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DOI: 10.1016/j.biombioe.2016.05.021

Document Version

Accepted author manuscript

Link to publication record in Manchester Research Explorer

Citation for published version (APA):

Kiss, A. A., Lange, J. P., Schuur, B., Brilman, D. W. F., van der Ham, A. G. J., & Kersten, S. R. A. (2016). Separation technology–Making a difference in biorefineries. *Biomass and Bioenergy*, *95*, 296-309. https://doi.org/10.1016/j.biombioe.2016.05.021

Published in: Biomass and Bioenergy

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1	Separation technology - Making a difference in biorefineries
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12	Keywords: Separation technology, Biomass conversion, Process intensification, Biorefinery
13	Highlights
14 15 16 17	 Separation and purification accountable for largest part of costs in biorefineries Hybrid technologies based on process intensification ready to make a big difference Energy efficient separation technologies that require low CapEx and OpEx
18	Abstract

19 In the quest for a sustainable bio-based economy, biorefineries play a central role as they 20 involve the sustainable processing of biomass into marketable products and energy. This 21 paper aims to provide a perspective on applications of separations that can make a great 22 difference in biorefineries, by significantly reducing the costs and thus making the processes 23 competitive without subsidies. A parallel is drawn between bio-refinery and petro-refinery, to 24 highlight the specific separation challenges encountered in biorefineries and point out the 25 impact of separations on the total costs. Existing and foreseen separations in biorefineries are 26 reviewed, and the upcoming challenges in the bio-domain (additional to current fossil) are 27 identified. Relevant industrial examples are provided to illustrate the tremendous eco-28 efficiency benefits of well-designed separation processes based on process intensification 29 principles (e.g. reactive separations, dividing-wall column, affinity and trigger-enhanced 30 separations). These examples also illustrate the low sustainability of several bio-separations 31 currently practiced, in terms of high relative energy requirements, large amounts of gypsum 32 co-production and/or excess use of caustic.

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1 1. Introduction

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2 Biomass is nature's way of storing solar energy and is considered a renewable alternative to 3 fossil resources. Biorefineries involve the sustainable processing of bioresources into a 4 spectrum of marketable products and energy (IEA Bioenergy Task 42). However, this is not 5 an entirely new concept, as biomass converting technologies (sugar, starch, pulp & paper) 6 have been around for a long time and these can be partly considered as biorefineries. As the 7 biorefinery concept evolved, a variety of criteria is used now for the taxonomy: technological 8 implementation status, type of raw material used, main intermediate produced, conversion 9 process applied, or a combination of these features (de Jong and Jungmeier, 2015).

Remarkably, the nature is fully able to operate efficiently with a mix of reactants that lead to a mix of products. Hence Mother Nature does not offer pure chemicals to the humankind. The chemical industry, on the other hand, developed along the line of using almost pure raw materials (obtained in pre-treatment steps) that are converted into a mix of products which are separated afterwards into pure components/intermediates. These are then combined to make materials with well-tuned and controlled structures and properties. Thus, purity is (so far) the key to control these processes.

17 A comparison between the oil refineries and biorefineries will help to put things into context 18 and show the role of separation technology in both cases. Figure 1 illustrates this analogy. A 19 classical refinery transforms fossil sources (oil & gas) into energy, fuels, and chemicals. The 20 raw materials are converted first into building blocks, from which more valuable 21 intermediates and end-user products are obtained. The separation and purification steps in oil 22 refineries typically use distillation technologies (along with liquid extraction, crystallization, 23 absorption, adsorption, membranes) that can account for 40-50% of the total costs (Kiss, 24 2013). A biorefinery has quite similar functions to a classic refinery, but in this case the 25 feedstock used is biomass instead of oil. In addition to the production of energy, fuels and 26 chemicals, a biorefinery may also produce bioproducts such as food for humans and livestock. 27 On the way to chemicals, the biomass is converted to biochemical building blocks, a set of 28 functional molecules that are more suitable for organic synthesis. It is worth noting that all 29 separation technologies applied in biorefineries are derivatives from the petro-chemical 30 industry. As the balance in properties of the streams is different than the ones in petro-31 chemical industry, a different balance/emphasis in separation technologies could be expected. 32 The biomass pre-treatment step (involving mainly phase separations, but also size reduction, 33 removing dirt/sand, etc.) already leads to some primary products, and it is responsible for 20-

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40% of the total costs (Ramaswamy et al., 2013). Afterwards the conditioned biomass is used

1 on dedicated technology platforms where it is converted into products. These platforms are 2 primarily determined by the chemistry routes and the feedstock composition (de Jong et al., 3 2012). Several technology platforms are available, such as: *biogas* (methane from anaerobic 4 digestion), syngas (a mixture of CO and H₂ from gasification for Fischer-Tropsh synthesis), 5 hydrogen (by steam reforming, water electrolysis, or fermentation), C6 sugars (hydrolysis of 6 sucrose, starch, cellulose and hemicellulose), C5 sugars (hydrolysis of hemicellulose), lignin 7 (from lignocellulosic biomass processing), pyrolysis oil or bio-crude (hydrothermal 8 liquefaction oil, as obtained by a thermo-chemical biomass-to-liquid technology), oils and 9 fats (from oil crops, algae and waste oils), organic juice (liquid after pressing wet biomass), 10 *electricity and heat* (internal use or to grid). These platforms produce in most cases liquid or 11 gas mixtures of components that require expensive separation steps contributing to the largest 12 part of the total costs (Ramaswamy et al., 2013; de Jong and Jungmeier, 2015).

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14 While there are many recent books and reviews tackling the topic of biorefinery – mainly the 15 pre-treatment and conversion steps (Centi and van Santen, 2007; Kamm et al., 2010; 16 FitzPatrick et al., 2010; Cherubini, 2010; Naik et al., 2010; Pandey and Kim, 2011; 17 Bridgwater 2012; Aresta et al., 2012; Stuart and El-Halwagi, 2012; Pandey et al., 2015; and 18 others) – there are only very few publications dedicated to the (role of) separation technology 19 in biorefineries (Huang et al., 2008; Ramaswamy et al., 2013). These latter papers give an 20 overview of available separation technologies and discuss some applications, but no critical 21 evaluation is made and no boundary conditions are provided for the actual applicability.

22 This perspective paper aims to fill this gap, by identifying the key challenges (caused by 23 reactivity, polarity, dilution, complex matrix) and foreseen separations in the envisaged 24 biorefineries, raising some warning signals with regard to several unsustainable separations 25 that should be avoided, while indicating when certain separations are applicable or not by 26 using relevant examples. An overview is given with regard to applications of separation 27 technologies that can make a great difference in biorefineries by simplifying processes (less 28 equipment) and reducing the operating costs (lower energy requirements), making the 29 biorefineries more competitive even without any subsidy. The focus of this work is on the 30 efficient separation (meaning molecular separation, not phase separation) of fluid mixtures 31 using various technologies (e.g. reactive separations, advanced distillation, affinity and 32 trigger-enhanced separations, etc). Several showcases are presented to illustrate the impact of 33 separation on eco-efficiency.

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1 2. Challenges and opportunities for separations in biorefineries

2 In addition to biomass conversion steps (including pre-treatment), separation and purification 3 of biomass converted components into products is of utmost importance for biorefinery 4 applications. Compared to conventional chemical processes, the separation in biorefineries 5 may be severely hindered by factors, such as (in case of water-based biorefineries): low feed 6 concentration, product inhibition issues, and/or low product yield leading to very diluted 7 (aqueous) streams that resemble more waste water streams than typical chemical reactor 8 effluents. Note that we assume mainly water-based biorefinery streams throughout this paper. 9 However, we will occasionally consider other type of biorefinery stream where appropriate, 10 e.g. streams that consist of pyrolysis or liquefaction oils and lipids.

11 Moreover, the presence of water and oxygenated compounds forming complex matrices and 12 azeotropes constitutes an additional difficulty as compared to oil refineries. Another key 13 aspect is the production of high-volume low-value biofuels and/or low-volume high-value 14 chemicals. The economy of scale in the biorefinery domain is at a different level as compared to fossil based refineries, as the mass yield of product to feed is typically much worse (with 15 16 very few exceptions) thus making the CapEx per kg product even more important. In many 17 cases the separation part is the crucial factor determining the commercial success of 18 biorefineries, as it accounts for the largest part of the total costs (Ramaswamy et al., 2013; 19 Kiss et al., 2015).

20 To summarize, the challenges in separation technology for biorefineries relate to the presence 21 of reactive mixtures, often temperature restricted to 150 °C, polarity of components present in 22 the mixture, often much diluted aqueous solutions, and a complex matrix of organics that 23 often contains also inorganic compounds with a detrimental effect on extraction. Examples of 24 separation challenges foreseen in the biorefineries include among others:

25 Concentration of oxygenates from water (aqueous streams), as for example acids (e.g. 26 acetic, lactic, succinic, levulinic) and/or their corresponding salts; light oxygenates 27 (e.g. alcohols, carbonyls); heavy oxygenates (e.g. sugars)

- 28 • Removal of sugars from a mix of phenols (e.g. from pyrolysis oil)
- 29 Separation of lights from heavies, with similar molecular structure (e.g. bio-oil) •
- Decontamination of sugar (e.g. removal of furanics/phenolics for fermentation, and 30 31 removal of amino acids/ash for chemo catalysis)

32 Fractionating (algal/microbial) biomass in lipids, proteins, carbohydrate rich fractions • 33 With the risk of oversimplification, the novelty of many separation challenges relies on two

main characteristics of the streams, namely their thermal instability and high dilution. These
challenges are partly compensated by an important opportunity: bio-based components are
generally highly polar and functional hence offer opportunities for intermolecular affinities.

4 Thermal instability is a key issue in biorefineries since it hinders the workhorse of separation 5 technologies, the classic distillation. Complex and reactive mixtures (prone to fouling) are 6 obtained from processing of lignocellulose or sugar streams, and they contain hundreds of 7 components as found in liquefaction and pyrolysis oil, or the sugar product stream from acid or hydrogenation/hydrogenolysis processes. The thermal instability results mainly from the 8 9 presence high functionalized molecules, which contain reactive functionalities such as 10 hydroxyls, aldehydes, ketones and carboxylic acid groups or furanic rings. The challenge is 11 further worsened by the fact that biorefinery streams often consist of complex mixtures 12 containing hundreds of components and often come at low pH (Lange, 2015). Dewatering the 13 product may worsen the situation by increasing the concentration of reactive components and 14 contained acids. This makes subsequent separation at high temperature (e.g. by distillation), 15 unfavorable compared to alternative low-temperature separations (e.g. LLX, permeation). But 16 a few alternative distillation technologies may still be worth considering, e.g. vacuum 17 distillation, molecular distillation, short-path distillation, pass-through distillation. Such an 18 example is the separation of phenolics from bio-oil (Elkasabi et al., 2014).

19 Diluted (aqueous) solutions require typically a pre-concentration before the actual separation 20 and purification. Indeed biomass conversion often proceeds in diluted liquid phase e.g. as 21 diluted sugar stream (e.g. sugar juice or starch/cellulose hydrolysate) or derivatives from such 22 stream (e.g. after conversion of the diluted sugars to a variety of oxygenates such as alcohols, 23 polyols, acids or furanics). High dilution often results in large reaction processing equipment 24 and expensive separation schemes. The abundant solvent (generally water) is generally lighter 25 than the desired product hence, re-concentration by solvent evaporation can be an expensive 26 endeavor (due to the high enthalpy of vaporization of water). Alternative, non-distillative re-27 concentration may include precipitation (e.g. of acid salts), extraction (e.g. of furanics and 28 phenolics) or ultrafiltration (e.g. of bio-oils and other high molecular weight products).

Large feed variability is expected in the feed streams entering the separation or purification stages in a biorefinery compared to the low variability in a regular oil refinery. This is clearly the outcome of the large diversity in terms of properties and content that the raw biomass may possess. Such variability may greatly affect the performance of the separation and purification processes and would require a very careful design of the processes in order to achieve their targets. As variability may affect some process configurations more than other factors, with a
 direct economic impact, this issue has to be considered in the separation process selection.

3 Feed detoxification is a peculiar case of separation with diluted feed. Microorganisms used

4 for sugar fermentation are often sensitive to low concentration of toxins such as acetate,

5 furanic or phenolics. In contrast, chemical catalysts are more sensitive to either basic minerals

6 (e.g. for acid-catalysis) and/or N-, S- and Cl-components (e.g. for hydrogenation) contained in

7 the feed (Lange, 2015). New and inexpensive technologies for feed detoxification will likely

8 become of prime importance. Owing to the low concentration of toxins, feed detoxification

9 will likely include ion-exchange, adsorption, extraction or precipitation.

These separation challenges are particularly encountered in the front end of the biorefineries. Once these intermediate streams are upgraded to well-defined and thermally stable platform molecules, conventional distillation may come back as very effective approach. This does not mean that separation research ends here as biorefineries offer a wealth of opportunities in advanced distillations, e.g. azeotropic, extractive or reactive distillations.

15

16 **3.** Applications of separations in biorefineries

17 This section gives an overview of the main applications of key separation technologies in 18 biorefineries, conveniently grouped here according to the separation mechanism. Additional 19 details about each of these technologies are provided in the books of Seader et al. (2011), de 20 Haan and Bosch (2013), Kiss (2013) and in particular Ramaswamy et al. (2013) who puts 21 them in the context of biorefineries.

22

23 **3.1 Phase-change separations**

24 **Distillation** processes are used in biorefineries for the separation and dehydration of alcohols 25 (bioethanol and biobutanol), purification of biodiesel, isolation of volatile organic compounds 26 (essential oils) and phytochemicals from biomass (extract), concentration of chemicals in 27 pyrolysis oil and separation of various fractions (alcohols, aldehydes, ketones, acids, 28 phenolics and sugars). A particular case is the steam distillation process that is used for the 29 direct separation of the desirable components from solid (not liquid) biomass feedstock (Bergeron et al., 2012; Ramaswamy et al., 2013). Distillation is a prime candidate for 30 31 purification of final low molecular weight products, but it is not convenient for high-boiling 32 components, particularly when these are highly functionalized and thus temperature sensitive. 33 **Precipitation and crystallization** applications to biorefineries include: ethanol precipitation 34 for recovery of hemicelluloses from the pretreated hydrolyzates (pre-hydrolysis liquor) and

spent liquors from pulp mills, precipitation by acidification using CO₂ or sulfuric acid applied
to extract lignin from kraft black liquor, separation of succinic acid from fermentation broth
or proteins from aqueous solutions (Huang et al., 2008; Ramaswamy et al., 2013).

Filtration is an established solid-liquid separation technology (Sparks and Chase, 2015). As solid biomass is typically the starting feedstock in biorefineries, a number of solid-liquid separation tasks (including filtration) are involved, such as the separation of prehydrolyzate and post-distillation slurries. Hence the use of efficient and cost-effective filtration processes is important for improving the overall process performance (Ramaswamy et al., 2013).

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10 **3.2 Affinity-based separations**

11 Liquid-liquid extraction (LLX) plays an important role in biorefineries, being used for 12 separating biofuels (bioalcohols) and chemicals (carboxylic acids) from dilute mixtures 13 (fermentation broths), extraction of acetic acid from biomass hydrolysates using mixed 14 solvents, extraction of 5-hydroxymethylfurfural (HMF) from an aqueous reaction solution 15 using methyl isobutyl ketone (MIBK) as solvent, removing inhibitors from biomass hydro-16 lyzates and various impurities (soap, methanol, and glycerol) from biodiesel, extraction of 17 chemicals (aqueous extractions, or extraction with hydrophobic-polar solvent and antisolvent) 18 of fast pyrolysis bio-oils, and extraction of succinic, maleic, lactic, and itaconic acids with 19 ionic liquids, or by reactive extraction using amines (Huang et al., 2008; Ramaswamy et al., 20 2013). The main advantage of using LLX is in the recovery of (diluted) components that are 21 boiling at higher temperature than the solvent (water) and are thermo-sensitive. Extraction 22 avoids or minimizes the need to distil out huge amounts of water, which is very energy 23 intensive. Depending on concentration and nature of the solute, reactive extraction with e.g. 24 amines could be required due to the low extractability with physical solvents such as MIBK. 25 Relatively low boiling solutes (e.g. acetic acid) can be stripped readily from such composite 26 solvents (amines diluted in MIBK, 1-octanol or other diluents). For higher boiling solutes 27 (e.g. lactic acid), direct thermal regeneration is not feasible due to stability issues of both 28 solvent and solute. In such cases, extraction – back-extraction is applied (Krzyzaniak et al., 29 2014), aiming at concentrating the solution to the max, prior to further thermal purification.

30 *Supercritical fluid extraction* (SFE) using CO_2 is very suitable for extracting hydrophobic 31 constituents from biomass, e.g. recovery of value added phytochemicals (pigments, phenolics, 32 carotenoids) and lipids from microalgae (Huang and Ramaswamy, 2012). The advantages of 33 SFE are speed (no surface tension, low viscosities, fast diffusivity) and selectivity (properties 34 of a sc-fluid can be altered by varying pressure and temperature). But the requirements for high pressures increases the process costs compared to conventional LLX, so SFE process should be used only where the advantages are major and offset the drawbacks. In many cases SFE is used with the intent to avoid water distillation. Notably, the evaporation of water costs in terms of thermal energy 2.26 MJ kg⁻¹, while the CO₂ re-compression to supercritical conditions requires electricity up to 0.54 MJ kg⁻¹ (i.e. ~1.3 MJ kg⁻¹ equivalent thermal energy when considering electricity generation).

Solid-liquid extraction (SLE) technology includes classic solid-liquid extraction, ultrasoundand microwave-assisted extraction, as well as pressurized subcritical liquid extraction.
Biomass contains value-added co-products such as bioactive compounds and phytochemicals
(phenolics, terpenes, sterols, enzymes, polysaccharides, alkaloids, toxins, and pigments) that
can be extracted using SLE prior to or during conversion (Huang and Ramaswamy, 2012).

12 *Absorption* is used for the removal of acid gases (H_2S and CO_2) from syngas prior to further 13 conversion into methanol and diesel fuel, or CO_2 capturing. A particular method is *reactive* 14 *absorption* that combines the absorption of gases in liquid solutions with simultaneous 15 chemical reactions. This method is used for gas treatment and purification, removal of 16 harmful substances, and the production of various industrial chemicals (Yildirim et al., 2012).

17 Adsorption can be used in biorefineries for the efficient removal of inhibitors from biomass 18 hydrolysate, separation and purification of biofuels and chemicals (dehydration of bioalcohols 19 with molecular sieve), removal of impurities (glycerol, methanol, free fatty acids, soap, 20 catalyst, metals, water and glycerides) from the raw biodiesel using magnesium silicate 21 (Magnesol®) or magnesium silicate and bentonite as adsorbent (Ramaswamy et al., 2013).

Simulated moving bed (SMB) is often used for separation processes by adsorption in the biodomain: e.g. purification of glycerol from biodiesel production (using the Ambersep BD50 resin, or gel-type acidic ion-exchange resin beads) where the raffinate stream contains salts and organic impurities including FFAs, purification of oligosaccharides (made up of xylose and arabinose units), isolation of lactic acid from acetic acid, separation of sugars (glucose and xylose) and EmimAc (IL) from the biomass hydrolyzate (Ramaswamy et al., 2013).

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29 **3.3 Size/charge-based separations**

30 *Ion exchange* (IEX) can be used in biorefineries for the removal of inhibitors from biomass 31 hydrolysate (acid, salts), purification of biodiesel to remove impurities (FFA, glycerol, 32 methanol, and soap using IEX resin Lewatit® GF202), separation of carboxylic acids, 33 purification of succinic acid (IEXs are used for simultaneous acidification and crystallization) and of xylose from biomass prehydrolyzates (Ramaswamy et al., 2013). Fortunately, IEX also

2 trap other organics by means of adsorption.

3 Membrane separations, such as microfiltration (0.050-10 µm), ultrafiltration (1-100 nm), 4 and *nanofiltration* (< 2 nm) can be used in biorefineries for the separation of biofuels and 5 chemicals, depending on the molecules to be separated: e.g. removal of inhibitors (acetic 6 acid), algal biomass harvesting, separation of hemicelluloses from biomass hydrolyzates (or 7 process water of pulp mills), lignin recovery from pulp mill waste liquors or biomass 8 prehydrolysis liquor, biodiesel separation and purification (Atadashi et al., 2011), separation 9 of liquid mixtures (carboxylic acids recovery from dilute solutions), gas separation and 10 purification (He et al., 2012; Ramaswamy et al., 2013). Membrane pervaporation/pertraction 11 is a particular case – a highly selective, economical, safe, and eco-friendly technology – being 12 a promising method for liquid-liquid separations in biorefineries, with applications such as the 13 removal of inhibitory products from fermentation broth (Huang et al. 2008).

Electrodialysis is used in biorefineries for the separation of organic acids or carboxylic acids (acetic acid, oxalic acid, citric acid, gluconic acid, and succinic acid) from their fermentation broths (Huang et al., 2007), recovery of basic components such as mono-ethanol amine (de Groot et al., 2011), bipolar membrane electrodialysis for the production of organic bases (de Groot et al., 2011) and of lactic acid by continuous fermentation with an integrated product recovery process (Strathmann, 2010), as well as recovery of gluconic, ascorbic and succinic acids from their sodium salts (Wang et al., 2011; Ramaswamy et al., 2013).

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22 **3.4 Reactive separations**

Mostly applied to equilibrium reactions (to drive the reaction to complete conversions) and insitu product removal (where the products are removed to avoid the bio-/catalyst poisoning), reactive separation processes make use of process intensification principles to combine the reaction and separation step in single unit, thus leading to significant advantages in terms of eco-efficiency: high conversion/yield, enhanced selectivity, high productivity, improved energy efficiency, and less equipment (Schmidt-Traub and Gorak, 2010; Kiss, 2013).

Reactive distillation (RD) is applied in biorefineries to the production of succinate esters, fatty esters & biodiesel (Kiss, 2014), upgrade of flash pyrolysis oil, esterification of succinic and acetic acid from fermentation of biomass carbohydrates (Orjuela et al., 2011), esterification of glycerol to produce triacetin (Ramaswamy et al., 2013).

33 *Reaction-membrane separations* include membrane (bio)reactors, bioreactor-membrane per-

34 vaporation/distillation. Membrane reactors can be used for biodiesel production (Kiss, 2014),

while fermentation-membrane pervaporation systems are used in butanol production (in situ
 product-recovery technology). For succinic acid production a fermentation-bipolar membrane
 electrodialysis system can be used (Ramaswamy et al., 2013).

Extractive fermentation applications in biorefineries include for example the extraction of butanol from the fermentation broth (Dhamole et al., 2012). Proposed configurations consist of fermentation integrated with in-situ product removal, as well as external product removal in an extraction column with a recycle of product-lean broth. Membrane-assisted solvent extraction can also be used for recovery and separation of organic acids, biofuels, and other chemicals (Ramaswamy et al., 2013).

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11 **3.5 Technology selection**

Based on our (industrial) experience we can draw the following recommendation of selectingseparation technologies and give the following initial rules of thumb.

14 \rightarrow Distillation is a strong option under the following conditions:

- o The temperature of thermal degradation of all components largely needs to
 exceed the reachable boiling point for distillates (at atmospheric or vacuum
 conditions).
- 18 The difference in boiling point of products to separate needs to exceed 5 °C.
 - The concentration of the distillate needs to exceed 10 % wt of the feed stream.
- Affinity separation is usable as an economical separation or pre-concentration method
 under specific conditions:
 - Product and medium need to show significant differences in chemical affinity on at least one specific scale, e.g. acidity/basicity, polarity, H-bonding, etc.
- 24 o Low concentration of solute can be affordable if the affinity gap is sufficiently
 25 large to be exploited in an economical way.
- 26 o The extractant needs to provide a moderate level of bonding, i.e. not too weak
 27 to ensure efficient extraction but at the same time not too strong to allow
 28 efficient regeneration (Jongmans et al., 2012).
- 29 o Interactions of impurities (trace compounds) in the feed and the extractant need
 30 to be minimized.
- 31 o Very critical is an affordable sorbent recovery concept. This can be distillation
 32 (for extraction), thermal desorption (for adsorption), depressurization (for
 33 adsorption), or back extraction with a medium that is convenient for
 34 downstream processing.

 \rightarrow Permeation is a viable option for separation when the right materials are available

- The product and medium need to show large difference in diffusivity. This is often related to molecular weight. However it can also imply difference in chemical affinity, when the diffusion proceeds via selective 'dissolution' in polymeric membrane, or in charge, i.e. in size of the solvated ion.
 - The permeable materials should be able to cope with the large feed variability.
 - The availability of affordable permeable materials (able to operate at extreme conditions) is critical to such processes and must be evaluated at early stages.
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10 **4. Evaluation of separation processes**

11 Among others, process systems engineering (PSE) can contribute with shortcut methods that 12 allow the techno-economic evaluations of separation processes. This section briefly provides 13 some short cut methods to evaluate the cost of distillation and sustainability of separations. It 14 is worth mentioning that mainly distillation and extraction, as well as micro/ultra/nano-15 filtration are used on large scale production in biorefineries, while other separation methods 16 are still in research and development stage. However, in case of extraction the costs of 17 recovery (often by distillation) determine the overall costs, while the cost of filtration largely 18 depends on the cost of membranes (i.e. designated membrane area and type of material).

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20 **4.1 Evaluation of distillation processes**

The cost of process segments, including complex distillation trains, was reported to be largely dictated by the duty of their major equipment (Lange et al., 1996, Lange, 2001). Indeed, the investment cost (inside battery limit) of process segments of fuel and chemical plants was shown to correlate with their overall energy transfer duty according to the following equation (cost updated to 2014):

26 Investment cost (ISBL, M 2014) = 4.7 * (exchange duty [MW])^{0.55} (1)

27 Process flow modeling programs, which are now common tools for engineers, easily provide 28 the equipment duties required by this equation. However, a well-converged and optimized 29 process flow model may already be too demanding for a preliminary cost estimate. Simpler 30 though cruder estimates may then become handy. A few decades ago, Rudd et al. (1973) 31 eluded on such crude estimate by proposing to use the ratio 'feed flow / boiling-point 32 difference' as indicator for distillation cost, e.g. for selecting the cheapest sequence of 33 distillation columns for complex systems. However, they did not provide the support for this 34 indicator, or any specific factor for converting this ratio into distillation cost. Building on this

1 concept, the new concept of *distillation resistance* (R_d) – detailed in another dedicated paper 2 (Lange, 2016) – can be taken as good proxy for the overall duty of a complex distillation and 3 the overall distillation costs. This concept assumes that the thermal duty is determined by the 4 fraction of the top streams (which are vaporized and then condensed), and the difference in 5 boiling points of components to separate (which determine the reflux ratios). R_d is defined as 6 the sum of the individual ratio $(F_i/\Delta T_i)$ of all condensable distillates, where F_i is the mass 7 fraction [%wt on total feed intake] of each component and ΔT_i is the temperature difference 8 [°C] between its boiling point and that of the first heavier component in the feed.

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$$R_d [100/°C] = \sum F_i [\%wt] / \Delta T_i [°C]$$

(2)

- 10 Simple distillations show $R_d < 1$ and a duty of $\sim 1 \text{ GJ t}^{-1}$ feed whereas demanding distillations
- 11 show R_d =3-7 and a duty of 3-8 GJ t⁻¹ feed. Reasonable linear regressions are proposed to
- 12 relate R_d with reboiler and total duties, which allow estimating the OpEx and overall CapEx.
- 13 Thermal duty [GJ t⁻¹ feed] = $1.1 \times R_d$ (3)
- 14 Firing duty [GJ t⁻¹ feed] = $0.6 \times R_d$ (4)
- 15 OpEx [t^{-1} feed] = 5 [GJ^{-1}] * Firing duty [GJ t^{-1} feed] = 3×R_d (5)
- 16 CapEx $[10^6 \,\text{\$}, 2014] = 4.7 \,\text{*} \,(\text{Thermal duty} \,[\text{MW}])^{0.55}$ (6)
- The concept of distillation resistance is based on an average distillation quality. Such assumption is reasonable when evaluating complex distillation trains that separate more than 5 streams. But care should be taken when stringent requirements are made on product purity or product recovery, particularly when the evaluation is limited to a single and demanding distillation. Also, when process intensification methods are employed (e.g. DWC technology) the capital and operating expenditures should be reduced by about 25% (Kiss, 2013).
- Besides the cost estimation of distillation, the energy efficiency of such processes can be also estimated. Pleşu et al. (2015) proposed a simple equation that is easily usable in calculations to evaluate the distillation sequence energy efficiency (DSE) for any alternative. DSE is calculated as the sum of feed molar fractions (x_i) multiplied by the product of column efficiencies – which are equal to the Carnot efficiency for distillates, 100% for bottoms product. In case of side stream products with boiling point higher than that of the feed, the efficiency is 100% - otherwise it corresponds to the Carnot efficiency of the column.

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$$DSE = \sum_{i=1}^{O} \left(x_i \prod_{C=1}^{N} \eta_C \right) \text{ (maximize)}$$
 (7)

31 where $\eta_{\rm C}$ is the Carnot efficiency of the column: $\eta_{\rm C} = (T_{\rm bottom} - T_{\rm distillate}) / T_{\rm bottom}$). Once DSE 32 is calculated, all the alternative solutions can be sorted from higher to lower efficiency. Another simple criterion used especially when comparing various distillation alternatives with each other is the $N_S \times (RR+1)$ which is directly proportional to the total annual costs (Kiss, 2013). This product includes the number of stages (N_S) that is proportional to the column height h_c, and the reflux ratio + distillate (RR+1) factor that is proportional to the column diameter d_c. Furthermore, d_c is proportional to the production rate, while the reflux ratio + distillate (RR+1) multiplied with the distillate rate (D) and the heat of vaporization (ΔH^{vap}) gives the energy requirements of the distillation column:

8
$$C_{\text{shell}} = f_p \left(M \& S/280 \right) d_c^{1.066} h_c^{0.802}$$
 (8)

9
$$C_{\text{hex (reboiler/condenser)}} = (M\&S/280) c_x A^{0.65}$$
 (9)

10
$$C_{\text{energy}} = (RR+1) D \Delta H^{\text{vap}} hours/year$$
 (10)

where f_p and c_x are cost factors, M&S is the Marshall & Swift equipment cost index, A is the heat exchange area. Note that in contrast to the previous methods, such analysis requires a short-cut or detailed modeling of the column, but is presumably more accurate.

14

15 **4.2** Evaluation of alternative separation processes

16 The correlation between energy exchange duty and CapEx discussed above is not limited to 17 distillation but can be extended to other separation technologies that are energy intensive, i.e. 18 that require more than 10 MW energy exchange. Large-scale extraction likely meets these 19 requirements when the solvent is regenerated by means of distillation. Indeed, the solvent 20 distillation column is likely to dominate the cost of the whole extraction train. The cost 21 contribution of the extraction may then be accounted for by addition of a modest cost penalty 22 of up to 25%. By extension, the concept of distillation resistance R_d may also be of value for a 23 first estimate of CapEx. However, the accuracy might become questionable as only two 24 components are separated.

25 In cases of separation train based on extraction and back extraction, the correlation between 26 exchange duty and CapEx might not apply. The same is expected for other, more energy-lean 27 separation technologies e.g. based on crystallization, precipitation, membrane permeation or 28 ion-exchange. Other indicators need to be developed for such applications. One alternative 29 indicator could be the mass transfer flux involved in the separation. This approach was 30 proposed for evaluating the recovery of butanol, lactic acid and phenol from fermentation 31 broth by means of adsorption, pervaporation, extraction and pertraction (Oudshoorn et al., 32 2010). The mass transfer flux was used to estimate the interfacial area and the vessel volume 33 required for separation. Various equations are then used to derive CapEx and OpEx from the 34 estimated separation volume. When applicable, the regeneration of the auxiliary phase was accounted for by doubling the CapEx and OpEx, with the assumption that regeneration
 follows the same limitations (e.g. same time constant) as the separation itself.

- 3 In case of membrane filtration, Pilutti and Nemeth (2003) reported capital costs of membrane
- 4 systems starting at 198-462 m^{-3} (0.75-1.75 gal^{-1}) for a permeate capacity of 157-315 $m^{3} h^{-1}$
- 5 (1-2 MGD, million gallons per day), but dropping fast to about 105 m⁻³ (0.4 gal⁻¹) at 788
- 6 $m^3 h^{-1}$ (5 MGD) and even 53-106 \$ m^{-3} (0.2-0.4 \$ gal⁻¹) at capacities of 1577-6309 $m^3 h^{-1}$ (10-
- 7 40 MGD). The capital costs include membranes, skids, racks, compressors, blowers, pumps,
- piping, instrumentation, controls, and other components needed for a complete and operable
 system. However, the operating costs are more difficult to estimate as they depend on many
- 10 factors including water quality, flux, recovery, pretreatment, and cost of consumables.
- 11 In a more recent study, Movahed (2010) estimated MF/UF capital costs in the range of 160-
- 12 320 $\text{\$ per m}^3 \text{day}^{-1}$ (0.6-1.2 \$ per GPD) permeate capacity, and operating & maintenance costs
- 13 (O&M) costs of 0.08-0.1 \mbox{m}^{-3} (0.3-0.4 \mbox{Kgal}^{-1}) with overall water costs of 0.1-0.16 \mbox{m}^{-3}
- 14 (0.4-0.6 \$ Kgal⁻¹). These costs include equipment, piping, controls, membrane replacement,
- 15 chemicals and power but do not include building, site work, and finished water pumping.
- 16

17 **4.3** Critical evaluation of sustainability

Sustainability of processes can be evaluated based on various metrics, such as the life-cycle analysis (LCA), eco-cost value (EVR) ratio, (socio-)eco-efficiency analysis, or the AIChE sustainability index (Dimian et al., 2014). It should be realized here that all separation must be economically and environmentally sensible. For instance, one needs to ensure that the value of the targeted product exceeds the recovery cost. This may imply that the energy required for recovering and purifying a bio-fuel component needs to remain a modest fraction of the heating value of the product itself.

25 One specific example of a bio-fuel is n-butanol, a fuel with an energy content of 36 MJ kg⁻¹. 26 Garcia-Chavez et al. (2012) calculated that a traditional thermal separation (two-distillation 27 columns with a decanter after the first column to pass the heterogeneous azeotrope) uses about 21.3 MJ kg⁻¹ to concentrate the n-butanol out of a 1 wt% solution, which is almost 60% 28 29 of the energy content of butanol. In contrast, a process based on liquid extraction would only 30 cost about 15% of the energy content, of which the majority was spent on evaporation of co-31 extracted water. Even less energy consuming (3.76 MJ kg⁻¹, 10.4% of the energy content of n-32 butanol at a comparable feed concentration range of 0.4-1.2 wt%) is the dual extraction process proposed by Kurkijärvi et al (2014), which benefits from one more hydrophilic 33 34 solvent and one more hydrophobic solvent to reduce the losses of solvent to the raffinate and

1 simultaneously reduces the heat required to evaporate co-extracted water. The efficiency of 2 extraction processes is thus highly dependent on the selectivity of the solvent and the more 3 selective, the less energy spent on evaporation of water. A similar conclusion can be drawn 4 for a wider spectrum of separation technologies based on the overview papers of Oudshoorn 5 et al. (2009) and Huang et al. (2014). Oudshoorn et al. (2009) considered a variety of 6 technologies to recover n-butanol from fermentation broth and developed short-cuts for 7 estimating their energy requirement. Not surprisingly, the separation selectivity appeared to 8 largely dictate the energy efficiency of separation across all technologies. In fact, few 9 technologies appeared to have an energy requirement that is lower than ~10% of the heating value of n-butanol (36 MJ kg⁻¹). Figure 2 – based on the work of Oudshoorn et al. (2009) – 10 11 gives an overview of these technologies for butanol recovery.

12 Next to biofuels, also production of chemicals using fermentation is hindered by the typically 13 low concentrations. To illustrate the effect of the concentration of the solute in the fermen-14 tation broth on energy demand of the recovery, Figure 3 plots the steam costs as function of 15 the broth concentration when a traditional distillation would be used to separate the binary 16 acetic acid / water mixture. When the steam cost is higher or equal to the product price then 17 the process is obviously not economically feasible. A typical value aimed for is to have 18 energy costs that are about or even less than 10% of the product price. It clearly follows that it 19 is economically infeasible and unsustainable to recover acetic acid by distillation from the 20 various fermentation processes, e.g. from a glucose fermentation to produce acetic acid in 5 21 wt% concentration, or from a fermented wastewater with only 1 wt%. In traditional liquid 22 extraction processes so called physical solvents are used, for example using ethyl acetate as in 23 the text book example of Seader et al. (2011). With these solvents, distributions below unity 24 are observed (IJmker et al. 2014), and the reduction in heat duty of approximately a factor of 25 three is mainly due to the easier recovery by distillation than the initial process. For example, 26 Seader et al. (2011) make advantageous use from a heterogeneous azeotrope. However, 27 Figure 3 clearly shows that these physical solvents will not enable economic processes to 28 recover acids from fermented wastewater, as the energy costs are higher than 10% of the 29 product price. Chemically active solvents contain an extractant that complexes the acid, 30 resulting in much higher distribution coefficients (Krzyzaniak et al., 2013; IJmker et al. 2014; 31 Reyhanitash et al., 2015), but recovery of the acids from these complex-forming solvents is 32 not as easy as from a physical solvent and typically involves a back-extraction after which 33 further treatment is required. This is a nice example of the dilemma mentioned earlier of 34 selecting an extractant that is neither too weak nor too strong. For the future developments in

this field, there is a clear need for innovative process concepts, e.g. in the direction of CO₂enhanced extractions (Reyhanitash et al., 2015), that make a step reduction in the energy demand such that the recovery costs become much lower than the product price.

4 New developments on the liquid-liquid extraction of organic acids are not only important for 5 the reduction of the energy usage during the recovery of the solvent, but also with respect to 6 the co-generation of large amounts of gypsum as byproduct from fermentation-based acid 7 productions. In the traditional approach the fermentation is treated with Ca(OH)₂ to maintain 8 microbial activity also at higher acid concentrations, while the extraction is favored at low 9 pH. Hence the treatment with H_2SO_4 boosts the extraction efficiency, but co-generates large 10 amounts of gypsum, e.g. for the industrial fermentative production of lactic acid this is about 11 one ton of gypsum per ton of lactic acid, which is obviously unsustainable.

12 Although the focus in this section was limited to a single biofuel (n-butanol) and a single 13 chemicals category (organic acids), the presented challenges in sustainable separations are 14 certainly valid for a wide range of biorefineries, that typically deal with highly diluted streams 15 and large amounts of water.

16

17 **5.** Case studies of separations

This section provides a selection of showcases, applicable to biofuels and chemicals, whichillustrate the great impact of innovative separations in biorefineries.

20

21 **5.1 Advanced distillation**

22 Distillation remains a powerful separation technology, particularly at the high-value end of 23 the biorefinery for the separation and purification of thermally stable and well defined 24 components. New developments show much promises. Kiss (2013) illustrated the beneficial 25 use of dividing-wall column (DWC) technology in the production of biofuels, leading to 26 significant capital and energy savings. Of particular interest is bioethanol, a renewable fuel 27 produced by various routes (corn-to-ethanol, sugarcane-to-ethanol, integrated lignocellulosic 28 biomass-to-ethanol) in which the raw materials undergo several pre-treatment steps before 29 entering the fermentation stage. All these technologies produce diluted bioethanol (typically 30 5-12 % wt ethanol) that is further concentrated to reach the requirements of the international 31 bioethanol standards. To reach the purity targets, an energy demanding separation is needed in 32 practice, in order to overcome the presence of the binary azeotrope ethanol-water (95.63 % wt 33 ethanol). The separation is typically carried out by distillation, the first step being a pre-34 concentration distillation column (PDC) that increases the ethanol content from 5-12% up to

1 91-94 %wt. The second step consists of the ethanol dehydration, up to concentrations 2 exceeding the azeotropic composition. Quite a number of separation alternatives are available 3 as described in the literature: pervaporation, adsorption, pressure-swing distillation, extractive 4 distillation, azeotropic distillation, and hybrid methods combining these options (Vane, 2008). 5 Among them, extractive distillation (ED) is still the option of choice in case of large scale 6 production of bioethanol fuel. Typically, ED is performed in a sequence of two columns, one 7 being the extractive distillation column (EDC) which separates ethanol, while the other one is 8 the solvent recovery column (SRC) that recovers the mass separating agent that is recycled 9 back in the process. Further improvements to the extractive distillation process were 10 proposed, with the aim to increase the energy efficiency of bioethanol purification. One that 11 stands out is a novel heat pump assisted extractive distillation process, based on mechanically 12 driven heat pumps (Kiss and Infante Ferreira, 2016). This process efficiently combines vapor 13 recompression (VRC) with dividing-wall column technology that allows the combination of 14 all functions (three classic columns) into just one column – flowsheet shown in Figure 4 (Luo 15 et al., 2015). Table 1 provides more details about the key performance indicators, including the total investment, operating and annual costs (Luo et al., 2015). Due to the use of a 16 17 compressor and a larger side-reboiler required by the VRC system, the total investment cost 18 of this VRC E-DWC process is about 29% higher than for the classical process, but this is 19 compensated by the significant energy savings, which exceed 60% at a direct comparison. The specific energy requirements are only 4.46 MJ kg⁻¹ (1.24 kWh kg⁻¹) ethanol for the novel 20 21 VRC assisted E-DWC, thus energy savings of over 50% are possible as compared to other 22 classic alternatives described in literature (Baeyens et al., 2015).

23

24 **5.2 Reactive separations**

Another opportunity of a bio-separation is reactive separation, being reactive distillation or reactive extraction, to circumvent difficulties in separation or in reaction (e.g. thermodynamic equilibrium or secondary product degradation).

Reactive distillation is an established technology, used for example in the recovery of acetic acid from aqueous liquors. Since acetic acid (normal boiling point of 118°C) is higher boiling than water, recovering acetic acid by traditional distillation from dilute streams is not economical due to the large amount of water that needs to be removed overhead. One method relies on recovering the acetic acid in a valuable form, such as acetate. When a reactive distillation process is carried out, in which the acetic acid reacts with methanol, the formed methyl acetate (n.b.p. 57 °C) is then recovered and easily separated over the top (Agreda and Zoeller, 1993). More recently, Le et al. (2015) described a heterogeneous azeotropic
 distillation schemes in a DWC for a feed mixture of water, acetic acid and an organic
 component (isobutyl acetate) that acts as an entrainer. Remarkable, the proposed Petlyuk
 DWC system proposed achieves energy savings of about 20%.

5 Biodiesel production by reactive separations received significant attention during the past 6 decade (Kiss, 2014). Being a mixture of fatty acid methyl esters (FAME), biodiesel is a 7 renewable fuel used complementary to petro-diesel fuel. Its main synthesis routes are by 8 either trans-esterification of tri-alkyl glycerides (TAG) or esterification of free fatty acids 9 (FFA). Both routes use catalysts (homogeneous or solid acid / base catalysts) and can be 10 applied in many types of industrial production processes (e.g. batch, continuous, supercritical, 11 enzymatic, multi-step, reactive separations).

12
$$TAG + 3 MeOH \rightleftharpoons 3 FAME + Glycerol$$
 (trans-esterification) (11)

13
$$FFA + MeOH \rightleftharpoons FAME + H_2O$$
 (esterification) (12)

14 The trans-esterification is mainly base catalyzed, while the esterification is catalyzed by acids 15 - although alternative acid/base catalysts could be used but at prohibitive reaction rates. The reaction time can be dramatically shortened by increasing the liquid-liquid interfacial area by 16 17 various process intensification techniques - e.g. static mixers, micro-channels reactors, 18 microwaves assisted reactors, ultrasound assisted reactors, rotating / spinning tube reactors 19 and centrifugal contactors (Qui et al., 2010) - or by integrating the reaction and separations 20 steps to pull the equilibrium to full conversions, e.g. reactive distillation (Kiss and Bildea, 21 2012), reactive absorption, reactive extraction, reactive membrane separators (Kiss, 2014), 22 and centrifugal contactors (Kraai et al., 2008). After the FAME synthesis stage, there are 23 several down-stream processing steps required for catalyst neutralization and salt removal, 24 alcohol recovery and recycle, as well as glycerol and biodiesel purification. Among the 25 process intensification alternatives investigated, reactive distillation is the most promising 26 option. Figure 5 illustrates a heat integrated reactive distillation process for fatty acids 27 esterification, and a simpler one similar to reactive absorption (reactive column without 28 reboiler and condenser). Table 2 provides the process parameters, whereas Figure 6 (Kiss and 29 Bildea, 2012) compares the energy requirements for a classic two-step process based on pre-30 treatment of free fatty acids and trans-esterification of glycerides versus reported reactive 31 separation processes based on esterification of waste oils with high FFA content (Kiss, 2014). 32 The figures are worth noting, especially considering the on-going quest on increasing the eco-33 efficiency of biodiesel production. The specific energy use in reactive separation processes is significantly lower than the FAME purification step alone in the conventional process. On top
of the energy savings, the reactive separation processes benefit from lower investment costs
and reduced plant footprint due to less equipment being used (Kiss and Bildea, 2012).

4 Levulinic acid can be converted to nylon intermediates adipic acid or caprolactam. Key 5 processing step include the hydrogenation to γ -valerolactone (gVL), transesterification to 6 alkyl pentenoate and e.g. methoxy carbonylation to methyl adipate. The transesterification can 7 be performed under reactive distillation conditions. The concept was demonstrated to achieve 8 >95 mol% yield using various homogeneous and heterogeneous acids immersed in gVL with 9 continuous feed of MeOH and stripping of Me-pentenoate entrained with MeOH vapor – as 10 shown in Figure 7 (Lange et al., 2007).

11 Reactive distillation comes very handy for coupling levulinate ester with furfural to produce 12 C_{10} oxygenates that can be subsequently converted to C_{10} hydrocarbons by hydro-13 deoxygenation (Lange et al., 2012). The coupling step is typically performed with an excess 14 base (e.g. NaHCO₃) and is followed by an acidification step to generate the acid form of the 15 furfurylidene-levulinic acid intermediate. However, reactive distillation allows the use of a 16 catalytic amount of base, e.g. solid base, by stripping water out of the medium and thereby 17 avoiding hydrolysis of the ester and subsequent neutralization of the basic catalyst.

18 Reactive extraction is a very promising technology for the conversion of bio-based feedstock.
19 Reactive extraction of cellulose-derived levulinic and formic acids with butene was proposed
20 by Gürbüz et al. (2011) to obtain levulinate and formate esters, thereby allowing for recovery
21 and recycle of sulfuric acid. The esters were converted over a dual-catalyst-bed system to
22 GVL and 2-butanol, followed by production of butene to be recycled for reactive extraction
23 and to be converted to liquid fuels by oligomerization.

24 Another example of reactive extraction can be found in the conversion of sugars to furans. For 25 instance, bi-phasic systems were explored for the conversion of fructose to HMF, as reviewed 26 by Kuster (1990). Similarly, Moreau et al. (1998) converted xylose to furfural using HY and 27 HMOR zeolites at 170°C using biphasic systems (water/toluene or the less effective water/MIBK). Beyond improved yield, biphasic operation also allows to recover the diluted 28 29 furfural by solvent extraction rather than by distillation of the water-rich furfural/water 30 azeotrope. Bi-phasic systems were further developed for both furfural and HMF by the group 31 of Dumesic who proposed an integrated process based on sugar dehydration, furanics 32 recovery and furanics upgrading (Roman-Leshkov et al., 2006). It is worth noting the recent 33 identification of alkylphenols, which were reported to be effective extractant for furfural even at low extractant/water ratio and could be regenerated by simple distillation of furfural (Azadi
 et al., 2012). Such solvent could be produced from the lignin waste-product of the biorefinery.

3

4 **5.3 Triggered affinity separations**

5 The emphasis of this section is not so much on the extraction part that is typically the focus of 6 most studies, but rather on the subsequent 'non-distillative' separation.

7 Extractions with operational swings. Following the reasoning above, the usefulness of 8 thermal separations in biorefineries is limited to situations where reasonable concentrations of 9 the desired products are obtained, e.g. bio-ethanol productions with yields of 5-12%. For 10 more diluted systems, it is generally better to apply affinity separation, such as (reactive) 11 liquid-liquid extraction. Often, and especially when the distribution ratios - defined as 12 [solute]_{extract} / [solute]_{raffinate} – are low, the concentration in the extract phase is low and the 13 heat duty for direct thermal recovery from the solvent extremely high. Therefore, extraction – 14 back-extraction cycles are commonly used, in which achieving a high concentration factor is 15 aimed for in order to facilitate further purification, e.g. by crystallization. The traditional 16 approach to increase the ratio between the distribution ratio in the back-extraction stage and 17 the distribution ratio in the extraction stage is to apply a temperature swing, but also diluent 18 swings may be applied (Krzyzaniak et al., 2013). The drawback of applying a diluent swing is 19 the need for an additional recovery step, in which the preferably low boiling diluent is evaporated before the solvent is sent back to the primary extraction process. More recently, 20 21 the use of CO_2 was reported to boost the distribution of acetic acid in an extraction from very 22 dilute (1 wt%) aqueous solutions resembling fermented aqueous wastewater (Reyhanitash et 23 al., 2015). Using this approach, distribution ratios could be increased up to 7-fold, and the 24 ratio of the acetic acid to co-extracted water increased from 1wt% to 34wt%. In addition to 25 the large concentration factor, a second benefit of using CO_2 instead of a volatile organic 26 diluent is that instead of a distillation, the added CO_2 is simply removed from the system by 27 depressurization which has a lower cost penalty than water evaporation.

Extractions with temperature induced phase splitting. It is also possible to induce a phase split in a homogeneous system by applying a trigger. Temperature induced phase splitting is a concept that makes use of changes in miscibility with temperature. Due to a temperature change, the miscibility reduces, resulting in a phase split creating two liquid phases. Two temperature-dependent phase splitting events are known, one achieved when exceeding the lower critical solution temperature (LCST), and the other when cooling below the upper critical solution temperature (UCST). Both LCST-behavior and UCST behavior may be

1 exploited in combination with liquid extraction. LCST behavior is mostly utilized in aqueous 2 two-phase systems (ATPS) – see Figure 8 and Wohlfarth (2004) for more examples. ATPS 3 systems have been applied for a range of applications interesting for biorefineries, e.g. for 4 fractraction of salts (Milosevic et al., 2014), proteins (Grilo et al., 2016) and alkaloids (Freire 5 et al. 2012). After the extraction, the solvent recovery is achieved by an induced phase split 6 due to a mild temperature increase. Especially in biorefineries, the mildness of this technique 7 valuable due to the sensitivity of many molecules (e.g. proteins), which in such ATPS may be 8 isolated without losing their functionality. Examples of ATPS application include extraction 9 of fatty acids (Glembin et al., 2014), and proteins (Desai et al., 2014) in algae biorefineries. 10 Monteillet et al. (2014) showed another application of LCST behavior, where they combined 11 LCST with a magnetoresponse to create multiresponsive ionic liquid emulsions capable of 12 extracting β -carothene.

13 A very recent application of UCST behavior was recently reported by Kumar et al. (2015), 14 who applied the induced phase splitting upon cooling in the fractionation of a complex bio-oil 15 stream generated by thermal liquefaction of lignocellulosic biomass. This approach was then 16 applied to recover the light fraction of the bio-oil for recycling as liquefaction medium – see 17 Figure 9 (Kumar et al., 2015). This approach is based on hot extraction (T~70 °C) of the light 18 fraction of the oil with a suitable extraction solvent followed by cold (T~25 $^{\circ}$ C) de-mixing of 19 the light fraction and the extraction solvent. The study illustrated the selection of the 20 extraction solvent and definition of required solvent properties, showed the potential of 21 multistage extraction / regeneration for the bio-oil produced by direct thermal liquefaction, 22 extended the concept to fractionate a petroleum crude oil, discussed the theoretical basis of 23 the fractionation using polymer solution theory, and showed a low energy requirement of the 24 extraction process by means of process simulation, i.e., an equivalent of ~1% of the biomass 25 intake (Kumar et al., 2015).

Extractions with CO₂-induced phase splitting. The concept of splitting phases by bubbling CO₂ through a homogeneous system was first reported by Jessop et al (2005), and following this seminal paper, many publications appeared on extraction of lipids from natural sources such as soy bean (Phan et al., 2009) and microalgae (Boyd et al., 2012).

30 Du et al. (2013) have worked out a conceptual process scheme (Figure 10) for the extraction 31 of lipids from microalgae using CO_2 switchable secondary amines. It was shown that with the 32 secondary amines it was actually possible to extract the lipids from wet algae. However, to 33 make the benchmark hexane extraction efficient, drying of the algae prior to extraction was 34 needed for good extraction efficiency. This type of process with its ability to extract from wet algae was further examined and compared with other technologies to extract lipids from algae (Du et al., 2015). Although to date the research on this topic is ongoing, and detailed aspects of process elements like solvent recovery are still under investigation, the study showed that among the other technologies investigated (such as hexane extraction and extraction with supercritical CO₂), using CO₂-switchable solvents was the only approach to yield more energy (37.8 MJ kg⁻¹ lipids) than the separation costs (19.8 MJ kg⁻¹ lipids).

7

8 **6.** Conclusions

9 Separations in biorefineries are responsible for the largest part of the total costs and hence any 10 major improvements in separations can make the difference between a commercial success 11 and failure. By reviewing the existing and foreseen separations in biorefineries, identifying 12 and discussing the additional challenges in the bio-domain, and providing an overview of 13 applications of separation technologies in biorefineries, this perspective paper concludes that 14 there are many opportunities to improve separations in biorefineries. A selection of relevant 15 examples related to biofuels and chemicals proved that separation technology can make a big difference in biorefineries, by considerably reducing the overall energy requirements and the 16 17 production costs, thus increasing competitiveness of bio-based fuels and chemicals.

18

19 Acknowledgement

20 The vivid and useful discussions with all the participants of the 1st biorefinery conference

- 21 (Biorefinery I: Chemicals and Materials from Thermo-Chemical Biomass Conversion and
- 22 *Related Processes*, 27 Sep 2 Oct 2015, Chania, Crete, Greece) are gratefully acknowledged.
- 23

24 **References**

- Agreda V. H., Zoeller J. R., Acetic acid and its derivatives, Marcel Dekker, New-York,
 USA, 1993.
- Aresta M., Dibenedetto A., Dumeignil F., Biorefinery: From biomass to chemicals and
 fuels, de Gruyter, Berlin, Germany, 2012.
- 3. Atadashi I., Aroua M. K., Aziz A. A., Biodiesel separation and purification: A review,
 Renewable Energy, 36 (2011), 437-443.
- 4. Azadi P., Carrasquillo-Flores R., Pagán-Torres Y. J., Gürbüz E. I., Farnood R., Dumesic J.
- A., Catalytic conversion of biomass using solvents derived from lignin, Green Chemistry,
 14 (2012), 1573-1576.
- 34 5. Baeyens J., Kang Q., Appels L., Dewil R., Lv Y., Tan T., Challenges and opportunities in

- improving the production of bio-ethanol, Progress in Energy and Combustion Science, 47
 (2015), 60-88.
- Bergeron C., Carrier D. J., Ramaswamy S., Biorefinery co-products: Phytochemicals,
 primary metabolites and value-added biomass processing, John Wiley & Sons, Chichester,
 UK, 2012.
- 7. Boyd A. R., Champagne P., McGinn P. J., MacDougall K. M., Melanson J. E., Jessop P.
 G., Switchable hydrophilicity solvents for lipid extraction from microalgae for biofuel
 production, Bioresource Technology, 118 (2012), 628-632.
- 8. Bridgwater A. V., Review of fast pyrolysis of biomass and product upgrading, Biomass
 and Bioenergy, 38 (2012), 68-94.
- 9. Centi G., van Santen R. A., Catalysis for renewables: From feedstock to energy
 production, Wiley-VCH, Weinheim, Germany, 2007.
- 13 10. Cherubini F., The biorefinery concept: Using biomass instead of oil for producing energy
 14 and chemicals, Energy Conversion and Management, 51 (2010), 1412-1421.
- 15 11. Christensen S. P., Donate F. A., Frank T. C., LaTulip R. J., Wilson L. C., Mutual
 solubility and lower critical solution temperature for water + glycol ether systems, Journal
 of Chemical Engineering Data, 50 (2005), 869-877.
- 18 12. de Groot M. T., Bos A. A. C. M., Lázaro A. P., de Rooij R. M., Bargeman G.,
 19 Electrodialysis for the concentration of ethanolamine salts, Journal of Membrane Science,
 20 371 (2011), 75-83.
- 13. de Groot M. T., de Rooij R. M., Bos A. A. C. M., Bargeman G., Bipolar membrane
 electrodialysis for the alkalinization of ethanolamine salts, Journal of Membrane Science,
 378 (2011), 415-424.
- 14. de Haan A. B., H. Bosch, Industrial separation processes, De Gruyter, Berlin, Germany,
 2013.
- 15. de Jong E., Higson A., Walsh P., Wellish M., Bio-based chemicals: Value-added products
 from biorefineries, International Energy Agency (IEA) report, Bioenergy Task 42, 2012.
- 16. de Jong E., Jungmeier G., Biorefinery concepts in comparison to petrochemical refineries,
 in Pandey A., Höfer R., Taherzadeh M., Nampoothiri M., Larroche C. (Eds), Industrial
- 30 biorefineries & white biotechnology, Elsevier, Waltham, USA, 2015.
- 31 17. Desai, R. K., Streefland, M., Wijffels, R. H., Eppink, M. H. M., Extraction and stability of
 32 selected proteins in ionic liquid based aqueous two phase systems, Green Chemistry, 16
 33 (2014), 2670-2679.
- 34 18. Dhamole P. B., Wang Z., Liu Y., Wang B., Feng H., Extractive fermentation with non-

ionic surfactants to enhance butanol production. Biomass and Bioenergy, 40 (2012), 112 119.

- 3 19. Dimian A.C., Bildea C.S., Kiss A.A., Integrated design and simulation of chemical
 4 processes, 2nd edition, Elsevier, Amsterdam, The Netherlands, 2014.
- 5 20. Du Y., Schuur B., Samorì C., Tagliavini E., Brilman D. W. F., Secondary amines as
 6 switchable solvents for lipid extraction from non-broken microalgae, Bioresource
 7 Technology, 149 (2013), 253-260.
- 8 21. Du Y., Schuur B., Kersten S. R. A., Brilman D. W. F., Opportunities for switchable
 9 solvents for lipid extraction from wet algal biomass: An energy evaluation, Algal
 10 Research, 11 (2015), 271-283.
- 22. Elkasabi Y., Mullen C. A., Boateng A. A., Distillation and isolation of commodity
 chemicals from bio-oil made by tail-gas reactive pyrolysis, ACS Sustainable Chemistry &
 Engineering, 2 (2014), 2042-2052.
- 14 23. FitzPatrick M., Champagne P., Cunningham M. F., Whitney R. A., A biorefinery
 15 processing perspective: Treatment of lignocellulosic materials for the production of valueadded products, Bioresource Technology, 101 (2010), 8915-8922.
- 17 24. Freire M. G., Cláudio A. F. M., Araújo J. M. M., Coutinho J. A. P., Marrucho I. M.,
 18 Canongia Lopes J. N., Rebelo L. P. N., Aqueous biphasic systems: A boost brought about
 19 by using ionic liquids, Chemical Society Reviews, 41 (2012), 4966-4995.
- 25. Garcia-Chavez L. Y., Garsia C. M., Schuur B., de Haan A.B., Biobutanol recovery using
 nonfluorinated task-specific ionic liquids, Industrial & Engineering Chemistry Research,
 51 (2012), 8293-8301.
- 23 26. Glembin P., Racheva R., Kerner M., Smirnova I., Micelle mediated extraction of fatty
 acids from microalgae cultures: Implementation for outdoor cultivation, Separation and
 Purification Technology, 135 (2014), 127-134.
- 26 27. Grilo A. L., Aires-Barrosa M. R., Azevedo A. M., Partitioning in aqueous two-phase
 27 systems: Fundamentals, applications and trends, Separation & Purification Reviews, 45
 28 (2016), 68-80.
- 28. Gürbüz E. I., Alonso D. M., Bond J. Q., Dumesic J. A., Reactive extraction of levulinate
 esters and conversion to γ-valerolactone for production of liquid fuels, 4 (2011), 357-361.
- 29. He Y., Bagley D. M., Leung K. T., Liss S. N., Liao B.-Q., Recent advances in membrane
 technologies for biorefining and bioenergy production. Biotechnology Advances, 30
 (2012), 817-858.
- 34 30. Huang C., Xu T., Zhang Y., Xue Y., Chen G., Application of electrodialysis to the

- production of organic acids: State-of-the-art and recent developments, Journal of
 Membrane Science, 288 (2007), 1-12.
- 3 31. Huang H. J., Ramaswamy S., Tschirner U. W., Ramarao B. V., A review of separation
 technologies in current and future biorefineries, Separation and Purification Technology
 62 (2008), 1-21.
- 32. Huang H. J., Ramaswamy S., Separation and purification of phytochemicals as coproducts in biorefineries, 37-54, in Bergeron C., Carrier D. J., Ramaswamy S. (Eds),
 Biorefinery co-products: Phytochemicals, primary metabolites and value-added biomass
 processing, John Wiley & Sons, Chichester, UK, 2012.
- 33. Huang H. J., Ramaswamy S., Liu Y., Separation and purification of biobutanol during
 bioconversion of biomass, Separation and Purification Technology, 132 (2014), 513-540.
- 34. IJmker H. M., Gramblicka M., Kersten S. R. A., van der Ham A. G. J., Schuur B., Acetic
 acid extraction from aqueous solutions using fatty acids, Separation and Purification
 Technology, 125 (2014), 256-263.
- 35. Jessop P. G., Heldebrant D. J., Li X., Eckert C. A., Liotta C. L., Green chemistry:
 Reversible nonpolar-to-polar solvent, Nature, 436 (2005), 1102-1102.
- 17 36. Jongmans M. T. G., Londono A., Mamilla S. B., Pragt H. J., Aaldering K. T. J., Bargeman
- 18 G., Nieuwhof M. R., ten Kate A. J. B., Verwer P., Kiss A. A., van Strien C. J. G., Schuur
- 19 B., de Haan A. B., Extractant screening for the separation of dichloroacetic acid from
- 20 monochloroacetic acid by extractive distillation, Separation & Purification Technology,
 21 98 (2012), 290-297.
- 37. Kamm B., Gruber P. R., Kamm M., Biorefineries Industrial processes and products:
 status quo and future directions, Wiley-VCH, Weinheim, Germany, 2010.

38. Kiss A. A., Novel applications of dividing-wall column technology to biofuel production
 processes, Journal of Chemical Technology and Biotechnology, 88 (2013), 1387-1404.

- 39. Kiss A.A., Advanced distillation technologies Design, control and applications. Wiley,
 Chichester, UK, 2013.
- 40. Kiss A. A., Process intensification technologies for biodiesel production Reactive
 separation processes, Springer, Heidelberg, Germany, 2014.
- 41. Kiss A. A., Bildea C. S., A review on biodiesel production by integrated reactive
 separation technologies, Journal of Chemical Technology and Biotechnology, 87 (2012),
 861-879.
- 42. Kiss A. A., Grievink J., Rito-Palomares M., A systems engineering perspective on process
 integration in industrial biotechnology, Journal of Chemical Technology and Bio-

- 1 technology, 90 (2015), 349-355.
- 43. Kiss A. A., Infante Ferreira C. A., Heat pumps in chemical process industry, CRC-Press
 (Taylor & Francis Group), Boca Raton (FL), US, 2016.
- 4 44. Kurkijärvi A., Lehtonen J., Linnekoski J., Novel dual extraction process for acetone–
 butanol-ethanol fermentation, Separation and Purification Technology, 124 (2014), 18-25.
- 45. Kuster B. F. M., 5-Hydroxymethylfurfural (HMF) A review focusing on its manufacture,
 Starch Starke, 42 (1990), 314-321.
- 46. Kraai G. N., van Zwol F., Schuur B., Heeres H. J., de Vries J. G., Two-phase
 (bio)catalytic reactions in a table-top centrifugal contact separator, Angewandte Chemie,
 120 (2008), 3969-3972.
- 47. Kraai G. N., van Zwol F., Schuur B., Heeres H. J., de Vries J. G., Two-phase
 (bio)catalytic reactions in a table-top centrifugal contact separator. Angewandte Chemie International Edition, 47 (2008), 3905-3908.
- 48. Krzyżaniak A., Leeman M., Vossebeld F., Visser T. J., Schuur B., de Haan A. B., Novel
 extractants for the recovery of fermentation derived lactic acid, Separation and
 Purification Technology, 111 (2013), 82-89.
- 49. Krzyżaniak A., Schuur B., de Haan A. B., Equilibrium studies on lactic acid extraction
 with N,N-didodecylpyridin-4-amine (DDAP) extractant, Chemical Engineering Science,
 109 (2014), 236-243.
- 50. Kumar S., Lange J-P., van Rossum G., Kersten S. R.A., Bio-oil fractionation by
 temperature-swing extraction: Principle and application, Biomass and Bioenergy, 83
 (2015), 96-104.
- 51. Kumar S., Lange J-P., van Rossum G., Kersten S. R. A., Liquefaction of lignocellulose in
 fractionated light bio-oil: Proof of concept and techno-economic assessment, ACS
 Sustainable Chemistry & Engineering, 3 (2015), 2271-2280.
- 52. Lange J-P., Fuels and chemicals manufacturing: Guidelines for understanding and
 minimising the production costs, CatTech, 5(2001), 82-95.
- 53. Lange J-P., Distillation in biorefineries When is it affordable?, ACS Sustainable
 Chemistry & Engineering (2016), *Submitted for publication*.
- 54. Lange J-P., Renewable feedstocks: The problem of catalyst deactivation and its
 mitigation, Angewandte Chemie International Edition, 54 (2015), 13186-13197.
- 55. Lange J-P., Tijm P. J. A., Processes for converting methane to liquid fuels: Economic
 screening through energy management, Chemical Engineering Science, 51 (1996), 23792387.

- 56. Lange J-P., Vestering J. Z., Haan R. J., Towards 'bio-based' nylon: conversion of γ valerolactone to methyl pentenoate under catalytic distillation conditions, Chemical
 Communications, 33 (2007), 3488-3490.
- 57. Lange J-P., van der Heide E., van Buijtenen J., Price R. J., Furfural a promising platform
 for lignocellulosic biofuels, ChemSusChem, 5 (2012), 150-166.
- 58. Le Q-K., Halvorsen I. J., Pajalic O., Skogestad S., Dividing wall columns for
 heterogeneous azeotropic distillation, Chemical Engineering Research and Design, 99
 (2015), 111-119.
- 59. Luo H., Bildea C. S., Kiss A. A., Novel heat-pump-assisted extractive distillation for
 bioethanol purification, Industrial & Engineering Chemistry Research, 54 (2015), 22082213.
- 60. Milosevic M., Staal K. J. J., Bargeman G., Schuur B., de Haan A. B., Fractionation of
 aqueous sodium salts by liquid-liquid extraction in aqueous two phase systems, Separation
 and Purification Technology, 125 (2014), 208-215
- 15 61. Movahed B., Introduction and applications of low pressure membranes (MF/UF), AMTA
 16 Technology Transfer Workshop, Knoxville, Tennessee, May 4-6, 2010.
- Monteillet H., Workamp M., Li X., Schuur B., Kleijn J. M., Leermakers F. A. M., Sprakel
 J., Multi-responsive ionic liquid emulsions stabilized by microgels, Chemical Communications, 50 (2014), 12197-12200.
- 63. Moreau C., Durand R., Peyron D., Duhamet J., Rivalier P., Selective preparation of
 furfural from xylose over microporous solid acid catalysts, Industrial Crops & Products, 7
 (1998), 95-99.
- 64. Naik S. N., Goud V. V., Rout P. K., Dalai A. K., Production of first and second generation
 biofuels: A comprehensive review, Renewable and Sustainable Energy Reviews, 14
 (2010), 578-597.
- 65. Orjuela A., Kolah A., Lira C. T., Miller D. J., Mixed succinic acid / acetic acid
 esterification with ethanol by reactive distillation, Industrial and Engineering Chemistry
 Research, 50 (2011), 9209-9220.
- 66. Oudshoorn A., van der Wielen L. A. M., Straathof A. J. J., Assessment of options for
 selective 1-butanol recovery from aqueous solution, Industrial & Engineering Chemistry
 Research, 48 (2009), 7325-7336.
- 32 67. Oudshoorn A., van den Berg C., Roelands C. P. M., Straathof A. J. J., van der Wielen L.
 33 A. M., Short-cut calculations for integrated product recovery options in fermentative
 34 production of bio-bulk chemicals, Process Biochemistry, 45 (2010), 1605-1615.

1	68. Pandey M. P., Kim C. S., Lignin depolymerization and conversion: A review of
2	thermochemical methods, Chemical Engineering & Technology, 34 (2011), 29-41.
3	69. Pandey A., Höfer R., Taherzadeh M., Nampoothiri M., Larroche C., Industrial bio-
4	refineries & white biotechnology, Elsevier, Waltham, USA, 2015.
5	70. Phan L., Brown H., White J., Hodgson A., Jessop P. G., Soybean oil extraction and
6	separation using switchable or expanded solvents, Green Chemistry, 11 (2009), 53-59.
7	71. Pilutti M., Nemeth J. E, Technical and cost review of commercially available MF/UF
8	membrane products, International Desalination Association, 2003. (BAH03-029).
9	72. Pleşu V., Bonet Ruiz A. E., J. Bonet, Llorens J., Iancu P., Shortcut assessment of
10	alternative distillation sequence schemes for process intensification, Computers &
11	Chemical Engineering, 83 (2015), 58-71.
12	73. Qiu Z. Y., Zhao L. N., Weather L., Process intensification technologies in continuous
13	biodiesel production, Chemical Engineering and Processing, 49 (2010), 323-330.
14	74. Ramaswamy S., Huang H-J., Ramarao B. V., Separation and purification technologies in
15	biorefineries, Wiley, Chichester, UK, 2013.
16	75. Reyhanitash E., Zaalberg B., IJmker H. M., Kersten S. R. A., B. Schuur, CO ₂ -enhanced
17	extraction of acetic acid from fermented wastewater, Green Chemistry, 17 (2015), 4393-
18	4400.
19	76. Roman-Leshkov Y., Chheda J. N., Dumesic J. A., Phase modifiers promote efficient
20	production of hydroxymethylfurfural from fructose, Science, 312 (2006), 1933-1937.
21	77. Rudd D. F., Powers G. J., Siirola J. J., Process synthesis, Prentice-Hall, Englewood Cliffs,
22	USA, 1973.
23	78. Schmidt-Traub H., Gorak A., Integrated reaction and separation operations: Modelling
24	and experimental validation, Springer, Berlin, Germany, 2010.
25	79. Seader J. D., Henley E. J., Roper D. K., Separation process principles, 3rd edition, Wiley,
26	Hoboken NJ, 2011.
27	80. Sparks T., Chase G., Filters and filtration handbook, 6th edition, Butterworth-Heinemann,
28	Oxford, UK, 2015.
29	81. Strathmann H., Electrodialysis, a mature technology with a multitude of new applications.
30	Desalination, 264 (2010), 268-288.
31	82. Stuart P. R., El-Halwagi M. M., Integrated biorefineries: Design, analysis, and
32	optimization, CRC Press, Boca Raton, USA, 2012.
33	83. Vane L. M., Separation technologies for the recovery and dehydration of alcohols from
34	fermentation broths, Biofuels, Bioproducts and Biorefining, 2 (2008), 553-588.

- 84. Wang Y., Huang C., Xu T. Which is more competitive for production of organic acids,
 ion-exchange or electrodialysis with bipolar membranes? Journal of Membrane Science
 374 (2011), 150-156.
- 4 85. Wohlfarth C., CRC Handbook of thermodynamic data of aqueous polymer solutions, CRC
 5 Press, Boca Raton, USA, 2004.
- 86. Yildirim O., Kiss A. A., Huser N., Lessmann K., Kenig E. Y., Reactive absorption in
 chemical process industry: A review on current activities, Chemical Engineering Journal,
 213 (2012), 371-391.
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1 Tables

2

3 **Table 1.** Bioethanol dehydration process comparison in terms of key performance indicators

Key performance indicator	Classic process	E-DWC process	VRC E- DWC process	Difference vs. classic (%)
Equipment cost breakdown (k\$)				
 – column shells (incl. internals) 	1,103	1,111	912	
 – condensers (heat exchangers) 	1,335	1,073	71	
– reboilers (heat exchangers)	885	1,442	356	
 process-process heat exchangers 	137	_	1,503	
 – compressor (VRC) 	—	_	1,632	
Total investment costs, TIC (k\$)	3,462	3,626	4,477	+29.3
Total operating costs, TOC (k\$ yr ⁻¹)	5,784	5,355	4,221	-27.0
Total annual costs, TAC (k\$ yr ⁻¹)	6,130	5,718	4,668	-23.8
CO_2 emissions (kg CO_2 t ⁻¹ product)	288.94	288.31	173.04	-40.1 (-61.1)
			(112.35)	
Thermal energy use (MJ kg ⁻¹ product)	7.45	7.45	2.88	-61.1
Electrical energy use (MJ kg ⁻¹ product)	n/a	n/a	0.50	n/a
Equivalent energy requirements (MJ kg ⁻¹)	7.45	7.45	4.46	-40.1

4 * Values given in parenthesis are for the case when electricity is generated from renewable sources.

Table 2. Comparison between integrated reactive-absorption vs reactive-distillation processes
 (at a plant capacity of 1250 kg h⁻¹ fatty esters)

Equipment / Parameter / Units		HI-RD	RA	HI-RA
Reactive column – reboiler duty (heater), kJ s ⁻¹	136	136	n/a	n/a
HEX-1 heat duty (fatty acid heater), kJ s ⁻¹	95	0	108	27
HEX-2 heat duty (methanol heater), kJ s ⁻¹	8	0	65	0
Reactive column – condenser duty (cooler), kJ s ⁻¹	- 72	- 72	n/a	n/a
HEX-3 water cooler/decanter, kJ s ⁻¹	- 6	- 6	- 77	0
COOLER heat duty (biodiesel cooler), kJ s ⁻¹	- 141	- 38	- 78	- 14
FLASH heat duty (methanol recovery), kJ s ⁻¹	0	0	0	0
Compressor power (electricity), kJ s ⁻¹	0.6	0.6	0.6	0.6
Reactive column, number of reactive stages	10	10	10	10
Feed stage number, for acid / alcohol streams	3 / 10	3 / 10	1 / 15	1 / 15
Reactive column diameter, m	0.4	0.4	0.4	0.4
Reflux ratio (mass ratio R/D), kg kg ⁻¹	0.10	0.10	n/a	n/a
Boil-up ratio (mass ratio V/B), kg kg ⁻¹	0.12	0.12	n/a	n/a
Productivity, kg ester kg ^{-1} catalyst h ^{-1}	20.4	20.4	19.2	19.2
Energy requirements per ton biodiesel, MJ t ⁻¹ FAME	688.3	391.6	498.2	77.7
Steam consumption, kg steam t ⁻¹ FAME	295	168	214	34

4

1	Figure captions (auto-updated)
2	
3	Figure 1. Analogy between petroleum refinery (left) and biorefinery (right)
4	
5	Figure 2. Estimated energy requirements (as percent of heating value of butanol) and
6	selectivity (defined as (XBuOH/XH2O)prod / (XBuOH/XH2O)feed) for the recovery of 2
7	% wt 1-butanol from aqueous solution. The numbers x/y °C represent the temperature for
8	recovery and regeneration (based on the data from Oudshoorn et al., 2009).
9	
10	Figure 3. Steam costs for distillation of acetic acid and water as function of the molar fraction
11	of acetic acid in the feed (a reflux ratio of 3 and a steam price of 6 €/GJ were assumed)
12	
13	Figure 4. Vapor recompression (VRC) assisted extractive dividing-wall column (E-DWC) for
14	bioethanol concentration and dehydration
15	
16	Figure 5. Heat integrated reactive distillation (top) and reactive absorption (bottom) processes
17	for biodiesel production
18	
19	Figure 6. Energy requirements for a conventional two-step process based on FFA pre-
20	treatment and trans-esterification (top) versus reactive separation processes (bottom)
21	
22	Figure 7. Trans-esterification of γ - $\omega\alpha\lambda\epsilon\rhoo\lambda\alpha\chi\tau$ ove with methanol to methyl pentenoates
23	(200 °C, lactone : pTSA = 50:1 molar ratio, MeOH feed rate = 11 mol molpTSA-1 h-1)
24	
25	Figure 8. Lower critical solution temperature (LCST) phase diagram for glycol ethers (based
26	on data from Christensen et al., 2005). Squares: water + diethylene glycol 2-methyl-1-butyl
27	ether, circles: water + diethylene glycol n-pentyl ether, triangles: water + triethylene glycol n-
28	heptyl ether
29	
30	Figure 9. Process block diagram of direct liquefaction followed by extraction of the light oil
31	and subsequent recovery of the extraction solvent
32	
33	Figure 10. Conceptual process for extraction of lipids from microalgae with a secondary
34	amine solvent. After the extraction stage, a CO2-induced phase splitting stage allows recovery
35	of the oil from the solvent, after which the solvent is regenerated by bubbling nitrogen.



9Figure 2. Estimated energy requirements (as percent of heating value of butanol) and10selectivity (defined as $(X_{BuOH}/X_{H2O})_{prod} / (X_{BuOH}/X_{H2O})_{feed})$ for the recovery of 2 % wt 1-11butanol from aqueous solution. The numbers x/y °C represent the temperature for recovery12and regeneration (based on the data from Oudshoorn et al., 2009).

Selectivity





Figure 3. Steam costs for distillation of acetic acid and water as function of the molar fraction
of acetic acid in the feed (a reflux ratio of 3 and a steam price of 6 €/GJ were assumed)





Figure 4. Vapor recompression (VRC) assisted extractive dividing-wall column (E-DWC) for
 bioethanol concentration and dehydration



Figure 5. Heat integrated reactive distillation (top) and reactive absorption (bottom) processes
for biodiesel production



Figure 6. Energy requirements for a conventional two-step process based on FFA pretreatment and trans-esterification (top) versus reactive separation processes (bottom)

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3Figure 7. Trans-esterification of γ-valerolactone with methanol to methyl pentenoates (2004°C, lactone : pTSA = 50:1 molar ratio, MeOH feed rate = 11 mol mol_{pTSA}⁻¹ h⁻¹)



Figure 8. Lower critical solution temperature (LCST) phase diagram for glycol ethers (based
 on data from Christensen et al., 2005). Squares: water + diethylene glycol 2-methyl-1-butyl
 ether, circles: water + diethylene glycol n-pentyl ether, triangles: water + triethylene glycol n heptyl ether





Figure 10. Conceptual process for extraction of lipids from microalgae with a secondary amine solvent. After the extraction stage, a CO₂-induced phase splitting stage allows recovery of the oil from the solvent, after which the solvent is regenerated by bubbling nitrogen.

filtration

water recycle

solids