

Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

Felis catus Papillomavirus Type 2
Infection and Skin Cancer in
Domestic Cats

A thesis presented in partial fulfilment of the requirements for the
degree of

Doctor of Philosophy

in

Veterinary Science

at Massey University, Manawatū,
New Zealand

Neroli Anne Thomson

2017

Abstract

Felis catus papillomavirus type 2 (FcaPV-2) is a virus which commonly infects the skin of domestic cats. While most infections are asymptomatic, there is growing evidence that FcaPV-2 may play a role in the development of a subset of feline cutaneous squamous cell carcinomas (SCCs).

In the first part of this thesis, the natural history of FcaPV-2 infection was investigated with the aim of determining when cats become infected with the virus. A real-time PCR assay was developed to quantify FcaPV-2 DNA in feline skin swabs. This assay was then used to measure the FcaPV-2 DNA load in serial samples from two populations of cats. Results from these studies showed that most kittens are exposed to FcaPV-2 in the first few days of life. Additionally, the primary source of exposure is likely to be direct contact with other cats in the household, particularly their queen, as some of the queens appeared to be shedding large amounts of virus. FcaPV-2 mRNA was also detected in some of the kittens, confirming that they had become infected with FcaPV-2 soon after birth.

The aim of the second part of this thesis was to determine the quantity and transcriptional activity of the FcaPV-2 DNA present in feline cutaneous SCCs in order to determine if the virus was involved in cancer development or just present as an innocent bystander. Real-time PCR assays were developed to measure FcaPV-2 gene expression in SCCs and the results clearly distinguished two subsets of feline cutaneous SCCs. The majority of the SCCs had low copy numbers of FcaPV-2 DNA and no FcaPV-2 gene expression, suggesting the virus was an incidental finding. In contrast, around a third of the SCCs had detectable FcaPV-2 gene expression and high copy numbers of FcaPV-2 DNA, similar to that found in the FcaPV-2-induced premalignant lesions. There was also a significant association between FcaPV-2 gene expression and alterations in a host cell cycle regulatory protein (p16). Taken together, these results strongly suggest that FcaPV-2 played a role in the development of around a third of the feline cutaneous SCCs.

The results from the studies reported in this thesis support a causative role of FcaPV-2 in a proportion of feline cutaneous SCCs. However, as infection of cats is common and appears to occur early in life, there may be little opportunity to prevent SCC development by preventing FcaPV-2 infection.

Acknowledgements

The journey to the publication of this thesis, while trying at times, has been made worthwhile by the wonderful people I have met along the way.

My utmost thanks go to my primary supervisor John Munday, who supported me from start to end, and mostly managed to stop me from taking myself too seriously along the way. In all seriousness though John, you've been an excellent supervisor, thank-you.

I am also very grateful to Keren Dittmer and Magda Dunowska, for their enthusiasm, patience and technical skills. Countless times I barged into Keren's office eager to share a new idea or result, before realising that she probably had more important work to do. Thank-you Keren for sharing my excitements and frustrations. To Magda, I am most grateful for the time and effort you put in to help me get the first real-time PCR assay working well, it proved to be immensely useful.

This thesis would not have been possible without the many people who helped me collect samples. In particular, I am very grateful to the people who let me collect swabs and hair plucks from their cats, and to the veterinarians who sent me biopsies. Genevieve Rogerson and Robyn Jarrett deserve special mention, thank-you both. The staff at the Feline Centre for Nutrition at Massey University, and Adrienne French, at New Zealand Pathology Limited, were also very helpful in this regard.

I would also like to acknowledge the financial support I received from a Massey Doctoral Scholarship and from the grants which funded the research presented in this thesis, including the Morris Animal Foundation, Maurice and Phyllis Paykel Trust, IVABs Postgraduate Research Fund and the Massey University Research Fund.

Finally, I could not have done this without the love and support of my family, especially my mother Barbara, who provided lots of moral support and proofread this thesis. Nor could I have done this without the encouragement of my partner Sam; thank-you Sam for all the ways you made me smile, even when I was tired and grumpy, throughout this journey.

Table of Contents

Chapter 1: Literature Review	1
1.1 Introduction to papillomaviruses	3
1.1.1 Papillomavirus structure and taxonomy	3
1.1.2 The spectrum of PV-induced disease	5
1.2 Epidemiology of PV infection	9
1.2.1 The high-risk HPVs	9
1.2.2 Cutaneous HPVs	12
1.2.3 Epidemiology of PV infection in animals	14
1.3 Molecular mechanisms.....	17
1.3.1 The normal PV lifecycle	17
1.3.2 High-risk HPV-induced cancer	21
1.3.3 Host factors in high-risk HPV-induced cancer	25
1.3.4 Beta HPVs and skin cancer	28
1.3.5 PV-induced cancer in animals.....	30
1.3.6 Summary of the natural history of PV infection.....	33
1.4 PV infections of domestic cats	35
1.4.1 The discovery of PV-induced lesions in cats.....	35
1.4.2 Early studies on feline viral plaques and Bowenoid <i>in situ</i> carcinomas	36
1.4.3 PCR and sequencing of the first feline PV	37
1.4.4 PCR of papillomas, FVPs and BISCs.....	38
1.4.5 Sequencing of the second feline PV	39
1.4.6 FcaPV-2 in squamous cell carcinomas.....	40
1.4.7 Sequencing of the third and fourth feline PVs	42
1.4.8 Feline PVs in oral lesions	43
1.4.9 Alterations in host cell cycle regulatory proteins p16, pRb and p53	44
1.4.10 Feline sarcoids	47
1.5 Conclusion	49
1.6 References.....	51
Chapter 2: Timing and Source of Exposure to FcaPV-2.....	69
2.1 Introduction	69
2.2 Methods	71
2.2.1 Sample collection	71
2.2.2 DNA extraction	71

2.2.3 Generation of recombinant plasmids as standards for qPCR	72
2.2.4 Quantitative PCR assay	73
2.2.5 Quantitative PCR assay validation	74
2.2.6 Normalisation.....	74
2.2.7 Statistical analysis	75
2.3 Results.....	76
2.3.1 Sample collection	76
2.3.2 Performance of the qPCR assay	76
2.3.3 Testing of the samples	78
2.4 Discussion	82
2.5 References.....	86
 Chapter 3: Timing of FcaPV-2 Infection in a Cat Colony.....	89
3.1 Introduction.....	89
3.2 Methods	91
3.2.1 Selection of cats	91
3.2.2 Swab collection for DNA analysis.....	91
3.2.3 Sampling time points	91
3.2.4 Swabs for RNA analysis	92
3.2.5 DNA and RNA extraction	93
3.2.6 Real-time PCR.....	93
3.2.7 Statistical analysis	94
3.3 Results.....	95
3.3.1 FcaPV-2 DNA load in the swab samples.....	95
3.3.2 FcaPV-2 RNA in swab and tissue samples.....	97
3.4 Discussion	99
3.5 References.....	103
 Chapter 4: FcaPV-2 Gene Expression in SCCs.....	105
4.1 Introduction.....	105
4.2 Methods	107
4.2.1 Sample Collection.....	107
4.2.2 RNA extraction and cDNA synthesis	107
4.2.3 Development of real-time PCR assays for feline reference genes.....	108
4.2.4 Reference gene validation study.....	109
4.2.5 Development of real-time PCR assays for FcaPV-2 gene expression.....	110

4.2.6 Quantification of FcaPV-2 DNA	110
4.2.7 Immunohistochemistry for p16.....	111
4.2.8 Analysis of real-time PCR data.....	111
4.3 Results	112
4.3.1 Sample collection	112
4.3.2 Performance of the real-time PCR assays	112
4.3.3 Reference gene selection	113
4.3.4 FcaPV-2 gene expression.....	115
4.3.5 Quantification of FcaPV-2 DNA	116
4.3.6 Immunohistochemistry for p16.....	117
4.4 Discussion.....	119
4.5 References.....	122
 Chapter 5: FcaPV-2 Gene Expression in FFPE SCCs.....	125
5.1 Introduction	125
5.2 Methods	127
5.2.1 Sample collection	127
5.2.2 RNA extraction and cDNA synthesis.....	127
5.2.3 Real-time PCR assays for FcaPV-2 gene expression	128
5.2.4 Reference genes	128
5.2.5 Quantification of FcaPV-2 DNA	131
5.2.6 Immunohistochemistry for p16.....	131
5.2.7 Statistical analysis.....	131
5.3 Results	132
5.3.1 Sample selection and quality.....	132
5.3.2 FcaPV-2 gene expression.....	133
5.3.3 FcaPV-2 viral load	136
5.3.4 P16 immunohistochemistry	137
5.4 Discussion.....	139
5.5 References.....	143
 Chapter 6: The Physical State of FcaPV-2 DNA in SCCs	145
6.1 Introduction	145
6.2 Experiment 1 Methods	147
6.2.1 Identification of repeat elements in the cat genome	147
6.2.2 Samples and DNA extraction	148

6.2.3 Long range PCR	149
6.2.4 Next generation sequencing.....	150
6.3 Experiment 1 Results and Interpretation	151
6.3.1 Long-range PCR	151
6.3.2 Next generation sequencing.....	152
6.4 Experiment 2 Methods	156
6.4.1 Sample selection	156
6.4.2 Nested PCR.....	157
6.5 Experiment 2 Results and Interpretation	158
6.6 Discussion	160
6.7 References.....	165
 Chapter 7: General Discussion	167
7.1 The timing of FcaPV-2 infection in cats.....	168
7.2 Comparison of the natural history of FcaPV-2 infection with other species	170
7.3 FcaPV-2 gene expression in SCCs	173
7.4 Cumulative evidence for a role of FcaPV-2 in feline skin cancer.....	176
7.4.1 Transitional lesions between PV-induced premalignant lesions and cancer	176
7.4.2 The detection of PV DNA in a high proportion of cancers.....	176
7.4.3 PV infection as a risk factor for the development of cancer	177
7.4.4 PV-induced immortalisation of cell lines	178
7.4.5 Interactions between PV and host proteins	179
7.4.6 Deregulated PV E6/E7 gene expression.....	180
7.4.7 PV-induced cancer models in transgenic mice	181
7.4.8 PV vaccine trials	182
7.5 Reducing the incidence of FcaPV-2 associated disease	183
7.5.1 Vaccination.....	183
7.5.2 PV-targeted immunotherapy.....	186
7.6 Final summary	188
7.7 References.....	190
 Appendix A: List of Publications and Statements of Contribution	195

List of Figures

Chapter 1

Figure 1.1. Bovine Papillomavirus Capsid	3
Figure 1.2. Genome organisation of <i>Felis catus</i> papillomavirus type 2 (FcaPV-2) and human papillomavirus type 16 (HPV-16), with accession numbers and genome size.....	3
Figure 1.3. Schematic representation of PV genera	4
Figure 1.4. Cutaneous papilloma from a dog.....	5
Figure 1.5. Patterns of PV gene expression in the normal PV lifecycle	18
Figure 1.6. High-risk HPV gene expression in an in situ cancer (CIN3)	22
Figure 1.7. Proposed mechanism of PV-induced cancer.....	24

Chapter 2

Figure 2.1. Amplification plot and standard curve for the newly developed qPCR FcaPV-2 assay	77
---	----

Chapter 3

Figure 3.1. Copies of FcaPV-2 DNA per swab over time	96
---	----

Chapter 4

Figure 4.1. Reference gene stability in NormFinder	113
Figure 4.2. Candidate reference genes ranked according to their GeNorm M values after the removal of RPS7 and RPL17 (lower values indicate higher stability).	115
Figure 4.3. p16 immunohistochemistry.....	118

Chapter 5

Figure 5.1. Reference gene stability in NormFinder	129
Figure 5.2. Candidate reference genes ranked according to their GeNorm M values (lower values indicate higher stability).	130
Figure 5.3. Log normalised relative quantity of FcaPV-2 E6/E7 and L1/L2 gene expression in premalignant lesions and SCCs that had detectable FcaPV-2 gene expression (pos-SCCs)	136
Figure 5.4. Log transformed viral load (copies of FcaPV-2 DNA per copy of reference gene DNA) in FcaPV-2 E6/E7-positive and -negative SCCs, premalignant lesions and normal skin SCCs	137
Figure 5.5. p16 immunohistochemistry and haematoxylin and eosin (H&E) stained sections	138

Chapter 6

Figure 6.1. Proportion of the cat (<i>F. catus</i>) genome that is comprised of repeated elements and low complexity DNA sequences	147
Figure 6.2. Long-range PCR products after gel electrophoresis.....	151
Figure 6.3. The intense 11 kbp band for SCC 1	152
Figure 6.4. Manually annotated contigs from NGS relative to the published genome sequence for FcaPV-2 (EU_796884)	155
Figure 6.5. Gel electrophoresis of FcaPV-2 reverse transcribed mRNA transcripts.....	158

List of Tables

Chapter 1

Table 1.1. Rates of detection of PV DNA from SCCs using PCR (PV positive/ total tested)	41
--	----

Chapter 2

Table 2.1. Coefficient of variation (%) as a measure of precision (intra-assay) and reproducibility (inter-assay) of the newly developed FcaPV-2 assay	78
Table 2.2. FcaPV-2 DNA detected in swab and hair pluck samples at the different time points	78
Table 2.3. <i>Felis catus</i> papillomavirus type 2 (FcaPV-2) DNA loads in swabs and hair pluck samples from individual cats	81

Chapter 3

Table 3.1. FcaPV-2 early (E7) and late (L1) gene expression in swab samples	98
Table 3.2. FcaPV-2 early (E7) and late (L1) gene expression in post-mortem tissue samples.....	98

Chapter 4

Table 4.1. Reference gene primers and assay performance.....	109
Table 4.2. FcaPV-2 DNA load, p16 immunostaining and FcaPV-2 gene expression in SCCs and controls.....	116
Table 4.3. FcaPV-2 DNA copy number and E6/E7 expression compared to p16 status	117

Chapter 5

Table 5.1. FcaPV-2 E6/E7 gene expression by SCC location.....	132
Table 5.2. Details of individual FFPE SCCs.....	134
Table 5.3. Details of individual samples: premalignant skin lesions and normal skin samples.....	135

Chapter 6

Table 6.1. FcaPV-2 DNA load, p16 immunostaining and FcaPV-2 gene expression of selected SCCs from chapter 4.....	149
Table 6.2. Single nucleotide polymorphisms between the two contigs and the previously published sequence for FcaPV-2 (EU796884)	154
Table 6.3. FcaPV-2 DNA load, p16 immunostaining and FcaPV-2 gene expression in SCCs and controls.....	157

Abbreviations

Common Abbreviations

28Sr	Reference gene coding for the 28s ribosomal sub-unit
ABL2	Abelson proto-oncogene 2 non-receptor tyrosine kinase RNA reference gene
ACTB	Beta actin RNA reference gene
ANOVA	Analysis of variance statistical method
B2M	Beta-2 microglobulin RNA reference gene
BISC	Bowenoid in situ carcinoma
CI	Confidence interval
CIN	Cervical intraepithelial neoplasia
Cq	Number of PCR cycles when threshold reached
CV	Coefficient of variation
DSH	Domestic short hair
EV	Epidermodysplasia verruciformis
FFPE	Formalin fixed paraffin embedded
FIV	Feline immunodeficiency virus
FVP	Feline viral plaque
GAPDH	Glyceraldehyde 3-phosphate dehydrogenase RNA reference gene
GUSB	Beta glucuronidase RNA reference gene
H&E	Haematoxylin and eosin stain
IgG	Immunoglobulin G
MHC	Major histocompatibility molecules
NRQ	Normalised relative quantity
ORF	Open reading frame
p16	Cyclin dependant kinase inhibitor p16 ^{INK4A}
p53	Tumour suppressor p53 protein
pRb	Retinoblastoma protein
PCR	Polymerase chain reaction
PV	Papillomavirus
qPCR	Quantitative PCR
RPL17	Ribosomal protein L17 RNA reference gene
RPS7	Ribosomal protein S7 RNA reference gene
RPS19	Ribosomal protein S19 RNA reference gene
RT	Reverse transcriptase
SCC	Squamous cell carcinoma
SNP	Single nucleotide polymorphism
VLP	Virus-like particle
YWHAZ	Tyrosine 3-monooxygenase/ 5 tryptophan 5-monooxygenase activation protein zeta

Papillomavirus Abbreviations

		Phylogeny- genus	Tissue infected
Domestic cats			
FcaPV-1	<i>Felis catus</i> papillomavirus type 1 Formerly <i>Felis domesticus</i> papillomavirus 1	Lambdapapillomavirus	oral mucosa
FcaPV-2	<i>Felis catus</i> papillomavirus type 2 Formerly <i>Felis domesticus</i> papillomavirus 2	Dyothetapapillomavirus	skin
FcaPV-3	<i>Felis catus</i> papillomavirus type 3	Taupapillomavirus	skin
FcaPV-4	<i>Felis catus</i> papillomavirus type 4	Taupapillomavirus	unknown
Humans			
HPV-1	Human papillomavirus type 1	Chipapillomavirus	skin
HPV-2	Human papillomavirus type 2	Alphapapillomavirus	skin
HPV-4	Human papillomavirus type 4	Gammapapillomavirus	skin
HPV-5	Human papillomavirus type 5	Betapapillomavirus	skin
HPV-6	Human papillomavirus type 6	Alphapapillomavirus	genital mucosa
HPV-8	Human papillomavirus type 8	Betapapillomavirus	skin
HPV-9	Human papillomavirus type 9	Betapapillomavirus	skin
HPV-11	Human papillomavirus type 11	Alphapapillomavirus	genital mucosa
HPV-16	Human papillomavirus type 16*	Alphapapillomavirus	genital mucosa
HPV-17	Human papillomavirus type 17	Betapapillomavirus	skin
HPV-18	Human papillomavirus type 18*	Alphapapillomavirus	genital mucosa
HPV-27	Human papillomavirus type 27	Alphapapillomavirus	skin
HPV-38	Human papillomavirus type 38	Betapapillomavirus	skin
HPV-57	Human papillomavirus type 57	Alphapapillomavirus	skin
HPV-76	Human papillomavirus type 76	Betapapillomavirus	skin
HPV-93	Human papillomavirus type 93	Betapapillomavirus	skin
Domestic dogs			
CPV-1	<i>Canis familiaris</i> oral papillomavirus Formerly COVP	Lambdapapillomavirus	oral mucosa
CPV-2	<i>Canis familiaris</i> papillomavirus type 2	Taupapillomavirus	skin
Domestic cattle			
BPV-1	<i>Bos taurus</i> papillomavirus type 1	Deltapapillomavirus	skin
BPV-2	<i>Bos taurus</i> papillomavirus type 2	Deltapapillomavirus	skin
BPV-3	<i>Bos taurus</i> papillomavirus type 3	Xipapillomavirus	skin
BPV-4	<i>Bos taurus</i> papillomavirus type 4	Xipapillomavirus	oral/ oesophageal mucosa
BPV-13	<i>Bos taurus</i> papillomavirus type 13	Deltapapillomavirus	skin
BPV-14	<i>Bos taurus</i> papillomavirus type 14	Deltapapillomavirus	skin
Horses			
EcPV-2	<i>Equus caballus</i> papillomavirus type 2	Dyiotapapillomavirus	genital mucosa
Rabbits			
SfPV-1	<i>Sylvilagus floridanus</i> papillomavirus type 1 Formerly cottontail rabbit papillomavirus	Kappapapillomavirus	skin
OcPV-1	<i>Oryctolagus cuniculus</i> papillomavirus type 1	Kappapapillomavirus	oral mucosa
Mice			
MnPV-1	<i>Mastomys natalensis</i> papillomavirus type 1	Iotapapillomavirus	skin
MmuPV-1	<i>Mus musculus</i> papillomavirus type 1	Pipapillomavirus	skin