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# DEVELOPMENT AND RESPONSES OF EQUINE MUSCULO-SKELETAL CONNECTIVE TISSUES

A Collection of Published Papers Presented in Application

for the

Degree of Doctor of Science at Massey University

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November 2006

# Declaration

I declare that the material to be examined in this thesis has not been submitted by me to any other university for the award of any degree.

> Elwyn C Firth November 2006

## Acknowledgements

The scientific papers presented here show my interest in a particular area of equine research, and also in my active collaboration with many different colleagues over a period of some 25 years. The work was conducted when I was a staff member at Utrecht University, The Netherlands, and it continued at Massey University after I returned to New Zealand in 1988.

The part played by others in the work recorded here is indicated by their authorship or in the acknowledgement section of the papers. Although in almost all cases the papers are the result of studies which I conceived, planned and was actively engaged in, more than half do not bear my name as first author. This is because almost all the work involved collaboration with colleagues outside my own academic organisation and country of residence, and much of it involved postgraduate students who were engaged in research projects which I supervised.

A most enjoyable feature of research is engaging with other minds, and I thank all with whom I have had the great pleasure of working in this way. The joy of discovering new things is exceeded only by the pleasure of discovering them with other scientists. I state this here, because such fulfilment is rarely communicated to young people, who apparently find it difficult to contemplate enjoyment in science when they are choosing a career.

For their assistance and collaboration in my research work, I express my deep appreciation to all past and present staff in the above institutions and in other groups in which I have worked. In particular, I thank Professors J T Vaughan, A W Kersjes, Neil Waters, Des Feilden and Grant Guilford, the first three of whom appointed me, and all of whom have been generous in assisting with access to university facilities and have encouraged and supported me in research in many different ways.

Finally, for their love and support I thank my dear family, Sharmian, Jos and Tom, to whom I dedicate this work.

#### Introduction

This application for assessment for examination for the Doctor of Science degree (Massey University) consists of a collection of papers published in international peerreviewed scientific journals. The work was conducted over the last 25 years at Utrecht University, The Netherlands and Massey University, New Zealand. Also in the list of publications are papers (#) originally published in my PhD thesis (1983), and peer-reviewed abstracts of selected presentations at international conferences (\*). They are included not for assessment, but to provide the reader with an insight into the origins of the work and the direction in which it is continuing.

My research focus began to take shape in the 1980s, when I first became interested in the reasons for the site predisposition of infective disease within bone, and in the differences in the responses of bone to mechanical forces. This collection of papers thus focuses on the responses of the musculo-skeletal connective tissues - bone, tendon and cartilage - to various influences. This research area remains important to both equine and human health, since the ways in which connective tissues respond to growth, disease, exercise and lack of exercise largely determine their subsequent structural integrity, and if and when they will be injured during the exertions of athletic training and competition. The work presented here does not include my research in fields less related to the subject of the development and responses of connective tissues.

### **Summary**

The focus of these studies has been the response of bone, cartilage and tendon to growth, exercise, and infective and aseptic disease. Study of the behaviour of these connective tissues has centred on their responses to various stimuli, which can be classified as either natural (for instance, infective disease, aseptic disease, growth, natural exercise), or induced under laboratory conditions (forced exercise, alterations in nutrition).

This collection of papers, entitled "Development and responses of equine musculo-skeletal connective tissues", thus has three major sections: infection, cartilage development, and exercise. However, division of the papers into separate chapters would be inappropriate because these influences overlap to some extent. For instance, normal growth and development requires exercise, and the propensity to cartilage and bone infection in foals is related to the stage of maturation of the tissues, which in turn may be related to exercise.

The publications are presented in the order in which the candidate successively conducted studies of how the above stimuli act on connective tissues. The studies appear in the reference list in the order in which they appear in this summary.

### Infection

The initial work was through the clinical, radiological and pathological study of many foals which apparently had infective polyarthritis, but which were shown to also be afflicted with osteomyelitis. Until then, it had been assumed that bone infection was a sequel of chronic infection of articular cartilage extending into the underlying bone. Epiphyseal osteomyelitis was found to be common in foals, often present in subchondral bone without evidence of infection at the surface of joint cartilage. The epiphyseal form of haematogenous osteomyelitis is rare in children, and had not been described previously in foals. The then new classification system of the various combinations of infective arthritis, epiphyseal osteomyelitis (also referred to as metaphysitis), and tarsal osteomyelitis (1,2,3, 4) is still used. These disease entities were recognized in calves and the same classification system was applicable (5).

The reason for predilection of the osteomyelitis for particular joints, and for particular sites within those joints (6) was studied with specific reference to the vascular anatomy (7,8), cartilage thickness (9), cartilage maturation (10), cartilage morphology (11), and the varying shape of the physis in the growing animal (12). Besides the classically described bone infection originating at the junction of physeal cartilage and metaphyseal bone (metaphysitis), infection was also demonstrated to originate on the epiphyseal side of the physeal cartilage.

Transphyseal vessels were a significant route of extension of infection, allowing extension of bone infection from one side of the physis to the other. Epiphyseal bone infection was shown to originate at the chondro-osseous junction of the deepest layer of cartilage and the underlying immature bone. The infection almost always began at sites with thick epiphyseal cartilage, and the latter was found to be where intra-cartilaginous vessel arcades persist for longest and the process of expansion of the secondary ossification centre of the epiphysis lasts longer than in areas of thin cartilage. This relationship of the predilection site of haematogenous epiphyseal bone infection to cartilage thickness was a novel finding, largely explaining why the disease occurs in specific sites, and only in foals up to a few weeks old.

The special nature of the terminal vascular arrangements in the thickest unossified epiphyseal cartilage led to the first studies of the tissue pressures in bone of both foals (13,14) and older ponies (15), since the conditions for bacterial colonization most likely include low pressure and low blood flow velocity in the areas shown to be susceptible to establishment of infection.

Consideration of the clinical implications of studies up to that time prompted investigation of the early diagnosis of infective metaphysitis in foals (16), and the features of metaphysitis caused by Rhodococcus sp (17). This resulted in a hypothesis that transphyseal vessels probably have a reserve function in the repair of metaphyseal retained cartilage and possibly other lesions on either side of the physis (18). A separate form of bone infection, diaphyseal osteitis, was also documented in terms of its frequent prevalence, relationship to penetrative trauma or other sources of infection, predilection site, and management (19).

At the time, the rationale for clinical treatment of infective arthritis relied on investigation of synovial fluid concentrations of systemically administered antibiotics, using normal horses, and in some laboratories in horses with induced bacterial infection. The latter was undesirable, and was replaced by a model using endotoxin-induced inflammation (20,21) which could be accurately quantified and minimized in terms of severity and duration. This allowed induction of extremely mild bone inflammation (16) which is still used in orthopaedic research, thus improving the welfare of animals. The interest in pursuing treatment led to collaborative work investigating aspects of optimizing factors to increase concentrations of antibiotic at the focus of bone infection (22,23,24,25,26,27). This was a prime focus before other methods such as regional intravenous, intraarticular, and regional intra-osseous administration were examined in detail.

#### **Cartilage Development and Osteochondrosis in foals**

Careful pathological examination of bone sections of very young foals without evidence of infection revealed the incidental finding of small retained cartilage lesions (28) at particular sites of the epiphyseal and metaphyseal aspect of the physeal cartilage (12). Therefore, induction of cartilage retention was attempted so that the relationship of retained cartilage to angular limb deviation could be studied. After altering the foot conformation by applying a plastic wedge on the outside (lateral) wall, the bone strain increased on the lateral aspect of the third metacarpal bone by 100%, and decreased by 40% on the medial aspect (compared to values before the wedge was applied). Within 10 days, the strains had equalized (29) and the hoped for non-invasive model of angular limb deviation did not eventuate. However, the work was the stimulus for the author, many years later, to study the effects of altered forces on connective tissues.

From the studies conducted to this stage, involving dissection and examination of hundreds of foal joints, it was clear that the sites with thickest cartilage were those of predilection to not only bacterial infection but also to articular osteochondrosis. The latter disease is part of the Developmental Orthopaedic Disease complex, is said to be the biggest problem facing horse breeders world-wide, and is still increasing in prevalence. This stimulated particular interest in the stage of skeletal maturity at birth and in subsequent development as immature structures develop into those capable of sustaining the forces applied during athletic locomotion. The degree of maturation at birth and subsequent growth and development also determine the likelihood of infection (4) in the first weeks of life. Also, abnormal endochondral ossification remains the acknowledged reason for the development of lesions of osteochondrosis.

At the time, it was vigorously contended that dietary concentration of copper was of central importance in the pathogenesis of equine osteochondrosis. This was of considerable importance to the New Zealand horse-breeding industry, since most of our pastures have much lower copper concentration than is recommended by the National Research Council and by feed companies in the Northern Hemisphere. Feeding copper to late pregnant mares resulted in higher liver copper stores in their newborn foals, compared to those of non-supplemented mares (30) and some foals from supplemented mares had much greater storage of copper, which they retained for longer (31). Osteochondrosis lesion scores were lower in foals whose dams had been supplemented with copper, but the lesions were very small and of unlikely clinical significance (32). The need for supplemental copper appears to have been overstated, at least in terms of pathogenesis of osteochondrosis, although higher copper status of the foal was associated with superior healing of early osteochondrosis lesions in Dutch Warmblood horses (33). The growth rate at 3-4 months was significantly higher in foals with more severe osteochondrosis, and in severely affected young horses the bone mineral density was less (perhaps indicating greater bone fragility) than in the same sites in horses with less severe osteochondrosis (34). The mechanisms through which growth affects cartilage to cause osteochondrosis lesions, and why some lesions heal and others do not, remain open questions.

Because of the known effect of concentrate feeding regimens on the risk of osteochondrosis, study was undertaken to determine if the dietary requirements of New Zealand thoroughbred foals could be realized when raised on well-managed pasture alone. Although accurate measurement of the dietary intake of individual animals at pasture remains impossible, very good estimates of intake and requirements of weanlings (35), yearlings (36) and mares (37), and of the total body mineral composition of foals (38) and of mare's milk (39) provided a scientific basis for Thoroughbred pasture nutrition. Because of the possible relationship between excess body weight and pathogenesis of juvenile orthopedic

diseases, whole body fat content was estimated in Thoroughbred weanlings (40). Enlargements above the fetlock area, commonly referred to as epiphysitis and said to be a form of osteochondrosis, were common and devoid of indications of being inflammatory, instead appearing to be a normal response of bone tissue (41). In these same 5 month old foals, only subtle articular osteochondrosis lesions could be detected (42). In other foals, higher growth rates appeared to be associated more with season-related nutritional effects (43) than with puberty, the onset of which occurred in both fillies and colts within a very narrow interval in spring (43,44,45).

#### Exercise

Various factors induce change in the structure, composition and resistance to deformation of connective tissues. In an age in which most horses are used for intermittent athletic or recreational purposes, the most obvious and significant factor affecting equine tissues appears to be exercise, in the form of athletic preparation and training and competition, or **lack** of exercise through confinement in either pasture or stall. If the training stimulus is too severe, or if adaptive change to a suitable stimulus is inappropriate, then accumulation of micro-damage can result in injury, of either subtle or catastrophic nature. The equine scientific literature indicates that muscle appears to be less often injured than tendon, bone and joint cartilage, so attention was thus given to the latter three tissues.

The response of tissues to growth, ageing, or exercise were documented in crosssectional study of tendons of wild (46) and domesticated (47,48) horses, and articular cartilage and subchondral bone of wild horses (49). Controlled studies involving treadmill training showed obvious change in the collagen properties in the central but not the peripheral region of the superficial digital flexor tendon of exercised horses that were exercised for 19 months, but not 19 weeks (50), compared to control horses which were walked and trotted only (51,52).

In carpal bones, the bone mineral density was greater in the dorsal compressive load path of the carpal bones, compared to the more palmar regions of the joint. In some regions of interest, mean density was more than 30% greater in the trained compared to the control group (53), and such changes were evident in horses trained for only 19 weeks (54). Foals bred from osteochondrosis-afflicted parents were sprint-exercised and compared to a group of foals kept in a large stall, and to another group which was at pasture. Again, the exercise, and lack of it in the boxed group, resulted in clear differences in bone mineral content in the distal radius and in the third carpal bone (34).

Neither of the exercise regimens in the above studies were typical of management used in New Zealand. The Massey University Grass Exercise Study (MUGES) was the first to determine the effect of conventional training exercise conducted at a racetrack on bone, tendon, and joint cartilage (55). The exact distance that each horse worked at various velocities was determined (56). The cross sectional area (57) and volume (58) of the superficial digital flexor and common digital extensor tendons were slightly greater in the trained than untrained group; the lower density of the former tendon is a novel finding. Wear lines and osteochondral fragmentation in articular cartilage were few and mild in the carpus, but obvious in the metacarpo-phalangeal joints (55), associated with altered collagen biochemical characteristics (59). Carpal hyaline cartilage was thicker in trained horses, which indicated that cartilage responded to increased load (60). Bone accretion rate in the metacarpal diaphysis was determined by confocal microscopy after serial injection of bone markers (61). Diaphyseal size increased after bone density increased, leading to the first recognition that different gaits or gait velocity may stimulate different bone properties contributing to bone strength (62). After the training period of only 12-14 weeks, increased bone density in the third metacarpal condyle was established to be via deposition of new bone on pre-existing internal surfaces without prior resorption (63), in the form of progressively thicker columnar structures in the third carpal bone (60). Other novel findings included the physical communication between cartilage and bone through the calcified articular cartilage (64).

Because of the deficiencies demonstrated in one conventional imaging modality (65) and reviewed in others (66), more sophisticated imaging modalities are required to study changes in horses that are training and/or growing. Also, spontaneous lesions that would have prevented successful racing careers had been observed in untrained young horses, and worse lesions were found in horses that had been trained, although their training was at lower intensity than usually utilized in commercial training. These articular cartilage changes are probably the most important factor causing the high wastage in Thoroughbred training and racing, and require more sensitive detection techniques (55, 62).

Collaborative work (67) gave support to the hypothesis that injury and wastage might be as dependant on the factors, summarized in an invited review (68), which affect tissue development and health before training, and not just on what occurred during training. This required monitoring of cartilage, bone and tendon development in foals exposed from a very early age to exercise of a lower intensity than the regimen known to have caused long-term damage to articular cartilage in the confined foals described above. In the largest study of its kind, in collaboration with Dutch, English and American investigators, 16 of 32 foals raised on pasture were exercised on a grass track 5 days per week until 18 months of age, and the other 16 exercised only spontaneously in the paddock. An imaging method relatively new to equine research, peripheral quantitative computed tomography, was used to measure bone mineral content of the proximal phalanx, third metacarpal bone and distal radius at 0, 2, 4, 12 and 18 months of age. The early conditioning exercise resulted in a significant increase in bone size and strength and mineral content in the trained group compared to the control group, and the effect of spelling after race training of the horses as two and three year olds was determined (69). Computed tomography in the standing horse (as opposed to the anaesthetized laterally recumbent animal) is thus a rational option for quantifying the effects of growth, early exercise, athletic competition and withdrawal from competition, but magnetic resonance imaging is required for assessing articular cartilage.

The foals exposed to early exercise were also studied *ex vivo* to ensure that the conditioning regimen had caused no ill-effect on the most susceptible connective

tissue, articular cartilage. The features of articular calcified cartilage were shown, using combined forms of microscopy (70), to be different in conditioned and control foals (71). In the hyaline cartilage of the third metacarpal bone condylar surface, conditioning exercise was associated with a lower proportion of dead chondrocytes than in the cartilage of the control foals, which had exercised freely at pasture (72). This is an important observation and indicates that cartilage responds like other musculo-skeletal system tissues, that moderate exercise in the young is beneficial, and that managing horses in paddocks may not be as natural as it appears, in that some degree of sedentarism occurs.

The studies in this thesis began with the neonatal foal, later concentrated on the training of the young equine athlete, and most recently returned to the study of exercise in the very young as a way of altering the capability of tissues to later athletic training. However, the function of tissues largely depends on their structure, and structure is determined by many interacting influences, including genetic factors which can be modified in gestational or early post-natal life by various environmental stimuli, including maternal size and constraint, maternal and fetal nutritional status, and the pre- and post-natal exercise environments. Study in this exciting arena of very early growth and development, and its effect on later musculo-skeletal health, has begun (73, 74).

### List of publications

Five papers (#) originally published in the candidate's PhD thesis (1983) and peer-reviewed abstracts of selected presentations at international conferences (\*) are included not for assessment, but to provide the reader with an insight into the origins of the work and the direction in which it is continuing.

- 1. Firth EC, Dik KJ, Goedegebuure SA, Hagens FM, Verberne LRM, Merkens HW, Kersjes AW. (1980) Polyarthritis and bone infection in foals. *Zbl Vet Med. B* 27, 102-124.
- 2. \*Goedegebuure SA, Dik KJ, Firth EC, Merkens HW. (1980) Polyarthritis and polyosteomyelitis in foals. *Veterinary Pathology* 17,651.
- 3. #Firth EC. (1983) Current concepts of infectious polyarthritis in foals. Commissioned article. Equine Veterinary Journal 15, 5-9.
- 4. Firth EC, Goedegebuure SA, Dik KJ, Poulos PW. (1985) Tarsal osteomyelitis in foals. *Veterinary Record* 116, 261-266.
- 5. Firth EC, Kersjes AW, Dik KJ, Hagens FM. (1987) Haematogenous osteomyelitis in cattle. *Veterinary Record* 120, 148-152.
- 6. Firth EC, Goedegebuure SA. (1988) The site of focal osteomyelitis lesions in foals. *Veterinary Quarterly 10, 99-108*.
- 7. #Firth EC, Poulos PW. (1982) Blood vessels in the developing growth plate of the equine distal radius and metacarpus. *Research in Veterinary Science 33*, 159-166.

- 8. #Firth EC, Poulos PW. (1983) Microangiographic studies of metaphyseal vessels in young foals. *Research in Veterinary Science* 34, 231-235.
- 9. Firth EC, Greydanus Y. (1987) Cartilage thickness measurement in foals. Research in Veterinary Science 42, 35-46.
- 10. #Firth EC, Hartman W. (1983) An in vitro study on joint fitting and cartilage thickness in the radiocarpal joint of foals. *Research in Veterinary Science 34*, 320-326.
- 11. Firth EC, Poulos P. (1993) Vascular characteristics of the cartilage and subchondral bone of the distal radial epiphysis of the young foal. New Zealand Veterinary Journal 41(3),73-77.
- 12. Firth EC, Hodge H. (1997) Physeal form of the longbones of the foal. Research in Veterinary Science 62(3), 217-221.
- 13. Stolk PWT, Firth EC. (1988) Intra-osseous pressure in the equine third metatarsal bone. Cornell Veterinarian 78, 191-206.
- 14. Stolk PWT, Firth EC. (1990) Intra-Osseous Pressure and Pressure Pulse Gradients along the Equine Third Metatarsal Bone. Cornell Veterinarian 80, 317-328.
- 15. Stolk PWT, Firth EC. The relationship between intra-articular and juxtaarticular intra-osseous pressures in the metatarsophalangeal region of the pony. *Veterinary Quarterly 16(2), 81-86.*
- 16. Firth EC. (1990) Diagnosis of metaphysitis in the foal. New Zealand Veterinary Journal 38(2), 66-68.
- 17. Firth EC, Alley MR, Hodge H. (1993) *Rhodococcus equi* associated osteomyelitis in foals. *Australian Veterinary Journal* 70, 304-308.
- 18. Firth EC, Klarenbeek A. (1995) Transphyseal vessel involvement in repair of metaphyseal retained cartilage. Australian Veterinary Journal 72, 452-455.
- 19. Firth EC. (1987) Bone sequestration in large animals. Australian Veterinary Journal 64, 65-69.
- 20. Firth EC, Seuren F, Wensing T. (1987) An induced synovitis disease model in the pony. *Cornwell Veterinarian 11, 107-118*.
- 21. Firth EC, Klein WR, Nouws JFM. (1988) Effect of induced synovial inflammation on pharmacokinetics and synovial concentration of sodium ampicillin and kanamycin sulphate after systemic administration in ponies. *Journal Veterinary Pharmacology & Therapeutic 11*, 56-62.
- 22. Firth EC, Nouws JFM, Driessens F, Schmaetz P, Peperkamp K, Klein WR. (1986) Effect of injection site on the pharmacokinetics of procaine penicillin in the horse. *American Journal of Veterinary Research* 47, 2380-84.

- 23. Nouws JFM, Firth EC, Vree TB, Baakman M. (1987) Pharmacokinetics and renal clearance of sulfamethazine, sulfadiazine and their N4-acetyl and hydroxy metabolites in horses. *American Journal of Veterinary Research* 48, 392-402.
- 24. Firth EC, Nouws JFM, Klein WR, Driessens F. (1990) The effect of phenylbutazone on plasma disposition of penicillin G in the horse. Journal Veterinary Pharmacology Therapeutic 13, 179-185.
- 25. Firth EC, Whittem T, Nouws J. (1993) Kanamycin concentrations in synovial fluid after intramuscular administration in the horse. *Australian Veterinary Journal* 80, 324-325.
- 26. Anderson BH, Firth EC, Whittem T. (1995) The disposition of gentamicin in equine plasma, synovial fluid and lymph. *Journal Veterinary Pharmacology & Therapeutics 18, 124-131.*
- 27. Whittem T, Firth EC, Hodge H, Turner K. (1996) Pharmocokinetic interactions between repeated dose phenylbutazone and gentamicin in the horse. *Journal Veterinary Pharmacology & Therapeutics 19, 454-459.*
- 28. #Firth EC, Poulos PW. (1984) Retained cartilage in the distal radial physis of foals. Veterinary Pathology 21, 10-17.
- 29. Firth EC, Schamhardt HC, Hartman W. (1988) Measurements of bone strain in foals with altered foot conformation. American Journal of Veterinary Research 49, 261-265.
- 30. Pearce SG, Grace ND, Firth EC, Wichtel JJ, Holle SA, Fennessy PF. (1998) Effect of copper supplementation on copper status of pregnant mares and foals. Equine Veterinary Journal 30, 200-203.
- 31. Gee EK, Morel PCH, Mogg TD, Firth EC, Grace ND, Fennessy PF. (2004) Liver copper kinetics in Thoroughbred foals at pasture from birth to 160 days of age. *New Zealand Journal of Agricultural Research 47, 109-118.*
- 32. Pearce SG, Firth EC, Grace ND, Fennessy PF. (1998) Effect of copper supplementation on the evidence of developmental orthopaedic disease in pasture-fed New Zealand thoroughbreds. *Equine Veterinary Journal 30*, 211-218.
- 33. van Weeren PR, Knaap J, Firth EC. (2003) The influence of liver copper status of mare and newborn foal on the development of osteochondrotic lesions. *Equine Veterinary Journal 35(1), 67-71*.
- 34. Firth EC, van Weeren PR, Pfeiffer DU, Delahunt J, Barneveld A. (1999) Effect of age, exercise and growth rate on bone mineral density (BMD) in the third carpal bone and distal radius of Dutch warmblood foals with osteochondrosis. Equine Veterinary Journal 31, 74-78.
- 35. Grace ND, Firth EC, Shaw<sup>b</sup> HL, Gee<sup>b</sup> EK. (2002) Digestible energy intake, dry matter digestibility and effect of increased calcium intake on bone parameters of grazing Thoroughbred weanlings in New Zealand. New Zealand Veterinary Journal 51(4) 165-173.

- 36. Grace ND<sup>•</sup>, Gee<sup>§</sup> EK, Firth<sup>§</sup> EC, Shaw<sup>§</sup> HL. (2002) Digestible energy intake, dry matter digestibility, and mineral status of grazing New Zealand Thoroughbred yearlings. *New Zealand Veterinary Journal 50, 63-69*.
- 37. Grace ND, Shaw HL, Gee EK, Firth EC. (2002) Determination of the digestible energy intake and apparent absorption of macroelements in pasture-fed lactating Thoroughbred mares. New Zealand Veterinary Journal 50(5), 182-185.
- 38. Grace ND, Pearce SG, Firth EC, Fennessy PF. (1999) Content and distribution of macro- and microminerals in the body of pasture-fed young horses. *Australian Veterinary Journal* 77, 172-176.
- 39. Grace ND, Pearce SG, Firth EC, Fennessy PF. (1999) Concentrations of macroand micro-elements in the milk of pasture-fed thoroughbred mares. *Australian Veterinary Journal* 77, 177-180.
- 40. Gee EK, Fennessy PF, Morel PCH, Grace ND, Firth EC, MoggTD. (2003) Chemical body composition of 20 Thoroughbred foals at 160 days of age, and preliminary investigation of techniques used to predict body fatness. New Zealand Veterinary Journal 51(3), 125-131.
- 41. Gee EK, Firth EC, Morel MCH, Fennessy PF, Grace ND, Mogg TD. (2005) Enlargements of the distal third metacarpus and metatarsus in Thoroughbred foals at pasture from birth to 160 days of age. *New Zealand Veterinary Journal* 53, 438-448.
- 42. Gee EK, Firth EC, Morel PCH, Fennessy PF, Grace ND, Mogg TD. (2005) Articular/epiphyseal osteochondrosis in Thoroughbred foals at 5 months of age: influences of foal growth and prenatal maternal copper supplementation. New Zealand Veterinary Journal 53(6), 449-457.
- 43. Brown-Douglas CG, Firth EC, Parkinson TJ, Fennessy PF. (2004) Onset of Puberty in pasture-raised Thoroughbreds born in spring and autumn. *Equine Veterinary Journal 36, 499-504.*
- 44. Brown-Douglas CG, Firth EC, Parkinson TJ, Fennessy PF. (2003) Bodyweights and growth rates of spring- and autumn-born Thoroughbred horses raised on pasture. New Zealand Veterinary Journal 53(5), 326-3.
- 45. Brown-Douglas CG, Firth EC, Parkinson TJ, Fennessy PF. (2005) The pituitary and testicular response to GnRH challenge between 4 and 14 months of age in Thoroughbred colts born in spring and autumn. *Animal Reproduction Science* 88, 279-98.
- 46. Patterson-Kane JC, Firth EC, Goodship AE, Parry DAD. (1997) Age-related differences in collagen crimp patterns in the superficial digital flexor tendon core region of untrained horses. *Australian Veterinary Journal* 75(1), 39-44.
- 47. Patterson-Kane JC, Parry DAD, Goodship AE, Firth EC. (1997) Exercise modifies the age-related change in crimp pattern in the core region of the equine superficial digital flexor tendon. New Zealand Veterinary Journal 45, 135-139. random
- 48. Patterson-Kane JC, Parry DAD, Birch HL, Goodship AE, Firth EC. (1997) An Age-Related Study of Morphology and Cross-Link Composition of Collagen

Fibrils in the Digital Flexor Tendons of Young Thoroughbred Horses. *Connective Tissue Research* 36, 253-260.

- 49. Cantley CEL, Firth EC, Delahunt JW, Pfeiffer DU, Thompson KG. (1999) Naturally occurring osteoarthritis in the metacarpophalangeal joints of wild horses. Equine Veterinary Journal 31, 73-81.
- 50. Firth EC. (2000) Tendinopathy in Domestic Animals. Sports Medicine and Arthroscopy Review (8), 105-113.
- 51. Patterson-Kane JC, Wilson AM, Firth EC, Parry DAD, Goodship AE. (1997) Comparison of collagen fibril populations in the superficial digital flexor tendons of exercised and non-exercised Thoroughbreds. *Equine Veterinary Journal* 29(2), 121-125.
- 52. Patterson-Kane JC, Wilson AM, Firth EC, Parry DAD, Goodship AE. (1998) Exercise related alterations in crimp morphology in the central regions of superficial digital flexor tendons from young thoroughbreds: a controlled study. Equine Veterinary Journal 30, 61-64.
- 53. Firth EC, Delahunt J, Wichtel JW, Birch HL, Goodship AE. (1999) Galloping exercise induces regional changes in bone density within the third and radial carpal bones of thoroughbred horses. *Equine Veterinary Journal* 31(2), 111-115.
- 54. Firth EC, Goodship AE, Delahunt J, Smith T. (1999) Osteoinductive response in the dorsal aspect of the carpus of young thoroughbreds in training occurs within months. *Equine Veterinary Journal Supplement 30*, 552-554.
- 55. Firth EC, Rogers CW, Perkins NR, Anderson BH. (2004) Musculoskeletal responses of two year old thoroughbred horses to early training. 1. Study design, and clinical, nutritional, radiological, and histological observations. New Zealand Veterinary Journal 52, 261-271.
- 56. Rogers CW, Firth EC. (2004) Musculoskeletal responses of two year old thoroughbred horses to early training. 2. Measurement error and effect of training stage on relationship of objective and subjective criteria of training workload. New Zealand Veterinary Journal 52, 272-279.
- 57. Perkins NR, Rogers CW, Firth EC, Anderson BH. (2004) Musculoskeletal responses of two year old thoroughbred horses to early training. 3. *In vivo* ultrasonographic assessment of superficial digital flexor tendon cross sectional area and echogenicity. *New Zealand Veterinary Journal 52, 280-284*.
- 58. Firth EC, Rogers CW, Anderson BH. (2004) Musculoskeletal responses of two year old thoroughbred horses to early training. 4. Physical properties of forelimb digital tendons. New Zealand Veterinary Journal 52, 285-292.
- 59. Brama PA, Tekoppele JM, Bank RA, Barneveld A, Firth EC, van Weeren PR. (2000) The influence of strenuous exercise on collagen characteristics of articular cartilage in Thoroughbreds age 2 years. *Equine Veterinary Journal 32*, 551-54.
- 60. Firth EC, Rogers CW. (2005) Musculoskeletal responses of 2-year-old Thoroughbred horses to early training. 7. Bone and articular cartilage response in the carpus. New Zealand Veterinary Journal 53, 113-122.

- 61. Firth EC, Rogers CW, Doube M, Jopson NB. (2005) Musculoskeletal responses of two year old thoroughbred horses to early training. 6. Bone parameters in radius, third metacarpal and third metatarsal bones of young racetrack-trained thoroughbreds. *New Zealand Veterinary Journal* 53, 101-112.
- 62. Firth EC, Rogers CW. (2005) Musculoskeletal responses of 2-year-old Thoroughbred horses to early training. Conclusions. New Zealand Veterinary Journal 53, 377-383.
- 63. Boyde A, Firth EC. (2005) Musculoskeletal responses of two year old Thoroughbreds to training. 8. Quantitative backscattered electron SEM and confocal fluorescence microscopy of the epiphysis of the third metacarpal bone. New Zealand Veterinary Journal 53, 123-132.
- 64. Boyde A, Firth EC. (2004) Articular Calcified Cartilage in the third metacarpal bone of two year old Thoroughbred racehorses. *Journal of Anatomy 205, 491-500*.
- 65. Secombe CJ, Firth EC, Perkins NR, Bailey D, Anderson BH. (2004) The quantitative assessment of photodensity of the third carpal bone in the horse. *New Zealand Veterinary Journal* 52, 70-75.
- 66. Firth EC. (2004) Problems in quantifying bone response to exercise in horses: a review. New Zealand Veterinary Journal 52, 216-229.
- 67. Smith RKE, Birch HL, Patterson-Kane J, Firth EC, Williams L, Cherdchutham W, van Weeren WR, Goodship A. (1999) Should equine athletes commence training during skeletal development?: changes in tendon matrix associated with development, ageing, function and exercise. *Equine Veterinary Journal Supplement 30, 201-209.*
- 68. Firth EC. (2006) The response of bone, articular cartilage, and tendon to exercise in the horse. *Journal of Anatomy 206, 513-526.*
- 69. \*Firth EC, Rogers CW, van Weeren R, Barneveld A, Kawcak CE, McIlwraith CW, Goodship AE, Smith RWK. (2006) Change in bone parameters in Thoroughbred horses months after training has ceased. Abstract: International Conference on Equine Exercise Physiology, August 2006, Fontainebleau, France.
- 70. Doube M, Firth EC, Boyde A. (2005) Registration of Confocal Scanning Laser Microscopy and Quantitative Backscattered Electron Images for the Temporospatial Quantification of Mineralization Density in 18-Month Old Thoroughbred Racehorse Articular Calcified Cartilage. Scanning 27, 219-226.
- 71. \*Doube M, Firth EC, Boyde A. (2005) Preconditioning exercise and anatomic site relate to mineralisation and thickness in equine third metacarpal articular calcified cartilage. Abstract 34th European symposium in Calcified Tissues, Prague, May 2006. Calcified Tissue International 78, Suppl 1, 47.
- 72. \*Dykgraaf S, Firth EC, Rogers CW, Kawcak C. (2005) Effect of exercise on chondrocyte viability and subchondral bone sclerosis of the distal third metacarpal and metatarsal bones of young Thoroughbred horses. *Abstract American Association of Equine Practitioners, Annual Focus Symposium, Louisville July, 2004.*

- 73. \*Blair HT, Kenyon PR, Jenkinson CMC Peterson SW, Mackenzie DDS, Johnson PL, Morris ST, Firth EC. (2006). Does dam size and feeding during pregnancy affect fetal organ weight? *Early Human Development 82:548*.
- 74. \*Firth EC, Rogers CW, Vickers M, Kenyon PR, Jenkinson CMC, Blair HT, Johnson PL, Mackenzie DDS, Peterson SW. (2006). Does maternal size and feeding during pregnancy affect lean mass and bone content in sheep? *Early Human Development* 82:551.