

# 1 Towards a quantitative indicator of feather **disruption** following the cleansing of 2 oiled birds

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## REVISED MANUSCRIPT

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### Abstract

13

14 A computer-based imaging method for determining feather microstructure coherency  
15 following a cleansing treatment, was developed, calibrated and trialled on Mallard  
16 Duck (*Anas platyrhynchos*) feathers. The feathers were initially contaminated with a  
17 light crude oil and then cleansed by either detergent (Deacon 90) treatment or,  
18 alternatively, by magnetic particle technology (MPT) using iron powder. The  
19 imaging method provides a single quantitative parameter for the coherence of feather  
20 microstructure and the results confirm that MPT treatment imparts less disruption to  
21 the feather microstructure than detergent treatment. It is proposed that this imaging  
22 method can be developed and implemented for the assessment of **feather disruption**  
23 **and possibly damage**, either for the trialling of different treatment protocols, or as a  
24 tool during the rehabilitation process, along with other such indicators, to give a more  
25 comprehensive assessment of feather condition than is currently available.

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### Key Words

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29  
30 Computer-imaging; feather microstructure coherency; magnetic particle cleansing;  
31 oil-soaked birds

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### Highlights

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- 35  
36 • Computer-based imaging method developed to quantitatively assess feather  
37 **coherency**.
- 38 • Imaging method successfully applied in studying cleansing of oil-soaked feathers  
39 and to feather **coherency** assessment.
- 40 • Cleansing feathers magnetically imparts less **disruption** to feather integrity than  
41 detergent methods.

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48 **1. Introduction**

49

50 Magnetic particle technology (MPT) has demonstrated great utility in a range of  
51 discipline areas (Safarikova & Safarik, 2001) and is a convenient and quick means by  
52 which oil-soaked wildlife can be cleansed (Orbell *et al.*, 2007). This innovative  
53 approach has been investigated and developed for the clean-up of oil-soaked species  
54 such as the Mallard Duck (*Anas platyrhynchos*) and Little Penguin (*Eudyptula minor*)  
55 (Orbell *et al.*, 2004). This technology has the advantage of being portable and enables  
56 oil to be removed immediately in the field, upon first encounter, either directly or with  
57 the aid of pre-treatment agents in cases where the oil residues are highly weathered  
58 and/or persistent (Ngeh *et al.*, 2012).

59

60 Two further advantages of MPT technology over detergent-based techniques for oil  
61 removal are its high efficiency and its purported ability to invoke less damage to the  
62 feather microstructure (Orbell *et al.*, 2007). In relation to exploring the removal  
63 efficiency of MPT it has been necessary to develop a quantitative assay (Orbell *et al.*,  
64 1997) that can be used to compare the efficiency of MPT technology with those  
65 efficiencies offered by detergent cleansing. To this end, computer-assisted analyses  
66 such as those developed for the sequestering (Bigger *et al.*, 2010) and the sequential  
67 pick-up (Bigger *et al.*, 2013) of chemical contaminants by MPT have been used to  
68 process gravimetric laboratory data to quantitatively and objectively determine the  
69 efficiency of oil removal under a variety of conditions.

70

71 The quantitative assessment of feather condition [as indicated by its coherency](#), on the  
72 other hand, remains an area still to be developed. It has long been recognized that  
73 feather condition, as manifested in the ability of feathers to repel water, is a key factor  
74 governing the decision as to when to release a rehabilitated bird back into the wild  
75 (Ngeh, 2002). Work on the quantitative assessment of feather condition includes  
76 early studies that applied a theory of water repellence developed by Cassie and Baxter  
77 (1944) for woven fabrics and textiles, to the structure of a feather vane (Rijke, 1968;  
78 Rijke, 1970; Rijke *et al.*, 2000). It was proposed that the water repellence of contour  
79 feathers is mathematically related to the radius of the feather barb and the half-  
80 distance between the axes of the barb (Stephenson, 1997; Stephenson & Andrews,  
81 1997).

82

83 Further to these early studies, there appears to be very little available literature on the  
84 quantitative assessment of feather condition following, say, cleansing or other  
85 rehabilitation treatments. Those that have been reported and that provide a  
86 quantitative, or at least semi-quantitative, assessment of feather condition, have been  
87 developed to varying extents. For example, the presence of preening oils in feathers  
88 has long been recognized as an important factor in enabling the feather to repel water  
89 (Elder, 1954; Stettenheim, 1972; Elowson, 1984). Based on this observation, the use  
90 of gas chromatography to quantify levels of preening oils and waxes in feathers taken  
91 from rehabilitating birds has been explored as a possible method leading to the

92 assessment of feather condition (Murray, 1962; Odham & Stenhagen, 1971). Other  
93 studies on the assessment of feather condition include those that report the semi-  
94 quantitative assessment of wing feather mite infestations on songbirds (Behnke *et al.*,  
95 1999; Carleton & Proctor, 2010) and the use of infrared thermography to assess laying  
96 hen feather coverage (Zhao *et al.*, 2013).

97  
98 A notable and more recent contribution is the work of O'Hara and Morandin (2010)  
99 who developed a barbule amalgamation index that is calculated from measurements  
100 made of feather rami taken from micrograph images. This index was used as an  
101 indicator of feather condition following exposure to oil sheens. A quantitative  
102 assessment such as this can be of significant value in the overall assessment of the  
103 condition of a bird such as a Little Penguin and, hence, in determining when to release  
104 it from a rehabilitation facility. To date, the standard international practice for  
105 deciding on the water repellency of the plumage has been to place the penguin in a  
106 pool at various stages in the rehabilitation process, monitor its behaviour and  
107 buoyancy, inspect the degree of water penetration into the plumage, and make a  
108 subjective assessment accordingly (Stocker, 2000; Department of Primary Industries,  
109 2012). Clearly, to have a high rehabilitation success rate such practice requires a high  
110 level of experience and expertise in judging when the bird's plumage is fully water-  
111 proof and ready for release.

112  
113 In view of the need for the further development of quantitative methods for the  
114 assessment of feather condition, this paper describes a computer-assisted imaging  
115 method that can be used to provide a quantitative indication of feather microstructure  
116 coherency following treatment by detergent or other cleansing actions. It is proposed  
117 that the coherence of the feather structure is an important factor that should be  
118 considered along with other factors such as the levels of preening oils and waxes  
119 when considering the water resistance and thermal insulating properties of the feather.  
120 As such, the coherence of the feather structure can be explored as an important  
121 indicator of feather condition. An indicator such as this may also have potential  
122 future use in a range of veterinary and husbandry applications.

## 124 **2. Methods**

### 126 2.1 Materials and Feather Characterization

127  
128 Samples of breast feathers of the Mallard Duck (*Anas platyrhynchos*) were used in  
129 this study<sup>1</sup>. These feathers were initially contaminated with a light crude oil (Esso,  
130 Australia, Ltd.; viscosity 11.4 cP) by immersion in the oil for 1 min. Randomly  
131 selected and uncontaminated duck feathers were used as a control. A 5% (v/v)

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<sup>1</sup> The breast feathers of the Mallard Duck are ideal for this *proof of principle* study since they have a well-defined 2-D grid microstructure (Orbell *et al.*, 1999). It is appreciated, however, that feather microstructure is highly variable from one species to another and a specific microstructure parameter would need to be developed for each individual feather type.

132 cleansing detergent solution was prepared by mixing 10 mL of Decon 90™ with 190  
133 mL of distilled water. Iron particles having an average maximum dimension of 0.21  
134 mm were obtained from Hoganas, Sweden (grade C100.29). The particles were  
135 removed from the feathers using a "laboratory magnetic tester" supplied by Alpha  
136 Magnetics, Victoria, Australia (Orbell *et al.*, 1997).

137

### 138 2.2 Feather Treatment and Optical Microscopy

139

140 The detergent cleansing technique involved holding a clustered sample of 5 or 6 oiled  
141 feathers by their quills (calami) and agitating in the detergent solution for a period of  
142 10 min. The feathers were then rinsed by agitating them in distilled water for 5 min  
143 and were left to dry in air for one week. The magnetic particle cleansing treatment  
144 involved completely covering a cluster of oiled feathers with the iron particles in a  
145 Petri dish and removing all the particles with 1 to 3 continuous sweeps of the  
146 laboratory magnetic tester.

147

148 Treated single feather samples were placed on a glass plate and clamped on the stage  
149 of a Nikon optical microscope (Labophot Model 248625). Micrographs of the  
150 samples were obtained using Nikon Model 401 SLR camera fitted to the microscope.

151

### 152 2.3 Grid-Generating Algorithm

153

154 A Monte Carlo computer program was written to calculate the area distribution  
155 histogram of the quadrilateral elements in a randomly disrupted two-dimensional grid  
156 as a function of the extent of the imposed disruption to the grid. In the program, a  
157 square grid of side dimensions,  $S$ , comprising  $n \times n$  elements was generated as a series  
158 of  $n + 1$  horizontal and vertical intervals drawn between the sets of points  $P_1$  and  $P_2$   
159 whose coordinates are:  $P_{H,1}[x_H(i, 1), y_H(i, 1)]$ ;  $P_{H,2}[x_H(i, 2), y_H(i, 2)]$ ;  $P_{V,1}[x_V(i, 1),$   
160  $y_V(i, 1)]$  and  $P_{V,2}[x_V(i, 2), y_V(i, 2)]$  and where  $1 \leq i \leq n + 1$ . The subscripts  $H$  and  $V$   
161 represent the horizontal and vertical components respectively. The allowed  
162 deviations of the termini of each interval from their original positions on the grid is  
163 given by:

164

$$165 \quad d = f \times S / (2n) \quad (1)$$

166

167 where  $0 < f < 1$ . The parameter,  $f$ , is a constant that is set before the program is run  
168 and controls the extent of random disruption to the grid. The horizontal and vertical  
169 intervals within the perimeter of the square  $S$  are randomly disrupted by resetting the  
170 coordinates of their termini to:  $P_{H,1}[0, i \times S/n + k \times d]$ ;  $P_{H,2}[S, i \times S/n + k \times d]$ ;  $P_{V,1}[i$   
171  $\times S/n + k \times d, 0]$  and  $P_{V,2}[i \times S/n + k \times d, S]$ , where  $2 \leq i \leq n$  and the constant,  $k$ , is  
172 generated randomly by the computer and lies within the limits  $0 \leq k \leq 1$ . The sign of  
173  $k$  is also generated randomly by the computer.

174 A matrix of intercepts  $\{x_{\text{int}}(i, j), y_{\text{int}}(i, j)\}$  is generated for the intersection of each pair  
 175 of intervals for  $\{(i, j): 2 \leq (i, j) \leq n\}$  using the following system of linear equations:

176  
 177 
$$d_1 = x_{\text{H}}(i, 2) - x_{\text{H}}(i, 1) \quad (2)$$

178  
 179 
$$m_1 = [y_{\text{H}}(i, 2) - y_{\text{H}}(i, 1)]/d_1, \text{ for } d_1 \neq 0 \quad (3)$$

180  
 181 
$$d_2 = x_{\text{V}}(j, 2) - x_{\text{V}}(j, 1) \quad (4)$$

182  
 183 
$$m_2 = [y_{\text{V}}(j, 2) - y_{\text{V}}(j, 1)]/d_2, \text{ for } d_2 \neq 0 \quad (5)$$

184  
 185 
$$d_3 = m_2 - m_1 \quad (6)$$

186  
 187 
$$x_{\text{int}}(i, j) = [m_2 x_{\text{V}}(j, 1) - m_1 x_{\text{H}}(i, 1) + y_{\text{H}}(i, 1) - y_{\text{V}}(j, 1)]/d_3, \text{ for } d_3 \neq 0 \quad (7)$$

188  
 189 
$$y_{\text{int}}(i, j) = m_1 [x_{\text{int}}(i, j) - x_{\text{H}}(i, 1)] + y_{\text{H}}(i, 1) \quad (8)$$

190  
 191 The area,  $A$ , of a quadrilateral  $Q(P_1, P_2, P_3, P_4)$  defined by the points  $P_1(x_1, y_1)$ ,  
 192  $P_2(x_2, y_2)$ ,  $P_3(x_3, y_3)$ , and  $P_4(x_4, y_4)$  is given by equation (9) if  $P_1, P_2, P_3$  and  $P_4$  lie in  
 193 sequential order on the perimeter (McLanaghan & Levy, 1996):

194  
 195 
$$A = [(x_1 y_2 - x_2 y_1) + (x_2 y_3 - x_3 y_2) + (x_3 y_4 - x_4 y_3) + (x_4 y_1 - x_1 y_4)]/2 \quad (9)$$

196  
 197 The area of each quadrilateral in the randomly disrupted grid is calculated  
 198 systematically from the matrix of intercepts (see equations (7) and (8)) and stored in  
 199 the array  $A(i)$  where  $1 < i < n^2$ . The array  $A(i)$  is then used to generate a frequency-  
 200 area histogram upon multiple Monte Carlo iterations of the above algorithm.

201  
 202 2.4 Image Analysis Algorithm

203  
 204 Optical micrographs of feather images in the form of black and white photographic  
 205 jpeg files were analysed using an imaging algorithm designed to systematically count  
 206 white pixel areas and compile a frequency-area (count) histogram. The jpeg  
 207 photographic standard stores images as a two-dimensional array of pixel information,  
 208 each pixel having a red, blue and green ( $R, B, G$ ) component along with "alpha"  
 209 channel information associated with each pixel that determines its transparency.

210  
 211 The devised image processing algorithm firstly removes grey-scale shading (where  $R$ ,  
 212  $G, B$  values are all equal but not equal to zero) from the image file by systematically  
 213 examining the  $R, G, B$  information associated with each pixel and setting the pixel to  
 214 either "white" ( $R = 255, G = 255, B = 255$ ) or "black" ( $R = 0, G = 0, B = 0$ ) in  
 215 accordance with a ( $R_t, G_t, B_t$ ) threshold set by the user. The algorithm then counts the  
 216 number of sequential white pixels in each line of the image array and compiles a

217 histogram of the frequency of a given number of sequential white pixels. The latter is  
218 deemed to be directly related to the distribution of areas appearing within the grid  
219 pattern created by the feather microstructure.

220

221 Provision was made in the software to execute averaging cycles on the raw histogram  
222 data. In each cycle the average frequency of two successive frequencies in the  
223 histogram is calculated and recorded as the frequency of the upper of the two  
224 corresponding area ranges. This process has the effect of smoothing the distribution  
225 for the convenient analysis of the associated characteristic parameters. For the  
226 systems reported in the current work only one data averaging cycle was found to be  
227 necessary.

228

### 229 **3. Results and Discussion**

230

#### 231 3.1 Analysis of Computer-Generated Grids

232

233 Figure 1 shows three grids that were generated by the Monte Carlo computer  
234 algorithm using three levels of grid disruption. The different extent of disruption to  
235 the coherence of the grid pattern can be clearly seen as the disruption factor,  $f$ , is  
236 increased from zero to 1.0.

237

238 >>>Insert Figure 1

239

240 The distribution of the individual quadrilateral areas comprising the grid as a function  
241 of the degree of disruption of the grid was explored by generating multiple grids at a  
242 given extent of disruption ( $f$ ) and accumulating the frequencies in the area distribution  
243 histogram. The areas of the individual quadrilaterals comprising the grid were  
244 calculated in accordance with equation (9). The results are shown in Figure 2 for a  
245 selection of  $20 \times 20$  grids generated with various values of the  $f$  parameter and where  
246 each frequency is the cumulated frequency after 100 iterations of the Monte Carlo  
247 grid disruption cycle and where a resolution of 50 area channels was used.

248

249 >>>Insert Figure 2

250

251 It is clear from Figure 2 that the width of the theoretical area distribution increases as  
252 the extent of disruption to the grid increases. There is also a noticeable shift in the  
253 maximum towards a lower value of the mean area with increasing values of  $f$  and  
254 possible evidence of a bimodal distribution of the quadrilateral grid areas at the  
255 extreme  $f = 1$  value. The observation that there is a general broadening of the  
256 distribution with increasing  $f$  values supports the notion that the width of the area  
257 distribution function is an indicator of the extent of disruption to the coherence of the  
258 grid. To test the latter notion, the ratio  $w/h$  where  $w$  is the width of the distribution  
259 taken across the distribution at a frequency corresponding to half the maximum  
260 frequency,  $h$ , was plotted as a function of  $f$ . This plot is shown in Figure 3 for the grid

261 generating conditions used to create the data shown in Figure 2 and calculating data  
262 for  $f$  values ranging from 0.1 to 1.0 at increments of 0.1.

263

264 >>>Insert Figure 3

265

266 The data in Figure 3 suggest that a high degree of correlation exists between the  $w/h$   
267 ratio and the extent of disruption to the coherence of the grid. The relationship  
268 appears to be non-linear with the sensitivity of the  $w/h$  ratio as a measure of grid  
269 coherency increasing with an increasing extent of disruption. It is also clear that the  
270 distribution of the separate areas that comprise the overall grid can, in principle, be  
271 used to measure the overall coherence of the grid pattern.

272

### 273 3.2 Image Analysis

274

#### 275 3.2.1 Image Analysis of Computer-Generated Grids

276

277 Having established theoretically the relationship between the  $w/h$  ratio and the  $f$   
278 parameter, images of the computer-generated grids were analysed using the image  
279 analysis algorithm to produce area distribution data that could be compared with those  
280 data calculated directly from the grid-generating algorithm.

281

282 Figure 4 shows a selection of area distribution plots where the data were obtained by  
283 applying the image analysis algorithm to grid images ( $20 \times 20$  grids;  $200 \times 200$   
284 pixels) that were, in turn, generated by the grid-generating algorithm. The histogram  
285 data were processed using one averaging cycle.

286

287 >>>Insert Figure 4

288

289 Inspection of Figure 4 reveals similar behaviour to that observed in Figure 2 and  
290 suggests that the image analysis algorithm may be used to determine area distribution  
291 data from a micrograph image and subsequently the  $w/h$  ratio for these data. To  
292 explore this further a series of images of computer-generated grids with values of the  $f$   
293 parameter ranging from 0.1 to 1.0 at increments of 0.1 was processed using the image  
294 analysis algorithm and the  $w/h$  ratio for each grid was determined. Figure 5 shows a  
295 plot of the  $w/h$  ratio as a function of the  $f$  parameter.

296

297 >>>Insert Figure 5

298

299 Figure 5 reveals a high degree of correlation between the two variables, as was also  
300 observed in the case of Figure 3, and the relationship between the two variables is,  
301 once again, non-linear. The correspondence between the pixel areas as determined by  
302 the image analysis algorithm (i.e. a linear pixel-counting routine) and the actual areas  
303 of the grid quadrilaterals (i.e. calculated in accordance with equation (9)) can be  
304 confirmed by plotting the corresponding  $w/h$  ratios derived from each of these

305 techniques for the same series of computer-generated disrupted grids. Such a plot is  
306 presented in Figure 6 where  $(w/h)_{\text{img anal}}$  is the  $w/h$  ratio determined from the image  
307 analysis algorithm and  $(w/h)_{\text{calc}}$  is the  $w/h$  ratio determined by calculation in  
308 accordance with equation (9).

309

310 >>>Insert Figure 6

311

312 The linearity of the plot in Figure 6 confirms that the image analysis algorithm  
313 produces a reliable representation of the area distribution histogram of a disrupted  
314 two-dimensional grid and can therefore be explored further for use in determining the  
315  $w/h$  ratio in the case of a feather micrograph image.

316

### 317 3.2.2 Benchmarking Image Analysis Algorithm

318

319 Previous preliminary work on feather image analysis (Ryan, 2005; Ngeh, 2002)  
320 calibrated the area distributions of feather grid patterns that were determined utilizing  
321 software that was commercially available at the time (Ryan, 2005) against  
322 distributions that were determined from a manual "cut and weigh" (CW) method. The  
323 latter involved carefully cutting out the individual grid elements from an enlarged  
324 print of the micrograph of the feather and weighing the individual pieces to indirectly  
325 determine the separate areas (Ryan, 2005). These data were then used to compile the  
326 area distribution histogram. The CW method is clearly quite tedious and can be  
327 considered somewhat subjective in that it can be difficult to treat consistently the  
328 "grey scale" regions of the image. Nonetheless, the method does provide a reasonable  
329 benchmark distribution to test the image analysis algorithm and from which to make  
330 meaningful comparisons. Furthermore, the method can overcome a difficulty that  
331 was experienced when using the commercially available area imaging software where  
332 the software did not always distinguish correctly each of the individual area  
333 components comprising the grid-like pattern. It is therefore suggested that the pixel  
334 counting algorithm described in the current work can provide a more reliable and  
335 consistent analysis of the image as it systematically counts every sequence of white  
336 pixels in the image and, as such, does not rely on the correct initial identification of  
337 the separate area boundaries.

338

339 The image analysis algorithm was further tested on the image of a feather that was  
340 previously characterized (Ryan, 2005) using the CW method described above. Figure  
341 7 shows the area distribution histograms of the grid of a duck feather sample where  
342 the respective histograms were independently obtained using the image analysis  
343 algorithm ( $158 \times 290$ -pixel micrograph) and the CW method.

344

345 >>>Insert Figure 7

346



347 To directly compare the two histograms, both the frequency and area domains were  
348 normalized with the frequency domain being normalized at the maximum frequency.  
349 Each distribution was fitted with a 6th order polynomial function to subsequently  
350 achieve a consistent analysis of its characteristic  $w/h$  parameter. Moreover, Figure 7  
351 shows a satisfactory extent of superposition of the two distributions and further  
352 analyses of these reveals a  $w/h$  ratio calculated *via* the data obtained from the image  
353 analysis algorithm,  $(w/h)_{\text{img}} = 0.962$  and that obtained *via* the CW method,  $(w/h)_{\text{CW}} =$   
354  $0.955$ . There is only a very small difference (less than 1%) between these two values.  
355

### 356 3.2.3 Image Analysis of Treated Feathers

357

358 Figure 8 shows micrograph images ( $100 \times 290$  pixels) of duck feather samples  
359 following treatment with detergent or magnetic particles along with the control  
360 sample (no treatment). In each case the image on the left of each pair is the original  
361 image and the image on the right of the pair is that which was obtained using the  
362 image analysis algorithm following grey scale removal. These images clearly show  
363 the extents of disruption to the grid patterns that have been imparted by the different  
364 treatments where the treatment using magnetic particles seemingly imparts less  
365 disruption to the grid pattern than that of the detergent treatment. *It is important to*  
366 *note however, that such disruption to the grid pattern following these different*  
367 *treatments provides an indication of the feather condition at the time of treatment. It*  
368 *does not necessarily indicate the condition of the feather in the longer term when, for*  
369 *example, preening and the return of preening oils may improve feather condition.*  
370

371 >>>Insert Figure 8

372

373 The images shown in Figure 8 were further analysed using the software and the  
374 respective area distribution histograms were produced (see Figure 9). The plots in  
375 Figure 9 confirm quantitatively the observations that can be made by visual inspection  
376 of Figure 8. The profiles of the control sample and the sample treated with magnetic  
377 particles are similar and have  $w/h$  ratios of 0.561 and 0.600 respectively. This  
378 represents a difference of *ca.* 7%. The  $w/h$  ratio for the detergent-treated sample is  
379 significantly greater (0.743) than either of the latter (difference of *ca.* 20%).  
380

381 >>>Insert Figure 9

382

383 The distribution histogram for the detergent treated sample exhibits a long "tail" that  
384 corresponds to the larger open areas in its grid-like pattern. These are apparent in the  
385 corresponding micrograph image and reflect the greater extent of disruption to the  
386 grid pattern caused by the detergent treatment as compared with the magnetic particle  
387 treatment (see Figure 8). A very strong correlation between the  $w/h$  ratio for feathers  
388 treated with detergent and the concentration of detergent has also been shown to exist

389 (Orbell *et al.*, 2007) in the case where the *w/h* ratio was determined manually using  
390 the CW method.

391

#### 392 **4. Conclusions**

393

394 The image analysis algorithm developed in this work provides a reliable quantitative  
395 means of determining the coherency of a two-dimensional grid-like pattern that  
396 reflects the feather microstructure of the Mallard Duck. The algorithm produces a  
397 result that is consistent with theoretical computer-generated grids as well as with the  
398 pattern observed in the micrographs of the feather microstructure. This analysis  
399 technique successfully identified differences in the grid coherency of the feather  
400 structure following different cleansing treatments and, as such, is proposed as a  
401 possible quantitative indicator of the condition of this type of feather [at the point in](#)  
402 [time following the treatment. The effects of residual oil, metal, detergent or other](#)  
403 [contaminants on the outcome produced by the technique has not been taken into](#)  
404 [account and is yet to be explored.](#)

405

406 Nonetheless, this work provides an important proof of principle, namely that such  
407 indicators could be useful for the routine quantitative assessment of feather condition,  
408 either for the trialling of different treatment protocols, as a tool during the  
409 rehabilitation process, or as an assessment of feather condition in veterinary studies  
410 along with other indicators, to give a more complete and objective assessment of  
411 feather condition than is currently possible. [Furthermore, it is suggested that the](#)  
412 [method will be a valuable tool in helping to answer some remaining key questions](#)  
413 [such as: how much if any, of the feather disruption is permanent and how much can](#)  
414 [be reversed by preening and what is the impact of this disruption on a whole living](#)  
415 [animal and its feathers in a variety of different environments and life stages. Work](#)  
416 [along these lines is continuing in our laboratory.](#)

417

418

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425

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514 **Figure Captions**

515

516 **Fig 1.** Two-dimensional grids generated by the Monte Carlo computer algorithm and  
517 that were randomly disrupted using disruption factors of: (i)  $f = 0$ , (ii)  $f = 0.6$   
518 and (iii)  $f = 1.0$ .

519

520 **Fig 2.** Selection of area distribution histograms for the quadrilaterals comprising  
521 computer-generated  $20 \times 20$  grids subjected to different extents of random  
522 disruption as determined by the disruption parameter  $f$  where:  $f = 0.3$  (filled  
523 circles),  $f = 0.6$  (open circles) and  $f = 0.9$  (filled squares).

524

525 **Fig 3.** Plot of the ratio  $w/h$  versus the disruption parameter  $f$  for a computer-  
526 generated grid where the grid component areas were calculated from equation  
527 (9). Calculation parameters:  $20 \times 20$  grid; 100 iterations; 50 channels area  
528 resolution.

529

530 **Fig 4.** Selection of area distribution histograms for  $20 \times 20$  grids produced by the  
531 grid generating algorithm and where the images were processed by the image  
532 analysis algorithm using 1 data averaging cycle. Disruption parameters:  $f =$   
533  $0.3$  (filled circles),  $f = 0.6$  (open circles) and  $f = 0.9$  (filled squares).

534

535 **Fig 5.** Plot of the ratio  $w/h$  versus the disruption parameter  $f$  for area distribution  
536 histogram data derived from computer-generated  $20 \times 20$  grids where the  
537 images were processed by the image analysis algorithm using 1 data averaging  
538 cycle.

539

540 **Fig 6.** Plot of  $(w/h)_{\text{img anal}}$  versus  $(w/h)_{\text{calc}}$  where the corresponding ratios were  
541 determined for a series of computer-generated grids with the disruption  
542 parameter  $f$  varying between 0.1 and 1.0 at increments of 0.1.

543

544 **Fig 7.** Area distribution histograms of the grid-like pattern of a Mallard Duck feather  
545 micrograph where the distributions were independently determined using: (i)  
546 the image analysis algorithm (filled circles) and (ii) the CW method (open  
547 circles).

548

549 **Fig 8.** Micrograph images ( $5\times$ ) of Mallard Duck feather samples following treatment  
550 with: (i) detergent and (ii) magnetic particles. The control sample (no  
551 treatment) is also shown. In each pair of images, the image on the left is the  
552 original and the one on the right is the corresponding image with its grey scale  
553 removed.

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556 **Fig 9.** Area distribution histograms derived from the image analysis algorithm (5  
557 data averaging cycles) for Mallard Duck feather samples (Figure 8) that were  
558 subjected to: (i) detergent (filled squares) and (ii) magnetic particle (open  
559 circles) treatment. The control sample (filled circles) is shown for  
560 comparison.  
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562 **Figure 1**

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582 **Figure 2**

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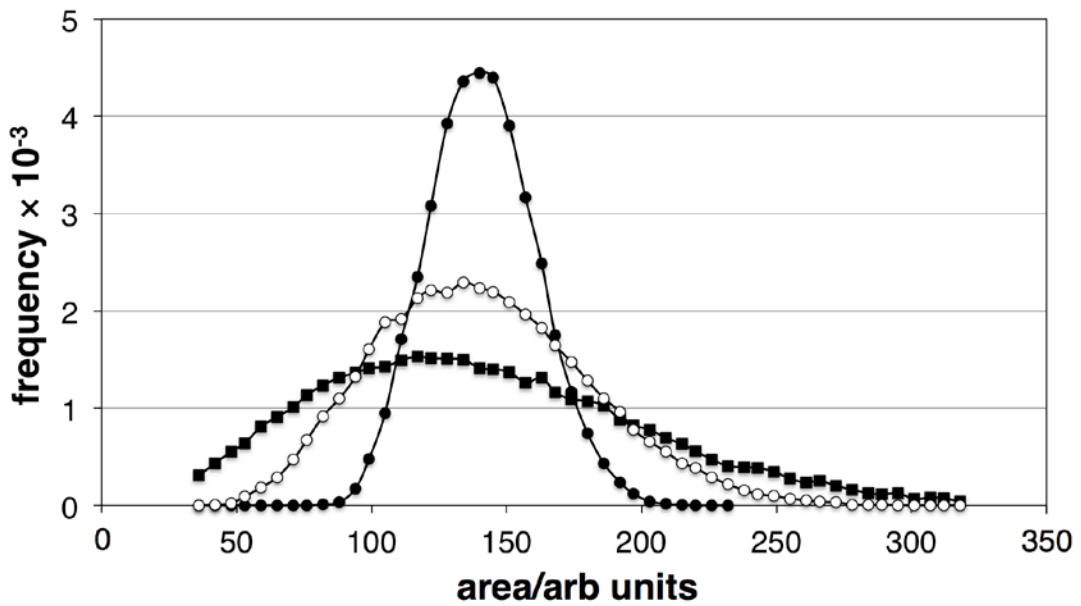
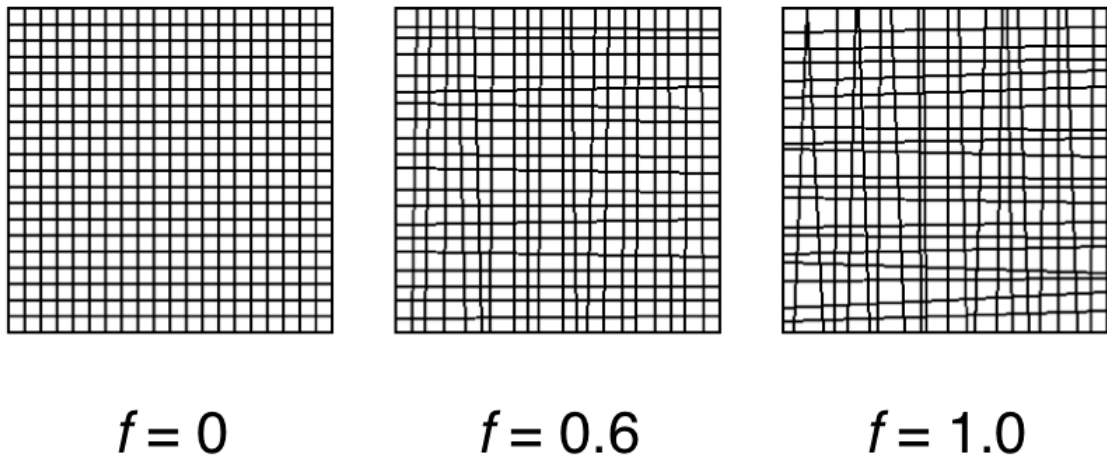
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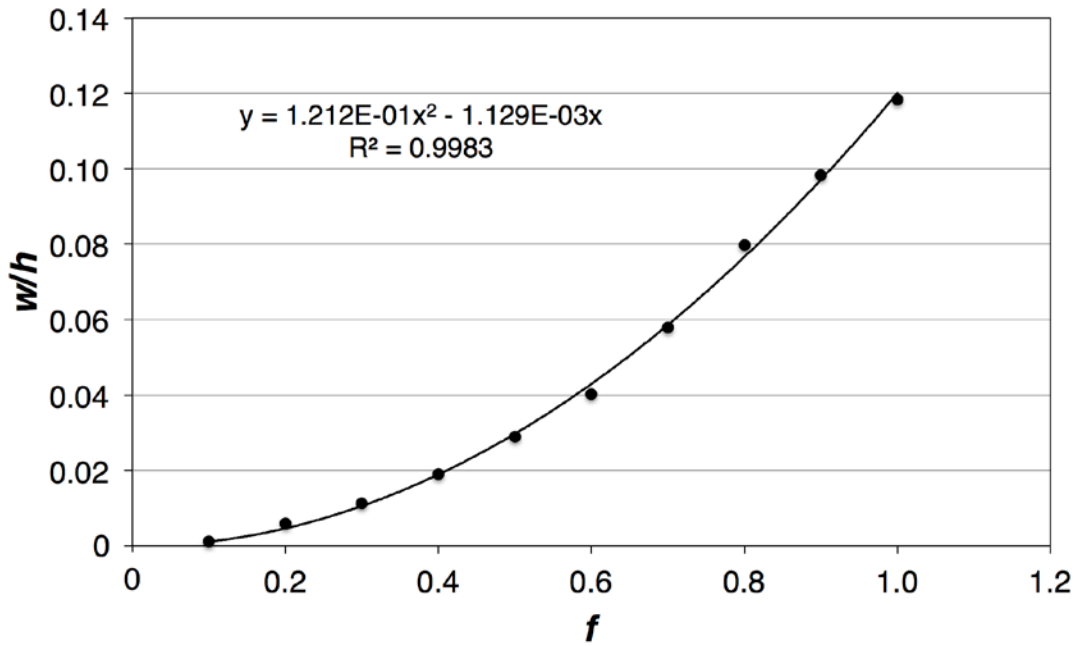
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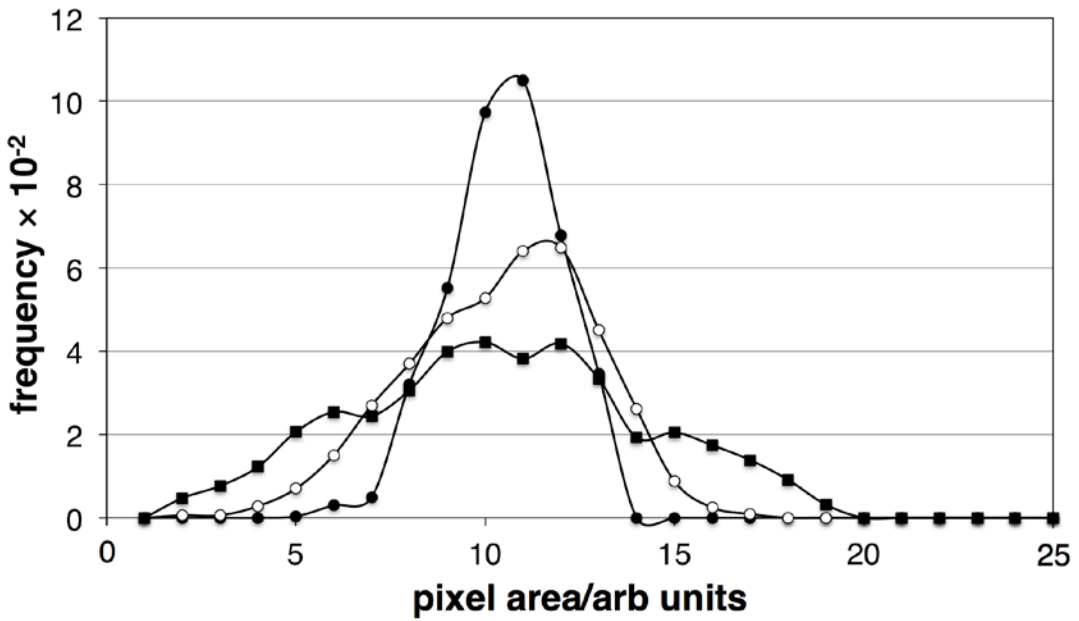
601 **Figure 3**

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623 **Figure 4**

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645 **Figure 5**

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667 **Figure 6**

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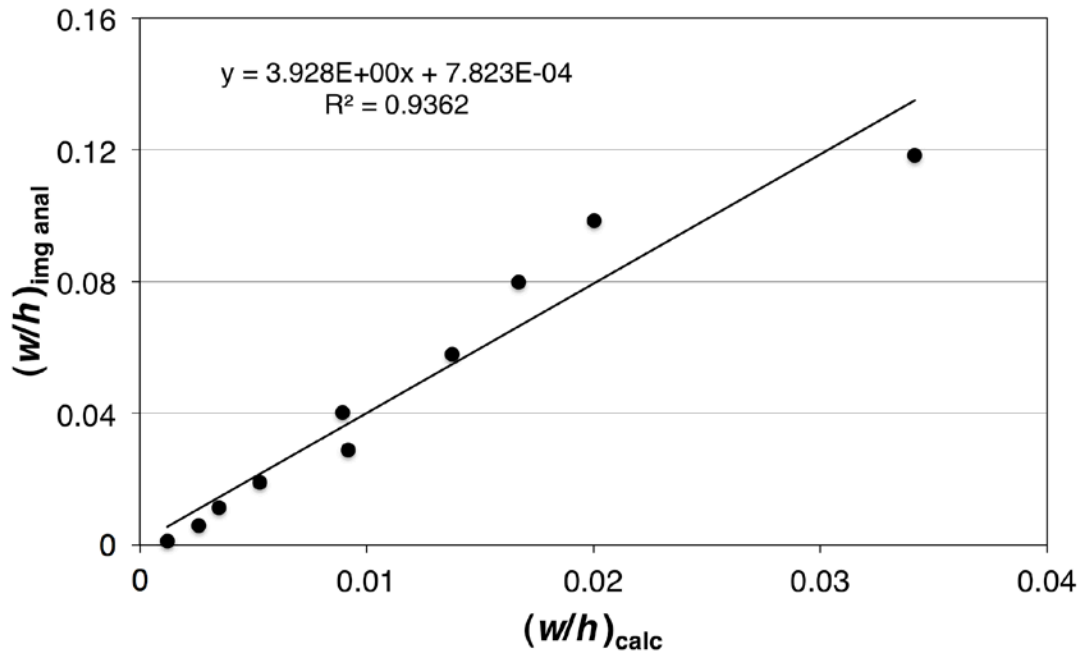
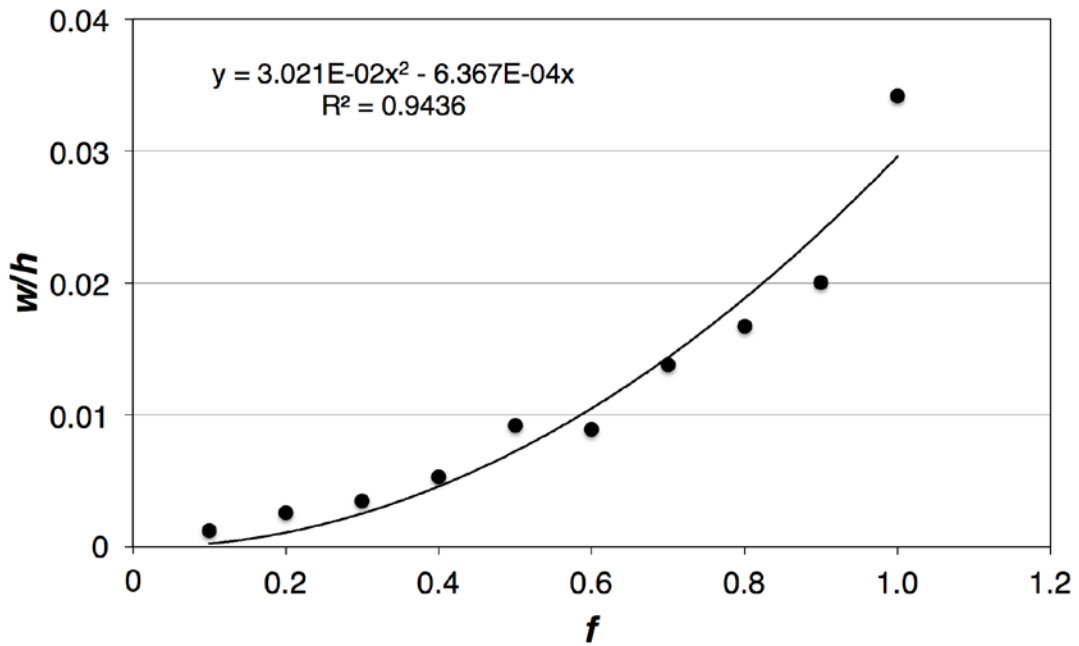
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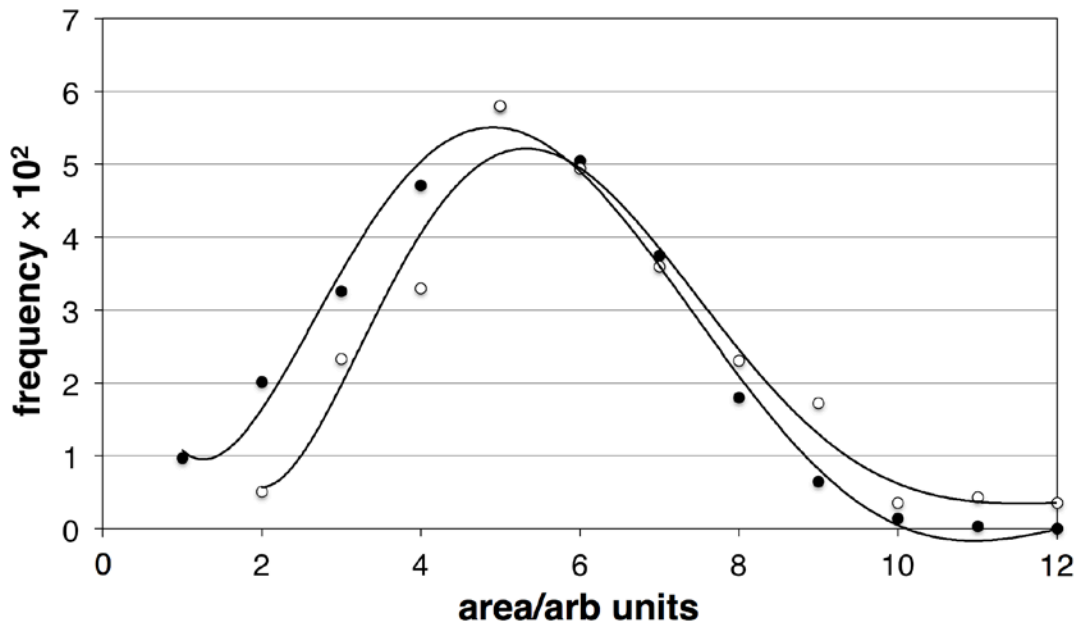
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689 **Figure 7**



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711 **Figure 8**

