

Copyright is owned by the Author of the thesis. Permission is given for a copy to be downloaded by an individual for the purpose of research and private study only. The thesis may not be reproduced elsewhere without the permission of the Author.

# ACARICIDE RESISTANCE AND GENETIC AFFINITIES OF SOME SELECTED POPULATIONS OF <u>TETRANYCHUS</u> <u>URTICAE</u> KOCH

IN NEW ZEALAND.

A thesis

presented in partial fulfilment

of the requirements for the Degree

of

Master of Horticultural Science

in

Entomology

at Massey University, New Zealand.

S.H. Ong 1973

#### ABSTRA CT

A study of resistance to acaricides in a number of populations of the two-spotted spider mite, Tetranychus urticae, in New Zealand had been carried out. Natural genetic and cytoplasmic incompatibilities between populations were also investigated with a view to possible biological control of the pest. Facets of acaricide resistance that were studied included multi-resistance, crossresistance, negatively correlated resistance and the inheritance of resistance. Chemicals used included an organophosphate representative (parathion-methyl), a carbamate (formetanate), an ungrouped compound (tricyclohexyltin hydroxide) and an organochlorine (dicofol). Cross-resistance was demonstrated between parathion-methyl and formetanate in five populations obtained from widely separate areas of New Zealand. The resistance to parathion of three strains was found to be inherited as a single dominant character and transmissible by both sexes. Cytoplasmic factors (or nucleo-cytoplasmic interactions) and minor genes were found to contribute slightly to the expression of total resistance. No resistance to tricyclohexyltin hydroxide (Plictran) and dicofol (Kelthane) was detected.

High degrees of incompatibility (haploid egg lethality) were observed in the hybrids of crosses between the various populations. Chromosomal rearrangements in balanced heterozygous conditions, in conjunction with the cytoplasm, were considered to be important factors determining the interpopulational sterilities. The interpopulational incompatibility phenomenon was found to be multifactorial and not associated with the resistance factor. The egg

ii

mortalities of some backcross series which remained constantly high in spite of several crossings, implicated that the introduction of normal males to a resistant mite population in an enclosed area (e.g. in a glasshouse) might be a worthwhile proposition in the integrated control of spider mites. Backcross hybrids, on allowing to multiply randomly, were capable of forming new gene combinations, leading consequently to the formation of new strains which were genetically different from the original parents used in the backcross series.

### ACKNOWLEDGEMENTS

I would like to sincerely thank Dr. G.H. Ballantyne (Lecturer, Microbiology and Genetics, Massey University) for his supervision and assistance in the research project and the writing up of this thesis.

My gratitude also goes to the Levin Horticultural Research Institute, Mr. Lines (Levin), Mr. D. Slade (Lecturer in Horticulture, Massey University) and Dr. G.H. Ballantyne who help in one way or another in supplying the various mite samples for the present research.

I would also like to express my appreciation to Ivon-Watkins-Dow Ltd. for kindly supplying some of the miticides.

Finally, I would like to thank the Microbiology and Genetics Department, Massey University, for allowing me access to the facilities in the laboratories.

iv

## CONTENTS

																PAGE
ABSTRACT			• •	•	•					•	•	•	•	•		ii
A CKNOWLEDGEMENT	5		•••	•	•	•••	•		• •	•	•	÷	•		•	iv
LIST OF FIGURES		• •		٠	•	•••	•	•		•	•	•	•	•	• ,	viii
LIST OF TABLES			•••	•	•	• •				•	•	•	ł	•	•	ix
LIST OF PLATES	• • • •	•••	•••	•	•	• •		•	• •	•	•	•	•	•	•	x
CHAPTER 1 IN	TRODUCT	ION	• •	٠	•	•••	•	•	• • •	٠	•	•	÷	•	•	1
CHAPTER 2 RE	VIEW OF	THE	LIT	ERA	TUI	RE	•	•	•••	•	•	•	•	•	•	4
2.	l In	trodi	ucti	on	•	• •	•	•	• •	٠	•	٠	•	•	•	4
2.	2 De:	fini	tion	S						•	•	•	•		•	4
2.	3 Or:	igin	of	res	ist	tan	ce			•			•	•	•	7
2.	4 Der	velop	omen	t c	of 1	res	ist	tan	ce					•	•	10
	2.	4.1	Pat men											-		10
	2.	4.2	Fac	tor	'S 8	aff	ect	tin	g t	he	de	eve	10	p-q		
			men	t c	of 1	res	ist	tan	ce.	•	•	•	•	٠	•	11
2.	5 Sta	abil:	ity	of	res	sis	tar	nce	•	•	٠	٠	•	•	•	14
2.		e dos	1000				12.07.5				nte	erp	re	eta	-	
		on ai									•	• •	•	Ċ.	•	18
		5.1	in	the	DN	1-1	ine	э.								20
	2.0	5.2	Lim mor													21
	2.0	5.3	Mis	int	er	pre	tat	tic	n o	of t	the	r	es	sp-	•	
	2.0	5.4	ons DM-	lin	ie :	in (	ger	net	ic	sti	ıdi	les		of		22
0	<b>a</b> 0		res													22
2.		neti														23
2.		ocher	nica	1 g	ene	eti	CS	of	re	si	sta	nc	е	•	•	25
2.	9 Bai	sic (	gene	tic	s	of	spi	ide	r m	ite	es	•	•	•	•	27
2.	10 In	compa	atib	ili	ty	in	Τ.	ur	tic	ae	•	•	•	٠	•	29
2.	ll Ser	<b>x-r</b> a	tio	in	<u>T.</u> 1	urt:	ica	<u>le</u>	• •	•	•	•	•	•	•	33
2.	12 Se:	x-de	term	ina	ti	on	in	<u>T</u> .	urt	ica	10	•	•	•	·	35
2.	13 Me	thod	s of	ge	enet	tic	c	ont	rol	•		•	•	•	•	37
2.	14 Gen	neti	c in	com	ipa-	tib	ili	ity	as	a	pc	oss	ił	le	)	
	me	ans (	of c	ont	rol	l i	n s	spi	der	m	ite	s		•		40

ν

LIFE HISTORY AND BEHAVIOUR OF MITES . . . . CHAPTER 3 42 3.1 Life cycle of T.urticae . . . . . . . 42 Environmental factors affecting develop-3.2 45 Seasonal life cycle and behaviour of 3.3 46 CHAPTER 4 49 The strains of spider mites . . . . . 4.1 49 4.2 Maintainance, isolation and disposal of 49 4.3 Toxicological testing . . . . . . . . 51 Selection technique (leaf-dip). . . . . 4.4 53 4.5 Mother x son inbreeding . . . . . . . 54 4.6 Mite crossing and handling methods: Reciprocal crosses . . . . . . . . . 56 4.7 Backcrossing with selection . . . . . 57 4.8 Construction of DM-lines. . . . . . . 58 Acaricidal Materials. . . . . . . . . . 4.9 58 CHAPTER 5 60 5.1 Toxicological study . . . . . . . . 60 Toxicological response of the 5.1.1 original strains . . . . . . . 60 Toxicological response of the 5.1.2 homozygous backcrossed resistant strains. . . . . . . . . . . . . . 67 Toxicological response of the 5.1.3 homozygous backcrossed strain 74 5.2 Inheritance of resistance . . . . . . 74 5.2.1 Reciprocal crosses between resistant and normal strains . . . . 75 5.2.2 Backcrosses between the resistant F1 and normal . . . . . . 80 Repeated backcrosses with selec-5.2.3 80 tion . . . . . . . . . . . . . . . . Incompatibility . . . . . . . . . . . 5.3 84 5.3.1 Intrapopulational crosses. . . . 84 Interpopulational crosses. . . . 5.3.2 86 Repeated backcrossing with selec-5.3.3 89 tion . . . . . . . . . . . . . . Intrapopulational and interpop-5.3.4 ulational crosses of hemozygeus backcrossed resistant strains . .

PAGE

94

PAGE

CHAPTER	6	DISCL	ISSION .	•	•	•••			•	•	•	•	•	•	•	•	•	•	98
		6.1	Toxico	108	gica	al 1	rea	cti	ons	5	•	•	•	•	•	•			93
		6.2	Geneti																
			bility	٥	•	•••	•	• •	•	•	•		٠	•	•	•	•	•	103
CHAPTER	7	CONCI	USIONS	•	•	•••		• •	•	•	•	•	•	•	•	•	•	•	111
REFERENCE	S			•	•		•		•	•	•	•	•	•	•	•	•	•	114
APPENDIX	1	• • •		•	•	•••	•		•	•	•	•	•	•	•	•	•	•	129
APPENDIX :	2															•		٠	141

vii

## LIST OF FIGURES

FIGUR	E	PAGE
l.	Toxicity of parathion-methyl to several isolates of <u>T.urticae</u>	61
2.	Toxicity of formetanate to several isolates of <u>T.urticae</u>	64
3.	Toxicity of tricyclohexyltin hydroxide to several isolates of <u>T.urticae</u>	66
4.	Response of the homozygous backcrossed resistant strains to parathion-methyl	68
5.	Response of the homozygous backcrossed resistant strains to formetanate	70
6.	Response of the homozygous backcrossed resistant strains to tricyclohexyltin hydroxide	73
7.	Dosage-mortality lines for the F, progeny LINES x PN reciprocal crosses	76
8.	Dosage-mortality lines for the F <sub>1</sub> progeny LHRS x PN reciprocal crosses	77
9.	Dosage-mortality lines for the F <sub>1</sub> progeny SF x PN reciprocal crosses	78
10.	Dosage-mortality lines for the B, progeny F (PN4 x LINES ) x PNO F (PN0 x LINES ) x PNO	81
11.	Dosage-mortality lines for the B <sub>1</sub> progeny F1(PN\$ x LHRS&) x PN& F <sub>1</sub> (PN& x LHRS\$) x PN&	82
12.	Dosage-mortality lines for the B <sub>1</sub> progeny F <sub>1</sub> (PN <sup>2</sup> x SP3) x PN3 F <sub>1</sub> (PN3 x SP2) x PN3	83
13.	Graph showing the change in percentage egg sterility with increasing number of backrosses	91
14.		• 92

viii

## LIST OF TABLES

TABLE		PAGE
l.	Summary of the dosage-mortality response of the PN, ETTR, LHRS, LINES and SP strains of <u>T.urticae</u> to parathion-methyl	62
2.	Summary of the dosage-mortality response of the PN, ETTR, LHRS, LINES and SP strains of <u>T.urticae</u> to formetanate	65
3.	Summary of the dosage-mortality response of the PN, ETTR, LHRS, LINES and SP strains of <u>T.urticae</u> to tricyclohexyltin hydroxide (Plictran)	65
4.	Summary of the dosage-mortality response of the HNR.PN, SP.PN, LHRS.PN and LINES.PN backcrossed strains to parathion-methyl	69
5.	Summary of the dosage-mortality response of the HNR.PN, SP.PN, LHKS.PN and LINES.PN backcrossed strains to formetanate	71
6.	Summary of the dosage-mortality response of the HNR.PN, SP.PN, LHRS.PN and LINES.PN backcrossed strains to tricyclohexyltin hydroxide (Plictran)	72
7.	Summary of the dosage-mortality data for the F <sub>1</sub> progeny from reciprocal crosses of the LINES and PN strains to parathion-methyl	75
8.	Summary of the dosage-mortality data for the F <sub>1</sub> progeny from reciprocal crosses of the LHRS and PN strains to parathion-methyl	79
9.	Summary of the dosage-mortality data for the F <sub>1</sub> progeny from reciprocal crosses of the SP and PN strains to parathion-methyl	79
10.	Intrapopulational and interpopulational crosses of six strains of <u>T.urticae</u>	85
11.	Intrapopulational and interpopulational crosses of selected homozygous backcrossed resistant strains of <u>I.urticae</u>	95
12.	Intrapopulational and interpopulational crosses of selected homozygous backcrossed resistant strains of <u>T.urticae</u>	96

E

## LIST OF PLATES

PLATE		PAGE
l.	Leaf cultures on a plastic tray	52a
2.	Trays of leaf cultures kept in long daylength environment	52a
3.	Glass slides with double-sticking tapes containing mites placed dorsally	52 b

x

#### CHAPTER 1

#### INTRODUCTION

'Can insects become resistant to sprays?. That now historical question was asked by Melander (1914) in 1914. Following the introduction of DDT in the early 1940s, the answer was clearly, yes, and the problem of resistance has had a profound influence ever since on the orientation of entomological research. Such influence is obvious since the development of resistant populations of insects will threaten man's hopes for improvement of his health standards and protection of his food reserves. While only 8 insect species were known to have developed resistance prior to 1940 (Brown, 1961a), the number of resistant strains began a sharp upward trend soon after the introduction and use of DDT and other synthetic organic insecticides. The history of insecticide resistance since then virtually parallels the history of insecticide development. Up to 1967, resistance had developed in 224 species of insects and acarines. Of these 97 are of public health or veterinary importance and 127 attack field or forest crops or stored products (Brown, 1972).

In similar vein, the control of spider mites (Acarina: Tetranychidae) did not constitute a problem until about two decades ago when resistance to agricultural insecticides became widespread.

The first case of resistance in mites probably occurred in 1937, when Compton and Kearns (1937) found inadequate control by Selocide sprays of a two-spotted spider mite population. Selocide resistance was a prelude to the unlimited resistance development which set in after the war, following the use of synthetic insecticides. In many situations, the Tetranychids were promoted from the role of a minor pest to that of a major one as a result of the use of DDT (Helle, 1965a).

An extraordinary genetic potential to adapt to various environments, plus subjection to the high selection pressure encountered in the commercial growing areas, are factors that make the frequency of the resistance phenomenon high in the family Tetranychidae. For these very reasons, the two-spotted spider mite, <u>Tetranychus</u> <u>urticae</u>, has recently been the most difficult to control among all the pests that confront horticulturists and agriculturists (Naegele and Jefferson, 1964). In spite of control measures, many orchardists and ornamentalists suffer economic loss due to defoliation, reduced tree vigour, poor fruit colour, or small fruit brought on as a result of mite feeding. The major problem in the chemical control of spider mites throughout the world is the continued development of strains resistant to the common chemical compounds. Each year, the problem becomes more severe with the number of non-effective types ever increasing, especially the organophosphorus compounds.

The resistance of insects of medical interest, such as <u>Anopheles, Aedes</u> and <u>Musca</u>, had been extensively studied for many years (Brown, 1960). Research on the resistance of agricultural pests had been done but on a smaller scale. As the chemical control of spider mites threatens to develop into a neck and neck race between the chemical industry and the resistance response of the mites, the desirability of an exhaustive investigation into the biological background of the organism, and the physiological and genetical base of the resistance become evident.

Genetic principles and methodology have been invaluable both from the point of view of understanding the development, spread and regression of resistance, and in providing pure strains for funda-

2

mental investigations on the interrelationships of genes, enzymes and toxicological responses to insecticides. From the practical standpoint, knowledge of the genetic identity of phenotypes has made possible the detection of genes for resistance in field strains prior to the use of insecticides or during the course of control operations, thus indicating the advisability of change to another insecticide. Additionally, information from hybridization, indicating reproductive barriers or genetic isolation, obtained during the course of studies on the genetics of resistance, has generated considerable interest in the feasibility of genetic control of insect populations.

With the various problems in mind, the aims of the present research are:

- to determine the distribution of acaricide resistance among the two-spected spider mite populations in certain selected areas of New Zealand.
- to determine the effectiveness of particular groups of acaricides under laboratory conditions.
- to study cross-resistance, multiple-resistance and negatively-correlated resistance patterns in the resistant populations chosen.
- to study the mechanism of inheritance of the resistances in these populations.
- 5) to study the patterns of genetic and cytoplasmic incompatibility that occurs among the chosen populations of spider mites.

3