

Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

Goat and cow casein derived ingredients and their interactions with iron

A thesis presented in partial fulfilment of the requirements for the degree of

Doctor of Philosophy

in

Food Technology

Massey University, Palmerston North, New Zealand

Alice Małgorzata Smialowska



Abstract

The objective of this study was to gain a fundamental understanding of how goat casein micelles and the products of casein proteins behave when fortified with iron.

Iron fortified skim milk was characterised by analysing the mass balance of micellar/non micellar fractions, chemical changes, micellar size changes and internal structure. Two treatments were examined to determine where in the processing line the addition of iron might best be added to a milk system. On average, at least 72% of the iron is bound to the micellar phase across the treatments and iron concentrations. Small angle X-ray scattering (SAXS) indicated that internal changes, mainly at the location of the colloidal calcium phosphate, occurred with iron addition.

Casein was extracted from goat milk using isoelectric precipitation however the extraction was more difficult than using cow milk. Iron fortification of the caseinates resulted in a tendency for oxidation and precipitation of the proteins to occur causing the formation of large aggregates. The caseinates could not stabilise the same amounts of iron to that of an intact casein micelle.

Phosphopeptides were isolated by adding calcium and ethanol to caseinate digests. There was an increase in serine, glutamic acid and isoleucine residues compared to caseinate. There was an increase in phosphorus from 7.8 ± 0.3 mg P/ g solids to 45.4 ± 2.4 mg P/ g solids in the isolate. The phosphopeptides were composed of smaller, more hydrophilic peptides compared to the full digest prior to precipitation. Ferrous sulfate was then investigated for use as the precipitant, instead of calcium. The peptides produced similar trends in terms of amino acid profile changes, phosphorus concentration increase and yield. Immobilised metal affinity chromatography was also investigated however this had a low throughput that may not be effective at process scale.

The effect of heating, cooling, ionic strength of the solution, holding time, iron loading, processing order and use in a model milk system were investigated to simulate potential industrial processing conditions using the calcium - extracted phosphopeptides. It was found that goat peptide isolates were able to bind 54.4 ± 0.5 mg Fe/ g protein compared to goat milk of 4.3 ± 0.1 mg Fe/ g protein. The optimal conditions for binding were found to be at pH 6.7 in a low ionic strength solution,

around 37 °C. There was a potential synergistic effect of adding the peptides to milk in terms of iron binding capacity. There were few differences in the amount of iron that could be bound comparing cow and goat derived phosphopeptides under the tested conditions.

The oxidation potential of ingredients was determined using malondialdehyde (MDA) as an oxidation product marker. There was a reduction in oxidation when iron was bound to milk or peptides compared to free ferrous sulfate in solution with intact goat milk performing the best producing $0.46 \pm 0.04 \,\mu g$ MDA/mL after 3 days at 30 °C compared to the blank of $1.25 \pm 0.16 \,\mu g$ MDA/mL. The goat peptides produced non-significantly different levels of MDA compared to the blank containing no ferrous sulfate.

Caco-2 cell lines are a way of approximating how systems may function in an intestine in terms of nutrient absorption. Iron absorption was improved in the order of casein hydrolysates > caseinate > skim milk for goat milk. In contrast, cow milk appeared to perform better without any modifications to the proteins. On an equal iron filtrate basis after the digestion and intestinal phase, calcium- precipitated goat phosphopeptides produced a response of 9.64 ± 0.94 ng ferritin/ nM iron. This response was greater than all other treatments with the exception of goat milk fortified with 5 mM iron and ascorbic acid with 12.30 ± 1.23 ng ferritin/ nM iron.

This work covers a wide range of milk products and iron interactions and has helped to build a fundamental understanding of goat milk protein functionality. The underpinning considerations to a manufacturing setting may allow further development of large scale ingredient production for the improved stability of iron fortified systems.

Acknowledgments

I would firstly like to thank my supervisor Dr. Alistair Carr for allowing me to pursue a PhD. His discussions and ideas have made for a very interesting and exciting project. This sense of humour made the long stints in the Synchrotron labs and during the whole project a lot more colourful. Thank you for carefully reading my thesis and asking valid questions which have enriched the discussion.

To my co-supervisor Dr. Lara Matia-Merino, thank you for your hard work and support during my studies. She has provided focus to the project and kept me motivated when I was struggling. Her directness helped move work along which I am grateful for while her advice and personal support showed her true caring nature.

Dr. Bridget Ingham: my co-supervisor who had her work cut out for her. I am thankful that I was able to incorporate techniques like SAXS in my work as it allowed me to experience a whole different world in science, not to mention travelling to the Synchrotrons. It has to be mentioned that she has great patience in explaining the theory, equations and models on numerous occasions and assessing my fitted models many times over.

I would like to acknowledge Dairy Goat Co-operative, specifically Dr. Colin Prosser for giving me the opportunity to undertake a PhD by providing the funding for the whole project.

Thanks to Bob (Dr. Robin Stewart) for firstly performing the Caco-2 cell assays for the part of the project and also showing me how the method works (and making me thankful that none of my work was as temperamental and demanding). Also, for being a great office buddy and sharing a few good yarns.

Thank you to the Laboratory Technicians in the School: Steve Glasgow, Micelle Tamehana, Janiene Gilliland, Warwick Johnson, Garry Radford and John Edwards (SEAT) for help using equipment, making orders and running methods (as well as some amusing chats). Trevor Loo from the Fundamental Sciences department taught me how to carry out digestions and gave some good pointers in this area so I must acknowledge how much this helped me. Along the same track, thanks to Don Otter for helping me with mass spectrometry, if anything, I have learnt how to read the

output even if results didn't eventuate. Finally, thank you to Maggie Zou who helped me run the HPLC and showing me the ins and outs of this technique.

Thank you to Prof. Dick FitzGerald for the useful discussions regarding IMAC; this helped me immensely to achieve results for this section of work.

Thank you to the Australian Synchrotron and the Lawrence Berkeley Advanced Light Source, specifically Nigel Kirby and Cheng Wang for helping our team in running the equipment and input in analysis.

Thank you to Circo Acrofit Studios and the people there that make it so much fun. My time in Palmerston North has been made more fulfilling when I knew class was coming up. This place was great for exercise and humour and I have made some awesome friends here.

Thank you to Kate Donohue who initially convinced me to carry out a postgraduate degree and being an excellent support through the shared experience of doing a PhD. Also, for turning me to the 'R side' of statistics, making me pretty confident in plot coding but also giving insight into statistical analysis. Our time together as flatmates was super fun as evidenced by us being inseparable during weekend shenanigans.

I need to also thank my parents for motivating and encouraging me (along with financial support) during this time. Thanks for being proud of me and being okay with me delaying becoming a 'real adult' and stretching out student life for as long as possible.

Table of Contents

1	Int	troduction	1
2	Lit	terature review	5
	2.1	Casein micelle composition and structure	5
	2.1.	.1 Casein	5
	2.1.	.2 Milk Salts	7
	2.1.	.3 Milk processing	
	2.2	Comparison of cow and goat milk composition	
	2.2.	.1 Protein	
	2.2.	.2 Physicochemical properties of goat casein micelles	
	2.2.	.3 Minerals	
	2.2.	.4 Nutritional value differences of cow and goat milk	14
	2.3	Natively Occurring Iron in milk	16
	2.3.	.1 Iron Concentration in milk	16
	2.3.	.2 Native Iron chelates in milk- Lactoferrin and Transferrin	16
	2.4	Iron Fortification of Milk	17
	2.4.	.1 Iron chemistry	17
	2.4.	.2 Iron salts for fortification	19
	2.4.	.3 Iron binding and effects on casein	
	2.5	Casein Phosphopeptides	
	2.5.	.1 Digestion of casein and peptide analysis	
	2.5.	.2 Phosphopeptides and their interaction with iron	
	2.5.	Animal and human studies using casein phosphopeptide- iron con	mplexes and
	thei	ir absorption	
	2.5.	.4 Current commercial phosphopeptide ingredients	
	2.6	Techniques	
	2.6.	.1 Small angle X-ray scattering	
	2.6.	2 Particle characterisation	
	2.6.	.3 Immobilised metal ion affinity chromatography	
	2.6.	.4 Lipid oxidation	
	2.6.	.5 Caco-2 cells	
	2.7	Conclusion	
3	Ma	aterials and methods	43
	3.1	Composition of skim milk powders	
	3.2	Studies on Iron fortified- skim milks: - sample preparation	

3.2.	.1 Nomenclature	44
3.2.	.2 Iron fortified dry blended milk	44
3.2.	.3 Iron fortified wet blended milk	44
3.2.	.4 Casein micelle partition	45
3.2.	.5 Moisture content of pellet	45
3.2.	.6 Calcium removal from cow and goat skim milk	45
3.3 cow ai	Studies on sodium caseinates- Isoelectric precipitation of sodium casein nd goat skim milk powders	nate from
3.3.	.1 Cow caseinate preparation	46
3.3.	.2 Goat casein preparation	46
3.3.	.3 Iron fortification of caseinates	46
3.4	Studies on cow and goat caseinate hydrolysates	47
3.4.	.1 Hydrolysis of cow and goat caseinate using trypsin	47
3.4.	.2 Selective precipitation of phosphopeptides using calcium chloride	47
3.4.	.3 Selective precipitation of phosphopeptides using ferrous sulfate	49
3.4.	.4 Immobilised metal affinity chromatography for phosphopeptide isola	ation 49
3.5	Studies on cow and goat phosphopeptides	
3.5.	.1 Sample preparation	
3.5.	.2 Iron chelation measurement	
3.6	Mineral and protein estimation	
3.6.	.1 Kjeldahl method	53
3.6.	.2 Lowry method	
3.6.	.3 Iron binding analysis	
3.6.	.4 Phosphorus assay	55
3.6.	.5 Calcium content	
3.7	Characterisation Techniques	
3.7.	.1 Dynamic Light scattering	
3.7.	.2 Zeta potential	
3.7.	.3 Turbidity	
3.7.	.4 Small angle X-ray Scattering (SAXS)	
3.7.	.5 OPA for Degree of hydrolysis	
3.7.	.6 RP-HPLC analysis	60
3.7.	.7 Amino acid composition	63
3.7.	.8 Protein- mineral oxidation activity	63
3.7.	9 Caco-2 cell lines	65
3.8	Statistical analysis	65

4	Iro	on fortification of goat and cow skim milk	67
2	4.1	Introduction	67
2	4.2	Results and Discussion	69
	4.2.1	1 Cow and goat skim milk characterisation	69
	4.2.2	2 Iron fortification of goat and cow skim milk	71
	4.2.3	3 Elemental analysis of wet blended goat and cow milk	85
	4.2.4	4 SAXS analysis of dry blended and wet blended iron fortified goat and c 90	ow milk
	4.2.5	5 Characterisation of iron fortified goat and cow milk with calcium d 109	epletion
2	4.3	Conclusion	114
5	Iso	lation and Addition of iron to caseinate solutions	117
4	5.1	Introduction	117
4	5.2	Results and Discussion	119
	5.2.1	1 Isoelectric precipitation of sodium caseinate from cow and goat milk.	119
	5.2.2	2 Iron fortification of cow and goat caseinate	126
4	5.3	Conclusion	134
6	Iso	lation and characterisation of phosphopeptides by sel	ective
pr	ecipi	itation and IMAC	137
(5.1	Introduction	137
(5.2	Results and Discussion	140
	6.2.1	1 Digestion of cow and goat caseinate- Rate of hydrolysis	140
	6.2.2	2 RP-HPLC of hydrolysed cow and goat caseinates	143
	6.2.3	3 Iron chelation of digests using Ferrozine	150
	6.2.4	4 Selective precipitation of phosphoserine containing peptides using 154	calcium
	6.2.5	5 Yield of phosphopeptides from the caseinate digests	155
	6.2.6	6 Calcium content	157
	6.2.7	7 Phosphorus content	157
	6.2.8	8 Amino acid analysis	158
	6.2.9	9 RP-HPLC of cow and goat calcium precipitated digests	161
	6.2.1	10 Selective precipitation of phosphoserine containing peptides using iron	n 163
	6.2.1	11 Iron content	165
	6.2.1	12 Phosphorus content	166
	6.2.1	13 Amino acid composition	166
	6.2.1	14 HPLC of cow and goat iron precipitated digests	171

	6.2.	.15 Isolation of phosphopeptides using IMAC	173
	6.3	Conclusion	183
7	Ch	naracterisation of iron binding capacity of cow and	goat
p	hospl	hopeptides	185
	7.1	Introduction	185
	7.2	Results and Discussion	187
	7.2.	.1 Calcium content	187
	7.2.	.2 Iron binding of phosphopeptides at various calcium levels	188
	7.2.	.3 Determination of isoelectric point of ion-exchanged peptides	189
	7.2.	.4 Iron binding at various pH, ionic strength and temperatures	190
	7.2.	.5 Effect of different buffer solutions at pH 6.7	198
	7.2.	.6 Effect of holding time on iron binding	200
	7.2.	.7 Testing the maximum capacity of iron binding	208
	7.2.	.8 Iron binding in a milk system	213
	7.3	Conclusion	221
8	Sta	ability and absorption of iron- phosphopeptide complexes .	223
	8.1	Introduction	223
	8.2	Results and Discussion	225
	8.2. pho	.1 Oxidative stability of cow and goat skim milk and cow and osphopeptides in a linoleic acid emulsion	goat 225
	8.2. goat	.2 Caco-2 cell line analysis of bioavailability and absorption of iron from cov t milk and their products	<i>w</i> and231
	8.3	Conclusion	244
9	Со	onclusions and Recommendations	247
	9.1	Iron fortification of cow and goat milk	247
	9.1.	.1 Recommendations	250
	9.2	Isoelectric precipitation of sodium caseinate	252
	9.2.	.1 Recommendations	253
	9.3 phospl	Comparison of calcium, iron precipitated and IMAC eluted cow and hopeptides	goat 253
	9.3.	.1 Recommendations	256
	9.4	Iron chelation capacity of phosphopeptides	256
	9.4.	.1 Calcium removal techniques	256
	9.4. cone	.2 Iron binding capacity of cow and goat peptides under various proce ditons 257	ssing
	9.4.	.3 Recommendations	259

9.5	Oxi	dative Stability and Caco-2 cell analysis of cow and goat milk products .	260
9.5.	.1	Anti-oxidant properties of cow and goat milk and phosphopeptides	260
9.5.	.2	Bioavailability of iron chelates using Caco-2 cells	261
9.5.	.3	Recommendations	262
9.6	Sun	nmary	263
10 Lis	st of	publications	
11 Re	fere	nces	269
A Ti	me s	eries of cow and goat caseinate digestion	
A.1	Chr	omatograms from section 6.2.2- 1: 200 E:S ratio	287
A.2	Chr	omatograms from section 6.2.2- 1: 200 E:S ratio	291
B M	etho	d development of IMAC: testing iron and buffers	297
B.1	Intro	oduction	297
B.1	.1	Method	297
B.1	.2	Results	298
B.2	Was	shing and elution chromatographs of section 6.2.15	304
B.3	Ref	erences	307

Table of Figures

Figure 2-1: Phosphoserine cluster containing three phosphorylated serines followed
Eisens 2.2: Schemetic of SAVS schemetic form Schecklesson & Sinch (2011) and
Kikhney & Svergun, (2015)
Figure 3-1: Protocol for hydrolysis and for the isolation of calcium precipitated phosphopeptides
Figure 3-2: Response of iron detection over three days to assess stability of reagents.
Figure 3-3: Phosphorus standard curve showing colour profile
Figure 3-4: Buffer gradient for RP-HPLC elution of casein and caseinates, buffer A: 95% H2O, 5% acetonitrile, 0.01% trifluroacetic acid; Buffer B: 95% acetonitrile, 5% H2O, 0.08% trifluroacetic acid
Figure 3-5: Buffer gradient for RP-HPLC elution of caseinate digests, buffer A: 95% H2O, 5% acetonitrile, 0.01% trifluroacetic acid; Buffer B: 95% acetonitrile, 5% H2O, 0.08% trifluroacetic acid
Figure 3-6: Buffer gradient for HPLC elution of hydrolysates, buffer A: 95% H2O, 5% acetonitrile, 0.01% trifluroacetic acid; Buffer B: 95% acetonitrile, 5% H2O, 0.08% trifluroacetic acid.
Figure 3-7: Standard curve of acid hydrolysed 1,1,3,3 tetraethoxypropane (TEP) for the determination of malondialdehyde content
Figure 4-1: Cow and goat skim milk casein micelle size distribution by intensity; error bars indicate standard error, n=3
Figure 4-2: pH of cow and goat skim milk at 3.5% (w/w) protein as 0, 5, 10, 15 and 20 mM of ferrous sulfate are added; error bars indicate standard error, n=373
Figure 4-3: Box plot of dry blended cow and goat milk fortified with 0, 5, 10, 15 or 20 mM ferrous sulfate showing particle size distribution, significant differences are indicated in Table 4-4, n>3

Figure 4-4: Size distribution of goat milk fortified with 0, 5, 10, 15 and 20 mM iron by Intensity in dry blended iron fortified milk; error bars indicate standard error, n>3.

Figure 4-5: Size distribution of cow milk fortified with 0, 5, 10, 15 and 20 mM iron by Intensity in dry blended iron fortified milk; error bars indicate standard error, n>3.

by Intensity in wet blended iron fortified milk; error bars indicate standard error, n>3.

Figure 4-8: Size distribution of goat milk fortified with 0, 5, 10, 15 and 20 mM iron by Intensity in wet blended iron fortified milk; error bars indicate standard error, n > 3.

Figure 4-9: Partition of iron in micellar phase after ultra-centrifugation at 90,000 xg, 1 hour, 20 °C shown as a fraction of the total iron detected in the micellar phase (open shapes) and the amount of iron bound to the micellar fraction (filled shapes) in dry blended iron fortified milk; error bars indicate standard error, n=3, arrows indicate related axis to points.

Figure 4-10: Partition of iron in micellar phase after ultra-centrifugation at 90,000 xg, 1 hour, 20 °C shown as a fraction of the total iron detected in the micellar phase (open shapes) and the amount of iron bound to the micellar fraction in (filled shapes) wet blended iron fortified milk; error bars indicate standard error, n=3, arrows indicate related axis to points.

Figure 4-17: Representation of how a casein micelle looks in a SAXS model.......94

Figure 4-21: Scattering curves of calcium (bottom) and iron (top) of energy scans.

Figure 4-24: Intensity plots (A, C) and Kratky plots (B, D) of milk treatments from SAXS. Figure 1 plots show dry blended milks measured at 4°C; Figure 2 plots show wet blended milks at 4°C; Figure 3 plots show wet blended milks at 37°C. For all Figures, A and B are cow milk samples and C and D are goat milk samples. Colours;

black= 0 mM, red= 5 mM, purple= 10 mM, blue= 15 mM and grey= 20 mM, indicate Figure 4-25: Schematic of how casein protein strands associate in native milk (top) and iron may interact with the colloidal calcium phosphate (bottom), either with the iron becoming part of the CCP amorphous structure (left) or precipitating onto the Figure 4-26: Particle size of cow milk particle size by intensity (Figure A) and number (Figure B) and goat milk by intensity (Figure C) and number (Figure D) with $0 (\Box)$, 5 (\circ), 10 (Δ), 15 (+) and 20 (x) mM iron; error bars indicate standard error, n=3. ... 111 Figure 4-27: Z-average particle sizes of calcium depleted cow and goat milks fortified Figure 4-28: Pellets of cow and goat, 0, 5, 10. 15 and 20 mM iron addition; from left to right: (a) 0 mM cow, (b) 0 mM goat, (c) 5 mM cow, (d) 5 mM goat, (e) 10 mM cow, (f) 10 mM goat, (g) 15 mM cow, (h) 15 mM goat, (i) 20 mM cow, (j) 20 mM goat. Figure 4-29: Moisture content (g water/ g pellet), A; protein content of serum (mg protein/ mL serum, B; iron content of either serum phase or starting milk solution, C; and iron content of pellet (mg Fe/ g pellet), D, of cow and goat calcium depleted milk

Figura	$5 1 \cdot C$	our drim	mille prooi	nitating at	nU16		120
riguic.	J-1. U	OW SKIIII	mink preci	pitating at	pm 4.0.	 	120

Figure 5-2: Cov	w casein curd after	collection.	 20

Figure 5-5: Goat casein curd after isoelectric precipitation and centrifugation. 123

Figure 5-7: RP-HPLC chromatogram of Alanate 180 using a 5 % to 60 % buffer B gradient at 214 nm. 125

Figure 5-9a: Cow caseinate (3.0 %) fortified with 5 mM, 11 mM and 22 mM ferrous sulfate with an unadjusted pH after fortification
Figure 5-10: Particle size by intensity (A, B), volume (C, D) and number (E, F) of cow (A, C, E) and goat (B, D, F) caseinate fortified with 0 (\Box), 5 (\circ), 11 (Δ), and 22 (+) mM ferrous sulfate; error bars indicate standard error, n=3
Figure 5-11: Protein solubility of cow and goat caseinate with iron addition of 0, 5, 11 and 22 mM ferrous sulfate after centrifugation at 10,800 xg for 20 minutes; error bars indicate standard error, n=3
Figure 5-12: Iron partition of iron fortified cow and goat caseinate; error bars indicate standard error, n=3
Figure 6-1: Degree of hydrolysis versus time of 1:50 enzyme to protein cow and goat caseinate samples; error bars indicate standard error, n=3
Figure 6-2: Degree of hydrolysis curve of cow and goat caseinate with a 1:200 enzyme to protein ratio; error bars indicate standard error, n=3
Figure 6-3: Chromatograms of cow caseinate digests at 0, 2, and 18 hours with 1:50 enzyme to protein ratio measured at 280 nm, bandwidth 8 nm
Figure 6-4: Chromatograms of goat caseinate digests at 0, 2 and 18 hours with 1:50 enzyme to protein ratio measured at 280 nm, bandwidth 8 nm
Figure 6-5: Chromatograms of cow caseinate digests at 2 and 18 hours with a 1:200 enzyme to protein ratio measured at 280 nm, bandwidth 8 nm
Figure 6-6: Chromatograms of cow caseinate digests at 2 and 18 hours with a 1:200 enzyme to protein ratio measured at 280 nm, bandwidth 8 nm
Figure 6-7: Fe binding capacity versus degree of hydrolysis of goat and cow caseinate hydrolysate at a 1:50 enzyme to protein ratio; error bars indicate standard error, n=3.
Figure 6-8: Fe binding capacity versus degree of hydrolysis of goat and cow caseinate hydrolysate at a 1:200 enzyme to protein ratio; error bars indicate standard error, n=3.
Figure 6-9: Isoelectric precipitated cow (left) and goat (right) caseinate hydrolysate after filtration.

Figure 6-10: Isoelectric precipitated cow (left) and goat (right) caseinate hydrolysate
after filtration and calcium addition at 100 mM final concentration and ethanol addition to $50 \% (v/v)$ final volume
Figure 6-11: Percentage change of amino acids after calcium and ethanol precipitation.
Figure 6-12: Amino acid composition of cow and goat calcium and ethanol precipitated peptides
Figure 6-13: Chromatograms of cow caseinate (blue), cow phosphopeptide (green), goat caseinate (red) and goat phosphopeptide (purple) at 214 nm, bandwidth 8 nm, reference 360 nm, bandwidth 50 nm
Figure 6-14: Chromatograms of cow caseinate (blue), cow phosphopeptide (green), goat caseinate (red) and goat phosphopeptide (purple) at 280 nm, bandwidth 8 nm, reference 360 nm, bandwidth 50 nm
Figure 6-15: Acid clarified cow (left) and goat (right) caseinate digests after ferrous sulfate addition of 90 mM final concentration
Figure 6-16: cow caseinate (left) and goat caseinate (right) digests after iron and ethanol addition
Figure 6-17: Percentage change of amino acid composition of cow and goat iron precipitated peptides with respect to their respective caseinate starting material 167
Figure 6-18: Comparison of amino acid composition of cow and goat iron precipitated peptides in mg/ 100 mg protein
Figure 6-19: Compositions of cow caseinate, calcium precipitated and iron precipitated peptides (top) and composition of goat caseinate, calcium precipitated and iron precipitated peptides (bottom), in mg/ 100 mg protein
Figure 6-20: Possible covalent bonding arrangements of iron to phosphoserine. A minimum of two oxygen groups are required to stabilise the iron (top) however the iron could bind to three oxygen atoms and could join two residues (bottom) 171
Figure 6-21: Ionic bonding of calcium and the phosphoserine group

Figure 6-22: cow caseinate digest (green) and phosphopeptide (blue); goat caseinate digest (maroon) and phosphopeptide (teal) at 214 nm, bandwidth 8 nm, reference 360 nm, bandwidth 50 nm. 173

Figure 6-23: Combined chromatograms of calcium and iron precipitated phosphopeptides: calcium- cow caseinate (green), iron- cow caseinate (maroon), calcium- goat caseinate (blue), iron- goat caseinate (teal) at 214 nm, bandwidth 8 nm, Figure 6-24: Protein content of cow and goat peptide eluted fractions in mg/mL; error Figure 6-25: Chromatogram of cow caseinate digest at 214 nm, bandwidth 8 nm, reference 360 nm, bandwidth 50 nm. 176 Figure 6-26: Chromatogram of goat caseinate digest at 214 nm, bandwidth 8 nm, Figure 6-27: Chromatogram of cow washing buffer peptides at 214 nm, bandwidth 8 Figure 6-28: Chromatogram of goat washing buffer peptides at 214 nm, bandwidth 8 Figure 6-29: Chromatogram of cow elution buffer peptides at 214 nm, bandwidth 8 Figure 6-30: Chromatogram of goat elution buffer peptides at 214 nm, bandwidth 8 Figure 6-31: Chromatogram of cow phosphate buffer peptides at 214 nm, bandwidth Figure 6-32: Chromatogram of goat phosphate buffer peptides at 214 nm, bandwidth Figure 6-33: Iron content of cow and goat eluted fractions in μ M; error bars indicate standard error. 182 Figure 6-34: Amino acid content (mg/100mg protein) of caseinates and IMAC

Figure 7-1: Zeta potential of cow and goat phosphopeptides (0.2 mg/mL) after calcium removal with ion exchange; error bars indicate standard error, n=3
Figure 7-2: Fraction of available ferrous iron out of the total iron added as a function of temperature in buffers at pH 2.75, 4.60, 6.7 with 50 mM and 400 mM NaCl in the absence of protein; error bars indicate standard error, n=3
Figure 7-3: Iron bound by cow phosphopeptides as a function of temperature in buffers at pH 2.75, 4.60, 6.70 at 50 mM and 400 mM NaCl after blank subtraction of iron in respective buffer; error bars indicate standard error, n=3
Figure 7-4: Iron bound by goat phosphopeptides as a function of temperature in buffers at pH 2.75, 4.60, 6.70 at 50 mM and 400 mM NaCl after blank subtraction of iron in respective buffer; error bars indicate standard error, n=3
Figure 7-5: Iron binding of cow and goat phosphopeptides after ion exchange with non-detectable calcium content in imidazole buffer, pH 6.7, 50 mM or 400 mM NaCl; error bars indicate standard error, n=3
Figure 7-6: Fraction of available ferrous iron out of the total iron added as a function of temperature in various buffers adjusted at pH 6.70, 50 mM NaCl; error bars indicate standard error, n=3
Figure 7-7: Iron binding of cow and goat phosphopeptides with various buffers at pH 6.70, 50 mM NaCl; error bars indicate standard error, n=3
Figure 7-8: Effect of holding time at different temperatures on blank imidazole, pH 6.70, 50 mM NaCl on the availability of ferrous iron; error bars indicate standard error, n=3.
Figure 7-9: Effect of holding time on the iron binding capacity at various temperatures at pH 6.7, 50 mM NaCl, in the presence of calcium; error bars indicate standard error, n=3.
Figure 7-10: Effect of holding time on the iron binding capacity at various temperatures at pH 6.7, 50 mM, in the absence of calcium; error bars indicate standard error, n=3.
Figure 7-11: Effect of temperature ramping on blank buffers imidazole, pH 6.70, 50 mM with time with respect to iron oxidation; error bars indicate standard error, n=3.

Figure 7-13: Effect of iron loading in blank imidazole buffer on fraction of ferrous iron availability with increasing temperature; error bars indicate standard error, n=3.

Figure 7-16: Binding capacity of cow and goat peptide and milk mixtures with different fractions of peptides and different ordering of addition at a fixed iron loading of 28 μ M in the presence (dialyzed) of calcium; error bars indicate standard error, n=3.

Figure 8-2: Schematic of intact casein proteins (top left) and micelles (top right) surrounding a fat globule with hydrophobic sections interacting while hydrophilic sections partition iron-bound sections away from the fat. Conversely, short hydrophilic peptides (bottom, amino acids represented by coloured circles) that contain a high concentration of iron allow a close proximity of iron to the fat increasing the chance of interaction over time. 230

Figure 8-8: Ferritin response of all cow milk derived treatments on an equal iron filtrate basis of ng Ferritin/ nM iron; error bars indicate standard error, n=7...... 242

Figure 8-9: Ferritin response of all goat milk derived treatments on an equal iron filtrate basis of ng Ferritin/ nM iron; error bars indicate standard error, n=7...... 242

Figure 8-10: Comparison of calcium and iron precipitated cow and goat phosphopeptides on an equal iron filtrate basis; error bars indicate standard error, n=7.

Figure 9-1: Summary of the protein fractions produced and analysed for iron binding and characterisation. 265

Figure B-1: Replicates of pH 4.0, I=0 buffers showing the iron concentration and pH
of the eluents
Figure B-2: Iron concentration and pH of eluents using buffer at low pH (3.0) and low
ionic strength (I=0) sample
Figure B-3: Iron concentration and pH of eluents using buffer using high ionic strength
samples (I=100) with buffer of pH 3.0 or 4.0
Figure B-4: Iron concentration of eluents using an alkaline eluent (buffer) (pH 8.0 or
pH 9.0) with low ionic strength
Figure B-5: Iron leaching of treatments that produced the lowest amount of leaching
during washing and elution

Table of Tables

Table 2-1: Anionic clusters of calcium sensitive caseins with charge by Swaisgood et al., (2008)
Table 2-2: Composition of goat, cow and human milk by Jandal, (1996) and Park et al., (2007)
Table 2-3: Comparison of casein compositions between cow and goat milk expressedas % of total casein12
Table 2-4: Calcium and phosphorus content of cow and goat milk in mg/100g ormg/100mL14
Table 2-5: Proportions of where native iron exists in whole and skim milk, byGaucheron (2000)16
Table 2-6: Iron forms permitted in the use of food according to Hurrell (1985), Allenet al., (2006) and Australia New Zealand Food Standards Code (2013).20
Table 2-7: Characteristics of purified casein phosphopeptides (CPP) adapted fromKibangou et al., (2005)25
Table 2-8: Commercial products derived from casein phosphopeptides for various purposes
Table 3-1: Composition of cow skim milk powder
Table 3-2: Composition of goat skim milk powder
Table 3-3: Cow and goat skim milk powder crude protein content
Table 4-1: Cow and goat skim milk powder crude protein content, n=3
Table 4-2: Z- average size of cow and goat skim milk after centrifugation 10,000 xg, 45 minutes and filtration at 0.22 μ M, n=3
Table 4-3: Zeta potential of cow and goat skim milk at 1 mg/mL in SMUF, n=3 71
Table 4-4: Z-average particle diameter (nm) of cow and goat milk fortified with ferrous sulfate; superscripts with different letters indicate significantly different values at 95% confidence $n \ge 3$.
<i>w > 0 / 0 contractice</i> , if <i>S</i>

Table 4-5: Z-average particle diameter (nm) of wet blend cow and goat milk fortified
with ferrous sulfate; superscripts with different letters indicate significantly different
values at 95% confidence, n>3
Table 6-1: Difference in amino acid composition of cow and goat caseinate versus
cow and goat calcium precipitated peptides
Table 7-1: Comparison of the iron binding capacity of peptides before and after
dialysis in 40 mM sodium formate, pH 3.0 and ion exchanged samples using Amberlite
IRC-50, letters that are the same indicate no significant difference at 95 % confidence,
n=3