

Biogeosciences, 13, 5245–5257, 2016  
www.biogeosciences.net/13/5245/2016/  
doi:10.5194/bg-13-5245-2016

© Author(s) 2016. CC Attribution 3.0 License.



# Effect of cover crops on greenhouse gas emissions in an irrigated field under integrated soil fertility management

Guillermo Guardia<sup>1</sup>, Diego Abalos<sup>2</sup>, Sonia García-Marco<sup>1</sup>, Miguel Quemada<sup>1</sup>, María Alonso-Ayuso<sup>1</sup>, Laura M. Cárdenas<sup>3</sup>, Elizabeth R. Dixon<sup>3</sup>, and Antonio Vallejo<sup>1</sup>

<sup>1</sup>ETSI Agrónomos, Technical University of Madrid, Ciudad Universitaria, 28040 Madrid, Spain

<sup>2</sup>School of Environmental Sciences, University of Guelph, Guelph, Ontario, N1G 2W1, Canada

<sup>3</sup>Rothamsted Research, North Wyke, Devon, EX20 2SB, UK

Correspondence to: Guillermo Guardia ([guillermo.guardia@upm.es](mailto:guillermo.guardia@upm.es))

Received: 27 January 2016 – Published in Biogeosciences Discuss.: 29 March 2016

Revised: 30 August 2016 – Accepted: 2 September 2016 – Published: 20 September 2016

**Abstract.** Agronomical and environmental benefits are associated with replacing winter fallow by cover crops (CCs). Yet, the effect of this practice on nitrous oxide (N<sub>2</sub>O) emissions remains poorly understood. In this context, a field experiment was carried out under Mediterranean conditions to evaluate the effect of replacing the traditional winter fallow (F) by vetch (*Vicia sativa* L.; V) or barley (*Hordeum vulgare* L.; B) on greenhouse gas (GHG) emissions during the intercrop and the maize (*Zea mays* L.) cropping period. The maize was fertilized following integrated soil fertility management (ISFM) criteria. Maize nitrogen (N) uptake, soil mineral N concentrations, soil temperature and moisture, dissolved organic carbon (DOC) and GHG fluxes were measured during the experiment. Our management (adjusted N synthetic rates due to ISFM) and pedo-climatic conditions resulted in low cumulative N<sub>2</sub>O emissions (0.57 to 0.75 kg N<sub>2</sub>O-N ha<sup>-1</sup> yr<sup>-1</sup>), yield-scaled N<sub>2</sub>O emissions (3–6 g N<sub>2</sub>O-N kg aboveground N uptake<sup>-1</sup>) and N surplus (31 to 56 kg N ha<sup>-1</sup>) for all treatments. Although CCs increased N<sub>2</sub>O emissions during the intercrop period compared to F (1.6 and 2.6 times in B and V, respectively), the ISFM resulted in similar cumulative emissions for the CCs and F at the end of the maize cropping period. The higher C : N ratio of the B residue led to a greater proportion of N<sub>2</sub>O losses from the synthetic fertilizer in these plots when compared to V. No significant differences were observed in CH<sub>4</sub> and CO<sub>2</sub> fluxes at the end of the experiment. This study shows that the use of both legume and nonlegume CCs combined with ISFM could provide, in addition to the advantages reported in previous studies, an opportunity to maximize agronomic

efficiency (lowering synthetic N requirements for the subsequent cash crop) without increasing cumulative or yield-scaled N<sub>2</sub>O losses.

## 1 Introduction

Improved resource-use efficiencies are pivotal components of sustainable agriculture that meets human needs and protects natural resources (Spiertz, 2010). Several strategies have been proposed to improve the efficiency of intensive irrigated systems, where nitrate (NO<sub>3</sub><sup>-</sup>) leaching losses are of major concern, during both cash crop and winter fallow periods (Quemada et al., 2013). In this sense, replacing winter intercrop fallow with cover crops (CCs) has been reported to decrease NO<sub>3</sub><sup>-</sup> leaching via retention of post-harvest surplus inorganic nitrogen (N) (Wagner-Riddle and Thurtell, 1998), consequently improving N use efficiency of the cropping system (Gabriel and Quemada, 2011). Furthermore, the use of CCs as green manure for the subsequent cash crop may further increase soil fertility and N use efficiency (Tonitto et al., 2006; Veenstra et al., 2007) through slow release of N and other nutrients from the crop residues, leading to a saving in synthetic fertilizer.

From an environmental point of view, N fertilization is closely related to the production and emission of nitrous oxide (N<sub>2</sub>O) (Davidson and Kanter, 2014), a greenhouse gas (GHG) with a molecular global warming potential ca. 300 times that of carbon dioxide (CO<sub>2</sub>) (IPCC, 2007). Nitrous oxide released from agricultural soils is mainly generated

by nitrification and denitrification processes, which are influenced by several soil variables (Firestone and Davidson, 1989). Thereby, modifying these parameters through agricultural management practices (e.g., fertilization, crop rotation, tillage or irrigation) aiming to optimize N inputs can lead to strategies for reducing the emission of (N<sub>2</sub>O) (Ussiri and Lal, 2013). In order to identify the most effective GHG mitigation strategies, side effects of methane (CH<sub>4</sub>) uptake and CO<sub>2</sub> emission (i.e., respiration) from soils, which are also influenced by agricultural practices (Snyder et al., 2009), need to be considered.

To date, the available information linking GHG emission and maize–winter CC rotation in the scientific literature is scarce. The most important knowledge gaps include effects of plant species selection and CC residue management (i.e., retention, incorporation or removal) (Basche et al., 2014). Cover crop species may affect N<sub>2</sub>O emissions in contrasting ways by influencing abiotic and biotic soil factors. These factors include mineral N availability in soil and the availability of carbon (C) sources for the denitrifier bacterial communities, soil pH, soil structure and microbial community composition (Abalos et al., 2014). For example, nonlegume CCs such as winter cereals could contribute to a reduction of N<sub>2</sub>O emissions due to their deep roots, which allow them to extract soil N more efficiently than legumes (Kallenbach et al., 2010). Conversely, it has been suggested that the higher C : N ratio of their residues as compared to those of legumes may provide energy (C) for denitrifiers, thereby leading to higher N<sub>2</sub>O losses in the presence of mineral N-NO<sub>3</sub><sup>-</sup> from fertilizers (Sarkodie-Addo et al., 2003). In this sense, the presence of cereal residues can increase the abundance of denitrifying microorganisms (Gao et al., 2016), thus enhancing denitrification losses when soil conditions are favorable (e.g., high NO<sub>3</sub><sup>-</sup> availability and soil moisture after rainfall or irrigation events, particularly in fine-textured soils) (Stehfest and Bouwman 2006; Baral et al., 2016). Furthermore, winter CCs can also abate indirect gaseous N losses through the reduction of leaching and subsequent emissions from water resources (Feyereisen et al., 2006). Thus, the estimated N<sub>2</sub>O mitigation potential for winter CCs ranges from 0.2 to 1.1 kg N<sub>2</sub>O ha<sup>-1</sup> yr<sup>-1</sup> according to Ussiri and Lal (2013).

In a CC–maize rotation system, mineral fertilizer application to the cash crop could have an important effect on N use efficiency and N losses from the agro-ecosystem. Different methods for calculating the N application rate (e.g., conventional or integrated) can be employed by farmers, affecting the amount of synthetic N applied to soil and the overall effect of CCs on N<sub>2</sub>O fluxes. Integrated soil fertility management (ISFM) (Kimani et al., 2003) provides an opportunity to optimize the use of available resources, thereby reducing pollution and costs from overuse of N fertilizers (conventional management). ISFM involves the use of inorganic fertilizers and organic inputs, such as green manure, and aims to maximize agronomic efficiency (Vanlauwe et al., 2011). When applying this technique to a CC–maize crop rotation,

the N fertilization rate for maize is calculated taking into account the background soil mineral N and the expected available N from mineralization of CC residues, which depends on residue composition. Differences in soil mineral N during the cash crop phase may be significantly reduced if ISFM practices are employed, affecting the GHG balance of the CC–cash crop cropping system.

Only one study has investigated the effect of CCs on N<sub>2</sub>O emissions in Mediterranean cropping systems (Sanz-Cobena et al., 2014). These authors found an effect of CC species on N<sub>2</sub>O emissions during the intercrop period. After 4 years of CC (vetch, barley or rape)–maize rotation, vetch was the only CC species that significantly enhanced N<sub>2</sub>O losses compared to fallow, mainly due to its capacity to fix atmospheric N<sub>2</sub> and because of higher N surplus from the previous cropping phases in these plots. In this study a conventional fertilization (same N synthetic rate for all treatments) was applied during the maize phase; how ISFM practices may affect these findings remains unknown. Moreover, the relative contribution of mineral N fertilizer, CC residues and/or soil mineral N to N<sub>2</sub>O losses during the cash crop has not been assessed yet. In this sense, stable isotope analysis (i.e., <sup>15</sup>N) represents a way to identify the source and the dominant processes involved in N<sub>2</sub>O production (Arah, 1997). Stable isotope techniques have been used in field studies evaluating N leaching and/or plant recovery in systems with cover crops (Bergström et al., 2001; Gabriel and Quemada, 2011; Gabriel et al., 2016). Furthermore, some laboratory studies have evaluated the effect of different crop residues on N<sub>2</sub>O losses using <sup>15</sup>N techniques (Baggs et al., 2003; Li et al., 2016), but to date, no previous studies have evaluated the relative contribution of cover crops (which include the aboveground biomass and the decomposition of root biomass) and N synthetic fertilizers to N<sub>2</sub>O emissions under field conditions. A comprehensive understanding of the N<sub>2</sub>O biochemical production pathways and nutrient sources is crucial for the development of effective mitigation strategies.

The objective of this study was to evaluate the effect of two different CC species (barley and vetch) and fallow on GHG emissions during the CC period and during the following maize cash crop period in an ISFM system. An additional objective was to study the contribution of the synthetic fertilizer and other N sources to N<sub>2</sub>O emissions using <sup>15</sup>N-labeled fertilizer. We hypothesized that (1) the presence of CCs instead of fallow would affect N<sub>2</sub>O losses, leading to higher emissions in the case of the legume CC (vetch) in accordance with the studies of Basche et al. (2014) and Sanz-Cobena et al. (2014) and (2) in spite of the ISFM during the maize period, which theoretically would lead to similar soil N availability for all plots, the distinct composition of the CC residues would affect N<sub>2</sub>O emissions. In order to test these hypotheses, a field experiment was carried out using the same management system for 8 years, measuring GHGs during the 8th year. To gain a better understanding of the effect of the management practices tested on the overall GHG budget of a

cropping system, CH<sub>4</sub>, CO<sub>2</sub> and yield-scaled N<sub>2</sub>O emissions were also analyzed during the experimental period. The relative contribution of each N source (synthetic fertilizer or soil endogenous N, including N mineralized from the CCs) to N<sub>2</sub>O emissions was also evaluated by <sup>15</sup>N-labeled ammonium nitrate in a parallel experiment.

## 2 Materials and methods

### 2.1 Site characteristics

The study was conducted at “La Chimenea” field station (40°03' N, 03° 31' W; 550 m a.s.l.), located in the central Tajo River basin near Aranjuez (Madrid, Spain), where an experiment involving cover-cropping systems and conservation tillage has been carried out since 2006. Soil at the field site is a silty clay loam (Typic Calcixerept; Soil Survey Staff, 2014). Some of the physicochemical properties of the top 0–10 cm soil layer, as measured by conventional methods, were as follows: pH<sub>H<sub>2</sub>O</sub>, 8.16; total organic C, 19.0 g kg<sup>-1</sup>; CaCO<sub>3</sub>, 198 g kg<sup>-1</sup>; clay, 25 %; silt, 49 %; and sand, 26 %. Bulk density of the topsoil layer determined in intact core samples (Grossman and Reinsch, 2002) was 1.46 g cm<sup>-3</sup>. Average ammonium (NH<sub>4</sub><sup>+</sup>) content at the beginning of the experiment was 0.42 ± 0.2 mg N kg soil<sup>-1</sup> (without differences between treatments). Nitrate concentrations were 1.5 ± 0.2 mg N kg soil<sup>-1</sup> in fallow and barley and 0.9 ± 0.1 mg N kg soil<sup>-1</sup> in vetch. Initial dissolved organic C (DOC) contents were 56.0 ± 7 mg C kg soil<sup>-1</sup> in vetch and fallow and 68.8 ± 5 mg C kg soil<sup>-1</sup> in barley. The area has a Mediterranean semiarid climate, with a mean annual air temperature of 14 °C. The coldest month is January, with a mean temperature of 6 °C, and the hottest month is August, with a mean temperature of 24 °C. During the last 30 years, the mean annual precipitation has been approximately 350 mm (17 mm from July to August and 131 mm from September to November).

Hourly rainfall and air temperature data were obtained from a meteorological station located at the field site (CR10X, Campbell Scientific Ltd., Shepshed, UK). A temperature probe inserted 10 cm into the soil was used to measure soil temperature. Mean hourly temperature data were stored on a data logger.

### 2.2 Experimental design and agronomic management

Twelve plots (12 m × 12 m) were randomly distributed in four replications of three cover-cropping treatments, including a cereal and a legume: (1) barley (B) (*Hordeum vulgare* L. ‘Vanessa’), (2) vetch (V) (*Vicia sativa* L. ‘Vereda’), and (3) traditional winter fallow (F). Cover crop seeds were broadcast by hand over the stubble of the previous crop and covered with a shallow cultivator (5 cm depth) on 10 October 2013, at a rate of 180 and 150 kg ha<sup>-1</sup> for B and V, respectively. The cover-cropping phase finished on 14 March 2014

following local practices, with an application of glyphosate (N-phosphonomethyl glycine) at a rate of 0.7 kg a.e. ha<sup>-1</sup>. Even though the safe use of glyphosate has been under discussion for many years (Chang and Delzell, 2016), it was used in order to preserve the same killing method in all the campaigns in this long-term experiment under conservation tillage management. All of the CC residues were left on top of the soil. Thereafter, a new set of N fertilizer treatments was set up for the maize cash crop phase. Maize (*Zea mays* L., Pioneer P1574, FAO Class 700) was directly drilled on 7 April 2014 in all plots, resulting in a plant population density of 7.5 plants m<sup>-2</sup>; harvesting took place on 25 September 2014. The fertilizer treatments consisted of ammonium nitrate applied on 2 June at three rates: 170, 140 and 190 kg N ha<sup>-1</sup> in F, V and B plots, respectively, according to ISFM practices. For the calculation of each N rate, the N available in the soil (which was calculated following soil analysis as described below), the expected N uptake by maize crop, and the estimated N mineralized from V and B residues were taken into account, assuming that crop requirements were 236.3 kg N ha<sup>-1</sup> (Quemada et al., 2014). Estimated N use efficiency of maize plants for calculating N application rate was 70 % according to the N use efficiency obtained during the previous years in the same experimental area. Each plot received P as triple superphosphate (45 % P<sub>2</sub>O<sub>5</sub>, Fertiberia<sup>®</sup>, Madrid, Spain) at a rate of 69 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup>, and K as potassium chloride (60 % K<sub>2</sub>O, Fertiberia<sup>®</sup>, Madrid, Spain) at a rate of 120 kg K<sub>2</sub>O ha<sup>-1</sup> just before sowing maize. All N, P and K fertilizers were broadcast by hand, and immediately after N fertilization the field was irrigated to prevent ammonia volatilization. The main crop previous to sowing CCs was sunflower (*Helianthus annuus* L. ‘Sambro’). Neither the sunflower nor the CCs were fertilized.

In order to determine the amount of N<sub>2</sub>O derived from the N fertilizers, double-labeled ammonium nitrate (<sup>15</sup>NH<sub>4</sub><sup>+</sup><sup>15</sup>NO<sub>3</sub><sup>-</sup>, 5 at. % <sup>15</sup>N, from Cambridge Isotope Laboratories, Inc., Massachusetts, USA) was applied on 2 m × 2 m subplots established within each plot at a rate of 130 kg N ha<sup>-1</sup>. In order to reduce biases due to the use of different N rates (e.g., apparent priming effects or different mixing ratios between the added and resident soil N pools) the same amount of N was applied for all treatments. In each subplot, the CC residue was also left on top of the soil. This application took place on 26 May by spreading the fertilizer homogeneously with a hand sprayer, followed by an irrigation event.

Sprinkler irrigation was applied to the maize crop at a total amount of 688.5 mm in 31 irrigation events. Sprinklers were installed in a 12 m × 12 m framework. The water doses to be applied were estimated from the crop evapotranspiration (ET<sub>c</sub>) of the previous week (net water requirements). This was calculated daily as ET<sub>c</sub> = K<sub>c</sub> × ET<sub>o</sub>, where ET<sub>o</sub> is reference evapotranspiration calculated by the FAO Penman–Monteith method (Allen et al., 1998) using data from the meteorological station located in the experimental field. The

crop coefficient ( $K_c$ ) was obtained using the relationship for maize in semiarid conditions (Martínez-Cob, 2008).

Two different periods were considered for data reporting and analysis: Period I (from CC sowing to N fertilization of the maize crop) and Period II (from N fertilization of maize to the end of the experimental period, after maize harvest).

### 2.3 GHG emissions sampling and analyzing

Fluxes of  $N_2O$ ,  $CH_4$  and  $CO_2$  were measured from October 2013 to October 2014 using opaque, manually operated circular static chambers as described in detail by Abalos et al. (2013). One chamber (diameter 35.6 cm, height 19.3 cm) was located in each experimental plot. The chambers were hermetically closed (for 1 h) by fitting them into stainless steel rings, which were inserted at the beginning of the study into the soil to a depth of 5 cm in order to minimize the lateral diffusion of gases and to avoid the soil disturbance associated with the insertion of the chambers in the soil. The rings were only removed during management events. Each chamber had rubber sealing tape to guarantee an airtight seal between the chamber and the ring and was covered with a radiant barrier reflective foil to reduce temperature gradients between inside and outside. A rubber stopper with a three-way stopcock was placed in the wall of each chamber to take gas samples. Greenhouse gas measurements were always made with barley/vetch plants inside the chamber. During the maize period, gas chambers were set up between maize rows.

During Period I, GHGs were sampled weekly or every 2 weeks. During the first month after maize fertilization, gas samples were taken twice per week. Afterwards, gas sampling was performed weekly or fortnightly, until the end of the cropping period. To minimize any effects of diurnal variation in emissions, samples were always taken at the same time of day (10:00–12:00), which is reported as a representative time (Reeves and Wang, 2015).

Measurements of  $N_2O$ ,  $CO_2$  and  $CH_4$  emissions were made at 0, 30 and 60 min to test the linearity of gas accumulation in each chamber. Gas samples (100 mL) were removed from the headspace of each chamber by syringe and transferred to 20 mL gas vials sealed with a gastight neoprene septum. The vials were previously flushed in the field using 80 mL of the gas sample. Samples were analyzed by gas chromatography using a HP-6890 gas chromatograph equipped with a headspace autoanalyzer (HT3), both from Agilent Technologies (Barcelona, Spain). Inert gases were separated by HP Plot-Q capillary columns. The gas chromatograph was equipped with a  $^{63}Ni$  electron-capture detector (micro-ECD) to analyze  $N_2O$  concentrations, and with a flame ionization detector (FID) connected to a methanizer to measure  $CH_4$  and  $CO_2$  (previously reduced to  $CH_4$ ). The temperatures of the injector, oven and ECD were 50, 50 and 350 °C, respectively. The accuracy of the gas chromatographic data was 1 % or better. Two gas standards comprising a mixture of gases (high standard with  $1500 \pm 7.50$  ppm

$CO_2$ ,  $10 \pm 0.25$  ppm  $CH_4$  and  $2 \pm 0.05$  ppm  $N_2O$  and low standard with  $200 \pm 1.00$  ppm  $CO_2$ ,  $2 \pm 0.10$  ppm  $CH_4$  and  $200 \pm 6.00$  ppb  $N_2O$ ) were provided by Carbueros Metálicos S.A. and Air Products SA/NV, respectively, and used to determine a standard curve for each gas. The response of the GC was linear within 200–1500 ppm for  $CO_2$  and 2–10 ppm  $CH_4$  and quadratic within 200–2000 ppb for  $N_2O$ .

The increases in  $N_2O$ ,  $CH_4$  and  $CO_2$  concentrations within the chamber headspace were generally (80 % of cases) linear ( $R^2 > 0.90$ ) during the sampling period (1 h). Therefore, emission rates of fluxes were estimated as the slope of the linear regression between concentration and time (after corrections for temperature) and from the ratio between chamber volume and soil surface area (MacKenzie et al., 1998). Cumulative  $N_2O$ ,  $CH_4$  and  $CO_2$  emissions per plot during the sampling period were estimated by linear interpolations between sampling dates, multiplying the mean flux of two successive determinations by the length of the period between sampling and adding that amount to the previous cumulative total (Sanz-Cobena et al., 2014). The measurement of  $CO_2$  emissions from soil, including plants in opaque chambers, only includes ecosystem respiration and not photosynthesis (Meijide et al., 2010).

### 2.4 $^{15}N$ isotope analysis

Gas samples from the subplots receiving double-labeled AN fertilizer were taken after 60 min of static chamber closure 1, 4, 9, 11, 15, 18, 22 and 25 days after fertilizer application. Stable  $^{15}N$  isotope analysis of  $N_2O$  contained in the gas samples was carried out on a cryo-focusing gas chromatography unit coupled to a 20/20 isotope ratio mass spectrometer (both from SerCon Ltd., Crewe, UK). Ambient samples were taken occasionally as required for the subsequent isotopic calculations. Solutions of 6.6 and 2.9 at. % ammonium sulfate  $[(NH_4)_2SO_4]$  were prepared and used to generate 6.6 and 2.9 at. %  $N_2O$  (Laughlin et al., 1997), which were used as reference and quality control standards. In order to calculate the atom percent excess (APE) of the  $N_2O$  emitted in the subplots, the mean natural abundance of atmospheric  $N_2O$  from the ambient samples (0.369 at. %  $^{15}N$ ) was subtracted from the measured enriched gas samples. To obtain the  $N_2O$  flux that was derived from fertilizer ( $N_2O - N_{diff}$ ), the following equation was used (Senbayram et al., 2009):

$$N_2O - N_{diff} = N_2O - N \times \left( \frac{N_2O\_APE_{sample}}{APE_{fertilizer}} \right), \quad (1)$$

in which “ $N_2O - N$ ” is the  $N_2O$  emission from soil, “ $N_2O\_APE_{sample}$ ” is the  $^{15}N$  at. % excess of emitted  $N_2O$ , and “ $APE_{fertilizer}$ ” is the  $^{15}N$  at. % excess of the applied fertilizer (Senbayram et al., 2009).

### 2.5 Soil and crop analyses

In order to relate gas emissions to soil properties, soil samples were collected at 0–10 cm depth during the growing sea-

son on almost all gas-sampling occasions, particularly after each fertilization event. Three soil cores (2.5 cm diameter and 15 cm length) were randomly sampled close to the ring in each plot, and then mixed and homogenized in the laboratory. Soil  $\text{NH}_4^+$  and  $\text{NO}_3^-$  concentrations were analyzed using 8 g of soil extracted with 50 mL of KCl (1 M), and measured by automated colorimetric determination using a flow injection analyzer (FIAS 400 Perkin Elmer) provided with a UV-visible spectrophotometer detector. Soil (DOC) was determined by extracting 8 g of homogeneously mixed soil with 50 mL of deionized water (and subsequently filtered) and analyzed with a total organic C analyzer (multi N/C 3100 Analytik Jena) equipped with an IR detector. The water-filled pore space (WFPS) was calculated by dividing the volumetric water content by total soil porosity. Total soil porosity was calculated according to the following relationship: soil porosity =  $(1 - \text{soil bulk density}/2.65)$ , assuming a particle density of  $2.65 \text{ g cm}^{-3}$  (Danielson and Sutherland, 1986). Gravimetric water content was determined by oven-drying soil samples at  $105^\circ\text{C}$  with a Sartorius® MA30.

Four  $0.5 \text{ m} \times 0.5 \text{ m}$  squares were randomly harvested from each plot before killing the CC by applying glyphosate. Aerial biomass was cut by hand at soil level, dried, weighed and ground. A subsample was taken for determination of total N content. From these samples the CC biomass and N contribution to the subsequent maize were determined.

At maize harvest, two 8 m central rows in each plot were collected and weighed in the field following separation of grain and straw. For aboveground N uptake calculations, N content was determined in subsamples of grain and biomass. Total N content of maize and CC subsamples was determined with an elemental analyzer (TruMac CN, Leco).

## 2.6 Calculations and statistical analysis

Yield-scaled  $\text{N}_2\text{O}$  emissions and N surplus in the maize cash crop were calculated as the amount of  $\text{N}_2\text{O}$  emitted (considering the emissions of the whole experiment, i.e., Period I and Period II) per unit of aboveground N uptake and taking the difference between N application and aboveground N uptake, respectively (van Groenigen et al., 2010).

Statistical analyses were carried out with Statgraphics Plus 5.1. Analyses of variance were performed for all variables during the experiment (except climatic ones), for both periods indicated in Sect. 2.2. Data distribution normality and variance uniformity were previously assessed by the Shapiro–Wilk test and Levene’s statistic, respectively, and transformed ( $\log_{10}$ , root square, arcsin or inverse) before analysis when necessary. Means of soil parameters were separated by Tukey’s honest significance test at  $P < 0.05$ , while cumulative GHG emissions, yield-scaled  $\text{N}_2\text{O}$  emissions and N surplus were compared by the orthogonal contrasts method at  $P < 0.05$ . For non-normally distributed data, the Kruskal–Wallis test was used on non-transformed data to evaluate differences at  $P < 0.05$ . Linear correlations were carried out to

determine relationships between gas fluxes and WFPS, soil temperature, DOC,  $\text{NH}_4^+$  and  $\text{NO}_3^-$ . These analyses were performed using the mean/cumulative data of the replicates of the CC treatments ( $n = 12$ ), and also for all the dates when soil and GHG were sampled, for Period I ( $n = 16$ ), Period II ( $n = 11$ ) and the whole experimental period ( $n = 27$ ).

## 3 Results

### 3.1 Cover crop (Period I)

#### 3.1.1 Environmental conditions and WFPS

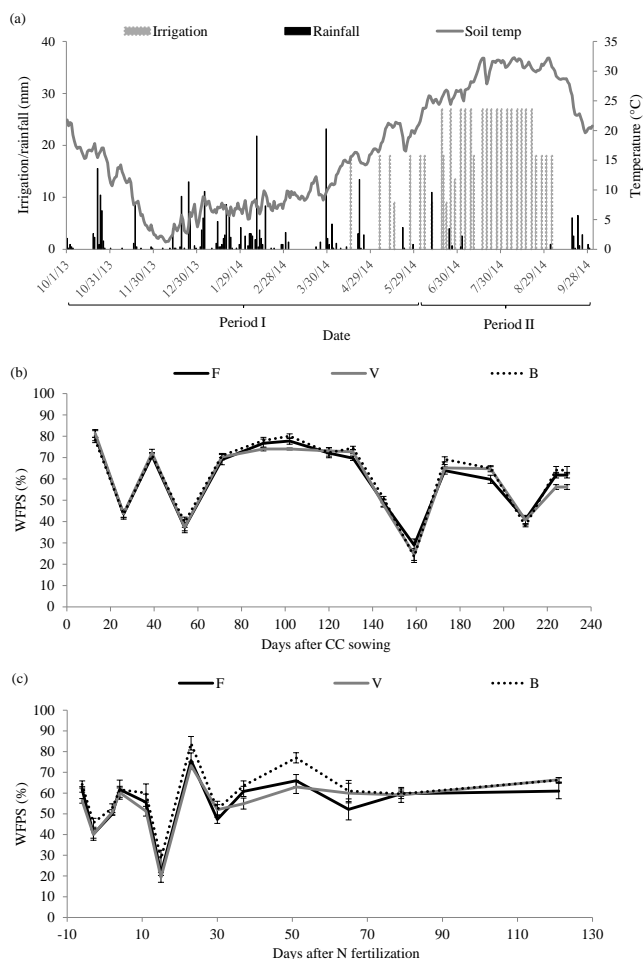
Mean soil temperature during the intercrop period was  $8.8^\circ\text{C}$ , ranging from  $1.8^\circ\text{C}$  (December) to  $15.5^\circ\text{C}$  (April) (Fig. 1a), which were typical values in the experimental area. Mean soil temperature during maize cropping period was  $24.6^\circ\text{C}$ , which was also a standard value for this region. The accumulated rainfall during this period was 215 mm, whereas the 30-year mean is 253 mm. WFPS ranged from 40 to 81 % (Fig. 1b). No significant differences were observed for WFPS mean values between the different treatments ( $P > 0.05$ ).

#### 3.1.2 Mineral N and DOC and cover crop residues

Topsoil  $\text{NH}_4^+$  content was below  $5 \text{ mg N kg soil}^{-1}$  most of the time in Period I, although a peak was observed after maize sowing (55 days after CC kill date) (Fig. 2a), with the highest values reached in B ( $50 \text{ mg N kg soil}^{-1}$ ). Mean  $\text{NH}_4^+$  content was significantly higher in B than in F ( $P < 0.05$ ), but daily  $\text{NH}_4^+$  concentrations between treatments were only significantly different between treatments on one sampling date (210 days after CC sowing). Nitrate content increased after CC killing, reaching values above  $25 \text{ mg N kg soil}^{-1}$  in the V treatment (Fig. 2c). Mean  $\text{NO}_3^-$  content during Period I was significantly higher in the V plots than in the B and F plots ( $P < 0.001$ ). Dissolved organic C ranged from 60 to  $130 \text{ mg C kg soil}^{-1}$  (Fig. 2e). Average topsoil DOC content was significantly higher in B than in V and F (10 and 12 %, respectively,  $P < 0.01$ ) but differences were only observed on some sampling dates. The total amount of cover crop biomass left on the ground was  $540.5 \pm 26.5$  and  $1106.7 \pm 93.6 \text{ kg dry matter ha}^{-1}$  in B and V, respectively. Accordingly, the total N content of these residues was  $11.0 \pm 0.6$  and  $41.3 \pm 4.5 \text{ kg N ha}^{-1}$  in B and V, respectively.

#### 3.1.3 GHG fluxes

Nitrous oxide fluxes ranged from  $-0.06$  to  $0.22 \text{ mg N m}^{-2} \text{ d}^{-1}$  (Fig. 3a) in Period I. The soil acted as a sink for  $\text{N}_2\text{O}$  at some sampling dates, especially for the F plots. Cumulative fluxes at the end of Period I were significantly greater in CC treatments compared to F (1.6 and 2.6 higher in B and V, respectively) ( $P < 0.05$ ; Table 1). Net  $\text{CH}_4$



**Figure 1.** Daily mean soil temperature (°C) rainfall and irrigation (mm) (a) and soil WFPS (%) in the three cover crop (CC) treatments (fallow, F; vetch, V; and barley, B) during Period I (b) and II (c). Vertical lines indicate standard errors.

uptake was observed in all intercrop treatments, and daily fluxes ranged from  $-0.60$  to  $0.25 \text{ mg C m}^{-2} \text{ d}^{-1}$  (data not shown). No significant differences were observed between treatments in cumulative  $\text{CH}_4$  fluxes at the end of Period I ( $P > 0.05$ ; Table 1). Carbon dioxide fluxes (data not shown) remained below  $1 \text{ g C m}^{-2} \text{ d}^{-1}$  during the intercrop period. The greatest fluxes were observed in B, although differences in cumulative fluxes were not significant ( $P > 0.05$ ; Table 1) in the whole intercrop period, but soil respiration was increased in B, with respect to F, from mid-February to the end of Period I. Nitrous oxide emissions were significantly correlated to  $\text{CO}_2$  fluxes ( $P < 0.01$ ,  $n = 17$ ,  $r = 0.69$ ) and soil temperature ( $P < 0.05$ ,  $n = 17$ ,  $r = 0.55$ ).

## 3.2 Maize crop (Period II)

### 3.2.1 Environmental conditions and WFPS

Mean soil temperature ranged from  $19.6$  (reached in September) to  $32.3$  °C (reached in August) with a mean value of  $27.9$  °C (Fig. 1a). Total rainfall during the maize crop period was  $57 \text{ mm}$ . WFPS ranged from  $19$  to  $84$  % (Fig. 1c). Higher mean WFPS values ( $P < 0.01$ ) were measured in B during some sampling dates.

### 3.2.2 Mineral N and DOC

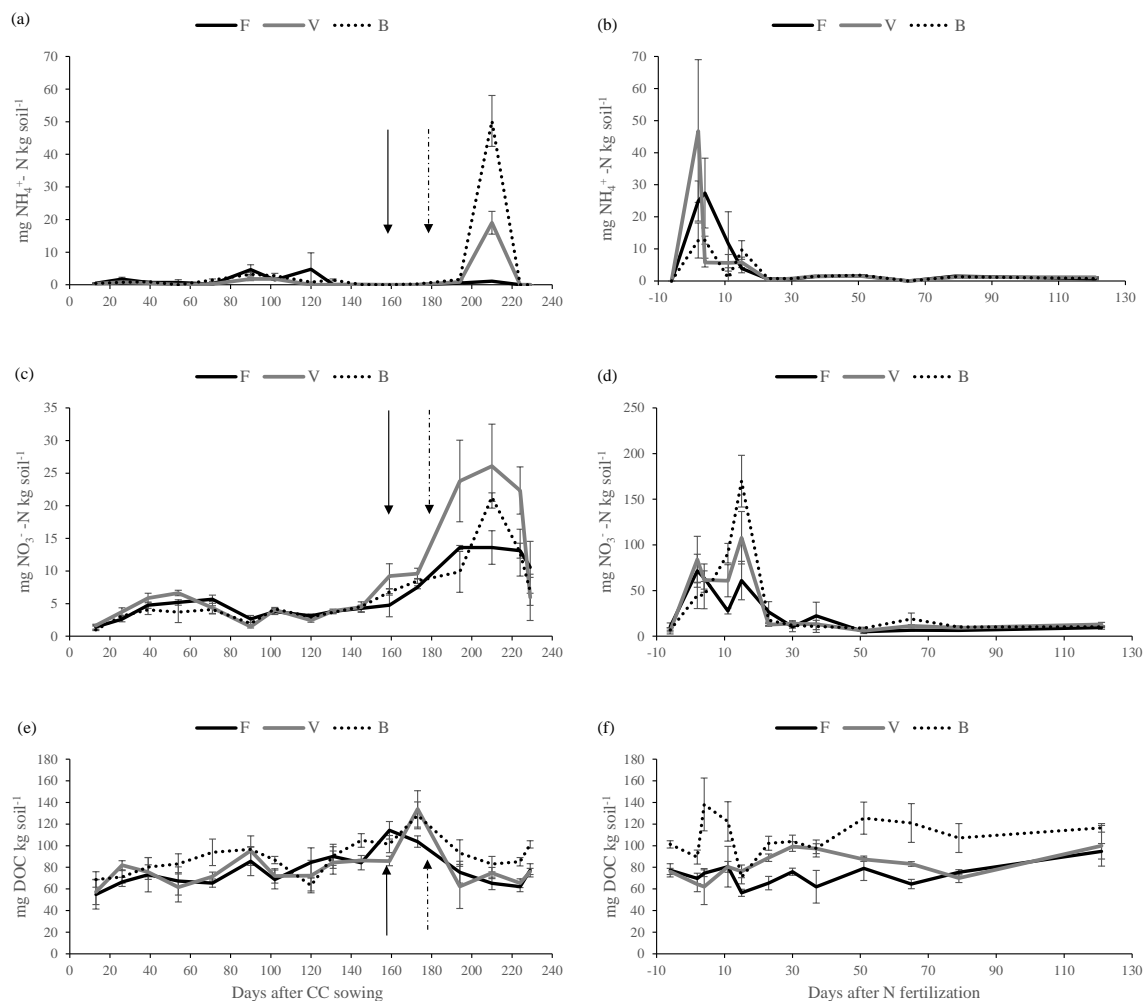
Topsoil  $\text{NH}_4^+$  content increased rapidly after N fertilization (Fig. 2b), decreasing to values below  $10 \text{ mg N kg soil}^{-1}$  from 15 days after fertilization to the end of the experimental period. Nitrate concentrations (Fig. 2d) also peaked after AN addition, reaching the highest value ( $170 \text{ mg N kg soil}^{-1}$ ) 15 days after fertilization in B ( $P < 0.05$ ). No significant differences ( $P > 0.05$ ) between treatments were observed in average soil  $\text{NH}_4^+$  or  $\text{NO}_3^-$  during the maize phase. Dissolved organic C ranged from  $56$  to  $138 \text{ mg C kg soil}^{-1}$  (Fig. 2f). Average topsoil DOC content was  $26$  and  $44$  % higher in B than in V and F, respectively ( $P < 0.001$ ).

### 3.2.3 GHG fluxes, yield-scaled $\text{N}_2\text{O}$ emissions and N surplus

Nitrous oxide fluxes ranged from  $0.0$  to  $5.6 \text{ mg N m}^{-2} \text{ d}^{-1}$  (Fig. 3b). The highest  $\text{N}_2\text{O}$  emission peak was observed 1–4 days after fertilization for all plots. Other peaks were subsequently observed until 25 days after fertilization, particularly in B plots, where  $\text{N}_2\text{O}$  emissions 23 and 25 days after fertilization were higher ( $P < 0.05$ ) than those of F and V (Fig. 3b). No significant differences in cumulative  $\text{N}_2\text{O}$  fluxes were observed between treatments throughout or at the end of the maize crop period (Table 1), although fluxes were numerically higher in B than in V ( $0.05 < P < 0.10$ ). Daily  $\text{N}_2\text{O}$  emissions were significantly correlated with  $\text{NH}_4^+$  topsoil content ( $P < 0.05$ ,  $n = 12$ ,  $r = 0.84$ ).

As in the previous period, all treatments were  $\text{CH}_4$  sinks, without significant differences between treatments ( $P > 0.05$ ; Table 1). Respiration rates ranged from  $0.15$  to  $3.0 \text{ g C m}^{-2} \text{ d}^{-1}$ ; no significant differences ( $P > 0.05$ ; Table 1) were observed among the  $\text{CO}_2$  values for the different treatments. Yield-scaled  $\text{N}_2\text{O}$  emissions and N surplus are shown in Table 1. No significant differences were observed between treatments, although these values were generally lower in V than in B ( $0.05 < P < 0.15$ ).

Considering the whole cropping period (Period I and Period II),  $\text{N}_2\text{O}$  fluxes significantly correlated with WFPS ( $P < 0.05$ ,  $n = 12$ ,  $r = 0.61$ )  $\text{NH}_4^+$  ( $P < 0.05$ ,  $n = 27$ ,  $r = 0.84$ ) and  $\text{NO}_3^-$  ( $P < 0.05$ ,  $n = 27$ ,  $r = 0.50$ ).



**Figure 2.** (a, b)  $\text{NH}_4^+$ -N, (c, d)  $\text{NO}_3^-$ -N and (e, f) DOC concentrations in the 0–10 cm soil layer for the three CC treatments (fallow, F; vetch, V; and barley, B) during both cropping periods. The black arrows indicate the time of spraying glyphosate over the CCs. The dotted arrows indicate the time of maize sowing. Vertical lines indicate standard errors.

### 3.2.4 Fertilizer-derived $\text{N}_2\text{O}$ emissions

The proportion (%) of  $\text{N}_2\text{O}$  losses from ammonium nitrate, calculated by isotopic analyses, is represented in Fig. 4. The highest percentages of  $\text{N}_2\text{O}$  fluxes derived from the synthetic fertilizer were observed 1 day after fertilization, ranging from 34 % (V) to 67 % (B). On average, almost 50 % of  $\text{N}_2\text{O}$  emissions in the first sampling event after N synthetic fertilization came from other sources (i.e., soil endogenous N, including N mineralized from the CCs). The mean percentage of  $\text{N}_2\text{O}$  losses from synthetic fertilizer throughout all sampling dates was 2.5 times higher in B compared to V ( $P < 0.05$ ) and was positively correlated with DOC concentrations ( $P < 0.05$ ,  $n = 12$ ,  $r = 0.71$ ). There were no significant differences between V and F ( $P > 0.05$ ).

## 4 Discussion

### 4.1 Role of CCs in $\text{N}_2\text{O}$ emissions: Period I

Cover crop treatments (V and B) increased  $\text{N}_2\text{O}$  losses compared to F, especially in the case of V (Table 1). These results are consistent with the meta-analysis of Basche et al. (2014), which showed that, overall, CCs increase  $\text{N}_2\text{O}$  fluxes (compared to bare fallow), with highly significant increments in the case of legumes and a lower effect in the case of non-legume CCs. In the same experimental area, Sanz-Cobena et al. (2014) found that V was the only CC significantly affecting  $\text{N}_2\text{O}$  emissions. The greatest differences between treatments were observed at the beginning (13–40 days after CC sowing) and at the end of this period (229 days after CC sowing) (Fig. 3a). On these dates, the mild soil temperatures and the relatively high moisture content were more suitable for soil biochemical processes, which may trigger

**Table 1.** Total cumulative N<sub>2</sub>O-N, CH<sub>4</sub>-C and CO<sub>2</sub>-C fluxes; N surplus; and yield-scaled N<sub>2</sub>O emissions in the three CC treatments (fallow, F; vetch, V; and barley, B) at the end of both cropping periods. *P* values were calculated with Student's *t* test and *df* = 9.

Treatment		N <sub>2</sub> O kg N <sub>2</sub> O-N ha <sup>-1</sup>	CH <sub>4</sub> kg CH <sub>4</sub> -C ha <sup>-1</sup>	CO <sub>2</sub> kg CO <sub>2</sub> -C ha <sup>-1</sup>	Surplus kg N ha <sup>-1</sup>	Yield-scaled N <sub>2</sub> O emissions g N <sub>2</sub> O-N kg aboveground N uptake <sup>-1</sup>
	F	0.05	-0.30	443.02		
	V	0.13	-0.28	463.01		
	B	0.08	-0.24	582.13		
	SE	0.03	0.07	46.33		
End of Period I	Estimate	-11.48	-11.45	-134.37		
	F vs. CCs <i>t</i> test	-2.5	-0.61	-1.00		
	<i>P</i> value	0.03*	0.56	0.34		
	Estimate	5.29	-6.23	-127.50		
	V vs. B <i>t</i> test	1.99	-0.57	-1.64		
	<i>P</i> value	0.08	0.58	0.14		
	F	0.57	-0.46	2595.07	31.47	4.21
	V	0.48	-0.33	2778.84	13.72	3.06
	B	0.74	-0.35	2372.07	55.94	5.64
	SE	0.10	0.08	177.35	15.30	0.85
End of Period II	Estimate	-7.46	-23.69	83.36	-3.16	-0.12
	F vs. CCs <i>t</i> test	-0.30	-1.25	0.19	-0.08	-0.14
	<i>P</i> value	0.77	0.24	0.86	0.94	0.89
	Estimate	-26.59	2.08	417.8	-38.67	-2.59
	V vs. B <i>t</i> test	-1.90	0.19	1.62	-1.79	-2.16
	<i>P</i> value	0.09	0.85	0.14	0.11	0.06

\* and SE denote significant at *P* < 0.05 and the standard error of the mean, respectively.

N<sub>2</sub>O emissions (Fig. 1a, b) (Firestone and Davidson, 1989). Average topsoil NO<sub>3</sub><sup>-</sup> was significantly higher in V (Fig. 2b), which was the treatment that led to the highest N<sub>2</sub>O emissions. Legumes such as V are capable of biologically fixing atmospheric N<sub>2</sub>, thereby increasing soil NO<sub>3</sub><sup>-</sup> content with the potential to be denitrified. Furthermore, the mineralization of the most recalcitrant fraction of the previous V residue (which supplies nearly 4 times more N than the B residue, as indicated in Sect. 3.1.2) together with high C-content sunflower residue could also explain higher NO<sub>3</sub><sup>-</sup> contents in V plots (Frimpong et al., 2011) and higher N<sub>2</sub>O losses from denitrification (Baggs et al., 2000). After the CC kill date, N release from decomposition of roots and nodules and faster mineralization of V residue compared to that of B (shown by NO<sub>3</sub><sup>-</sup> in soil in Fig. 2c) are the most plausible explanations for the N<sub>2</sub>O increases at the end of the intercrop period (Fig. 3a) (Rochette and Janzen, 2005; Wichern et al., 2008).

Some studies (e.g., Justes et al., 1999; Nemecek et al., 2008) have pointed out that N<sub>2</sub>O losses can be reduced with the use of CCs, due to the extraction of plant-available N unused by previous cash crop. However, in our study lower N<sub>2</sub>O emissions were measured from F plots without CCs during the intercrop period. This may be a consequence of higher NO<sub>3</sub><sup>-</sup> leaching in F plots (Gabriel et al., 2012; Quemada et al., 2013), limiting the availability of the substrate for denitrification. Frequent rainfall during the intercrop period (Fig. 1a) and the absence of N uptake by CCs may have led to N losses

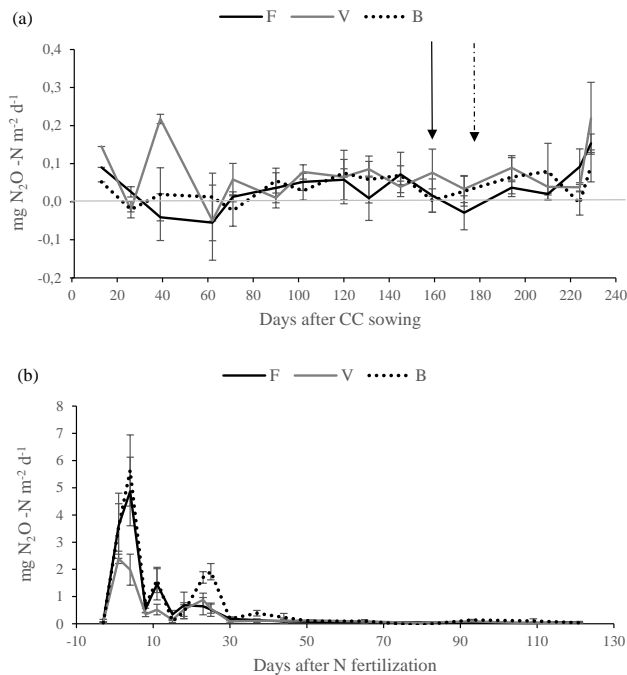
through leaching, resulting in low concentrations of soil mineral N in F plots.

Nitrous oxide emissions were low during this period but in the range of those reported by Sanz-Cobena et al. (2014) in the same experimental area. Total emissions during Period I represented 8, 10 and 21 % of total cumulative emissions in F, B and V, respectively (Table 1). The absence of N fertilizer application to the soil combined with the low soil temperatures during winter – which were far from the optimum values for nitrification and denitrification (25–30 °C) processes (Ussiri and Lal, 2013) – may have caused these low N<sub>2</sub>O fluxes. The significant positive correlation between soil temperature and N<sub>2</sub>O fluxes during this period highlights the key role of this parameter as a driver of soil emissions (Schindlbacher et al., 2004; García-Marco et al., 2014).

#### 4.2 Role of CCs in N<sub>2</sub>O emissions: Period II

Isotopic analysis during Period II, in which ISFM was carried out, showed that a significant proportion of N<sub>2</sub>O emissions came from endogenous soil N or the mineralization of crop residues, especially after the first few days following N fertilization (Fig. 4). In this sense, even though an interaction between crop residue and N fertilizer application has been previously described (e.g., in Abalos et al., 2013), the similar proportion of N<sub>2</sub>O losses coming from fertilizer in B and F (without residue) 1 day after N fertilization revealed the importance of soil mineral N contained in the micropores for

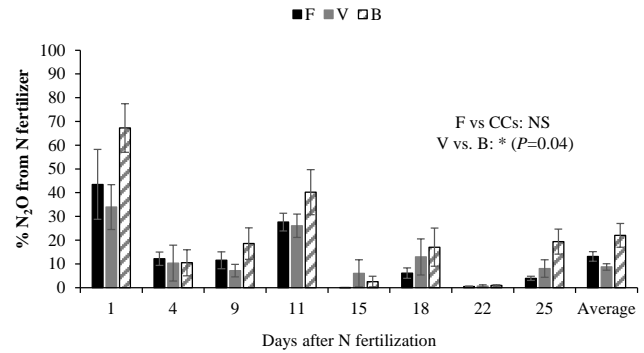




**Figure 3.**  $\text{N}_2\text{O}$  emissions for the three CC treatments (fallow, F; vetch, V; and barley, B) during Period I (a) and II (b). The black arrows indicate the time of spraying glyphosate over the CCs. The dotted arrows indicate the time of maize sowing. Vertical lines indicate standard errors.

the  $\text{N}_2\text{O}$  bursts after the first irrigation events, with respect to the N released from CC residues.

As we hypothesized, the different CCs played a key role in the  $\text{N}_2\text{O}$  emissions during Period II. Barley plots had higher  $\text{N}_2\text{O}$  emissions than fallow or V-residue plots (at the 10% significance level; Table 1). Further, a higher proportion of  $\text{N}_2\text{O}$  emissions was derived from the fertilizer in B-residue than in V-residue plots (Fig. 4). These results are in agreement with those of Baggs et al. (2003), who reported a higher percentage of  $\text{N}_2\text{O}$  derived from the  $^{15}\text{N}$ -labeled fertilizer using a cereal (ryegrass) as surface mulching instead of a legume (bean), in a field trial with zero-tillage management. The differences between B and V in terms of cumulative  $\text{N}_2\text{O}$  emissions and in the relative contribution of each source to these emissions (fertilizer- or soil-N) could be explained by: (i) the higher C : N residue of B ( $20.7 \pm 0.7$  while that of V was  $11.1 \pm 0.1$ , according to Alonso-Ayuso et al., 2014) may have provided an energy source for denitrification (Sarkodie-Addo et al., 2003), favoring the reduction of the  $\text{NO}_3^-$  supplied by the synthetic fertilizer and enhancing  $\text{N}_2\text{O}$  emissions, as supported by the positive correlation of DOC with the proportion of  $\text{N}_2\text{O}$  coming from the synthetic fertilizer; (ii)  $\text{NO}_3^-$  concentrations, which tended to be higher in B during the maize cropping phase, could have led to incomplete denitrification and larger  $\text{N}_2\text{O}/\text{N}_2$  ratios (Yamulki and Jarvis, 2002); (iii) the easily mineralizable V



**Figure 4.** Proportion of  $\text{N}_2\text{O}$  losses (%) that come from N synthetic fertilizer during Period II, for the three CC treatments (fallow, F; vetch, V; and barley, B). Vertical lines indicate standard errors. “NS” and \* denote not significant and significant at  $P < 0.05$ , respectively.

residue (with low C : N ratio) provided an additional N source for soil microorganisms, thus decreasing the relative amount of  $\text{N}_2\text{O}$  derived from the synthetic fertilizer (Baggs et al., 2000; Shan and Yan, 2013); and (iv) V plots were fertilized with a lower amount of immediately available N (i.e., ammonium nitrate) than B plots, which could have resulted in better synchronization between N release and crop needs (Ussiri and Lal, 2013) in V plots. Supporting these findings, Bayer et al. (2015) recently concluded that partially supplying the maize N requirements with winter legume cover crops can be considered a  $\text{N}_2\text{O}$  mitigation strategy in subtropical agro-ecosystems.

The mineralization of B residues resulted in higher DOC contents for these plots compared to the F or V plots ( $P < 0.001$ ). This was observed in both Period I (as a consequence of soil C changes after the 8-year cover-cropping management) and Period II (due to the CC decomposition). Although in the present study the correlation between DOC and  $\text{N}_2\text{O}$  emissions was not significant, positive correlations have been previously found in other low-C Mediterranean soils (e.g., Vallejo et al., 2006; López-Fernández et al., 2007). Some authors have suggested that residues with a high C : N ratio can induce microbial N immobilization (Frimpong and Baggs, 2010; Dendooven et al., 2012). In our experiment, a  $\text{N}_2\text{O}$  peak was observed in B plots 20–25 days after fertilization (Fig. 3b) after a remarkable increase of  $\text{NO}_3^-$  content (Fig. 2d), which may be a result of a remineralization of previously immobilized N in these plots.

The positive correlation of  $\text{N}_2\text{O}$  fluxes and soil  $\text{NO}_3^-$  content and WFPS during the whole cycle further supports the importance of denitrification process for explaining  $\text{N}_2\text{O}$  losses in this agro-ecosystem (Davidson et al., 1991; García-Marco et al., 2014). However, the strong positive correlation of  $\text{N}_2\text{O}$  with  $\text{NH}_4^+$  indicated that nitrification was also a major process leading to  $\text{N}_2\text{O}$  fluxes, and showed that the continuous drying-wetting cycles during a summer irri-

gated maize crop in a semiarid region can lead to favorable WFPS conditions for both nitrification and denitrification processes (Fig. 1c) (Bateman and Baggs, 2005). Emission factors ranged from 0.2 to 0.6 % of the synthetic N applied, which were lower than the IPCC default value of 1 %. As explained above, ecological conditions during the intercrop period (rainfall and temperature) and maize phase (temperature) could be considered normal (based on the 30-year average) in Mediterranean areas. Aguilera et al. (2013) obtained a higher emission factor for high (1.01 %) and low (0.66 %) water-irrigation conditions in a meta-analysis of Mediterranean cropping systems. We hypothesized that management practices may have contributed to these low emissions, but other inherent factors such as soil pH should also be considered. Indeed, a higher  $\text{N}_2\text{O}/\text{N}_2$  ratio has been associated with acidic soils, so lower  $\text{N}_2\text{O}$  emissions from denitrification could be expected in alkaline soils (Mørkved et al., 2007; Baggs et al., 2010).

#### 4.3 Methane and $\text{CO}_2$ emissions

As is generally found in non-flooded arable soils, all treatments were net  $\text{CH}_4$  sinks (Snyder et al., 2009). No significant differences were observed between treatments in any of the two periods (Table 1), which is similar to the pattern observed by Sanz-Cobena et al. (2014). Some authors (Dunfield and Knowles, 1995; Tate, 2015) have suggested an inhibitory effect of soil  $\text{NH}_4^+$  on  $\text{CH}_4$  uptake. Low  $\text{NH}_4^+$  contents during almost all of the CC and maize cycle may explain the apparent lack of this inhibitory effect (Banger et al., 2012). However, during the dates when the highest  $\text{NH}_4^+$  contents were reached in V and B (225 days after CC sowing) (Fig. 3a),  $\text{CH}_4$  emissions were significantly higher for these plots (0.12 and 0.16  $\text{mg CH}_4\text{-C m}^{-2} \text{d}^{-1}$  for V and B, respectively) than for F ( $-0.01 \text{ mg CH}_4\text{-C m}^{-2} \text{d}^{-1}$ ) (data not shown). Similarly, the  $\text{NH}_4^+$  peak observed 2 days after fertilization (Fig. 3b) decreased in the order  $\text{V} > \text{F} > \text{B}$ , the same trend as  $\text{CH}_4$  emissions (which were 0.03,  $-0.04$  and  $-0.63 \text{ mg CH}_4\text{-C m}^{-2} \text{d}^{-1}$  in V, F and B, respectively; data not shown). Contrary to Sanz-Cobena et al. (2014), the presence of CCs did not increase  $\text{CO}_2$  fluxes (Table 1) during the whole of Period I (which was longer than the period considered by these authors), even though higher fluxes were associated with B (but not V) with respect to F plots in the last phase of the intercrop. This was probably as a consequence of higher root biomass and plant respiration rates in the cereal (B) than in the legume (V). Differences from fall to early winter were not significant, since low soil temperatures limited respiration activity. The decomposition of CC residues and the growth of the maize rooting system resulted in an increase in  $\text{CO}_2$  fluxes during Period II (Oorts et al., 2007; Chirinda et al., 2010), although differences between treatments were not observed.

#### 4.4 Yield-scaled emissions, N surplus and general assessment

Yield-scaled  $\text{N}_2\text{O}$  emissions ranged from 1.74 to 7.15  $\text{g N}_2\text{O-N kg aboveground N uptake}^{-1}$ , which is about 1–4 times lower than those reported in the meta-analysis of van Groenigen et al. (2010) for a fertilizer N application rate of 150–200  $\text{kg ha}^{-1}$ . Mean N surpluses of V and F (Table 1) were in the range (0–50  $\text{kg N ha}^{-1}$ ) recommended by van Groenigen et al. (2010), while the mean N surplus in B (55  $\text{kg N ha}^{-1}$ ) was also close to optimal. In spite of higher  $\text{N}_2\text{O}$  emissions in V during Period I (which accounted for a low proportion of total cumulative  $\text{N}_2\text{O}$  losses during the experiment), these plots did not emit greater amounts of  $\text{N}_2\text{O}$  per kg of N taken up by the maize plants, and even tended to decrease yield-scaled  $\text{N}_2\text{O}$  emissions and N surplus (Table 1).

Adjusting fertilizer N rate to soil endogenous N led to lower  $\text{N}_2\text{O}$  fluxes than previous experiments where conventional N rates were applied (e.g., Adviento-Borbe et al., 2007; Hoben et al., 2011; Sanz-Cobena et al., 2012; Li et al., 2015), in agreement with the study by Migliorati et al. (2014). Moreover,  $\text{CO}_2$  equivalent emissions associated with manufacturing and transport of N synthetic fertilizers (Lal, 2004) can be reduced when low synthetic N input strategies, such as ISMF, are employed. Our results highlight the critical importance of the cash crop period on total  $\text{N}_2\text{O}$  emissions and demonstrate that the use of nonlegume and – particularly – legume CCs combined with ISFM may provide an optimum balance between GHG emissions from crop production and agronomic efficiency (i.e., lowering synthetic N requirements for a subsequent cash crop, and leading to similar yield-scaled  $\text{N}_2\text{O}$  emissions as fallow).

The use of CCs has environmental implications beyond effects on direct soil  $\text{N}_2\text{O}$  emissions. For instance, CCs can mitigate indirect  $\text{N}_2\text{O}$  losses (from  $\text{NO}_3^-$  leaching). In the study by Gabriel et al. (2012), conducted in the same experimental area,  $\text{NO}_3^-$  leaching was reduced (on average) by 30 and 59 % in V and B, respectively. Considering an emission factor of 0.075 from N leached (De Klein et al., 2006), indirect  $\text{N}_2\text{O}$  losses from leaching could be mitigated by  $0.23 \pm 0.16$  and  $0.45 \pm 0.17 \text{ kg N ha}^{-1} \text{ yr}^{-1}$  if V and B are used as CCs, respectively. Furthermore, the recent meta-analysis of Poepflau and Don (2015) revealed a C sequestration potential of  $0.32 \pm 0.08 \text{ Mg C ha}^{-1} \text{ yr}^{-1}$  with the introduction of CCs. These environmental factors, together with  $\text{CO}_2$  emissions associated with CC sowing and killing, should be assessed in future studies in order to confirm the potential in CCs for increasing both the agronomic and the environmental efficiency of irrigated cropping areas.

## 5 Conclusions

Our study confirmed that the presence of CCs (particularly V) during the intercrop period increased N<sub>2</sub>O losses, but the contribution of this phase to cumulative N<sub>2</sub>O emissions, considering the whole cropping cycle (intercrop–cash crop), was low (8–21 %). The high influence of the maize crop period over total N<sub>2</sub>O losses was due to not only N synthetic fertilization, but also CC residue mineralization and especially endogenous soil N. The type of CC residue determined the N synthetic rate in an ISFM system and affected the percentage of N<sub>2</sub>O losses coming from N fertilizer/soil N, as well as the pattern of N<sub>2</sub>O losses during the maize phase (through changes in soil NH<sub>4</sub><sup>+</sup>, NO<sub>3</sub><sup>-</sup> and DOC concentrations). By employing ISFM, similar N<sub>2</sub>O emissions were measured from CCs and F treatments at the end of the whole cropping period, resulting in low yield-scaled N<sub>2</sub>O emissions (3–6 g N<sub>2</sub>O-N kg aboveground N uptake<sup>-1</sup>) and N surplus (31 to 56 kg N ha<sup>-1</sup>). Replacing winter F with CCs did not significantly affect CH<sub>4</sub> uptake or respiration rates, during either intercrop or maize cropping periods. Our results highlight the critical importance of the cash crop period on total N<sub>2</sub>O emissions, and demonstrate that the use of nonlegume and – particularly – legume CCs combined with ISFM could be considered an efficient practice from both environmental and agronomic points of view, leading to similar N<sub>2</sub>O losses per kilogram of aboveground N uptake to bare fallow.

## 6 Data availability

Our raw data will be accessible through the repository of the Technical University of Madrid (UPM) <http://oa.upm.es/contact/>.

*Acknowledgements.* The authors are grateful to the Spanish Ministry of Economy and Innovation and the Community of Madrid for their economic support through projects AGL2012-37815-C05-01-AGR and the Agrisost-CM project (S2013/ABI- 2717). We also thank the technicians and researchers at the Department of Chemistry and Agricultural Analysis of the Agronomy Faculty (Technical University of Madrid, UPM). Rothamsted Research is grant funded by the Biotechnology and Biological Sciences Research Council (BBSRC), UK.

Edited by: E. Veldkamp

Reviewed by: two anonymous referees

## References

- Abalos, D., Sanz-Cobena, A., Garcia-Torres, L., van Groenigen, J. W., and Vallejo, A.: Role of maize stover incorporation on nitrogen oxide emissions in a non-irrigated Mediterranean barley field, *Plant Soil*, 364, 357–371, 2013.

- Abalos, D., Deyn, G. B., Kuyper, T. W., and van Groenigen, J. W.: Plant species identity surpasses species richness as a key driver of N<sub>2</sub>O emissions from grassland, *Glob. Change Biol.*, 20, 265–275, 2014.
- Adviento-Borbe, M. A. A., Haddix, M. L., Binder, D. L., Walters, D. T., and Dobermann, A.: Soil greenhouse gas fluxes and global warming potential in four high-yielding maize systems, *Glob. Change Biol.*, 13, 1972–1988, 2007.
- Aguilera, E., Lassaletta, L., Sanz-Cobena, A., Garnier, J., and Vallejo, A.: The potential of organic fertilizers and water management to reduce N<sub>2</sub>O emissions in Mediterranean climate cropping systems. A review, *Agr. Ecosyst. Environ.*, 164, 32–52, 2013.
- Allen, R. G., Raes, L. S., and Smith, D. M.: Crop evapotranspiration, Guidelines for computing crop water requirements, Irrigation and Drainage, Paper 56, Rome, Italy, FAO, 1998.
- Alonso-Ayuso, M., Gabriel, J. L., and Quemada, M.: The kill date as a management tool for cover cropping success, *Plos One*, 9, e109587, doi:10.1371/journal.pone.0109587, 2014.
- Arah, J. R. M.: Apportioning nitrous oxide fluxes between nitrification and denitrification using gas-phase mass spectrometry, *Soil Biol. Biochem.*, 29, 1295–1299, 1997.
- Baggs, E. M., Rees, R. M., Smith, K. A., and Vinten, A. J. A.: Nitrous oxide emission from soils after incorporating crop residues, *Soil Use Manage.*, 16, 82–87, 2000.
- Baggs, E. M., Stevenson, M., Pihlatie, M., Regar, A., Cook, H., and Cadisch, G.: Nitrous oxide emissions following application of residues and fertiliser under zero and conventional tillage, *Plant Soil*, 254, 361–370, 2003.
- Baggs, E. M., Smales, C. L., and Bateman, E. J.: Changing pH shifts the microbial sources as well as the magnitude of N<sub>2</sub>O emission from soil, *Biol. Fert. Soils*, 46, 793–805, 2010.
- Banger, K., Tian, H., and Lu, C.: Do nitrogen fertilizers stimulate or inhibit methane emissions from rice fields?, *Glob. Change Biol.*, 18, 3259–3267, 2012.
- Baral, K. R., Arthur, E., Olesen, J. E., and Petersen, S. O.: Predicting nitrous oxide emissions from manure properties and soil moisture: An incubation experiment, *Soil Biol. Biochem.*, 97, 112–120, 2016.
- Basche, A. D., Miguez, F. E., Kaspar, T. C., and Castellano, M. J.: Do cover crops increase or decrease nitrous oxide emissions? A meta-analysis, *J. Soil Water Conserv.*, 69, 471–482, 2014.
- Bateman, E. J. and Baggs, E. M.: Contributions of nitrification and denitrification to N<sub>2</sub>O emissions from soils at different water-filled pore space, *Biol. Fert. Soils*, 41, 379–388, 2005.
- Bayer, C., Gomes, J., Zanatta, J. A., Vieira, F. C. B., de Cássia Piccolo, M., Dieckow, J., and Six, J.: Soil nitrous oxide emissions as affected by long-term tillage, cropping systems and nitrogen fertilization in Southern Brazil, *Soil Till. Res.*, 146, 213–222, 2015.
- Bergström, L. F. and Jokela, W. E.: Ryegrass Cover Crop Effects on Nitrate Leaching in Spring Barley Fertilized with (15)NH<sub>4</sub>(15)NO<sub>3</sub>, *J. Environ. Qual.*, 30, 1659–1667, 2001.
- Chang, E. T. and Delzell, E.: Systematic review and meta-analysis of glyphosate exposure and risk of lymphohematopoietic cancers, *J. Environ. Sci. Heal. B*, 51, 402–434, 2016.
- Chirinda, N., Olesen, J. E., Porter, J. R., and Schjønning, P.: Soil properties, crop production and greenhouse gas emissions from organic and inorganic fertilizer-based arable cropping systems, *Agr. Ecosyst. Environ.*, 139, 584–594, 2010.

- Danielson, R. E. and Sutherland, P. L.: Porosity, in: *Methods of Soil Analysis. Part I. Physical and Mineralogical Methods*, edited by: Klute, A., Agronomy Monograph 9, American Society of Agronomy, Soil Science Society of America, Madison, WI, 443–461, 1986.
- Davidson, E. A.: Fluxes of nitrous oxide and nitric acid from terrestrial ecosystem, in: *Microbial production and consumption of greenhouse gases: Methane, Nitrous oxide and Halomethane*, edited by: Rogers, J. E. and Whitman, W. B., American Society of Microbiology, Washington, 219–236, 1991.
- Davidson, E. A. and Kanter, D.: Inventories and scenarios of nitrous oxide emissions, *Environ. Res. Lett.*, 9, 105012, doi:10.1088/1748-9326/9/10/105012, 2014.
- De Klein, C., Novoa, R. S. A., Ogle, S., Smith, K. A., Rochette, P., Wirth, T. C., Mc Conket, B. G., Walsh, M., Mosier, A., Rypdal, K., and Williams, S. A.: IPCC guidelines for national greenhouse gas inventories, Volume 4, Chapter 11: N<sub>2</sub>O emissions from managed soils, and CO<sub>2</sub> emissions from lime and urea application, Technical Report 4-88788-032-4, Intergovernmental Panel on Climate Change, 2006.
- Dendooven, L., Patino-Zúñiga, L., Verhulst, N., Luna-Guido, M., Marsch, R., and Govaerts, B.: Global warming potential of agricultural systems with contrasting tillage and residue management in the central highlands of Mexico, *Agr. Ecosyst. Environ.*, 152, 50–58, 2012.
- Dunfield, P. and Knowles, R.: Kinetics of inhibition of methane oxidation by nitrate, nitrite, and ammonium in a humisol, *Appl. Environ. Microb.*, 61, 3129–3135, 1995.
- Feyereisen, G. W., Wilson, B. N., Sands, G. R., Strock, J. S., and Porter, P. M.: Potential for a rye cover crop to reduce nitrate loss in southwestern Minnesota, *Agron. J.*, 98, 1416–1426, 2006.
- Firestone, M. K. and Davidson, E. A.: Microbiological basis of NO and N<sub>2</sub>O production and consumption in soil, in: *Exchange of Trace Gases between Terrestrial Ecosystems and the Atmosphere* Chichester, edited by: Andeae, M. O. and Schimel, D. S., Wiley, 7–21, 1989.
- Frimpong, K. A. and Baggs, E. M.: Do combined applications of crop residues and inorganic fertilizer lower emission of N<sub>2</sub>O from soil?, *Soil Use Manage.*, 26, 412–424, 2010.
- Frimpong, K. A., Yawson, D. O., Baggs, E. M., and Agyarko, K.: Does incorporation of cowpea-maize residue mixes influence nitrous oxide emission and mineral nitrogen release in a tropical luvisol?, *Nutr. Cycl. Agroecosys.*, 91, 281–292, 2011.
- Gabriel, J. L. and Quemada, M.: Replacing bare fallow with cover crops in a maize cropping system: yield, N uptake and fertiliser fate, *Eur. J. Agron.*, 34, 133–143, 2011.
- Gabriel, J. L., Muñoz-Carpena, R., and Quemada, M.: The role of cover crops in irrigated systems: Water balance, nitrate leaching and soil mineral nitrogen accumulation, *Agr. Ecosyst. Environ.*, 155, 50–61, 2012.
- Gabriel, J. L., Alonso-Ayuso, M., García-González, I., Hontoria, C., and Quemada, M.: Nitrogen use efficiency and fertiliser fate in a long-term experiment with winter cover crops, *Eur. J. Agron.*, 79, 14–22, 2016.
- Gao, J., Xie, Y., Jin, H., Liu, Y., Bai, X., Ma, D., Zhu, Y., Wang, C., and Guo, T.: Nitrous Oxide Emission and Denitrifier Abundance in Two Agricultural Soils Amended with Crop Residues and Urea in the North China Plain, *Plos One*, 11, e0154773, doi:10.1371/journal.pone.0154773, 2016.
- García-Marco, S., Ravella, S. R., Chadwick, D., Vallejo, A., Gregory, A. S., and Cárdenas, L. M.: Ranking factors affecting emissions of GHG from incubated agricultural soils, *Eur. J. Soil Sci.*, 65, 573–583, 2014.
- Grossman, R. B. and Reinsch, T. G.: 2.1 Bulk density and linear extensibility. *Methods of Soil Analysis. Part 4: Physical Methods*, Soil Science Society of America, Madison, USA, 201–228, 2002.
- Hoben, J. P., Gehl, R. J., Millar, N., Grace, P. R., and Robertson, G. P.: Nonlinear nitrous oxide (N<sub>2</sub>O) response to nitrogen fertilizer in on-farm corn crops of the US Midwest, *Glob. Change Biol.*, 17, 1140–1152, 2011.
- IPCC: Climate change 2007. The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change, edited by: Solomon, S., Qin, D., Manning, M., Chen, Z., Marquis, M., Averyt, K. B., Tignor, M., and Miller, H. L., Cambridge University Press, p. 996, 2007.
- Justes, E., Mary, B., and Nicolardot, B.: Comparing the effectiveness of radish cover crop, oilseed rape volunteers and oilseed rape residues incorporation for reducing nitrate leaching, *Nutr. Cycl. Agroecosys.*, 55, 207–220, 1999.
- Kallenbach, C. M., Rolston, D. E., and Horwath, W. R.: Cover cropping affects soil N<sub>2</sub>O and CO<sub>2</sub> emissions differently depending on type of irrigation, *Agr. Ecosyst. Environ.*, 137, 251–260, 2010.
- Kimani, S. K., Nandwa, S. M., Mugendi, D. N., Obanyi, S. N., Ojiem, J., Murwira, Herbert K., and Bationo, A.: Principles of integrated soil fertility management, in: *Soil fertility management in Africa: A regional perspective*, edited by: Gichuri, M. P., Bationo, A., Bekunda, M. A., Goma, H. C., Mafongoya, P. L., Mugendi, D. N., Murwira, H. K., Nandwa, S. M., Nyathi, P., and Swift, M. J., Academy Science Publishers (ASP), Centro Internacional de Agricultura Tropical (CIAT), Tropical Soil Biology and Fertility (TSBF), Nairobi, KE, 51–72, 2003.
- Lal, R.: Carbon emission from farm operations, *Environ. Int.* 30, 981–990, 2004.
- Laughlin, R. J., Stevens, R. J., and Zhuo, S.: Determining nitrogen-15 in ammonium by producing nitrous oxide, *Soil Sci. Soc. Am. J.*, 61, 462–465, 1997.
- Li, N., Ning, T., Cui, Z., Tian, S., Li, Z., and Lal, R.: N<sub>2</sub>O emissions and yield in maize field fertilized with polymer-coated urea under subsoiling or rotary tillage, *Nutr. Cycl. Agroecosys.*, 102, 397–410, 2015.
- Li, X., Sørensen, P., Olesen, J. E., and Petersen, S. O.: Evidence for denitrification as main source of N<sub>2</sub>O emission from residue-amended soil, *Soil Biol. Biochem.*, 92, 153–160, 2016.
- López-Fernández, S., Diez, J. A., Hernaiz, P., Arce, A., García-Torres, L., and Vallejo, A.: Effects of fertiliser type and the presence or absence of plants on nitrous oxide emissions from irrigated soils, *Nutr. Cycl. Agroecosys.*, 78, 279–289, 2007.
- MacKenzie, A. F., Fan, M. X., and Cadrin, F.: Nitrous oxide emission in three years as affected by tillage, corn-soybean-alfalfa rotations, and nitrogen fertilization, *J. Environ. Qual.*, 27, 698–703, 1998.
- Martínez-Cob, A.: Use of thermal units to estimate corn crop coefficients under semiarid climatic conditions, *Irrigation Sci.*, 26, 335–345, 2008.

- Meijide, A., Cárdenas, L. M., Sánchez-Martín, L., and Vallejo, A.: Carbon dioxide and methane fluxes from a barley field amended with organic fertilizers under Mediterranean climatic conditions, *Plant Soil*, 328, 353–367, 2010.
- Migliorati, M. D. A., Scheer, C., Grace, P. R., Rowlings, D. W., Bell, M., and McGree, J.: Influence of different nitrogen rates and DMPP nitrification inhibitor on annual N<sub>2</sub>O emissions from a subtropical wheat–maize cropping system, *Agr. Ecosyst. Environ.*, 186, 33–43, 2014.
- Mørkved, P. T., Dörsch, P., and Bakken, L. R.: The N<sub>2</sub>O product ratio of nitrification and its dependence on long-term changes in soil pH, *Soil Biol. Biochem.*, 39, 2048–2057, 2007.
- Nemecek, T., von Richthofen, J. S., Dubois, G., Casta, P., Charles, R., and Pahl, H.: Environmental impacts of introducing grain legumes into European crop rotations, *Eur. J. Agron.*, 28, 380–393, 2008.
- Oorts, K., Merckx, R., Gréhan, E., Labreuche, J., and Nicolardot, B.: Determinants of annual fluxes of CO<sub>2</sub> and N<sub>2</sub>O in long-term no-tillage and conventional tillage systems in northern France, *Soil Till. Res.*, 95, 133–148, 2007.
- Poeplau, C. and Don, A.: Carbon sequestration in agricultural soils via cultivation of cover crops – A meta-analysis, *Agr. Ecosyst. Environ.*, 200, 33–41, 2015.
- Quemada, M., Baranski, M., Nobel-de Lange, M. N. J., Vallejo, A., and Cooper, J. M.: Meta-analysis of strategies to control nitrate leaching in irrigated agricultural systems and their effects on crop yield, *Agr. Ecosyst. Environ.*, 174, 1–10, 2013.
- Quemada, M., Gabriel, J. L., and Zarco-Tejada, P.: Airborne hyperspectral images and ground-level optical sensors as assessment tools for maize nitrogen fertilization, *Remote Sens.*, 6, 2940–2962, 2014.
- Reeves, S. and Wang, W.: Optimum sampling time and frequency for measuring N<sub>2</sub>O emissions from a rain-fed cereal cropping system, *Sci. Total Environ.*, 530, 219–226, 2015.
- Rochette, P. and Janzen, H. H.: Towards a revised coefficient for estimating N<sub>2</sub>O emissions from legumes, *Nutr. Cycl. Agroecosys.*, 73, 171–179, 2005.
- Sanz-Cobena, A., Sánchez-Martín, L., García-Torres, L., and Vallejo, A.: Gaseous emissions of N<sub>2</sub>O and NO and NO<sub>3</sub><sup>-</sup> leaching from urea applied with urease and nitrification inhibitors to a maize (*Zea mays*) crop, *Agr. Ecosyst. Environ.*, 149, 64–73, 2012.
- Sanz-Cobena, A., García-Marco, S., Quemada, M., Gabriel, J. L., Almendros, P., and Vallejo, A.: Do cover crops enhance N<sub>2</sub>O, CO<sub>2</sub> or CH<sub>4</sub> emissions from soil in Mediterranean arable systems?, *Sci. Total Environ.*, 466, 164–174, 2014.
- Sarkodie-Addo, J., Lee, H. C., and Baggs, E. M.: Nitrous oxide emissions after application of inorganic fertilizer and incorporation of green manure residues, *Soil Use Manage.*, 19, 331–339, 2003.
- Schindlbacher, A., Zechmeister-Boltenstern, S., and Butterbach-Bahl, K.: Effects of soil moisture and temperature on NO, NO<sub>2</sub>, and N<sub>2</sub>O emissions from European forest soils, *J. Geophys. Res.-Atmos.*, 109, D17302, doi:10.1029/2004JD004590, 2004.
- Senbayram, M., Chen, R., Mühling, K. H., and Dittert, K.: Contribution of nitrification and denitrification to nitrous oxide emissions from soils after application of biogas waste and other fertilizers, *Rapid Commun. Mass Sp.*, 23, 2489–2498, 2009.
- Shan, J. and Yan, X.: Effects of crop residue returning on nitrous oxide emissions in agricultural soils, *Atmos. Environ.*, 71, 170–175, 2013.
- Snyder, C. S., Bruulsema, T. W., Jensen, T. L., and Fixen, P. E.: Review of greenhouse gas emissions from crop production systems and fertilizer management effects, *Agr. Ecosyst. Environ.*, 133, 247–266, 2009.
- Soil Survey Staff: Keys to Soil Taxonomy, 12th Edn., USDA-Natural Resources Conservation Service, Washington DC, USA, 2014.
- Spiertz, J. H. J.: Nitrogen, sustainable agriculture and food security. A review, *Agron. Sustain. Dev.*, 30, 43–55, 2010.
- Stehfest, E. and Bouwman, L.: N<sub>2</sub>O and NO emission from agricultural fields and soils under natural vegetation: summarizing available measurement data and modeling of global annual emissions, *Nutr. Cycl. Agroecosys.*, 74, 207–228, 2006.
- Tate, K. R.: Soil methane oxidation and land-use change—from process to mitigation, *Soil Biol. Biochem.*, 80, 260–272, 2015.
- Tonitto, C., David, M. B., and Drinkwater, L. E.: Replacing bare fallows with cover crops in fertilizer-intensive cropping systems: A meta-analysis of crop yield and N dynamics, *Agr. Ecosyst. Environ.*, 112, 58–72, 2006.
- Ussiri, D. and Lal, R.: Soil emission of nitrous oxide and its mitigation, Springer, the Netherlands, 2013.
- Vallejo, A., Skiba, U. M., García-Torres, L., Arce, A., López-Fernández, S., and Sánchez-Martín, L.: Nitrogen oxides emission from soils bearing a potato crop as influenced by fertilization with treated pig slurries and composts, *Soil Biol. Biochem.*, 38, 2782–2793, 2006.
- van Groenigen, J. W., Velthof, G. L., Oenema, O., van Groenigen, K. J., and van Kessel, C.: Towards an agronomic assessment of N<sub>2</sub>O emissions: a case study for arable crops, *Eur. J. Soil Sci.*, 61, 903–913, 2010.
- Vanlauwe, B., Kihara, J., Chivenge, P., Pypers, P., Coe, R., and Six, J.: Agronomic use efficiency of N fertilizer in maize-based systems in sub-Saharan Africa within the context of integrated soil fertility management, *Plant Soil*, 339, 35–50, 2011.
- Veenstra, J. J., Horwath, W. R., and Mitchell, J. P.: Tillage and cover cropping effects on aggregate-protected carbon in cotton and tomato, *Soil Sci. Soc. Am. J.*, 71, 362–371, 2007.
- Wagner-Riddle, C. and Thurtell, G. W.: Nitrous oxide emissions from agricultural fields during winter and spring thaw as affected by management practices, *Nutr. Cycl. Agroecosys.*, 52, 151–163, 1998.
- Wichern, F., Eberhardt, E., Mayer, J., Joergensen, R. G., and Müller, T.: Nitrogen rhizodeposition in agricultural crops: methods, estimates and future prospects, *Soil Biol. Biochem.*, 40, 30–48, 2008.
- Yamulki, S. and Jarvis, S.: Short-term effects of tillage and compaction on nitrous oxide, nitric oxide, nitrogen dioxide, methane and carbon dioxide fluxes from grassland, *Biol. Fert. Soils*, 36, 224–231, 2002.