer (Bowen et al., 2002; Chen et al., 2001; Jatoi et al., 2007; Kim et al., 2003; Kucuk et al., 2001; Rao et al., 1999), data on the efficacy of lycopene supplementation in prostate cancer patients during radiation therapy are lacking. To our knowledge, no studies have examined the effects of lycopene supplementation in

men undergoing radiation therapy for the treatment of prostate cancer. Several researchers have proposed the urgency in evaluating the effectiveness of lycopene supplementation in prostate cancer patients undergoing radiation and androgen oblation therapy (Bowen, 2005; Davis, et al., 2005). Thus, the proposed study evaluated the impact of daily supplementation of three different doses of lycopene on serum lycopene levels, PSA, and select markers of inflammation, and on the onset and severity of specific physical side effects (urinary frequency and urgency, proctitis, and diarrhea) and performance status in patients with localized prostate cancer during radiotherapy.

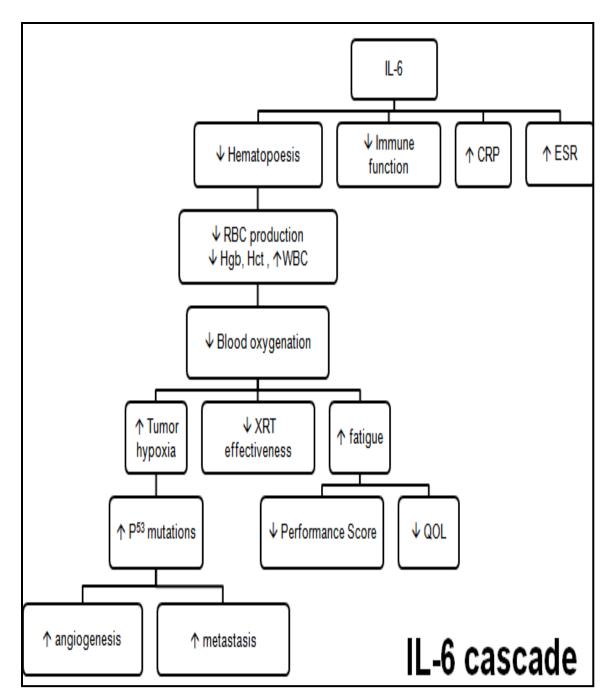


Figure 2.1. Cascade of events reported with increased IL-6 expression. IL-6 = interleukin-6, CRP = c-reactive protein, ESR = erythrocyte sedimentation rate, RBC = red blood cells, Hgb = hemoglobin, Hct = hematocrit, WBC = white blood count, XRT = radiation therapy, QOL = quality of life

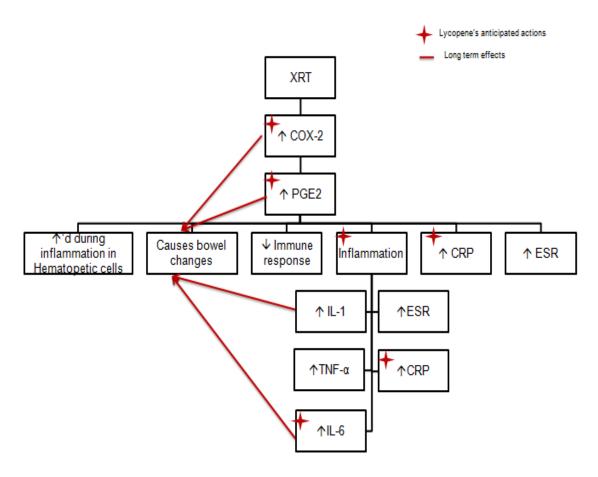


Figure 2.2. Radiation therapy related cascade. XRT = radiation therapy, COX-2 = cyclooxygenase-2, PGE2 = prostaglandin E2, CRP = c-reactive protein, ESR = erythrocyte sedimentation rate, IL-1 = interleukin-1, TNF- α = tumor necrosis factor- α , IL-6 = interleukin-6

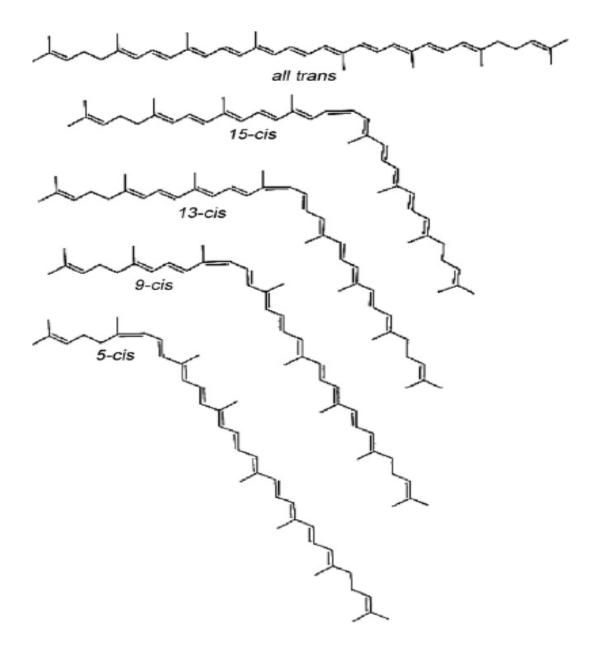


Figure 2.3: All-trans and cis isomers of lycopene (Source – (Rao & Rao, 2007)).

TABLE 2.1. Cell and animal research studies implicating inflammation in prostate tumorigenesis

Study	Animal/ cell line	Tests/Assays	Results
(Quintar et al., 2010)	Rat model	Prostatic tissues immuno- stained for PBP, ACTA2, ErbB1, & ErbB2 receptors, TUNEL, & cell proliferation markers. Dot & Western blots for PBP, ACTA2, ErbB1, ErbB2, & TGFb1	Post infection prostatic epithelium hypertrophied with ↑ PBP, ErbB1 & ErbB2 in a time dependent fashion. @ 72 hrs post infection, the epithelium showed apoptosis & atrophy with ↓ in PBP & ErbB receptors. ↓ in ACTA2 at 72 hr post infection indicating atrophic & proliferative changes promoted by even early stage prostatic inflammation
(Wong, Bray, & Ho, 2009)	LNCaP prostate cancer epithelial cells	Western blots, RNA isolation, cDNA synthesis, & real time quantitative PCR, ELISA, Flow cytometry	LNCaP cells show local proinflam response (NF-kB activation, & ↑ TNFa, IL-1b, & IL-6 expr. in activated macrophage conditioned media). Sig. ↑ VCAM-1 & nuclear estrogen receptor-a indicating potential link between chronic inflam. & its involvement in PC
(Fujita et al., 2002)	LNCaP cells	RNA Extraction and RT-PCR, Quantitative Real-Time PCR, western blot, COX assay, luciferase assay, ELISA	COX-2 mRNA & protein & COX activity in LNCaPCOX-2 cells sig. ↑ compared with parent and control-transfected cells & ↑ both proliferation in vitro & tumor growth rate in vivo. But pro-tumor effect not associated with androgen receptor level/activity. ↑ VEGF seen in LNCaP-COX-2 cells suggesting tumor growth in vivo stimulated by angiogenesis induced by COX-2.
(XH. Liu, et al., 2002)	Human PIN cell line	immunoprecipitation, immunoblotting, ELISA, nuclear extract preparation & electrophoretic mobility shift assay	PGE2 demonstrates autocrine upregulation by stimulating soluble IL-6 receptor release, gp130 dimerization, Stat-3 protein phosphorylation, & DNA binding activity, leading to ↑ PIN cell growth. COX-2 inhibitor ♥ cell growth. IL-6 neutralizing antibodies diminished PGE2-stimulated PIN cell growth indicating that ↑ COX-2/PGE2 leads to PC dev. thru activation IL-6 signaling pathway

TABLE 2.2. Human tissue studies implicating inflammation in prostate tumorigenesis

Study	n	Study Design & Population	Tests/Assays	Results
(Ravenna	20	human normal	real-time PCR and	RAGE, P2X7R, COX2, NOS2, PTX3, EGFR, ERa,
et al.,	pairs	and primary	Western blot analysis	(but not ERb) up-regulated in tumor samples.
2009)		prostate tissue	in prostate samples	Western blot analysis showed nuclear translocation of the NF-kB subunit p65.
(Bouraoui	47	Tissues from 5	Western blot &	in BPH, IL-1α, IL-6 and TNF expressed in pts with
et al.,		normal, 25 BPH	immune-histochemistry	PSA levels of 0-4 or 4-20 only, but not >20 ng/ml
2008)		and 17 human PC	of IL-1, IL-6 & TNF-α, PSA	In PC these cytokines only expressed in patients with PSA serum levels > 4 ng/ml.
(Khor et	586	Prostate tissue	Immunohistochemical	intensity of COX-2 staining an indep. predictor of
al., 2007)		from men treated	staining; any failure,	distant mets (p=0.0004); biochem. failure (p=0.008);
		with radiation	local failure, distant	& any failure (p=0.011). higher the expression of
		(XRT) and both	metastasis,	COX-2, greater the chance of failure. COX-2 over-
		short (STAD)or	biochemical failure,	expression most discriminating for those receiving
		long term	overall mortality, cause	STAD vs LTAD. COX-2 expr. sig. associated with
		androgen	specific mortality	biochemical failure, distant mets & any failure. LTAD
		deprivation		might overcome effects of COX-2 overexpr. COX-2
		(LTAD)		expr. may be useful in selecting pts needing LTAD.
(Cohen et	60	Prostate cancer	immune-	At 62-mths follow-up, COX-2 staining predicted
al., 2006)		specimens from	histochemistry, slide	progression with 82.4% sensitivity & 81.3%
		men post radical	grading; preoperative	specificity. Sensitivity (86.4%) & specificity (86.7%)
		prostatectomy	PSA, stage, Gleason	improved at ≥100-months follow-up., preop. PSA,
			sum (GS), margin,	EPE, margin, SV invasion & high COX-2 expr. were
			extraprostatic	sig. predictors of biochemical recurrence (p < 0.05).
			extension (EPE);	In multivariate analysis, preop PSA & COX-2 were
			seminal vesicle (SV)	indep. prognostic indicators. Pts with PSA > 7 ng/ml
			invasion	& ↑ COX-2 expr. had highest prob. of recurrence.

TABLE 2.2. Human tissue studies implicating inflammation in prostate tumorigenesis, cont'd

Study	n	Study Design & Population	Tests/Assays	Results
(Wang, Bergh, & Damber, 2004)	45	BPH samples.	immunohistochemist ry; double staining;	↑ COX-2 expression seen in all prostate luminal epithelial samples; highest proliferation index found in COX-2 +ve stained epithelium.COX-2 expression associated with Bcl-2 immunostaining in atrophic lesions (P<0.0001). T lymphocytes & macrophages predominant inflammatory cells related to the COX-2 expression in prostate epithelium.
(Hughes et al., 2002)	106	PC pts taking part in the Phase III RTOG 86-10 trial	Immunohistochemic al staining for COX-2	99% of slides expressed COX-2, samples with GS 7-10 had 81% COX-2 expression, GS 2-6 had only 69%
(Gupta, Srivastava, Ahmad, Bostwick, & Mukhtar, 2000)	12 PC tissue samples 12 control	Diagnosis of PC	semi-quanti. reverse transcription-PCR, immunoblotting, & immunohistochemist ry	Mean COX-2 mRNA levels 3.4-x ↑ in PC tissue compared with the paired benign tissue. COX-2 protein over- expressed in 10 of 12 samples;

TABLE 2.3. Clinical trials implicating inflammation in prostate tumorigenesis

Study	n	Study Design & Population	Tests/Assays	Results
(Salinas,	1001	Case control	Assessment of	Significant ♥ (21%) in PC risk seen among current
et al.,	PC	study	aspirin and other	users of aspirin compared with nonusers. Long-term
2010)	cases,		NSAID use,	use of aspirin & daily use of low-dose aspirin also
	942 age		Genotyping	associated with ♥ risk. no evidence that the association
	matched			with aspirin use varied by disease aggressiveness, but
	controls			there was effect modification. PC risk not related to use
(0.1)	070	NI (. I	D P	of non aspirin NSAIDs or acetaminophen.
(Schenk	676	Nested case-	Baseline serum was	↑ CRP associated with ↑ BPH risk (for quartile 4 vs.
et al.,	cases	control study	analyzed for CRP,	quartile 1, OR =1.40, 95% CI: 1.04, 1.88; which
2010)	683	from placebo	TNF-a, sTNF-RI &	attenuated after control for BMI (OR = 1.30, 95% CI:
	control	arm of the PC	sTNF-RII, IL-6, &	0.95, 1.75). Low sTNFRII & IL-6 associated with
		Prevention Trial	interferon c	BPH risk; associations only in men aged <65 years.
(Rebbeck,	1090	Caucasian PC	Genotype analysis	only remaining significant associations involved
et al.,		cases, history		CYP3A43 P340A genotypes & history of BPH on both
2008)		of BPH		Gleason grade (interaction p-value = 0.026) and tumor
				stage (interaction p-value = 0.017).
(Di	3942	Retrospective	Histopathology	↑% of pts with inflammation associated with BPH. FAA
Silverio et	case	histopathology	examination	(p = 0.027), PIN (p = 0.036) & incidental PC ↑ sig. with
al., 2003)		examination		age. Distribution on inflammatory aspects and AAH
				varied sig. (p = 0.002) based on prostate volume.

TABLE 2.3. Clinical trials implicating inflammation in prostate tumorigenesis, cont'd

Study	n	Study Design & Population	Tests/Assays	Results
(Nickel, Downey, Young, & Boag, 1999)	80	diagnosis of BPH	immunostained for leukocyte common antigen; computerized image-analysis system.	Inflammation present in all pts but only 1.1% of mean tissue surface area involved, 44% prostate specimens showed bacterial growth. no sig. diff. between inflammation pattern, volume or grade of inflammation in those catheterized or not (P=0.15) or culture +ve (pathogenic or not) & culture-negative cases (P=0.06). amount, degree or distribution of inflammation didn't sig. correlate with total PSA or PSA density

Table 2.4. Cell and animal research studies implicating COX-2 upregulation in radiation therapy of the prostate

Study	Animal/ cell line	Tests/Assays	Results
(Keskek, et al., 2006)	Sprague -Dawley rats	abdominal irradiation, Cyclooxygenase-2 Immunostaining, Myeloperoxidase Activity, Malondialdehyde Assay, Histopathology	COX-2 ↑ sig. in vascular endothelial cells on days 4 & 14 post exposure. COX-2 ↑ in fibroblasts immediately post irradiation & remained sig. ↑ during the entire study (<i>P</i> < 0.001), there was a peak COX-2 expression on day 14 similar to that observed in endothelial cells. Irradiation significantly ↑ intestinal epithelial damage, MPO activity, & MDA levels compared to the control group in a time-dependent fashion. Treatment with rofecoxib significantly ▶ these ↑ except on day 4
(Li et al., 2003)	PC3 cell line	Glutathione Assay, DCF, ROS assay, PGE2 monoclonal Immunoassay Kit	L-Buthionine sulfoximin ♥ cellular GSH & ↑ cellular ROS in PC-3 cells, whereas lipoic acid & NAC ↑ GSH level & ♥ cellular ROS. Both radiation & H202 similarly up-regulated COX-2 & PGE2 in PC-3 cells. ↑ greater when PC-3 cells pretreated with BSO. Pretreatment with a-lipoic acid or NAC for 24 h, up-regulated both radiation- & H202-induced COX2 & ♥ PGE2 production.
(Steinauer , et al., 2000)	PC3 cell line	Western blot analysis for COX-2 protein expression. PGE ₂ measured using Monoclonal Immunoassay Kit. flow cytometry.	dose-dependent ↑ in COX-2 of 37.0%, 79.7%, & 97.5% post irradiation with 5, 10, and 15 Gy, respectively. PGE₂ ↑ in irradiated cells than controls; ↓ PGE₂ levels in cells irradiated in the presence of NS-398. no differences in cell cycle distribution or apoptosis between cells irradiated in the presence or absence of NS-398.

TABLE 2.5. Summary of lycopene research in cell/animal studies

Study	Methods	Lycopene dose	animal/Cell line	Results
(Polívková,	AMES test, bone	(30 mg and 300	TA98,	Sig., dose dependent, antimutagenic effects of
et al., 2010)	marrow micronucleus	mg per plate)	TA100,	lycopene noted at various conc. Of AFB1 & IQ in
	test		BALB/c	TA98 & TA100. ↑ protective effects seen in AFB1 &
		0, 25 or 50	mice	IQ compared to MNU. Mice treated for 3 days with 25
		mg/kg for		or 50 mg/kg of lycopene prior to mutagen admin.
		animal study		show a sig. ♥ micronuclei numbers & tomato puree
				showed a stronger, dose-depend. antimutagenic
				effect compared with similar doses of pure lycopene
(Konijeti, et	Serum IGF-I, IGFBP-	28 mg/kg	TRAMP	PC incidence sig. Ψ in LB group compared to
al., 2010)	3 measured by	lycopene from	mice	controls (60% vs. 95%, P=0.0197) but not between
	ELISA, serum &	tomato paste		TP & control groups (80% vs. 95%, P=0.34). no diff
	prostate lycopene &	(TP) or		in prostate wts seen between groups. Both total
	α-tocopherol & liver	lycopene		serum & prostate lycopene levels ↑in LB & TP
	tissue deoxy-	beadlets (LB)		groups but not control group. ratio of 5-cis-lycopene
	guanosine & 8-			to trans-lycopene in serum was sig. \uparrow in LB group
	hydroxydeoxyguanosi			compared to TP group (P=0.0001). sig. ♥ oxidative
	ne measured by			DNA damage in livers of mice fed LB & TP diets but
	HPLC			not control group.
(Kanagaraj,	IGF-I, IGFBP-3 and	20, 40 & 60 µM	PC-3 cells	lycopene treatment showed a sig. ♥ in cell
et al., 2007)	IGF-I receptors in	lycopene		proliferation. 40 µM lycopene sig. ↑ IGFBP-3.
	lycopene-treated cells,	treated for 24,		Lycopene induced apoptosis; & DNA fragmentation
	Annexin V & PI	48, 72		absent after 24 hrs but not after 48 hrs. sig. in the
	binding studies	& 96 hrs		IGF-IR post cell treatment with lycopene, IGF-I

TABLE 2.5. Summary of lycopene research in cell/animal studies, cont'd

Study	Methods	Lycopene dose	animal/Cel I line	Results
(Ivanov, et al., 2007)	Proliferation assay, bromodeoxyuridine incorporation and flow cytometric analysis of cellular DNA content	0-100μΜ	LNCaP, PC3	LNCaP & PC-3 cells undergo mitotic arrest accumulating in G0/G1 phase - block in G1/S transition due to ♥ cyclin D1 & E, CDK4 & Rb phosphorylation. Corresponding with ♥ IGF-1 receptor expression & activation, ↑ IGFBP-2 expression & ♥AKT activation
(Srinivas an et al., 2007)	TBARS, single cell gel electrophoresis; levels of SOD, catalase, glutathione peroxidase ceruloplasmin, vitamin A, C, E & urinc acid	1.86, 9.31, 18.62 µM	hepatocyte s isolated from rat (Sprague- Dawley) liver	pretreatment with lycopene demonstrated a sig. ♥ in TBARS and DNA damage and a sig. ♠ in glutathione, vitamin A, C, E, uric acid and ceruloplasmin. Max protection to hepatocytes noted at 9.31 µM lycopene
(Gunasek era et al., 2007)	Inhibition of cell growth, time course studies, Mitogenic assays for growth factor studies	0, 0.04, 0.4, 5, 10, 20 μM	Dunning R3327AT3 or AT3 cells and DTE	both lycopene & lutein inhibited malignant AT3 cells in a dose and time-dep. manner, but not DTE cells. Lycopene showed more robust response than lutein & no synergistic or additive effect observed with both.
(Gitenay et al., 2007)	Mass Spec, RNA extravtion, real time PCR, western blot	50 mg/kg	Male Wistar rats,PC3A R human cell line	Cx43 expression sig. post exposure to RTS or YTS for 48 hrs compared to control, LBS effect not sig. diff. similar lycopene levels in cells incubated with RTS & LBS, while YTS contained no lycopene.
(Limpens et al., 2006)	HPLC-prostate and liver to detect lycopene and a,G, D tocopherol	Ly/Vit E-5 or 50 mg/kg; Ly+E - 5 mg/kg	PC-346C	none of the treatments sig. ♥ tumor vol., Lycopene+vit E @5 mg/kg sig. ♥ tumor growth, ↑ median survival by 40%. Lycopene levels ↑ in a dose dep.manner

TABLE 2.5. Summary of lycopene research in cell/animal studies, cont'd

Study	Methods	Lycopen e dose	animal/Cell line	Results
(A. Liu, et al., 2006)	subcellular fractionation of LNCaP cells using centrifugation and liquid chromatography-tandem mass spectrometry	1.48 Mmol/L	LNCaP, PC- 3, and DU145	After 24 hrs of incubation with 1.48 Mmol/L lycopene, LNCaP cells accumulated most lycopene & only one to express PSA. Levels of PSA ♥ 55% in LNCaP cells after 1.48 Mmol/L lycopene. lycopene not found to be a ligand for ligand-binding domain of the human androgen receptor. majority of lycopene (55%) localized in the nuclear membranes, followed by nuclear matrix (26%), & then microsomes (19%) & none in the cytosol.
(Hwang & Bowen, 2004)	effects of lycopene on cell growth or survival, cell cycle progression, and apoptosis	0, 0.1, 1, and 5 μ <i>M</i>	LNCaP	Lycopene at 1 μ <i>M</i> inhibited cell growth by 31% compared to placebo post 48-hr incubation. Lycopene at 5 μ <i>M</i> \uparrow number of cells in G2/M phase of cell cycle from 13% to 28% & \checkmark S-phase cells from 45% to 29%, no shifts in cell cycle seen in placebotreated groups. Apoptosis observed at 5 μ <i>M</i> lycopene @ late stages during 24 &48 hour treatments.
(Obermuller- Jevic et al., 2003)	Cellular uptake, thymidine assay, flow cytometry, western blot	0.1- 5.0µM	PrEC (Human prostate epithelial cells)	Dose dependent inhibition of cell proliferation noted; synchronized cells treated with 0.5µM lycopene showed minimal change, but cells treated with 5.0µM showed a significant accumulation in G0/G1 with no expression of cyclin D1, cyclin E was unaffected.

TABLE 2.5. Summary of lycopene research in cell/animal studies, cont'd

Study	Methods Lycope ne dose		animal/Cell line	Results		
(Yaping, Wenli, Weile, & Ying, 2003)	croton oil-induced mouse ear edema model, & glass slide method	0.1, 0.5, 1, 2 g/kg body wt		Administration of lycopene for four days was associated with ♥ swelling of the treated ear similar to amoxycillin. Lycopene also ↑ coagulation time		
(Guttenplan , et al., 2001)	DNA isolation, mutagenesis assay, lycopene assay	0.5, 1 mmol/kg	LacZ male mice	inhibition of spontaneous mutagenesis in prostate and colon observed at 1 mmol/kg of lycopene- rich tomato oleoresin (LTO), benzo[a]pyrene (BaP)-induced mutagenesis inhibited by LTO in prostate.		
(Forssberg, Lingen, Ernster, & Lindberg, 1959)	lycopene administered pre 48-12 hrs) & post (7-24 hrs) treatment	0.5-2 mg inj. Intraperit oneally	159 - lycop; 190 - control	lycopene improves survival when given before irradiation & at certain time after. females tolerated \(\bar{\textstyle } \) lycopene dose in combination with radiation better than males (males did better at smaller doses); radiation tolerance varies in different stages of menstrual cycle;		

TABLE 2.6. Summary of lycopene trials in healthy subjects

study	n	Lyco dose	food vs pill	duratio n	Participg Populat.	end point	Results
(Talvas et al., 2010)	30	0, 16 mg	Tomato paste, purified lycopen e	1 wk supplem entation, 2 wk washout period	Health men, 50- 70 yrs	Differentiat e the effect of tomato matrix from tomato paste or purified lycopene	serum lycopene ↑ after consuming red tomato paste & purified lycopene. No change in lipid profile, antioxidant status, PSA & IGF-I after consumption of tomato paste & lycopene. Sig. ↑ in IGFBP-3 & Bax:BcI-2 ratio & ✔of cyclin-D1, p53, & Nrf-2 in men consuming red tomato paste compared with serum collected after first washout period. Sig. ↑ of IGFBP-3, c-fos, & uPAR in men consuming purified lycopene compared with placebo
(Graydon et al., 2007)	20	15 mg	pill	4 weeks	healthy men 18- 60 yrs	Serum IGF-1, IGFBP-3 & Iycopene pre & post supplemnt	↑ change in serum lycopene in treatment group compared to control. no diff in median change in IGF-1 or IGFBP-3 between treatment & control groups. Change in lycopene corresponded with the change in IGFBP-3 in treatment group (r=0.78; P=0.008; n=10)
(Goyal, Chopra, Lwaleed, Birch, & Cooper, 2007)	6	22.8 mg	Cream of tomato soup	2 weeks	Healthy men	Blood/ seminal lycopene & antioxidant capacity pre & post supplemt	statistically sig. ↑ in blood & seminal plasma lycopene levels post supplementation, with a strong +ve correlation (<i>r</i> =0.84, <i>P</i> <0.05). no measurable ↑ in total radical scavenging capacity of semen.

TABLE 2.6. Summary of lycopene trials in healthy subjects, cont'd

study	n	Lyco pene dose	food vs pill	duration	Participg Populat.	end point	Results
(Riso, Brusamol ino, Martinetti , & Porrini, 2006)	20	5.7 mg	Lyc-o- mato drink	26 days	Healthy young men and women	Blood carotenoid, IGF-1 and IGFBP-3 estimation	tomato drink ↑ plasma lycopene, phytoene, phytofluene, & bcarotene (P < 0.05). No sig. effect of tomato drink on IGF-1. changes in lycopene pre & post each experimt period inversely & sig. correlated with IGF-1 (r = -0.33, P < 0.05). No correlation with other carotenoids. sig. ▶ serum IGF-1 (- 5.7%) with highest plasma lycopene (P<0.05). No change with placebo
(Gustin, et al., 2004)	25	10, 30, 60, 90, 120 mg	30 gm tomato paste + 5 ml olive oil	28 days	healthy men 18- 45 yrs	3 parameters of lycopene & isomers after single dose of lycopene; toxicity profile	headache, nausea, diarrhea most common adverse events, toxicity grade 2 or less; max serum lycopene reached in 15-32.6 hrs; half life 28-61.6 hrs; lower doses (10, 30) had the largest fin systemic exposure parameters
(Allen et al., 2003)	36	21 mg, 12 mg, 17 mg	sauce, soup, juice (v8 tomato or veggie)	6 week lycopene free diet- 2 wk washout+ 4 week Tx	healthy men and women; 18-65 yr	impact of single daily svgs of 3 tomato prod. on blood & BMC conc. of carotenoid & lycopene isomer	serum & BMC lycopene ↑ with all three (sauce, soup, juice); lycopene plateaued after 2 weeks.

TABLE 2.6. Summary of lycopene trials in healthy subjects, cont'd

study	n	Lyco pene dose	food vs pill	duration	Particip Populat.	end point	Results
(Hadle y, et al., 2002)	60	35 mg, 23 mg, 25 mg	cond. soup (35 mg), RTS soup (23 mg), V8 veg juice (25 mg)	1 wk lycopene free diet, 15 day lycopene suppleme ntation	healthy >40 yr old men & women	rate & magnitude of plasma lycopene after 1 wk of lycopene free diet & then with 3 diff food sources of lycopene; assess impact on biomarkers of oxidative damage	plasma conc of lycopene ♥ 35% (p<0.0001) during washout period, lycopene ↑ 123% cond. soup, 57% RTS soup, 112% V8. No significant change seen in urinary 8-OH-2'-dG & 8-epi-PGF2α, but ex-vivo lipoprotein oxidation lag period (measure of antiox. capacity) ↑ significantly
(A V Rao & Shen, 2002)	12	5, 10, 20 mg	ketchup & lyc-o- mato	2 week washout, then 2 week interventio n	healthy men & women, non- smokers, no MVI	serum lycopene, oxidative biomarkers (MDA)	dose dependent ↑ in serum lycopene with both pill and food, levels slightly higher with pill but diff not stat. Sig. MDA ↓ & reduced thiols ↑ with both pill and food all doses.
(Porrini & Riso, 2000)	9	7 mg	25 gm tomato paste +	14 days	healthy women	plasma and lymphocyte carotenoid conc.; lymphocyte resistance to oxidative stress	Inverse relationship between plasma lycopene conc. and lymphocyte lycopene conc.and oxidative DNA damagesmall amts of lycopene over short duration can \uparrow carotenoid conc and resistance of lymphocytes to oxidative stress.

TABLE 2.6. Summary of lycopene trials in healthy subjects, cont'd

study	n	Lyco pene dose	food vs pill	duration	Particip Populat.	end point	Results
(Bohm & Bitsch, 1999)	22	5 mg	tomatoe s, tomato juice and pill	2 wks of washout, 6 weeks of supplemt	adult females (20-27 yrs)	serum lycopene, lipid panel, antioxidant capacity of plasma	no change with tomato intake, but TJ (0.22) and oleoresin (0.25) ↑ serum lycopene conc. No affect on the lipid status or antioxidant capacity in any of the 3 groups
(Paeta u et al., 1998)	15	70-75 mg	tomato juice, and lycopen e oleoresi n & beadlets	4 weeks each, 6 week washout period between Tx	healthy men and women 33-61 yr	lycopene plasma response from food and supplements; changes in lycopene oxidation products	lycopene level ↑ with all 3 Tx, no stat significant diff among the three; levels plateaued after 1 wk for supplements and after 2 wks for TJ. TJ ↑ other tomato carotenoids (phytofluene, phytoene). Cyclolycopene (lycopene metabolite) ↑ most with TJ.
(Stahl & Sies, 1992)	6	0.35, 1.25, 2.5µmol /kg	tomato juice - heated and unheate d	single dose (absorption , dose dep), once a day x 4 days)	,	absorption studies, dose dependent response, accumulation	Absorption: Peak conc seen within 24-48 hrs post consumption of heated TJ; 1/2 life 2-3 days; Dose Dep: lycopene uptake was dose dep, but not very linear; higher absorption with smaller amounts. Accumulation: of lycopene & its isomers very linear

TABLE 2.7. Summary of lycopene trials in subjects with prostate involvement

study	N	Lycopene dose (mg)	food vs pill	durati on	patient population	end point	results
(Barber et al., 2006)	37	10 mg	lycoplus (with vit C, E etc)	1 year	PC pts; confirm with biopsy, no Tx, watchful waiting	change in PSA velocity with lycopene supplementat ion	lycopene inhibits DNA synthesis in PEC, results inconclusive!! Large ↑ in PSA doubling time but not stat. sig. Data skewed since some pts eliminated had sig. ♥ in PSA
(Edinger & Koff, 2006)	43	13 mg	50 gm T paste	10 wks	BPH, ↑ PSA	PSA	Significant Ψ in PSA
(Mohanty, et al., 2005)	40	0, 4 mg BID	Lyc-o- mato	1 year	HGPIN	PSA, serum lycopene, prostate biopsy, DRE	lycopene can delay or prevent HGPIN from progressing to PC, inverse relationship between lycopene and PSA
(Chang et al., 2005)	118 cases 52 contrl	NA	NA	2 years	Cases- histologically confirmed localized PC Controls- PSA<4ng/ml , -ve DRE	Plasma carotenoid	50% less risk for men with ↑ plasma levels of α-carotene, trans-β-carotene, β-cryptoxanthin, lutein & zeaxanthin. No Sig. associations for total lycopene, all-trans lycopene, & cis-lycopene isomer peaks 2, 3, & 5, high levels of cis-lycopene isomer peak inversely associated with risk.
(Jian, Du, Lee, & Binns, 2005)	cases -130, 274 contrl	5 mg	lycoplus (5 mg lycopene + other antox	I year	Cases- PC controls- hospital inpts without cancer	prostate cancer dev	prostate risk ♥ with ↑ing intake of lycopene, α & β-carotene, β-cryptoxanthine, lutein,& zeaxanthine

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TABLE 2.7. Summary of lycopene trials in subjects with prostate involvement, cont'd

study	N	Lycopene dose (mg)	food vs pill	duration	patient population	end point	Results
(Ansari & Gupta, 2004)	20	10 mg	pill	3 mths	Metastatic hormone refractory PC	ECOG, Dz response (complete or partial vs stable or prog. dz), bone pain, LUTS	complete response-5%, partial -30%, stable dz -50%) & dz prog15%. ECOG Grade 0-25%, Grade I-50% & Grade II-25%. ♥ ECOG from Grade I-0-35% & Grade II-I-15%, ↑ in 15% & unchanged – 35%. LUTS improved in 61% of pts. No supplmt related intol./ toxicity seen.
(H. Kim, et al., 2003)	32-tx; 34-ct	30 mg	3 pasta dishes	3 weeks	prostate cancer, BPH	PSA, lycopene, bax, bcl-2,	no change in bcl-2, but bax ♥ in cancer, ↑ in apoptotic cells in BPH & cancer; PSA ♥
(Bowen et al., 2002)	32	30 mg	3 pasta dishes	3 weeks	localized PC awaiting prostatectom y	PSA, leukocyte oxidative DNA damage	Sig. ↑ in serum & prostate lycopene; leukocyte oxidative DNA & PSA sig. ↓ with lycopene;
(Kucuk et al., 2002)	26	30 mg	lyc-o- mato	3 weeks	PC	PSA, IGF-1, IGFBP-3, prostate tissue	Tx group pts had smaller tumors, ♥ involvement of surgical margins, PSA ♥ 18%; IGF-1 & IGFBP-3 ♥ sig.

TABLE 2.7. Summary of lycopene trials in subjects with prostate involvement, cont'd

study	N	Lycop ene dose (mg)	food vs pill	durati on	patient population	end point	Results
(van Breemen et al., 2002)	32	30 mg	Tomato sauce based pasta dishes	3 weeks	T1/T2 PC patients waiting prostatectomy	Cis & trans lycopene in serum & prostate tissue	Total lycopene ↑ 2.0-fold in serum & 3.0-fold in prostate tissue post suppl. Mean all-trans-lycopene in prostate tissue ↑ to 22.7% (from 12.4%) of total lycopene post suppl., serum only a 2.8% ↑but statistically sig.
(Chen et al., 2001)	32	30 mg	3 pasta dishes	3 weeks	PC pts waiting prostatectomy	PSA, leukocyte oxidative DNA damage	Sig.♠ in serum & prostate lycopene; leukocyte oxidative DNA & PSA sig. ♥ with lycopene
(Kucuk, et al., 2001)	26	30 mg	lyc-o- mato	3 weeks	localized PC patients waiting prostatectomy	PSA, connexin43, bax, bcl-2	serum lycopene ♥ post intervention. Lycopene suppl. may ♥ PC growth; bcl-2, bax same between groups, IGF-1 ♥ in both groups

TABLE 2.7. Summary of lycopene trials in subjects with prostate involvement, cont'd

study	N	Lycop ene dose (mg)	food vs pill	durati on	patient population	end point	Results
(Lu et al., 2001)	65 cases 132 contro	NA	NA	4 years	Cases- pathologically confirmed PC Control – healthy cancer free men	Plasma tocopherols, retinol & carotenoids, NCI FFQ	Sig. inverse associations with PC observed with plasma lycopene [OR, 0.17; 95% CI, 0.04–0.78; P for trend, 0.0052] and zeaxanthin (OR,0.22; 95% CI, 0.06–0.83; P for trend, 0.0028) Borderline associations found for lutein (OR, 0.30; 95% CI, 0.09 –1.03; P for trend, 0.0064) & b-cryptoxanthin (OR, 0.31; 95% CI, 0.08 –1.24; P for trend, 0.0666).
(A Venket Rao, Fleshner, & Agarwal, 1999)	12 cases 12 age match edcon trols	NA	NA	NA	Cases - Histologically conformed & untreated PC Controls- untreated muscle invasive bladder cancer, but no BPH or PC	Serum & tissue lycopene, lutein, cryptoxanthin, β-carotene, lipid and protein oxidation	Sig ♥ serum & tissue lycopene but not other carotenoids noted in cases; while no diff in serum lipid peroxidation noted, serum protein thiol sig ♥ in cases. Role of lycopene in preventing oxidative damage and ♥ risk of PC should be evaluated further.

TABLE 2.8. Cell and animal research studies investigating lycopene and radiation

Study	animal/ Cell line	Test/Assays	Lycope ne dose	Results
(Saada, Rezk, & Eltahaw y, 2010)	Male albino rats	6 Gy γ-radiation (XRT); Ileum tissues for xanthine oxidase; thiobarbituric acid reactive substances (TBARS); xanthine dehydrogenase; Superoxide dismutase (SOD), catalase (CAT), serotonin (5-HT), dopamine (DA), glutathione (GSH); norepinephrine (NE), epinephrine (EPI); monoamine-oxidase (MAO)	5 mg/kg body weight	Irradiated animals showed sloughing villi, ulcers, & ruptured goblet cells, shrinkage of submucosa layers & fibroblasts. Histopathological changes associated with sig. ↑ in TBARS & change in xanthine oxidoreductase system (XOR). sig. ♥ in reduced GSH, SOD & CAT seen. XRT also induced a sig. ♥ in level of: 5-HT, DA, NE, & EPI associated with an ↑ in MAO activity. Lycopene pretreatment sig. ↑ the oxidant/antioxidant status, associated with sig. regeneration of small intestine, & improved monoamines levels. Thus, lycopene may protect small intestine against radiation-induced damage.
(Andic, et al., 2009)	Wistar albino rats	8 Gy abdominal and pelvic radiation; Study endpoints: weight loss, diarrhea, duration of diarrhea, survival, & plasma level of TBARS.	5 mg/kg body wt/day	Rats receiving RT only had sig. ↑ wt. loss rate compared to the lycopene + RT group (<i>P</i> = 0.001). Plasma TBARS levels after RT were also sig. ↑ in the RT only group compared to lycopene + RT group (<i>P</i> = 0.001). Lycopene supplementation sig. ↓ wt. loss & prevented oxidative stress in rats treated with abdomino-pelvic radiation.

TABLE 2.8. Cell and animal research studies investigating lycopene and radiation, cont'd

Study	animal/ Cell line	Test/Assays	Lycope ne dose	Results
(Srinivas an, Devipriy a, Kalpana, & Menon, 2009)	Culture d human lympho cytes	γ-radiation at 1, 2 and 4 Gy. The cellular changes were estimated by using TBARS, hydroperoxides (HP), the antioxidants SOD, CAT, GPx & GSH. The DNA damage was analyzed by cytokinesis blocked micronucleus assay (CBMN), dicentric aberration (DC) and translocation frequency.	1, 5 & 10μg/ml	Diff. doses of γ-radiation led to sig. ♠ in # of DC, micronuclei (MN), translocation frequency, TBARS & HP level, but levels of GSH & antiox. enzymes sig. ♥ compared with control. Max. damage to lymphocytes @ 4Gy. Lycopene pretreatment (1, 5 &10µg/ml) sig. ♥ frequency of MN, DC & translocation when compared with control. level of TBARS, HP were also ♥ & activities of SOD, CAT & GPx were sig. ♠ along with GSH levels when compared with control. 5µg/ml lycopene more effective than other two doses-offering protection to normal lymphocytes against γ-radiation-induced cellular damage.
(Srinivas an, et al., 2007)	cultured rat hepatoc ytes	Cellular changes estimated using TBARS, SOD, CAT, glutathione peroxidase (GPx), GSH, ceruloplasmin, vitamins A, E, C and uric acid. DNA damage analysed by single cell gel electrophoresis (comet assay).	1.86, 9.31 & 18.62 µM	↑ severity of DNA damage observed with ↑ in γ-radiation dose (1, 2 & 4 Gy) in cultured rat hepatocytes. TBARS ↑ sig. while levels of GSH, vitamins C, E and A, ceruloplasmin, uric acid & antiox.enzymes sig. In γ-irradiated groups. Max. damage to hepatocytes observed at 4 Gy irradiation. Pretreatment with lycopene (1.86, 9.31& 18.62 μM) showed a sig. In levels of TBARS & DNA damage. Antiox.enzymes ↑ sig. along with the levels of GSH, vitamins A, E, C, uric acid & ceruloplasmin. Max. protection of hepatocytes observed at 9.31 μM lycopene pretreatment.

References

- Agarwal, S., & Rao, A. V. (2000). Tomato lycopene and its role in human health and chronic disease. *Canadian Medical Association Journal*, *163*(6), 739-744.
- Akmansu, M., Unsal, D., Bora, H., & Elbeg, S. (2005). Influence of locoregional radiation treatment on tumor necrosis factor-[alpha] and interleukin-6 in the serum of patients with head and neck cancer. *Cytokine*, *31*(1), 41-45.
- Allen, C. M., Schwartz, S. J., Craft, N. E., Giovannucci, E. L., De Groff, V. L., & Clinton, S. K. (2003). Changes in plasma and oral mucosal lycopene isomer concentrations in healthy adults consuming standard servings of processed tomato products. *Nutrition & Cancer*, 47(1), 48.
- Andic, F., Garipagaoglu, M., Yurdakonar, E., Tuncel, N., & Kucuk, O. (2009).

 Lycopene in the Prevention of Gastrointestinal Toxicity of Radiotherapy.

 Nutrition and Cancer, 61(6), 784 788.
- Andreyev, J. (2007). Gastrointestinal symptoms after pelvic radiotherapy: a new understanding to improve management of symptomatic patients. *The Lancet Oncology, 8*(11), 1007-1017.
- Ansari, M. S., & Gupta, N. P. (2004). Lycopene: A novel drug therapy in hormone refractory metastatic prostate cancer. *Urologic Oncology: Seminars and Original Investigations*, 22(5), 415 -420.

- Balkwill, F., & Mantovani, A. (2001). Inflammation and cancer: back to Virchow?

 Lancet, 357, 539 545.
- Barber, N. J., Zhang, X., Zhu, G., Pramanik, R., Barber, J. A., Martin, F. L., et al. (2006). Lycopene inhibits DNA synthesis in primary prostate epithelial cells in vitro and its administration is associated with a reduced prostate-specific antigen velocity in a phase II clinical study. *Prostate Cancer & Prostatic Diseases*, *9*(4), 407-413.
- Bardia, A., Platz, E. A., Yegnasubramanian, S., De Marzo, A. M., & Nelson, W. G. (2009). Anti-inflammatory drugs, antioxidants, and prostate cancer prevention. *Current Opinion in Pharmacology*, *9*(4), 419-426.
- Ben-Baruch, A. (2006). Inflammation-associated immune suppression in cancer:

 The roles played by cytokines, chemokines and additional mediators.

 Seminars in Cancer Biology, 16(1), 38-52.
- Bohm, V., & Bitsch, R. (1999). Intestinal absorption of lycopene from different matrices and interactions to other carotenoids, the lipid status and the antioxidant capacity of human plasma. *European Journal of Nutrition, 38*, 118-125.
- Bouraoui, Y., Ricote, M., García-Tuñón, I., Rodriguez-Berriguete, G., Touffehi, M., Rais, N. B., et al. (2008). Pro-inflammatory cytokines and prostate-specific antigen in hyperplasia and human prostate cancer. *Cancer Detection and Prevention*, *32*(1), 23 32.

- Bowen, P. (2005). Selection of Surrogate Endpoint Biomarkers to Evaluate the Efficacy of Lycopene/Tomatoes for the Prevention/Progression of Prostate Cancer. *J. Nutr.*, 135(8), 2068S-2070S.
- Bowen, P., Chen, L., Stacewicz-Sapuntzakis, M., Duncan, C., Sharifi, R., Ghosh,
 L., et al. (2002). Tomato Sauce Supplementation and Prostate Cancer:
 Lycopene Accumulation and Modulation of Biomarkers of Carcinogenesis.
 Exp Biol Med, 227(10), 886-893.
- Bramley, P. M. (2000). Is lycopene benefecial to human health? *Phytochemistry*, *54*, 233-236.
- Bunker, C., McDonals, A., Evans, R., de la Rosa, N., Boumosleh, J., & Patrick, A. (2007). A randomized trial of lycopene supplementation in Tobago men with high prostate cancer risk. *Nutrition & Cancer*, *57*(2), 130-137.
- Cancer Facts and Figures 2010. (2010). Retrieved May 14, 2010, from http://www5.cancer.org/docroot/STT/content/STT_1x_Cancer_Facts_Figures_2010.asp
- Chang, S., Erdman, J. W., Clinton, S. K., Vadiveloo, M., Strom, S. S., Yamamura, Y., et al. (2005). Relationship Between Plasma Carotenoids and Prostate Cancer. *Nutrition and Cancer*, *53*(2), 127 134.
- Chen, L., Stacewicz-Sapuntzakis, M., Duncan, C., Sharifi, R., Ghosh, L., van Breemen, R., et al. (2001). Oxidative DNA damage in prostate cancer patients consuming tomato sauce based entrees as a whole-food intervention. *Journal of National Cancer Institute*, 93, 1872-1879.

- Christensen, E., Pintilie, M., Evans, K. R., Lenarduzzi, M., Ménard, C., Catton, C. N., et al. (2009). Longitudinal Cytokine Expression during IMRT for Prostate Cancer and Acute Treatment Toxicity. *Clinical Cancer Research*, 15(17), 5576-5583.
- Clark, P. E., Hall, M. C., Borden, J. L. S., Miller, A. A., Hu, J. J., Lee, W. R., et al. (2006). Phase I-II prospective dose-escalating trial of lycopene in patients with biochemical relapse of prostate cancer after definitive local therapy.

 Urology, 67(6), 1257-1261.
- Clinton, S. K. (1998). Lycopene: Chemistry, biology, and implications for human health and disease. *Nutrition Reviews*, *56*(2), 35-51.
- Clinton, S. K. (2005). Tomatoes or Lycopene: a Role in Prostate Carcinogenesis? *J. Nutr.*, *135*(8), 2057S-2059.
- Cohen, B. L., Gomez, P., Omori, Y., Duncan, R. C., Civantos, F., Soloway, M. S., et al. (2006). Cyclooxygenase-2 (COX-2) expression is an independent predictor of prostate cancer recurrence. *International Journal of Cancer,* 119(5), 1082-1087.
- Cole, A. T., Slater, K., Sokal, M., & Hawkey, C. J. (1993). In vivo rectal inflammatory mediator changes with radiotherapy to the pelvis. *Gut*, *34*(9), 1210-1214.
- Culig, Z., Steiner, H., Bartsch, G., & Hobisch, A. (2005). Interleukin-6 regulation of prostate cancer cell growth. *Journal of Cellular Biochemistry*, 95(3), 497-505.

- Danilko, K. V., Korytyna, G. F., Azhmadishina, L. Z., Yanbaeva, D. G., Zagidullin,
 S. Z., & Victorova, T. V. (2007). Association of polymorphism of cytokine
 genes (IL1B, IL1RN, TNFA, LTA, IL6, IL8, and, IL10) with chronic
 Obstructive Pulmonary Disease. *Molecular Biology*, 41(1), 22-31.
- Davis, C., Clevidence, B., Swanson, C. A., Ziegler, R. G., Dwyer, J. T., & Milner, J. A. (2005). A Research Agenda for Lycopene/Tomato Supplementation and Cancer Prevention. *Journal of Nutrition*, 135(8), 2074S.
- De Marzo, A. M., Platz, E. A., Sutcliffe, S., Xu, J., Gronberg, H., Drake, C. G., et al. (2007). Inflammation in prostate carcinogenesis. *Nature Reviews, 7*, 256 269.
- De Stefano, D., Maiuri, M. C., Simeon, V., Grassia, G., Soscia, A., Cinelli, M. P., et al. (2007). Lycopene, quercetin and tyrosol prevent macrophage activation induced by gliadin and IFN-[gamma]. *European Journal of Pharmacology*, *566*(1-3), 192-199.
- Di Silverio, F., Gentile, V., De Matteis, A., Mariotti, G., Giuseppe, V., Antonio
 Luigi, P., et al. (2003). Distribution of Inflammation, Pre-Malignant Lesions,
 Incidental Carcinoma in Histologically Confirmed Benign Prostatic
 Hyperplasia: A Retrospective Analysis. *European Urology, 43*(2), 164 175.
- Dorai, T., & Aggarwal, B. B. (2004). Role of chemopreventive agents in cancer therapy. *Cancer Letters*, *215*, 129-140.

- Dubois, R. N., Abramson, S. B., Crofford, L., Gupta, R. A., Simon, L. S., A. Van De Putte, L. B., et al. (1998). Cyclooxygenase in biology and disease. FASEB J., 12(12), 1063-1073.
- Duchesne, G. M. (2001). Radiation for prostate cancer. *The Lancet Oncology,* 2(2), 73-81.
- Edinger, M. S., & Koff, W. J. (2006). Effect of the consumption of tomato paste on plasma prostate-specific antigen levels in patients with benign prostate hyperplasia. *Brazilian Journal of Medical and Biological Research*, 39, 1115-1119.
- Ford, E. S., Liu, S., Mannino, D. M., Giles, W. H., & Smith, S. J. (2003). C-reactive protein concentration and concentrations of blood vitamins, carotenoids, and selenium among United States adults. *European Journal of Clinical Nutrition*, 57(9), 1157-1163.
- Forssberg, A., Lingen, C. H. R., Ernster, L., & Lindberg, O. (1959). Modification of the x-irradiation syndrome by lycopene. *Experimental Cell Research*, 16, 7-14.
- Fujita, H., Koshida, K., Keller, E. T., Takahashi, Y., Yoshimito, T., Namiki, M., et al. (2002). Cyclooxygenase-2 promotes prostate cancer progression. *The Prostate*, *53*(3), 232-240.
- Giovannucci, E. (1999). Tomatoes, tomato-based products, lycopene and cancer: Review of the Epidemiologic literature. *Journal of National Cancer Institute*, *91*(4), 317 331.

- Giovannucci, E. (2002). A Review of Epidemiologic Studies of Tomatoes,

 Lycopene, and Prostate Cancer. *Experimental Biology and Medicine*,

 227(10), 852-859.
- Gitenay, D., Lyan, B., Talvas, J., Mazur, A., Georgé, S., Caris-Veyrat, C., et al. (2007). Serum from rats fed red or yellow tomatoes induces Connexin43 expression independently from lycopene in a prostate cancer cell line. [doi: DOI: 10.1016/j.bbrc.2007.10.030]. *Biochemical and Biophysical Research Communications*, 364(3), 578-582.
- Goyal, A., Chopra, M., Lwaleed, B. A., Birch, B., & Cooper, A. J. (2007). The effects of dietary lycopene supplementation on human seminal plasma. BJU International, 99(6), 1456-1460.
- Graydon, R., Gilchrist, S. E. C. M., Young, I. S., Obermuller-Jevic, U.,
 Hasselwander, O., & Woodside, J. V. (2007). Effect of lycopene
 supplementation on insulin-like growth factor-1 and insulin-like growth
 factor binding protein-3: a double-blind, placebo-controlled trial. *Eur J Clin Nutr, 61*(10), 1196.
- Grivennikov, S. I., Greten, F. R., & Karin, M. (2010). Immunity, Inflammation, and Cancer. [doi: DOI: 10.1016/j.cell.2010.01.025]. *Cell, 140*(6), 883-899.
- Gunasekera, R. S., Sewgobind, K., Desai, S., Dunn, L., Black, H. S., McKeehan,
 W. L., et al. (2007). Lycopene and Lutein Inhibit Proliferation in Rat
 Prostate Carcinoma Cells. *Nutrition and Cancer*, 58(2), 171 177.

- Guns, E., & Cowell, S. (2005). Drug Insight: lycopene in the prevention and treatment. *Nature Clinical Practice in Urology*, *2*(1), 38-43.
- Gupta, S., Srivastava, M., Ahmad, N., Bostwick, D. G., & Mukhtar, H. (2000).

 Over-expression of cyclooxygenase-2 in human prostate adenocarcinoma. *The Prostate*, 42(1), 73-78.
- Gustin, D. M., Rodvold, K. A., Sosman, J. A., Diwadkar-Navsariwala, V.,
 Stacewicz-Sapuntzakis, M., Viana, M., et al. (2004). Single-Dose
 Pharmacokinetic Study of Lycopene Delivered in a Well-Defined Food-Based Lycopene Delivery System (Tomato Paste-Oil Mixture) in Healthy
 Adult Male Subjects. *Cancer Epidemiol Biomarkers Prev.*, 13(5), 850-860.
- Guttenplan, J., Chen, M., Kosinska, W., Thompson, S., Zhao, Z., & Cohen, L. A. (2001). Effects of a lycopene-rich diet on spontaneous and benzo[a]pyrene-induced mutagenesis in prostate, colon and lungs of the lacZ mouse. *Cancer Letters, 164*, 1-6.
- Hadley, C. W., Miller, E. C., Schwartz, S. J., & Clinton, S. K. (2002). Tomatoes, Lycopene, and Prostate Cancer: Progress and Promise. *Exp Biol Med,* 227(10), 869-880.
- Hauer-Jensen, M., Wang, J., Boerma, M., Fu, Q., & Denham, J. W. (2007).

 Radiation damage to the gastrointestinal tract: mechanisms, diagnosis, and management. *Current Opinion in Supportive and Palliative Care, 1*(1), 23-29.

- Haverkamp, J., Charbonneau, B., & Ratliff, T. L. (2008). Prostate inflammation and its potential impact on prostate cancer: A current review. *Journal of Cellular Biochemistry*, 103(5), 1344-1353.
- Heber, D. (2004). Phytochemicals beyond antioxidation. *Journal of Nutrition, 134*, 3175s-3176s.
- Helzlsouer, K. J., Erlinger, T. P., & Platz, E. A. (2006). C-reactive protein levels and subsequent cancer outcomes: Results from a prospective cohort study. *European Journal of Cancer*, *42*(6), 704-707.
- Hovdenak, N., Fajardo, L. F., & Hauer-Jensen, M. (2000). Acute radiation proctitis: a sequential clinicopathologic study during pelvic radiotherapy.

 International Journal of Radiation Oncology*Biology*Physics, 48(4), 1111-1117.
- Hovdenak, N., Karlsdottir, Aacute, sa, oslash, rbye, H., et al. (2003). Profiles and Time Course of Acute Radiation Toxicity Symptoms during Conformal Radiotherapy for Cancer of the Prostate. *Acta Oncologica, 42*(7), 741 748.
- Huang, M. T., Ghai, G., & Ho, C. T. (2004). Inflammatory Process and Molecular Targets for Antiinflammatory Nutraceuticals. *Comprehensive Reviews in Food Science and Food Safety*, *3*(4), 127-139.
- Hughes, L., Heydon, K., Edmonds, P., Roach, M., Rosenthal, S. A., Hammond, M. E. H., et al. (2002). *Cyclooxygenase 2 (COX-2) Expression in Locally*

- Advanced Prostate Cancer: Secondary Analysis of Radiation Therapy
 Oncology Group (RTOG) 86-10.
- Hwang, E.-S., & Bowen, P. E. (2004). Cell Cycle Arrest and Induction of Apoptosis by Lycopene in LNCaP Human Prostate Cancer Cells. *Journal of Medicinal Food, 7*(3), 284-289.
- Ivanov, N. I., Cowell, S. P., Brown, P., Rennie, P. S., Guns, E. S., & Cox, M. E. (2007). Lycopene differentially induces quiescence and apoptosis in androgen-responsive and -independent prostate cancer cell lines. *Clinical Nutrition*, *26*(2), 252-263.
- Jatoi, A., Burch, P., Hillman, D., Vanyo, J. M., Dakhil, S., Nikcevich, D., et al. (2007). A Tomato-Based, Lycopene-Containing Intervention for Androgen-Independent Prostate Cancer: Results of a Phase II Study from The North Central Cancer Treatment Group. *Urology*, 69(2), 289-294.
- Jian, L., Du, C.-J., Lee, A. H., & Binns, C. W. (2005). Do dietary lycopene and other carotenoids protect against prostate cancer? *International Journal of Cancer*, *113*(6), 1010-1014.
- Johnke, R. M., Edwards, J. M., Evans, M. J., Nangami, G. N., Bakken, N. T. G., Kilburn, J. M., et al. (2009). Circulating cytokine levels in prostate cancer patients undergoing radiation therapy:influence of neoadjuvant total androgen suppression. *In Vivo*, *23*, 827-834.
- Kanagaraj, P., Vijayababu, M. R., Ravisankar, B., Anbalagan, J., Aruldhas, M. M., & Arunakaran, J. (2007). Effect of lycopene on insulin-like growth

- factor-I, IGF binding protein-3 and IGF type-I receptor in prostate cancer cells. *Journal of Cancer Research and Clinical Oncology*, 133, 351-359.
- Keskek, M., Gocmen, E., Kilic, M., Gencturk, S., Can, B., Cengiz, M., et al. (2006). Increased Expression of Cyclooxygenase-2 (COX-2) in Radiation-Induced Small Bowel Injury in Rats. *Journal of Surgical Research*, 135(1), 76-84.
- Khachik, F., Carvalho, L., Bernstein, P. S., Muir, G. J., Zhao, D.-Y., & Katz, N. B. (2002). Chemistry, Distribution, and Metabolism of Tomato Carotenoids and Their Impact on Human Health. *Experimental Biology and Medicine*, 227(10), 845-851.
- Khor, L.-Y., Bae, K., Pollack, A., Hammond, M. E. H., Grignon, D. J.,
 Venkatesan, V. M., et al. (2007). COX-2 expression predicts prostatecancer outcome: analysis of data from the RTOG 92-02 trial. *The Lancet Oncology*, 8(10), 912 920.
- Kim, H., Bowen, P., Chen, L., Duncan, C., Ghosh, L., Sharifi, R., et al. (2003).

 Effects of tomato sauce consumption on apoptotic cell death in prostate benign hyperplasia and carcinoma. *Nutrition & Cancer, 47*(1), 40-47.
- Kim, L., Rao, A. V., & Rao, L. G. (2002). Effect of Lycopene on Prostate LNCaP Cancer Cells in Culture. *Journal of Medicinal Food*, *5*(4), 181-187.
- Kirschenbaum, A., Liu, X.-H., Yao, S., & Levine, A. C. (2001). The role of cyclooxygenase-2 in prostate cancer. [doi: DOI: 10.1016/S0090-4295(01)01255-9]. *Urology*, 58(2, Supplement 1), 127-131.

- König, J. E., Senge, T., Allhoff, E. P., & König, W. (2004). Analysis of the inflammatory network in benign prostate hyperplasia and prostate cancer.

 The Prostate, 58(2), 121-129.
- Konijeti, R., Henning, S., Moro, A., Sheikh, A., Elashoff, D., Shapiro, A., et al. (2010). Chemoprevention of prostate cancer with lycopene in the TRAMP model. *The Prostate*, 70(14), 1547-1554.
- Kramer, G., Mitteregger, D., & Marberger, M. (2007). Is Benign Prostatic

 Hyperplasia (BPH) an Immune Inflammatory Disease? *European Urology,*51(5), 1202-1216.
- Kucuk, O., Sarkar, F., Sakr, W., Djuric, Z., Pollak, M., Khachik, F., et al. (2001).
 Phase II randomized clinical trial of lycopene supplementation before radical prostatectomy. *Cancer Epidemiol Biomarkers and Prevention*, 10, 861-868.
- Kucuk, O., Sarkar, F. H., Djuric, Z., Sakr, W., Pollak, M. N., Khachik, F., et al.(2002). Effects of Lycopene Supplementation in Patients with LocalizedProstate Cancer. *Exp Biol Med*, 227(10), 881-885.
- Kun, Y., Lule, U. S., & Xiao-Lin, D. (2006). Lycopene: Its Properties and Relationship to Human Health. Food Reviews International, 22(4), 309 -333.
- Kuroda, K., Nakashima, J., Kanao, K., Kikuchi, E., Miyajima, A., Horiguchi, Y., et al. (2007). Interleukin 6 Is Associated with Cachexia in Patients with Prostate Cancer. *Urology, 69*(1), 113-117.

- Kurzrock, R. (2001). The role of cytokines in cancer-related fatigue. *Cancer,* 92(S6), 1684-1688.
- Lamprecht, M., Oettl, K., Schwaberger, G., Hofmann, P., & Greilberger, J. F. (2007). Several Indicators of Oxidative Stress, Immunity, and Illness Improved in Trained Men Consuming an Encapsulated Juice Powder Concentrate for 28 Weeks. *J. Nutr.*, 137(12), 2737-2741.
- Larsen, A., Bjorge, B., Klementsen, B., Helgeland, L., Wentzel-Larsen, T., Fagerhol, M., et al. (2007). Time patterns of changes in biomarkers, symptoms and histopathology during pelvic radiotherapy. *Acta Oncologica*, *46*(5), 639-650.
- Lee, A., Thurnham, D. I., & Chopra, M. (2000). Consumption of tomato products with olive oil but not sunflower oil increases the antioxidant activity of plasma. *Free Radical Biology and Medicine*, *29*(10), 1051-1055.
- Lehrer, S., Diamond, E. J., Mamkine, B., Droller, M. J., Stone, N. N., & Stock, R. G. (2005). C-reactive protein is significantly associated with prostate-specific antigen and metastatic disease in prostate cancer. *BJU International*, *95*(7), 961-962.
- Lewanski, C. R., & Gullick, W. J. (2001). Radiotherapy and cellular signalling. *The Lancet Oncology*, 2(6), 366-370.
- Li, L., Steinauer, K. K., Dirks, A. J., Husbeck, B., Gibbs, I., & Knox, S. J. (2003).

 Radiation-Induced Cyclooxygenase 2 Up-Regulation Is Dependent on

- Redox Status in Prostate Cancer Cells. *Radiation Research*, *160*(6), 617-621.
- Lieberman, R. (2001). Prostate cancer chemoprevention: Strategies for designing efficient clinical trials. *Urology*, *57*(4, Supplement 1), 224-229.
- Limpens, J., Schroder, F. H., de Ridder, C. M. A., Bolder, C. A., Wildhagen, M. F., Obermuller-Jevic, U. C., et al. (2006). Combined Lycopene and Vitamin E Treatment Suppresses the Growth of PC-346C Human Prostate Cancer Cells in Nude Mice. *J. Nutr., 136*(5), 1287-1293.
- Liu, A., Pajkovic, N., Pang, Y., Zhu, D., Calamini, B., Mesecar, A. L., et al. (2006). Absorption and subcellular localization of lycopene in human prostate cancer cells. *Mol Cancer Ther*, *5*(11), 2879-2885.
- Liu, X.-H., Kirschenbaum, A., Lu, M., Yao, S., Klausner, A., Preston, C., et al. (2002). Prostaglandin E2 Stimulates Prostatic Intraepithelial Neoplasia
 Cell Growth through Activation of the Interleukin-6/GP130/STAT-3
 Signaling Pathway. Biochemical and Biophysical Research
 Communications, 290(1), 249-255.
- Lu, Q.-Y., Hung, J.-C., Heber, D., Go, V. L. W., Reuter, V. E., Cordon-Cardo, C., et al. (2001). Inverse Associations between Plasma Lycopene and Other Carotenoids and Prostate Cancer. *Cancer Epidemiol Biomarkers Prev*, 10(7), 749-756.

- Matulka, R. A., Hood, A. M., & Griffiths, J. C. (2004). Safety evaluation of a natural tomato oleoresin extract derived from food-processing tomatoes. Regul Toxicol Pharmacol, 39(3), 390-406.
- McBride, W. H., Chiang, C.-S., Olson, J. L., Wang, C.-C., Hong, J.-H., Pajonk, F., et al. (2004). A Sense of Danger from Radiation. *Radiation Research*, *162*(1), 1-19.
- McClain, R. M., & Bausch, J. (2003). Summary of safety studies conducted with synthetic lycopene. *Regul Toxicol Pharmacol*, *37*(2), 274-285.
- McMillan, D. C., Talwar, D., Sattar, N., Underwood, M., St J O'Reilly, D., & McArdle, C. (2002). The relationship between reduced vitamin antioxidant concentrations and the systemic inflammatory response in patients with common solid tumours. *Clinical Nutrition*, *21*(2), 161-164.
- Milas, L., & Hanson, W. R. (1995). Eicosanoids and radiation. *European Journal of Cancer*, 31(10), 1580-1585.
- Mohanty, N. K., Saxena, S., Singh, U. P., Goyal, N. K., & Arora, R. P. (2005).

 Lycopene as a chemopreventive agent in the treatment of high-grade prostate intraepithelial neoplasia. *Urol Oncol, 23*(6), 383 385.
- Mollà, M., & Panés, J. (2007). Radiation-induced intestinal inflammation. *World Journal of Gastroenterology, 13*(22), 3043-3046.
- Nakashima, J., Tachibana, M., Horiguchi, Y., Oya, M., Ohigashi, T., Asakura, H., et al. (2000). Serum Interleukin 6 as a Prognostic Factor in Patients with Prostate Cancer. *Clin Cancer Res, 6*(7), 2702-2706.

- Neill, M. G., & Fleshner, N. (2006). An update on chemoprevention strategies in prostate cancer for 2006. *Current Opinion in Urology, 16*, 132-137.
- Nickel, J. C., Downey, J., Young, I., & Boag, S. (1999). Asymptomatic inflammation and/or infection in benign prostatic hyperplasia. *BJU International*, *84*(9), 976-981.
- Obermuller-Jevic, U. C., Olano-Martin, E., Corbacho, A. M., Eiserich, J. P., van der Vliet, A., Valacchi, G., et al. (2003). Lycopene Inhibits the Growth of Normal Human Prostate Epithelial Cells in Vitro. *J. Nutr., 133*(11), 3356-3360.
- Okunieff, P., Chen, Y., Maguire, D., & Huser, A. (2008). Molecular markers of radiation-related normal tissue toxicity. [10.1007/s10555-008-9138-7].

 Cancer and Metastasis Reviews, 27(3), 363-374.
- Paetau, I., Khachik, F., Brown, E. D., Beecher, G. R., Kramer, T. R., Chittams, J., et al. (1998). Chronic ingestion of lycopene-rich tomato juice or lycopene supplements significantly increases plasma concentrations of lycopene and related tomato carotenoids in humans. *Am J Clin Nutr, 68*(6), 1187-1195.
- Philip, M., Rowley, D. A., & Schreiber, H. (2004). Inflammation as a tumor promoter in cancer induction. [doi: DOI:
 10.1016/j.semcancer.2004.06.006]. Seminars in Cancer Biology, 14(6), 433-439.

- Polívková, Z., Šmerák, P., Demová, H., & Houška, M. (2010). Antimutagenic Effects of Lycopene and Tomato Purée. *Journal of Medicinal Food, 13*(6), 1-8.
- Porrini, M., & Riso, P. (2000). Lymphocyte Lycopene Concentration and DNA

 Protection from Oxidative Damage Is Increased in Women after a Short

 Period of Tomato Consumption. *J. Nutr., 130*(2), 189-192.
- Prasad, K. N., Cole, W. C., Kumar, B., & Prasad, K. C. (2002). Pros and cons of antioxidant use during radiation therapy. *Cancer Treatment Reviews*, 28(2), 79-91.
- Puri, T., Julka, P. K., Goyal, S., Nair, O., Sharma, D. N., & Rath, G. K. (2005).

 Role of natural lycopene and phytonutrients along with radiotherapy and chemotherapy in high grade gliomas. *J Clin Oncol (Meeting Abstracts)*, 23(16_suppl), 1561.
- Quintar, A. A., Doll, A., Leimgruber, C., Palmeri, C. M., Roth, F. D., Maccioni, M., et al. (2010). Acute inflammation promotes early cellular stimulation of the epithelial and stromal compartments of the rat prostate. *The Prostate*, 70(11), 1153-1165.
- Rafi, M. M., Yadav, P. N., & Reyes, M. (2007). Lycopene Inhibits LPS-Induced
 Proinflammatory Mediator Inducible Nitric Oxide Synthase in Mouse
 Macrophage Cells. *J Food Sci, 72*(1), S069-S074.

- Rao, A. V., & Agarwal, S. (1999). Role of lycopene as antioxidant carotenoid in the prevention of chronic diseases: A review. *Nutrition Research*, 19(2), 305-323.
- Rao, A. V., & Agarwal, S. (2000). Role of antioxidant lycopene in cancer and heart disease. *Journal of American College of Nutrition, 19*(5), 563-569.
- Rao, A. V., & Ali, A. (2007). Biologically Active Phytochemicals in Human Health:

 Lycopene. *International Journal of Food Properties*, 10(2), 279 288.
- Rao, A. V., Fleshner, N., & Agarwal, S. (1999). Serum and tissue lycopene and biomarkers of oxidation in prostate cancer patients: a case-control study.

 Nutrition & Cancer, 33(2), 159-164.
- Rao, A. V., & Rao, L. G. (2007). Carotenoids and human health. *Pharmacological Research*, *55*(3), 207-216.
- Rao, A. V., Ray, M. R., & Rao, L. G. (2006). Lycopene. *Adv Food Nutr Res, 51*, 99-164.
- Rao, A. V., & Shen, H. (2002). Effect of low dose lycopene intake on lycopene bioavailability and oxidative stress. *Nutr Res, 22*(10), 1125 1131.
- Ravenna, L., Sale, P., Di Vito, M., Russo, A., Salvatori, L., Tafani, M., et al. (2009). Up-regulation of the inflammatory-reparative phenotype in human prostate carcinoma. *The Prostate, 69*(11), 1245-1255.
- Rebbeck, T. R., Rennert, H., Walker, A. H., Panossian, S., Tran, T., Walker, K., et al. (2008). Joint effects of inflammation and androgen metabolism on

- prostate cancer severity. *International Journal of Cancer, 123*(6), 1385-1389.
- Riso, P., Brusamolino, A., Martinetti, A., & Porrini, M. (2006). Effect of a tomato drink intervention on insulin-like growth factor (IGF)-1 serum levels in healthy subjects. *Nutrition & Cancer*, *55*(2), 157.
- Royuela, M., Ricote, M., Parsons, M. S., García-Tuñón, I., Paniagua, R., & De Miguel, M. P. (2004). Immunohistochemical analysis of the IL-6 family of cytokines and their receptors in benign, hyperplasic, and malignant human prostate. *The Journal of Pathology*, 202(1), 41-49.
- Saada, H. N., Rezk, R. G., & Eltahawy, N. A. (2010). Lycopene protects the structure of the small intestine against gamma-radiation-induced oxidative stress. *Phytotherapy Research*, *24*(S2), S204-S208.
- Salinas, C. A., Kwon, E. M., FitzGerald, L. M., Feng, Z., Nelson, P. S., Ostrander,
 E. A., et al. (2010). Use of aspirin and other nonsteroidal antiinflammatory
 medications in relation to prostate cancer risk. *American Journal of Epidemiology*, 172(5), 578-590.
- Samant, R., & Gooi, A. C. C. (2005). Radiotherapy basics for family physicians.

 Potential toll for symptom relief. *Canadian Family Physician*, *51*, 1496-1501.
- Schenk, J. M., Kristal, A. R., Neuhouser, M. L., Tangen, C. M., White, E., Lin, D. W., et al. (2010). Biomarkers of Systemic Inflammation and Risk of

- Incident, Symptomatic Benign Prostatic Hyperplasia: Results From the Prostate Cancer Prevention Trial. *Am. J. Epidemiol.*, 171(5), 571-582.
- Schwarz, S., Obermuller-Jevic, U. C., Hellmis, E., Koch, W., Jacobi, G., & Biesalski, H.-K. (2008). Lycopene Inhibits Disease Progression in Patients with Benign Prostate Hyperplasia. *Journal of Nutrition*, *138*(1), 49-53.
- Sengupta, A., Ghosh, S., Das, R. K., Bhattacharjee, S., & Bhattacharya, S. (2006). Chemopreventive potential of diallylsulfide, lycopene and theaflavin during chemically induced colon carcinogenesis in rat colon through modulation of cyclooxygenase-2 and inducible nitric oxide synthase pathways. [Article]. *European Journal of Cancer Prevention August*, 15(4), 301-305.
- Sgambato, A., & Cittadini, A. (2010). Inflammation and cancer: a multifaceted link. *European Review for Medical and Pharmacological Sciences, 14*, 263-268.
- Shi, J., Kakuda, Y., & Yeung, D. (2004). Antioxidative properties of lycopene and other carotenoids from tomatoes: Synergistic effects. *Biofactors, 21*(1-4), 203.
- Shi, J., & Maguer, M. L. (2000). Lycopene in Tomatoes: Chemical and Physical Properties Affected by Food Processing. *Critical Reviews in Food Science and Nutrition*, *40*(1), 1 42.
- Shi, J., Qu, Q., Kakuda, Y., Yeung, D., & Jiang, Y. (2005). Stability and Synergistic Effect of Antioxidative Properties of Lycopene and Other

- Active Components. *Critical Reviews in Food Science and Nutrition, 44*(7), 559 573.
- Simone, C., B. II, Simone, N., L., Simone, V., & Simone, C., B. . (2007).

 Antioxidants and other nutrients do not interfere with chemotherapy or radiation therapy and can increase kill and increase survival, part 1.

 Alternative Therapies in Health and Medicine, 13(1), 22-28.
- Smyth, E. M., Grosser, T., Wang, M., Yu, Y., & FitzGerald, G. A. (2009).

 Prostanoids in health and disease. *J. Lipid Res., 50*(Supplement), S423-428.
- Song, M., & Kellum, J. A. (2005). Interleukin-6. *Critical Care Medicine*, 33(12), S463-S465.
- Srinivasan, M., Devipriya, N., Kalpana, K. B., & Menon, V. P. (2009). Lycopene:

 An antioxidant and radioprotector against [gamma]-radiation-induced
 cellular damages in cultured human lymphocytes. *Toxicology*, *262*(1), 43-49.
- Srinivasan, M., Sudheer, A. R., Pillai, K. R., Kumar, P. R., Sudhakaran, P. R., & Menon, V. P. (2007). Lycopene as a natural protector against [gamma]-radiation induced DNA damage, lipid peroxidation and antioxidant status in primary culture of isolated rat hepatocytes in vitro. *Biochimica et Biophysica Acta (BBA) General Subjects, 1770*(4), 659-665.

- Stahl, W., & Sies, H. (1992). Uptake of lycopene and its geometrical isomers is greater from heat-processed than from unprocessed tomato juice in humans. *J Nutr*, *122*, 2161-2166.
- Steinauer, K. K., Gibbs, I., Ning, S., French, J. N., Armstrong, J., & Knox, S. J. (2000). Radiation induces upregulation of cyclooxygenase-2 (COX-2) protein in PC-3 cells. *International Journal of Radiation*Oncology*Biology*Physics, 48(2), 325-328.
- Steiner, G. E., Djavan, B., Kramer, G., Handisurya, A., Newman, M., Lee, C., et al. (2004). The Picture of the Prostatic Lymphokine Network Is Becoming Increasingly Complex. *Reviews in Urology*, *4*(4), 171-177.
- Steuber, T., Helo, P., & Lilja, H. (2007). Circulating biomarkers for prostate cancer. *World Journal of Urology*, *25*(2), 111-119.
- Talvas, J., Caris-Veyrat, C., Guy, L., Rambeau, M., Lyan, B., Minet-Quinard, R., et al. (2010). Differential effects of lycopene consumed in tomato paste and lycopene in the form of a purified extract on target genes of cancer prostatic cells. *Am J Clin Nutr*, *91*(6), 1716-1724.
- Teh, B. S., Amosson, C. M., Mai, W. Y., McGary, J., Grant, W. H., & Butler, E. B. (2004). Intensity Modulated Radiation Therapy (IMRT) in the Management of Prostate Cancer. *Cancer Investigation*, 22(6), 913 924.
- Trumbo, P. R. (2005). Are there adverse effects of lycopene exposure. *The Journal of Nutrition*, *135*(8), 2060S-2061S.

- Twillie, D. A., Eisenberger, M. A., Carducci, M. A., Hseih, W.-S., Kim, W. Y., & Simons, J. W. (1995). Interleukin-6: A candidate mediator of human prostate cancer morbidity. *Urology*, *45*(3), 542-549.
- van Breemen, R. B., Xu, X., Viana, M. A., Chen, L., Stacewicz-Sapuntzakis, M., Duncan, C., et al. (2002). Liquid Chromatography-Mass Spectrometry of *cis* and all-*trans*-Lycopene in Human Serum and Prostate Tissue after Dietary Supplementation with Tomato Sauce. *J. Agric. Food Chem., 50*(8), 2214-2219.
- Wagenlehner, F. M. E., Elkahwaji, J. E., Ferran, A., Bjerklund-Johansen, T., Naber, K. G., Hartung, R., et al. (2007). The role of inflammation and infection in the pathogenesis of prostate carcinoma. *BJU International*, 100(4), 733-737.
- Walsh, D., Mahmoud, F., & Barna, B. (2003). Assessment of nutritional status and prognosis in advanced cancer: interleukin-6, C-reactive protein, and the prognostic and inflammatory nutritional index. Supportive Care in Cancer, 11(1), 60-62.
- Wang, W., Bergh, A., & Damber, J.-E. (2004). Chronic inflammation in benign prostate hyperplasia is associated with focal upregulation of cyclooxygenase-2, Bcl-2, and cell proliferation in the glandular epithelium.

 The Prostate, 61(1), 60-72.

- Wang, W., Bergh, A., & Damber, J.-E. (2005). Cyclooxygenase-2 Expression

 Correlates with Local Chronic Inflammation and Tumor Neovascularization
 in Human Prostate Cancer. *Clin Cancer Res*, 11(9), 3250-3256.
- Weisburger, J. H. (2002). Lycopene and Tomato Products in Health Promotion. *Exp Biol Med*, 227(10), 924-927.
- Wertz, K. (2009). Lycopene Effects Contributing to Prostate Health. *Nutrition and Cancer*, *61*(6), 775 783.
- Wertz, K., Siler, U., & Goralczyk, R. (2004). Lycopene:modes of action to promote prostate health. *Arch Biochem Biophys*, *430*, 127-134.
- Williams, C. S., Mann, M., & DuBois, R. N. (1999). The role of cyclooxygenases in inflammation, cancer, and development. *Oncogene*, *18*(55), 7908-7916.
- Wilson, J., & Balkwill, F. (2002). The role of cytokines in the epithelial cancer microenvironment. [doi: DOI: 10.1006/scbi.2001.0419]. *Seminars in Cancer Biology*, 12(2), 113-120.
- Withers, H. R. (1992). Biological Basis of Radiation Therapy for Cancer. *The Lancet*, 339(8786), 156-159.
- Wong, C. P., Bray, T. M., & Ho, E. (2009). Induction of proinflammatory response in prostate cancer epithelial cells by activated macrophages. *Cancer Letters*, *276*(1), 38 46.
- Yaping, Z., Wenli, Y., Weile, H., & Ying, Y. (2003). Anti-inflammatory and anticoagulant activities of lycopene in mice. *Nutrition Research*, *23*(11), 1591.

CHAPTER III DIETARY AND SERUM LYCOPENE LEVELS IN PROSTATE CANCER PATIENTS UNDERGOING IMRT

Submitting to: Cancer Causes & Control

Abstract

Objectives: To determine the effect of radiotherapy and supplementing three different amounts of tomato juice on serum lycopene levels in men with prostate cancer.

Methods: Dietary lycopene intake was calculated using the National Cancer Institute (NCI) Diet History Questionnaire. Gastrointestinal (GI) tolerance of tomato juice was evaluated using the NCI Cancer Therapy Evaluation Program: Common Toxicity Criteria v 2.0. Serum lycopene levels were determined using liquid chromatography-mass spectrometry. Both serum and dietary lycopene were used as covariates in data analysis.

Results: Daily tomato juice supplementation (4, 8 or 12 oz) was tolerated without any adverse GI effects. Serum lycopene decreased in control group participants, while increasing from 0.33±0.11 μg/mL to 0.41± 0.12 μg/ in the intervention group. No correlation between serum and dietary lycopene was observed. Control group participants lost weight, while participants in the intervention group did not.

Major conclusions: Tomato juice, a food source of lycopene (a potent antioxidant) is well tolerated and does increase serum lycopene levels in men with prostate cancer undergoing radiotherapy. Weight change should be monitored and evaluated in prostate cancer patients during radiotherapy. Larger clinical trials are needed to validate tomato juice use as a way to increase serum/dietary lycopene intake during radiotherapy in men with prostate cancer.

Introduction

Dietary phytochemicals (such as lycopene) may halt carcinogenesis by suppressing the initiating transforming and inflammatory processes (Dorai & Aggarwal, 2004). Epidemiological studies (Gann et al., 1999; Giovannucci, 1999, 2005; Vogt et al., 2002) have documented an inverse relationship between intake of lycopene rich foods and the development of prostate cancer. Researchers have also evaluated the impact of lycopene in patients with benign prostate hypertrophy (Edinger & Koff, 2006; Schwarz et al., 2008), and prostate cancer patients prior to initiating treatment (Bowen et al., 2002; Chen et al., 2001; Jatoi et al., 2007; Kim et al., 2003; Kucuk et al., 2001; Rao, Fleshner, & Agarwal, 1999) and after failure of treatment (Clark et al., 2006). Data on the effectiveness of lycopene supplementation during radiation therapy are lacking and researchers have proposed an urgency in evaluating the effectiveness of lycopene supplementation in prostate cancer patients undergoing radiation and androgen oblation therapies (Clinton, 2005; Davis et al., 2005), in order to formulate evidence-based recommendations for this population.

Serum antioxidant levels have been used to evaluate nutritional status and oxidative stress, since many of these antioxidants are essential and are utilized in physiological defense mechanisms (Polidori, Stahl, Eichler, Niestroj, & Sies, 2001). Oxidative stress generated secondary to chemotherapy and radiotherapy may further decrease tissue antioxidant levels, thereby increasing oxidative stress (Simone, Simone, Simone, & Simone, 2007), and may worsen cancer

treatment related side effects and existing subclinical or clinical nutrient deficiencies (Kucuk, 2002). A relatively new trend in cancer research is utilizing chemopreventive agents either by themselves or as adjuncts to halt disease progression, prevent secondary cancers or reduce treatment toxicities (Kucuk, 2002). In a case-controlled study, researchers demonstrated that prostate cancer patients have significantly lower (44%; p < 0.004) serum lycopene levels compared to matched controls (Rao, et al., 1999). Clinical trials evaluating the impact of lycopene supplementation during radiotherapy are limited, and since carotenoids have been reported to be preferentially utilized in oxidative states (Polidori, et al., 2001), we were interested in determining the level of serum lycopene in prostate cancer patients undergoing radiation therapy, and further evaluating the impact of tomato juice supplementation on serum lycopene levels. Consequently, we conducted this randomized controlled trial to evaluate the impact of three different volumes of tomato juice on serum lycopene levels during radiation therapy in men with localized prostate cancer. We also obtained diet history information to evaluate routine dietary lycopene intake among participants of this study. This study was part of a Phase I clinical trial evaluating the impact of three different volumes of tomato juice on various inflammatory markers and selected side effects of radiation therapy.

Materials and Methods

This randomized controlled trial conducted in men newly diagnosed with localized prostate cancer consisted of four study arms (control group participants

consumed their normal diets, and three intervention groups who received 4, 8 or 12 ounces (oz) of tomato juice in addition to their routine dietary intake). Participants were instructed to refrain from making any changes to their diet or consuming any nutritional (vitamin, mineral or other nutraceutical) supplements for the duration of the study. All participants received image guided intensity modulated radiation therapy (IMRT) to the prostate gland (treatment volume included prostate alone or prostate and seminal vesicles). In order to minimize time spent at the cancer center and avoid unnecessary follow-up visits, all assessments, blood draws and tomato juice administration (intervention groups only) were scheduled to coincide with each patient's physician, procedure planning or radiation therapy appointment times.

Seventeen men with newly diagnosed localized prostate cancer, receiving ≥ 72 Gray (Gy) of external radiation to the prostate alone or prostate and seminal vesicles completed this pilot study. Sample recruitment was consecutive and nonprobabilistic, since we included all patients meeting the eligibility criteria who were referred to the Hayworth Cancer Center at High Point Regional Health System (HPRHS) between April 2009 and October 2010. Participant screening was conducted in two stages: a preliminary screening was conducted by the radiation oncologist (BF) at the time of initial consult based on treatment area, type and dose. Patients meeting preliminary screening criteria signed a Health Insurance Portability and Accountability Act (HIPAA) form granting access to their medical information for detailed medical record review. In addition, patient

interviews were also conducted (MD) to thoroughly screen potential participants based on the study eligibility criteria. The Institutional Review Boards at the University of North Carolina Greensboro (UNCG) and HPRHS approved the study protocol. All participants provided written informed consent prior to enrolling in the study.

The *eligibility criteria* included – histologically confirmed localized prostate adenocarcinoma with no lymph node involvement or metastasis; normal immune, liver and renal function; and Eastern Cooperative Oncology Group (ECOG) performance score of 0 or 1. Exclusion criteria included: post prostatectomy, chemotherapy or other prior treatment for prostate cancer. Patients who received hormone therapy or participated in other treatment based clinical trials concurrently while receiving radiation therapy were also not eligible to participate in this study. Patients who were allergic to tomato products or red dye; had preexisting uncontrolled gastro-esophageal reflux disease and other malabsorptive disorders, or hyperkalemia; or routinely consumed (and unwilling to stop these supplements during treatment) fiber, saw palmetto, lycopene, omega-3 fatty acids/fish oil, EPA/DHA, vitamin C, E, A; β-carotene, flaxseeds and flaxseed oil supplements were also ineligible to participate in this study. Any potential participants consuming these specified supplements at the time of the initial interview were asked to discontinue these supplements until the end of treatment. This allowed for a wash-out period of several weeks before radiation treatment commenced. Verbal confirmation was obtained on the first day of

treatment to insure that the participants had discontinued use of supplements as previously instructed.

Names of each group (control, 4, 8, or 12 oz) and the participant number for that group were written on a piece of folded paper. The patient or accompanying family member then picked one of the folded pieces of paper and handed it to the investigator (MD) who transcribed the group assignment, writing it on the participant screening form and informing the participant and family members.

Assigned volume of tomato juice (4, 8 or 12 oz) was initiated two days prior to participants receiving the first dose of radiation therapy and continued daily until the last day of treatment. Participants in the intervention groups were provided with the measured amount of their assigned volume of tomato juice for consumption over the weekend and on holidays. Verbal confirmation of tomato juice consumption was obtained at the subsequent treatment day. Participants were instructed to consume tomato juice with either a meal or a snack with at least five grams of fat to facilitate lycopene absorption. During treatment days, participants were asked to consume their assigned volume of tomato juice and crackers (providing at least five grams of fat) after their radiation treatment, prior to leaving the cancer center, which was observed by the researcher (MD). Caloric contribution of the tomato juice and crackers was 121 kcal (4 oz), 146 kcal (8 oz) and 171 kcal (12 oz) daily (Table 3.3). Participants were asked to maintain their normal diets during the study. We used the National Cancer

Institute (NCI) Diet History Questionnaire (DHQ) ("Diet History Questionnaire,") to obtain participant's diet history and routine dietary lycopene intake once during the study period. The DHQ was given to the participants prior to starting radiation therapy and all participants except one returned the completed DHQ within two weeks of starting treatment. Participants were weighed by the nursing staff on the first day of treatment (baseline) and then once a week during treatment on a stationary platform scale in the radiation therapy department at HPRHS.

In order to evaluate (gastrointestinal) tolerance of tomato juice supplementation, the NCI Cancer Therapy Evaluation Program: Common Toxicity Criteria (CTC) v 2.0 ("Cancer therapy evaluation program. Common Toxicity Criteria, v2.0," 1999) was used. The CTC evaluates adverse events on a scale of 0-5 (TABLE 3.1) and any participant exhibiting toxicity grade ≥3 would have been withdrawn from the study by the researchers.

Standardized planning and filming protocol was followed, and participants were treated on a Linear Accelerator 2100 iX (Varian Medical Systems, CA). A diagnostic image obtained prior to each treatment was approved by the treating Radiation Oncologist. The prescribed radiation dose ranged between 72.50-79.20 Gy and treatment days varied between 29-44 days. Consequently, daily fractions ranged from 1.8-2.5 Gy.

Participants provided blood samples at baseline, midpoint (end of three weeks of treatment) and on the last day of treatment. Whole blood was collected in aluminum foil wrapped vacutainer tubes, and transported on ice to the

laboratory at UNCG. In order to minimize loss of lycopene, tubes were wrapped in aluminum foil immediately after collection and blood samples were processed under a yellow light. After centrifuging (3000 rpm x 20 min at 4°C) whole blood, the separated serum was divided into aliquot tubes labeled with patient ID, time point of blood draw (baseline, midpoint or endpoint) and stored at -80°C until analyzed. Serum and tomato juice samples were shipped under dry ice to Dr. Wei Jia's laboratory at the Kannapolis Research Campus, NC, for lycopene analysis using liquid chromatography-mass spectrometry (LC-MS).

Stock solution for lycopene standard was prepared by dissolving ~1 mg of the standard in 1 mL of chloroform followed by storage at -80 °C. Fresh calibration solutions were prepared each day from the stock solution. Calibration curves were obtained using freshly prepared lycopene standard solutions in the range of 0.005-10 μ g/mL in acetonitrile/methyl tert-butyl ether (MTBE) (1:1, v/v). Aliquots of 10 μ L of each standard solution were injected onto the column for LC analysis, and calibration curves were constructed by linear regression analysis of the area versus the concentration of lycopene.

Lycopene from tomato juice was extracted with an extraction mixture (hexane, methanol, acetone, 2:1:1, v/v/v, containing 2.5% butylated hydroxytoluen (BHT)). Tomato juice (200 μ L) was extracted with the extraction mixture (600 μ L). Samples were vortexed for one min and then centrifuged for 5 min at 13,000 rpm at 4°C. Aliquots of 10 μ L of the supernatant were diluted to 1 mL with the mobile phase for the HPLC injection. To prepare serum for lycopene

analysis, 300 μ L serum was mixed with same amount of ethanol. The mixture was extracted twice with 600 μ L of hexane containing 100mg/L BHT. The hexane extracts were collected after being centrifuged at 13,000 rpm for 5 min (4°C), combined, and evaporated to dryness under vacuum. The extract was reconstituted in 100 μ L of acetonitrile/MTBE (1:1, v/v). An aliquot of 10 μ L was injected onto the LC for lycopene measurement. An Agilent HPLC 1200 system equipped with a binary solvent delivery manager and a sample manager (Agilent Corporation, Santa Clara, CA, USA) is used with chromatographic separations performed on a 4.6 × 150 mm 5 μ m Agilent ZORBAX Eclipse XDB-C18 chromatography column. The flow rate was 1 mL/min. Elution solvent A was acetonitrile and solvent B was MTBE. The LC elution conditions are optimized with isocratic acetonitrile/MTBE (45:55, v/v). The column is maintained at 20°C. The detection wavelength was set at 472 nm. A 10 μ L aliquot reference standards and 10 μ L samples were injected onto the column, respectively.

On the last day of treatment, the researcher (MD) asked each participant about his reasons for participating in the study, whether he was glad to have participated, and if he would recommend any changes to the study protocol.

Descriptive statistics were used to analyze key participant and cancer related characteristics. Between group differences were detected using the non-parametric Wilcoxon Rank Sum analysis. We also conducted Spearman's correlation to determine any associations between dietary and serum lycopene levels and select lifestyle characteristics, and stepwise regression to detect the

strength of this relationship. Repeated measures were used to evaluate within group change over time. One-sided level of significance was established at p ≤ 0.05. Between-group comparisons with p between 0.051-0.10 were identified as demonstrating a trend towards significance.

Results

The flow diagram describes participant screening, final enrollment, attrition and reasons for participant exclusion (Figure 3.1). Seventeen men between the ages of 61 and 77 years completed this pilot study. Seventy-one percent of participants had clinical stage T1c prostate adenocarcinoma, followed by 18% with stage T2a and 6% each with stage T2b and T2c (data not shown). The largest proportion of our participants was non-Hispanic Whites (71%), followed by African-Americans (24%) and 6% of Peruvian descent. Despite a higher prevalence of prostate cancer reported among African-Americans ("Cancer Facts and Figures 2010," 2010), only 24% of our participants were African-American. This may be due to poor/limited access to health care and/or poor likelihood of localized prostate cancer diagnosis in African-Americans ("Cancer Facts and Figures 2010," 2010; Jemal, Siegel, Xu, & Ward, 2010). Height was statistically different between the control group and 12 oz group participants (p = 0.026); however, no differences were detected among groups for weight or body mass index (BMI). A majority (82%) of study participants in our study were married, and 71% reported less than or equivalent to a high school education (Table 3.2).

The most frequently (88%) observed medical diagnoses among study participants were hypercholesterolemia and hypertension. Thirty-five percent of participants had a diagnosis of diabetes but did not report following a "strict" diet for diabetes management/control. Only about 6% of participants reported still smoking compared with 65% of participants who reported consuming alcohol. One participant in the 4 oz treatment group reported changing his diet and activity significantly in the months prior to starting radiation therapy, but this was unrelated to his cancer diagnosis (data not shown). Overall, 71% of participants reported consuming one or more over-the-counter herbal/nutritional supplements at the time of enrollment. A multivitamin was the most commonly consumed (47%) supplement followed by fish oil/omega-3 fatty acid (29%). Other nutritional supplements consumed included vitamin D (18%), fiber (12%), vitamin C (12%), and 6% reported consuming either vitamin E, flax seeds, saw palmetto or coenzyme Q10 (Table 3.2). Twelve percent of the participants also reported using "eye" vitamin drops. Participants discontinued use of all supplements for the duration of the study. No statistically significant differences were detected between groups for supplement use. We did, however, find a significant positive correlation between supplement use and diagnosis of hypertension (r = 0.556, p = 0.009, n = 17) and observed a significant trend between supplement use and education level (r = 0.337, p = 0.093, n = 17).

Participants in all three treatment groups tolerated the tomato juice well with no reported gastrointestinal side effects (nausea, vomiting, or heartburn)

(data not shown). Participants in the intervention groups received 121 kcal (4 oz), 146 kcal (8 oz) and 171 kcal (12 oz) daily from the tomato juice and crackers (Table 3.3). When asked, some participants reported changes in their daily meal pattern or quantity of food intake in the meal closest to the time they received their tomato juice supplement at the cancer center. Not surprisingly, lycopene from tomato juice was statistically different between groups (Table 3.3). Macro and select micro nutrient analysis obtained from participant-reported DHQs are reported in Table 3.4. While a large variation was noted in energy intakes reported by participants among the four groups, caloric intake between only the control group and 8 oz group was statistically significant (p = 0.0476). Calculated dietary lycopene intake between the control group and 12 oz group (p = 0.0357) and 4 and 12 oz groups (p = 0.0286) were statistically significant. Intervention group participants received an estimated additional 8 mg, 18 mg, or 28 mg daily dose of lycopene from their assigned volume (4, 8 or 12 oz per day) of tomato juice. Total dietary lycopene (diet+supplemetal) intake was also statistically significantly different between groups (Table 3.4). We found a significant positive correlation (r = 0.517; p = 0.017; n = 17) with education level and reported dietary lycopene intake and a significant trend between hypertension and reported dietary lycopene intake (r = 0.335, p = 0.094, n = 17), indicating that participants with higher education and a diagnosis of hypertension reported higher intake of dietary lycopene.

No significant differences in body weight were detected among participants in any group at the beginning of the study. While very little variation in body weight was observed among the 4 oz and the 12 oz intervention group participants from the beginning to the end of the study, participants in the control group lost nine pounds while participants in the 8 oz group gained about eight pounds (Table 3.5).

Mean serum lycopene level for all study participants at baseline was 0.30 μg/mL and ranged between 0 to 1.04 μg/mL. When baseline levels were evaluated by study group, participants in the 8 oz group had the highest baseline lycopene level (0.51±0.17 µg/mL). Two participants in both 4 and 12 oz groups had no detectable serum lycopene at baseline. We detected a significant decrease (p = 0.009) in serum lycopene levels among control group participants over time, but not any of the intervention groups. However, overall serum lycopene levels increased with daily tomato juice supplementation. Intervention group mean serum lycopene increased from 0.33±0.11 µg/mL at baseline to 0.41± 0.12 µg/mL at endpoint. Participants in the 4 oz group demonstrated a progressive, yet non-significant decrease, while 8 oz group participants demonstrated a significant increase (p = 0.056 comparing percent change from baseline to midpoint and p = 0.095 comparing percent change from baseline to endpoint) in serum lycopene throughout treatment. All participants in the 12 oz group had measureable levels at midpoint and endpoint versus only one at baseline. We detected significant between-group differences in serum lycopene

levels at various time points (Table 3.4). Total lycopene intake per kilogram of body weight varied considerably: 0.07 mg/Kg (control group), 0.14 mg/Kg (4 oz), 0.27 mg/kg (8oz) and 0.53 mg/kg (12 oz group). These values were statistically significantly different (p < 0.01) between groups.

A significant positive correlation was detected between serum lycopene, weight (r = 0.525; p = 0.015; n = 17) and BMI (r = 0.541; p = 0.012; n = 17), and a negative correlation between serum lycopene and prior nutritional supplement use (r = -0.464; p = 0.030; n = 17). While not statistically significant, we also observed a negative trend between serum lycopene, smoking (r = -0.359; p = 0.078; n = 17) and diagnosis of hypertension (r = -0.356; p = 0.080; n = 17). However, in the final stepwise regression model, BMI (but not weight) and smoking were the two variables that explained 56% of the variance [F(2, 14) = 8.833, p = 0.003] in serum lycopene level.

The majority of participants (65%) reported taking part in the study because they "liked the idea of tomato juice helping reduce side effects of treatment." Forty seven percent of the participants also reported participating in the hopes that the results of the study would "help other people." Ninety-four percent of participants reported being glad that they participated in our clinical trial. Only one participant was dissatisfied, primarily because he was randomized into the control group and he wanted to participate in one of the intervention groups. While several participants remarked on the length of the DHQ while

completing the questionnaire, only one participant suggested shortening the DHQ during the exit interviews.

Discussion

We evaluated food based lycopene (tomato juice) supplementation in this randomized control trial in men undergoing radiation therapy for localized prostate cancer. The primary goal of this study was to evaluate serum lycopene levels during prostate radiation therapy and the impact of three different volumes of tomato juice administered daily on serum lycopene levels during radiation therapy. Secondary goals were to determine reported dietary lycopene intake in participants enrolled in our study and their reasons for participating in this clinical trial. We were able to demonstrate an increase in serum lycopene levels with daily tomato juice supplementation in men with prostate cancer undergoing radiation therapy. Serum lycopene levels ranging from 0.43 µmol/L in BPH (Schwarz, et al., 2008) to 0.64 µmol/L in prostate cancer patients (Bowen, et al., 2002) have been reported, similar to levels (0.55 µmol/L or 0.30 µg/mL) that we obtained at baseline. While Mayne et al (Mayne et al., 1999) have reported an association between serum and dietary lycopene intake, we did not observe this association. Inaccurate reporting of dietary lycopene, inconsistent digestion and absorption have been postulated as some of the reason why a poor correlation between dietary intake and serum levels of lycopene may be observed (Hadley, Miller, Schwartz, & Clinton, 2002). We also did not detect an association between alcohol intake, ethnicity, marital status or age as reported by Porrini and Riso (Porrini & Riso, 2005). While an optimum time (before or after radiation exposure) for initiating an antioxidant supplement has not been identified (S. L. Brown et al., 2010), we choose to initiate tomato juice supplement two days before radiation therapy started. Researchers have reported that lycopene reaches maximum concentration in the serum 15-48 hours after consumption (Gustin et al., 2004; Stahl & Sies, 1992), and our goal was to insure the presence of lycopene in the serum of participants at the commencement of radiation therapy, to evaluate the impact of radiation induced oxidative stress on serum lycopene levels with daily tomato juice intake during radiation treatment.

Dietary lycopene intake was estimated once using the DHQ. Mean reported dietary lycopene intake among participants was 7.24±1.48 mg/day (range 1.56-23.59 mg/day). Matulka et al (Matulka, Hood, & Griffiths, 2004) reported mean daily intake of lycopene to be about 8.2 mg/day, and lycopene intake ranging between 0.32-15.03 mg/d (Yong et al., 1994) has also been reported. We observed great variability among our participants in the reported consumption of lycopene. We found significant differences in reported dietary lycopene intake between the 12 oz group and 4 oz and control groups. Since most participants completed the DHQ within the first few weeks of treatment, we do not believe that the tomato juice supplement accounts for any of the differences in calculated dietary lycopene consumption. We found a positive correlation between education and dietary lycopene intake, perhaps indicating that participants with higher education included more variety (higher fruits and

vegetables) in their diet. While we did not find a correlation between education and dietary supplement use, a positive trend was observed, indicating that these men may be trying to lead a more "healthier" lifestyle, as reported by Wiygul et al who also reported similar supplement use in men diagnosed with prostate cancer (Wiygul et al., 2005).

We observed some weight loss in control group participants, while intervention group participants either maintained or gained weight (Table 3.5). Participants in the intervention groups received additional 121 kcal (4 oz), 146 kcal (8 oz) and 171 kcal (12 oz) daily (Table 3.4), which may account for perhaps 1.47 pounds weight gain in the 4 oz group, 1.61 pounds in the 8 oz group and 1.89 pounds in the 12 oz group if they maintained their usual dietary intake. Since we did not routinely monitor dietary intake beyond verbally ascertaining several times during the study that participants were maintaining their usual diet intake, we are unable to quantify whether any change in weight was related to the calories provided by the tomato juice and crackers or to other factors (such as fluid shifts/changes). In future studies, an isocaloric beverage with same amount of crackers should be provided to the control group participants to insure no discrepancies in caloric and volumetric intake. Similar trends in weight change have been reported in animal studies. Andic et al (Andic, Garipagaoglu, Yurdakonar, Tuncel, & Kucuk, 2009) reported weight loss in rats receiving radiation therapy without supplemental lycopene, and speculated that this may be a result of radiation induced anorexia and nausea. While control group

participants did not report any changes in appetite, other factors such as fatigue or biochemical changes (cytokine expression) may have contributed to their weight loss and should be monitored in these patients (Plata-Salamán, 1996).

Participants in our study tolerated the tomato juice supplemented daily during radiation therapy, as evidenced by no reported GI side effects (heartburn, nausea, or vomiting). Based on the responses received during the exit interview, our participants were enthusiastic about participating in a food-based trial, in order to reduce side effects of treatment. Reasons for participation were similar to those reported by Jatoi et al (Jatoi, et al., 2007), however, our participants did not report any adverse effects with daily tomato juice supplementation. Our participants had received no prior treatment for prostate cancer, and their disease was also localized, unlike the participant demographics reported by Jatoi et al (Jatoi, et al., 2007). Reasons for differences in tomato juice tolerance remain unclear.

This study has several strengths. To our knowledge, this is the first clinical trial to evaluate serum lycopene levels, tolerance of food-based lycopene supplementation and its impact on serum lycopene levels in men with localized prostate cancer undergoing radiation therapy. Lycopene intake was estimated using the DHQ. The DHQ was selected for estimating lycopene rich foods since it has been tested with older adults (Subar et al., 2001) and because it would require less work for the participants as compared to other available methods such as maintaining a food diary. It calculates average intake over a 12 month

period so that day-to-day or seasonal variations in dietary intake are included in the estimated amounts. Since we used a whole food approach instead of an isolated nutrient, there is a potential for greater beneficial effects due to the synergistic effect of all nutrients present (Norman et al., 2003) in tomato juice. We chose tomato juice as a vehicle for lycopene delivery because it is a processed tomato product, convenient to administer and consume. Additionally, this is a safe and effective way to increase consumption of vegetable intake among a population of men who typically have poor reported intake of fruits and vegetables.

This study had several limitations as well. These include a small sample size, which may have prevented us from detecting statistical significance and limits generalizability of these results. Due to variations in scheduling radiation treatments, blood was also not drawn from participants after an overnight fast.

Serum lycopene levels may be influenced by any lycopene intake in a recent meal, however, researchers have demonstrated that the serum concentration of carotenoids does not change significantly for up to four hours after a meal (E. D. Brown, Rose, Craft, Seidel, & Smith, 1989; Mejia & Arroyave, 1983; Mejia, Pineda, Noriega, Benitez, & Falla, 1984). Additionally, we only measured total lycopene content in the serum. Cis and trans isomers of lycopene should also be evaluated to determine clinical correlates.

To conclude, tomato juice supplementation to increase dietary lycopene intake as a means to improve serum lycopene is feasible and easily accepted

among this sample of prostate cancer patients undergoing radiation therapy. Since lower serum lycopene levels have been reported in prostate cancer patients (Rao, et al., 1999) and antioxidants may decrease due to oxidative stress generated by radiation therapy, processed tomato juice supplementation can be used to increase serum lycopene levels in men with prostate cancer undergoing radiation therapy. While we did not observe a dose-dependent response to tomato juice supplementation, we did observe a variable response to increased intake of tomato juice. Although we did not detect an association between dietary and serum lycopene, this lack of association may perhaps be explained by tissue lycopene uptake which we did not measure. These biomarker results need to be validated in larger clinical trials.

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Table 3.1. Supplement (tomato juice) toxicity evaluation ("Cancer therapy evaluation program. Common Toxicity Criteria, v2.0," 1999)

Criteria	Grade	Grade 1	Grade 2	Grade 3	Grade 4
Weight loss	None (<5%)	5-<10% of baseline, intervention not indicated	10-<20% of baseline; nutrition support indicated	> 20% of baseline; TF or TPN indicated	
Nausea	None	Able to eat reasonable intake	Intake significantly decreased but can eat	No significant intake	
Vomiting	None	1 episode in 24 hours over pretreatment	2-5 episodes in 24 hours; IV fluids indicated < 24 hrs	≥ 6 episodes in 24 hours, IV fluids or TPN indicated > 24 hrs	Life threatenin g conseque nces
Heartburn	None	Mild	Moderate	Severe	-

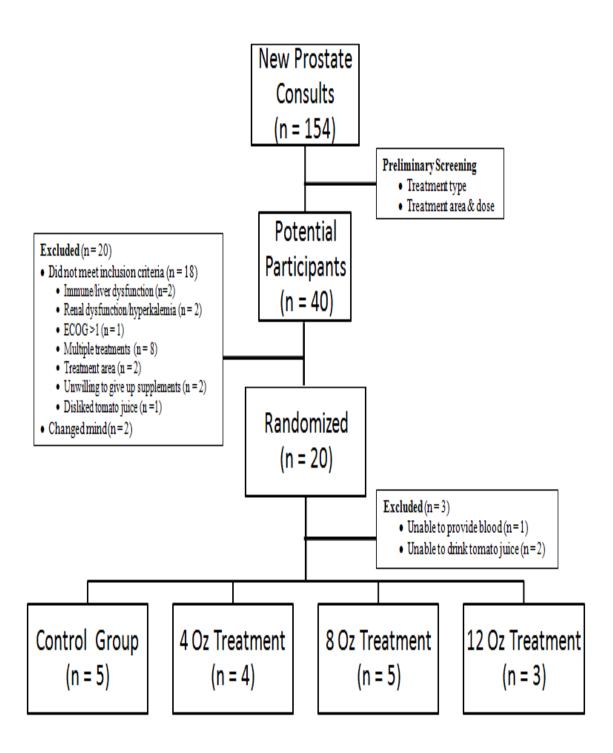


Figure 3.1: Flow diagram representing recruitment and randomization of study participants

Table 3.2. Participant characteristics. Data are reported as mean \pm standard error of the mean or number (%).

end of the mean of humber (70).	Control	4 Oz	8 Oz	12 Oz
	(n = 5)	(n=4)	(n = 5)	(n = 3)
Age	69.40±1.91	66.25±1.89	68.20±2.22	72.33±3.28
Height*	66.40± 0.87*	70.50±1.44	70.60±1.36	71.17±0.93*
Weight BMI (kg/m²) Ethnicity	167±13.11 27±1.97	193±14.61 27±1.30	221±30.74 30±3.22	168±28.41 23±4.10
African American	2 (40)	1 (25)	1 (20)	0
Caucasian	2 (40)	3 (75)	4 (80)	3 (100)
Other	1 (20)	0	0	0
Education				
< High School	1 (20)	2 (50)	2 (40)	0
High School	3 (60)	1 (25)	1 (20)	2 (67)
> High School	1 (20)	1 (25)	2 (40)	1 (33)
Marital Status	4 (00)	4 (400)	4 (00)	0 (07)
Married	4 (80)	4 (100)	4 (80)	2 (67)
Divorced	1 (20)	0	1 (20)	1 (33)
Diabetes	2 (40)	2 (50)	1 (20)	4 (22)
Yes No	2 (40)	2 (50)	1 (20)	1 (33)
Hypertension	3 (60)	2 (50)	4 (80)	2 (67)
Yes	5 (100)	4 (100)	4 (80)	2 (67)
No	0	0	1 (20)	1 (33)
Hypercholesterolemia	Ü	Ü	. (20)	. (66)
Yes	4 (80)	4 (100)	4 (80)	3 (100)
No	1 (20)	0	1 (20)	0
Routine use of supplements	` ,		,	
Yes	4 (80)	3 (75)	4 (80)	1 (33)
No	1 (20)	1 (25)	1 (20)	2 (67)
Supplements Evaluated				
Fiber [†]	0	0	2 (40)	0
Multivitamin [†]	3 (60)	1 (25)	4 (80)	1 (33)
Saw Palmetto [†]	0	0	1 (20)	0
Vitamin C [†]	0	1 (25)	1 (20)	0
Vitamin D [†]	2 (40)	0	1 (20)	0
Vitamin E [†]	0	0	1 (20)	0
Omega-3 Fatty acids/Fish oil	1 (20)	1 (25)	3 (60)	0
Flax seeds OR flaxseed oil	0	1 (25)	0	0
Coenzyme Q10 [†]	0	1 (25)	0	0
Eye Vitamin [†]	0	0	2 (40)	0

 $^{^{\}dagger}$ Data reported indicate affirmative responses from participants using these supplements. * p < 0.05 (one sided)

Table 3.3: Nutritional contribution from crackers and tomato juice provided to participants daily during IMRT

	4 Oz (n = 4)	8 Oz (n = 5)	12 Oz (n = 3)
6 crackers			
Calories (kcal)	96	96	96
Fat (gm)	5.4	5.4	5.4
Sodium (mg)	162	162	162
Potassium (mg)	12	12	12
Tomato Juice			
Calories (kcal)	25	50	75
Sodium (mg)	340	680	1020
Potassium (mg)	215	430	645
Lycopene (mg)	7.85 ^{ab}	17.52 ^{ac}	28.09 ^{bc}

ac; p = 0.011 (between group differences)

b; p < 0.001 (between group differences)

Table 3.4: Reported nutritional information obtained using the Diet History Questionnaire ("Diet History Questionnaire,"), measured and percent change in serum lycopene levels. Data are reported as mean ± standard error of the mean.

Nutrients	Control	4 Oz	8 Oz	12 Oz
	(n = 5)	(n = 4)	(n = 5)	(n = 3)
Energy (Kcal)	1634±276.8 ^a	2202±496.3	3015±668.2 ^a	2296±969.4
Protein (gm)	56±7.3	89±25.0	84±21.01	72±31.2
Total Fat (gm)	63±10.9	81±18.8	88±16.0	86±45.7
Saturated fat (gm)	17±3.42	25±6.75	27±4.82	23±10.1
Monounsaturated fat (gm)	24±4.02	32±6.84	38±8.20	34±18.5
Polyunsaturated fat (gm)	17±3.36	18±3.75	17±2.90	23±14.6
Cholesterol (mg)	187±39.95	278±110.50	306±63.48	160±56.4
Dietary Fiber	19±8.16	21±2.89	20±3.08	23±8.9
Dietary Lycopene	4.77±1.31 ^b	4.00±1.23 ^c	9.35±3.88	12.19±3.58 ^{bc}
(mg)		_		
Total (diet+TJ)	4.77 ^{de}	11.85 ^{fg}	26.87 ^{dfh}	40.27 ^{egh}
dietary lycopene				
(mg)				
Measured Serum				
Lycopene (µg/mL)				:
Baseline	0.209±0.07	0.295±0.21	0.514±0.17 ^l	0.082±0.08 ¹
Midpoint	0.137±0.05 ^J	0.272±0.08 ^k	0.534±0.09 ^{jkl}	0.069±0.01 ¹
Endpoint	0.167 ± 0.07^{m}	0.257±0.07	0.622 ± 0.25^{m}	0.253±0.20
Percent Change				
Serum lycopene				
Midpoint-Baseline	-135.21	-572.15	8.83	-30.74
Midpoint-Endpoint	-21.87	-3.34	-53.33	3.15
Endpoint-Baseline	-151.29	-97.79	-47.15	-15.21

p<0.05 (one sided)

Between group differences

 $^{abchil}p < 0.05 \quad ^{deg}p \leq 0.001 \quad ^{fm}p < 0.03 \quad \quad ^{j}p < 0.005 \quad \quad ^{k}p < 0.01$

Table 3.5: Measured weekly weight of participants undergoing IMRT. Data are reported as mean \pm standard error of the mean.

	Control (n = 5)	4 Oz (n = 4)	8 Oz (n = 5)	12 Oz (n = 3)
Starting Weight (lbs)	167±13.11	193±14.61	221±30.74	168±28.42
Week 1 (lbs)	167±12.94	195±14.01	220±29.75	168±26.82
Week 2 (lbs)	167±13.09	194±13.81	221±30.42	168±26.39
Week 3 (lbs)	168±13.52	194±14.08	221±30.10	169±27.02
Week 4 (lbs)	166±13.00	195±14.60	223±30.46	168±26.91
Week 5 (lbs)	167±13.37	195±13.91	222±30.48	168±27.06
Week 6 (lbs)	158±17.33	195±13.67	224±38.66	168±27.15
Week 7 (lbs)	158±17.19	195±13.74	229±38.20	168±26.24
Weight Change (lbs)	-9 lbs	+2 lbs	+8 lbs	None

One sided level of significance (p<0.05). No significant between group differences detected.

Lbs = pounds

References

- Andic, F., Garipagaoglu, M., Yurdakonar, E., Tuncel, N., & Kucuk, O. (2009).

 Lycopene in the Prevention of Gastrointestinal Toxicity of Radiotherapy.

 Nutrition and Cancer, 61(6), 784 788.
- Bowen, P., Chen, L., Stacewicz-Sapuntzakis, M., Duncan, C., Sharifi, R., Ghosh,
 L., et al. (2002). Tomato Sauce Supplementation and Prostate Cancer:
 Lycopene Accumulation and Modulation of Biomarkers of Carcinogenesis.
 Exp Biol Med, 227(10), 886-893.
- Brown, E. D., Rose, A., Craft, N., Seidel, K. E., & Smith, J. C., Jr. (1989).

 Concentrations of carotenoids, retinol, and tocopherol in plasma, in response to ingestion of a meal. *Clin Chem*, *35*(2), 310-312.
- Brown, S. L., Kolozsvary, A., Liu, J., Jenrow, K. A., Ryu, S., & Kim, J. H. (2010).

 Antioxidant Diet Supplementation Starting 24 Hours after Exposure

 Reduces Radiation Lethality. [doi: 10.1667/RR1716.1]. *Radiation*Research, 173(4), 462-468.
- Cancer Facts and Figures 2010. (2010). Retrieved May 14, 2010, from http://www5.cancer.org/docroot/STT/content/STT_1x_Cancer_Facts_Figures_2010.asp

- Cancer therapy evaluation program. Common Toxicity Criteria, v2.0. (1999).

 Retrieved [Accessed October 12, 2007
- Chen, L., Stacewicz-Sapuntzakis, M., Duncan, C., Sharifi, R., Ghosh, L., van Breemen, R., et al. (2001). Oxidative DNA damage in prostate cancer patients consuming tomato sauce based entrees as a whole-food intervention. *Journal of National Cancer Institute*, 93, 1872-1879.
- Clark, P. E., Hall, M. C., Borden, J. L. S., Miller, A. A., Hu, J. J., Lee, W. R., et al. (2006). Phase I-II prospective dose-escalating trial of lycopene in patients with biochemical relapse of prostate cancer after definitive local therapy.

 *Urology, 67(6), 1257-1261.
- Clinton, S. K. (2005). Tomatoes or Lycopene: a Role in Prostate Carcinogenesis? *J. Nutr.*, *135*(8), 2057S-2059.
- Davis, C., Clevidence, B., Swanson, C. A., Ziegler, R. G., Dwyer, J. T., & Milner, J. A. (2005). A Research Agenda for Lycopene/Tomato Supplementation and Cancer Prevention. *Journal of Nutrition*, 135(8), 2074S.
- Diet History Questionnaire. Retrieved Accessed November 6, 2007, from http://riskfactor.cancer.gov/DHQ/about/index.html
- Dorai, T., & Aggarwal, B. B. (2004). Role of chemopreventive agents in cancer therapy. *Cancer Letters*, *215*, 129-140.

- Edinger, M. S., & Koff, W. J. (2006). Effect of the consumption of tomato paste on plasma prostate-specific antigen levels in patients with benign prostate hyperplasia. *Brazilian Journal of Medical and Biological Research*, 39, 1115-1119.
- Gann, P. H., Ma, J., Giovannucci, E. L., Willett, W. C., Sacks, F. M., Hennekens,
 C. H., et al. (1999). Lower prostate cancer risk in men with elevated
 plasma lycopene levels: Result of a prospective analysis. *Cancer Research*, *59*, 1225-1230.
- Giovannucci, E. (1999). Tomatoes, tomato-based products, lycopene and cancer: Review of the Epidemiologic literature. *Journal of National Cancer Institute*, 91(4), 317 331.
- Giovannucci, E. (2005). Tomato Products, Lycopene, and Prostate Cancer: A

 Review of the Epidemiological Literature. *The Journal of Nutrition, 135*(8),

 S2030-S2031.
- Gustin, D. M., Rodvold, K. A., Sosman, J. A., Diwadkar-Navsariwala, V.,
 Stacewicz-Sapuntzakis, M., Viana, M., et al. (2004). Single-Dose
 Pharmacokinetic Study of Lycopene Delivered in a Well-Defined FoodBased Lycopene Delivery System (Tomato Paste-Oil Mixture) in Healthy
 Adult Male Subjects. *Cancer Epidemiol Biomarkers Prev, 13*(5), 850-860.

- Hadley, C. W., Miller, E. C., Schwartz, S. J., & Clinton, S. K. (2002). Tomatoes, Lycopene, and Prostate Cancer: Progress and Promise. *Exp Biol Med,* 227(10), 869-880.
- Jatoi, A., Burch, P., Hillman, D., Vanyo, J. M., Dakhil, S., Nikcevich, D., et al. (2007). A Tomato-Based, Lycopene-Containing Intervention for Androgen-Independent Prostate Cancer: Results of a Phase II Study from The North Central Cancer Treatment Group. *Urology*, 69(2), 289-294.
- Jemal, A., Siegel, R., Xu, J., & Ward, E. (2010). Cancer Statistics, 2010. *CA Cancer J Clin*, 60(5), 277-300.
- Kim, H., Bowen, P., Chen, L., Duncan, C., Ghosh, L., Sharifi, R., et al. (2003).

 Effects of tomato sauce consumption on apoptotic cell death in prostate benign hyperplasia and carcinoma. *Nutrition & Cancer, 47*(1), 40-47.
- Kucuk, O. (2002). Cancer chemoprevention. *Cancer and Metastasis Reviews,* 21, 189-197.
- Kucuk, O., Sarkar, F., Sakr, W., Djuric, Z., Pollak, M., Khachik, F., et al. (2001).
 Phase II randomized clinical trial of lycopene supplementation before radical prostatectomy. *Cancer Epidemiol Biomarkers and Prevention*, 10, 861-868.

- Matulka, R. A., Hood, A. M., & Griffiths, J. C. (2004). Safety evaluation of a natural tomato oleoresin extract derived from food-processing tomatoes. Regul Toxicol Pharmacol, 39(3), 390-406.
- Mayne, S. T., Cartmel, B., Silva, F., Kim, C. S., Fallon, B. G., Briskin, K., et al. (1999). Plasma lycopene concentrations in humans are determined by lycopene intake, plasma cholesterol concentrations and selected demographic factors. *J. Nutr.*, 129, 849-854.
- Mejia, L. A., & Arroyave, G. (1983). Determination of vitamin A in blood. Some practical considerations on the time of collection of the specimens and the stability of the vitamin. *Am J Clin Nutr, 37*(1), 147-151.
- Mejia, L. A., Pineda, O., Noriega, J. F., Benitez, J., & Falla, G. (1984).
 Significance of postprandial blood concentrations of retinol, retinol- binding protein, and carotenoids when assessing the vitamin A status of children.
 Am J Clin Nutr, 39(1), 62-65.
- Norman, H. A., Butrum, R. R., Feldman, E., Heber, D., Nixon, D., Picciano, M. F., et al. (2003). The Role of Dietary Supplements during Cancer Therapy. *J. Nutr.*, 133(11), 3794S-3799.
- Plata-Salamán, C. R. (1996). Anorexia during acute and chronic disease.

 Nutrition, 12(2), 69-78.

- Polidori, M. C., Stahl, W., Eichler, O., Niestroj, I., & Sies, H. (2001). Profiles of antioxidants in human plasma. *Free Radical Biology and Medicine*, *30*(5), 456-462.
- Porrini, M., & Riso, P. (2005). What Are Typical Lycopene Intakes? *The Journal of Nutrition*, 135(8), S2042-2046S.
- Rao, A. V., Fleshner, N., & Agarwal, S. (1999). Serum and tissue lycopene and biomarkers of oxidation in prostate cancer patients: a case-control study.

 Nutrition & Cancer, 33(2), 159 -164.
- Schwarz, S., Obermuller-Jevic, U. C., Hellmis, E., Koch, W., Jacobi, G., & Biesalski, H.-K. (2008). Lycopene Inhibits Disease Progression in Patients with Benign Prostate Hyperplasia. *Journal of Nutrition*, *138*(1), 49-53.
- Simone, C., B. II, Simone, N., L., Simone, V., & Simone, C., B. . (2007).

 Antioxidants and other nutrients do not interfere with chemotherapy or radiation therapy and can increase kill and increase survival, part 1.

 Alternative Therapies in Health and Medicine, 13(1), 22-28.
- Stahl, W., & Sies, H. (1992). Uptake of lycopene and its geometrical isomers is greater from heat-processed than from unprocessed tomato juice in humans. *J Nutr,* 122, 2161-2166.

- Subar, A., Thompson, F. E., Kipnis, V., Midthune, D., Hurwitz, P., McNutt, S., et al. (2001). Comparative Validation of the Block, Willett, and National Cancer Institute Food Frequency Questionnaires. *American Journal of Epidemiology*, 154(12), 1089-1099.
- Vogt, T. M., Mayne, S. T., Graubard, B. I., Swanson, C. A., Sowell, A. L.,
 Schoenberg, J. B., et al. (2002). Serum Lycopene, Other Serum
 Carotenoids, and Risk of Prostate Cancer in US Blacks and Whites. *Am. J. Epidemiol.*, 155(11), 1023-1032.
- Wiygul, J. B., Evans, B. R., Peterson, B. L., Polascik, T. J., Walther, P. J., Robertson, C. N., et al. (2005). Supplement use among men with prostate cancer. *Urology*, *66*(1), 161-166.
- Yong, L. C., Forman, M. R., Beecher, G. R., Graubard, B. I., Campbell, W. S., Reichman, M. E., et al. (1994). Relationship between dietary intake and plasma concentrations of carotenoids in premenopausal women: application of the USDA-NCI carotenoid food-composition database. *Am J Clin Nutr*, 60(2), 223-230.

CHAPTER IV

EFFECT OFTOMATO JUICE SUPPLEMENTATION ON INFLAMMATORY RESPONSE IN MEN UNDERGOING IMRT FOR PROSTATE CANCER

Submitting to: Cancer Letters

Abstract

Purpose: Evaluate selected inflammatory mediator levels in serum and the impact of different volumes of tomato juice on these, in prostate cancer patients undergoing radiotherapy.

Principle results: All participants exhibited inflammation at baseline. Increased c-reactive protein (CRP) and interleukin-6 (IL-6) was observed in control group, while decreases in serum CRP, IL-6 and prostaglandin E2 (PGE2) levels were observed in intervention groups. Serum tumor necrosis factor-α was not detected in these participants at any time point.

Major Conclusions: Systemic inflammation was observed pretreatment and tomato juice supplementation appears to decrease serum CRP, IL-6 and PGE2 levels in localized prostate cancer patients undergoing radiotherapy.

Introduction

Inflammation is now one of the acknowledged causes of carcinogenesis (Baniyash, 2006). However, the link between prostate cancer and inflammation is still considered "suggestive" (De Marzo, Marchi, Epstein, & Nelson, 1999; De Marzo et al., 2007; Palapattu et al., 2005; Schottenfeld & Beebe-Dimmer, 2006; Sciarra et al., 2008; Wagenlehner et al., 2007) since a direct causal relationship has not yet been demonstrated (Haverkamp, Charbonneau, & Ratliff, 2008). Biopsies obtained from patients with benign prostate hypertrophy and prostate cancer (Lehrer et al., 2005) do provide evidence of the presence of chronic inflammation in prostate carcinogenesis. Several pro-inflammatory mediators such as cytokines, chemokines, inflammatory enzymes, etc., have been implicated in fostering chronic inflammation (Aggarwal, Shishodia, Sandur, Pandey, & Sethi, 2006). Increased levels of c-reactive protein (CRP), interleukin-6 (IL-6), cyclooxygenase-2 (COX-2) and other inflammatory markers have been reported in various stages of prostate carcinogenesis (McArdle, McMillan, Sattar, Wallace, & Underwood, 2004; Pfitzenmaier et al., 2003). Researchers have recently also reported the up-regulation of several inflammatory markers in men with prostate cancer undergoing radiation therapy (Christensen et al., 2009; Johnke et al., 2009).

CRP, an acute phase protein (Ford, Liu, Mannino, Giles, & Smith, 2003; Helzlsouer, Erlinger, & Platz, 2006; Lehrer, et al., 2005), is a sensitive, yet non-specific marker of inflammation and is reported to be elevated in cancer patients

(Walsh, Mahmoud, & Barna, 2003). Radiation therapy has also been proposed as an independent cause of CRP elevation (Cengiz, Akbulut, Atahan, & Grigsby, 2001). CRP levels are inversely associated with circulating levels of lycopene along with other carotenoids, retinoids and antioxidants (Ford, et al., 2003; McMillan et al., 2002).

Tumor necrosis factor- α (TNF- α) is a key player in the body's inflammatory response, and can promote carcinogenesis by facilitating the growth of initiated cancer cells and by involving surrounding inflammatory cells (Balkwill, 2006). TNF- α is recognized not only for its role in carcinogenesis, but also for its ability to induce other inflammatory mediators such as NF-kB (Balkwill, 2006), and ultimately the acute phase response along with increasing prostaglandin, leukotriene and collegenase synthesis (Huang, Ghai, & Ho, 2004). TNF- α may induce a cascade of secondary mediators such as IL-1 that may be responsible for some of the physiological effects observed in various inflammatory conditions, independent of serum TNF- α levels (Kevin J. Tracey & Cerami, 1994).

IL-6 is a pleiotropic cytokine (Song & Kellum, 2005) that is involved in several key cellular functions such as proliferation, differentiation, angiogenesis and apoptosis (Culig, Steiner, Bartsch, & Hobisch, 2005), and has also been implicated in the development and progression of several tumors including the prostate (Culig, et al., 2005; Stark et al., 2009). Researchers have reported higher IL-6 levels in patients with prostate cancer, and have linked IL-6 along

with TNF-α and IL-1 with cancer progression (Bouraoui et al., 2008). While TNF-α and IL-1 stimulate the release of IL-6, it remains in the plasma longer than TNF-α and IL-1 and, consequently, is considered a key indicator of the activation of pro-inflammatory cytokines (Song & Kellum, 2005). Researchers have also recently demonstrated that IL-6 is upregulated in the plasma of men with prostate cancer undergoing radiation therapy (Christensen, et al., 2009; Johnke, et al., 2009).

Prostaglandin E2 (PGE2) is a key player in the immune suppression associated with inflammation (Ben-Baruch, 2006). It is synthesized by COX-2 and together they have demonstrated great potential as targets for cancer therapy (Ben-Baruch, 2006). COX-2 and PGE2 are over-expressed in both prostate intraepithelial neoplasia and prostate cancer (Kirschenbaum, Liu, Yao, & Levine, 2001). Radiation therapy may also induce inflammation and up-regulate PGE2 (Dorai & Aggarwal, 2004; Milas & Hanson, 1995; Steinauer et al., 2000) and other inflammatory markers. COX-2 can be induced by other proinflammatory cytokines such as TNF-α and IL-1 (Keskek et al., 2006). Sustained levels of COX-2 may play a key role in radiation-induced intestinal side effects/toxicities, by amplifying toxicities and by increasing their duration and severity (Keskek, et al., 2006). Upregulation of PGE2 has been implicated in radiation induced inflammatory changes in the bowel (Cole, Slater, Sokal, & Hawkey, 1993). Enhanced patient response to radiation therapy has been

proposed with the inhibition of COX-2 levels in prostate cancer patients (Khor et al., 2007).

Radiotherapy targets normal tissues and organs along with the cancer tissues by generating free radicals (Prasad, Cole, Kumar, & Prasad, 2002), inducing oxidative stress through generation of reactive oxygen species (ROS) and subsequent imbalance between pro-oxidants and antioxidants in the cell (Srinivasan et al., 2007). Tissue antioxidant levels can become depleted as a result of oxidative stress generated secondary to chemotherapy and radiotherapy, thereby increasing the level of oxidative stress (Simone, Simone, Simone, & Simone, 2007). Radiation therapy has been reported to induce cytokine expression (Friedman, 2002) along with triggering the acute phase response (Cengiz, et al., 2001; Koc, Taysi, Sezen, & Bakan, 2003). Researchers hypothesize that expression of radiation induced cytokines may be tissue specific (Stone, Coleman, Anscher, & McBride, 2003) and that these may serve as useful indicators of toxicity to the cells and tissues during radiation therapy for prostate cancer (Christensen, et al., 2009). Antioxidants can modulate cytokine levels in tissues, consequently reducing the inflammatory response (Okunieff et al., 2008). Several mechanisms have been proposed to explain the anti-inflammatory actions of tomato products/lycopene. These include antioxidative action, inhibition of IL-6, induction of phase II enzymes (Wertz, 2009), and modulation of the cyclo-oxygenase pathway (De Stefano et al., 2007; Sengupta, Ghosh, Das, Bhattacharjee, & Bhattacharya, 2006). We conducted this randomized controlled

trial to evaluate the levels of select serum markers of inflammation (CRP, TNF-α, IL-6 and PGE2) in men with localized prostate cancer undergoing radiation therapy. We were also interested in determining the impact of different volumes of tomato juice, dietary and serum lycopene on these select serum markers of inflammation during radiation therapy in this patient population.

Materials and Methods

Seventeen of 20 eligible patients with localized prostate cancer recruited between April 2009 and October 2010 completed the study. Participants were recruited consecutively at High Point Regional Health System's (HPRHS) Hayworth Cancer Center in High Point, North Carolina. Preliminary screening based on treatment area, type and dose was performed by the Radiation Oncologist (BF) at the time of initial consult. Participants meeting these criteria were referred to the primary investigator (MD) for detailed medical record review and patient interviews. Health Insurance Portability and Accountability Act and informed consent forms were obtained from participants. Institutional Review Board approvals for the study protocol were obtained at the University of North Carolina at Greensboro (UNCG) and HPRHS.

Patients were eligible for this study if they had a new diagnosis of localized prostate cancer and were scheduled to receive ≥ 72 Gray (Gy) external image guided intensity modulated radiation therapy (IMRT) to the prostate alone or prostate and seminal vesicles included in the treatment volume. Patients were ineligible to participate in this study if they had received prior treatment for

prostate cancer or were scheduled to receive other treatments concurrently with IMRT; or if they had abnormal liver, renal or immune function; hyperkalemia; uncontrolled gastro-esophageal reflux disease and other malabsorptive disorders; allergy to tomato products or red dye; ECOG score > 1; continued use of fiber supplements and other nutraceuticals with antioxidant properties (multivitamin, omega-3 fatty acids/fish oil, EPA/DHA, lycopene, vitamin C, E, A; B-carotene, flaxseeds and flaxseed oil) or action on the prostate (saw palmetto), and unwillingness to discontinue these for the study duration. Participant assessments, blood draws and tomato juice administration were scheduled during each patient's planning procedure and radiation therapy appointment time. Potential participants picked one out of four folded pieces of paper with different group name printed on each. The investigator (MD) recorded the group and individual participant ID on the participant screening form and informed the participant and family members of group assignment.

We used the National Cancer Institute (NCI) Diet History Questionnaire (DHQ) to obtain participant diet information and estimate dietary lycopene intake. The DHQ is a 124 item questionnaire that has been tested for reliability and validity. It is easy to use, takes about an hour to complete and includes questions on food portion sizes as well as dietary supplements ("Diet History Questionnaire,").

Tomato juice supplementation commenced two days before the first dose of radiation therapy and continued daily till the last day of treatment. During

treatment days, participants consumed tomato juice and crackers (providing at least five grams of fat, to facilitate lycopene absorption) prior to leaving the cancer center, in the presence of the researcher (MD). Tomato juice was provided to participants for consumption over weekends and holidays with the instruction to consume it with food providing at least five grams of fat.

Suggestions for snack and meals containing at least five grams of fat were discussed with participants. Verbal verification of tomato juice consumption was obtained from participants at their subsequent treatment day.

Blood samples were collected at baseline, at the end of three weeks of treatment (midpoint) and on the last day of treatment (endpoint). In order to protect serum lycopene from light, serum tubes for whole blood collection were wrapped in aluminum foil. Due to the length of transport time (20 min) from collection site to processing site at UNCG, collected blood was transported on ice. Blood samples were processed (centrifuged at 3000 rpm x 20 min at 4°C) under a yellow light to further minimize loss of lycopene. Separated serum, divided into aliquot tubes, was stored at -80°C until analyzed. Each aliquot tube was labeled with time point of blood draw (baseline, midpoint or endpoint) and patient ID. Standard Enzyme-Linked Immuno Sorbent Assay (ELISA) kits were used to test for serum PGE2, CRP, TNF- α, and IL-6 levels at the UNCG laboratory using the manufacturer's (R&D) instructions. We also used control samples for each kit to establish quantitative controls. Optical density was measured at 450nM using the SynergyTM HT (Bio-Tek Instrument, Inc) multi-

detection microplate reader. The minimum detectable limits for PGE2, CRP, TNF-α, and IL-6 were 30.9 pg/mL, 0.010 ng/mL, 1.6 pg/mL and 0.70 pg/mL respectively. Since the CRP levels were not normally distributed, we log transformed the serum levels. We chose not to use PSA as an endpoint in this study, even though it has been used as an endpoint in several lycopene clinical trials (Ansari & Gupta, 2004; Barber et al., 2006; Edinger & Koff, 2006; Vaishampayan et al., 2007). Since radiation therapy results in a decline in PSA levels post treatment, we would not be able to identify if the observed effect was due to radiation therapy or lycopene.

Serum samples for lycopene analysis were packed in dry ice and transported to Dr. Wei Jia's laboratory at the Kannapolis Research Campus, NC. Samples were analyzed for lycopene using high performance liquid chromatography and mass spectrometry (LC-MS).

All study participants received image guided IMRT, on a Linear Accelerator 2100 iX (Varian Medical Systems, CA) following same standardized planning and filming protocol. For treatment, participants were placed supine on the treatment table with their head resting on a square sponge and legs placed in a "W" shaped cushion. Patients were instructed to keep hands away from the treatment area by placing their hands on their chest. Daily treatment fractions were delivered utilizing specified photons with a prescribed field treatment plan. Each treatment day a diagnostic image was obtained by radiation therapy staff

and approved by the treating radiation oncologist to verify proper positioning of the isocenters.

Key participant and cancer related characteristics were analyzed using descriptive statistics. Inflammatory mediator data are reported as mean \pm standard error of the mean. Between-group differences were examined using the Wilcoxon rank sum test, and within-group differences were detected with repeated measures. Spearman's rho correlation was used to evaluate an association between the inflammatory mediators, serum lycopene and select lifestyle (weight, age) and cancer related characteristics (PSA, gleason sum (GS), treatment dose, daily fractions). One-sided level of significance was established at p \leq 0.05, with significant trends observed between 0.051-0.10.

Results

Forty seven percent of participants had a GS score 7, and 71% participants had tumor stage T1cN0M0. The proportion of participants who received 2 Gy, 1.8 Gy and 2.5 Gy fractions daily were 53%, 29% and 18% respectively. Treatment dose ranged between 72.50-79.20 Gy (Table I). Serum lycopene levels during radiation therapy

Overall serum lycopene at baseline was 0.30 μ g/mL, and 0.33 \pm 0.11 μ g/mL in the intervention group at baseline. We did observe a small non-significant increase in serum lycopene levels with daily tomato juice supplementation, in the intervention group towards the end of the study (0.41 \pm 0.12 μ g/mL). A progressive statistically significant decrease (p = 0.009) in serum lycopene was observed in

control group participants (Table II). Participants with GS 6 had lower serum lycopene levels at baseline (0.21±0.11 µg/mL) compared to participants with GS 7 (0.39±0.11 µg/mL).

CRP was detected in the serum of all participants at baseline. Participants in the 12 oz group had the lowest CRP, while participants in the 8 oz group had the highest CRP levels at baseline (Table II). Additionally, participants with GS 6 had higher CRP levels at baseline compared with participants with GS 7 (8.24±0.59 vs 8.10±0.14). No statistically significant within group differences were detected. While an increase in CRP was observed in control group participants throughout treatment, CRP levels decreased in the intervention groups. A statistically significant difference (p = 0.018) at midpoint was observed between control group and 12 oz group participants (Table II). While we did not detect a statistical significance, we did observe a trend towards significance (p = 0.071) in the baseline CRP levels between 8 and 12 oz groups, and midpoint levels (p = 0.057) between 4 and 12 oz groups. A statistically significant positive correlation was detected between serum IL-6 and CRP levels at baseline (r = 0.417, p = 0.048, n = 17) and endpoints (r = 0.566, p = 0.009, n = 0.00917).

No TNF-α levels were detected in the sera of study participants. IL-6 was detected in the serum in all but two participants (n=1 each control group and 8 oz groups) at baseline. Overall baseline IL-6 among study participants was 2.15 pg/mL. Unlike CRP, participants with GS 7 had higher IL-6 levels at baseline

 $(2.46\pm0.76 \text{ vs } 1.87\pm0.54 \text{ pg/mL})$. Highest IL-6 level at baseline was observed in the 4 oz group. Interestingly, a progressive increase in IL-6 levels was observed in control group participants, while a progressive decrease was observed in the 4 oz group means. While IL-6 levels in both 8 and 12 oz participants at end point were lower than baseline (Table II), the within group differences for 12 oz group only were statistically significant when comparing percent change at baseline and endpoint with midpoint (p = 0.014). While we did not detect a statistical significance, we did observe a trend towards significance (p = 0.0952) in the midpoint IL-6 levels between 4 and 8 oz groups.

Control group participants had the highest level of PGE2 at all time points. No PGE2 was detected in the serum of two participants (n=1 each, 4 and 8 oz groups) at any time point. Participants with GS 6 had higher PGE2 levels at baseline compared with participants with GS 7 (514.56±120.26 vs 440.08 ± 217.68 pg/mL). While PGE2 levels in the 4 oz group were lower at endpoint compared to baseline (Table II), both 8 and 12 oz participants demonstrated a progressive decrease, but within group differences for only the 12 oz group were statistically significant (p = 0.001) when comparing percent change at baseline with midpoint and endpoint (p = 0.003). We did observe a trend towards significance in the baseline PGE2 levels between 4 oz and control groups (p = 0.0952) and between 4 and 12 oz groups (p = 0.0857). We detected no correlation between inflammatory markers, cancer characteristics (PSA, GS,

radiation dose or daily fractions) and dietary or serum lycopene (data not reported).

Discussion

This study evaluated the levels of selected inflammatory mediators during radiation therapy of the prostate and the impact of lycopene supplementation via tomato juice on these mediators of inflammation. Overall, control group participants demonstrated a progressive but non-significant increase in CRP and IL-6 levels and a slight non-significant decrease in PGE2 level throughout treatment. High CRP levels in patients undergoing radiation therapy have been reported by other researchers (Cengiz, et al., 2001; Koc, et al., 2003). Since CRP is virtually undetectable in healthy adults the midpoint elevations observed in the 4 oz group participants may also represent an increase in the acute phase response. A considerable decline in the endpoint CRP levels was observed in the 8 oz group (percent change -2976% between baseline and endpoint), and a much lower magnitude of response was observed in the 4 (change -64%) and 12 oz (change -70%) group CRP levels between baseline and endpoint. Daily lycopene consumed as tomato juice may explain these results. An inverse relationship has been previously been reported between CRP and serum lycopene by Ford et al (Ford, et al., 2003). Participants with lower GS had higher CRP levels. The clinical implications of this remain unclear at the present time.

Serum IL-6 levels mirrored CRP levels in all groups. A progressive, non-significant increase in IL-6 levels was observed in control group and a non-

significant decrease observed in the intervention group participants. Higher IL-6 levels observed in participants with higher GS have also been reported by Alcover et al. (Alcover et al., 2010). Other researchers have also reported elevated IL-6 levels in prostate cancer patients undergoing IMRT (Christensen, et al., 2009; Johnke, et al., 2009). However, Johnke et al (Johnke, et al., 2009) reported normalization of IL-6 levels to pre-treatment levels after about two weeks of IMRT, results that we did not observed in this study. IL-6 levels increased progressively until the end in our control group participants. While we did observe an elevation at midpoint (end of 3 weeks of treatment) in the 8 and 12 oz groups, endpoint levels were below baseline and we observed a progressive decline in 4 oz group participants, as well. This may indicate a physiological role of tomato juice supplementation, even though we did not observe a dose dependent response.

We observed high PGE2 levels at baseline prior to participants receiving radiation therapy, confirming role of inflammation in carcinogenesis (Kirschenbaum, et al., 2001). Baseline PGE2 levels were highest among control and 12 oz groups, almost four times higher than participants in the 4 oz group. The non-significant decrease observed in intervention group participants overtime, likely indicates a beneficial effect of tomato juice to these participants. Lack of PGE2 levels observed in two patients may be due to gene polymorphisms (Zhang, Dhakal, Lang, & Kadlubar, 2010).

While we did not detect any TNF- α in the serum of our participants, researchers have reported higher TNF-α levels in prostate cancer patients undergoing IMRT (Christensen, et al., 2009). However, some researchers have also reported that detecting serum TNF-α may be problematic, perhaps as a result of a short half-life or binding to its competitive inhibitor soluble receptor (Bossola et al., 2000). TNF-α levels may also be suppressed by non steroidal anti-inflammatory agents, production of TGF-β, IL-10 and PGE2 (K. J. Tracey & Cerami, 1993), and binding with receptors (Kevin J. Tracey & Cerami, 1994). Akimoto et al reported 85% of prostate cancer patients in their study had TNF-α levels below the detectable limit (Akimoto, Okumura, & Fuse, 1998). However, TNF-α may induce a cascade of secondary mediators such as IL-1, that may be responsible for some of the physiological effects observed in various inflammatory conditions, independent of the serum TNF-α levels (Kevin J. Tracey & Cerami, 1994). Researchers have also reported lower TNF-α levels in patients with localized disease compared with metastatic prostate cancer (Michalaki, Syrigos, Charles, & Waxman, 2004). Since we did not observe any TNF-α in the serum of our participants, we cannot conclusively state that TNF-α was not induced. Future studies may need to test TNF receptors, IL-1 and other downstream secondary inflammatory mediators.

Increase in CRP at mid point may be reflective of acute radiation toxicity among those participants (data not reported). A progressive decline in CRP and PGE2 and a statistically significant within group differences when comparing

baseline and endpoint with midpoint (p = 0.014) for the 12 oz group may perhaps be a result of higher lycopene intake. This relationship needs to be evaluated further in prostate cancer patients undergoing radiation therapy. We did not detect a correlation between the different volumes of tomato juice intake and the levels of specific inflammatory mediators in our study. As we did not exclude participants for using anti-inflammatory medication, future studies should control for this variable as has also been recommended by other researchers (Grainger et al., 2008).

While recent publications have demonstrated higher serum levels of different cytokines during radiation therapy, we believe this is the first study evaluating the impact of food-based lycopene supplementation on select inflammatory mediators (CRP, TNF- α , IL-6 and PGE2) during radiotherapy in prostate cancer patients. Impact of dietary lycopene supplementation on the level of inflammatory mediators during radiotherapy also needs to be evaluated in men with prostate cancer receiving hormone therapy along with more aggressive stages of cancer.

This study has several limitations. In order to minimize disruption of daily activities among the study participants, blood draws were scheduled around the time of their prescheduled radiation treatment appointment. Consequently, patients were not in a fasted state for the blood draws. The impact of non-fasted state on these test parameters remains unknown. A small sample size of this study limits generalization of these results and may have prevented us from

detecting statistical significance. Due to the emerging research interest in cytokine expression during radiation therapy in prostate cancer patients, specific reference values relating to the magnitude of increase that can be expected in this population are lacking.

In conclusion, as expected, systemic inflammation was observed in men with localized prostate cancer prior to treatment. Despite the small sample size and mixed inflammatory mediator levels observed among the treatment group, our findings suggest that 8-12 oz tomato juice may be helpful in blunting the inflammatory response during radiation therapy in this population. Larger clinical trials are needed to validate these results and to ascertain if food based lycopene supplementation is an acceptable adjunct during radiation therapy.

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Table 4.1. Participant cancer related characteristics. Data are reported as mean ± standard error of the mean.

± standard error of	Control	4 Oz	8 Oz	12 Oz
	(n = 5)	(n = 4)	(n = 5)	(n = 3)
PSA	7.33±1.83	5.97±0.88	7.09±0.94	7.79±1.27
0-4.99 ng/mL	3 (60)	2 (50)	0	0
5-9.99 ng/mL	1 (20)	2 (50)	4 (80)	3 (100)
10-14.99 ng/mL	1 (20)	0	1 (20)	0
Gleason Sum*	6.40±0.25	6.75±0.25	6.40±0.25	6.33±0.33
6	3 (60)	1 (25)	3 (60)	2 (67)
7	2 (40)	3 (75)	2 (40)	1 (33)
Tumor Stage				
$T1_{c}N_{0}M_{0}$	4 (80)	3 (75)	3 (60)	2 (67)
$T2_{a}N_{0}M_{0}$	1 (20)	1 (25)	0	1 (33)
$T2_bN_0M_0$	0	0	1 (20)	0
$T2_{c}N_{0}M_{0}$	0	0	1 (20)	0
Treatment Dose	74.60±0.86	78.40±0.80	76.58±1.25	77.33±0.67
(Gy)	(72.50-76.00)	(76.00-79.20)	(72.50-79.20)	(76.00-78.00)
Treatment Days	34±2.21	43±1.50	39±2.75	39±0.33

Level of significance (p<0.05). No statistical significance detected.

Table 4.2. Serum lycopene and inflammatory markers at three time points in all participants. Data are reported as mean ± standard error of the mean

participants. Data a	Baseline	Midpoint	Endpoint
Lycopene (µg/mL)			
Control*	0.209±0.07	0.137±0.05 ^b	0.167±0.07 ^e
4 oz	0.295±0.21	0.272±0.08 ^c	0.257±0.07
8 oz	0.514±0.17 ^a	0.534±0.09 ^{bcd}	0.622±0.25 ^e
12 oz	0.082±0.08 ^a	0.069±0.01 ^d	0.253±0.20
Log CRP			
Control	7.65±0.49	8.57±0.51 ^f	8.34±1.04
4 oz	7.96±0.25	8.07±0.14	7.53±0.36
8 oz	9.22±0.78	8.92±0.96	8.02±0.53
12 oz	7.60±0.32	7.37±0.22 ^f	7.18±0.10
IL-6 (pg/mL)			
Control	1.433±0.517	1.911±0.406	2.655±1.383
4 oz	3.442±1.263	1.844±0.891	1.580±0.586
8 oz	2.289±0.909	5.888±3.181	1.914±0.519
12 oz	1.374±0.619	2.094±0.398	1.073±0.171
PGE2 (pg/mL)			
Control	707.47±307.80	684.81±428.24	664.58±428.64
4 oz	188.03±78.42	277.21±185.76	170.40±98.38
8 oz	351.13±176.75	183.79±162.06	98.31±40.99
12 oz	702.16±221.48	510.71±101.29	244.56±69.61

Statistical significance: p < 0.05 one-sided

^{*}within group difference p = 0.009
a p <0.05 (between group differences)
c p <0.01 (between group differences)

^b p <0.005 (between group differences) ^{def} p <0.03 (between group differences)

References

- Aggarwal, B. B., Shishodia, S., Sandur, S. K., Pandey, M. K., & Sethi, G. (2006).

 Inflammation and cancer: How hot is the link? *Biochemical Pharmacology*,

 72(11), 1605 1621.
- Akimoto, S., Okumura, A., & Fuse, H. (1998). Relationship Between Serum levels of Interleukin-6, Tumor Necrosis Factor-α and Bone Turnover Markers in Prostate Cancer Patients. *Endocrine Journal*, *45*(2), 183-189.
- Alcover, J., Filella, X., Luque, P., Molina, R., Izquierdo, L., Auge, J. M., et al. (2010). Prognostic value of IL-6 in localized prostate cancer. *Anticancer Research*, *30*, 4369-4372.
- Ansari, M. S., & Gupta, N. P. (2004). Lycopene: A novel drug therapy in hormone refractory metastatic prostate cancer. *Urologic Oncology: Seminars and Original Investigations*, 22(5), 415 -420.
- Balkwill, F. (2006). TNF-α in promotion and progression of cancer. *Cancer and Metastasis Reviews*, *25*, 409 416.

- Baniyash, M. (2006). Chronic inflammation, immunosuppression and cancer:

 New insights and outlook. [doi: DOI: 10.1016/j.semcancer.2005.12.002].

 Seminars in Cancer Biology, 16(1), 80-88.
- Barber, N. J., Zhang, X., Zhu, G., Pramanik, R., Barber, J. A., Martin, F. L., et al. (2006). Lycopene inhibits DNA synthesis in primary prostate epithelial cells in vitro and its administration is associated with a reduced prostate-specific antigen velocity in a phase II clinical study. *Prostate Cancer & Prostatic Diseases*, *9*(4), 407-413.
- Ben-Baruch, A. (2006). Inflammation-associated immune suppression in cancer:

 The roles played by cytokines, chemokines and additional mediators.

 Seminars in Cancer Biology, 16(1), 38-52.
- Bossola, M., Muscaritoli, M., Bellantone, R., Pacelli, F., Cascino, A., Sgadari, A., et al. (2000). Serum tumour necrosis factor-α levels in cancer patients are discontinuous and correlate with weight loss. *European Journal of Clinical Investigation*, *30*(12), 1107-1112.
- Bouraoui, Y., Ricote, M., García-Tuñón, I., Rodriguez-Berriguete, G., Touffehi, M., Rais, N. B., et al. (2008). Pro-inflammatory cytokines and prostate-specific antigen in hyperplasia and human prostate cancer. *Cancer Detection and Prevention*, 32(1), 23 32.

- Cengiz, M., Akbulut, S., Atahan, I. L., & Grigsby, P. W. (2001). Acute phase response during radiotherapy. *International Journal of Radiation Oncology*Biology*Physics*, 49(4), 1093-1096.
- Christensen, E., Pintilie, M., Evans, K. R., Lenarduzzi, M., Ménard, C., Catton, C. N., et al. (2009). Longitudinal Cytokine Expression during IMRT for Prostate Cancer and Acute Treatment Toxicity. *Clinical Cancer Research*, 15(17), 5576-5583.
- Cole, A. T., Slater, K., Sokal, M., & Hawkey, C. J. (1993). In vivo rectal inflammatory mediator changes with radiotherapy to the pelvis. *Gut*, *34*(9), 1210-1214.
- Culig, Z., Steiner, H., Bartsch, G., & Hobisch, A. (2005). Interleukin-6 regulation of prostate cancer cell growth. *Journal of Cellular Biochemistry*, *95*(3), 497-505.
- De Marzo, A. M., Marchi, V. L., Epstein, J. I., & Nelson, W. G. (1999).

 Proliferative Inflammatory Atrophy of the Prostate: Implications for Prostatic Carcinogenesis. *Am J Pathol*, *155*(6), 1985-1992.
- De Marzo, A. M., Platz, E. A., Sutcliffe, S., Xu, J., Gronberg, H., Drake, C. G., et al. (2007). Inflammation in prostate carcinogenesis. *Nature Reviews, 7*, 256 269.

- De Stefano, D., Maiuri, M. C., Simeon, V., Grassia, G., Soscia, A., Cinelli, M. P., et al. (2007). Lycopene, quercetin and tyrosol prevent macrophage activation induced by gliadin and IFN-[gamma]. *European Journal of Pharmacology*, *566*(1-3), 192-199.
- Diet History Questionnaire. Retrieved Accessed November 6, 2007, from http://riskfactor.cancer.gov/DHQ/about/index.html
- Dorai, T., & Aggarwal, B. B. (2004). Role of chemopreventive agents in cancer therapy. *Cancer Letters*, *215*, 129-140.
- Edinger, M. S., & Koff, W. J. (2006). Effect of the consumption of tomato paste on plasma prostate-specific antigen levels in patients with benign prostate hyperplasia. *Brazilian Journal of Medical and Biological Research*, 39, 1115-1119.
- Ford, E. S., Liu, S., Mannino, D. M., Giles, W. H., & Smith, S. J. (2003). C-reactive protein concentration and concentrations of blood vitamins, carotenoids, and selenium among United States adults. *European Journal of Clinical Nutrition*, 57(9), 1157-1163.
- Friedman, E. J. (2002). Immune Modulation by Ionizing Radiation and its

 Implications for Cancer Immunotherapy. [Article]. *Current Pharmaceutical Design*, 8(19), 1765-1780.

- Grainger, E. M., Kim, H. S., Monk, J. P., Lemeshow, S. A., Gong, M., Bahnson, R. R., et al. (2008). Consumption of dietary supplements and over-the-counter and prescription medications in men participating in the Prostate Cancer Prevention Trial at an academic center. *Urologic Oncology:*Seminars and Original Investigations, 26(2), 125 132.
- Haverkamp, J., Charbonneau, B., & Ratliff, T. L. (2008). Prostate inflammation and its potential impact on prostate cancer: A current review. *Journal of Cellular Biochemistry*, 103(5), 1344-1353.
- Helzlsouer, K. J., Erlinger, T. P., & Platz, E. A. (2006). C-reactive protein levels and subsequent cancer outcomes: Results from a prospective cohort study. *European Journal of Cancer*, *42*(6), 704-707.
- Huang, M. T., Ghai, G., & Ho, C. T. (2004). Inflammatory Process and Molecular Targets for Antiinflammatory Nutraceuticals. *Comprehensive Reviews in Food Science and Food Safety, 3*(4), 127-139.
- Johnke, R. M., Edwards, J. M., Evans, M. J., Nangami, G. N., Bakken, N. T. G., Kilburn, J. M., et al. (2009). Circulating cytokine levels in prostate cancer patients undergoing radiation therapy:influence of neoadjuvant total androgen suppression. *In Vivo*, 23, 827-834.

- Keskek, M., Gocmen, E., Kilic, M., Gencturk, S., Can, B., Cengiz, M., et al. (2006). Increased Expression of Cyclooxygenase-2 (COX-2) in Radiation-Induced Small Bowel Injury in Rats. *Journal of Surgical Research*, 135(1), 76-84.
- Khor, L.-Y., Bae, K., Pollack, A., Hammond, M. E. H., Grignon, D. J.,
 Venkatesan, V. M., et al. (2007). COX-2 expression predicts prostatecancer outcome: analysis of data from the RTOG 92-02 trial. *The Lancet Oncology*, 8(10), 912 920.
- Kirschenbaum, A., Liu, X.-H., Yao, S., & Levine, A. C. (2001). The role of cyclooxygenase-2 in prostate cancer. [doi: DOI: 10.1016/S0090-4295(01)01255-9]. *Urology*, 58(2, Supplement 1), 127-131.
- Koc, M., Taysi, S., Sezen, O., & Bakan, N. (2003). Levels of some Acute-PhaseProteins in the Serum of Patients with Cancer during Radiotherapy.Biological & Pharmaceutical Bulletin, 26(10), 1494-1497.
- Lehrer, S., Diamond, E. J., Mamkine, B., Droller, M. J., Stone, N. N., & Stock, R.
 G. (2005). C-reactive protein is significantly associated with prostate-specific antigen and metastatic disease in prostate cancer. *BJU International*, 95(7), 961-962.

- McArdle, P. A., McMillan, D. C., Sattar, N., Wallace, A. M., & Underwood, M. A. (2004). The relationship between interleukin-6 and C-reactive protein in patients with benign and malignant prostate disease. *British Journal of Cancer*, *91*(10), 1755-1757.
- McMillan, D. C., Talwar, D., Sattar, N., Underwood, M., St J O'Reilly, D., & McArdle, C. (2002). The relationship between reduced vitamin antioxidant concentrations and the systemic inflammatory response in patients with common solid tumours. *Clinical Nutrition*, *21*(2), 161-164.
- Michalaki, V., Syrigos, K., Charles, P., & Waxman, J. (2004). Serum levels of IL-6 and TNF-[alpha] correlate with clinicopathological features and patient survival in patients with prostate cancer. *British Journal of Cancer*, *90*(12), 2312-2316.
- Milas, L., & Hanson, W. R. (1995). Eicosanoids and radiation. *European Journal of Cancer*, 31(10), 1580-1585.
- Okunieff, P., Swarts, S., Keng, P., Sun, W., Wang, W., Kim, J., et al. (2008).

 Antioxidants Reduce Consequences of Radiation Exposure. *Advances in Experimental Medicine and Biology, 614*, 165-178.

- Palapattu, G. S., Sutcliffe, S., Bastian, P. J., Platz, E. A., De Marzo, A. M., Isaacs, W. B., et al. (2005). Prostate carcinogenesis and inflammation: emerging insights. *Carcinogenesis*, *26*(7), 1170-1181.
- Pfitzenmaier, J., Vessella, R., Higano, C. S., Noteboom, J. L., Wallace Jr, D., & Corey, E. (2003). Elevation of cytokine levels in cachectic patients with prostate carcinoma. *Cancer*, *97*(5), 1211-1216.
- Prasad, K. N., Cole, W. C., Kumar, B., & Prasad, K. C. (2002). Pros and cons of antioxidant use during radiation therapy. *Cancer Treatment Reviews*, 28(2), 79-91.
- Schottenfeld, D., & Beebe-Dimmer, J. (2006). Chronic inflammation: A common and important factor in the pathogenesis of neoplasia. *CA Cancer J Clin,* 56, 69 83.
- Sciarra, A., Mariotti, G., Salciccia, S., Gomez, A. A., Monti, S., Toscano, V., et al. (2008). Prostate growth and inflammation. *The Journal of Steroid Biochemistry and Molecular Biology*, *108*(3-5), 254-260.
- Sengupta, A., Ghosh, S., Das, R. K., Bhattacharjee, S., & Bhattacharya, S. (2006). Chemopreventive potential of diallylsulfide, lycopene and theaflavin during chemically induced colon carcinogenesis in rat colon through modulation of cyclooxygenase-2 and inducible nitric oxide

- synthase pathways. [Article]. European Journal of Cancer Prevention August, 15(4), 301-305.
- Simone, C., B. II, Simone, N., L., Simone, V., & Simone, C., B. . (2007).

 Antioxidants and other nutrients do not interfere with chemotherapy or radiation therapy and can increase kill and increase survival, part 1.

 Alternative Therapies in Health and Medicine, 13(1), 22-28.
- Song, M., & Kellum, J. A. (2005). Interleukin-6. *Critical Care Medicine*, *33*(12), S463-S465.
- Srinivasan, M., Sudheer, A. R., Pillai, K. R., Kumar, P. R., Sudhakaran, P. R., & Menon, V. P. (2007). Lycopene as a natural protector against [gamma]-radiation induced DNA damage, lipid peroxidation and antioxidant status in primary culture of isolated rat hepatocytes in vitro. *Biochimica et Biophysica Acta (BBA) General Subjects, 1770*(4), 659-665.
- Stark, J. R., Li, H., Kraft, P., Kurth, T., Giovannucci, E., Stampfer, M. J., et al. (2009). Circulating prediagnostic interleukin-6 and C-reactive protein and prostate cancer incidence and mortality. *International Journal of Cancer,* 124(11), 2683-2689.
- Steinauer, K. K., Gibbs, I., Ning, S., French, J. N., Armstrong, J., & Knox, S. J. (2000). Radiation induces upregulation of cyclooxygenase-2 (COX-2)

- protein in PC-3 cells. *International Journal of Radiation*Oncology*Biology*Physics, 48(2), 325-328.
- Stone, H. B., Coleman, C. N., Anscher, M. S., & McBride, W. H. (2003). Effects of radiation on normal tissue: consequences and mechanisms. *The Lancet Oncology*, *4*(9), 529-536.
- Tracey, K. J., & Cerami, A. (1993). Tumor Necrosis Factor, Other Cytokines and Disease. *Annual Review of Cell Biology*, *9*(1), 317-343.
- Tracey, K. J., & Cerami, A. (1994). Tumor necrosis factor: A pleiotropic cytokine and therapeutic target. *Annual Reviews in Medicine*, *45*, 491 503.
- Vaishampayan, U., Hussain, M., Banerjee, M., Seren, S., Sarkar, F. H., Fontana, J., et al. (2007). Lycopene and Soy Isoflavones in the Treatment of Prostate Cancer. *Nutrition and Cancer*, *59*(1), 1 7.
- Wagenlehner, F. M. E., Elkahwaji, J. E., Ferran, A., Bjerklund-Johansen, T., Naber, K. G., Hartung, R., et al. (2007). The role of inflammation and infection in the pathogenesis of prostate carcinoma. *BJU International*, 100(4), 733-737.
- Walsh, D., Mahmoud, F., & Barna, B. (2003). Assessment of nutritional status and prognosis in advanced cancer: interleukin-6, C-reactive protein, and

- the prognostic and inflammatory nutritional index. Supportive Care in Cancer, 11(1), 60-62.
- Wertz, K. (2009). Lycopene Effects Contributing to Prostate Health. *Nutrition and Cancer*, *61*(6), 775 783.
- Zhang, J., Dhakal, I., Lang, N., & Kadlubar, F. (2010). Polymorphisms in inflammatory genes, plasma antioxidants, and prostate cancer risk. Cancer Causes and Control, 21(9), 1437-1444.

CHAPTER V

IMPACT OF TOMATO JUICE SUPPLEMENTATION ON ACUTE TREATMENT TOXICITY AND PERFORMANCE STATUS DURING IMRT FOR PROSTATE CANCER

Submitting to: The Cancer Journal: The Journal of Principles & Practice of Oncology

Abstract

Purpose: to evaluate the impact of three different volumes of tomato juice on several frequently reported acute side effects (diarrhea, proctitis and urinary urgency and frequency) of radiation therapy and performance status in men with localized prostate cancer.

Materials and Methods: The National Cancer Institute's Cancer Therapy

Evaluation Program: Common Toxicity Criteria v 2.0 was used to evaluate
tolerance of tomato juice; and Common Terminology Criteria for Adverse Events
was used to evaluate severity of side effects related to radiation therapy.

Performance status was assessed using the Eastern Cooperative Oncology

Group scale.

Results: Men with localized prostate cancer (tumor grade T_{1c-2c}N₀M₀) scheduled to receive external radiation to the prostate alone or prostate and seminal vesicles were recruited between April 2009 and October 2010. Seventy one percent of participant tumors were staged T_{1c}N₀M₀. Daily radiation fractions varied between 1.8-2.5 Gray. Control group participants reported higher (worse) performance status scores than participants in the intervention groups. A strong positive correlation was detected between performance status, reported alcohol intake and diagnosis of hypertension at baseline. No differences in urinary frequency and urgency were detected between control and intervention groups.

However, tomato juice supplementation (especially 12 oz tomato juice) appeared to have a protective effect on gastrointestinal acute toxicity (diarrhea and proctitis) for the first three weeks of treatment.

Discussion: Daily tomato juice supplementation during intensity modulated radiation therapy in men with localized prostate cancer resulted in a gastrointestinal protective effect for the first three weeks of treatment and in performance status though out treatment. These results need to be validated in larger clinical trials using 12 ounce tomato juice or equivalent tomato products.

Introduction

In 2010, 217,730 new cases of prostate cancer were projected in the US and more than 90% of these cases were expected to be loco-regional cases with 100% five-year survival (Jemal, Siegel, Xu, & Ward, 2010). This is significant considering primary treatment strategies in addition to active surveillance include surgery and radiation therapy, ("Cancer Facts and Figures 2010," 2010) and reducing side effects of therapy will enhance quality of life (Andic, Garipagaoglu, Yurdakonar, Tuncel, & Kucuk, 2009). Side effects of radiation therapy may be either acute or chronic. Acute side effects of radiation therapy are those observed during or within three months of radiation, while symptoms occurring after three months are classified as chronic(Hauer-Jensen, Wang, Boerma, Fu, & Denham, 2007) or late effects (Stone, Coleman, Anscher, & McBride, 2003). Despite improved biochemical control, intensity modulated radiation therapy (IMRT) may still cause toxicity (Christensen et al., 2009). While the timeliness, severity and type of expression is dependent on the site of the irradiated tissue and cytokine cascade activation (Bentzen, 2006; Stone, et al., 2003), rate of tissue renewal, presence of microvascular (diabetes, hypertension) auto-immune or collagen vascular diseases, and radiation dose (Andreyev, 2007; Okunieff, Chen, Maguire, & Huser, 2008) may also impact treatment tolerance. Preexisting medical conditions such as hypertension (HTN), coronary artery disease (CAD) and diabetes (DM) may actually increase the potential for higher toxicity

upon exposure to radiation therapy (Chon & Loeffler, 2002; Houterman, Janssen-Heijnen, Hendrikx, Berg, & Coebergh, 2006; Okunieff, Chen, et al., 2008).

The most common acute toxicity symptoms of pelvic radiation are gastrointestinal (GI) and genitourinary (GU) symptoms. De Meerleer et al reported that the incidence of acute GI toxicity was observed in about 37% of patients (Grade 1: 44% and Grade 2: 29%), while 38% patients reported mild to moderately severe GU toxicity (Grade 1: 47%, Grade 2: 36% and Grade 3: 7%) (De Meerleer et al., 2004). Other researchers (Fonteyne, Villeirs, Lumen, & De Meerleer, 2009; Lips et al., 2008) have reported similar incidence of acute toxicity in prostate cancer patients undergoing radiation therapy. Symptoms of early radiation toxicity may be subtle and transient, but they can impact quality of life and functional status of patients considerably during treatment (Hauer-Jensen, et al., 2007), even in patients with milder symptoms (Christiansen et al., 2007). While acute toxicities may be transient and of short duration, it is imperative to treat or minimize these, since acute toxicities have been reported to be one of the significant predictors of long term "chronic" toxicities (Denham et al., 1999; Hovdenak et al., 2003; O'Brien, 2001; Schultheiss et al., 1997).

Researchers have proposed that various cytokines may be predictive, prognostic or diagnostic markers of radiation toxicity (Okunieff, Chen, et al., 2008). The release of a "cytokine storm" has been suggested to occur immediately after tissue irradiation, and that the intensity of these cytokines may be a predictor of toxicities in patients (Okunieff, Chen, et al., 2008). Higher levels

of serum interleukin-6 (IL-6) has been reported in prostate cancer patients undergoing IMRT (Christensen, et al., 2009), and increased levels of IL-6 along with tumor necrosis factor-α (TNF-α) have been reported in men with prostate cancer who develop radiation-induced proctitis (Christiansen, et al., 2007). Other researchers have reported elevation of IL-6 and other cytokines in patients with radiation induced pathologies (Indaram, Visvalingam, Locke, & Bank, 2000; McBride, 1995). Prostaglandins have also been implicated in radiation induced enteritis, but the results are not conclusive (Lifshitz, Savage, Taylor, Tewfik, & Van Orden, 1982).

Antioxidants can modulate cytokine levels in tissues, consequently reducing the inflammatory response (Okunieff et al., 2008). The potential for using radioprotective compounds to limit radiation—induced toxicity by protecting normal tissues and enhancing the therapeutic benefits has been explored by researchers and clinicians (Grdina, Murley, & Kataoka, 2002; Weiss & Landauer, 2003). Synthetic antioxidants such as amifostine have been successfully used in head and neck, ovarian, and non-small cell lung cancer patients to reduce treatment related toxicities (Grdina, et al., 2002). This radioprotective effect is not limited to synthetic antioxidants (Weiss & Landauer, 2003). Several phytochemicals (genistein, caffeine etc.) Have been identified that have both antioxidant as well as radioprotective effects in vivo (Weiss & Landauer, 2003). We limited our focus to lycopene which is a potent antioxidant and anti-inflammatory agent (Huang, Ghai, & Ho, 2004; Rafi, Yadav, & Reyes, 2007).

Researchers (Saada, Rezk, & Eltahawy, 2010; Srinivasan, Devipriya, Kalpana, & Menon, 2009; Srinivasan et al., 2007) have demonstrated that lycopene can protect cells against γ-radiation induced cellular damage and decrease acute GI side effects (diarrhea) observed during pelvic radiation therapy (Andic, et al., 2009). We undertook this study to evaluate the impact of a food based source of lycopene (tomato juice) on the frequently reported acute GI (diarrhea and proctitis) and GU (urinary urgency and frequency) side effects and performance status in men undergoing radiation therapy for localized prostate cancer.

Materials and Methods

Participants were selected consecutively and non-probabilistically at High Point Regional Health System's (HPRHS) Hayworth Cancer Center in High Point, NC between April 2009 and October 2010. Of the 154 new prostate cancer referrals to the cancer center, 40 participants met the preliminary screening criteria based on treatment area, type and dose. Eighteen participants did not meet the other eligibility criteria and two participants expressed no interest in participating in the study. Of the twenty participants who were randomized into four treatment arms (control, 4, 8 or 12 oz tomato juice), two dropped out due to inability to consume the assigned volume of tomato juice (n = 1 each from 8 and 12 oz groups) and one participant (4 oz group) was withdrawn by the investigators due to inability to obtain access for blood draws. The study protocol was approved by the Institutional Review Boards at the University of North Carolina at Greensboro (UNCG) and HPRHS. All participants signed a Health

Insurance Portability and Accountability Act (HIPAA) and informed consent forms prior to study enrollment.

The *eligibility criteria* included: newly diagnosed localized prostate cancer patients scheduled to receive ≥72 Gray (Gy) IMRT, treatment volume included prostate alone or prostate and seminal vesicles; no concurrent hormones, chemotherapy or other treatments for cancer; Eastern Cooperative Oncology Group (ECOG) performance score of 0 or 1. Patients were ineligible to participate in this study if they had pre-existing uncontrolled gastro-esophageal reflux disease and other malabsorptive disorders, renal or liver disease; compromised immune system; hyperkalemia; allergy to tomato products or red dye; or routinely used and unwilling to stop these supplements- fiber, saw palmetto, lycopene, omega-3 fatty acids/fish oil, EPA/DHA, vitamin C, E, A; β-carotene, flaxseeds and flaxseed oil supplements. In order to minimize time spent at the cancer center and to avoid additional follow-up visits, assessments, blood draws and tomato juice administration were scheduled around each patient's appointment times (planning, radiation therapy, etc.).

A Linear Accelerator 2100 ix (Varian Medical Systems, CA) was used to provide image guided IMRT to all participants in the study. Standardized planning and treatment protocols were followed for radiation therapy delivery under close supervision of the treating radiation oncologist. Researchers (MD) assessed performance status weekly using the ECOG performance scale. The NCI Cancer Therapy Evaluation Program: Common Toxicity Criteria (CTC) v 2.0 was used to

assess any GI adverse effects (nausea, vomiting, heartburn) from tomato juice supplementation, and the NCI Common Terminology Criteria for Adverse Events (CTCAE) was used to evaluate severity of side effects (urinary frequency and urgency, diarrhea and proctitis) related to radiation therapy.

Participants in the three intervention groups commenced tomato juice consumption two days before treatment and continued drinking it daily until the last day of treatment. During week days, tomato juice and crackers containing five grams of fat (to aid in the absorption of lycopene) were provided at the cancer center. The researcher (MD) observed participants drinking the juice and eating the crackers each treatment day. Participants were provided containers of their assigned volume of tomato juice for weekends and holidays with the instructions to consume tomato juice with either a meal or a snack containing at least five grams of fat. A few examples of snacks and meals containing five grams of fat were discussed with participants. Verbal confirmation of tomato juice consumption was obtained on the next radiation therapy day.

Data are reported as group means ± standard error of the mean. We utilized the Wilcoxon rank sum test to detect between-group differences for performance status and side-effects of treatment, and repeated measures were used to detect within-group differences at different time points. Chi-square was used to detect differences in proportions. We also conducted Spearman's rho correlation to evaluate the association of performance status scores and treatment side effects with select lifestyle characteristics. Differences were

considered statistically significant at one-sided p ≤ 0.05, and a trend towards significance was observed with p between 0.051-0.10.

Results

Participants in our study were between 61 and 77 years of age, with no statistically significant differences in age distribution among participants in the four groups. Majority (71%) of our participants were Caucasian, 82% of participants were married, and 71% had at least a high school education. The TNM staging ranged from $t_{1c}n_0m_0$ to $t_{2c}n_0m_0$, with 71% of participant tumors in the t_{1c}n₀m₀ stage. Treatment radiation dose ranged from 7250-7920 cgy and treatment days varied between 29 (18%), 38 (41%), 39 (12%) and 44 (29%). Overall 88% participants had a diagnosis of HTN and hyperlipidema, 35% were diagnosed with CAD and DM. Ninety-four percent of participants reported either never smoking or quitting some time ago and 65% of reported currently consuming alcohol. Seventy-one percent of participants reported using one or more over-the-counter herbal/nutritional supplements prior to entering the study but agreed to stop taking them for the duration of the study (Table 5.1). No statistically significant between-group differences were detected for any of these variables.

Control group participants had higher ECOG scores with increasing cumulative doses of radiation. A consistent peak was observed at weeks 3-5 followed by a slight decline at week 6, as a result of one participant in this group completing therapy. No change in performance status was reported by

participants in the 12 oz group, while a slightly increased group mean was noted in the 8 oz group beginning week 6. However, no statistically significant betweengroup or within-group differences were detected (Table 5.2).

Utilizing the CTCAE measurement criteria, urinary frequency and urgency were evaluated using participant reported baseline or pretreatment levels. Statistically significant differences (p = 0.039) in urinary frequency were detected at week 5 between 4 and 8 oz groups. A significant trend was observed at week 5 between control group and 8 oz group participants (p = 0.059) and between 8 and 12 oz groups (p = 0.089). Urinary urgency was statistically significant at weeks 2 (p = 0.008) and 5 (p = 0.048) between control and 4 oz groups, and at weeks 5 (p = 0.004) and 7 (p = 0.029) between control and 8 oz groups. A significant trend was observed at week 2 between control group and 8 oz group participants (p = 0.083) (Table 5.3). No within-group differences were detected in control, 4 or 12 oz groups, but a trend towards significance was observed, in the 8 oz group for urinary frequency (p = 0.099) and urinary urgency (p = 0.069).

Statistically significant differences were observed in the reported incidence of diarrhea at weeks 6 and 7 (p = 0.029) between control and 8 oz groups. Participants in the 12 oz group reported a very minimal increase in stool frequency. While no within-group differences were observed in control and 12 oz groups for diarrhea, we observed a trend towards significance (p= 0.065) in 4 oz group and statistical significance (p = 0.000) in the 8 oz group. No incidence of proctitis was reported among participants in the 12 oz group. Despite reported

proctitis in the other three groups, no statistically significant differences were detected between or within-groups. Overall it appears that tomato juice supplementation exhibited a protective effect on reported diarrhea incidence during the first three weeks of treatment (Table 5.4).

The relationship between certain lifestyle factors (supplement usage, diagnosis of HTN, CAD, DM), tomato juice supplementation and symptoms was investigated using the Spearman's correlation. There was a positive correlation between ECOG score and alcohol intake (beginning week 3) and hypertension (baseline to week 7) and ECOG scores (Table 5.5). Prior dietary supplement use showed a statistically significant association with ECOG score at week 1 (r = 0.555, p = 0.010, n = 17) and 2 (r = 0.433, p = 0.041, n = 17), and a significant trend at baseline and weeks 3-5 (Data not shown). No significant correlation was observed between ECOG score and DM.

Discussion

The primary objective of this paper was to examine the impact of three different doses of tomato juice on the performance status and selected radiation therapy related side effects in men with localized prostate cancer compared with participants in the control group. We did not measure fatigue in our participants, but nursing staff observed higher fatigue in control group participants, which may be reflected in the higher ECOG scores observed in control group participants. While not statistically significant, these results are clinically relevant and have implications beyond our study population, if these results can be replicated in

larger trials. If consumption of tomato products can alleviate radiation therapy induced fatigue in patients, there is considerable potential for improved quality of life during treatment with daily consumption of at least 8-12 oz of tomato juice.

Overall no significant differences in urinary frequency and urgency were observed between control and intervention group participants. While some between-group differences were detected for both urinary frequency and urgency, these may not be good outcome measures. Recommendations to increase fluid intake during treatment, personal tolerance and individual level of discomfort would greatly impact the subjective assessment of these variables.

Tomato juice supplementation appears to have a GI protective effect for the first three weeks of treatment. While the incidence of diarrhea throughout radiation therapy appears to be lower in the control group, one participant reported taking Imodium® to manage his diarrhea, which resulted in a lower group mean for diarrhea (Table 5.4), but a higher weekly participant score (Grade 2: increased stool frequency, bleeding mucus discharge or rectal discomfort requiring medication; anal fissure) and group mean for proctitis (Table 5.4). Despite consistency in treatment planning and delivery, significant variability in the incidence and severity of side effects is not uncommon (Bentzen, 2006). Radiation therapy may induce new microscopic colitis or exacerbate preexisting GI conditions causing diarrhea observed during therapy. Such effects may account for the increase in the incidence of diarrhea observed in the 4 and 8 oz tomato juice groups. Participants in the 12 oz group demonstrated the best GI

protective response. No proctitis was observed in this group and the incidence of diarrhea was minimal. Only one participant in the 12 oz group reported higher stool frequency (change from every other day to daily bowel movement). Researchers have reported decreased severity of diarrhea and acute GI toxicity in rats that were supplemented with lycopene and underwent pelvic radiation (Andic, et al., 2009). Our small sample size may have precluded us from detecting statistical significance. We did detect a significant positive association between performance status (ECOG score) and the diagnosis of hypertension or consumption of alcohol and but not serum or total dietary lycopene, DM or CAD. Chon and Loeffler have also reported lower radiation tolerance in patients with a diagnosis of diabetes or hypertension (Chon & Loeffler, 2002).

To our knowledge, this is the very first study evaluating the tolerance of lycopene supplementation (from processed tomato juice) during radiation therapy in prostate cancer patients. We assessed the tolerance of three doses of tomato juice during radiotherapy and evaluated their impact on performance status as well as both GI and GU symptoms during radiotherapy. While we did not observe any GU protective effect, we did observe a GI protective effect in the intervention groups. Tomato juice was well tolerated by the participants of this study, with no reported incidence of nausea, vomiting or heartburn. This study also expands the scope of lycopene research from preventive to an effective adjunct during radiation therapy.

This study has several limitations. We used validated clinical assessment tools to monitor participant side effects of treatment and performance status. However, we did not find the criteria to be sensitive enough for the purposes of our study. For instance, when evaluating the incidence of diarrhea, participants with increased daily stool frequency of 1-3 stools were all classified on the CTCAE scale as grade 1 toxicity. Some of our participants reported no more than an additional stool per day over baseline, while other participants reported an increase of 2-3 stools per day. However, by using the CTCAE criteria, both groups of participants were classified as grade 1. More objective criteria to evaluate treatment toxicities need to be investigated for future clinical trials. Additionally, the small sample size of this study limits generalization of these results and also may have prevented us from detecting consistent statistically significant differences between groups.

Conclusions

Tomato juice supplementation had no effect on acute GU side effects of radiation treatment, but appeared to offer a GI protective effect in men with prostate cancer receiving at least 8-12 oz of tomato juice daily during radiation therapy. Larger clinical trials are needed to validate these results.

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Table 5.1. Frequency of lifestyle characteristics of study participants.

Table 6.11.1 Toquelley 61 me	Control	4 Oz	8 Oz	12 Oz
	(n = 5)	(n = 4)	(n = 5)	(n = 3)
Diabetes				_
Yes	2 (40)	2 (50)	1 (20)	1 (33)
No	3 (60)	2 (50)	4 (80)	2 (67)
Hypertension				
Yes	5 (100)	4 (100)	4 (80)	2 (67)
No	0	0	1 (20)	1 (33)
Cardiovascular Disease				
Yes	2 (40)	3 (75)	1 (20)	0
No	3 (60)	1 (25)	4 (80)	3 (100)
Hypercholesterolemia				
Yes	4 (80)	4 (100)	4 (80)	3 (100)
No	1 (20)	0	1 (20)	0
Smoker				
Yes	0	1 (25)	0	0
No	5 (100)	3 (75)	5 (100)	3 (100)
Alcohol Consumption				
Yes	3 (60)	1 (25)	5 (100)	2 (67)
No	2 (40)	3 (75)	0	1 (33)
Routine use of				
supplements				
Yes	4 (80)	3 (75)	4 (80)	1 (33)
No	1 (20)	1 (25)	1 (20)	2 (67)

p<0.05 (one-sided). No statistical significance detected.

Table 5.2. Reported weekly Eastern Cooperative Oncology Group performance status ("Eastern Cooperative Oncology Group Performance Status," 1982). Data are reported as mean ± standard error of the mean.

	Control	4 Oz		
	(n = 5)	(n = 4)	(n = 5)	(n = 3)
Week 1	0.20 ± 0.20	0.25 ± 0.25	0.20 ± 0.20	0.33 ± 0.33
Week 2	0.40 ± 0.25	0.25 ± 0.25	0.20 ± 0.20	0.33 ± 0.33
Week 3	0.60±0.25	0.25 ± 0.25	0.20±0.20	0.33 ± 0.33
Week 4	0.60 ± 0.25	0.25±0.25	0.20 ± 0.20	0.33 ± 0.33
Week 5	0.60 ± 0.25	0.25 ± 0.25	0.20 ± 0.20	0.33 ± 0.33
Week 6	0.33 ± 0.33	0.25 ± 0.25	0.25±0.25	0.33 ± 0.33
Week 7	0.67±0.67	0.25±0.25	0.25±0.25	0.33±0.33

p<0.05 (one-sided). No statistical significance detected.

Table 5.3. Reported weekly incidence of acute genitourinary side effects. Data are reported as mean ± standard error of the mean.

Urinary Control		4 Oz	8 Oz	12 Oz	
frequency	(n = 5)	(n = 4)	(n = 5)	(n = 3)	
Week 1	0.60 ± 0.60	0.25±0.25	0.40±0.25	0.00±0.00	
Week 2	0.40±0.25	0.75±0.25	0.60±0.25	0.00 ± 0.00	
Week 3	0.40±0.25	0.75 ± 0.25	0.40±0.25	0.33±0.33	
Week 4	0.40±0.25	0.50 ± 0.29	0.60±0.25	0.33 ± 0.33	
Week 5	0.40±0.25	0.25±0.25 ^a	1.20±0.20 ^a	0.33 ± 0.33	
Week 6	0.33±0.33	0.50 ± 0.29	1.00±0.41	0.00 ± 0.00	
Week 7	0.00 ± 0.00	0.75 ± 0.25	1.00±0.41	0.67±0.33	
Urinary Urgency					
Week 1	0.00 ± 0.00	0.50±0.29	0.40±0.25	0.33±0.33	
Week 2	0.00 ± 0.00^{b}	1.00±0.00 ^b	0.60±0.25	0.33 ± 0.33	
Week 3	0.40±0.25	0.50±0.29	0.80±0.20	0.67±0.33	
Week 4	0.40±0.25	0.75 ± 0.25	0.80±0.20	0.33±0.33	
Week 5	0.00 ± 0.00^{cd}	0.75±0.25 ^d	1.00±0.00 ^c	0.33±0.33	
Week 6	0.00 ± 0.00	0.75 ± 0.25	0.75±0.25	0.67±0.33	
Week 7	0.00±0.00 ^e	0.75±0.25	1.00±0.00 ^e	0.33±0.33	

Level of significance (p≤0.05).

 $^{bc}p \le 0.01$ $^{ade}p \le 0.05$

Table 5.4. Reported weekly incidence of acute gastrointestinal side effects. Data are reported as mean ± standard error of the mean.

Diarrhea	Control	4 Oz	8 Oz	12 Oz	
	(n = 5)	(n = 4)	(n = 5)	(n = 3)	
Week 1	0.20±0.20	0.00 ± 0.00	0.20±0.20	0.00±0.00	
Week 2	0.60±0.25	0.00 ± 0.00	0.40±0.25	0.00 ± 0.00	
Week 3	0.20±0.20	0.50 ± 0.29	0.60±0.25	0.00 ± 0.00	
Week 4	0.40±0.25	0.75±0.25	0.80±0.20	0.33±0.33	
Week 5	0.20±0.20	0.50 ± 0.29	0.80±0.20	0.33 ± 0.33	
Week 6	0.00 ± 0.00^{a}	0.75±0.25	1.25±0.00 ^a	0.33 ± 0.33	
Week 7	0.00 ± 0.00^{a}	0.75±0.25	1.00±0.00 ^a	0.33±0.33	
Proctitis					
Week 1	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00	
Week 2	0.00 ± 0.00	0.00 ± 0.00	0.20±0.20	0.00 ± 0.00	
Week 3	0.00 ± 0.00	0.00 ± 0.00	0.40±0.25	0.00 ± 0.00	
Week 4	0.00 ± 0.00	0.00 ± 0.00	0.20±0.20	0.00 ± 0.00	
Week 5	0.60 ± 0.40	0.50 ± 0.50	0.20±0.20	0.00 ± 0.00	
Week 6	0.67±0.67	0.50 ± 0.50	0.50±0.50	0.00 ± 0.00	
Week 7	0.67±0.67	0.50 ± 0.50	0.50 ± 0.50	0.00 ± 0.00	
Week 8	1.00±1.00	0.00 ± 0.00	0.67±0.67	0.00±0.00	

p≤0.05, one-sided

 $^{^{}a}$ p ≤ 0.05

Table 5.5: Spearman's rho correlations for ECOG score ("Eastern Cooperative Oncology Group Performance Status," 1982)

			,					
ECOG Score								
	Baseline	Wk1	Wk2	Wk3	Wk4	Wk5	Wk6	Wk7
	N=17	N=17	N=17	N=17	N=17	N=17	N=14	N=14
Alcohol	0.304	0.171	0.334	.485 [*]	.485 [*]	.485 [*]	.519 [*]	.536 [*]
Hypertension	.789**	.658**	.566**	.494*	.494*	.494*	.645**	.575 [*]

Correlation is significant at

**p≤0.01 level (1-tailed)

* p≤0.05 level (1-tailed)

References

- Andic, F., Garipagaoglu, M., Yurdakonar, E., Tuncel, N., & Kucuk, O. (2009).

 Lycopene in the Prevention of Gastrointestinal Toxicity of Radiotherapy.

 Nutrition and Cancer, 61(6), 784 788.
- Andreyev, J. (2007). Gastrointestinal symptoms after pelvic radiotherapy: a new understanding to improve management of symptomatic patients. *The Lancet Oncology, 8*(11), 1007-1017.
- Bentzen, S. M. (2006). Preventing or reducing late side effects of radiation therapy: radiobiology meets molecular pathology. *Nature Reviews Cancer*, *6*(9), 702-713.
- Cancer Facts and Figures 2010. (2010). Retrieved May 14, 2010, from http://www5.cancer.org/docroot/STT/content/STT_1x_Cancer_Facts_Figures_2010.asp
- Chon, B. H., & Loeffler, J. S. (2002). The Effect of Nonmalignant Systemic

 Disease on Tolerance to Radiation Therapy. *Oncologist*, 7(2), 136-143.
- Christensen, E., Pintilie, M., Evans, K. R., Lenarduzzi, M., Ménard, C., Catton, C. N., et al. (2009). Longitudinal Cytokine Expression during IMRT for Prostate Cancer and Acute Treatment Toxicity. *Clinical Cancer Research*, 15(17), 5576-5583.

- Christiansen, H., Saile, B., Hermann, R., Rave-Fränk, M., Hille, A., Schmidberger, H., et al. (2007). Increase of hepcidin plasma and urine levels is associated with acute proctitis and changes in hemoglobin levels in primary radiotherapy for prostate cancer. *Journal of Cancer Research and Clinical Oncology*, 133(5), 297-304.
- De Meerleer, G., Vakaet, L., Meersschout, S., Villeirs, G., Verbaeys, A.,

 Oosterlinck, W., et al. (2004). Intensity-modulated radiotherapy as primary
 treatment for prostate cancer: Acute toxicity in 114 patients. [doi: DOI:
 10.1016/j.ijrobp.2004.04.017]. International Journal of Radiation
 Oncology*Biology*Physics, 60(3), 777-787.
- Denham, J. W., O'Brien, P. C., Dunstan, R. H., Johansen, J., See, A., Hamilton, C. S., et al. (1999). Is there more than one late radiation proctitis syndrome? *Radiotherapy and Oncology*, *51*(1), 43-53.
- Eastern Cooperative Oncology Group Performance Status. (1982). Retrieved

 [Accessed August 11, 2007, from

 http://ecog.dfci.harvard.edu/general/perf_stat.html
- Fonteyne, V., Villeirs, G., Lumen, N., & De Meerleer, G. (2009). Urinary toxicity after high dose intensity modulated radiotherapy as primary therapy for prostate cancer. [doi: DOI: 10.1016/j.radonc.2009.03.013]. *Radiotherapy and Oncology*, 92(1), 42-47.
- Grdina, D. J., Murley, J. S., & Kataoka, Y. (2002). Radioprotectants: Current Status and New Directions. [Article]. *Oncology*, *63*, 2-10.

- Hauer-Jensen, M., Wang, J., Boerma, M., Fu, Q., & Denham, J. W. (2007).

 Radiation damage to the gastrointestinal tract: mechanisms, diagnosis, and management. *Current Opinion in Supportive and Palliative Care, 1*(1), 23-29.
- Houterman, S., Janssen-Heijnen, M. L. G., Hendrikx, A. J. M., Berg, H. A. v. d., & Coebergh, J. W. W. (2006). Impact of comorbidity on treatment and prognosis of prostate cancer patients: A population-based study. [doi: DOI: 10.1016/j.critrevonc.2005.08.003]. Critical Reviews in Oncology/Hematology, 58(1), 60-67.
- Hovdenak, N., Karlsdottir, Aacute, sa, oslash, rbye, H., et al. (2003). Profiles and Time Course of Acute Radiation Toxicity Symptoms during Conformal Radiotherapy for Cancer of the Prostate. *Acta Oncologica*, *42*(7), 741 748.
- Huang, M. T., Ghai, G., & Ho, C. T. (2004). Inflammatory Process and Molecular Targets for Antiinflammatory Nutraceuticals. *Comprehensive Reviews in Food Science and Food Safety, 3*(4), 127-139.
- Indaram, A. V. K., Visvalingam, V., Locke, M., & Bank, S. (2000). Mucosal cytokine production in radiation-induced proctosigmoiditis compared with inflammatory bowel disease. *The American Journal of Gastroenterology*, *95*(5), 1221-1225.
- Jemal, A., Siegel, R., Xu, J., & Ward, E. (2010). Cancer Statistics, 2010. *CA Cancer J Clin*, 60(5), 277-300.

- Lifshitz, S., Savage, J. E., Taylor, K. A., Tewfik, H. H., & Van Orden, D. E.

 (1982). Plasma prostaglandin levels in radiation induced enteritis.

 International Journal of Radiation Oncology*Biology*Physics, 8, 275-277.
- Lips, I., Dehnad, H., van Gils, C., Boeken Kruger, A., van der Heide, U., & van Vulpen, M. (2008). High-dose intensity-modulated radiotherapy for prostate cancer using daily fiducial marker-based position verification: acute and late toxicity in 331 patients. *Radiation Oncology, 3*(1), 15.
- McBride, W. H. (1995). Cytokine cascades in late normal tissue radiation responses. *International Journal of Radiation Oncology*Biology*Physics*, 33(1), 233-234.
- O'Brien, P. C. (2001). Radiation injury of the rectum. *Radiotherapy and Oncology*, *60*(1), 1-14.
- Okunieff, P., Chen, Y., Maguire, D., & Huser, A. (2008). Molecular markers of radiation-related normal tissue toxicity. [10.1007/s10555-008-9138-7].

 Cancer and Metastasis Reviews, 27(3), 363-374.
- Okunieff, P., Swarts, S., Keng, P., Sun, W., Wang, W., Kim, J., et al. (2008).

 Antioxidants Reduce Consequences of Radiation Exposure. *Advances in Experimental Medicine and Biology, 614*, 165-178.
- Rafi, M. M., Yadav, P. N., & Reyes, M. (2007). Lycopene Inhibits LPS-Induced Proinflammatory Mediator Inducible Nitric Oxide Synthase in Mouse Macrophage Cells. *J Food Sci, 72*(1), S069-S074.

- Saada, H. N., Rezk, R. G., & Eltahawy, N. A. (2010). Lycopene protects the structure of the small intestine against gamma-radiation-induced oxidative stress. *Phytotherapy Research*, *24*(S2), S204-S208.
- Schultheiss, T. E., Lee, W. R., Hunt, M. A., Hanlon, A. L., Peter, R. S., & Hanks, G. E. (1997). Late GI and GU complications in the treatment of prostate cancer. *International Journal of Radiation Oncology*Biology*Physics*, 37(1), 3-11.
- Srinivasan, M., Devipriya, N., Kalpana, K. B., & Menon, V. P. (2009). Lycopene:

 An antioxidant and radioprotector against [gamma]-radiation-induced
 cellular damages in cultured human lymphocytes. *Toxicology*, *262*(1), 43-49.
- Srinivasan, M., Sudheer, A. R., Pillai, K. R., Kumar, P. R., Sudhakaran, P. R., & Menon, V. P. (2007). Lycopene as a natural protector against [gamma]-radiation induced DNA damage, lipid peroxidation and antioxidant status in primary culture of isolated rat hepatocytes in vitro. *Biochimica et Biophysica Acta (BBA) General Subjects*, *1770*(4), 659-665.
- Stone, H. B., Coleman, C. N., Anscher, M. S., & McBride, W. H. (2003). Effects of radiation on normal tissue: consequences and mechanisms. *The Lancet Oncology*, *4*(9), 529-536.
- Weiss, J. F., & Landauer, M. R. (2003). Protection against ionizing radiation by antioxidant nutrients and phytochemicals. *Toxicology*, 189(1-2), 1-20.

CHAPTER VI

EPILOGUE

Nutrition care of the oncology patient has always been a passion of mine. This interest was sparked while I was doing my dietetic internship at the All India Institute of Medical Sciences in New Delhi, India. Patients were housed in cubicle like compartments in a large hall with glass partitions, sequestered from the outside world including friends and family. The diagnosis of cancer was a certain death sentence in the 1990's, and this, along with social isolation and ravages of the disease and treatment, led to poor nutritional intake and progressive weight loss. It boggles my mind to this day why the health care providers were surprised when they observed weight loss in these patients. While working as a dietitian in the US, the majority of the consults received for Medical Nutrition Therapy (MNT) were for "weight loss. Poor PO intake." However, I have observed more intensive monitoring of cancer patients receiving chemotherapy due to obvious impediments to intake such as nausea, vomiting and diarrhea. Radiation therapy patients, more specifically patients with head and neck cancer, were referred for MNT only after extensive weight loss. We are now more aware of the effects of treatment on normal tissue, cytokine milieu created as a result of treatment, and a host of other factors that contribute to decreased intake and the common outcomes of weight loss and overall treatment tolerance. Proactive nutrition care in cancer patients can help improve treatment tolerance and quality of life. Researchers are now beginning to explore nutrients as radiosensitizers to enhance the effect of radiation therapy on tumors while protecting normal tissue and reducing potential side effects of treatment.

My dissertation examined the effect of tomato juice, a rich source of lycopene, in prostate cancer patients undergoing radiation therapy. The overall goals of this project were to evaluate whether dietary lycopene increases serum lycopene levels, delays onset of some of the most common side effects of radiation therapy and consequently improves performance status during therapy. So why lycopene?

My interest in lycopene was piqued while researching the link between carotenoids and lung cancer for NTR 673:Research Methodology, in Spring 2007. Lycopene, a potent antioxidant, has been a source of fascination to researchers for several decades. Researchers as early as the 1950s have been utilizing lycopene to mitigate the effects of radiation (Forssberg, Lingen, Ernster, & Lindberg, 1959). Several epidemiological studies (Gann et al., 1999; Giovannucci, 1999, 2005; Vogt et al., 2002) have documented an inverse relationship between intake of lycopene rich foods and the development of prostate cancer. Using cell and animal studies as well as retrospective and prospective trials in humans, researchers have evaluated the impact of lycopene in patients with benign prostate hypertrophy (Edinger & Koff, 2006; Schwarz et al., 2008), high grade prostatic intraepithelial neoplasia (Mohanty, Saxena, Singh, Goyal, & Arora, 2005) and prostate cancer patients prior to initiating

treatment (Bowen et al., 2002; Chen et al., 2001; Jatoi et al., 2007; Kim et al., 2003; Kucuk et al., 2001; Rao, Fleshner, & Agarwal, 1999) or after treatment failure (Clark et al., 2006). However, there is paucity of research investigating the impact of lycopene/tomato products supplementation in prostate cancer patients undergoing treatment. Researchers have demonstrated several mechanisms of action for lycopene's effectiveness. Some of the mechanisms of action that are relevant to this project include inhibition of IL-6 expression and modulation of the cyclooxygenase pathway. Therefore, this research project was developed from research gaps identified in the areas of vulnerable populations, exposures, mechanisms and biomarkers (Davis et al., 2005).

Due to the exploratory nature of this research, we encountered several challenges. The primary challenge was lack of sufficient funding to approach multiple cancer centers for patient recruitment, compensating a professional laboratory for blood sample assessment, and providing compensation to participants. We also had to limit the number of inflammatory mediators that we could test due to limited funding. However, this project could not have been conducted without the funding support we did receive from the University of North Carolina at Greensboro's Faculty Grant.

Initially, when participant recruitment commenced, we had some challenges with potential participant appointment notifications. Since the lycopene study was outside the routine job responsibilities of the cancer center staff, I had to work with them to insure I was notified about patient appointment

times and any subsequent changes in their appointment schedule, in order not to miss meeting with the participants. After making a few repeat trips to the cancer center, and missing a few screening appointments, we devised a system of notification that overcame this obstacle.

We did not achieve our intended target of recruiting 40 total participants. This may have prevented us from observing statistical significance where statistical differences did exist. The possibility of reduced side effects from a food product consumed during radiation therapy did appear to be an incentive for at least some of the participants who volunteered for our study, as evidenced by participant enthusiasm and enrollment. It was a fascinating, educational experience working with the participants of this study. When asked during the exit interviews, 65% of participants reported enrolling in the study to reduce their side effects of treatment, and 47% of participants also reported participating for purely altruistic reasons. These men were extremely motivated and inquisitive about the impact of nutrition and nutritional supplements on prostate health and were further intrigued by the possibility of something as simple as a tomato making them "feel better" along with the possibility of "chemicals" in the blood that could predict their radiation response! Several participants requested additional information on risk vs benefits of supplements, and lowering sodium intake in their diet. Since the information requested would not have influenced the study results, information was provided to the participants when requested.

Due to the number of blood variables being tested, we had to insure that blood collection and processing procedures could be standardized. All collected blood samples were treated the same, thus, we did not add indomethacin to the blood samples to halt synthesis of prostaglandins as stipulated in the PGE₂ ELISA kit. Omitting this step may result in higher PGE₂ expression. Also considering the labile nature of the cytokines, blood processing methodology needs to be carefully considered, specifically the temperature at which blood samples should be processed. Since lycopene is affected by heat and light, we covered the collection tubes containing participant blood immediately with aluminum foil and placed them on ice post collection for transportation to the UNCG laboratory.

Based on the results of this study, participant comments and researcher observations, tomato juice was well tolerated by men over 60 years of age, diagnosed with prostate cancer, undergoing radiation therapy. Only two participants withdrew from the study due to inability to consume the assigned volume of tomato juice. We recognize that tomato juice may not be acceptable to all men as a vehicle for the delivery of dietary lycopene. Consequently, other tomato products providing similar amounts of lycopene may need to be substituted.

Participants in our study did demonstrate high systemic inflammation, which was not unexpected. We did not observe a significant correlation between inflammatory markers and side effects of treatment. The most noteworthy

difference that we did observe was performance status in the intervention groups. Control group participants reported higher fatigue than the intervention group participants and this was reflected in the reported performance status responses from week to week. Another challenge observed during the study was the measurement of urinary frequency and urgency. Considering the age of the participants and frequent complaints of difficulty urinating with this prostate pathology, and also since all participants in the study were instructed by the Radiation Oncologist to increase their fluid intake, it was difficult to truly assess if the urinary frequency and urgency complaints by participants were radiation therapy related. Consequently this may not be an appropriate endpoint for lycopene intervention trials. We observed weight loss in the control group and a protective/delayed gastrointestinal response with lycopene supplementation, confirming results from an animal study (Andic, Garipagaoglu, Yurdakonar, Tuncel, & Kucuk, 2009). Overall, the results of this study may help expand the role of lycopene research from preventive to therapeutic.

Strengths and Limitations

Strengths

- To our knowledge, this is the first study evaluating serum lycopene levels
 and the impact of food-based lycopene supplementation during
 radiotherapy in prostate cancer patients.
- Lycopene intake was assessed using a validated food frequency questionnaire, the National Cancer Institute's (NCI) diet history

- questionnaire (DHQ) which has been validated with older adults.

 Additionally, since the DHQ calculates average intake over a 12 month period, day-to-day or seasonal variations in intake are not reflected.
- 3. The DHQ was also selected for estimating lycopene rich foods over other available methods (food diary) mainly because it would require less work for the patients as compared to maintaining a food diary during their treatment regimen.
- Tolerance of food-based lycopene supplementation during radiotherapy was assessed using the NCI Cancer Therapy Evaluation Program:
 Common Toxicity Criteria (CTC) v 2.0.
- Impact of lycopene supplementation on both performance status and select side effects during radiotherapy was assessed.
- 6. Impact of total lycopene on key inflammatory markers was evaluated.
- 7. This study expands the scope of lycopene research from preventive to an effective adjunct during radiation therapy in patients with localized prostate cancer.

Limitations

1. In order to minimize disruption of daily activities among the study participants, blood draws were scheduled around the time of their prescheduled radiation treatment appointment. Consequently, patients were not in a fasted state for the blood draws. Several researchers have demonstrated that the serum concentration of carotenoids do not change

- significantly for up to four hours after a meal (Brown, Rose, Craft, Seidel, & Smith, 1989; Mejia & Arroyave, 1983; Mejia, Pineda, Noriega, Benitez, & Falla, 1984). However, the impact of non-fasted state on the other test parameters remains unknown.
- 2. We were unable to use food records to monitor intake of lycopene rich foods during study duration.
- While the DHQ require less work for the patients as compared to maintaining a food diary, it relies heavily on patient memory in its reporting.
- 4. Efficacy of lycopene supplementation was only evaluated in men with localized prostate cancer during radiotherapy. It also needs to be evaluated in prostate cancer patients receiving other treatments such as hormone therapy.
- We only measured total serum lycopene levels in our participants. Role of specific lycopene isomers (cis vs trans) should also be evaluated in future trials.
- 6. Small sample size of this study limits generalization of these results.

Future Work

Based on the results of this research study, a phase II trial utilizing 8-12 oz of tomato juice or equivalent quantity of tomato product should be conducted.

Participants in the 12 oz group demonstrated the least side effects of treatment and a progressive decline in CRP and PGE2 expression between the three time

points. Despite a small increase in IL-6 expression at midpoint, the endpoint results were lower than the baseline results in the 12 oz group. While participants in the 4 oz group demonstrated a progressive decline in IL-6 between the three time points, some participants did demonstrate other side effects of treatment. Lower fatigue was observed among all intervention group participants but not in the control group. This was an unexpected finding. While we did not measure fatigue scores, we speculate that it impacted the performance status scores observed. Since we excluded patients who concurrently received hormone therapy or were treated with brachytherapy, or received radiation to the whole pelvic bed, the impact of lycopene supplementation in these groups also needs to be investigated. Additionally, no information exists on the impact of this supplementation in reducing long term complications of radiation therapy and in survivors. A retrospective review of long-term side effects in our study participants should also be conducted to identify the impact of lycopene on these side effects.

Overall, patients who volunteered to participate in our study appeared to have a desire for more "natural" alternatives to manage side effects of radiation therapy. This research also provides valuable data on the impact of tomato juice as a source of the carotenoid lycopene on select markers of inflammation. Links exploring the relationship between cytokine expression, radiation side-effects and lycopene need to be explored further in larger clinical trials. We hope that the

results of this study will allow us to secure funding in the future to conduct trials with a larger sample of patients to validate these results.

References

- Andic, F., Garipagaoglu, M., Yurdakonar, E., Tuncel, N., & Kucuk, O. (2009).

 Lycopene in the Prevention of Gastrointestinal Toxicity of Radiotherapy.

 Nutrition and Cancer, 61(6), 784 788.
- Bowen, P., Chen, L., Stacewicz-Sapuntzakis, M., Duncan, C., Sharifi, R., Ghosh,
 L., et al. (2002). Tomato Sauce Supplementation and Prostate Cancer:
 Lycopene Accumulation and Modulation of Biomarkers of Carcinogenesis.
 Exp Biol Med, 227(10), 886-893.
- Brown, E. D., Rose, A., Craft, N., Seidel, K. E., & Smith, J. C., Jr. (1989).

 Concentrations of carotenoids, retinol, and tocopherol in plasma, in response to ingestion of a meal. *Clin Chem, 35*(2), 310-312.
- Chen, L., Stacewicz-Sapuntzakis, M., Duncan, C., Sharifi, R., Ghosh, L., van Breemen, R., et al. (2001). Oxidative DNA damage in prostate cancer patients consuming tomato sauce based entrees as a whole-food intervention. *Journal of National Cancer Institute*, 93, 1872-1879.
- Clark, P. E., Hall, M. C., Borden, J. L. S., Miller, A. A., Hu, J. J., Lee, W. R., et al. (2006). Phase I-II prospective dose-escalating trial of lycopene in patients with biochemical relapse of prostate cancer after definitive local therapy.

 Urology, 67(6), 1257-1261.

- Davis, C., Clevidence, B., Swanson, C. A., Ziegler, R. G., Dwyer, J. T., & Milner, J. A. (2005). A Research Agenda for Lycopene/Tomato Supplementation and Cancer Prevention. 135(8), 2074S.
- Edinger, M. S., & Koff, W. J. (2006). Effect of the consumption of tomato paste on plasma prostate-specific antigen levels in patients with benign prostate hyperplasia. *Brazilian Journal of Medical and Biological Research*, 39, 1115-1119.
- Forssberg, A., Lingen, C. H. R., Ernster, L., & Lindberg, O. (1959). Modification of the x-irradiation syndrome by lycopene. *Experimental Cell Research*, 16, 7-14.
- Gann, P. H., Ma, J., Giovannucci, E. L., Willett, W. C., Sacks, F. M., Hennekens,
 C. H., et al. (1999). Lower prostate cancer risk in men with elevated
 plasma lycopene levels: Result of a prospective analysis. *Cancer Research*, *59*, 1225-1230.
- Giovannucci, E. (1999). Tomatoes, tomato-based products, lycopene and cancer: Review of the Epidemiologic literature. *Journal of National Cancer Institute*, *91*(4), 317 331.
- Giovannucci, E. (2005). Tomato Products, Lycopene, and Prostate Cancer: A

 Review of the Epidemiological Literature. *The Journal of Nutrition, 135*(8),

 S2030-S2031.

- Jatoi, A., Burch, P., Hillman, D., Vanyo, J. M., Dakhil, S., Nikcevich, D., et al.
 (2007). A Tomato-Based, Lycopene-Containing Intervention for Androgen-Independent Prostate Cancer: Results of a Phase II Study from The North Central Cancer Treatment Group. *Urology*, 69(2), 289-294.
- Kim, H., Bowen, P., Chen, L., Duncan, C., Ghosh, L., Sharifi, R., et al. (2003).

 Effects of tomato sauce consumption on apoptotic cell death in prostate benign hyperplasia and carcinoma. *Nutrition & Cancer*, *47*(1), 40-47.
- Kucuk, O., Sarkar, F., Sakr, W., Djuric, Z., Pollak, M., Khachik, F., et al. (2001).
 Phase II randomized clinical trial of lycopene supplementation before radical prostatectomy. *Cancer Epidemiol Biomarkers and Prevention*, 10, 861-868.
- Mejia, L. A., & Arroyave, G. (1983). Determination of vitamin A in blood. Some practical considerations on the time of collection of the specimens and the stability of the vitamin. *Am J Clin Nutr, 37*(1), 147-151.
- Mejia, L. A., Pineda, O., Noriega, J. F., Benitez, J., & Falla, G. (1984).
 Significance of postprandial blood concentrations of retinol, retinol- binding protein, and carotenoids when assessing the vitamin A status of children.
 Am J Clin Nutr, 39(1), 62-65.

- Mohanty, N. K., Saxena, S., Singh, U. P., Goyal, N. K., & Arora, R. P. (2005).

 Lycopene as a chemopreventive agent in the treatment of high-grade prostate intraepithelial neoplasia. *Urol Oncol*, *23*(6), 383 385.
- Rao, A. V., Fleshner, N., & Agarwal, S. (1999). Serum and tissue lycopene and biomarkers of oxidation in prostate cancer patients: a case-control study.

 Nutrition & Cancer, 33(2), 159 -164.
- Schwarz, S., Obermuller-Jevic, U. C., Hellmis, E., Koch, W., Jacobi, G., & Biesalski, H.-K. (2008). Lycopene Inhibits Disease Progression in Patients with Benign Prostate Hyperplasia. *J. Nutr., 138*(1), 49-53.
- Vogt, T. M., Mayne, S. T., Graubard, B. I., Swanson, C. A., Sowell, A. L., Schoenberg, J. B., et al. (2002). Serum Lycopene, Other Serum Carotenoids, and Risk of Prostate Cancer in US Blacks and Whites. *Am. J. Epidemiol.*, 155(11), 1023-1032.

APPENDIX A. RECTUITMENT FLYER

Effect of Tomato Juice during Radiation Therapy in men with Prostate Cancer



Are you a man, who

- · has recently been diagnosed with prostate cancer,
- · has not undergone hormone therapy, surgery or other treatment for prostate cancer,
- is about to start radiation treatment,
 AND
- is interested in participating in a clinical trial?

If you responded YES to each of the above statements, then call us!

The purpose of this research is to investigate the effect of lycopene (a nutrient found in tomato juice) on radiation therapy related side effects in men with prostate cancer undergoing radiation treatment.

You will not be eligible to participate in this study if you

To be eligible to participate in this study you must

- Have a new diagnosis of localized prostate cancer
- · Have not yet initiated treatment;
- · Have normal immune, liver and kidney function;
- Be willing to sign informed consents
- Be willing to provide small blood samples (2 tablespoons three different times)

For additional information, please contact Dr. Bart Frizzell at 878 - 6036

Mri Datta at 307 - 9020

- Have metastasized prostate cancer or have previously received treatment for prostate cancer;
- · Are allergic to tomato products
- Are unwilling to provide blood samples
- Have certain medical conditions High blood potassium levels, uncontrolled gastro-esophageal reflux disease (GERD), malabsorption disorders, liver or kidney problems
- Routinely use the following supplements –MVI, fiber, saw palmetto, lycopene, Omega-3 Fatty acids/Fish oil, Vitamin C, E, A and B-carotene, flaxseeds or flaxseed oil supplements;
- · Are unable/unwilling to sign the informed consent

APPENDIX B. HEALTH INFORMATION PORTABILITY AND ACCOUNTABILITY ACT (HIPAA) FORM – ENGLISH

HIGH POINT REGIONAL HEALTH SYSTEM INSTITUTIONAL REVIEW BOARD

<u>HIPAA – AUTHORIZATION FOR RESEARCH</u>

For the Use and/or Disclosure of Protected Health Information

You may qualify for participation in a research study. This "HIPAA Authorization for Research" form gives you information about how health information collected about you for a research study would be used by researchers and disclosed to others involved in the research study. Your signature on this form permits High Point Regional to give your information to the research team.

Study Title:	Food-based lycopene supplementation in prostate cancer patients	
	<u>undergoing radiotherapy</u>	
Principal Investigator:Martha Taylor, PhD		
1	.	

I authorize the use and/or disclosure of my protected health information as described below:

- 1. The following entity/person(s) is *authorized to use or disclose* my information:
 - High Point Regional Health System 601 North Elm Street High Point, NC 27262 (336) 878-6000
- 2. The following entity/person(s) is *authorized to receive* my information:
 - The Researchers (members, agents or successors of the research team, such as the Principal Investigator, Co-Investigators and members of their research staff)
 - The sponsor of the research study, and its agents and contractors
 - Representatives of government organizations, review boards, and other persons who are required to watch over the safety and effectiveness of medical products and therapies and the conduct of research
- 3. Description of the information that may be used or disclosed:
 - Health information in my medical records that is relevant to the study
 (Taking part in this research study may involve collecting and disclosing health information that you consider confidential or private that directly identifies you.

Information in your medical record may include results of tests, procedures, interventions, interactions, questionnaires or surveys. All information may be used and possibly disclosed and re-disclosed to monitor your health status, to measure the effects of drugs/devices/procedures /interventions as stated in this study, to determine research results and outcomes, and possibly develop new drugs/devices/tests/procedures and commercial products.)

- 4. My information will be used or disclosed for the following *purposes*:
 - By the Researchers, among themselves and with other participating researchers to conduct the Research;
 - By the research Sponsor, as will be described in the Informed Consent Form;
 - By the representatives of government organizations, review boards, and other persons to watch over the safety and effectiveness of medical products and therapies and the conduct of research.

(Your health information may be used by, disclosed to and re-disclosed for: research, quality assurance, or regulatory purposes, by: members, agents or successors of the research team, such as the Principal Investigator, Co-Investigators and members of their research staff ("Researchers"); other researchers and their staff involved with this study at other medical centers, institutions, hospitals, central data centers, the U.S. Food and Drug Administration, the Department of Health and Human Services, the Federal Office of Human Research Protection, High Point Regional Health System and/or the sponsor of this research study.)

- 5. Study data that does not directly identify you may be published in medical journals or shared with others as part of scientific discussions.
- 6. You have the right to see and copy your personal health information related to the research study for as long as this information is held by the Investigator or a research institution. However, to ensure the scientific integrity of the study, you will not be able to review some of the study information until after the study has been complete.
- 7. This authorization will expire when the last component of this research study is completed. (Some research studies include more than one component, each of which may span a different time period; therefore, this authorization extends to the time when the last component is completed.)
- 8. I understand that I may revoke this authorization at any time. My revocation must be in writing and contain my signature. I am aware that my revocation is not effective to the extent that the entity/person(s) I have authorized to use or disclose my information have acted in reliance upon this authorization. To revoke my authorization, I understand that I need to contact the IRB Office at the address and phone number listed above in item #1 above and complete the appropriate paperwork.

I have been advised, and I understand, that this authorization is voluntary. I do not have to sign this authorization and my refusal to sign will not affect my abilities to obtain treatment from High Point Regional Health System, nor will it affect my eligibility for benefits. However, I understand that I will be required to sign this authorization form prior to receiving research-related treatment.
 I understand that if my information is disclosed to someone who is not required to comply with the federal privacy protection regulations, then such information may be re-disclosed and would no longer be protected.
 I certify that I have received a copy of this authorization.

Signature of Patient	Date
Print Name	
below:	by someone other than the patient, please indicate
Signature	Date
Print Name	Relationship to Patient

03-03; revised: 12-18-03; revised 01-09-06

APPENDIX C. HEALTH INFORMATION PORTABILITY AND ACCOUNTABILITY ACT (HIPAA) FORM – SPANISH

High Point Regional Health System Comité de Ética de Investigación Clínica

Autorización para Investigación-HIPAA

Para el Uso y/o Divulgación de Información Médica Protegida

Usted puede calificar para participar en un estudio de investigación. Este formulario "Autorización para Investigación de HIPPA" le da información acerca de cómo su información medica colectada para el estudio de investigación será usada por los investigadores y será divulgada a otros envueltos en el estudio de investigación. Su firma en este formulario permite a High Point Regional dar su información al equipo de investigación.

Título del Estudio:
Investigador Principal:Martha Taylor, PhD
Yo autorizo el uso y/o divulgación de mi información médica protegida como se describe abajo
1. La siguiente entidad/nersona/s) son autorizadas nara usar o divulgar mi información:

- 1. La siguiente entidad/persona(s) son autorizadas para usar o divulgar mi información:
 - High Point Regional Health System 601 North Elm Street High Point, NC 27262 (336) 878-6000
- 2. La siguiente entidad/persona(s) esta autorizada para recibir mi información:
 - Los investigadores (miembros, agentes o sucesores del equipo de investigación, tales como el Investigador Principal, Co-Investigadores y miembros del personal de investigación)
 - El patrocinador del estudio de investigación y sus agentes y contratistas
 - Representantes de organizaciones gubernamentales, comités de ética, y otras personas que les es requerido vigilar la seguridad y la efectividad de los productos médicos y terapias y el conducto de la investigación.
- 3. Descripción de la información que puede ser usada y divulgada:
 - Información médica en mis expedientes médicos que sea pertinente al estudio

(El tomar parte en este estudio puede envolver el colectar y divulgar información médica que usted considera confidencial o privada que directamente lo identifica. Información médica en su expediente médico puede incluir los resultados de exámenes, procedimientos, intervenciones, interacciones, cuestionarios o encuestas. Toda la

información puede ser usada y posiblemente divulgada y revelada para monitorear su estado de salud para medir los efectos de los medicamentos/aparatos/procedimientos/intervenciones como se ha expuesto en este estudio, para determinar los resultados y consecuencias y posiblemente desarrollar nuevos medicamentos/aparatos/exámenes/procedimientos y productos comerciales.)

- 4. Mi información será usada o divulgada con los siguientes propósitos:
 - Por los Investigadores, entre ellos y con otros investigadores participantes para conducir la Investigación.
 - Por el investigador Patrocinador, como será descrito en el Consentimiento Informado;
 - Por los representantes de organizaciones gubernamentales, comités de ética y otras personas que vigilan la seguridad y la efectividad de los productos médicos y terapias y el conducto de la investigación.

(Su información puedes ser usada, divulgada y revelada para: investigación, control de calidad o propósitos reguladores, por: miembros, agentes o sucesores de el equipo de investigación, tal como el Investigador Principal, Co-Investigadores y miembros del personal de investigación ("Investigadores"); otros investigadores y su personal envueltos con este estudio en sus centros médicos, instituciones, hospitales, centro de datos central, Administración de Alimentos y Fármacos de EE. UU., el Departamento de Salud y Servicios Humanos, La Oficina Federal para la Protección de los Seres Humanos en la Investigación, High Point Regional Health System y/o el patrocinador de este estudio de investigación.

- 5. Información del estudio que no lo identifique directamente puede se publicada en diarios médicos o compartida con otros en discusiones científicas.
- 6. Usted tiene el derecho de ver y copiar su información personal médica relacionada con la investigación mientras que el Investigador o la institución de investigación tenga esta información. Sin embargo, para asegurar la integridad científica de la investigación usted no podrá repasar alguna de la información de la investigación hasta después de que se haya completado la investigación.
- 7. Esta autorización se vence cuando el ultimó componente de esta investigación se complete. (Algunos estudios de investigación incluyen más de un componente, lo cual cada uno tiene un lapso de un período de tiempo diferente; por consiguiente, esta autorización se extiende al tiempo cuando el ultimó componente es completado.)
- 8. Yo entiendo que puedo revocar esta autorización en cualquier momento. Mi revocación debe ser por escrito y debe contener mi firma. Estoy al tanto de que mi revocación no es efectiva a la entidad/persona(s) que he autorizado y han actuado según esta autorización para usar y divulgar mi información. Para revocar mi autorización, yo

- entiendo que necesito contactar la oficina del IRB a la dirección y número de teléfono indicado arriba en el artículo #1 arriba y completar el papeleo apropiado.
- 9. Se me ha aconsejado y entiendo que esta autorización es voluntaria. No tengo que firmar esta autorización y mi rechazo a firmar no afectará mis habilidades a obtener tratamiento de High Point Regional Health System, ni tampoco afectará mi elegibilidad para beneficios. Sin embargo, yo entiendo que se requiere que firme esta autorización antes de recibir tratamiento relacionado a la investigación.
- 10. Yo entiendo que si mi información es divulgada a alguien que no se le requiera cumplir con las regulaciones federales de privacidad, después la información puede ser revelada y ya no seria protegida.

Yo certifico que he recibido una copia de e	esta autorización.
Firma del paciente	Fecha
Escriba su nombre en letra de molde	
En el caso de que este formulario sea com favor indíquelo abajo:	pletado por alguien que no es el paciente, po
Firma	
Escriba su nombre en letra de molde	Relación al paciente

APPENDIX D. CONSENT FORM – ENGLISH

HIGH POINT REGIONAL HEALTH SYSTEM INSTITUTIONAL REVIEW BOARD INFORMED CONSENT

What is a research study?

You are invited to be in a research study. Research studies are designed to gain scientific knowledge that may help other people in the future. You may or may not receive any benefit from being part of the study. There may also be risks associated with being part of research studies. You are being asked to take part in this study because you have been diagnosed with prostate cancer and are about to begin radiation therapy for it. Your participation in this study is voluntary. Please take your time to make your decision and ask your study doctor or the study staff to explain any words or information that you do not understand. You may also discuss the study with your friends and family.

Who is sponsoring the study?

The researchers do not hold a direct financial interest in the sponsor or the product being studied.

Why is the study being done?

This study is being done to test if lycopene, a natural substance found in tomatoes, tomato products and some other foods helps reduce radiation therapy related side effects in men with prostate cancer, receiving external beam radiation therapy.

In this study, the effect of three different amounts of tomato juice is being evaluated. In this study, you will receive 4 ounces, 8 ounces or 12 ounces of tomato juice.

How many people will take part in the study?

Forty men with prostate cancer about to initiate radiation therapy for treatment will be invited to participate in this study.

What is involved in the study?

If you agree to participate, we will need to check your blood work to make sure your kidneys and liver are functioning adequately and check your bloods lycopene level and other immune function related labs. This will require a blood sample (about 2 tablespoons of blood) to be drawn at the beginning of the study. These tests will be repeated at the end of three weeks of treatment and on the last day you receive your radiation treatment.

We will ask you to drink your assigned amount of tomato juice daily while you are receiving your radiation treatment, starting at least 2 days before you begin your treatment. On Friday of each week (during treatment), you will be provided with two additional doses of the beverage to take home with you. One dose of the beverage should be consumed on Saturday and the other on Sunday.

You will be asked to complete the National Cancer Institute's Diet History Questionnaire. This will help us estimate your usual intake of lycopene rich foods. There is no right or wrong answer, and you do not need to change your eating habits just because you are participating in this study. Please answer the questions honestly about how frequently you have been consuming specific foods.

At the end of each week, you receive treatment we will also ask you to report your level of fatigue and ability to carry out your daily activities and any symptoms you may be experiencing.

How long will I be in the study?

You will be in the study for the entire time you are receiving radiation therapy.

The researcher may decide to take you off the study if: 1) continued participation is not in your best medical interest, 2) health conditions occur which would make your participation possibly dangerous, or 3) new information becomes available.

While we encourage you to continue till the end of the study, you may of course, withdraw from the study at any time. This will not comprise or affect the quality of your care that you may receive from this institution. If you decide to stop participating at any time, we encourage you to talk with the investigator or study staff first to learn about any potential health or safety consequences.

What are the risks of the study?

Being in this study may or may not involve any risk to you. You should discuss the risk of being in this study with the study staff.

<u>Blood draw</u>: You may experience discomfort, bruising, and/or bleeding where the needle is inserted. There may be bruising at the puncture site when blood is drawn and occasionally some redness and swelling. Occasionally some people become dizzy, lightheaded or feel faint. Infection may occur on rare occasions.

Risks and side effects related to the study treatments:

Study supplement: Lycopene is a naturally substance found in tomatoes, tomato products and some other foods. Processed tomato juice, which is the chosen food to provide lycopene in this study, may potentially cause some of the following side effects:

- heartburn
- abdominal distention
- flatulence (gas)(Jatoi et al., 2007)
- nausea
- vomiting
- diarrhea

There may be other side effects that we cannot predict or are unexpected. If you have any unusual symptoms, report them immediately to your doctor and the study staff. You should also tell the research staff about all medications, vitamins and supplements you take and any medical conditions you have. This may help avoid side effects, interactions and other risks.

Will I benefit from taking part in the study?

You may or may not receive any benefit from taking part in this research study. We hope the information learned from this study will benefit other patients in the future. The benefit of participating in this study may be a decrease in radiation therapy related side effects.

Will my medical information be kept private?

We will do our best to make sure that the personal information in your medical record will be kept private. However, we cannot guarantee total privacy. Your personal information may be given out if required by law. If information from this study is published or presented at scientific meetings, your name and other personal information will not be used.

A record of your progress will be kept in a confidential file at the hospital or doctor's office where you receive treatment. Organizations that may inspect and/or copy your research and medical records (blood samples etc.) for quality assurance, research, and data analysis include groups such as:

Comprehensive cancer center at Wake Forest University (CCCWFU)

National Cancer Institute (NCI) and its representative

Food and Drug Administration (FDA)

Office of Human Research protections (OHRP)

Department of Health and Human Services (DHHS)

Institutional Review Board (IRB) at High Point Regional Health System

Possible other federal or state government agencies

If your record is used out given out for governmental purposes, it will be done under conditions that will protect your privacy to the fullest extent possible consistent with the laws relating to public disclosure of information and law-enforcement responsibilities of the agency. These agencies may review the research to see that it is being done safely and correctly.

You authorize the use of clinical information contained in your records, but any publication that includes such information or data shall not reveal your name, show

your picture or contain any other personally identifying information, except as otherwise required by law.

What are the costs of taking part in this study?

There are no costs to you for taking part in this study. All the study costs, including any study medications and procedures directly related to the study, will be paid for by the study. Costs or your regular medical care/treatment, which are not related to this study, will be your own responsibility.

You will not be paid to participate in this study.

What happens if I am injured because I took part in this study?

It is important that you tell your study doctor, Bart Frizzell, M.D. if you feel that you have been injured because if taking part in this study. You can tell the doctor in person or call him at 336-878-6036.

You will get medical treatment if you are injured as a result of taking part in this study. You and/or your health plan will be charged for this treatment. This study will not pay for medical treatment. Although no funds or monies have been set aside to compensate you in the event of injury or illness related to the study treatment or procedures, you do not waive any of your legal rights for compensation by signing this form.

You or your insurance company will be charged for continuing medical care and/or hospitalization.

What are my rights if I take part in this study?

Taking part in this study is voluntary. You may choose to take part, not to take part or may leave the study at any time. No matter what decision you make, there will be no penalty for you and you will not lose any of your regular benefits. Leaving the study will not affect your medical care or result in any penalty or loss of benefits to which you are entitled.

Even after you agree to take part in this study, you may withdraw at anytime. Before you withdraw, you should talk to one of the researchers. This will allow them to inform you of any medical problems that could result from stopping your treatment. Your decision to stop taking part in this study will not affect your medical treatment or your relationship with those treating you or with this institution. If you withdraw from the study, you will still be offered all available care that meets your medical condition. You are free to seek care from a doctor of your choice at any time. The investigators also have the right to stop your participation in the study at any time. This could happen if you lose too much weight or have side effects from either the radiation therapy or the beverage that prevents you from continuing in the study.

We will tell you about new information that may affect your health, welfare or willingness to stay in the study. You may be asked to sign another consent form in response to new information.

Who can answer my questions about the study?

For questions about the study or a research-related injury, contact your doctor, Bart Frizzell, M.D. at 336-878-6036.

You can contact the study coordinator, Mri Datta at 336-307-9020 if you have further questions.

For questions about your rights as a research participant, contact the High Point Regional health System Institutional Review Board (which is a group of people at the hospital where you receive treatment, who review the research to protect your rights) at 336-878-6207.

Participant Contract

I have been offered the opportunity to ask questions about this study and all questions have been answered to my satisfaction. The contents of this form have been explained to

me and I understand them. I agree to allow the research personnel specified above the access to my medical records.

It may be necessary for my doctor to contact me at a future date regarding new information about the treatment I received; therefore, I agree to notify my doctor of any change of address and/or telephone number.

My signature below means that I have voluntarily agreed to participate in this research study. I will be given a copy of all 6 pages of this consent. I have read it or it has been read to me, I may also request a copy of the study (complete study plan)

(Participant Name)	(Participant Signature)	(Date)
I certify that I have explained to the		1 1
potential benefits, and possible risk and have answered any questions t	1 1	n in the research study
Name of person obtaining consent)	(Signature)	(Date)

I,	, withdraw my consent to participate in this
study and refuse to have clinical da	ata collected from my medical record (s).
Participant name(please print	study ID#
Participant signature	Date
Witness signature	Date

APPENDIX E. CONSENT FORM - SPANISH

HIGH POINT REGIONAL HEALTH SYSTEM COMITÉ DE ÉTICA DE INVESTIGACIÓN CLÍNICA

CONSENTIMIENTO INFORMADO

¿Qué es un estudio de investigación?

Usted es invitado a participar en un estudio de investigación. Los estudios de investigación son diseñados para obtener conocimiento científico que pueda ayudar a otras personas en el futuro. Quizá reciba o no beneficios por ser parte del estudio. También pueden existir riesgos asociados por ser parte de los estudios de investigación. Se le ha pedido ser parte de este estudio porque usted ha sido diagnosticado con cáncer de la próstata y está a punto de recibir radio terapia para ella. Su participación en este estudio es voluntaria. Por favor de tomarse el tiempo para tomar su decisión y pregúntele a su doctor o a el personal del estudio que le expliquen cualquier palabra o información que no entienda. También puede hablar con su familia y amistades acerca del estudio.

¿Quién esta patrocinando el estudio?

Los investigadores no tienen ningún interés financiero al patrocinar el producto que se está estudiando.

¿Por qué se está haciendo este estudio?

Este estudio se esta haciendo para probar si el licopeno, una sustancia natural que se halla en tomates, productos de tomates y otros alimentos, puede ayudar a reducir los efectos secundarios relacionados con la radioterapia en hombres con cáncer de la próstata que reciben radioterapia.

En este estudio, se está evaluando el efecto de tres diferentes cantidades de jugo de tomate. En este estudio usted recibirá 4 onzas, 8 onzas y 12 onzas de jugo de tomate.

¿Cuántas personas participarán en el estudio?

Cuarenta hombres con cáncer de la próstata que están por iniciar la radioterapia serán invitados a participar en este estudio.

¿Qué envuelve el estudio?

Si usted está de acuerdo en participar, necesitamos revisar sus análisis de sangre para asegurarnos de que sus riñones e hígado estén funcionando adecuadamente y analizaremos en su sangre su nivel de licopeno y otras funciones inmunes. Esto requiere que tomemos una muestra de su sangre (como 2 cucharadas de sangre) al principio del estudio. Estos exámenes se repetirán al final de tres semanas de tratamiento y al último día que reciba la radioterapia. Le pediremos que tome a diario su bebida de jugo de tomate con la cantidad asignada mientras esté recibiendo su radioterapia, comenzando por lo menos 2 días antes de empezar su tratamiento. El viernes de cada semana (durante el tratamiento) se le proveerá con dos dosis adicionales de la bebida para que la lleve a casa. Debe consumir una dosis de la bebida el sábado y la otra el domingo.

Se le pedirá que llene el Cuestionario de Historial Alimenticio del Instituto Nacional de Cáncer. Esto nos ayudará a estimar su consumo de alimentos ricos en licopeno. No hay ninguna contestación correcta o incorrecta y no tiene que cambiar sus hábitos de alimentación solo porque va a participar en este estudio. Por favor de contestar todas las preguntas lo más honesto posible, de que tan frecuente ha estado consumiendo alimentos específicos.

1	Participante:

Al final de cada semana que reciba tratamiento también le pediremos que reporte su nivel de fatiga y habilidad de llevar a cabo sus actividades diarias y cualquier síntoma que experimente.

¿Cuánto tiempo participaré en el estudio?

Usted participará en el estudio todo el tiempo que reciba la radioterapia.

El investigador puede decidir removerlo del estudio si: 1) su participación no es lo más conveniente para su salud 2) ocurren condiciones de salud que harían su participación posiblemente peligrosa o 3) sale disponible información nueva.

Aunque le animamos a que continué hasta el final, usted puede retirarse del estudio en cualquier momento. Esto no comprometerá o afectará la calidad de cuidado que recibe de esta institución. Si decide no participar en cualquier momento le animamos a que hable con el investigador o personal del estudio primero para ver si hay alguna consecuencia potencial en su salud o seguridad.

¿Cuáles son los riesgos del estudio?

El participar en este estudio puede o no envolver riesgos a usted. Hable con el personal del estudio acerca de los riesgos por participar en el.

Extracción de sangre: Puede experimentar incomodidad, moretón y/o sangrado donde introducen la aguja. Pueden obtener un moretón en el sitio de la punción cuando extraigan la sangre y ocasionalmente enrojecimiento e hinchazón. Ocasionalmente algunas personas se marean o sienten que van a desmayar. En raras ocasiones puede producirse una infección.

Riesgos y efectos secundarios relacionados a los tratamientos del estudio:

Suplemento del estudio: Licopeno es una sustancia natural que se encuentra en tomates, productos de tomates y otros alimentos. El jugo de tomate procesado, el alimento escogido para proveer licopeno en este estudio, puede potencialmente causar los siguientes efectos secundarios:

- acidez/agruras
- distensión abdominal
- flatulencia (gas)
- náusea
- vomito
- diarrea

Puede haber otros efectos secundarios que no podemos predecir o que son inesperados. Si tiene algún síntoma inusual repórtelos de inmediato a su doctor o al el personal del estudio. También déjele saber a el personal de investigación sobre todo medicamento, vitaminas o suplementos que toma y cualquier condición de salud que tenga. Esto puede ayudar a evitar efectos secundarios, interacciones y otros riesgos.

¿Me beneficiaré por participar en el estudio?

Puede que si o que no reciba ningún beneficio al participar en el estudio de investigación. Esperamos que la información que obtengamos del estudio le podrá beneficiar a otros pacientes en el futuro. El beneficio de participar en este estudio puede ser una disminución en los efectos secundarios relacionados con la radioterapia.

2	Participante:

¿Se mantendrá privada mi información médica?

Haremos lo mejor para asegurarnos de que su información personal en su expediente médico se mantenga privada. Sin embargo, no le podemos garantizar privacidad total. Su información personal puede ser divulgada si es requerido por la ley. Si la información de este estudio es publicada o presentada en juntas científicas, no se usará su nombre o alguna otra información personal.

El expediente de su progreso se mantendrá en un archivo confidencial en el hospital o la oficina del doctor donde usted recibe el tratamiento. Las organizaciones que quizá inspeccionen y/o copien su estudio o expediente médico (muestras de sangre, etc.) para garantía de calidad, investigación y análisis de datos incluyen los grupos como:

Centro exhaustivo de cáncer en La Universidad Wake Forest (CCCWFU)

Instituto Nacional de Cáncer (NCI) y sus representantes

Administración de Alimentos y Fármacos (FDA)

Oficina para la Protección de los Seres Humanos en la Investigación (OHRP)

Departamento de Salud y Servicios Humanos (DHHS)

Comité de Ética de Investigación Clínica en High Point Regional Health System Posiblemente otras agencias gubernamentales federales o estatales

Si su expediente se libera con propósitos gubernamentales, será bajo condiciones que proteja su privacidad hasta donde las leyes relacionadas a la divulgación pública de información y las responsabilidades de la agencia a ejecutar la ley lo permitan. Estas agencias pueden examinar la investigación para ver si se esta haciendo de una manera segura y correcta.

Usted autoriza el uso de información clínica que contiene su expediente, pero cualquier publicación que incluya tal información o datos no deben revelar su nombre, mostrar su fotografía o contener cualquier otra información que lo identifique, a menos que se requiera por ley.

¿Cuáles con los costos por participar en este estudio?

No hay ningún costo a usted por participar en este estudio. Todos los costos del estudio, incluyendo cualquier medicamento o procedimientos directamente relacionados con el estudio, serán pagados por el estudio. Los costos de su tratamiento/cuidado médico regular, que no están relacionados con el este estudio, serán su responsabilidad. No se le pagará por participar en este estudio.

¿Qué pasa si me lastimo por participar en este estudio?

Es importante que le diga a su doctor, Bart Frizzell, , M.D. si siente que ha sido lastimado al participar en este estudio. Puede decírselo al doctor en persona o llamarlo al 336-878-6036. Usted recibirá tratamiento médico si es lastimado como resultado de participar en este estudio. Se le cobrará a usted y/o plan de salud por este tratamiento. Este estudio no pagará por tratamiento médico. Aunque no se ha reservado fondos o dinero para compensarlo en evento de ser lastimado o por enfermedad relacionada con el tratamiento o procedimientos del estudio, al firmar este formulario usted no esta renunciando sus derechos legales por compensación.

Se le cobrará a usted o su compañía de seguro por cuidado médico continuo y/o por ser hospitalizado.

3	Participante:

¿Cuáles son mis derechos si participo en este estudio?

El participar en este estudio es voluntario. Puede optar por participar, no participar o por dejar el estudio en cualquier momento. No importa cual sea su decisión, no habrá ninguna penalización y no perderá ninguno de sus beneficios regulares. El que deje el estudio no afectará su cuidado médico o resultará en ninguna penalización o pérdida de beneficios a los cuales tenga derecho.

Aun si concuerda en participar en este estudio, usted puede retirarse en cualquier momento. Antes de retirarse debe hablar con los investigadores. Esto les permitirá informarle de cualquier problema que pueda surgir como resultado por dejar el tratamiento. Su decisión de dejar de participar en el estudio no afectará su tratamiento médico o su relación con quienes lo atienden o con esta institución. Si se retira del estudio, aún se le ofrecerá todo cuidado disponible que reúna los requisitos a su condición médica. Tiene el derecho de buscar cuidado de cualquier doctor en cualquier momento. Los investigadores también tienen el derecho de detener su participación en este estudio en cualquier momento. Esto ocurre si baja demasiado de peso o tiene efectos secundarios, ya sea de la radioterapia o la bebida, que prevenga a que continúe en el estudio.

Le informaremos acerca de nueva información que pueda afectar su salud, bienestar o disposición de permanecer en el estudio. Puede que se le pida firmar otro consentimiento en reacción a la nueva información.

¿Quién puede contestar mis preguntas acerca del estudio?

Para preguntas acerca del estudio o herida/lesión relacionado con el estudio, contacte a su doctor, Bart Frizzell, M.D. al 336878-6036.

Puede contactar a la coordinadora del estudio, Mri Datta al 336-307-9020 si tiene más preguntas.

Para preguntas sobre sus derechos como participante de la investigación, contacte a el Comité de Ética de Investigación Clínica de High Point Regional Health System (lo cual es un grupo de personas en el hospital donde recibe tratamiento, quienes revisan el proyecto de investigación para proteger sus derechos: al 336-878-6207.

Contrato del Participante

Se me ha ofrecido la oportunidad de hacer preguntas acerca del estudio y todas las preguntas han sido contestadas a mi satisfacción. Se me ha explicado el contenido de este formulario y lo entiendo. Estoy de acuerdo en permitir el personal de la investigación especificado arriba a tener acceso a mi expediente médico.

Quizá sea necesario que mi doctor me contacte en el futuro sobre nueva información acerca del tratamiento que recibí; por lo tanto yo concuerdo en notificar a mi doctor con cualquier cambio de dirección y/o número de teléfono.

Mi firma abajo significa que yo voluntariamente concuerdo en participar en esta investigación. Se me dará una copia de las 5 páginas de este consentimiento. Yo lo he leído o se me ha leído, también puedo pedir una copia del estudio (plan completo del estudio)

(Nombre del Participante)	(Firma del Participante)	(Fecha)	
4		Participante:	

ombre de la persona obteniendo el consentimiento)	(Firma)	(Fecha)
		Participante:

Yo, consentimiento a participar en este estudio y me rehúso a que co expediente(s) médico.	, retiro mi olecten datos de mi
Nombre del participanteestudio	# de identificación de
(por favor escriba claramente su nombre en letra de molo	de)
Firma del participanteF	Fecha
Firma del testigo F	Fecha
6	Participante:

APPENDIX F. SCREENING FORM

Participant Information/Screening

Name:					Age:	
Ethnicity: 🗖	African Ame	rican 🛭 Cau	casiar	n ☐ Other_		
Height:	report	ed wt:	h	now long ago:_		
Weight chan	ge: 🛭 loss (#	#/time)		_ □ gain (#/tim	e)	
Marital Statu	s: 🖵 m	narried u wido	wed	☐ divorced	□ ne	ver married
Highest level of Education: □ < high school □ high school □ > high schoo						
Other signif	icant inform	nation:				
Group Assi	gnment:		P	articipant ID:		
No. Of	treatments	prescribed: _		_ Gy, over _		days
Labs	Date:	Results Date:		Medi □ HTN	ical I	History
PSA				□ DM		
Na				☐ CAD		
K				☐ Hyper chol		
Gluc						_
BUN				Meds & Supp		ents: Prescription
Cr.				B# 11 41		OTC
<u>Ca</u>				Medication	S	Supplements
albumin						
SGOT						
SGPT LDH						
Alk. Phos						
			_			

		Yes	No	
ECOG performance score 0 or 1				
Are you allergic to tomatoes or red food coloring?				
Are you allergic to any other foods? If yes, please sp	pecify foods allergic to			
Have you ever been diagnosed with				
Problems	with your immune function			
	liver disease			
kidney disease				
malabsorption disorders/ Irri	table bowel/ Celiac disease			
Uncontrolled gastro	o-esophageal reflux disease			
Are you currently taking any fiber supplements/pills: Metamuc	cil, Citrucel, Fibercon, Benefiber			
Do you exercise? If yes, how often?				
Do you smoke? If yes how many packs a day				
Do you drink alcohol? If yes, please specify how ma	any drinks			
· · · · · · · · · · · · · · · · · · ·	_drinks per month of			
Weekly drinks a week of Socially				
Since being diagnosed with cancer, have you made a	• • •			
Activity				
	Diet			
	Lifestyle			
Are you currently taking any of the following supple				
Multivitamin	supplement with lycopene			
	Saw Palmetto			
	Lycopene			
Vitamin A				
	Vitamin C			
	Vitamin E			
Beta-carotene				
	Omega-3 Fatty acids			
	Fish oil			
	Flax seeds			
	Flaxseed oil			
	EPA/DHA			
Other Su	applements (please specify)			
Are you willing to stop taking these supplements for the	duration of the study?			
Would you be willing to sign an informed consent for				
Notes:				

APPENDIX G. INSTRUCTIONS FOR STUDY PARTICIPANTS – ENGLISH

Instructions for tomato juice and prostate cancer study participants

- 1. Please **continue with your normal eating habits and meal patterns**. Do not make any changes in your diet just because you are participating in this study.
- 2. During your participation in the study, each Friday you will be provided with two beverage containers. Please consume the beverage in one container on Saturday and the other on Sunday.
- 3. Consume beverage within 30 minutes of eating a meal to aid in the digestion of the beverage.
- 4. **<u>Do not</u>** consume any of the following supplements for the duration of your treatment/study participation as these may change how your body handles the treatment you are receiving.

a. Multivitamin
b. Saw Palmetto
c. Lycopene
d. Vitamin A
d. Fish Oil
j. EPA/DHA
k. Flaxseeds

d. Beta-Carotene h. Omega-3 l. Flaxseed oil

- 5. Before taking any other nutritional supplement or vitamins, please check with your radiation doctor and the study coordinator. Some of these supplements may decrease the benefits of the radiation treatment you are receiving.
- 6. You will be asked to complete a **food frequency questionnaire** at the beginning of the study. This should take you about an hour to complete. Please answer all questions as accurately as you can and return <u>the questionnaire to the study</u> <u>coordinator, one week before your last treatment</u>. There is no right or wrong answer.
- 7. Please let your radiation doctor and study coordinator know if there are any changes in your bowel movements, bladder problems or you experience other problems such as nausea, vomiting, or heartburn.

APPENDIX H. INSTRUCTIONS FOR STUDY PARTICIPANTS – SPANISH

Instrucciones para los participantes en el estudio de cáncer de la próstata y jugo de tomate

- 1. Por favor **continúe con sus hábitos y patrones normales de alimentación.** No haga ningún cambio en su dieta solo porque esta participando en este estudio.
- 2. Durante su participación en el estudio, cada viernes se le proveerá con dos envases de bebidas. Por favor de consumir la bebida de un envase el sábado y la otra el domingo.
- 3. Consuma la bebida dentro de 30 minutos de haber comido para ayudar con la digestión de la bebida.
- 4. <u>No</u> consuma ninguno de los siguientes suplementos por la duración de su participación en el tratamiento/estudio, ya que estos pueden cambiar la manera como su cuerpo maneja el tratamiento que esta recibiendo.

a.	Multivitamínico	e. Vitamina A	i. Aceite de pescado
b.	Palma enana americana	f. Vitamina C	j. EPA/DHA
c.	Licopeno	g. Vitamina E	k. Linaza
d.	Beta Caroteno	h. Omega-3	 Aceite de linaza

- 5. Antes de tomar algún otro suplemento o vitaminas, por favor revise con su doctor de radiación y el coordinador del estudio. Algunos de estos suplementos pueden disminuir los beneficios del tratamiento de radiación que está recibiendo.
- 6. Se le pedirá completar un cuestionario de frecuencia de alimentos al principio del estudio. Le tomará una hora completarlo. Por favor de contestar todas las preguntas lo más preciso posible y entréguelo al coordinador del estudio, una semana antes de su último tratamiento. No hay ninguna contestación correcta o incorrecta.
- 7. Por favor déjele saber a su doctor de radiación y al coordinador del estudio si hay algún cambio en sus defecaciones, problemas de la vejiga o si experimenta otros problemas como náusea, vomito o ardor de estómago.

Se puede comunicar con la coordinadora del estudio Mri Datta al 307-9020

APPENDIX I. NATIONAL CANCER INSTITUTE DIET HISTORY QUESTIONNAIRE

NATIONAL INSTITUTES OF HEALTH

Diet History Questionnaire



GENERAL INSTRUCTIONS

- Answer each question as best you can. Estimate if you are not sure. A guess is better than leaving a blank.
- Use only a black ball-point pen. Do not use a pencil or felt-tip pen. Do not fold, staple, or tear the pages.
- Put an X in the box next to your answer.
- If you make any changes, cross out the incorrect answer and put an X in the box next to the correct answer. Also draw a circle around the correct answer.
- If you mark NEVER, NO, or DON'T KNOW for a question, please follow any arrows or instructions that direct you to the next question.

BEFORE TURNING THE PAGE, PLEASE COMPLETE THE FOLLOWING QUESTIONS.

Today's date:

MONTH D	YEAR		
☐ Jan ☐ Fe b ☐ Mar ☐ Ap r ☐ May ☐ J un ☐ Ju l ☐ Au g ☐ Sep ☐ Oct ☐ No v ☐ Dec		0123456789	☐ 2007 ☐ 2008 ☐ 2009 ☐ 2010 ☐ 2011

In wh	at month	were
vou h	orn?	

☐ Jan
☐ Fe b
☐ Mar
□Apr
☐ May
□Jun
□JuI
☐ Au g
☐ Sep
☐ Oct
□ No v
☐ Dec

In what year were you born?

Are you male or female?

☐Male ☐Female

BAR CODE LABEL OR SUBJECT ID HERE

1		er the past 12 months, how often did you drink		Οv	er the	past 12 months	
	ton	nato juice or vegetable juice?		1	Цом с	often did vou drink eth	er fruit drinks (such
		NEVER (GO TO QUESTION 2)		4.	as cra	nberry cocktail, Hi-C, iet or regular)?	
		1 time per month or less 2–3 times per month 1–2 times per week 3–4 times per week 5–6 times per week 1 time per day 2–3 times per day 4–5 times per day 6 or more times per day			-	EVER (GO TO QUESTI time per month or less 3 times per month	ON 5) 1 time per day 2–3 times per day
	1a.	Each time you drank tomato juice or vegetable juice , how much did you usually drink?			☐ 1- ☐ 3- ☐ 5-	2 times per week 4 times per week 6 times per week	☐ 4–5 times per day ☐ 6 or more times per day
	,	Less than ¾ cup (6 ounces) ¾ to 1¼ cups (6 to 10 ounces) More than 1¼ cups (10 ounces)		4	di _:	d you usually drink?] Less than 1 cup (8 our	uit drinks, how much
2		er the <u>past 12 months</u> , how often did you drink nge juice or grapefruit juice?] 1 to 2 cups (8 to 16 ou] More than 2 cups (16	
ſ		NEVER (GO TO QUESTION 3)		4		ow often were your fro ugar-free drinks?	uit drinks diet or
		1 time per month or less 2–3 times per month 2 2–3 times per day 2–3 times per day 4–5 times per day 3–4 times per week 5–6 times per week	,			Almost never or never About ¼ of the time About ½ of the time About ¾ of the time About ¾ of the time Almost always or alwa	
	2a.	Each time you drank orange juice or grapefruit juice , how much did you usually drink?		5.	(NOT	often did you drink mi l in coffee, NOT in cero late milk and hot choo	eal)? (Please include
	,	Less than ¾ cup (6 ounces) ¾ to 1¼ cups (6 to 10 ounces) More than 1¼ cups (10 ounces)			□ 1 f	EVER (GO TO QUESTI	☐ 1 time per day
3	oth mix	er the past 12 months, how often did you drink er 100% fruit juice or 100% fruit juice (such as apple, grape, pineapple, or ers)?			☐ 1- ☐ 3-	3 times per month 2 times per week 4 times per week 6 times per week	☐ 2–3 times per day ☐ 4–5 times per day ☐ 6 or more times per day
ſ		NEVER (GO TO QUESTION 4)				ach time you drank m ow much did you usua	
		1 time per month or less 2–3 times per month 1–2 times per week 3–4 times per week 5–6 times per week 1 time per day 2–3 times per day 4–5 times per day 6 or more times per day			5b. W	Less than 1 cup (8 our 1 to 1½ cups (8 to 12 o More than 1½ cups (1 hat kind of milk did y	ounces) 2 ounces)
	3a.	Each time you drank other fruit juice or fruit juice mixtures , how much did you usually drink?] Whole milk] 2% fat milk] 1 % fat milk] Skim, nonfat, or ½% fa	at milk
		Less than ¾ cup (6 ounces) ¾ to 1½ cups (6 to 12 ounces) More than 1½ cups (12 ounces)				Soy milk Rice milk Other	

Over the past 12 months			/	d.	pop diet or sugar-free?		
6	ene Ins	w often did you drink me ergy, or high-protein be tant Breakfast, Ensure, S ers?	everages such as			☐ Almost never or never ☐ About 1/4 of the time ☐ About 1/2 of the time ☐ About 3/4 of the time	
ſ	— _□	NEVER (GO TO QUESTION	ON 7)			Almost always or always	
		1 time per month or less 2–3 times per month 1–2 times per week 3–4 times per week 5–6 times per week	☐ 1 time per day ☐ 2–3 times per day ☐ 4–5 times per day ☐ 6 or more times per day		e.	How often were these soft drinks, soda, or pop caffeine-free? Almost never or never About 1/4 of the time About 1/2 of the time	
	6a.	Each time you drank m beverages , how much				☐ About ¾ of the time ☐ Almost always or always	
\ \ 2		Less than 1 cup (8 our 1 to 1½ cups (8 to 12 c More than 1½ cups (12	ounces) 2 ounces)	8.	8. Over the past 12 months, did you drink beer? NO (GO TO QUESTION 9) T YES		
1		er the past 12 months, d nks, soda, or pop?	ia you aririk sort		,		
ſ		NO (GO TO QUESTION 8)	8	a.	How often did you drink beer IN THE SUMMER ?	
	Γ^{\square}	YES			□NEVER		
	♦ 7a.	How often did you drink or pop IN THE SUMME ☐ NEVER				☐ 1 time per month or less ☐ 2–3 times per month ☐ 1–2 times per week ☐ 3–4 times per week ☐ 5–6 times per week ☐ 1 time per day ☐ 2–3 times per day ☐ 4–5 times per day ☐ 6 or more times ☐ per day	
		☐ 1 time per month or les☐ 2–3 times per month☐ 1–2 times per week☐ 3–4 times per week☐ 5–6 times per week	1 time per day 2–3 times per day 4–5 times per day 6 or more times per day	8	b.	How often did you drink beer DURING THE REST OF THE YEAR?	
	7b.	How often did you drink or pop DURING THE R ☐ NEVER				☐ 1 time per month or less ☐ 2–3 times per month ☐ 1–2 times per week ☐ 3–4 times per week ☐ 5–6 times per week ☐ 1 time per day ☐ 2–3 times per day ☐ 4–5 times per day ☐ 6 or more times ☐ per day	
	7c.	☐ 1 time per month or les☐ 2–3 times per month☐ 1–2 times per week☐ 3–4 times per week☐ 5–6 times per wee	2–3 times per day 4–5 times per day 6 or more times per day oft drinks, soda, or	8	SC.	Each time you drank beer , how much did you usually drink? Less than a 12-ounce can or bottle 1 to 3 12-ounce cans or bottles More than 3 12-ounce cans or bottles	
	,	12 to 16 ounces or 1 c	or less than 1 can or bottle				

Over the past 12 months	11b. How often did you eat oatmeal, grits, or other cooked cereal DURING THE REST
9. How often did you drink wine or wine coolers?	OF THE YEAR?
☐ NEVER (GO TO QUESTION 10)	□NEVER
☐ 1 time per month or less ☐ 1 time per day ☐ 2–3 times per month ☐ 2–3 times per day ☐ 1–2 times per week ☐ 4–5 times per day ☐ 3–4 times per week ☐ 6 or more times per day ☐ 5–6 times per week	☐ 1–6 times per year ☐ 7–11 times per year ☐ 1 time per month ☐ 2–3 times per month ☐ 1 time per week ☐ 2 times per week ☐ 3–4 times per week ☐ 5–6 times per week ☐ 1 time per day ☐ 2 or more times ☐ per day
9a. Each time you drank wine or wine coolers, how much did you usually drink? Less than 5 ounces or less than 1 glass 5 to 12 ounces or 1 to 2 glasses More than 12 ounces or more than 2 glasses	11c. Each time you ate oatmeal, grits, or other cooked cereal, how much did you usually eat?Less than ¾ cup
10. How often did you drink liquor or mixed drinks ?	☐ ¾ to 1¼ cups ☐ More than 1¼ cups
☐ NEVER (GO TO QUESTION 11)	12. How often did you eat cold cereal ?
□ 1 time per month or less □ 1 time per day □ 2–3 times per month □ 2–3 times per day □ 1–2 times per week □ 4–5 times per day □ 3–4 times per week □ 6 or more times per day □ 5–6 times per week 10a. Each time you drank liquor or mixed drinks, how much did you usually drink? □ Less than 1 shot of liquor □ 1 to 3 shots of liquor □ More than 3 shots of liquor □ More than 3 shots of liquor □ NO (GO TO QUESTION 12) □ YES □ 11a. How often did you eat oatmeal, grits, or	NEVER (GO TO QUESTION 13) 1–6 times per year
other cooked cereal IN THE WINTER? □ NEVER □ 1–6 times per winter □ 2 times per week □ 7–11 times per winter □ 3–4 times per week □ 1 time per month □ 5–6 times per week □ 2–3 times per month □ 1 time per day □ 1 time per week □ 2 or more times per day	12c. How often was the cold cereal you ate All Bran, Fiber One, 100% Bran, or Bran Buds? Almost never or never About ¼ of the time About ½ of the time About ¾ of the time Almost always or always

Over the past 12 months	13a. Each time you ate applesauce, how much did you usually eat?
12d. How often was the cold cereal you ate some other bran or fiber cereal (such as Cheerios, Shredded Wheat, Raisin Bran, Bran Flakes, Grape-Nuts, Granola, Wheaties, or Healthy Choice)?	☐ Less than ½ cup ☐ ½ to 1 cup ☐ More than 1 cup
□ Almost never or never □ About ½ of the time □ About ¾ of the time □ Almost always or always 12e. How often was the cold cereal you ate any other type of cold cereal (such as Corn Flakes, Rice Krispies, Frosted Flakes, Special K, Froot Loops, Cap'n Crunch, or others)? □ Almost never or never □ About ¼ of the time □ About ¾ of the time □ About ¾ of the time □ Almost always or always	14. How often did you eat apples? NEVER (GO TO QUESTION 15) 1–6 times per year
12f. Was milk added to your cold cereal?	frozen)? — NEVER (GO TO QUESTION 16)
□ NO (GO TO QUESTION 13) □ YES 12g. What kind of milk was usually added?	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 1 time per week ☐ 2 or more times per day
☐ Whole milk ☐ 2% fat milk ☐ 1% fat milk ☐ Skim, nonfat, or ½% fat milk ☐ Soy milk ☐ Rice milk ☐ Other	15a. Each time you ate pears , how many did you usually eat? Less than 1 pear 1 pear More than 1 pear
12h. Each time milk was added to your cold cereal, how much was usually added?	16. How often did you eat bananas? NEVER (GO TO QUESTION 17) 1–6 times per year
☐ ½ to 1 cup ☐ More than 1 cup 13. How often did you eat applesauce?	☐ 7-11 times per year ☐ 3-4 times per week ☐ 1 time per month ☐ 5-6 times per week ☐ 2-3 times per month ☐ 1 time per day ☐ 1 time per week ☐ 2 or more times per day
□ NEVER (GO TO QUESTION 14) □ 1–6 times per year □ 2 times per week □ 7–11 times per year □ 3–4 times per week □ 1 time per month □ 5–6 times per week □ 2–3 times per month □ 1 time per day □ 1 time per day □ 1 time per day	

Over the past 12 months	18c. Each time you ate peaches , nectarines , or plums , how much did you usually eat?
16a. Each time you ate bananas, how many did you usually eat?☐ Less than 1 banana☐ 1 banana☐ More than 1 banana	Less than 1 fruit or less than ½ cup 1 to 2 fruits or ½ to ¾ cup More than 2 fruits or more than ¾ cup 19. How often did you eat grapes ?
17. How often did you eat dried fruit , such as prunes or raisins (not including dried apricots)? NEVER (GO TO QUESTION 18) 1-6 times per year 2 times per week 7-11 times per year 3-4 times per week 1 time per month 5-6 times per week 2-3 times per month 1 time per day 1 time per week 2 or more times per day 17a. Each time you ate dried fruit , how much did you usually eat (not including dried apricots)? Less than 2 tablespoons 2 to 5 tablespoons More than 5 tablespoons More than 5 tablespoons 18. Over the past 12 months, did you eat peaches, nectarines, or plums?	NEVER (GO TO QUESTION 20) 1–6 times per year 2 times per week 7–11 times per year 3–4 times per week 1 time per month 5–6 times per week 2–3 times per month 1 time per day 1 time per week 2 or more times per day 19a. Each time you ate grapes , how much did you usually eat? Less than ½ cup or less than 10 grapes ½ to 1 cup or 10 to 30 grapes More than 1 cup or more than 30 grapes More than 1 cup or more than 30 grapes NO (GO TO QUESTION 21) NO (GO TO QUESTION 21)
NO (GO TO QUESTION 19) YES 18a. How often did you eat fresh peaches, nectarines, or plums WHEN IN SEASON? NEVER 1-6 times per season 2 times per week 3-4 times per week 5-6 times per week 1 time per month 1 time per day 2 or more times per day 18b. How often did you eat peaches, nectarines, or plums (fresh, canned, or frozen) DURING THE REST OF THE YEAR? NEVER 1-6 times per year 2 times per week 3-4 times per week 3-4 times per week 5-6 times per week 1 time per month 5-6 times per week 2-3 times per month 1 time per day 2 or more times per day	20a. How often did you eat fresh cantaloupe WHEN IN SEASON? NEVER 1-6 times per season 2 times per week 7-11 times per season 3-4 times per week 1 time per month 5-6 times per week 2-3 times per month 1 time per day 2 or more times per day 20b. How often did you eat fresh or frozen cantaloupe DURING THE REST OF THE YEAR? NEVER 1-6 times per year 2 times per week 7-11 times per year 3-4 times per week 1 time per month 5-6 times per week 2-3 times per month 1 time per day 2 or more times per day

Over the past 12 months	22. Over the <u>past 12 months</u> , did you eat strawberries?
20c. Each time you ate cantaloupe , how much did you usually eat?	□ NO (GO TO QUESTION 23)
Less than ¼ melon or less than ½ cup ¼ melon or ½ to 1 cup More than ¼ melon or more than 1 cup 21. Over the past 12 months, did you eat melon, other than cantaloupe (such as watermelon or honeydew)?	YES 22a. How often did you eat fresh strawberries WHEN IN SEASON? □ NEVER
NO (GO TO QUESTION 22) THE YES 21a. How often did you eat fresh melon, other than cantaloupe (such as watermelon or honeydew) WHEN IN SEASON?	☐ 1–6 times per season ☐ 2 times per week ☐ 7–11 times per season ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 1 time per week ☐ 2 or more times per day 22b. How often did you eat fresh or frozen strawberries DURING THE REST OF THE YEAR
□ NEVER □ 1–6 times per season □ 2 times per week □ 7–11 times per season □ 3–4 times per week □ 1 time per month □ 5–6 times per week □ 2 times per week □ 3–4 times per week □ 1 time per day □ 1 time per day □ 1 time per day □ 2 or more times □ 2 times per week □ 2 times per week	□ NEVER □ 1–6 times per year □ 2 times per week □ 7–11 times per year □ 3–4 times per week □ 1 time per month □ 5–6 times per week □ 2–3 times per month □ 1 time per day □ 1 time per week □ 2 or more times per day
21b. How often did you eat fresh or frozen melon, other than cantaloupe (such as watermelon or honeydew) DURING THE REST OF THE YEAR? NEVER 1-6 times per year 2 times per week 3-4 times per week 1 time per month 5-6 times per week 2-3 times per month 1 time per day 2 or more times per day 1 time per week 2 or more times per day 21c. Each time you ate melon other than cantaloupe, how much did you usually eat? Less than ½ cup or 1 small wedge ½ to 2 cups or 1 medium wedge More than 2 cups or 1 large wedge	22c. Each time you ate strawberries , how much did you usually eat? Less than ¼ cup or less than 3 berries ½ to ¾ cup or 3 to 8 berries More than ¾ cup or more than 8 berries 23. Over the past 12 months, did you eat oranges , tangerines, or tangelos? NO (GO TO QUESTION 24) YES 23a. How often did you eat fresh oranges, tangerines, or tangelos WHEN IN SEASON? NEVER
	☐ 1–6 times per season ☐ 2 times per week ☐ 7–11 times per season ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 2 or more times per day

Over the past 12 months	25. How often did you eat other kinds of fruit?
23b. How often did you eat oranges, tangerines, or tangelos (fresh or canned) DURING THE REST OF THE YEAR? NEVER 1-6 times per year	NEVER (GO TO QUESTION 26) 1–6 times per year
per day 23c. Each time you ate oranges, tangerines, or tangelos, how many did you usually eat? Less than 1 fruit 1 fruit More than 1 fruit	☐ Less than ¼ cup ☐ ¼ to ¾ cup ☐ More than ¾ cup 26. How often did you eat COOKED greens (such as spinach, turnip, collard, mustard, chard, or kale)? ☐ NEVER (GO TO QUESTION 27)
24. Over the past 12 months, did you eat grapefruit? NO (GO TO QUESTION 25) YES 24a. How often did you eat fresh grapefruit WHEN IN SEASON? NEVER 1-6 times per season 2 times per week 5-6 times per week 1 time per month 2-3 times per month 1 time per week 2 or more times per day 24b. How often did you eat grapefruit (fresh or canned) DURING THE REST OF THE YEAR? NEVER 1-6 times per year 7-11 times per year 1 time per month 2-3 times per month 2-3 times per month 1 time per day 2 times per week 2-1 time per month 2-3 times per year 1 time per day 2 times per week 2 or more times per week 2 or more times per day 24c. Each time you ate grapefruit, how much did you usually eat?	1-6 times per year 2 times per week 7-11 times per year 3-4 times per week 1 time per month 5-6 times per week 2-3 times per month 1 time per day 2 or more times per day 26a. Each time you ate COOKED greens, how much did you usually eat? Less than ½ cup ½ to 1 cup More than 1 cup We will ask about lettuce later.) NEVER (GO TO QUESTION 28) 1-6 times per year 2 times per week 7-11 times per year 3-4 times per week 1 time per month 5-6 times per week 2-3 times per month 1 time per day 2 or more times per day 27a. Each time you ate RAW greens, how much did you usually eat? Less than ½ cup ½ to 1 cup More than 1 cup
☐ Less than ½ grapefruit ☐ ½ grapefruit ☐ More than ½ grapefruit ■	

Over the past 12 months	31. How often did you eat string beans or green beans (fresh, canned, or frozen)?
28. How often did you eat coleslaw ?	
NEVER (GO TO QUESTION 29) □ 1–6 times per year □ 2 times per week □ 7–11 times per year □ 3–4 times per week □ 1 time per month □ 5–6 times per week □ 2–3 times per month □ 1 time per day □ 1 time per week □ 2 or more times per day 28a. Each time you ate coleslaw, how much did you usually eat? □ Less than ¼ cup □ ¼ to ¾ cup □ More than ¾ cup □ More than ¾ cup □ How often did you eat sauerkraut or cabbage (other than coleslaw)?	□ NEVER (GO TO QUESTION 32) □ 1–6 times per year □ 2 times per week □ 7–11 times per year □ 3–4 times per week □ 1 time per month □ 5–6 times per week □ 2–3 times per month □ 1 time per day □ 1 time per week □ 2 or more times per day 31a. Each time you ate string beans or green beans, how much did you usually eat? □ Less than ½ cup □ ½ to 1 cup □ More than 1 cup 32. How often did you eat peas (fresh, canned, or frozen)?
(otner tnan colesiaw)? NEVER (GO TO QUESTION 30) 1–6 times per year	□ NEVER (GO TO QUESTION 33) □ 1–6 times per year □ 2 times per week □ 7–11 times per year □ 3–4 times per week □ 1 time per month □ 5–6 times per week □ 2–3 times per month □ 1 time per day □ 1 time per week □ 2 or more times per day 32a. Each time you ate peas , how much did you usually eat? □ Less than ¼ cup □ ¼ to ¾ cup □ ¼ to ¾ cup □ More than ¾ cup 33. Over the past 12 months, did you eat corn ?
30. How often did you eat carrots (fresh, canned, or frozen)? NEVER (GO TO QUESTION 31) 1–6 times per year	NO (GO TO QUESTION 34) TYES 33a. How often did you eat fresh corn WHEN IN SEASON? NEVER 1-6 times per season 7-11 times per season 1 time per month 2-3 times per month 1 time per day 1 time per week 2 or more times per day
	画面が

Over the past 12 months	36. How often did you eat mixed vegetables ?
33b. How often did you eat corn (fresh, canned, or frozen) DURING THE REST OF THE YEAR ?	☐ NEVER (GO TO QUESTION 37)
□ NEVER□ 1–6 times per year□ 2 times per week	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day
☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2 times per week ☐ 2 times per week ☐ 1 time per day	1 time per week 2 or more times per day 36a. Each time you ate mixed vegetables , how
1 time per week 2 or more times per day	much did you usually eat?
33c. Each time you ate corn , how much did you usually eat?	☐ Less than ½ cup ☐ ½ to 1 cup ☐ More than 1 cup
☐ Less than 1 ear or less than ½ cup ☐ 1 ear or ½ to 1 cup	37. How often did you eat onions ?
More than 1 ear or more than 1 cup	☐ NEVER (GO TO QUESTION 38)
34. Over the past 12 months, how often did you eat broccoli (fresh or frozen)?	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day
NEVER (GO TO QUESTION 35)	☐ 1 time per week ☐ 2 or more times per day
☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week	37a. Each time you ate onions , how much did you usually eat?
☐ 2–3 times per month ☐ 1 time per day ☐ 2 or more times per day	☐ Less than 1 slice or less than 1 tablespoon☐ 1 slice or 1 to 4 tablespoons☐ More than 1 slice or more than 4 tablespoons
34a. Each time you ate broccoli , how much did you usually eat?	 Now think about all the cooked vegetables you ate in the past 12 months and how they were
Less than ¼ cup 1/4 to 1 cup More than 1 cup	prepared. How often were your vegetables COOKED WITH some sort of fat , including oil spray? (<i>Please do not include potatoes.</i>)
35. How often did you eat cauliflower or Brussels sprouts (fresh or frozen)?	☐ NEVER (GO TO QUESTION 39)
☐ NEVER (GO TO QUESTION 36)	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week
□ 1–6 times per year □ 2 times per week □ 7–11 times per year □ 3–4 times per week □ 1 time per month □ 5–6 times per week □ 2-3 times per month □ 1 time per day □ 1 time per week □ 2 or more times per day	☐ 2–3 times per month ☐ 1 time per day ☐ 2 or more times per day
35a. Each time you ate cauliflower or Brussels sprouts , how much did you usually eat?	
☐ Less than ¼ cup ☐ ¼ to ½ cup ☐ More than ½ cup	
↓	

Over the <u>past 12 months</u>	40. Over the <u>past 12 months</u> , how often did you eat sweet peppers (green, red, or yellow)?
38a. Which fats were usually added to your vegetables DURING COOKING ? (Please do not include potatoes. Mark all that apply.)	□ NEVER (GO TO QUESTION 41)
☐ Margarine ☐ Corn oil (including low-fat) ☐ Canola or rapeseed oil ☐ Butter (including ☐ Oil spray, such as Pam low-fat) ☐ or others ☐ Lard, fatback, or ☐ Other kinds of oils	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 1 time per week ☐ 2 or more times per day
☐ Lard, fatback, or ☐ Other kinds of oils bacon fat ☐ None of the above ☐ Olive oil	40a. Each time you ate sweet peppers , how much did you usually eat?
39. Now, thinking again about all the cooked vegetables you ate in the <u>past 12 months</u> , how often was some sort of fat, sauce, or dressing added AFTER COOKING OR AT THE TABLE ? (Please do not include potatoes.)	Less than ½ pepper ½ to ½ pepper More than ½ pepper 41. Over the past 12 months, did you eat fresh tomatoes (including those in salads)?
☐ NEVER (GO TO QUESTION 40)	NO (GO TO QUESTION 42)
☐ 1–6 times per year ☐ 3–4 times per week ☐ 7–11 times per year ☐ 5–6 times per week ☐ 1 time per month ☐ 1 time per day ☐ 2–3 times per month ☐ 2 times per day ☐ 1–2 times per week ☐ 3 or more times per day	YES 41a. How often did you eat fresh tomatoes (including those in salads) WHEN IN
39a. Which fats, sauces, or dressings were usually added AFTER COOKING OR AT THE TABLE? (Please do not include potatoes. Mark all that apply.)	SEASON?
☐ Margarine ☐ Salad dressing (including low-fat) ☐ Cheese sauce ☐ Butter (including ☐ White sauce low-fat) ☐ Other ☐ Lard, fatback, or	☐ 1–6 times per season ☐ 2 times per week☐ 7–11 times per season☐ 3–4 times per week☐ 1 time per month☐ 5–6 times per week☐ 2–3 times per month☐ 1 time per day☐ 1 time per week☐ 2 or more times☐ 2 or more times☐ 2 per day☐ 2 or more times☐ 2 times☐ 3–4 tim
39b. If margarine, butter, lard, fatback, or bacon fat was added to your cooked vegetables	41b. How often did you eat fresh tomatoes (including those in salads) DURING THE REST OF THE YEAR?
AFTER COOKING OR AT THE TABLE, how much did you usually add?	□ NEVER
☐ Did not usually add these ☐ Less than 1 teaspoon ☐ 1 to 3 teaspoons ☐ More than 3 teaspoons	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 1 time per week ☐ 2 or more times per day
39c. If salad dressing, cheese sauce, or white sauce was added to your cooked vegetables AFTER COOKING OR AT THE TABLE, how much did you usually add?	41c. Each time you ate fresh tomatoes , how much did you usually eat?
☐ Did not usually add these ☐ Less than 1 tablespoon ☐ 1 to 3 tablespoons ☐ More than 3 tablespoons	☐ Less than ¼ tomato ☐ ¼ to ½ tomato ☐ More than ½ tomato

Over the past 12 months	45. How often did you eat French fries, nome fries, hash browned potatoes, or tater tots?
42. How often did you eat lettuce salads (with or	•
without other vegetables)?	☐ NEVER (GO TO QUESTION 46)
☐ NEVER (GO TO QUESTION 43)	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week
☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day	☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 1 time per week ☐ 2 or more times per day
1 time per week 2 or more times per day	45a. Each time you ate French fries, home fries, hash browned potatoes, or tater tots how
42a. Each time you ate lettuce salads , how much did you usually eat?	much did you usually eat?
☐ Less than ¼ cup ☐ ¼ to 1¼ cups ☐ More than 1¼ cups	10 to 25 fries or ½ to 1 cup More than 25 fries or more than 1 cup
↓ 43. How often did you eat salad dressing (including	46. How often did you eat potato salad ?
low-fat) on salads?	☐ NEVER (GO TO QUESTION 47)
☐ NEVER (GO TO QUESTION 44)	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week
☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day	☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 1 time per week ☐ 2 or more times per day
☐ 1 time per week ☐ 2 or more times per day	46a. Each time you ate potato salad , how much did you usually eat?
43a. Each time you ate salad dressing on salads, how much did you usually eat?	Less than ½ cup
Less than 2 tablespoons 2 to 4 tablespoons	☐ More than 1 cup
☐ More than 4 tablespoons	47. How often did you eat baked, boiled, or mashed potatoes?
44. How often did you eat sweet potatoes or yams?	☐ NEVER (GO TO QUESTION 48)
☐ NEVER (GO TO QUESTION 45)	
☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 2 or more times per day	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 1 time per week ☐ 2 or more times per day
44a. Each time you ate sweet potatoes or yams , how much did you usually eat?	47a. Each time you ate baked, boiled, or mashed potatoes , how much did you usually eat?
☐ 1 small potato or less than ¼ cup ☐ 1 medium potato or ¼ to ¾ cup ☐ 1 large potato or more than ¾ cup	☐ 1 small potato or less than ½ cup☐ 1 medium potato or ½ to 1 cup☐ 1 large potato or more than 1 cup
1	

47b. How often was sour cream (including low-fat) added to your potatoes, EITHER IN COOKING OR AT THE TABLE? Almost never or never (GO TO QUESTION 47d) About ½ of the time About ½ of the time About ½ of the time Less than 1 tablespoons About ½ of the time About ½ of the	Over the	past 12 months	47h.	Each time cheese of added to your potate	
Almost never or never (GO TO QUESTION 47d) About ½ of the time	fa	at) added to your potatoes, EITHER IN		usually added? Less than 1 tables	
Almost always or always		About ¼ of the time	40.11	☐ More than 3 tables	•
47c. Each time sour cream was added to your potatoes, how much was usually added? Less than 1 tablespoons Less than 1 tablespoons Less than 3 tablespoons Less than 3 tablespoons Less than 3 tablespoons Less than 4 tablespoons Less than 5 tablespoons Less than 6 tablespoons Less than 6 tablespoons Less than 6 tablespoons Less than 6 tablespoons Less than 7 tablespoons Less than 1 tablespoon Less than 6 tablespoons Less than 1 tablespoon Less than 1 tablespoon		About ¾ of the time		•	
potatoes, how much was usually added? Less than 1 tablespoon 1 to 3 tablespoons More than 3 tablespoons More than 3 tablespoons More than 3 tablespoons More than 3 tablespoons Almost never or never About ½ of the time About ½ of the time About 5 of the time About	47c. E			_	_
1 time per week 2 or more times				7–11 times per year 1 time per month	☐ 3–4 times per week ☐ 5–6 times per week
47d. How often was margarine (including low-fat) added to your potatoes, EITHER IN COOKING OR AT THE TABLE? Almost never or never		1 to 3 tablespoons			☐ 1 time per day ☐ 2 or more times per day
Less than 1 tablespoon 1 to 5 tablespoons 2 times per were 3 times per year 2 times per were 3 times per were 49 times per were 4	→47d. H	low often was margarine (including low-fat)	48a.		alsa, how much did you
About ½ of the time About ½ of the time About ½ of the time Almost always or always		OOKING OR AT THE TABLE?		☐ 1 to 5 tablespoons	
Almost always or always		About ¼ of the time About ½ of the time	↓ 49. Ho		•
added to your potatoes, EITHER IN COOKING OR AT THE TABLE? Almost never or never 1 time per month 5-6 times per we 2-3 times per wenth 1 time per day 1 time per week 2 or more times 49a. Each time you ate catsup, how much do usually eat? Almost always or always 47f. Each time margarine or butter was added to your potatoes, how much was usually added? Never added Less than 1 teaspoon 1 to 3 teaspoons More than 3 teaspoons More than 3 teaspoons 1-6 times per year 2 times per week 2-3 times per year 2 times per week 3-4 times per				NEVER (GO TO QUE	STION 50)
About ⅓ of the time Usually eat? Less than 1 teaspoon 1 to 6 teaspoons More than 6 teaspoons More than 1 teaspoon More than 6 teaspoons More than 3 teaspoons Teaspoons More than 6 teaspoons More than 7 teaspoons More than 8 teaspoons More than 9 teaspoons More than 9 teaspoons More than 9 teaspoons More than 9 teaspoons More than 1 teaspoon More than 1 teaspoons More than 1 teaspoons More than 1 teaspoons More than 1 teaspoons More than 2 teaspoons More than 3 teaspoons More than 3 teaspoons More than 3 teaspoons More than 1 teaspoons More than 2 teaspoons More than 3 teaspoons More than 3 teaspoons More than 3 teaspoons More than 6 teaspoons	ad	dded to your potatoes, EITHER IN COOKING OR AT THE TABLE?		7–11 times per year 1 time per month 2–3 times per month	☐ 2 times per week ☐ 3–4 times per week ☐ 5–6 times per week ☐ 1 time per day ☐ 2 or more times per day
Less than 1 teaspoon 1 to 6 teaspoons More than 6 teaspoons dumplings? More than 6 teaspoons dumplings? More than 6 teaspoons More than 6 teaspoons More than 6 teaspoons dumplings? More than 6 teaspoons More than 6 teaspoons dumplings? dumplings? More than 6 teaspoons dumplings? dumplings? More than 6 teaspoons dumplings? dumplings du		About ¼ of the time About ½ of the time About ¾ of the time	49a.	Each time you ate c	
 Never added	yo	each time margarine or butter was added to our potatoes, how much was usually		1 to 6 teaspoons	
Less than 1 teaspoon 1 to 3 teaspoons More than 3 teaspoons 1-6 times per year 2 times per wee 7-11 times per year 3-4 times per wee 7-11 time per month 1 time per month 1 time per day 1 time per week 2 or more times	Г	_			tuffing, dressing, or
47g. How often was cheese or cheese sauce added to your potatoes, EITHER IN COOKING OR AT THE TABLE? □ 7-11 times per year □ 1 time per month □ 5-6 times per w □ 1 time per month □ 1 time per day □ 1 time per week □ 2 or more times		Less than 1 teaspoon 1 to 3 teaspoons			STION 51)
<u> </u>	a	dded to your potatoes, EITHER IN		7–11 times per year 1 time per month 2–3 times per month	☐ 2 times per week ☐ 3–4 times per week ☐ 5–6 times per week ☐ 1 time per day ☐ 2 or more times per day
Almost never or never (GO TO QUESTION 48) About ½ of the time About ½ of the time About ¾ of the time About ¾ of the time About ¾ of the time		About ¼ of the time About ½ of the time	50a.		
☐ Almost always or always ☐ Less than ½ cup ☐ ½ to 1 cup ☐ More than 1 cup				☐ ½ to 1 cup	

Over the past 12 months	53b. How often were the beans you ate refried
51. How often did you eat chili ?	beans, beans prepared with any type of fat, or with meat added?
NEVER (GO TO QUESTION 52) 1–6 times per year	Almost never or never About ¼ of the time About ½ of the time About ¾ of the time Almost always or always 54. How often did you eat other kinds of vegetables? NEVER (GO TO QUESTION 55) 1–6 times per year 2 times per week 7–11 times per year 3–4 times per week 1 time per month 5–6 times per week 2–3 times per month 1 time per day 1 time per week 2 or more times per day 54a. Each time you ate other kinds of vegetables, how much did you usually eat? Less than ¼ cup ¼ to ½ cup More than ½ cup
□ 2–3 times per month □ 1 time per day □ 1 time per week □ 2 or more times per day 52a. Each time you ate Mexican foods , how much did you usually eat? □ Less than 1 taco, burrito, etc. □ 1 to 2 tacos, burritos, etc. □ More than 2 tacos, burritos, etc. 53. How often did you eat cooked dried beans (such as baked beans, pintos, kidney, blackeyed peas, lima, lentils, soybeans, or refried beans)? (Please don't include bean soups or chili.)	55. How often did you eat rice or other cooked grains (such as bulgur, cracked wheat, or millet)? NEVER (GO TO QUESTION 56) 1–6 times per year 2 times per week 3–4 times per week 1 time per month 5–6 times per week 2–3 times per month 1 time per day 1 time per week 2 or more times per day 55a. Each time you ate rice or other cooked grains , how much did you usually eat?
NEVER (GO TO QUESTION 54) 1–6 times per year	□ Less than ½ cup □ ½ to 1½ cups □ More than 1½ cups 55b. How often was butter, margarine, or oil added to your rice IN COOKING OR AT THE TABLE? □ Almost never or never □ About ¼ of the time □ About ½ of the time □ About ¾ of the time □ Almost always or always

Over the past 12 months	56f. Each time syrup was added to your pancakes, waffles, or French toast, how
56. How often did you eat pancakes, waffles, or French toast?	much was usually added?
☐ NEVER (GO TO QUESTION 57)	☐ Less than 1 tablespoon☐ 1 to 4 tablespoons☐ More than 4 tablespoons
☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 1 time per week ☐ 2 or more times per day	57. How often did you eat lasagna, stuffed shells, stuffed manicotti, ravioli, or tortellini? (Please do not include spaghetti or other pasta.)
56a. Each time you ate pancakes, waffles, or French toast, how much did you usually eat? Less than 1 medium piece 1 to 3 medium pieces More than 3 medium pieces	NEVER (GO TO QUESTION 58) □ 1–6 times per year □ 2 times per week □ 7–11 times per year □ 3–4 times per week □ 1 time per month □ 5–6 times per week □ 2–3 times per month □ 1 time per day □ 1 time per week □ 2 or more times per day
56b. How often was margarine (including low-fat) added to your pancakes, waffles, or French toast AFTER COOKING OR AT THE TABLE ?	57a. Each time you ate lasagna, stuffed shells, stuffed manicotti, ravioli, or tortellini, how much did you usually eat? ☐ Less than 1 cup
☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ Almost always or always	☐ 1 to 2 cups ☐ More than 2 cups 58. How often did you eat macaroni and cheese? ☐ NEVER (GO TO QUESTION 59)
56c. How often was butter (including low-fat) added to your pancakes, waffles, or French toast AFTER COOKING OR AT THE TABLE ?	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 2 or more times per day
☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ Almost always or always 56d. Each time margarine or butter was added to your pancakes, waffles, or French toast, how much was usually added?	58a. Each time you ate macaroni and cheese, how much did you usually eat? Less than 1 cup 1 to 1½ cups More than 1½ cups 59. How often did you eat pasta salad or macaroni salad?
☐ Never added ☐ Less than 1 teaspoon ☐ 1 to 3 teaspoons ☐ More than 3 teaspoons	□ NEVER (GO TO QUESTION 60) □ 1–6 times per year □ 2 times per week □ 7–11 times per year □ 3–4 times per week
56e. How often was syrup added to your pancakes, waffles, or French toast? Almost never or never (GO TO QUESTION 57) About ¼ of the time About ½ of the time About ¾ of the time About ¾ of the time Almost always or always	☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 2 or more times per day ☐ 2 or more times per day ☐ 1 time per week ☐ 2 or more times per day

59a. Each time you ate pasta salad or macaroni	
salad, how much did you usually eat?	☐ NEVER (GO TO INTRODUCTION TO QUESTION 62)
☐ Less than ½ cup ☐ ½ to 1 cup ☐ More than 1 cup 60. Other than the pastas listed in Questions 57, 58,	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 2 or more times per day
and 59, how often did you eat pasta, spaghetti, or other noodles?	61a. Each time you ate bagels or English muffins , how many did you usually eat?
☐ NEVER (GO TO QUESTION 61)	
☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week	☐ Less than 1 bagel or English muffin☐ 1 bagel or English muffin☐ More than 1 bagel or English muffin☐
☐ 2–3 times per month ☐ 1 time per day ☐ 2 or more times per day	61b. How often was margarine (including low-fat) added to your bagels or English muffins?
60a. Each time you ate pasta, spaghetti, or other noodles, how much did you usually eat?	☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time
Less than 1 cup	☐ Almost always or always
☐ 1 to 3 cups ☐ More than 3 cups	61c. How often was butter (including low-fat) added to your bagels or English muffins?
60b. How often did you eat your pasta, spaghetti, or other noodles with tomato sauce or spaghetti sauce made WITH meat?	☐ Almost never or never ☐ About 1/2 of the time ☐ About 1/2 of the time
☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time	☐ About ¾ of the time ☐ Almost always or always
☐ About ¾ of the time ☐ Almost always or always	61d. Each time margarine or butter was added to your bagels or English muffins, how much was usually added?
60c. How often did you eat your pasta, spaghetti, or other noodles with tomato sauce or spaghetti sauce made WITHOUT meat?	☐ Never added ☐ Less than 1 teaspoon ☐ 1 to 2 teaspoons ☐ More than 2 teaspoons
☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ About ¾ of the time ☐ Almost always or always	61e. How often was cream cheese (including low-fat) spread on your bagels or English muffins?
60d. How often did you eat your pasta, spaghetti, or other noodles with margarine, butter, oil, or cream sauce?	Almost never or never (GO TO INTRODUCTION TO QUESTION 62) About ¼ of the time About ½ of the time About ¾ of the time
☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ Almost always or always	Almost always or always

Over the past 12 months	62d. Each time mayonnaise or mayonnaise-type dressing was added to your sandwich
61f. Each time cream cheese was added to your bagels or English muffins, how much was usually added?	breads or rolls, how much was usually added?
☐ Less than 1 tablespoon ☐ 1 to 2 tablespoons ☐ More than 2 tablespoons	☐ Less than 1 teaspoon ☐ 1 to 3 teaspoons ☐ More than 3 teaspoons
mane than 2 tablespeems	62e. How often was margarine (including low-fat) added to your sandwich bread or rolls?
The next questions ask about your intake of breads other than bagels or English muffins. First, we will ask about bread you ate as part of sandwiches only. Then we will ask about all other bread you ate.	☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ Almost always or always
62. How often did you eat breads or rolls AS PART OF SANDWICHES (including burger and hot dog rolls)?	62f. How often was butter (including low-fat) added to your sandwich bread or rolls?
☐ NEVER (GO TO QUESTION 63)	☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time
☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week	☐ About ¾ of the time ☐ Almost always or always
☐ 2–3 times per month ☐ 1 time per day ☐ 1 time per week ☐ 2 or more times per day	62g. Each time margarine or butter was added to your sandwich breads or rolls, how much was usually added?
62a. Each time you ate breads or rolls AS PART OF SANDWICHES , how many did you usually eat?	☐ Never added ☐ Less than 1 teaspoon
☐ 1 slice or ½ roll ☐ 2 slices or 1 roll ☐ More than 2 slices or more than 1 roll	☐ 1 to 2 teaspoons ☐ More than 2 teaspoons
62b. How often were the breads or rolls that you	63. How often did you eat breads or dinner rolls , NOT AS PART OF SANDWICHES ?
used for your sandwiches white bread (including burger and hot dog rolls)?	NEVER (GO TO QUESTION 64)
☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ Almost always or always	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 2 or more times per day
62c. How often was mayonnaise or mayonnaise-type dressing (including lowfat) added to your sandwich bread or rolls?	63a. Each time you ate breads or dinner rolls , NOT AS PART OF SANDWICHES , how much did you usually eat?
Almost never or never (GO TO QUESTION 62e) About ¼ of the time About ½ of the time	☐ 1 slice or 1 dinner roll ☐ 2 slices or 2 dinner rolls ☐ More than 2 slices or 2 dinner rolls
☐ About ¾ of the time ☐ Almost always or always	
▼ Question 62e appears in the next column	Y

Over the past 12 months	64. How often did you eat jam, jelly, or honey on bagels, muffins, bread, rolls, or crackers?
63b. How often were the breads or rolls you ate white bread?	☐ NEVER (GO TO QUESTION 65)
☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ Almost always or always	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 1 time per week ☐ 2 or more times per day
63c. How often was margarine (including low-fat) added to your breads or rolls?	64a. Each time you ate jam, jelly, or honey , how much did you usually eat?
☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ Almost always or always	☐ Less than 1 teaspoon ☐ 1 to 3 teaspoons ☐ More than 3 teaspoons 65. How often did you eat peanut butter or other nut butter ?
63d. How often was butter (including low-fat) added to your breads or rolls?	☐ NEVER (GO TO QUESTION 66)
☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ Almost always or always	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 1 time per week ☐ 2 or more times per day
63e. Each time margarine or butter was added to your breads or rolls, how much was usually added?	65a. Each time you ate peanut butter or other nut butter , how much did you usually eat?
 Never added Less than 1 teaspoon 1 to 2 teaspoons More than 2 teaspoons	Less than 1 tablespoon 1 to 2 tablespoons More than 2 tablespoons
63f. How often was cream cheese (including low-fat) added to your breads or rolls?	♦ 66. How often did you eat roast beef or steak IN SANDWICHES?
Almost never or never (GO TO QUESTION 64) About 1/4 of the time About 3/4 of the time About 3/4 of the time Almost always or always 63g. Each time cream cheese was added to your breads or rolls, how much was usually added? Less than 1 tablespoon 1 to 2 tablespoons More than 2 tablespoons	NEVER (GO TO QUESTION 67) 1–6 times per year

Over the <u>past 12 months</u>	69. How often did you eat other cold cuts or
67. How often did you eat turkey or chicken COLD CUTS (such as loaf, luncheon meat, turkey ham, turkey salami, or turkey pastrami)? (We will ask about other turkey or chicken later.)	luncheon meats (such as bologna, salami, corned beef, pastrami, or others, including low-fat)? (Please do not include ham, turkey, or chicken cold cuts.)
NEVER (GO TO QUESTION 68) 1–6 times per year	NEVER (GO TO QUESTION 70) 1–6 times per year
ham? (We will ask about other ham later.) NEVER (GO TO QUESTION 69) 1-6 times per year	free cold cuts or luncheon meats? (Please do not include ham, turkey, or chicken cold cuts.) Almost never or never About ½ of the time About ½ of the time About ¾ of the time Almost always or always 70. How often did you eat canned tuna (including in salads, sandwiches, or casseroles)? NEVER (GO TO QUESTION 71) 1-6 times per year 7-11 times per year 7-11 times per year 1 time per month 2-3 times per month 1 time per day 1 time per week 2 or more times per day 70a. Each time you ate canned tuna, how much did you usually eat? Less than ¼ cup or less than 2 ounces More than ½ cup or more than 3 ounces More than ½ cup or more than 3 ounces
	water-packed tuna? ☐ Almost never or never ☐ About ½ of the time ☐ About ¾ of the time ☐ About ¾ of the time ☐ About ¾ of the time ☐ Almost always or always

Over the past 12 months	73. How often did you eat ground beef in mixtures (such as meatballs, casseroles, chili, or
70c. How often was the canned tuna you ate	meatloaf)?
prepared with mayonnaise or other	
dressing (including low-fat)?	☐ NEVER (GO TO QUESTION 74)
Almost never or never	☐ 1–6 times per year ☐ 2 times per week
☐ About ¼ of the time ☐ About ½ of the time	☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week
About ¾ of the time	☐ 2–3 times per month ☐ 1 time per day
☐ Almost always or always	☐ 1 time per week ☐ 2 or more times per day
71. How often did you eat GROUND chicken or	73a. Each time you ate ground beef in mixtures ,
turkey? (We will ask about other chicken and	how much did you usually eat?
turkey later.)	☐ Less than 3 ounces or less than ½ cup
☐ NEVER (GO TO QUESTION 72)	3 to 8 ounces or ½ to 1 cup
☐ 1–6 times per year ☐ 2 times per week	☐ More than 8 ounces or more than 1 cup
☐ 7–11 times per year ☐ 3–4 times per week	74. How often did you eat hot dogs or frankfurters?
☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day	(Please do not include sausages or vegetarian
☐ 1 time per week ☐ 2 or more times per day	hot dogs.)
	☐ NEVER (GO TO QUESTION 75)
71a. Each time you ate GROUND chicken or	☐ 1–6 times per year ☐ 2 times per week
turkey, how much did you usually eat?	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week
	☐ 1 time per month ☐ 5–6 times per week
Less than 2 ounces or less than ½ cup 2 to 4 ounces or ½ to 1 cup	☐ 2–3 times per month ☐ 1 time per day ☐ 1 time per week ☐ 2 or more times per day
☐ More than 4 ounces or more than 1 cup	
▼ 72. How often did you eat beef hamburgers or	74a. Each time you ate hot dogs or frankfurters ,
cheeseburgers?	how many did you usually eat?
—	Less than 1 hot dog
☐ NEVER (GO TO QUESTION 73)	☐ 1 to 2 hot dogs ☐ More than 2 hot dogs
☐ 1–6 times per year ☐ 2 times per week	I more than 2 net dogs
☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week	74b. How often were the bet dogs or freakfurters
☐ 2–3 times per month ☐ 1 time per day	74b. How often were the hot dogs or frankfurters you ate light or low-fat hot dogs ?
☐ 1 time per week ☐ 2 or more times per day	
72a. Each time you ate beef hamburgers or	☐ Almost never or never ☐ About ¼ of the time
cheeseburgers, how much did you usually	About 1/2 of the time
eat?	About ³ / ₄ of the time
Less than 1 patty or less than 2 ounces	☐ Almost always or always
☐ 1 patty or 2 to 4 ounces	
☐ More than 1 patty or more than 4 ounces	
72b. How often were the beef hamburgers or	
cheeseburgers you ate made with lean	
ground beef?	
☐ Almost never or never	
About 1/4 of the time	
☐ About ½ of the time ☐ About ¾ of the time	
Almost always or always	

Over the past 12 months	77b. How often was the steak you ate lean steak?
75. How often did you eat beef mixtures such as beef stew, beef pot pie, beef and noodles, or beef and vegetables?	☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time
☐ NEVER (GO TO QUESTION 76)	☐ Almost always or always
☐ 1-6 times per year ☐ 2 times per week ☐ 7-11 times per year ☐ 3-4 times per week ☐ 1 time per month ☐ 5-6 times per week ☐ 2-3 times per month ☐ 1 time per day ☐ 1 time per week ☐ 2 or more times per day 75a. Each time you ate beef stew, beef pot pie, beef and noodles, or beef and vegetables,	78. How often did you eat pork or beef spareribs ? NEVER (GO TO QUESTION 79) 1–6 times per year 2 times per week 3–4 times per week 5–6 times per week 2–3 times per month 1 time per day
how much did you usually eat? Less than 1 cup 1 to 2 cups More than 2 cups 76. How often did you eat roast beef or pot roast? (Please do not include roast beef or pot roast in	☐ 1 time per week ☐ 2 or more times per day 78a. Each time you ate pork or beef spareribs , how much did you usually eat? ☐ Less than 4 ribs ☐ 4 to 12 ribs ☐ More than 12 ribs
sandwiches.) NEVER (GO TO QUESTION 77) 1–6 times per year 2 times per week	79. How often did you eat roast turkey , turkey cutlets , or turkey nuggets (including in sandwiches)?
☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 2 or more times per day 76a. Each time you ate roast beef or pot roast (including in mixtures), how much did you	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 1 time per week ☐ 2 or more times per day
usually eat? Less than 2 ounces 2 to 5 ounces More than 5 ounces	79a. Each time you ate roast turkey , turkey cutlets , or turkey nuggets , how much did you usually eat? (Please note: 4 to 8 turkey nuggets = 3 ounces.)
77. How often did you eat steak (beef)? (<i>Do not include steak in sandwiches</i>)	Less than 2 ounces 2 to 4 ounces More than 4 ounces
NEVER (GO TO QUESTION 78) □ 1–6 times per year □ 2 times per week □ 7–11 times per year □ 3–4 times per week □ 1 time per month □ 5–6 times per week □ 2–3 times per month □ 1 time per day □ 1 time per week □ 2 or more times per day	80. How often did you eat chicken as part of salads, sandwiches, casseroles, stews, or other mixtures?
77a. Each time you ate steak (beef), how much did you usually eat? Less than 3 ounces 3 to 7 ounces More than 7 ounces	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 2 or more times per day

Over the past 12 months	82. How often did you eat baked ham or ham steak?
80a. Each time you ate chicken as part of salads, sandwiches, casseroles, stews, or other mixtures, how much did you usually eat?	☐ NEVER (GO TO QUESTION 83)
Less than ½ cup ½ to 1½ cups More than 1½ cups	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 1 time per week ☐ 2 or more times per day
81. How often did you eat baked, broiled, roasted, stewed, or fried chicken (including nuggets)? (Please do not include chicken in mixtures.)	82a. Each time you ate baked ham or ham steak , how much did you usually eat?
NEVER (GO TO QUESTION 82) 1–6 times per year	□ Less than 1 ounce □ 1 to 3 ounces □ More than 3 ounces □ S3. How often did you eat pork (including chops, roasts, and in mixed dishes)? (Please do not include ham, ham steak, or sausage.) □ NEVER (GO TO QUESTION 84) □ 1-6 times per year □ 2 times per week □ 7-11 times per year □ 3-4 times per week □ 1 time per month □ 5-6 times per week □ 2-3 times per month □ 1 time per day □ 1 time per week □ 2 or more times per day 83a. Each time you ate pork , how much did you
81b. How often was the chicken you ate fried chicken (including deep fried) or chicken nuggets ? Almost never or never About 1/4 of the time	usually eat? Less than 2 ounces or less than 1 chop 2 to 5 ounces or 1 chop More than 5 ounces or more than 1 chop 84. How often did you eat gravy on meat, chicken, potatoes, rice, etc.?
☐ About ½ of the time ☐ About ¾ of the time ☐ Almost always or always 81c. How often was the chicken you ate WHITE meat? ☐ Almost never or never ☐ About ¼ of the time	NEVER (GO TO QUESTION 85) □ 1–6 times per year □ 2 times per week □ 7–11 times per year □ 3–4 times per week □ 1 time per month □ 5–6 times per week □ 2–3 times per month □ 1 time per day □ 1 time per week □ 2 or more times per day
☐ About ½ of the time ☐ About ¾ of the time ☐ Almost always or always	84a. Each time you ate gravy on meat, chicken, potatoes, rice, etc., how much did you usually eat?
81d. How often did you eat chicken WITH skin? Almost never or never About 1/4 of the time About 1/2 of the time About 3/4 of the time Almost always or always	Less than 1/2 cup 1/2 to 1/2 cup More than 1/2 cup

Over the past 12 months	87a. Each time you ate sausage , how much did you usually eat?
85. How often did you eat liver (all kinds) or liverwurst ?	☐ Less than 1 patty or 2 links☐ 1 to 3 patties or 2 to 5 links☐ More than 3 patties or 5 links
□ 1–6 times per year □ 2 times per week □ 7–11 times per year □ 3–4 times per week □ 1 time per month □ 5–6 times per week □ 2–3 times per month □ 1 time per day □ 1 time per week □ 2 or more times per day 85a. Each time you ate liver or liverwurst , how much did you usually eat? □ Less than 1 ounce	87b. How often was the sausage you ate light, low-fat, or lean sausage? Almost never or never About ½ of the time About ½ of the time About ¾ of the time About ¾ of the time Almost always or always 88. How often did you eat fish sticks or fried fish
1 to 4 ounces More than 4 ounces	(including fried seafood or shellfish)? ☐ NEVER (GO TO QUESTION 89)
86. How often did you eat bacon (including low-fat)? NEVER (GO TO QUESTION 87) 1-6 times per year	□ 1–6 times per year □ 2 times per week □ 7–11 times per year □ 3–4 times per week □ 1 time per month □ 5–6 times per week □ 2–3 times per month □ 1 time per day □ 1 time per week □ 2 or more times per day 88a. Each time you ate fish sticks or fried fish , how much did you usually eat? □ Less than 2 ounces or less than 1 fillet □ 2 to 7 ounces or 1 fillet □ More than 7 ounces or more than 1 fillet □ More than 7 ounces or more than 1 fillet □ NEVER (GO TO INTRODUCTION TO QUESTION 90) □ 1–6 times per year □ 2 times per week □ 7–11 times per year □ 3–4 times per week □ 1 time per month □ 5–6 times per week □ 2–3 times per month □ 1 time per day □ 1 time per week □ 2 or more times per day
87. How often did you eat sausage (including lowfat)?	89a. Each time you ate eat fish or seafood that was NOT FRIED , how much did you usually eat?
NEVER (GO TO QUESTION 88) 1–6 times per year	Less than 2 ounces or less than 1 fillet 2 to 5 ounces or 1 fillet More than 5 ounces or more than 1 fillet

Over the past 12 months	92. Over the past 12 months, did you eat soups?
Now think about all the meat, poultry, and fish you ate in the <u>past 12 months</u> and how they were prepared.	☐ NO (GO TO QUESTION 93) ☐ YES
90. How often was oil, butter, margarine, or other fat used to FRY, SAUTE, BASTE, OR MARINATE any meat, poultry, or fish you ate? (Please do not include deep frying.) NEVER (GO TO QUESTION 91) 1–6 times per year	92a. How often did you eat soup DURING THE WINTER? NEVER 1-6 times per winter 2 times per week 7-11 times per winter 3-4 times per week 1 time per month 5-6 times per week 2-3 times per month 1 time per day 1 time per week 2 or more times per day
☐ 1 time per week ☐ 2 or more times per day 90a. Which of the following fats were regularly used to prepare your meat, poultry, or fish? (Mark all that apply.)	92b. How often did you eat soup DURING THE REST OF THE YEAR?
☐ Margarine (including low-fat) ☐ Corn oil ☐ Dow-fat) ☐ Canola or rapeseed oil ☐ Butter (including low-fat) ☐ Oil spray, such as Pam or others ☐ Lard, fatback, or bacon fat ☐ Other kinds of oils ☐ None of the above	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 2 or more times per day
91. How often did you eat tofu , soy burgers , or soy meat-substitutes?	92c. Each time you ate soup , how much did you usually eat?
☐ NEVER (GO TO QUESTION 92)	☐ Less than 1 cup ☐ 1 to 2 cups ☐ More than 2 cups
☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 1 time per week ☐ 2 or more times per day 91a. Each time you ate tofu, soy burgers, or soy meat-substitutes, how much did you usually eat?	92d. How often were the soups you ate bean soups? Almost never or never About ¼ of the time About ½ of the time About ¾ of the time Almost always or always
Less than ¼ cup or less than 2 ounces ¼ to ½ cup or 2 to 4 ounces More than ½ cup or more than 4 ounces	92e. How often were the soups you ate cream soups (including chowders)? Almost never or never About 1/4 of the time About 1/2 of the time About 3/4 of the time About 3/4 of the time Almost always or always

Over the past 12 months	you usually eat?
92f. How often were the soups you ate tomato or vegetable soups ?	Fewer than 4 crackers ☐ 4 to 10 crackers ☐ More than 10 crackers
☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ Almost always or always	95. How often did you eat corn bread or corn muffins ?
92g. How often were the soups you ate broth soups (including chicken) with or without noodles or rice? Almost never or never About ¼ of the time About ½ of the time About ¾ of the time About ¾ of the time About ¾ of the time	NEVER (GO TO QUESTION 96) 1–6 times per year 2 times per week 7–11 times per year 3–4 times per week 1 time per month 5–6 times per week 2–3 times per month 1 time per day 1 time per week 2 or more times per day 95a. Each time you ate corn bread or corn muffins, how much did you usually eat?
93. How often did you eat pizza ? —	☐ Less than 1 piece or muffin ☐ 1 to 2 pieces or muffins ☐ More than 2 pieces or muffins
1-6 times per year 2 times per week 7-11 times per year 3-4 times per week 1 time per month 5-6 times per week 2-3 times per month 1 time per day 2 or more times per day 3a. Each time you ate pizza, how much did you usually eat? Less than 1 slice or less than 1 mini pizza 1 to 3 slices or 1 mini pizza More than 3 slices or more than 1 mini pizza More than 3 slices or more than 1 mini pizza 3b. How often did you eat pizza with pepperoni, sausage, or other meat? Almost never or never About ½ of the time About ½ of the time Almost always or always	96. How often did you eat biscuits? NEVER (GO TO QUESTION 97) 1–6 times per year 2 times per week 7–11 times per year 3–4 times per week 1 time per month 5–6 times per week 2–3 times per month 1 time per day 1 time per week 2 or more times per day 96a. Each time you ate biscuits, how many did you usually eat? Fewer than 1 biscuit 1 to 2 biscuits More than 2 biscuits More than 2 biscuits 97. How often did you eat potato chips, tortilla chips, or corn chips (including low-fat, fat-free, or low-salt)?
94. How often did you eat crackers ? NEVER (GO TO QUESTION 95) 1–6 times per year	NEVER (GO TO QUESTION 98) 1–6 times per year

Over the past 12 months	99a. Each time you ate pretzels , how many did you usually eat?			
97a. Each time you ate potato chips, tortilla chips, or corn chips , how much did you usually eat?	Fewer than 5 average twists 5 to 20 average twists More than 20 average twists			
☐ Fewer than 10 chips or less than 1 cup☐ 10 to 25 chips or 1 to 2 cups☐ More than 25 chips or more than 2 cups	100. How often did you eat peanuts , walnuts , seeds , or other nuts ?			
97b. How often were the chips you ate Wow chips or other chips made with fat substitute (Olean or Olestra)? Almost never or never About ¼ of the time About ¾ of the time About ¾ of the time Almost always or always 97c. How often were the chips you ate other lowfat or fat-free chips? Almost never or never About ¼ of the time About ¼ of the time About ¾ of the time Almost always or always	NEVER (GO TO QUESTION 101) 1–6 times per year			
fat)? NEVER (GO TO QUESTION 99) 1–6 times per year 2 times per week 7–11 times per year 3–4 times per week 2–3 times per month 1 time per day 1 time per week 2 or more times per day 98a. Each time you ate popcorn, how much did you usually eat? Less than 2 cups, popped 2 to 5 cups, popped More than 5 cups, popped More than 5 cups, popped 1–6 times per year 2 times per week 7–11 times per year 3–4 times per week 1 time per month 5–6 times per week 2–3 times per month 1 time per day 1 time per week 2 or more times per day 3 or more times 3 or m	1–6 times per year 2 times per week 7–11 times per year 3–4 times per week 1 time per month 5–6 times per week 2–3 times per month 1 time per day 2 or more times per day 101a. Each time you ate energy, high-protein, or breakfast bars, how much did you usually eat? Less than 1 bar 1 bar More than 1 bar More than 1 bar 102. How often did you eat yogurt (NOT including frozen yogurt)? NEVER (GO TO QUESTION 103) 1–6 times per year 2 times per week 7–11 times per year 3–4 times per week 1 time per month 5–6 times per week 2–3 times per month 1 time per day 1 time per week 2 or more times per day 1 time per week 2 or more times per day			

Over the past 12 months	104c. How often was the cheese you ate fat-free cheese ?		
102a. Each time you ate yogur t, how much did you usually eat? Less than ½ cup or less than 1 container ½ to 1 cup or 1 container More than 1 cup or more than 1 container	☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ About ¾ of the time ☐ Almost always or always		
½ to 1 cup or 1 container More than 1 cup or more than 1 container 103. How often did you eat cottage cheese (including low-fat)?	Almost always or always 105. How often did you eat frozen yogurt, sorbet, or ices (including low-fat or fat-free)? NEVER (GO TO QUESTION 106) 1-6 times per year 2 times per week 7-11 times per year 3-4 times per week 1 time per month 5-6 times per week 2-3 times per month 1 time per day 1 time per week 2 or more times per day 105a. Each time you ate frozen yogurt, sorbet, or ices, how much did you usually eat? Less than ½ cup or less than 1 scoop ½ to 1 cup or 1 to 2 scoops More than 1 cup or more than 2 scoops More than 1 cup or more than 2 scoops 1-6 times per year 2 times per week 7-11 times per year 3-4 times per week 1 time per month 5-6 times per week 1 time per month 2 times per week 2 and times per week 2 an		
☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ About ¾ of the time ☐ Almost always or always	low-fat, or fat-free ice cream or sherbet? ☐ Almost never or never ☐ About ½ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ Almost always or always		

Over the past 12 months	109. How often did you eat doughnuts, sweet rolls, Danish, or pop-tarts?		
107. How often did you eat cake (including low-fat or fat-free)?	☐ NEVER (GO TO QUESTION 110)		
NEVER (GO TO QUESTION 108) □ 1–6 times per year □ 2 times per week □ 7–11 times per year □ 3–4 times per week □ 1 time per month □ 5–6 times per week □ 2–3 times per month □ 1 time per day □ 1 time per week □ 2 or more times per day	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 1 time per week ☐ 2 or more times per day 109a. Each time you ate doughnuts, sweet rolls, Danish, or pop-tarts, how much did you		
107a. Each time you ate cake, how much did you usually eat? Less than 1 medium piece 1 medium piece More than 1 medium piece 107b. How often was the cake you ate light, lowfat, or fat-free cake?	usually eat? Less than 1 piece 1 to 2 pieces More than 2 pieces 110. How often did you eat sweet muffins or dessert breads (including low-fat or fat-free)?		
☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ Almost always or always	NEVER (GO TO QUESTION 111) 1–6 times per year		
108. How often did you eat cookies or brownies (including low-fat or fat-free)?	110a. Each time you ate sweet muffins or dessert breads , how much did you usually eat?		
NEVER (GO TO QUESTION 109) 1–6 times per year	☐ Less than 1 medium piece ☐ 1 medium piece ☐ More than 1 medium piece		
☐ 1 time per week ☐ 2 or more times per day 108a. Each time you ate cookies or brownies , how much did you usually eat? ☐ Less than 2 cookies or 1 small brownie ☐ 2 to 4 cookies or 1 medium brownie ☐ More than 4 cookies or 1 large brownie	110b. How often were the sweet muffins or dessert breads you ate light, low-fat, or fat-free sweet muffins or dessert breads? Almost never or never About 1/4 of the time About 1/2 of the time About 3/4 of the time Almost always or always		
108b. How often were the cookies or brownies you ate light, low-fat, or fat-free cookies or brownies?	111. How often did you eat fruit crisp, cobbler, or strudel?☐ NEVER (GO TO QUESTION 112)		
☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ Almost always or always	☐ 1–6 times per year ☐ 2 times per week ☐ 7–11 times per year ☐ 3–4 times per week ☐ 1 time per month ☐ 5–6 times per week ☐ 2–3 times per month ☐ 1 time per day ☐ 2 or more times per day		

Over the past 12 months	112e. How often were the pies you ate pecan p				
111a. Each time you ate fruit crisp, cobbler , or strudel , how much did you usually eat? ☐ Less than ½ cup ☐ ½ to 1 cup ☐ More than 1 cup	☐ Almost never or never ☐ About 1/4 of the time ☐ About 1/2 of the time ☐ About 3/4 of the time ☐ Almost always or always				
112. How often did you eat pie ?	113. How often did you eat chocolate candy?				
NEVER (GO TO QUESTION 113) 1–6 times per year	□ NEVER (GO TO QUESTION 114) □ 1–6 times per year □ 2 times per week □ 7–11 times per year □ 3–4 times per week □ 1 time per month □ 5–6 times per week □ 2–3 times per month □ 1 time per day □ 1 time per week □ 2 or more times per day 113a. Each time you ate chocolate candy , how much did you usually eat? □ Less than 1 average bar or less than 1 ounce □ 1 average bar or 1 to 2 ounces □ More than 1 average bar or more than 2 ounces □ 114. How often did you eat other candy ?				
The next four questions ask about the kinds of pie you ate. Please read all four questions before answering. 112b. How often were the pies you ate fruit pie (such as apple, blueberry, others)? Almost never or never About ½ of the time About ¾ of the time About ¾ of the time Almost always or always 112c. How often were the pies you ate cream, pudding, custard, or meringue pie? Almost never or never About ¼ of the time About ½ of the time About ¾ of the time About ¾ of the time About ¾ of the time About ¼ of the time About ¾ of the time	NEVER (GO TO QUESTION 115) 1–6 times per year				

Over the past 12 months	116. How many cups of coffee , caffeinated or decaffeinated, did you drink?
115a. Each time you ate eggs , how many did you usually eat?	☐ NEVER (GO TO QUESTION 117)
☐ 1 egg ☐ 2 eggs ☐ 3 or more eggs 115b. How often were the eggs you ate egg substitutes? ☐ Almost never or never	Less than 1 cup per
☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ Almost always or always	☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ Almost always or always
115c. How often were the eggs you ate egg	☐ Aimost always of always
whites only? Almost never or never	117. How many glasses of ICED tea, caffeinated or decaffeinated, did you drink?
☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time	☐ NEVER (GO TO QUESTION 118)
☐ Almost always or always 115d. How often were the eggs you ate regular	Less than 1 cup per 5-6 cups per week month 1 cup per day 1-3 cups per day 2-3 cups per day
whole eggs?	☐ 1 cup per week ☐ 4–5 cups per day ☐ 2–4 cups per week ☐ 6 or more cups per day
☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ Almost always or always	117a. How often was the iced tea you drank decaffeinated or herbal tea? ☐ Almost never or never ☐ About 1/4 of the time ☐ About 1/2 of the time
115e. How often were the eggs you ate cooked in oil, butter, or margarine?	☐ About ¾ of the time ☐ Almost always or always
☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ Almost always or always	118. How many cups of HOT tea , caffeinated or decaffeinated, did you drink? NEVER (GO TO QUESTION 119) Less than 1 cup per 5–6 cups per week
115f. How often were the eggs you ate part of egg salad? ☐ Almost never or never	month
☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ About ¾ of the time ☐ Almost always or always	118a. How often was the hot tea you drank decaffeinated or herbal tea?
	☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ Almost always or always

Over the past 12 months	usually use?		
119. How often did you add sugar or honey to your coffee or tea?	Regular powdered Low-fat or fat-free powdered		
☐ NEVER (GO TO QUESTION 120)	Regular liquid Low-fat or fat-free liquid		
□ Less than 1 time per month □ 5–6 times per week □ 1–3 times per month □ 2–3 times per day □ 1 time per week □ 4–5 times per day □ 2–4 times per week □ 6 or more times per day	122. How often was cream or half and half added to your coffee or tea?		
119a. Each time sugar or honey was added to your coffee or tea, how much was usually added? Less than 1 teaspoon 1 to 3 teaspoons	□ Less than 1 time per month □ 1 time per day □ 1–3 times per month □ 2–3 times per day □ 1 time per week □ 4–5 times per day □ 2–4 times per week □ 6 or more times per day		
▼ ☐ More than 3 teaspoons	122a. Each time cream or half and half was		
120. How often did you add artificial sweetener to	added to your coffee or tea, how much was usually added?		
your coffee or tea? NEVER (GO TO QUESTION 121)	☐ Less than 1 tablespoon ☐ 1 to 2 tablespoons ☐ More than 2 tablespoons		
☐ Less than 1 time per ☐ 5–6 times per week month ☐ 1 time per day ☐ 1–3 times per month ☐ 2–3 times per day ☐ 1 time per week ☐ 4–5 times per day	▼ 123. How often was milk added to your coffee or tea?		
☐ 2–4 times per week ☐ 6 or more times per day	☐ NEVER (GO TO QUESTION 124)		
120a. What kind of artificial sweetener did you usually use? ☐ Equal or aspartame ☐ Sweet N Low or saccharin	☐ Less than 1 time per ☐ 5–6 times per week month ☐ 1 time per day ☐ 2–3 times per day ☐ 1 time per week ☐ 4–5 times per day ☐ 2–4 times per week ☐ 6 or more times per day		
→ —	123a. Each time milk was added to your coffee or		
121. How often was non-dairy creamer added to your coffee or tea?	tea, how much was usually added?		
□ NEVER (GO TO QUESTION 122) □ Less than 1 time per □ 5–6 times per week month □ 1 time per day □ 1–3 times per month □ 2–3 times per day	Less than 1 tablespoon 1 to 3 tablespoons More than 3 tablespoons 123b. What kind of milk was usually added to your		
☐ 1 time per week ☐ 4—5 times per day ☐ 2—4 times per week ☐ 6 or more times per day	coffee or tea?		
121a. Each time non-dairy creamer was added to your coffee or tea, how much was usually used? Less than 1 teaspoon 1 to 3 teaspoons More than 3 teaspoons			

Over the past 12 months	125c. How often was the margarine you ate fat- free margarine ?			
124. How often was sugar or honey added to foods you ate? (Please do not include sugar in coffee, tea, other beverages, or baked goods.) NEVER (GO TO INTRODUCTION TO	☐ Almost never or never ☐ About ¼ of the time ☐ About ½ of the time ☐ About ¾ of the time ☐ Almost always or always			
1-6 times per year 2 times per week 7-11 times per year 3-4 times per week 7-11 times per year 3-4 times per week 1 time per month 5-6 times per week 2-3 times per month 1 time per day 1 time per day 1 time per week 2 or more times per day 124a. Each time sugar or honey was added to foods you ate, how much was usually added? Less than 1 teaspoon 1 to 3 teaspoons More than 3 teaspoons More than 3 teaspoons dessenting that you eat. If possible, please check the labels of these foods to help you answer. 125. Over the past 12 months, did you eat margarine? NO (GO TO QUESTION 126) YES 125a. How often was the margarine you ate regular-fat margarine (stick or tub)? Almost never or never About ½ of the time About 3/4 of the time Almost always or always 125b. How often was the margarine you ate light or low-fat margarine (stick or tub)? Almost never or never About ½ of the time About ¾ of the time Almost always or always Almost always or always	126. Over the past 12 months, did you eat butter? NO (GO TO QUESTION 127) YES 126a. How often was the butter you ate light or low-fat butter? Almost never or never About ½ of the time About ½ of the time About ¾ of the time Almost always or always 127. Over the past 12 months, did you eat mayonnaise or mayonnaise-type dressing? NO (GO TO QUESTION 128) YES 127a. How often was the mayonnaise you ate regular-fat mayonnaise? Almost never or never About ½ of the time About ½ of the time Almost always or always 127b. How often was the mayonnaise you ate light or low-fat mayonnaise? Almost never or never About ¼ of the time About ¾ of the time About ¾ of the time Almost always or always			
	· I			

Over the past 12 months	129b. How often was the cream cheese you ate light, low-fat, or fat-free cream cheese?			
127c. How often was the mayonnaise you ate fat-	iigitt, iow-iat, or iat-free cream cheese:			
free mayonnaise?	☐ Almost never or never ☐ About ¼ of the time			
☐ Almost never or never	About ½ of the time			
Almost never of never	About ¾ of the time			
About ½ of the time	Almost always or always			
About ¾ of the time	120 Over the neet 12 months, did you get saled			
☐ Almost always or always	130. Over the <u>past 12 months</u> , did you eat salad dressing?			
128. Over the past 12 months, did you eat sour	•			
cream?	NO (GO TO INTRODUCTION TO QUESTION 131)			
☐ NO (GO TO QUESTION 129)				
	130a. How often was the salad dressing you ate			
	regular-fat salad dressing (including oil			
128a. How often was the sour cream you ate	and vinegar dressing)?			
regular-fat sour cream?				
	☐ Almost never or never			
☐ Almost never or never	About ¼ of the time			
About 1/4 of the time	About ½ of the time			
About ½ of the time	About ¾ of the time			
About % of the time	Almost always or always			
☐ Almost always or always	130b. How often was the salad dressing you ate			
100h Haw often was the saw aream you at light				
128b. How often was the sour cream you ate light ,	light or low-fat salad dressing?			
low-fat, or fat-free sour cream?				
	☐ Almost never or never			
☐ Almost never or never	☐ About ¼ of the time			
About ¼ of the time	About ½ of the time			
About ½ of the time	☐ About ¾ of the time			
About ¾ of the time	☐ Almost always or always			
☐ Almost always or always				
▼	130c. How often was the salad dressing you ate			
129. Over the past 12 months, did you eat cream	fat-free salad dressing?			
cheese?	latilite salad diessing:			
cheese ?				
— —	☐ Almost never or never			
☐ NO (GO TO QUESTION 130)	About ¼ of the time			
	About ½ of the time			
	☐ About ¾ of the time			
♥				
129a. How often was the cream cheese you ate	The following two questions ask you to			
regular-fat cream cheese?	summarize your usual intake of vegetables and			
J	fruits. Please do not include salads, potatoes, or			
☐ Almost never or never	juices.			
About ¼ of the time	Janese.			
About ½ of the time	131 Over the nact 12 months, how many convince of			
About ½ of the time	131. Over the past 12 months, how many servings of			
☐ About 74 of the time	vegetables (not including salad or potatoes) did			
☐ All Host always of always	you eat per week or per day?			
	Less than 1 per week 2 per day			
	☐ 1–2 per week ☐ 3 per day			
	3–4 per week 4 per day			
	☐ 5–6 per week ☐ 5 or more per day			
	☐ 1 per day			

Over the past 12 months	The next questions are about your use of fiber supplements or vitamin pills.
132. Over the past 12 months, how many servings of fruit (not including juices) did you eat per week or per day? Less than 1 per week	135. Over the past 12 months, did you take any of the following types of fiber or fiber supplements on a regular basis (more than once per week for at least 6 of the last 12 months)? (Mark all that apply.) NO, didn't take any fiber supplements on a regular basis (GO TO QUESTION 136)
133. Over the past month, which of the following foods did you eat AT LEAST THREE TIMES? (Mark all that apply.) Avocado, guacamole Olives Cheesecake Oysters Chocolate, fudge, or butterscotch toppings or syrups Plantains Chow mein noodles Pork neckbones, hock, head, feet Dried apricots Pudding or custard Egg rolls Veal, venison, lamb Granola bars Whipped cream, regular Hot peppers Whipped cream, substitute Milkshakes or ice-cream sodas NONE	□ YES, psyllium products (such as Metamucil, Fiberall, Serutan, Perdiem, Correctol) □ YES, methylcellulose/cellulose products (such as Citrucel, Unifiber) □ YES, Fibercon □ YES, Bran (such as wheat bran, oat bran, or bran wafers) 136. Over the past 12 months, did you take any multivitamins, such as One-a-Day-, Theragran-, or Centrum-type multivitamins (as pills, liquids, or packets)? □ NO (GO TO INTRODUCTION TO QUESTION 138) □ YES
134. For ALL of the <u>past 12 months</u> , have you followed any type of vegetarian diet ?	137. How often did you take One-a-day-, Theragran-, or Centrum-type multivitamins?
NO (GO TO INTRODUCTION TO QUESTION 135) ☐ YES	Less than 1 day per month 1–3 days per month 1–3 days per week 4–6 days per week Every day
134a. Which of the following foods did you TOTALLY EXCLUDE from your diet? (Mark all that apply.) Meat (beef, pork, lamb, etc.) Poultry (chicken, turkey, duck) Fish and seafood Eggs Dairy products (milk, cheese, etc.)	137a. Does your multivitamin usually contain minerals (such as iron, zinc, etc.)? NO YES Don't know 137b. For how many years have you taken multivitamins? Less than 1 year 1-4 years
	5–9 years 10 or more years

Over the past 12 months	139. How often did you take Vitamin A (NOT as part of a multivitamin in Question 137)?
137c. Over the past 12 months, did you take any	,
vitamins, minerals, or other herbal	☐ NEVER (GO TO QUESTION 140)
supplements other than your multivitamin?	Less than 1 day per month
	1–3 days per month
□NO	1–3 days per week
<u> </u>	4–6 days per week
Thank you <i>very much</i> for completing this	☐ Every day
questionnaire! Because we want to be able to	120a When you took Vitamin A shout how much
use all the information you have provided, we	139a. When you took Vitamin A , about how much did you take in one day?
would greatly appreciate it if you would please	and you take in one day:
take a moment to review each page making sure	Less than 8,000 IU
that you:	□ 8,000–9,999 IU
Did not alsin any name and	10,000–14,999 IU
 Did not skip any pages and Crossed out the incorrect answer and circled 	☐ 15,000–24,999 IU ☐ 25,000 IU or more
the correct answer if you made any changes.	Don't know
the correct answer if you made any changes.	
YES (GO TO INTRODUCTION TO	139b. For how many years have you taken Vitamin A?
QUESTION 138)	Vitaliii A:
These last questions are about the vitamins,	Less than 1 year
minerals, or herbal supplements you took that are	☐ 1–4 years ☐ 5–9 years
NOT part of a One-a-day-, Theragran-, or	10 or more years
Centrum-type of multivitamin.	▼ □ No de mino yeans
Please include vitamins taken as part of an antioxidant supplement.	140. How often did you take Vitamin C (NOT as part of a multivitamin in Question 137)?
antioxidant supplement.	,
138. How often did you take Beta-carotene (NOT as	☐ NEVER (GO TO QUESTION 141)
part of a multivitamin in Question 137)?	Less than 1 day per month
	1–3 days per month
☐ NEVER (GO TO QUESTION 139)	☐ 1–3 days per week
Less than 1 day per month	4–6 days per week
1–3 days per month	☐ Every day
☐ 1–3 days per week	140a. When you took Vitamin C , about how much
4–6 days per week	did you take in one day?
☐ Every day	
138a. When you took Beta-carotene , about how	Less than 500 mg
much did you take in one day?	□ 500–999 mg
_	☐ 1,000–1,499 mg ☐ 1,500–1,999 mg
Less than 10,000 IU	2,000 mg or more
☐ 10,000–14,999 IU ☐ 15,000–19,999 IU	☐ Don't know
☐ 20,000—24,999 IU	
☐ 25,000 IU or more	140b. For how many years have you taken
☐ Don't know	Vitamin C?
138b. For how many years have you taken Beta-	Less than 1 year
carotene?	☐ 1–4 years
our otorio:	☐ 5–9 years
Less than 1 year	☐ 10 or more years
☐ 1–4 years	
5–9 years	
☐ 10 or more years	¹ ↓

Over the past 12 months	142b. For how many years have you taken Calcium or Calcium-containing antacids?			
141. How often did you take Vitamin E (NOT as part	Calcium of Calcium	il-containing antacids?		
of a multivitamin in Question 137)?	☐ Less than 1 year			
ora malaviami in Quodion 107).	☐ 1–4 years			
☐ NEVER (GO TO QUESTION 142)	☐ 5–9 years ☐ 10 or more years			
Less than 1 day per month	,			
1–3 days per month	The last two questions ask	you about other		
☐ 1–3 days per week	supplements you took more			
4–6 days per week				
☐ Every day	143. Please mark any of the f	ollowina sinale		
	supplements you took r			
141a. When you took Vitamin E , about how much	week (NOT as part of a			
did you take in one day?	137):			
Less than 400 IU	☐ B-6	☐ Folic acid/folate		
☐ 400–799 IU	☐ B-complex	Glucosamine		
☐ 800–999 IU	☐ Brewer's yeast	☐ Hydroxytryptophan (HTP)		
1,000 IU or more	Cod liver oil			
☐ Don't know	☐ Coenzyme Q	□ Niacin		
	Fish oil	☐ Selenium		
141b. For how many years have you taken Vitamin E?	(Omega-3 fatty acids)	Zinc		
	144. Please mark any of the f	ollowing herbal or		
Less than 1 year	botanical supplements			
1–4 years	once per week.	<u> </u>		
☐ 5–9 years	<u> </u>			
	☐ Aloe Vera	Ginger		
140 Have after did very take Calairuse on Calairuse	☐ Astragalus	☐ Ginkgo biloba		
142. How often did you take Calcium or Calcium-	Bilberry	Ginseng (American or		
containing antacids (NOT as part of a	☐ Cascara sagrada	Asian)		
multivitamin in Question 137)?	☐ Cat's claw	☐ Goldenseal		
	Cayenne	Grapeseed extract		
☐ NEVER (GO TO QUESTION 143)	Cranberry	☐ Kava, kava		
Less than 1 day per month	☐ Dong Kuai (Tangkwei)	☐ Milk thistle		
1–3 days per month	☐ Echinacea	☐ Saw palmetto☐ Siberian ginseng		
1 –3 days per week	☐ Evening primrose oil ☐ Feverfew	St. John's wort		
4–6 days per week	Garlic	☐ Valerian		
Every day	Garno	Other		
		_		
142a. When you took Calcium or Calcium-				
containing antacids, about how much	Thank you <u>very much</u> for con			
elemental calcium did you take in one day?	questionnaire! Because we v			
(If possible, please check the label for	all the information you have p			
elemental calcium.)	greatly appreciate it if you wo			
_	moment to review each page	making sure that you:		
Less than 500 mg				
☐ 500–599 mg	Did not skip any pages	and		
☐ 600–999 mg	Crossed out the incorre	ect answer and circled the		
1,000 mg or more	correct answer if you m	ade any changes.		
☐ Don't know				

APPENDIX J. WEEKLY SYMPTOM, TOXICITY AND ECOG ASSESSMENT FORM

Enrollment ID:

NCI Symptom and Toxicity Evaluation

Date:

Criteria	Pt	Grade	Grade 1	Grade 2	Grade 3	Grade 4
	response	0				
Weight		None	5-<10% of baseline,	10-<20% of baseline;	> 20% of baseline; TF or	
loss		(<5%)	intervention not	nutrition support	TPN indicated	
			indicated	indicated		
Nausea		None	Able to eat	Intake significantly	No significant intake	
			reasonable intake	decreased but can eat		
Vomiting		None	1 episode in 24	2-5 episodes in 24	\geq 6 episodes in 24 hours,	Life threatening
			hours over	hours; IV fluids	IV fluids or TPN indicated	consequences
			pretreatment	indicated < 24 hrs	> 24 hrs	
Heartburn		None	Mild	Moderate	Severe	-
Urinary		None	Increase in	Increase in frequency	frequency of urination	
frequency/			frequency or	or nocturia more	hourly or with more	
urgency			nocturia upto 2 x	than 2 x normal but	urgency, requiring catheter	
			normal	less frequent than		
				every hr		
Diarrhea		None	Increase of < 4	Increase of 4 – 6	Increase of ≥ 7 stools/day	Physiologic
(patients			stools/day over pre-	stools/day or	or incontinence; or need	consequences
without			treatment	nocturnal stools	for parenteral support for	requiring intensive
colostomy					dehydration	care; or hemodynamic
)						collapse
Proctitis			increased stool	increased stool	increased stool	perforation, bleeding
			frequency,	frequency, bleeding	frequency/diarrhea	or necrosis or other
			occasional blood	mucus discharge or	requiring parenteral	life threatening
		None	streaked stools or	rectal discomfort	support; rectal bleeding	complication
		None	rectal discomfort	requiring medication;	requiring transfusions or	requiring surgical
			(including	anal fissure	persistent mucus discharge,	intervention (eg
			hemorrhoids not		necessitating pads	colostomy)
			requiring meds			

Grade	Patient response	ECOG Criteria
0	ТООРОПОС	Fully active, able to carry on all pre-disease performance without restriction
1		Restricted in physically strenuous activity but ambulatory and able to carry out work of a light or sedentary nature, e.g., light house work, office work
2		Ambulatory and capable of all selfcare but unable to carry out any work activities. Up and about more than 50% of waking hours
3		Capable of only limited selfcare, confined to bed or chair more than 50% of waking hours
4		Completely disabled. Cannot carry on any selfcare. Totally confined to bed or chair

APPENDIX K. EXIT INTERVIEW FORM

Tomato Juice study participant evaluation

	apply).
	Liked the idea of tomato juice helping with radiation therapy side effects
	Wanted to participate in a research study
	Wanted to help student research
	Other (please specify)
respo	ou glad you participated in this study? (Please select only one nse). Yes Maybe, not sure No (please elaborate why not)
would	I on your experience participating in this study, what changes (if any) I you recommend the researchers make to improve participation in this for other men with prostate cancer undergoing radiation therapy?
	No changes
	Recommend the following changes
	a
	b
	c.

Thank you for your valuable participation in this research study.

APPENDIX L. SUPPLEMENTAL FIGURES



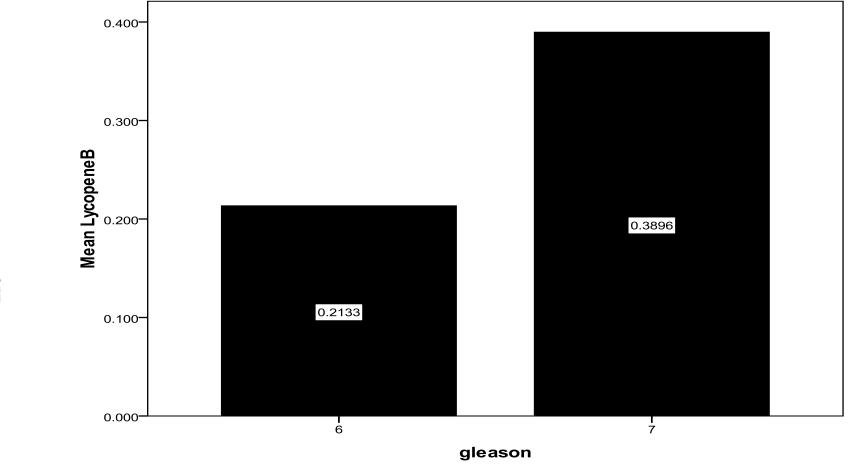


Figure 1. Serum lycopene levels at baseline among study [participants based on Gleason sum.

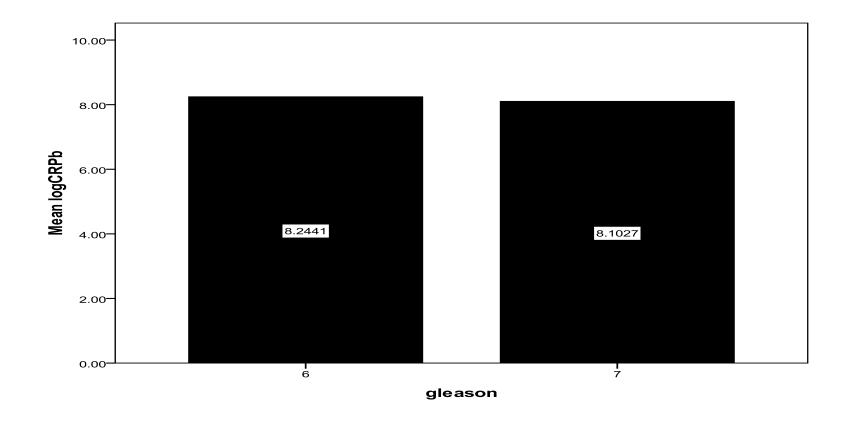


Figure 2. Serum log c-reactive protein levels at baseline based on Gleason Sum

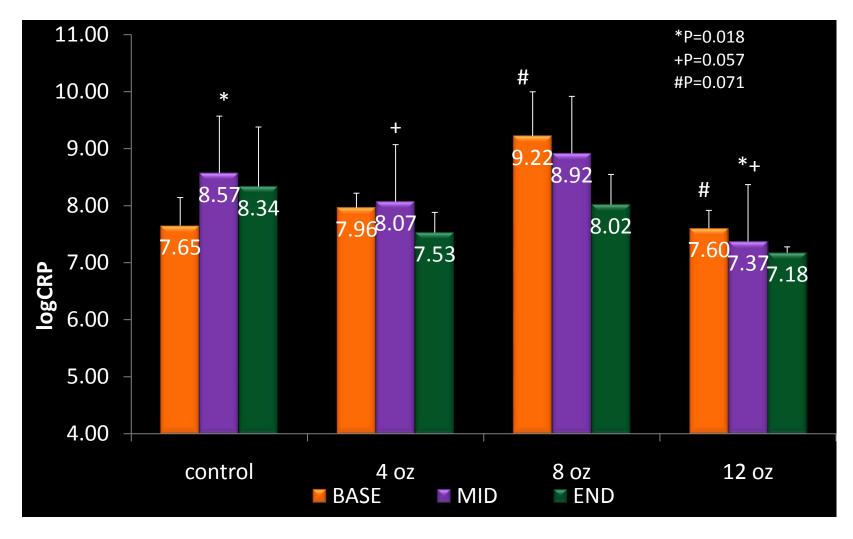


Figure 3. Serum log c-reactive protein levels among study groups at three time points.

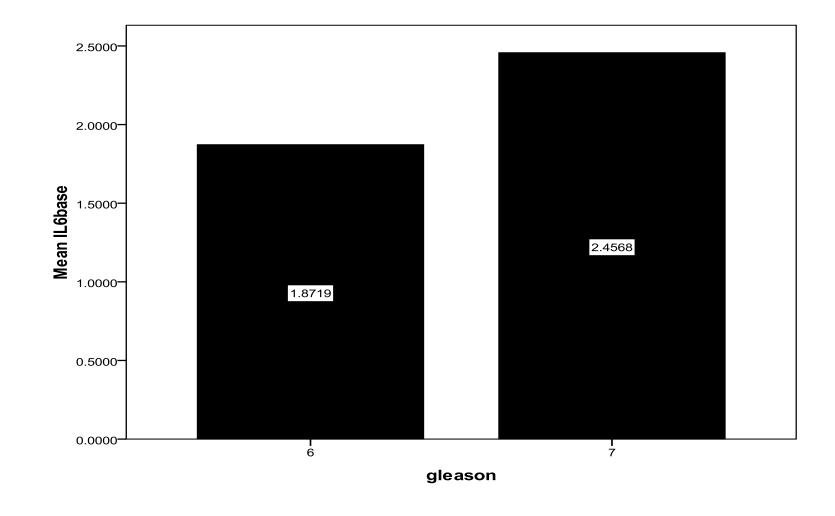


Figure 4. Serum interleukin-6 levels at baseline based on Gleason Sum

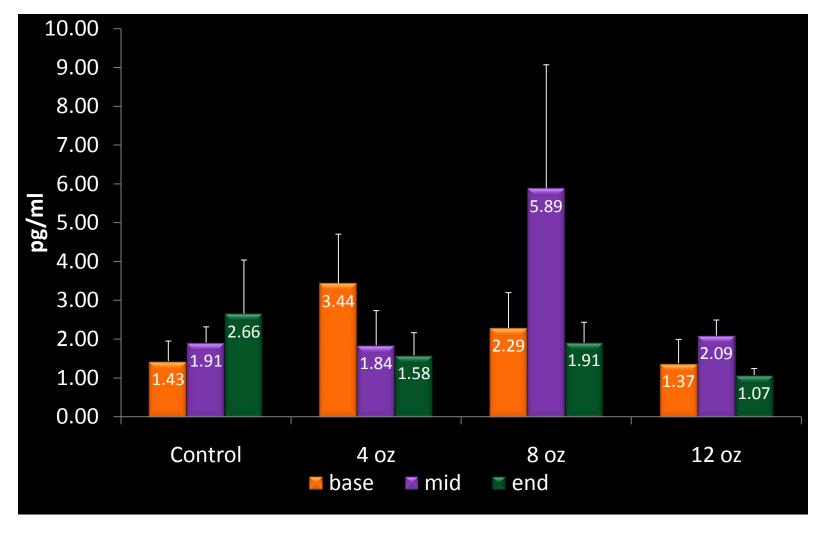


Figure 5. Serum Interleukin-6 levels among study groups at three time points.

Figure 6. Serum prostaglandin E2 levels at baseline based on Gleason Sum

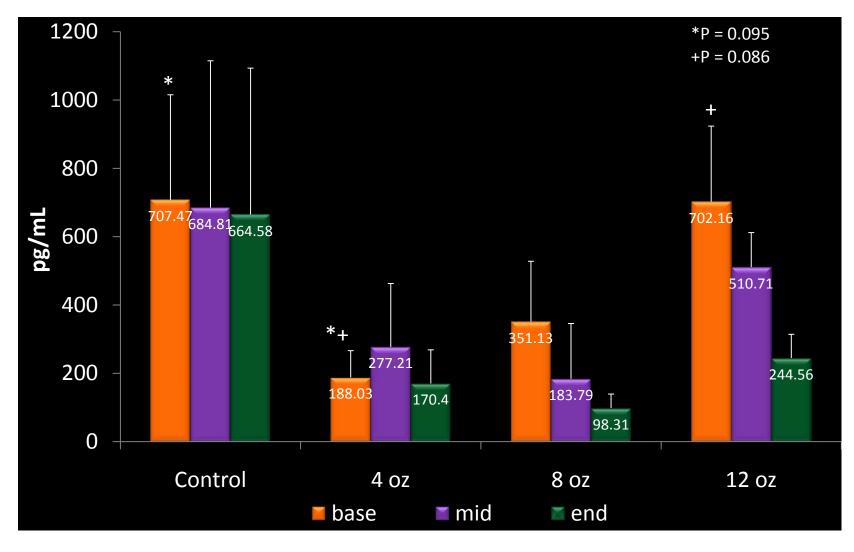


Figure 7. Serum prostaglandin E2 levels among study groups at three time points.

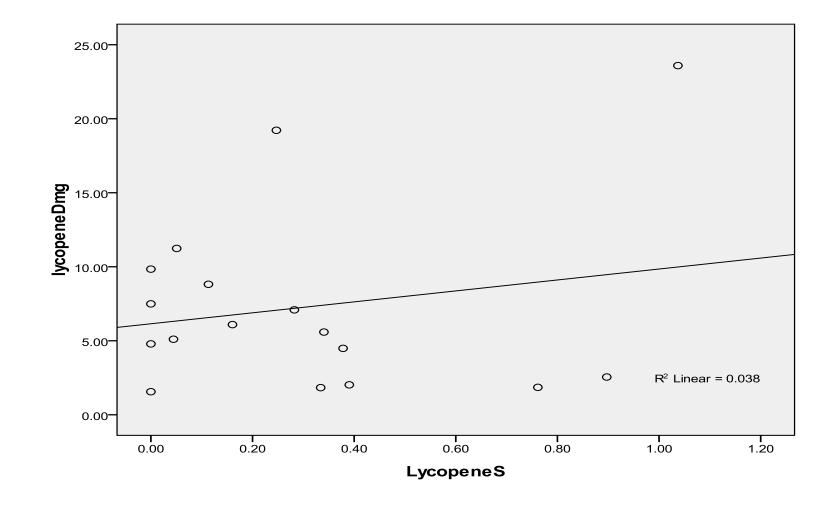


Figure 8. Scatter plot of serum lycopene plotted against total lycopene intake

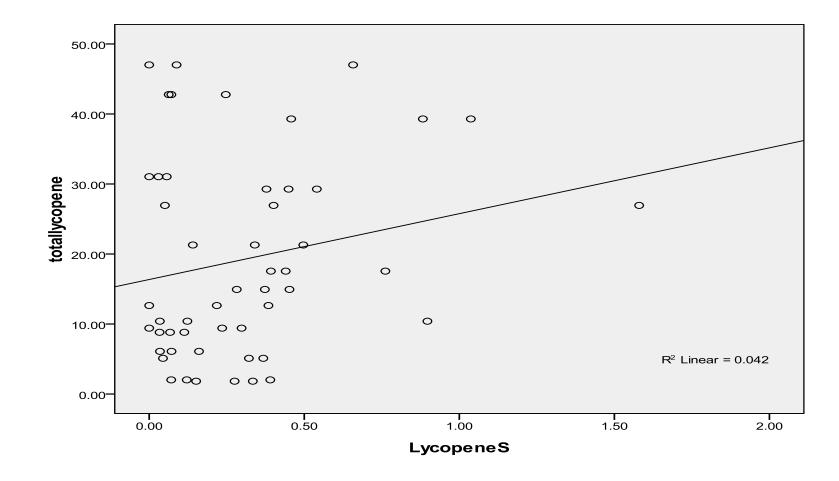


Figure 9. Scatter plot of serum lycopene plotted against total lycopene intake

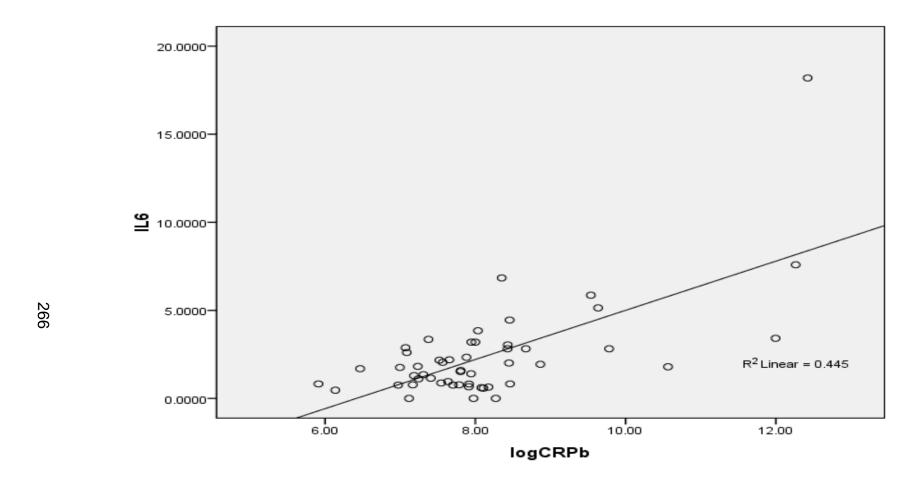


Figure 10. Scatter plot of serum CRP log transformed plotted against serum interleukin-6

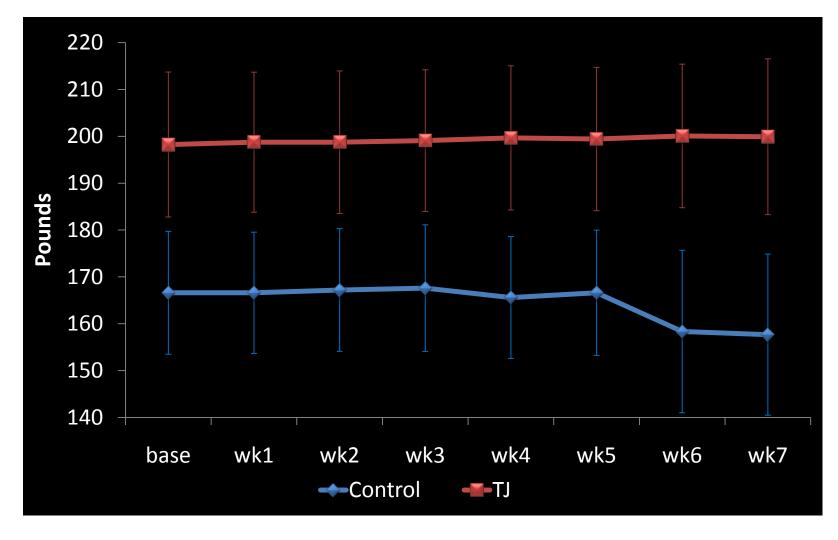


Figure 11. Weight change during treatment in control and all treatment groups

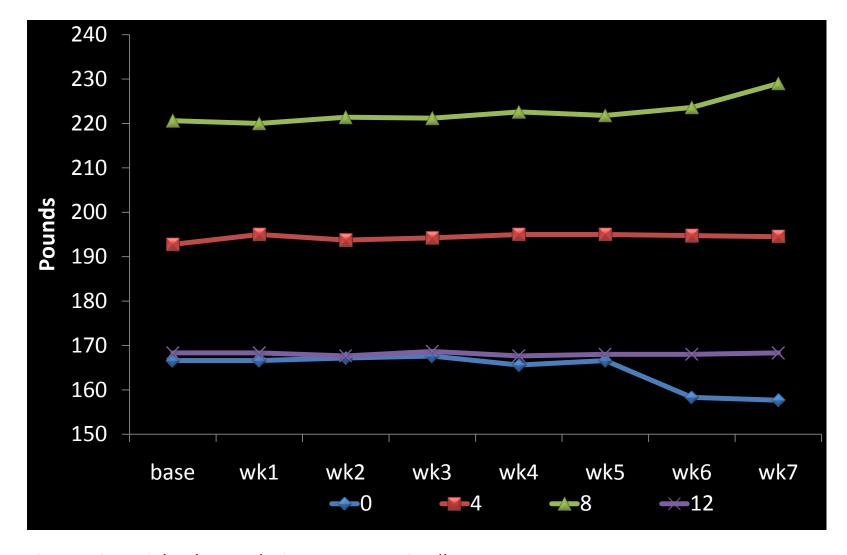


Figure 12. Weight change during treatment in all groups

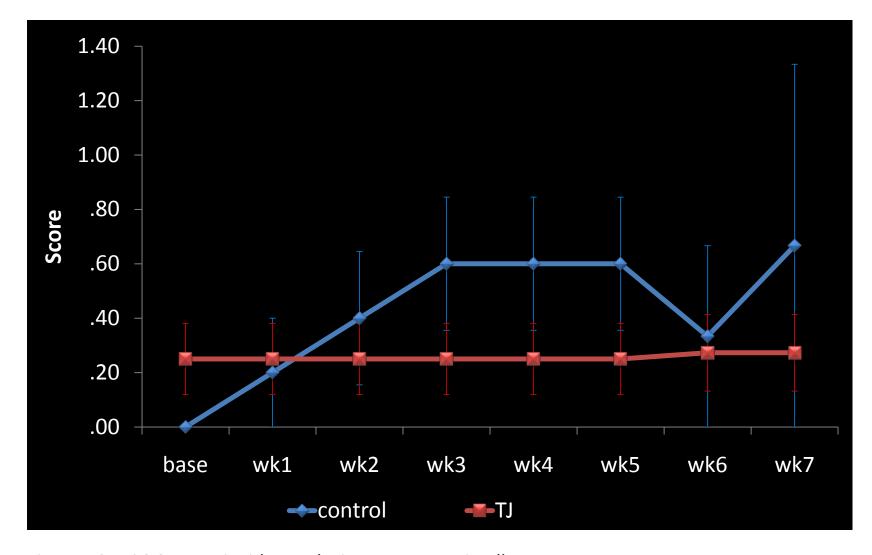


Figure 13. ECOG score incidence during treatment in all groups

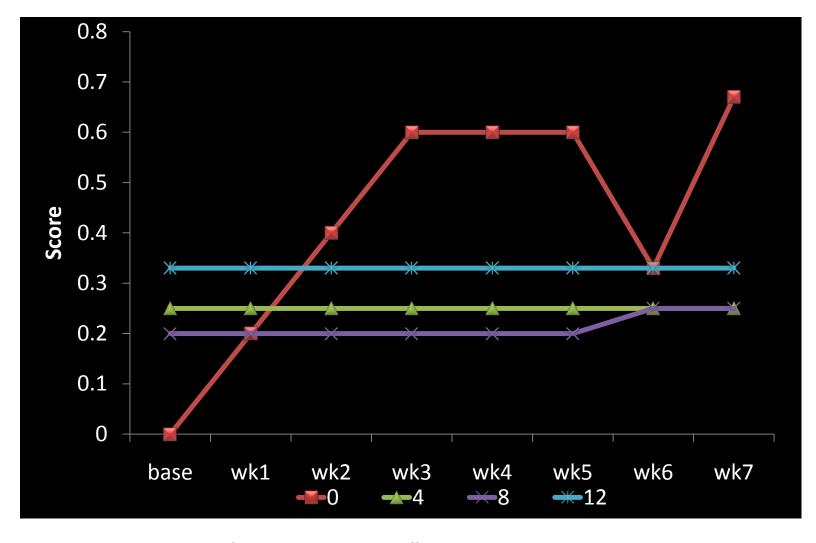


Figure 14. ECOG scores during treatment in all groups

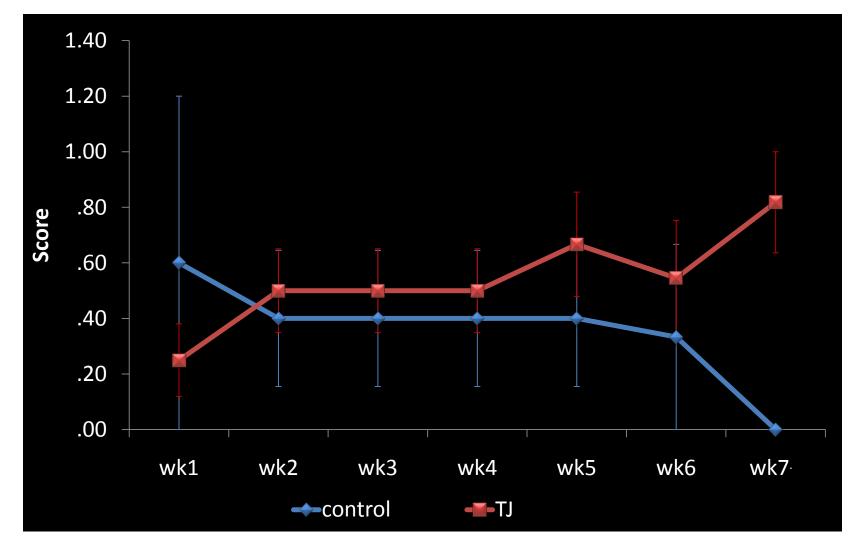


Figure 15. Urinary frequency during treatment in control and all treatment groups

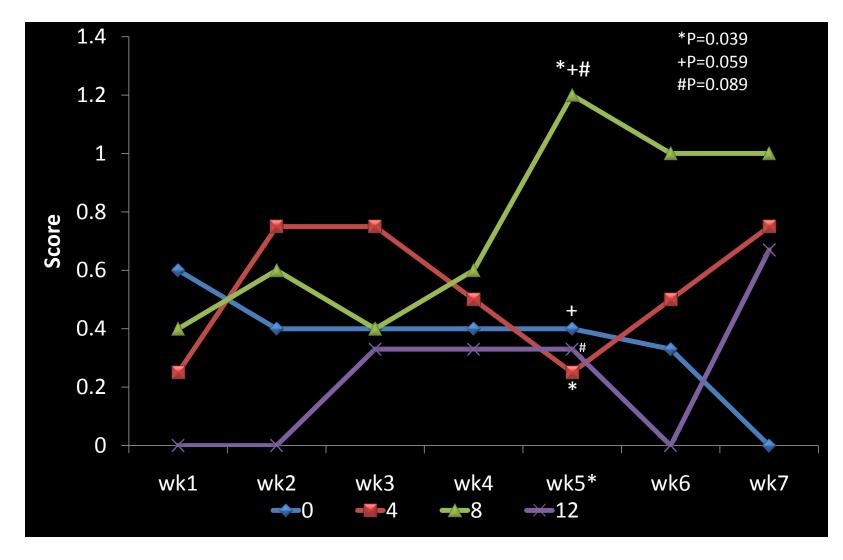


Figure 16. Urinary frequency incidence during treatment in all groups

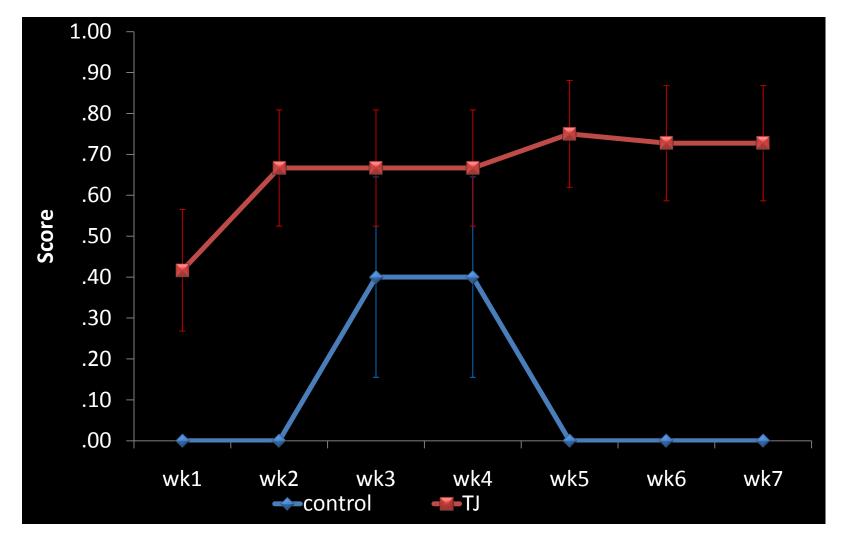


Figure 17. Urinary urgency during treatment in control and all treatment groups

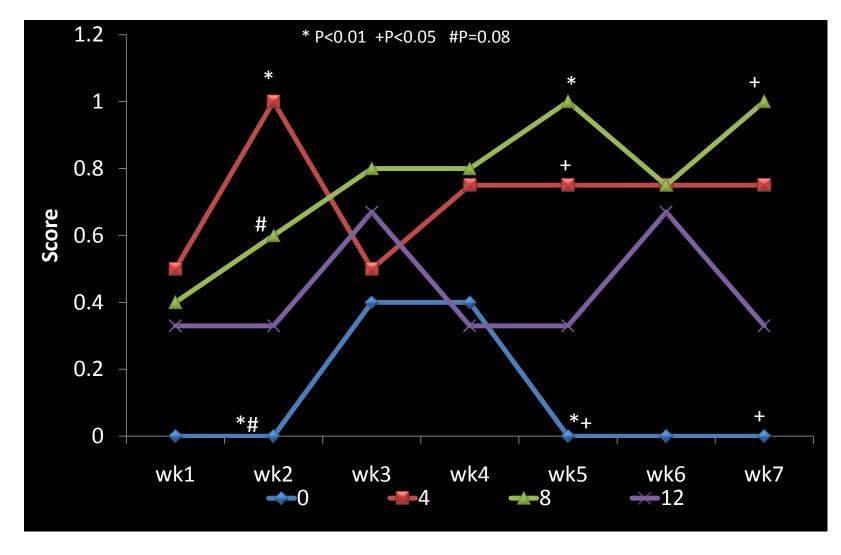


Figure 18. Urinary urgency incidence during treatment in all groups

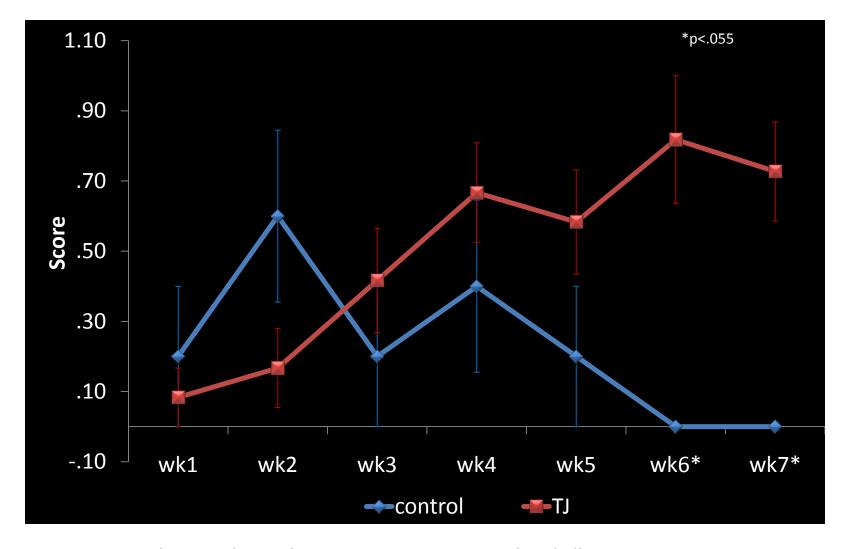


Figure 19. Diarrhea incidence during treatment in control and all treatment groups

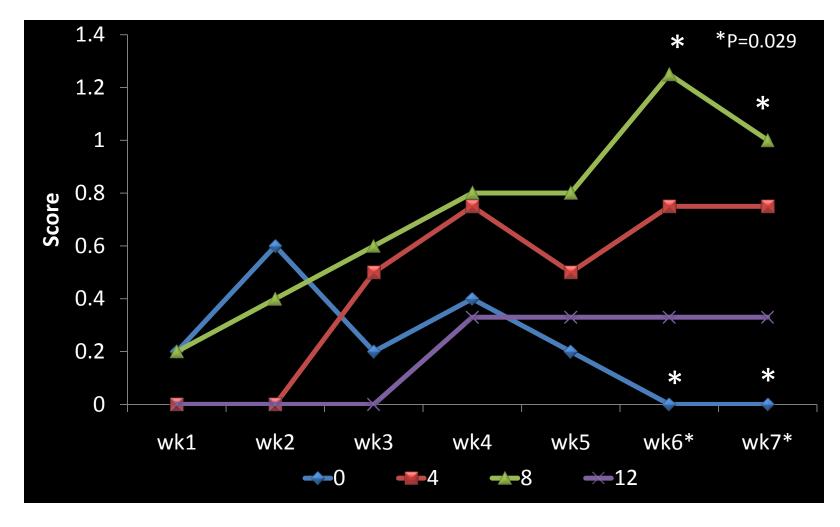


Figure 20. Diarrhea incidence during treatment in all groups

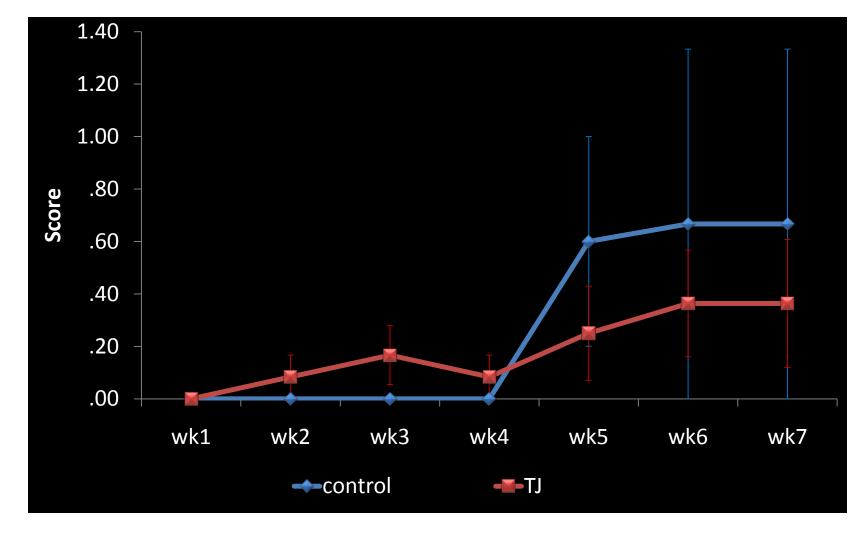


Figure 21. Proctitis incidence during treatment in control and all treatment groups

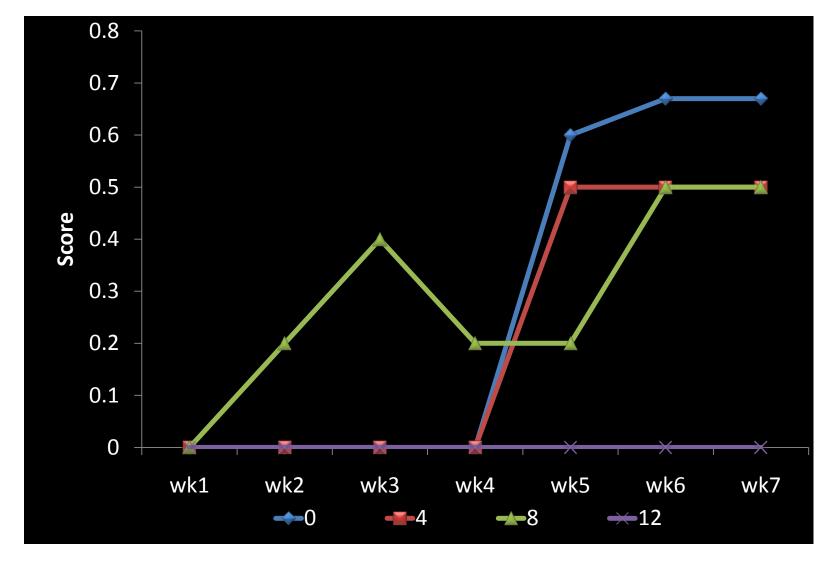


Figure 22. Proctitis incidence during treatment in all groups