

## COMMUNICATION

# Room Temperature Alkynylation of *H*-Phosphi(na)tes and Secondary Phosphine Oxides with Ethynylbenziodoxolone (EBX) Reagents

Cite this: DOI: 10.1039/x0xx00000x

Received 00th January 2012,

Accepted 00th January 2012

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DOI: 10.1039/x0xx00000x

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This is the peer reviewed version of the following article: Chem. Commun. 2014,50, 12923, which has been published in final form at <http://pubs.rsc.org/en/Content/ArticleLanding/2014/CC/C4CC06851C#!divAbstract>

**Highly efficient protocols for the alkynylation of *H*-phosphi(na)tes and secondary phosphine oxides with silyl, aryl and alkyl ethynyl-benziodoxolone (EBX) reagents are reported. Alkynyl phosphorus compounds were obtained in 69-93% yield without the need for a transition metal catalyst at room temperature under open flask conditions.**

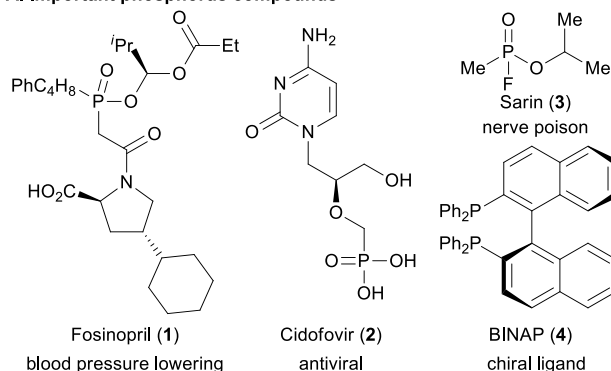
Phosphorus-based functional groups are important in biology, medicine and synthetic chemistry.<sup>1</sup> Synthetic phosphorus derivatives display a broad range of bioactivities, including drugs like the blood-pressure lowering fosinopril **1**<sup>2a</sup> and the antiviral cidofovir **2**<sup>2b</sup> and the nerve poison sarin **3** (Fig. 1, A). Furthermore, ligands such as BINAP **4** based on phosphorus play a privileged role in modern organic and organometallic chemistry. Phosphonium salts and phosphonic esters are also important building blocks to access alkenes via Wittig or Horner-Wadsworth-Emmons (HWE) reactions. The development of general methods for the synthesis of organophosphorus compounds is consequently an important goal in organic chemistry.

In this context, alkynyl phosphorus derivatives constitute an important class of building blocks because of the versatile reactivity of the triple bond (Fig 1, B).<sup>3</sup> Hydration of the alkyne leads to  $\beta$ -ketophosphonates, which are used in the HWE reaction.<sup>4</sup> The addition of other nucleophiles has been also intensively investigated.<sup>3b</sup> The use of alkynyl phosphorus in [3+2], [4+2] and [2+2+2] cycloadditions<sup>5</sup> gives access to important phosphorus cyclic compounds, including versatile chiral ligands for metal catalysis<sup>3b,5f-h</sup> or amino acids analogues for studies in chemical biology.<sup>5a</sup>

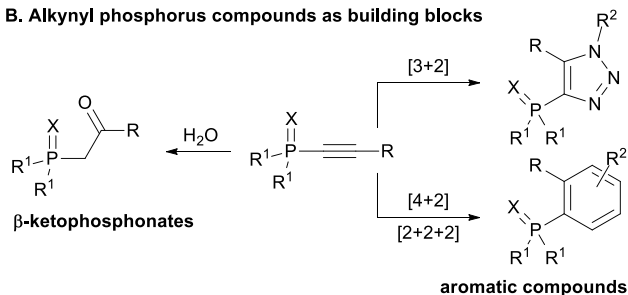
To date, four different approaches have been developed for the synthesis of alkynyl phosphorus compounds, including: (1) reaction of electrophilic alkynes or their precursors with nucleophilic phosphorus (III) reagents involving Michaelis-

Arbuzov and Michaelis-Becker reactions,<sup>4a,6</sup> (2) reaction of nucleophilic alkynes with electrophilic phosphorus,<sup>7</sup> (3)  $\beta$ -elimination of heteroatom-substituted vinylphosphonates,<sup>8</sup> and (4) metal-mediated reaction of *H*-phosphites with 1,1-dibromo alkenes, terminal acetylenes, propiolic acid derivatives, copper acetylides or bromo alkynes.<sup>9</sup>

### A. Important phosphorus compounds



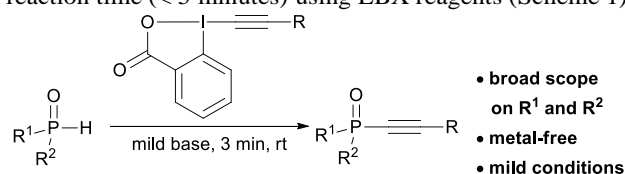
### B. Alkynyl phosphorus compounds as building blocks



**Fig. 1** Important phosphorus-containing bioactive molecules (A) and alkynyl phosphorus derivatives as important building blocks (B).

In addition, metal-catalyzed method to access alkynyl phosphines have also been reported.<sup>10</sup> Despite these different alternatives, there are currently no general access to alkynyl phosphorus compounds under mild conditions in absence of transition metals. Most works focus on only one class of phosphorus compounds, especially phosphonates.

In this context, alkynyl iodonium salts have been long known as electrophilic alkylation reagents,<sup>11</sup> but their use for the synthesis of alkynyl phosphorus compounds has been limited to phosphites<sup>6b</sup> and phosphonates salts.<sup>12</sup> Our group has discovered and exploited the properties of cyclic alkynyl benziodoxolone reagents, such as TIPS-ethynyl-benziodoxolone (TIPS-EBX, **6a**) for the alkylation of nucleophiles.<sup>13</sup> This class of reagents, first synthesized by Ochiai and Zhdankin,<sup>14</sup> has then later also been exploited by others.<sup>15</sup> Herein, we would like to report the successful alkylation of *H*-phosphites, *H*-phosphinates and secondary phosphine oxides at room temperature in open flask and a short reaction time (< 5 minutes) using EBX reagents (Scheme 1).



**Scheme 1** Our approach towards phosphorus alkylation.

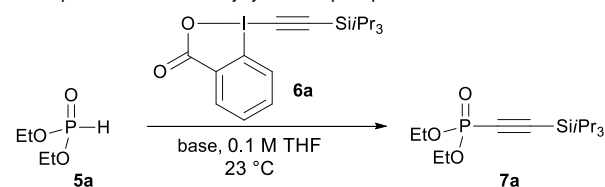
To start our studies, we selected 1,1,3,3-tetramethyl guanidine (TMG) for the preparation of alkyne **7a** from diethyl phosphinate **5a** with TIPS-EBX **6a**, as it has been the base of choice in the case of thiol nucleophiles.<sup>13h</sup> To our delight, 53% isolated yield of **7a** was obtained (Table 1, entry 1). Other amine bases such as 4-dimethylaminopyridine (4-DMAP), 1,4-diazabicyclo[2,2,2]octane (DABCO) and triazabicyclodecene (TBD) led to lower yields (entries 2-4). In contrast, the use of 1,8-diazabicycloundec-7-ene (DBU) gave product **7a** in 82% isolated yield (entry 5). A fast examination of base and reagent loading showed that 1.5 equivalents of DBU and 1.1 equivalents of TIPS-EBX **6a** were optimal (entries 6-8). Other solvents gave lower yields when compared to THF (entries 9-13). Alkynyl phosphite **7a** was already formed within 3 min in 90% isolated yield (entry 14). The formation of **7a** was not observed in the absence of DBU (entry 15).

With optimum conditions in hand the scope of *H*-phosphites and *H*-phosphinates was examined (Scheme 2).<sup>16</sup> Dimethyl and dibenzyl phosphites **6b** and **6c** gave the desired product **7b** and **7c** in 85% and 89% yield respectively. *H*-(*R*)-BINOL phosphite derivative **7d** was obtained in 86% yield. In addition, alkynes **7e** and **7f** were obtained from the corresponding phosphinates in 76% and 87% yield respectively. More complex (*R*)-phenyl menthyl and phenyl AZT *H*-phosphinates were also suitable substrates to deliver the products **7g** (71%) and **7h** (70%) as a mixture of diastereoisomers. However, phenylphosphinic acid **6i** could not be converted into alkyne **7i**. Substituted ethynyl-benziodoxolones (R-EBXs), such as Ph-EBX **6b**,<sup>13f</sup> *t*Bu-EBX **6c**,<sup>13f</sup> and *n*Hex-EBX **6d**<sup>15g</sup> could also be used in this direct alkylation successfully to give products **7j-l**. Notably, an 1,5-C-H insertion product resulting from a carbene intermediate was not observed in the alkylation of **5a** with *n*Hex-EBX **6d**, in contrast to what had been observed when ketoesters were used as nucleophiles with alkynyliodonium salts.<sup>17</sup>

We then turned our attention to the alkylation of secondary phosphine oxides (SPO) (Scheme 3).<sup>16</sup> The alkylation of this class of substrates has been much less studied in the past.

Indeed, to the best of our knowledge, only the alkylation of diphenyl phosphine oxide **8a** has been reported.<sup>6d,9e,9f</sup> Initially, the conditions used in alkylation of *H*-phosphites and *H*-phosphinates using DBU as a base (Table 1, entry 14) were tested for the alkylation of diphenyl phosphine oxide **8a**, but only 60% isolated yield of **9a** was obtained. TMG, which had been the second best base in the case of phosphites, gave better results and the alkylation product **9a** was isolated in 91% yield. This protocol could also be used in the synthesis of di-(4-tolyl) phosphinite **9b** (89%), di-(4-fluorophenyl) phosphinite **9c** (93%), di-(*n*-butyl) phosphinite **9d** (86%), and phenyl-*t*-butyl phosphinite **9e** (87%). Significantly, cyclic phosphinite **9f** can be made in 89% yield. However, bulky substituted phosphinite, such as **9g** and **9h** could not be synthesized. We further investigated the synthesis of ethynyl phosphinite bearing aryl or alkyl groups on the alkyne. Phenylethynyl phosphine **9i** and *t*-butylethynyl phosphine **9j** were obtained in 72% and 87% yield respectively. *n*-Hexyl ethynyl phosphine **9k** could also be obtained in 85% yield.

**Table 1** Optimization of the alkylation of phosphite **8a**.

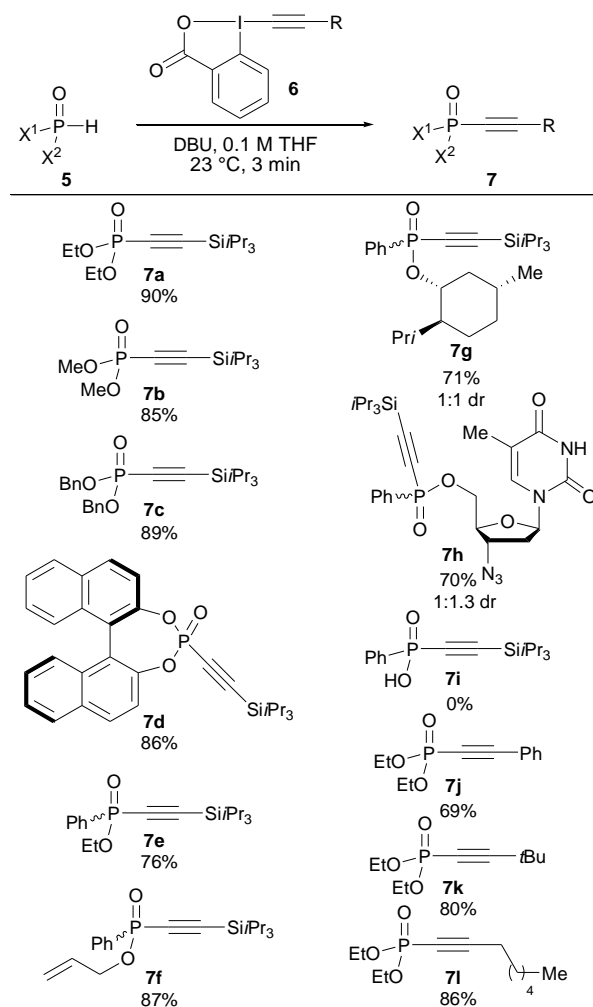


entry	equiv. <b>6a</b>	base (equiv.)	solvent	time (min)	yield (%) <sup>a</sup>
1	1.2	TMG (2.0)	THF	90	55 (53) <sup>b</sup>
2	1.2	4-DMAP (2.0)	THF	90	0
3	1.2	DABCO (2.0)	THF	90	30
4	1.2	TBD (2.0)	THF	90	0
5	1.2	DBU (2.0)	THF	90	85 (82) <sup>b</sup>
6	1.2	DBU (1.5)	THF	90	84
7	1.2	DBU (1.2)	THF	90	50
<b>8</b>	<b>1.1</b>	<b>DBU (1.5)</b>	<b>THF</b>	<b>90</b>	<b>89 (88)<sup>b</sup></b>
9	1.1	DBU (1.1)	THF	90	45
10	1.1	DBU (1.5)	<i>i</i> -PrOH	90	5
11	1.1	DBU (1.5)	TBME	90	22
12	1.1	DBU (1.5)	MeCN	90	61
13	1.1	DBU (1.5)	CH <sub>2</sub> Cl <sub>2</sub>	90	23
<b>14</b>	<b>1.1</b>	<b>DBU (1.5)</b>	<b>THF</b>	<b>3</b>	<b>92 (90)<sup>b</sup></b>
15	1.1	DBU (0)	THF	3 or 90	0

<sup>a</sup> 0.05 mmol **5a** was used. The yield is obtained from <sup>1</sup>H-NMR with 1,3,5-trimethoxybenzene as internal reference. <sup>b</sup> Isolated yield after purification on column chromatography.

Several plausible mechanisms could be envisaged for the direct alkylation reaction. For the reaction of alkynyliodonium salts with carbon nucleophiles, conjugate addition onto the β-carbon of ethynyl-benziodoxolone, followed by an α-elimination of the aryl iodide and 1,2-shift rearrangement to deliver the alkyne has been most often proposed.<sup>17</sup> The fact that no products resulting from C-H insertion had been observed in the case of aliphatic alkynes could be rationalized by a fast 1,2-shift of the phosphorus atom. Nevertheless, an alternative mechanism involving nucleophilic attack of phosphi(na)te/phosphine oxide onto the iodine atom of benziodoxolone, followed by C-P bond formation to give the alkyne product cannot be excluded at this stage. A single electron transfer (SET) from phosphi(na)te/phosphine oxide to the ethynyl-benziodoxolone, could also be proposed as a first step in the reaction.

Recombination on the iodine to give an I-P bond followed by reductive elimination or direct alkylation of the phosphorus atom would then lead to the observed product. However, when TEMPO was added to the reaction mixture under the standard reaction conditions (Table 1, entry 14), alkyne **7a** was still obtained in 85% yield. This result suggests that long-living radical intermediates are probably not involved. It is also important to notice that SET pathways often require further activation of the hypervalent iodine reagent by a Lewis or Brønsted acid to facilitate electron transfer, and the developed method proceeded under basic conditions.<sup>18</sup>

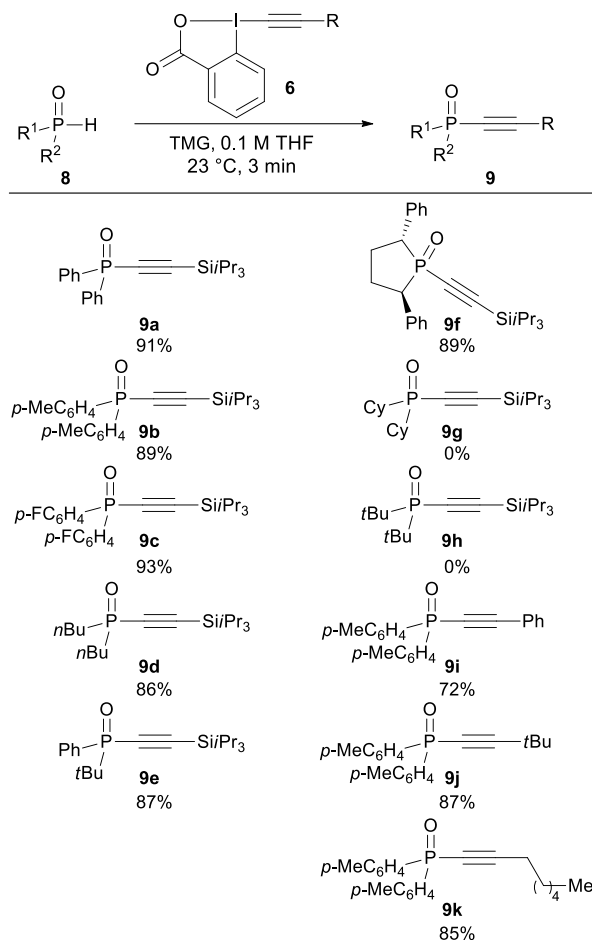


**Scheme 2** Scope of the alkylation of phosphites and phosphinates. Reaction conditions: 0.15–0.29 mmol of *H*-phosphite or *H*-phosphinate **5** (1.0 equiv.), 1.5 equiv. of DBU and 1.2 equiv. of R-EBX **6**, 0.1 M in THF at 23 °C. Isolated yield after purification on column chromatography is given. <sup>b</sup>2.5 equiv. of DBU was used in the case of **7i**.

## Conclusions

In conclusion, we have described very simple and general protocols for the alkylation of *H*-phosphites, -phosphinates, and secondary phosphine oxides using ethynyl-benziodoxolone reagents in good to excellent yield. The developed alkylation method proceeded at room temperature in a few minutes and did not require the use of transition metals. It could be applied to the synthesis of silyl, alkyl and aryl substituted alkynes and was efficient for a broad range of different substituents on

phosphorus. Further extension of the scope, application of the obtained building blocks in synthesis, and more detailed investigation of the reaction mechanism are currently ongoing in our laboratory and the results will be reported in due course.



**Scheme 3** Scope of the *H*-phosphine oxides. Reaction conditions: 0.15–0.29 mmol of *H*-phosphine oxide **8** (1.0 equiv.), 1.5 equiv. of TMG and 1.5 equiv. of R-EBX **6**, 0.1 M in THF at 23 °C. Isolated yield after purification on column chromatography is given.

## Notes and references

EPFL, F. Hoffmann-La Roche Ltd and ERC (Starting Grant iTools4MC, number 334840) are acknowledged for financial support. We also thank Dr. P. A. Donets and Prof. N. Cramer from the Laboratory of Asymmetric Catalysis and Synthesis at EPFL for donation of starting materials **5d**, **8e**, **8f** and **8g**. Dr. C. C. Chen dedicates this article to Prof. V. K. Aggarwal FRS at University of Bristol.

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Electronic Supplementary Information (ESI) available: [details of any supplementary information available should be included here]. See DOI: 10.1039/c000000x/

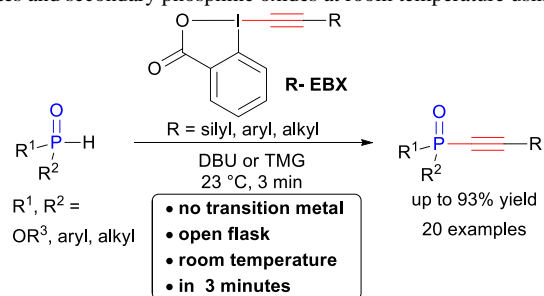
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## COMMUNICATION

## Table of Content Entry

We report the alkylation of *H*-phosphi(na)tes and secondary phosphine oxides at room temperature using ethynylbenziodoxolone (EBX) reagents.



# **Room Temperature Alkynylation of *H*-Phosphi(na)tes and Secondary Phosphine Oxides with Ethynylbenziodoxolones (EBX) Reagents**

**C. Chun Chen and Jerome Waser\***

**(60 pages)**

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4. Spectra of New Compounds	p. S17

## 1. General Method

All chemicals were purchased from Acros, Aldrich, Fluka, VWR, Aplichem, Maybrige, TCI or Merck and used as such unless stated otherwise. Chromatographic purification was performed as flash chromatography using Macherey-Nagel silica 40-63, 60 Å, using the solvents indicated as eluent with 0.1-0.5 bar pressure. TLC was performed on Merck silica gel 60 F<sub>254</sub> TLC glass plates or aluminium plates and visualized with UV light, permanganate stain, CAN stain or Anisaldehyde stain. Melting points were measured on a Büchi B-540 melting point apparatus using open glass capillaries, the data is uncorrected. <sup>1</sup>H-NMR spectra were recorded on a Bruker DPX-400 400 MHz spectrometer in chloroform-d, DMSO-d<sub>6</sub> or CD<sub>3</sub>OD, all signals are reported in ppm with the internal chloroform signal at 7.26 ppm, the internal DMSO signal at 2.50 ppm or the internal methanol signal at 3.30 ppm as standard. The data is being reported as (s = singlet, d = doublet, t = triplet, q = quadruplet, qi = quintet, m = multiplet or unresolved, br = broad signal, app = apparent, coupling constant(s) in Hz, integration, interpretation). <sup>13</sup>C-NMR spectra were recorded with <sup>1</sup>H-decoupling on a Bruker DPX-400 100 MHz spectrometer in chloroform-d, DMSO-d<sub>6</sub> or CD<sub>3</sub>OD, all signals are reported in ppm with the internal chloroform signal at 77.0 ppm, the internal DMSO signal at 39.5 ppm or the internal methanol signal at 49.0 ppm as standard. <sup>31</sup>P-NMR spectra were recorded on a Bruker DPX-400 162 MHz spectrometer in chloroform-d. <sup>19</sup>F-NMR spectra were recorded on a Bruker DPX-400 376 MHz spectrometer in chloroform-d. No internal standard was used for <sup>31</sup>P-NMR and <sup>19</sup>F-NMR spectra. Infrared spectra were recorded on a JASCO FT-IR B4100 spectrophotometer with an ATR PRO410-S and a ZnSe prisma and are reported as cm<sup>-1</sup> (w = weak, m = medium, s = strong, br = broad). Gas chromatographic and low resolution mass spectrometric measurements were performed on a Perkin-Elmer Clarus 600 gas chromatographer and mass spectrometer using a Perkin-Elmer Elite fused silica column (length: 30 m, diameter: 0.32 mm) and Helium as carrier gas. High resolution mass spectrometric measurements were performed by the mass spectrometry service of ISIC at the EPFL on a MICROMASS (ESI) Q-TOF Ultima API.

## 2. Synthesis of Reagents and Starting Materials

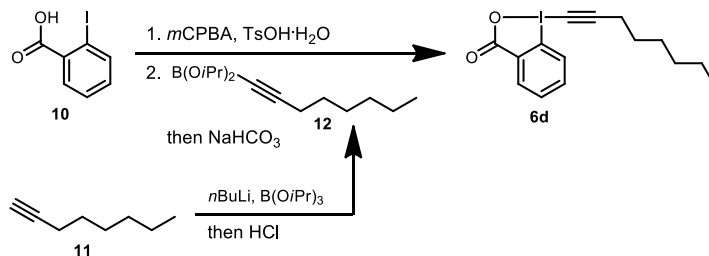
TIPS-EBX (**6a**), Ph-EBX (**6b**) and *t*Bu-EBX (**6c**) were made using our reported protocols.<sup>1</sup> *n*Hex-EBX (**6d**) was made using a reported procedure.<sup>2</sup>

Diethyl phosphite (**5a**), dimethyl phosphite (**5b**), dibenzyl phosphite (**5c**), ethyl phenylphosphinate (**5e**), phenyl phosphoric acid (**5i**), di-phenylphosphine oxide (**8a**), 3'-Azido-3'-deoxythymidine, Di-*tert*-butylphosphine oxide (**8h**) are commercially available from Sigma-Aldrich or TCI.

Dinaphtho[2,1-d:1',2'-f][1,3,2]dioxaphosphepine 4-oxide (**5d**), *tert*-butyl(phenyl)phosphine oxide (**8e**), (2*R*,5*R*)-2,5-diphenylphospholane 1-oxide (**8f**), and dicyclohexylphosphine oxide (**8g**) were generously given by Dr. Pavel Donets and Prof. Nicolai Cramer from the Laboratory of Asymmetric Catalysis and Synthesis at EPFL.

Allyl phenylphosphinate (**5f**),<sup>3</sup> (1*R*,2*S*,5*R*)-2-isopropyl-5-methylcyclohexyl phenylphosphinate (**5g**),<sup>4</sup> di-*p*-tolylphosphine oxide (**8b**),<sup>5</sup> bis(4-fluorophenyl)phosphine oxide (**8c**),<sup>4</sup> dibutylphosphine oxide (**8d**),<sup>4</sup> *t*-butyl(phenyl)phosphine oxide (**8e**), were made using reported literature procedures.

### Octynyl-1,2-benziodoxol-3(1*H*)-one (**6d**)



Following a slightly modified procedure,<sup>6</sup> a solution of 1-octyne (**11**) (747 mg, 6.78 mmol, 1.00 eq.) and dry diethyl ether (7.0 mL) was cooled to -78 °C, at which temperature 1.6 M *n*BuLi in hexanes (4.24 mL, 6.78 mmol, 1.00 eq.) was added dropwise. The mixture was stirred at -78 °C for 90 minutes and then cannulated into a to -78 °C pre-cooled solution consisting of triisopropyl borate (1.56 mL, 6.78 mmol, 1.00 eq.) and dry diethyl ether (7.0 mL). The reaction mixture was stirred at -78 °C for 2 hours, after which 2.0 M HCl in diethyl ether (3.73 mL, 7.46 mmol, 1.10 eq.) was added. The cooling bath was removed and the mixture was stirred for an additional 60 minutes. After filtration and solvent removal, Kugelrohr distillation (75 °C at 0.6 mbar) furnished pure diisopropyloct-1-ynylboronate (**12**, 940 mg, 3.95 mmol, 58% yield) as a colorless liquid. <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) δ 4.55 (hept, 2H, *J* = 6.2 Hz, CHO), 2.27 (t, 2H, *J* = 7.0 Hz,

<sup>1</sup> (a) Brand, J. P.; Waser, J. *Synthesis* **2012**, *44*, 1155. (b) Brand, J. P.; Chevalley, C.; Scopelliti, R.; Waser, J. *Chem. Eur. J.* **2012**, *18*, 5655.

<sup>2</sup> Meouma, J. B.; Olofsson, B. *Chem. Eur. J.* **2012**, *18*, 3690.

<sup>3</sup> Fourgeaud, P.; Midrier, C.; Vors, J.-P.; Volle, J.-N.; Pirat, J.-L.; Virieux, D. *Tetrahedron* **2010**, *66*, 758.

<sup>4</sup> Xu, Q.; Zhao, C.-Q.; Han, L.-B. *J. Am. Chem. Soc.* **2008**, *130*, 12648.

<sup>5</sup> Busacca, C. A.; Lorenz, J. C.; Grinberg, N.; Haddad, N.; Hrapchak, M.; Latil, B.; Lee, H.; Sabila, P.; Saha, A.; Sarvestani, M.; Chen, S.; Varsolona, R.; Wei, X.; Senanayake, C. H. *Org. Lett.* **2005**, *7*, 4277.

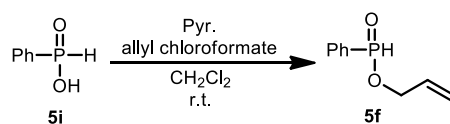
<sup>6</sup> Brown, H. C.; Bhat, N. G.; Srebnik, M. *Tetrahedron Lett.* **1988**, *29*, 2631.



propargylic CH<sub>2</sub>), 1.60-1.48 (m, 2H, CH<sub>2</sub>), 1.45-1.24 (m, 6H, CH<sub>2</sub>), 1.19 (d, 12H, *J* = 6.2 Hz, <sup>4</sup>Pr CH<sub>3</sub>), 0.89 (t, 3H, *J* = 6.9 Hz, hexyl CH<sub>3</sub>). The values of the <sup>1</sup>H NMR spectrum are in accordance with reported literature data.<sup>6</sup>

Following a slightly modified procedure,<sup>2</sup> 2-iodobenzoic acid (**10**) (692 mg, 2.79 mmol, 1.00 eq.), *para*-toluenesulfonic acid monohydrate (TsOH·H<sub>2</sub>O, 531 mg, 2.79 mmol, 1.00 eq.) and *meta*-chloroperoxybenzoic acid (*m*CPBA-70%, 756 mg, 3.07 mmol, 1.10 eq.) were dissolved in dichloromethane (4.5 mL) and 2,2,2-trifluoroethanol (4.5 mL). The mixture was stirred at room temperature under nitrogen for 1 hour, after which diisopropyloct-1-ynylboronate (**12**, 930 mg, 3.90 mmol, 1.40 eq.) was added in one portion. The reaction mixture was stirred for 2 hours at room temperature, filtered and concentrated *in vacuo*. The resulting oil was dissolved in dichloromethane (30 mL) and under vigorous stirring, saturated aq. NaHCO<sub>3</sub> (30 mL) was added. The mixture was stirred for 15 minutes, the two layers were separated and the aqueous layer was extracted with additional portions of dichloromethane (3 x 10 mL). The combined organic layers were dried over MgSO<sub>4</sub>, filtered and concentrated *in vacuo*. The crude product was purified by flash column chromatography (ethyl acetate) to afford **6d** (940 mg, 2.64 mmol, 95%) as a white solid. *R*<sub>f</sub> (EtOAc) = 0.25. <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) δ 8.42-8.35 (m, 1H, ArH), 8.20-8.13 (m, 1H, ArH), 7.78-7.69 (m, 2H, ArH), 2.59 (t, 2H, *J* = 7.1 Hz, CCCH<sub>2</sub>), 1.70-1.58 (m, 2H, CH<sub>2</sub>), 1.51-1.39 (m, 2H, CH<sub>2</sub>), 1.38-1.26 (m, 4H, CH<sub>2</sub>), 0.94-0.86 (m, 3H, CH<sub>2</sub>CH<sub>3</sub>). <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>) δ 166.7, 134.7, 132.5, 131.7, 131.6, 126.3, 115.7, 109.9, 39.4, 31.3, 28.7, 28.3, 22.6, 20.6, 14.1. The characterization data is in accordance with reported literature values.<sup>7</sup>

### Allyl phenylphosphinate (**5f**)

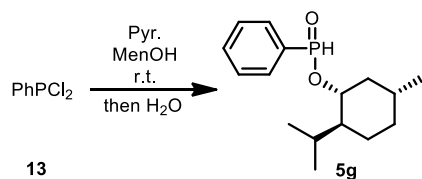


Product **5f** was made by using the known procedure reported by Virieux and co-workers.<sup>3</sup> Pyridine (3.0 mL, 36 mmol) was added to a stirring solution of phenylphosphinic acid (5.17 g, 36.4 mmol) and allyl chloroformate (4.0 mL, 36 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (70 mL) at room temperature. This reaction was stirred for 30 min. Once gas evolution was stopped, the resulting mixture was heated at reflux for 15 min, and cooled down to room temperature. 0.1 N aqueous solution (50 mL) was added to the resulting mixture, and stirred for 30 min. The organic layer was separated, washed with water (2 \* 50 mL), and dried over Na<sub>2</sub>SO<sub>4</sub>. The resulting solution was filtered and organic solvent was removed under reduced pressure. The remaining oil was distilled under vacuum to give pure **5f** (3.93 g, 21.6 mmol, 59%). <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) δ 7.90 – 7.85 (m, 2H, Ph H), 7.65 – 7.61 (m, 1H, Ph H), 7.65 (d, *J* = 400 Hz, 1H, PH), 7.56 – 7.52 (m, 2H, Ph H), 5.97 (ddtd, *J* = 17.0, 10.3, 5.6, 0.8 Hz, 1H, alkene CH), 5.46 – 5.35 (m, 1H), 5.28 (dt, *J* = 10.5, 1.1 Hz,

<sup>7</sup> Huang, H.; Zhang, G.; Gong, L.; Zhang, S.; Chen, Y. *J. Am. Chem. Soc.*, **2014**, *136*, 2280.

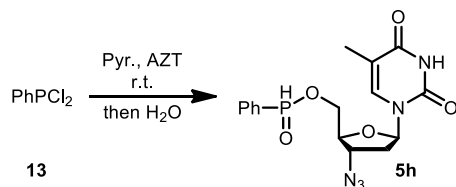
1H, alkene CH<sub>2</sub>), 4.60 (dddt, *J* = 16.1, 9.5, 5.5, 1.6 Hz, 2H, CH<sub>2</sub>O). The characterization data is in accordance with reported literature values.<sup>3</sup>

**(1*R*,2*S*,5*R*)-2-Isopropyl-5-methylcyclohexyl phenylphosphinate (5g)<sup>4</sup>**



Product **5g** was made by using the known procedure reported by Han and co-workers.<sup>4</sup> PhPCl<sub>2</sub> (**13**) (4.34 mL, 32.0 mmol) in dry Et<sub>2</sub>O (10 mL) was added to a mixture of pyridine (2.59 mL, 32.0 mmol) and *R*-menthol (5.0 g, 32 mmol) in dry Et<sub>2</sub>O (30 mL) at 0 °C, and the resulting solution was stirred at room temperature overnight. H<sub>2</sub>O (3 mL) was added, and the reaction mixture was washed with H<sub>2</sub>O and extracted with hexane (60 mL \* 3 times). The organic layer was separated, dried over MgSO<sub>4</sub>, and filtrated. The organic solvent was removed under reduced pressure to give a crude oil. The oil was distilled under vacuum to give the pure adduct **5g** (5.2 g, 19 mmol, 58%). The diastereomeric ratio is 1:1.6 calculated based on integration of peaks in <sup>31</sup>P-NMR. <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) δ 8.37 (d, *J* = 485 Hz, 1H, PH), 7.80 (ddt, *J* = 14.1, 8.2, 1.3 Hz, 2H, Ph H), 7.67 – 7.57 (m, 1H, Ph H), 7.56 – 7.47 (m, 2H, Ph H), 4.29 (td, *J* = 10.5, 4.5 Hz, 1H, CHO), 2.38 – 2.04 (m, 2H, CH or CH<sub>2</sub>), 1.79 – 1.62 (m, 2H, CH or CH<sub>2</sub>), 1.47 (ddt, *J* = 13.1, 6.7, 3.4 Hz, 2H, CH or CH<sub>2</sub>), 1.36 – 1.19 (m, 1H, CH or CH<sub>2</sub>), 1.04 – 0.78 (m, 11H, CH, CH<sub>2</sub> and CH<sub>3</sub>). <sup>31</sup>P-NMR (162 MHz, CDCl<sub>3</sub>) δ 21.3, 22.7. The characterization data is in accordance with reported literature values.<sup>4</sup>

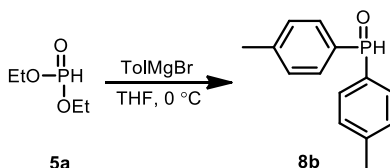
**((2*S*,3*S*,5*R*)-3-Azido-5-(5-methyl-2,4-dioxo-3,4-dihydropyrimidin-1(2*H*)-yl)tetrahydrofuran-2-yl)methyl phenylphosphinate (5h)**



Product **5h** was made by using the known procedure reported by Han and co-workers.<sup>4</sup> PhPCl<sub>2</sub> (**13**) (0.16 mL, 1.2 mmol) in dry Et<sub>2</sub>O (2 mL) was added to a mixture of pyridine (0.19 mL, 2.3 mmol) and AZT (0.30 g, 1.1 mmol) in dry Et<sub>2</sub>O (6 mL) at 0 °C, and the resulting solution was stirred at room temperature overnight. H<sub>2</sub>O (2 mL) was added, and the reaction mixture was washed with H<sub>2</sub>O (5 mL) and extracted with EtOAc (5 mL \* 5 times). The organic layer was separated, dried over MgSO<sub>4</sub>, and filtrated. The organic solvent was removed under reduced pressure to give a crude

gel. The crude gel was purified by column chromatography (MeOH/CH<sub>2</sub>Cl<sub>2</sub> 1/10) to give the product **5h** as a pale brown gel (0.20 g, 0.52 mmol, 47%, ca. 95% purity). **R<sub>f</sub>** 0.25 (MeOH/CH<sub>2</sub>Cl<sub>2</sub> 1/8, KMnO<sub>4</sub>). The diastereomeric ratio is 1:1 which was calculated based on integration of peaks in <sup>1</sup>H-NMR at 7.14 and 6.85 ppm. **<sup>1</sup>H-NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.78 (d, *J* = 490 Hz, 1H, PH), 7.67 (dd, *J* = 14.1, 8.3, 2.5, 1.4 Hz, 2H, Ph H), 7.52 – 7.45 (m, 1H, Ph H), 7.43 – 7.34 (m, 2H, Ph H), 7.14 (s, 0.5H, CH thymine, 1. diastereoisomer), 6.85 (s, 0.5H, CH thymine, 2. diastereoisomer), 6.08 (dt, *J* = 11.4, 6.5 Hz, 1H, CHON), 4.40 – 4.16 (m, 3H, CH<sub>2</sub>O and CHO), 3.94 (dq, *J* = 8.1, 3.7 Hz, 1H, CHN<sub>3</sub>), 2.35 – 2.17 (m, 2H, CH<sub>2</sub>), 1.70 (s, 1.5H, Me thymine, 1. diastereoisomer), 1.58 (s, 1.5H, Me thymine, 2. diastereoisomer). **<sup>13</sup>C-NMR** (101 MHz, CDCl<sub>3</sub>) δ 163.9, 150.3, 142.6, 136.8, 132.7 (d, *J* = 3.0 Hz), 132.1, 130.7 (d, *J* = 12.1 Hz), 128.6 (d, *J* = 13.9 Hz), 126.7, 111.2, 85.5 (d, *J* = 186.0 Hz), 60.9 (d, *J* = 183.5 Hz), 37.3, 12.5. **<sup>31</sup>P-NMR** (162 MHz, CDCl<sub>3</sub>) δ 22.2.<sup>8</sup> **IR** 3662(m), 2989(s), 2107(w), 1693(m), 1408(m), 1255(m), 1067(s), 894 (m).

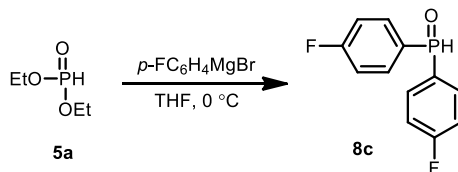
### Di-p-tolylphosphine oxide (**8b**)<sup>5</sup>



Product **8b** was made by using the known procedure reported by Busacca and co-workers.<sup>5</sup> A solution of diethylphosphite (**5a**) (1.5 mL, 12 mmol) in 10 mL THF was added dropwise to 1.0 M TolMgBr in THF (38.4 mL) at 0 °C. The resulting solution was stirred at 0 °C degree for 15 min and at room temperature for 30 min, and cooled down to 0 °C again before 0.1 N HCl aqueous solution (10 mL) was added. TBME (30 mL) was added to the resulting mixture and stirred for 5 min. The upper organic layer was decanted from the gel. CH<sub>2</sub>Cl<sub>2</sub> (35 mL) was subsequently added to the gel and stirred for 5 min. The resulting mixture was then filtered through a celite pad, washing the pad with CH<sub>2</sub>Cl<sub>2</sub> (35 mL). The organic layer was separated. All combined organic solutions were dried over MgSO<sub>4</sub> and the solvent was removed under reduced pressure to give a crude solid. The solid was recrystallized from boiling EtOAc (15 mL) to give the corresponding **8b** as a colorless solid (2.3 g, 9.8 mmol, 84%). **<sup>1</sup>H-NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.06 (d, *J* = 500 Hz, 1H, PH), 7.60 (ddd, *J* = 13.7, 8.1, 1.6 Hz, 4H, Tol CH), 7.35 – 7.29 (m, 4H, Tol CH), 2.43 (s, 6H, Me). The characterization data is *not* in accordance with reported literature values, as the tolyl Me peak was missing in the reported data.<sup>5</sup> The other peaks are corresponding.

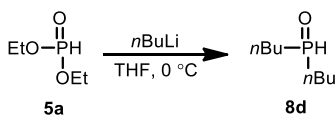
<sup>8</sup> The phosphorous signal of both diastereoisomers was overlapping.

### Bis(4-fluorophenyl)phosphine oxide (**8c**)<sup>4</sup>



Product **8c** was made by using the known procedure reported by Busacca and co-workers.<sup>5</sup> Diethylphosphite (**5a**) (1.0 mL, 7.7 mmol) in 8 mL THF was added dropwise to a solution of  $p\text{-FC}_6\text{H}_4\text{MgBr}$  in 10 mL THF (prepared from 0.64 g Mg (26 mmol) with 2.81 mL  $p\text{-FC}_6\text{H}_4\text{Br}$  (25.6 mmol)) at  $0\text{ }^\circ\text{C}$ . The resulting solution was stirred at  $0\text{ }^\circ\text{C}$  degree for 15 min and at room temperature for 30 min, and cooled down to  $0\text{ }^\circ\text{C}$  again before 0.1 N HCl aqueous solution (10 mL) was added. TBME (25 mL) was added to the resulting mixture and stirred for 5 min. The upper organic layer was decanted from the gel.  $\text{CH}_2\text{Cl}_2$  (25 mL) was subsequently added to the gel and stirred for 5 min. The resulting mixture was then filtered through a celite pad, washing the pad with  $\text{CH}_2\text{Cl}_2$  (25 mL). The organic layer was separated. All combined organic solutions were dried over  $\text{MgSO}_4$  and the solvent was removed under reduced pressure to give a crude oil. The crude oil was purified by column chromatography (pentane/EtOAc 2.5/1) to give the product **8c** as a colorless oil (1.4 g, 5.7 mmol, 73%). <sup>1</sup>H-NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.11 (d,  $J = 485$  Hz, 1H, PH), 7.72 (ddd,  $J = 13.1, 8.6, 5.5$  Hz, 4H, Ar CH), 7.28 – 7.19 (m, 4H, Ar CH). The characterization data is in accordance with reported literature values.<sup>5</sup>

### Dibutylphosphine oxide (**8d**)<sup>5</sup>



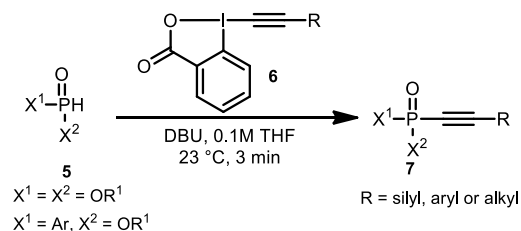
Product **8d** was made by using the known procedure reported by Busacca and co-workers.<sup>5</sup> A solution of diethylphosphite (**5a**) (3.0 mL, 23 mmol) in 10 mL THF was added dropwise to 2.5 M  $n\text{BuLi}$  in THF (30.7 mL, 77.0 mmol) at  $0\text{ }^\circ\text{C}$ . The resulting solution was stirred at  $0\text{ }^\circ\text{C}$  degree for 15 min and at room temperature for 30 min, and cooled down to  $0\text{ }^\circ\text{C}$  again before 0.1 N HCl aqueous solution (20 mL) was added. TBME (40 mL) was added to the resulting mixture and stirred for 5 min. The upper organic layer was decanted from the gel.  $\text{CH}_2\text{Cl}_2$  (50 mL) was subsequently added to the gel and stirred for 5 min. The resulting mixture was then filtered through a celite pad, washing the pad with  $\text{CH}_2\text{Cl}_2$  (50 mL). The organic layer was separated. All combined organic solutions were dried over  $\text{MgSO}_4$  and the solvent was removed under reduced pressure to give a crude gel. The gel was recrystallized from boiling hexane (15 mL) to give **8d** as a colorless solid (3.3 g, 20 mmol, 87%). <sup>1</sup>H-NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.59 – 6.19 (dm,  $J = 446$  Hz, 1H, PH), 1.92 – 1.51 (m, 8H,  $\text{CH}_2$ ), 1.51 – 1.37 (m, 4H,  $\text{CH}_2$ ), 0.93 (t,  $J = 7.3$  Hz, 6H,  $\text{CH}_3$ ). The characterization data is in accordance with reported literature values.<sup>5</sup>

### 3. Alkynylation Reaction

#### Optimization procedure:

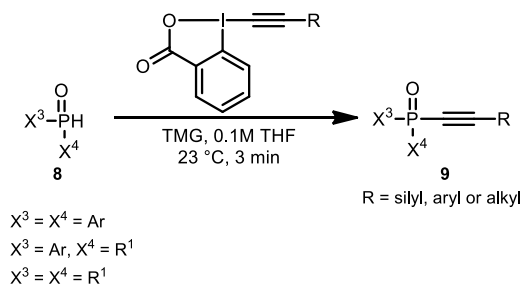
To a stirring solution of TIPS-EBX (**6a**) (1.1 – 1.2 equiv) and base (1.1 – 2.0 equiv) in solvent (0.1 M) was added diethyl phosphite **5a** (7.3 mg, 0.050 mmol, 1.0 equiv). The resulting solution was then stirred at r.t for 3 or 90 min. Subsequently 1M HCl (3 mL) and EtOAc (3 mL) were added to quench the resulting mixture. The organic layer was separated, and the aqueous layer was extracted with EtOAc (3 mL \* 3). All combined organic layers were washed with sat. NaHCO<sub>3</sub> (5 mL \* 2), dried over MgSO<sub>4</sub>, and removed under reduced pressure to give a crude adduct. Yield (%) based on <sup>1</sup>H-NMR with 1,3,5-trimethoxybenzene as internal reference. Pure product **7a** was further obtained by column chromatographic purification. *R<sub>f</sub>* 0.55 (pentane/EtOAc 1/1, KMnO<sub>4</sub>); pentane/EtOAc 1/1 was used as the eluting solvents for purification.

#### General procedure A:



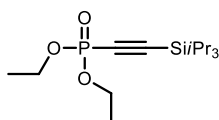
To a stirring solution of TIPS-, Ph-, *t*Bu- or *n*Hex-EBX **6** (1.1 equiv) and DBU (1.5 equiv) in THF (0.1 M) was added phosphite or phosphinate **5** (0.15–0.29 mmol, 1.0 equiv). The resulting solution was then stirred at r.t for 3 min. Subsequently 1M HCl (3 mL) and EtOAc (3 mL) were added to quench the resulting mixture. The organic layer was separated, and the aqueous layer was extracted with EtOAc (3 mL \* 3). All combined organic layers were washed with sat. NaHCO<sub>3</sub> (5 mL \* 2), dried over MgSO<sub>4</sub>, and removed under reduced pressure to give a crude adduct. The pure corresponding product was obtained by column chromatographic purification.

#### General procedure B:



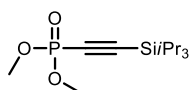
To a stirring solution of TIPS-, Ph-, *t*Bu- or *n*Hex-EBX **6** (1.5 equiv) and TMG (1.5 equiv) in THF (0.1 M) was added phosphine oxide **8** (0.15–0.29 mmol, 1.0 equiv). The resulting solution was then stirred at r.t for 3 min. Subsequently 1M HCl (3 mL) and EtOAc (3 mL) were added to quench the resulting mixture. The organic layer was separated, and the aqueous layer was extracted with EtOAc (3 mL \* 3). All combined organic layers were washed with sat. NaHCO<sub>3</sub> (5 mL \* 2), dried over MgSO<sub>4</sub>, and removed under reduced pressure to give a crude adduct. The pure corresponding product was obtained by column chromatographic purification.

### Diethyl ((triisopropylsilyl)ethynyl)phosphonate (**7a**)



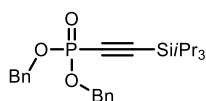
*H*-phosphite **5a** (45 mg, 0.32 mmol) and TIPS-EBX (**6a**) were used in general procedure A. **10a** was obtained as a colorless gel (91 mg, 0.28 mmol, 90% yield). **R<sub>f</sub>** 0.55 (pentane/EtOAc 1/1, KMnO<sub>4</sub>); pentane/EtOAc 1/1 was used as the eluting solvents for purification. **<sup>1</sup>H-NMR** (400 MHz, CDCl<sub>3</sub>) δ 4.15 (dd, *J* = 8.9, 7.1, 0.7 Hz, 4H, CH<sub>2</sub>O), 1.36 (td, *J* = 7.1, 0.7 Hz, 6H, ethyl CH<sub>3</sub>), 1.15 – 1.04 (m, 21H, TIPS). **<sup>13</sup>C-NMR** (101 MHz, CDCl<sub>3</sub>) δ 106.5 (d, *J* = 37.6 Hz), 96.4 (d, *J* = 269.7 Hz), 63.1 (d, *J* = 5.5 Hz), 17.7, 16.06 (d, *J* = 6.9 Hz), 10.8. **<sup>31</sup>P-NMR** (162 MHz, CDCl<sub>3</sub>) δ -8.5. **IR** 3669(s), 2998(s), 2361(s), 2180(w), 1408(s), 1252(s), 1079(s), 883(s), 819(m). **HRMS** (ESI) calcd for C<sub>15</sub>H<sub>32</sub>O<sub>3</sub>PSi<sup>+</sup> [M+H]<sup>+</sup> 319.1853, found 319.1857.

### Dimethyl ((triisopropylsilyl)ethynyl)phosphonate (**7b**)



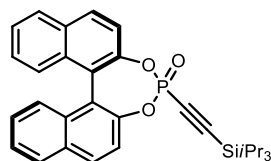
*H*-phosphite **5b** (35 mg, 0.32 mmol) and TIPS-EBX (**6a**) were used in general procedure A. **7b** was obtained as a colorless gel (79 mg, 0.27 mmol, 85% yield). **R<sub>f</sub>** 0.55 (pentane/EtOAc 1/1, KMnO<sub>4</sub>); pentane/EtOAc 1/1 was used as the eluting solvents for purification. **<sup>1</sup>H-NMR** (400 MHz, CDCl<sub>3</sub>) δ 4.12 (d, *J* = 8.9 Hz, 6H, Me), 1.45 – 1.17 (m, 21H, TIPS). **<sup>13</sup>C-NMR** (101 MHz, CDCl<sub>3</sub>) δ 107.9 (d, *J* = 37.9 Hz), 94.6 (d, *J* = 273.0 Hz), 53.4 (d, *J* = 5.6 Hz), 18.4, 10.8. **<sup>31</sup>P-NMR** (162 MHz, CDCl<sub>3</sub>) δ -5.2. **IR** 3674(s), 2989(s), 2361(s), 2180(w), 1251(s), 1081(s), 893(m), 822(m). **HRMS** (ESI) calcd for C<sub>13</sub>H<sub>28</sub>O<sub>3</sub>PSi<sup>+</sup> [M+H]<sup>+</sup> 291.1540, found 291.1541.

### Dibenzyl ((triisopropylsilyl)ethynyl)phosphonate (**7c**)



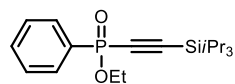
*H*-phosphite **5c** (88 mg, 0.32 mmol) and TIPS-EBX (**6a**) were used in general procedure A. **7c** was obtained as a colorless gel (126 mg, 0.283 mmol, 89% yield). **Rf** 0.7 (pentane/EtOAc 1/1, KMnO<sub>4</sub>); pentane/EtOAc 2/1 was used as the eluting solvents for purification. **<sup>1</sup>H-NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.49 – 7.30 (m, 10H, CH Ph), 5.14 (d, *J* = 8.5 Hz, 4H, Bn CH<sub>2</sub>), 1.25 – 0.97 (m, 21H, TIPS). **<sup>13</sup>C-NMR** (101 MHz, CDCl<sub>3</sub>) δ 135.5 (d, *J* = 7.5 Hz), 128.5, 128.5, 127.8, 107.8 (d, *J* = 38.3 Hz), 95.8 (d, *J* = 274.7 Hz), 68.5 (d, *J* = 5.2 Hz), 18.5, 10.8. **<sup>31</sup>P-NMR** (162 MHz, CDCl<sub>3</sub>) δ -8.0. **IR** 3674(s), 2972(s), 2902(s), 2361(s), 2180(w), 1933(w), 1454(m), 1394(s), 1251(s), 1056(s), 881(s), 785(w). **HRMS** (ESI) calcd for C<sub>25</sub>H<sub>36</sub>O<sub>3</sub>PSi<sup>+</sup> [M+H]<sup>+</sup> 443.2166, found 443.2172.

#### 4-((Triisopropylsilyl)ethynyl)dinaphtho[2,1-d:1',2'-f][1,3,2]dioxaphosphepine 4-oxide (**7d**)



*H*-phosphite **5d** (60 mg, 0.18 mmol) and TIPS-EBX (**6a**) were used in general procedure A. **7d** was obtained as a colorless gel (80 mg, 0.16 mmol, 86% yield). **Rf** 0.8 (pentane/EtOAc 1/1, KMnO<sub>4</sub>); pentane/EtOAc 3/1 was used as the eluting solvents for purification. **[α]<sub>D</sub><sup>23.0</sup>** -63.7 (c = 0.44, CHCl<sub>3</sub>). **<sup>1</sup>H-NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.09 – 7.92 (m, 4H, Ar H), 7.63 (dd, *J* = 8.8, 1.1 Hz, 1H, Ar H), 7.58 – 7.47 (m, 3H, Ar H), 7.46 – 7.29 (m, 4H, Ar H), 1.15 – 1.00 (m, 21H, TIPS). **<sup>13</sup>C-NMR** (101 MHz, CDCl<sub>3</sub>) δ 146.7 (d, *J* = 10.9 Hz), 145.4 (d, *J* = 9.1 Hz), 132.3 (d, *J* = 9.3 Hz), 131.9 (d, *J* = 4.9 Hz), 131.4, 130.9, 128.5 (d, *J* = 11.4 Hz), 127.2 (d, *J* = 28.1 Hz), 126.7 (d, *J* = 3.7 Hz), 125.9, 121.8 (dd, *J* = 13.6, 2.6 Hz), 120.9 (dd, *J* = 27.4, 3.0 Hz), 111.3 (d, *J* = 39.5 Hz), 93.2 (d, *J* = 294.2 Hz), 18.4, 10.8.<sup>9</sup> **<sup>31</sup>P-NMR** (162 MHz, CDCl<sub>3</sub>) δ -1.91. **IR** 3669(s), 2991(s), 2361(s), 2180(w), 1934(w), 1407(s), 1251(s), 1076(s), 893(s). **HRMS** (ESI) calcd for C<sub>31</sub>H<sub>34</sub>O<sub>3</sub>PSi<sup>+</sup> [M+H]<sup>+</sup> 513.2009, found 513.2003.

#### Ethyl phenyl((triisopropylsilyl)ethynyl)phosphinate (**7e**)

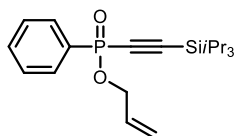


*H*-phosphinate **5e** (54 mg, 0.32 mmol) and TIPS-EBX (**6a**) were used in general procedure A. **7e** was obtained as a colorless gel (85 mg, 0.24 mmol, 76% yield). **Rf** 0.5 (pentane/EtOAc 1/1, KMnO<sub>4</sub>); pentane/EtOAc 2/1 was used as the eluting solvents for purification. **<sup>1</sup>H-NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.90 (ddd, *J* = 14.4, 7.5, 1.4 Hz, 2H, Ar H), 7.56 (dd, *J* = 7.5, 1.4 Hz, 1H, Ar H), 7.48 (d, *J* = 3.8 Hz, 2H, Ar H), 4.36 – 4.17 (m, 2H, ethyl CH<sub>2</sub>), 1.40 (t, *J* = 7.0 Hz, 3H, ethyl CH<sub>3</sub>), 1.21 – 0.98 (m, 21H, TIPS). **<sup>13</sup>C-NMR** (101 MHz, CDCl<sub>3</sub>) δ 132.6 (d, *J* = 3.1 Hz), 131.2 (d, *J* = 170.3 Hz), 130.9 (d, *J* = 11.3 Hz), 128.4 (d, *J* = 14.9 Hz), 109.2 (d, *J* = 26.4 Hz), 99.8 (d, *J* = 192.3 Hz), 62.3 (d, *J* = 6.6 Hz), 18.4, 16.3 (d, *J* = 6.9 Hz), 10.9. **<sup>31</sup>P-NMR** (162 MHz, CDCl<sub>3</sub>) δ 8.1. **IR** 3674(s), 3392(w), 3228(w), 2972(s), 2361(s), 2180(w), 1933(w), 1452(m), 1394(s),

<sup>9</sup> Not all aromatic signals were resolved.

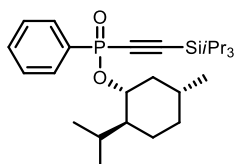
1250(s), 1053(s), 881(s), 801(m). **HRMS** (ESI) calcd for  $C_{19}H_{32}O_2PSi^+$   $[M+H]^+$  351.1904, found 351.1909.

### Allyl phenyl((triisopropylsilyl)ethynyl)phosphinate (**7f**)



*H*-phosphinate **5f** (58 mg, 0.32 mmol) and TIPS-EBX (**6a**) were used in general procedure A. **7f** was obtained as a colorless gel (100 mg, 0.285 mmol, 87% yield). **R<sub>f</sub>** 0.6 (pentane/EtOAc 1/1,  $KMnO_4$ ); pentane/EtOAc 1/1 was used as the eluting solvents for purification. **<sup>1</sup>H-NMR** (400 MHz,  $CDCl_3$ )  $\delta$  7.97 – 7.81 (m, 2H, Ph CH), 7.59 – 7.51 (m, 1H, Ph CH), 7.50 – 7.43 (m, 2H, Ph CH), 5.98 (ddt,  $J = 17.1, 10.7, 5.5$  Hz, 1H, alkene CH), 5.37 (dq,  $J = 17.1, 1.5$  Hz, 1H, alkene  $CH_2$ ), 5.23 (dq,  $J = 10.3, 1.3$  Hz, 1H, alkene  $CH_2$ ), 4.66 (ddt,  $J = 7.0, 4.1, 1.6$  Hz, 2H,  $CH_2O$ ), 1.19 – 0.98 (m, 21H, TIPS). **<sup>13</sup>C-NMR** (101 MHz,  $CDCl_3$ )  $\delta$  132.8 (d,  $J = 3.0$  Hz), 132.6 (d,  $J = 7.4$  Hz), 130.9 (d,  $J = 11.4$  Hz), 130.9 (d,  $J = 170.5$  Hz), 128.5 (d,  $J = 15.0$  Hz), 118.2, 109.7 (d,  $J = 26.8$  Hz), 99.5 (d,  $J = 193.5$  Hz), 66.4 (d,  $J = 6.2$  Hz), 18.4, 10.9. **<sup>31</sup>P-NMR** (162 MHz,  $CDCl_3$ )  $\delta$  8.6. **IR** 3676(s), 2964(s), 2361 (s), 2125(w), 1923(w), 1452(m), 1407(s), 1251(s), 1057(s), 881(s), 792(m). **HRMS** (ESI) calcd for  $C_{20}H_{32}O_2PSi^+$   $[M+H]^+$  362.1830, found 363.1904.

### (*1R,2S,5R*)-2-Isopropyl-5-methylcyclohexyl phenyl((triisopropylsilyl)ethynyl)phosphinate (**7g**)



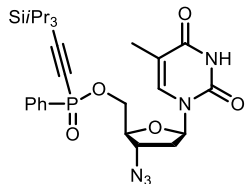
*H*-phosphinate **5g** (108 mg, 0.372 mmol) and TIPS-EBX (**6a**) were used in general procedure A. **7g** was obtained as a colorless gel (119 mg, 0.264 mmol, 71% yield). The diastereomeric ratio is 1:1, which was calculated based on integration of peaks in **<sup>31</sup>P-NMR**. **R<sub>f</sub>** 0.8 (pentane/EtOAc 1/1,  $KMnO_4$ ); pentane/EtOAc 2.5/1 was used as the eluting solvents for purification. **<sup>1</sup>H-NMR** (400 MHz,  $CDCl_3$ )  $\delta$  7.90 (dddd,  $J = 14.3, 8.4, 4.1, 1.4$  Hz, 2H, Ph CH), 7.59 – 7.52 (m, 1H, Ph CH), 7.48 (tdd,  $J = 6.8, 3.9, 1.4$  Hz, 2H, Ph CH), 4.46 (tdd,  $J = 10.8, 7.8, 4.4$  Hz, 1H, CHO), 2.62 – 2.34 (m, 1H, aliphatic H), 1.79 – 1.62 (m, 2H, aliphatic H), 1.36 – 1.21 (m, 2H, aliphatic H), 1.20 – 1.04 (m, 25H, aliphatic H and TIPS), 0.98 – 0.86 (m, 9H, aliphatic H and TIPS). **<sup>13</sup>C-NMR** (101 MHz,  $CDCl_3$ )  $\delta$  132.9 (d,  $J = 170.5$  Hz), 132.4, 132.4, 132.1, 130.9, 130.8, 130.6, 128.3 (d,  $J = 15.0$  Hz), 109.3 (d,  $J = 26.0$  Hz), 108.6 (d,  $J = 26.2$  Hz), 101.3 (d,  $J = 190.7$  Hz), 100.5 (d,  $J = 190.7$  Hz), 79.6 (d,  $J = 8.1$  Hz), 78.5 (d,  $J = 7.9$  Hz), 48.6, 48.6, 48.5, 43.7, 42.6, 34.1 (d,  $J = 2.8$  Hz), 31.7, 31.6, 25.8, 25.5, 23.1, 23.0, 21.9, 21.9, 21.0 (d,  $J = 2.0$  Hz), 18.5, 18.4, 17.7, 16.2, 15.6, 12.3, 11.0 (d,  $J = 1.9$  Hz).<sup>10</sup> **<sup>31</sup>P-NMR** (162 MHz,  $CDCl_3$ )  $\delta$  7.1, 5.1. **IR** 3674(s), 2988(s),

<sup>10</sup> Not all aliphatic signals were resolved.



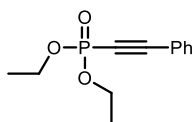
2361(s), 2180(w), 1453(m), 1407(s), 1250(s), 1079(s), 882(s), 787(m). **HRMS** (ESI) calcd for  $C_{27}H_{45}O_2PSi^+M+H$  461.2926, found 461.2998.

### 3'-Azido-3'-deoxythymidinyl phenyl((triisopropylsilyl)ethynyl)phosphinate (7h)



*H*-phosphinate **5h** (172 mg, 0.443 mmol) and TIPS-EBX (**6a**) were used in general procedure A. **7h** was obtained as a colorless gel (176 mg, 0.313 mmol, 70% yield). The diastereomeric ratio is 1:1.3, which was calculated based on average integration of peaks in  $^1H$ -NMR at 7.42 and 7.23 ppm, and  $^{31}P$ -NMR. **Rf** 0.45 (pentane/EtOAc 1/1,  $KMnO_4$ ); pentane/EtOAc 2.5/1 was used as the eluting solvents for purification.  $^1H$ -NMR (400 MHz,  $CDCl_3$ )  $\delta$  9.14 (s, 1H, NH), 7.90 (ddd,  $J = 14.5, 7.9, 6.4$  Hz, 2H, Ph CH), 7.64 (td,  $J = 7.4, 1.6$  Hz, 1H, Ph CH), 7.58 – 7.49 (m, 2H, Ph CH), 7.42 (s, 0.4H, CH thymine, 1. diastereoisomer), 7.23 (s, 0.6H), CH thymine, 2. diastereoisomer, 6.36 – 6.11 (m, 1H, CHON), 4.56 – 4.27 (m, 3H, CHO and  $CH_2O$ ), 4.12 (dd,  $J = 8.7, 3.4$  Hz, 1H,  $CHN_3$ ), 2.44 (dtd,  $J = 13.6, 6.4, 3.7$  Hz, 1H,  $CH_2$ ), 2.21 (ddt,  $J = 23.2, 14.1, 7.3$  Hz, 1H,  $CH_2$ ), 1.75 (s, 1.3H, Me thymine, 1. diastereoisomer), 1.64 (s, 1.7H, Me thymine, 2. diastereoisomer), 1.23 – 1.00 (m, 21H, TIPS).  $^{13}C$ -NMR ( $CDCl_3$ , 100MHz)  $\delta$  163.7, 163.6, 150.3, 150.2, 135.0, 134.8, 133.5, 133.5, 130.8 (d,  $J = 11.9$  Hz), 130.8 (d,  $J = 11.7$  Hz), 130.5, 129.2, 128.8 (d,  $J = 15.2$  Hz), 111.8 (d,  $J = 26.7$  Hz), 111.5, 111.4, 99.3 (d,  $J = 196.0$  Hz), 98.1 (d,  $J = 196.0$  Hz), 84.6, 84.6, 82.5 (d,  $J = 7.1$  Hz), 82.4 (d,  $J = 7.6$  Hz), 65.1, 65.0, 64.9, 60.8, 60.7, 37.7, 37.6, 18.4 (d,  $J = 1.9$  Hz), 12.3, 12.3, 10.9.  $^{31}P$ -NMR (162 MHz,  $CDCl_3$ )  $\delta$  10.4, 10.0. **IR** 3668(m), 2972(s), 2361(m), 2180(w), 1699(w), 1453(m), 1407(s), 1250(s), 1076(s), 882(m). **HRMS** (ESI) calcd for  $C_{19}H_{28}NSi^+$  (M+H) 572.2379, found 572.2570.

### Diethyl (phenylethynyl)phosphonate (7j)<sup>11</sup>

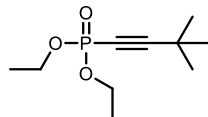


*H*-phosphite **5a** (26 mg, 0.18 mmol) and Ph-EBX (**6b**) were used in general procedure A. **7j** was obtained as a colorless gel (30 mg, 0.13 mmol, 69% yield). **Rf** 0.45 (pentane/EtOAc 1/1,  $KMnO_4$ ); pentane/EtOAc 1/1 was used as the eluting solvents for purification.  $^1H$ -NMR (400 MHz,  $CDCl_3$ )  $\delta$  7.60 – 7.52 (m, 2H, Ph CH), 7.50 – 7.32 (m, 3H, Ph CH), 4.37 – 4.13 (m, 4H, Et  $CH_2$ ), 1.40 (td,  $J = 7.1, 0.7$  Hz, 6H, Et  $CH_3$ ).  $^{13}C$ -NMR (101 MHz,  $CDCl_3$ )  $\delta$  132.6 (d,  $J = 2.5$  Hz), 130.7, 128.6, 119.6 (d,  $J = 5.6$  Hz), 99.1 (d,  $J = 52.9$  Hz), 79.8 (d, one peak mixing with  $CDCl_3$ ), 63.3 (d,  $J = 5.5$  Hz), 16.2 (d,  $J = 7.0$  Hz).  $^{31}P$ -NMR (162 MHz,  $CDCl_3$ )  $\delta$  -6.0. **IR**

<sup>11</sup> Wang, Y.; Gan, J.; Liu, L.; Yuan, H.; Gao, Y.; Liu Y.; Zhao, Y. *J. Org. Chem.* **2014**, *79*, 3678.

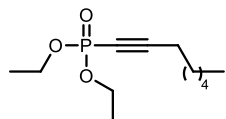
3674(s), 3344(m), 2973(s), 2361(s), 2180(w), 1452(s), 1374(s), 1251(s), 1051(s), 881(m). The characterization data is in accordance with reported literature values.<sup>11</sup>

### Diethyl (3,3-dimethylbut-1-yn-1-yl)phosphonate (7k)



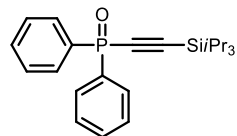
*H*-phosphite **5a** (46 mg, 0.32 mmol) and *t*Bu-EBX (**6c**) were used in general procedure A. **7k** was obtained as a colorless gel (58 mg, 0.27 mmol, 80% yield). **R<sub>f</sub>** 0.55 (pentane/EtOAc 1/1, KMnO<sub>4</sub>); pentane/EtOAc 1/1 was used as the eluting solvents for purification. **<sup>1</sup>H-NMR** (400 MHz, CDCl<sub>3</sub>) δ 4.17 – 4.02 (m, 4H, Et CH<sub>2</sub>), 1.33 (td, *J* = 7.1, 0.8 Hz, 6H, Et CH<sub>3</sub>), 1.25 (s, 9H, <sup>t</sup>Bu). **<sup>13</sup>C-NMR** (101 MHz, CDCl<sub>3</sub>) δ 110.2 (d, *J* = 50.8 Hz), 68.7 (d, *J* = 301.8 Hz), 62.8 (d, *J* = 5.5 Hz), 29.8, 27.9, 16.0. **<sup>31</sup>P-NMR** (162 MHz, CDCl<sub>3</sub>) δ -5.7. **IR** 3676(s), 2988(s), 2361(s), 2180(w), 1407(m), 1394(s), 1251(s), 1083(s), 893(m), 798(m). **HRMS** (ESI) calcd for C<sub>10</sub>H<sub>19</sub>O<sub>3</sub>P<sup>+</sup> (M+H) 219.1071, found 219.1150.

### Diethyl oct-1-yn-1-ylphosphonate (7l)



*H*-phosphite **5a** (23 mg, 0.16 mmol) and *n*Hex-EBX (**6d**) were used in general procedure A. **7l** was obtained as a colorless gel (34.5 mg, 0.14 mmol, 86% yield). **R<sub>f</sub>** 0.3 (pentane/EtOAc 1/2, KMnO<sub>4</sub>); pentane/EtOAc 1/1 was used as the eluting solvents for purification. **<sup>1</sup>H-NMR** (400 MHz, CDCl<sub>3</sub>) δ 4.29 – 3.99 (m, 4H, Et CH<sub>2</sub>), 2.33 (td, *J* = 7.2, 4.4 Hz, 2H, propargyl CH<sub>2</sub>), 1.66 – 1.49 (m, 2H, CH<sub>2</sub>), 1.47 – 1.15 (m, 12H, CH<sub>2</sub> and ethyl CH<sub>3</sub>), 0.88 (dd, *J* = 7.5, 6.1 Hz, 3H, Me). **<sup>13</sup>C-NMR** (101 MHz, CDCl<sub>3</sub>) δ 103.2 (d, *J* = 52.9 Hz), 70.4 (d, *J* = 303.2 Hz), 62.9 (d, *J* = 5.5 Hz), 31.1, 28.4, 27.4 (d, *J* = 2.2 Hz), 22.4, 19.2 (d, *J* = 4.5 Hz), 16.1 (d, *J* = 7.1 Hz), 14.0. **<sup>31</sup>P-NMR** (162 MHz, CDCl<sub>3</sub>) δ -6.1. **IR** 3662(s), 2988(w), 2207(w), 1394(m), 1251(m), 1057(s), 893(m). **HRMS** (ESI) calcd for C<sub>12</sub>H<sub>24</sub>O<sub>3</sub>P<sup>+</sup> (M+H) 247.1458, found 247.1470.

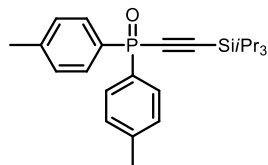
### Diphenyl((triisopropylsilyl)ethynyl)phosphine oxide (9a)



*H*-phosphine oxide **8a** (48.7 mg, 0.233 mmol) and TIPS-EBX (**6a**) were used in general procedure B. **9a** was obtained as a colorless gel (81 mg, 0.21 mmol, 91% yield). **R<sub>f</sub>** 0.5 (pentane/EtOAc 1/1, KMnO<sub>4</sub>); pentane/EtOAc 1/1 was used as the eluting solvents for

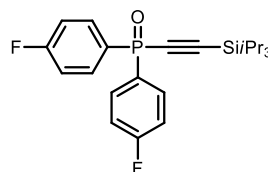
purification. **<sup>1</sup>H-NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.85 (ddd, *J* = 13.8, 8.3, 1.5 Hz, 4H, Ph CH), 7.55 – 7.48 (m, 2H, Ph CH), 7.48 – 7.41 (m, 4H, Ph CH), 1.21 – 1.00 (m, 21H, TIPS). **<sup>13</sup>C-NMR** (101 MHz, CDCl<sub>3</sub>) δ 133.2 (d, *J* = 120.9 Hz), 132.1 (d, *J* = 2.9 Hz), 130.9 (d, *J* = 11.2 Hz), 128.5 (d, *J* = 13.5 Hz), 113.4 (d, *J* = 18.8 Hz), 101.1 (d, *J* = 151.5 Hz), 18.5, 11.0. **<sup>31</sup>P-NMR** (162 MHz, CDCl<sub>3</sub>) δ 6.9. **IR** 3674(s), 2991(s), 2361(s), 2121(w), 1934(w), 1407(s), 1251(s), 1081(s), 893(s), 791(m). **HRMS** (ESI) calcd for C<sub>23</sub>H<sub>32</sub>OPSi<sup>+</sup> [M+H]<sup>+</sup> 383.1955, found 383.1957.

### Di-*p*-tolyl((triisopropylsilyl)ethynyl)phosphine oxide (**9b**)



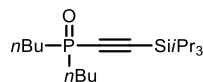
*H*-phosphine oxide **8b** (53.7 mg, 0.233 mmol) and TIPS-EBX (**6a**) were used in general procedure B. **9b** was obtained as a colorless gel (85 mg, 0.21 mmol, 89% yield). **R<sub>f</sub>** 0.45 (pentane/EtOAc 1/1, KMnO<sub>4</sub>); pentane/EtOAc 1/1 was used as the eluting solvents for purification. **<sup>1</sup>H-NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.72 (dd, *J* = 13.6, 8.1 Hz, 4H, Ar CH), 7.24 (dd, *J* = 8.1, 3.0 Hz, 4H, Ar CH), 2.36 (s, 6H, Me), 1.22 – 1.01 (m, 21H, TIPS). **<sup>13</sup>C-NMR** (101 MHz, CDCl<sub>3</sub>) δ 142.5 (d, *J* = 3.0 Hz), 130.8 (d, *J* = 11.6 Hz), 129.5, 129.2 (d, *J* = 13.8 Hz), 112.4 (d, *J* = 18.8 Hz), 101.5 (d, *J* = 150.4 Hz), 21.5, 18.4, 11.0. **<sup>31</sup>P-NMR** (162 MHz, CDCl<sub>3</sub>) δ 7.2. **IR** 3674(s), 3319(w), 3227(w), 2972(s), 2361(s), 2190(w), 1933(w), 1452(m), 1407(s), 1251(s), 1056(s), 893(s), 803(w). **HRMS** (ESI) calcd for C<sub>25</sub>H<sub>36</sub>OPSi<sup>+</sup> [M+H]<sup>+</sup> 411.2268, found 411.2267.

### Bis(4-fluorophenyl)((triisopropylsilyl)ethynyl)phosphine oxide (**9c**)



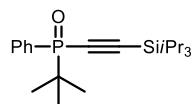
*H*-phosphine oxide **8c** (45 mg, 0.18 mmol) and TIPS-EBX (**9a**) were used in general procedure B. **9c** was obtained as a colorless gel (70 mg, 0.17 mmol, 93% yield). **R<sub>f</sub>** 0.5 (pentane/EtOAc 1/1, KMnO<sub>4</sub>); pentane/EtOAc 1/1 was used as the eluting solvents for purification. **<sup>1</sup>H-NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.90 – 7.76 (m, 4H, Ar CH), 7.15 (td, *J* = 8.7, 2.3 Hz, 4H, Ar CH), 1.20 – 1.03 (m, 21H, TIPS). **<sup>13</sup>C-NMR** (101 MHz, CDCl<sub>3</sub>) δ 165.3 (dd, *J* = 254.0, 3.5 Hz), 133.4 (dd, *J* = 12.9, 8.9 Hz), 129.0 (dd, *J* = 125.1, 3.4 Hz), 116.1 (dd, *J* = 21.6, 14.8 Hz), 114.3 (d, *J* = 19.6 Hz), 100.7 (d, *J* = 154.4 Hz), 18.48, 10.99. **<sup>31</sup>P-NMR** (162 MHz, CDCl<sub>3</sub>) δ 4.6. **<sup>19</sup>F-NMR** (376 MHz, CDCl<sub>3</sub>) δ -106.0. **IR** 3674(s), 3391(m), 3226(m), 2958(s), 2361 (s), 2125(w), 1933(w), 1082(s), 893(s), 802(w). **HRMS** (ESI) calcd for C<sub>23</sub>F<sub>2</sub>H<sub>30</sub>OPSi<sup>+</sup> [M+H]<sup>+</sup> 419.1766, found 419.1767.

### Dibutyl((triisopropylsilyl)ethynyl)phosphine oxide (**9d**)



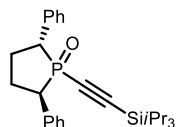
*H*-phosphine oxide **8d** (39 mg, 0.23 mmol) and TIPS-EBX (**6a**) were used in general procedure B. **9d** was obtained as a colorless liquid (69 mg, 0.20 mmol, 86% yield, 95% purity). **R<sub>f</sub>** 0.5 (MeOH/CH<sub>2</sub>Cl<sub>2</sub> 1/10, KMnO<sub>4</sub>); MeOH/CH<sub>2</sub>Cl<sub>2</sub> 3/100 was used as the eluting solvents for purification. **<sup>1</sup>H-NMR** (400 MHz, CDCl<sub>3</sub>) δ 1.95 – 1.76 (m, 4H, Bu CH<sub>2</sub>), 1.75 – 1.57 (m, 4H, Bu CH<sub>2</sub>), 1.46 (h, *J* = 7.3 Hz, 4H, Bu CH<sub>2</sub>), 1.16 – 1.02 (m, 21H, TIPS), 0.93 (t, *J* = 7.3 Hz, 6H, Me). **<sup>13</sup>C-NMR** (101 MHz, CDCl<sub>3</sub>) δ 109.3 (d, *J* = 15.6 Hz), 31.2 (d, *J* = 78.9 Hz), 24.2 (d, *J* = 3.7 Hz), 23.9 (d, *J* = 15.6 Hz), 18.4, 13.6, 10.9.<sup>12</sup> **<sup>31</sup>P-NMR** (162 MHz, CDCl<sub>3</sub>) δ 25.3. **IR** 3675(m), 3385(m), 2972(sh), 2902(s), 2361(s), 2180(w), 1921(w), 1453(m), 1394(s), 1251(s), 1051(sh), 881(s), 8020(w). **HRMS** (ESI) calcd for C<sub>19</sub>H<sub>40</sub>OPSi<sup>+</sup> [M+H]<sup>+</sup> 343.2581, found 343.2584.

### *tert*-Butyl(phenyl)((triisopropylsilyl)ethynyl)phosphine oxide (**9e**)



*H*-phosphine oxide **8e** (45 mg, 0.25 mmol) and TIPS-EBX (**6a**) were used in general procedure B. **9e** was obtained as a colorless gel (78 mg, 0.22 mmol, 87% yield). **R<sub>f</sub>** 0.45 (pentane/EtOAc 2/1, KMnO<sub>4</sub>); pentane/EtOAc 2/1 was used as the eluting solvents for purification. **<sup>1</sup>H-NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.86 (ddd, *J* = 11.9, 8.3, 1.4 Hz, 2H, Ph CH), 7.56 – 7.50 (m, 1H, Ph CH), 7.45 (tdd, *J* = 7.0, 3.1, 1.6 Hz, 2H, Ph CH), 1.27 – 1.04 (m, 30H, TIPS and <sup>t</sup>Bu). **<sup>13</sup>C-NMR** (101 MHz, CDCl<sub>3</sub>) δ 132.0, 131.9, 129.4 (d, *J* = 108.0 Hz), 128.0 (d, *J* = 12.2 Hz), 111.5 (d, *J* = 14.1 Hz), 99.5 (d, *J* = 132.5 Hz), 33.7 (d, *J* = 82.4 Hz), 23.7, 18.5, 11.0. **<sup>31</sup>P-NMR** (162 MHz, CDCl<sub>3</sub>) δ 29.8. **IR** 3674(s), 3390(w), 3226(w), 2988(s), 2361(s), 2195(w), 1933(w), 1452(m), 1394(s), 1251(s), 1056(s), 880(s). **HRMS** (ESI) calcd for C<sub>21</sub>H<sub>36</sub>OPSi<sup>+</sup> [M+H]<sup>+</sup> 363.2268, found 363.2268.

### (2*R*,5*R*)-2,5-Diphenyl-1-((triisopropylsilyl)ethynyl)phospholane 1-oxide (**9f**)

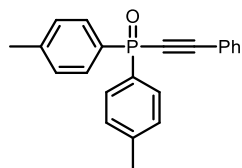


*H*-phosphine oxide **8f** (45 mg, 0.18 mmol) and TIPS-EBX (**6a**) were used in general procedure B. **9f** was obtained as a colorless gel (68 mg, 0.15 mmol, 89% yield). **R<sub>f</sub>** 0.6 (pentane/EtOAc 1/1,

<sup>12</sup> One of ethynyl carbons is not strong enough to be recorded.

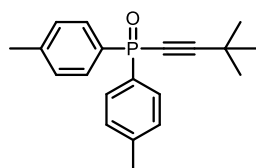
KMnO<sub>4</sub>); pentane/EtOAc 2/1 was used as the eluting solvents for purification.  $[\alpha]_D^{23.0} +2.2$  (c = 0.2, CHCl<sub>3</sub>). **<sup>1</sup>H-NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.36 – 7.13 (m, 10H, Ph CH), 3.57 (ddd, *J* = 24.7, 12.5, 7.1 Hz, 1H, CHPh), 3.35 (td, *J* = 12.2, 7.6 Hz, 1H, CHPh), 2.63 – 2.19 (m, 3H, CH<sub>2</sub>), 2.16 – 1.99 (m, 1H, CH<sub>2</sub>), 0.88 (m, 21H, TIPS). **<sup>13</sup>C-NMR** (101 MHz, CDCl<sub>3</sub>) δ 135.5 (dd, *J* = 43.1, 5.7 Hz), 128.8 (d, *J* = 5.5 Hz), 128.6 (dd, *J* = 10.5, 2.3 Hz), 127.7 (d, *J* = 5.4 Hz), 127.0 (dd, *J* = 21.4, 2.8 Hz), 112.8 (d, *J* = 13.5 Hz), 100.0 (d, *J* = 132.1 Hz), 50.5 (d, *J* = 33.4 Hz), 49.8 (d, *J* = 33.4 Hz), 31.1 (d, *J* = 9.5 Hz), 27.3 (d, *J* = 11.3 Hz), 18.3, 10.7. **<sup>31</sup>P-NMR** (162 MHz, CDCl<sub>3</sub>) δ 36.3. **IR** 3671(m), 2987(s), 2361(m), 2180(w), 1407(s), 1251(s), 1059(s), 893(m). **HRMS** (ESI) calcd for C<sub>27</sub>H<sub>38</sub>OPSi<sup>+</sup> [M+H]<sup>+</sup> 437.2424, found 437.2425.

### (Phenylethynyl)di-*p*-tolylphosphine oxide (**9i**)



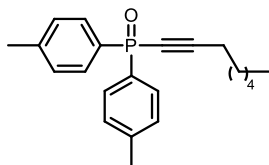
*H*-phosphine oxide **8b** (31 mg, 0.13 mmol) and Ph-EBX (**6b**) were used in general procedure B. **9i** was obtained as a colorless solid (32 mg, 0.10 mmol, 72% yield). **R<sub>f</sub>** 0.5 (pentane/EtOAc 1/1, KMnO<sub>4</sub>); pentane/EtOAc 1/1 was used as the eluted solvents for purification. **Mp** 78 °C; **<sup>1</sup>H-NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.78 (dd, *J* = 13.7, 8.1 Hz, 4H, Ar CH), 7.62 – 7.56 (m, 2H, Ar CH), 7.47 – 7.41 (m, 1H, Ar CH), 7.37 (tt, *J* = 6.7, 1.6 Hz, 2H, Ar CH), 7.29 (dd, *J* = 8.1, 3.0 Hz, 4H, Ar CH), 2.40 (s, 6H, Me). **<sup>13</sup>C-NMR** (101 MHz, CDCl<sub>3</sub>) δ 142.8 (d, *J* = 3.0 Hz), 132.5, 132.5, 131.0 (d, *J* = 11.7 Hz), 130.5, 129.4 (d, *J* = 13.9 Hz), 128.6, 120.2 (d, *J* = 4.1 Hz), 105.0 (d, *J* = 29.8 Hz), 83.3 (d, *J* = 169.0 Hz), 21.7. **<sup>31</sup>P-NMR** (162 MHz, CDCl<sub>3</sub>) δ 9.2. **IR** ν 3675(s), 3344(s), 2973(s), 2891(s), 2361(s), 2180(w), 1407(s), 1251(s), 1072(s), 894(s). **HRMS** (ESI) calcd for C<sub>22</sub>H<sub>20</sub>OP<sup>+</sup> [M+H]<sup>+</sup> 331.1246, found 331.1245.

### 3,3-Dimethylbut-1-yn-1-yl)di-*p*-tolylphosphine oxide (**9j**)



*H*-phosphine oxide **8b** (47 mg, 0.20 mmol) and *t*Bu-EBX (**6c**) were used in general procedure B. **9j** was obtained as a colorless gel (55 mg, 0.17 mmol, 87% yield). **R<sub>f</sub>** 0.4 (pentane/EtOAc 1/1, KMnO<sub>4</sub>); pentane/EtOAc 1/1 was used as the eluting solvents for purification. **<sup>1</sup>H-NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.68 (dd, *J* = 13.6, 8.0 Hz, 4H, Ar CH), 7.25 (dd, *J* = 8.0, 2.8 Hz, 4H, Ar CH), 2.38 (s, 6H, Ar Me), 1.31 (s, 9H, *t*Bu). **<sup>13</sup>C-NMR** (101 MHz, CDCl<sub>3</sub>) δ 142.3, 130.8 (d, *J* = 11.6 Hz), 130.6 (d, *J* = 124.1 Hz), 129.2 (d, *J* = 13.8 Hz), 116.1 (d, *J* = 28.8 Hz), 73.4 (d, *J* = 173.7 Hz), 30.0, 28.4, 21.6. **<sup>31</sup>P-NMR** (162 MHz, CDCl<sub>3</sub>) δ 7.9. **IR** 3674(s), 2988(s), 2361(s), 2177(w), 1933(w), 1452(m), 1407(s), 1252(s), 1230(s), 1066(s), 893(s), 778(w). **HRMS** (ESI) calcd for C<sub>20</sub>H<sub>24</sub>OP<sup>+</sup> (M+H) 311.1486, found 311.1565.

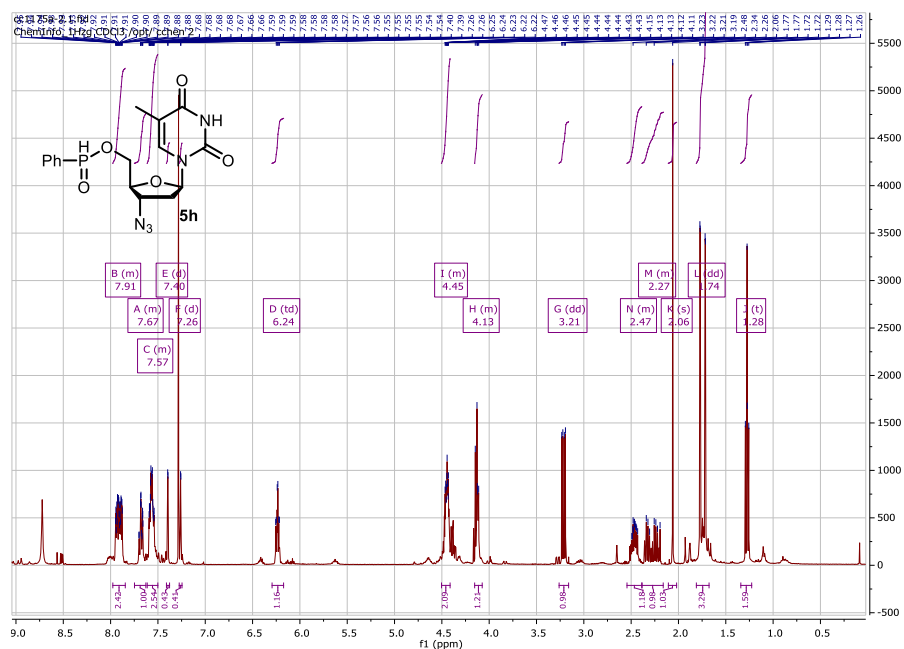
### Oct-1-yn-1-yl-di-p-tolylphosphine oxide (**9k**)



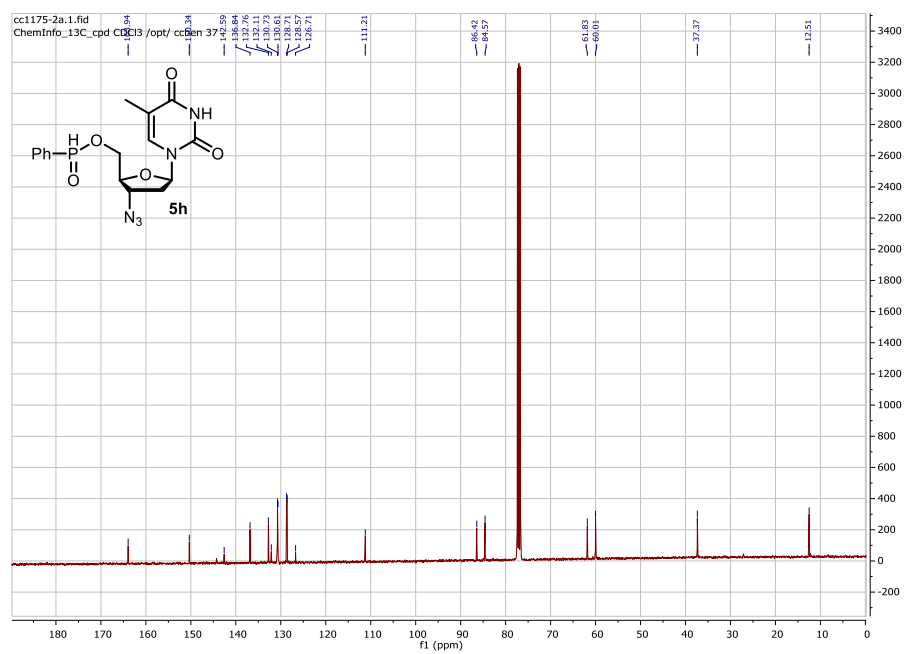
*H*-phosphine oxide **8b** (37.7 mg, 0.163 mmol) and *n*Hex-EBX (**6d**) were used in general procedure B. **9k** was obtained as a colorless gel (47 mg, 0.14 mmol, 85% yield). **R<sub>f</sub>** 0.35 (pentane/EtOAc 1/1, KMnO<sub>4</sub>); pentane/EtOAc 1/1 was used as the eluting solvents for purification. **<sup>1</sup>H-NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.72 (dd, *J* = 13.6, 8.1 Hz, 4H, Ar H), 7.33 – 7.21 (m, 4H, Ar H), 2.50 – 2.35 (m, 8H, 2\*CH<sub>3</sub>C<sub>6</sub>H<sub>4</sub> + CH<sub>2</sub>), 1.70 – 1.56 (m, 2H, CH<sub>2</sub>), 1.50 – 1.38 (m, 2H, CH<sub>2</sub>), 1.36 – 1.21 (m, 4H, CH<sub>2</sub>), 0.96 – 0.83 (m, 3H, Me). **<sup>13</sup>C-NMR** (101 MHz, CDCl<sub>3</sub>) δ 142.5 (d, *J* = 3.0 Hz), 130.9 (d, *J* = 11.6 Hz), 129.7, 129.2 (d, *J* = 13.8 Hz), 109.3 (d, *J* = 30.4 Hz), 75.2 (d, *J* = 174.4 Hz), 31.2, 28.5, 27.6 (d, *J* = 1.8 Hz), 22.5, 21.6, 19.8 (d, *J* = 3.2 Hz), 14.0. **<sup>31</sup>P-NMR** (162 MHz, CDCl<sub>3</sub>) δ 8.1. **IR** 3662(s), 2988(s), 2193(s), 1394(m), 1407(s), 1252(s), 1057(s), 893(s), 809(w). **HRMS** (ESI) calcd for C<sub>22</sub>H<sub>28</sub>OP<sup>+</sup> (M+H) 339.1872, found 339.1888

### 4. Spectra of New Compounds

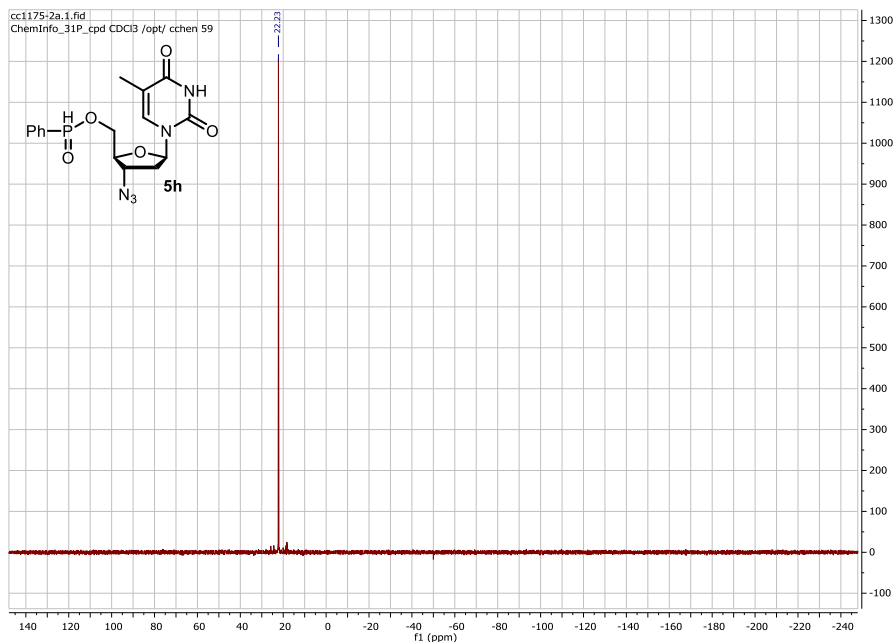
### <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) of compound 5h



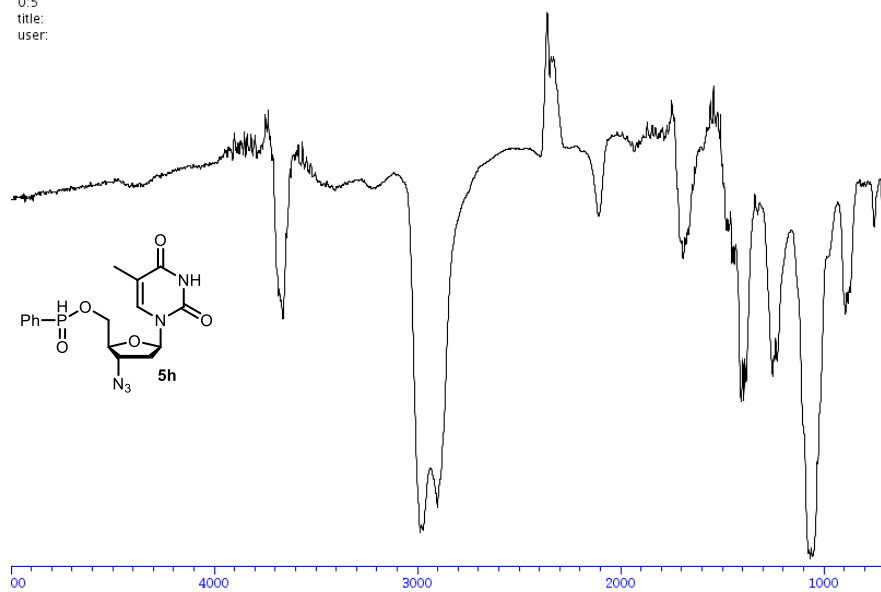
### <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>) of compound 5h



# <sup>31</sup>P-NMR (162 MHz, CDCl<sub>3</sub>) of compound **5h**

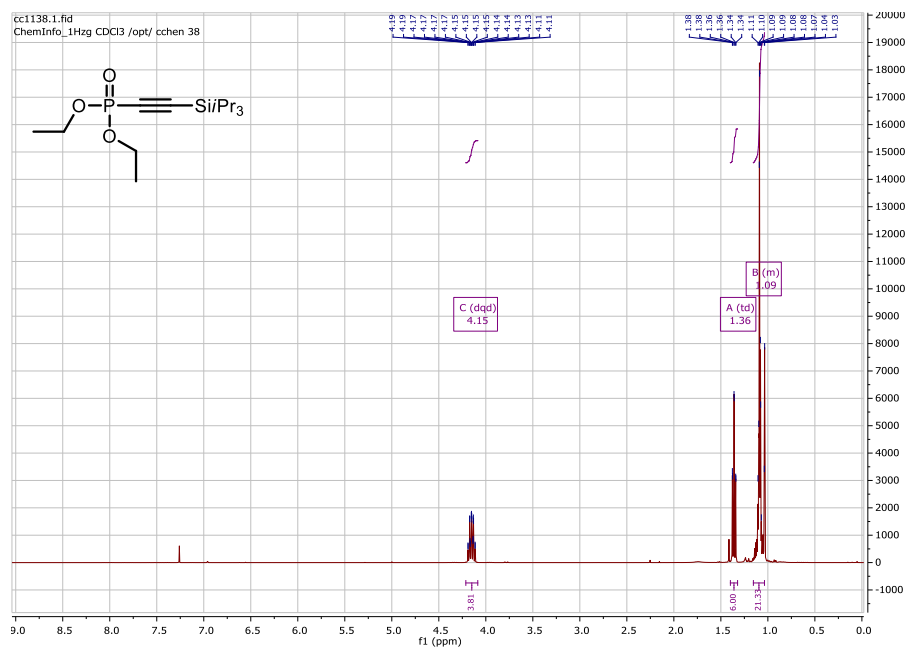


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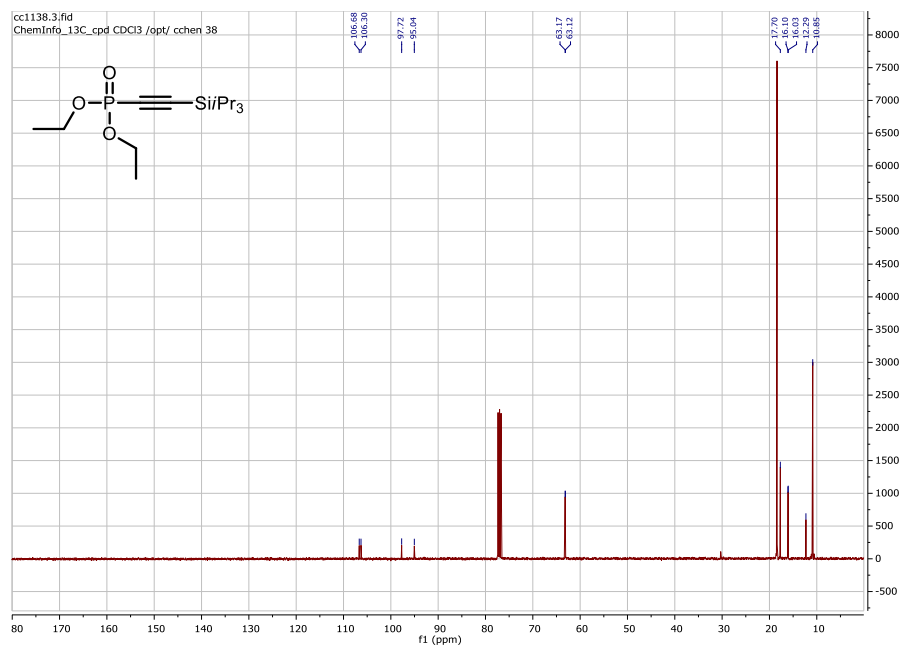




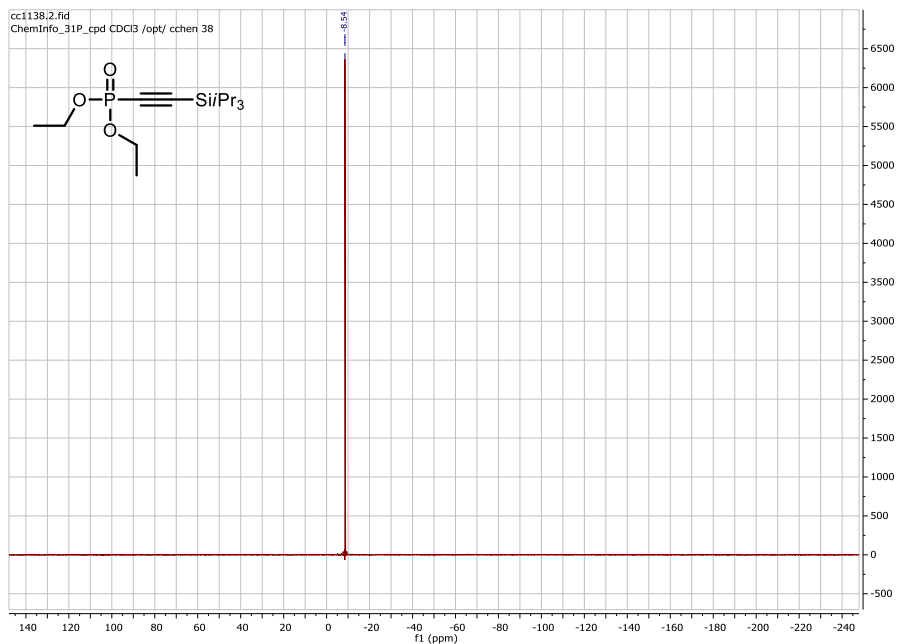
# <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) of compound 7a



# <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>) of compound 7a



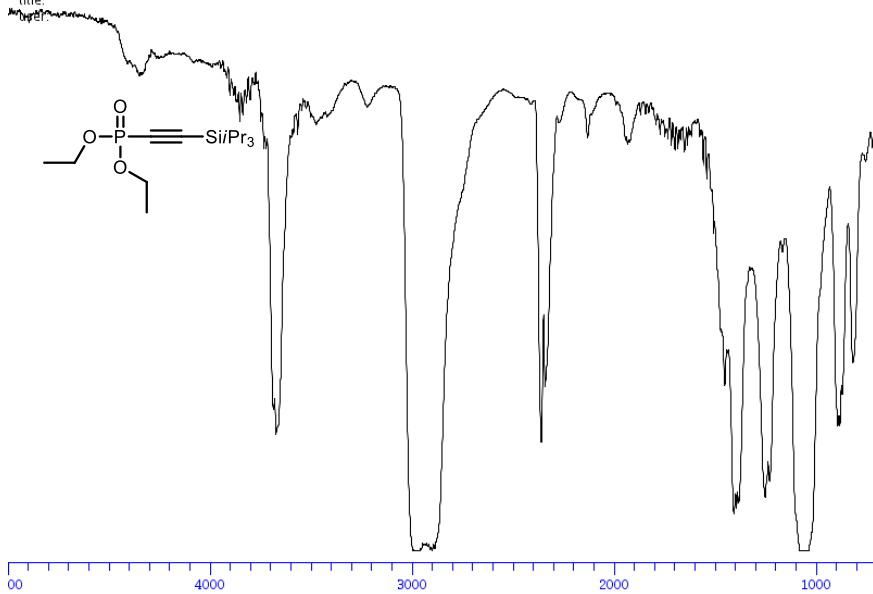
### <sup>31</sup>P-NMR (162 MHz, CDCl<sub>3</sub>) of compound **7a**



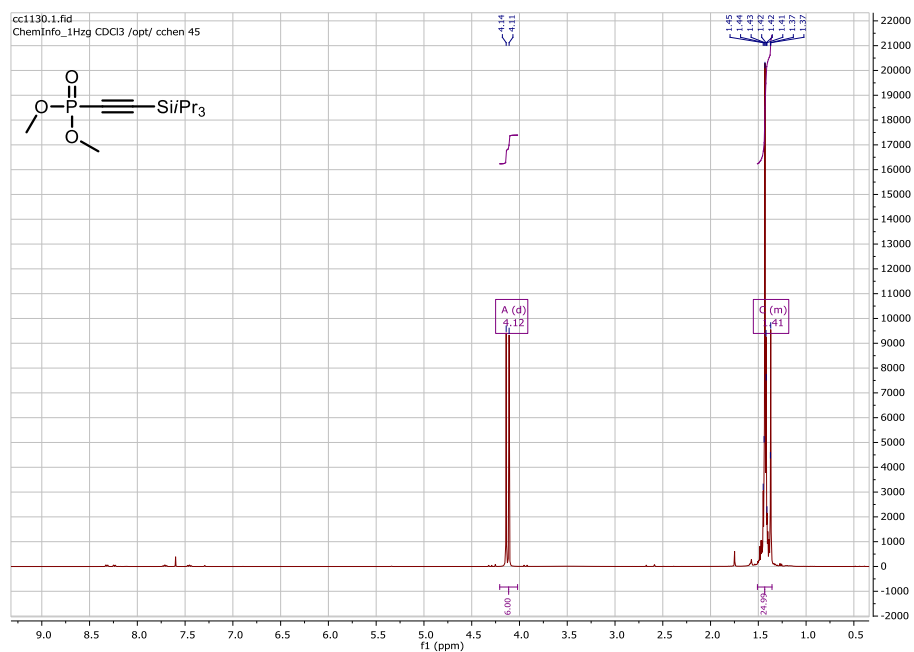
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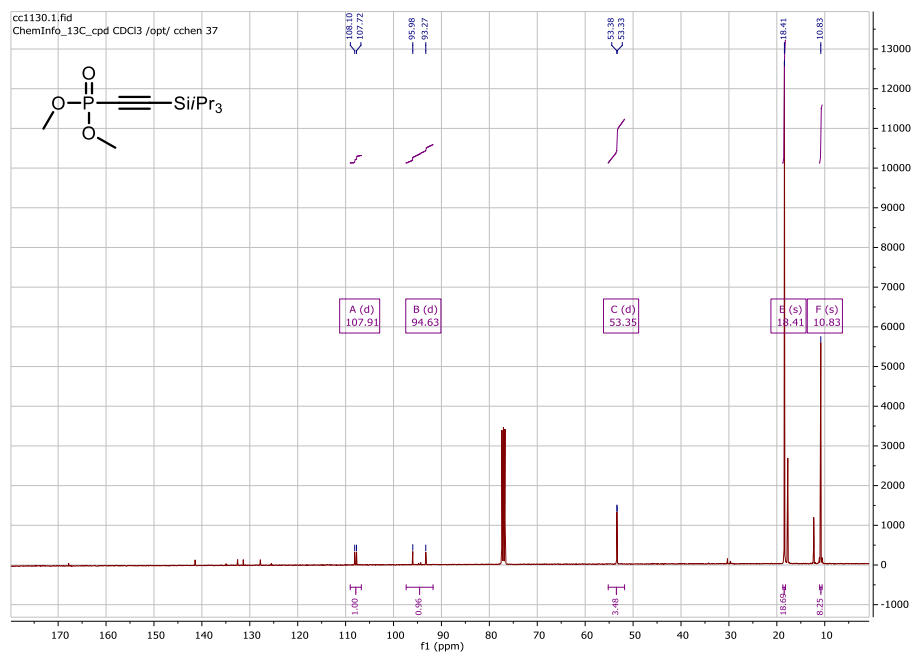
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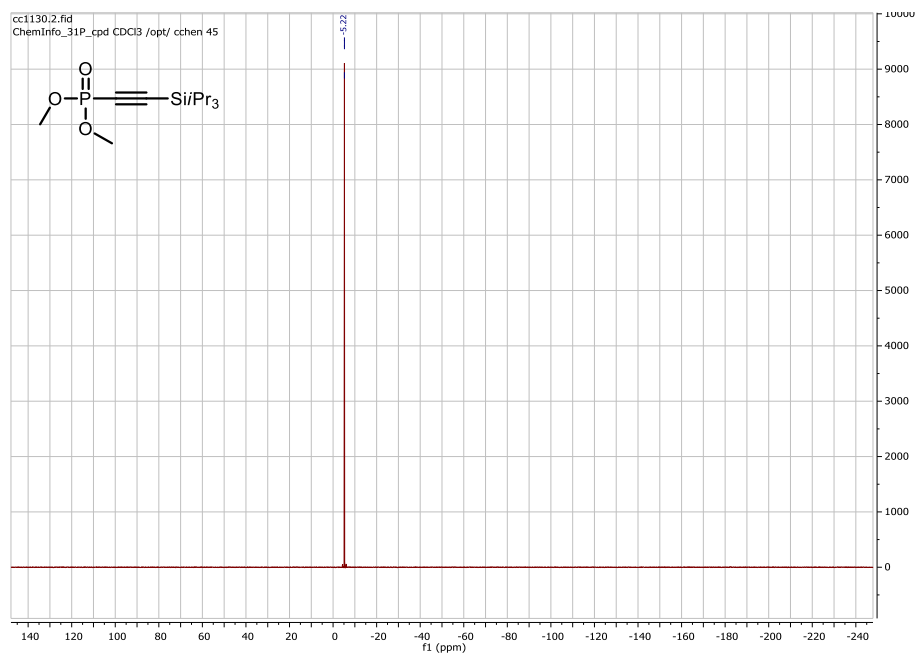
# <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) of compound **7b**



# <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>) of compound **7b**



# <sup>31</sup>P-NMR (162 MHz, CDCl<sub>3</sub>) of compound **7b**

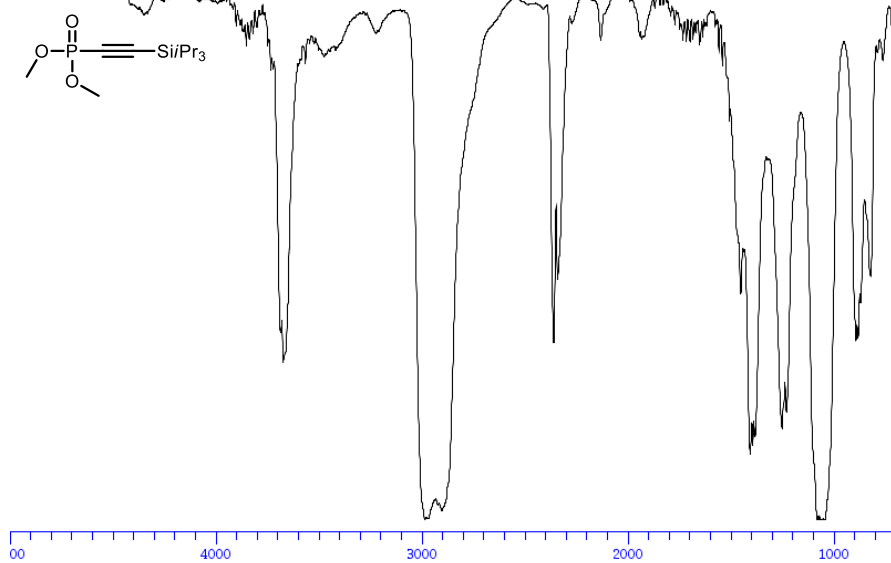


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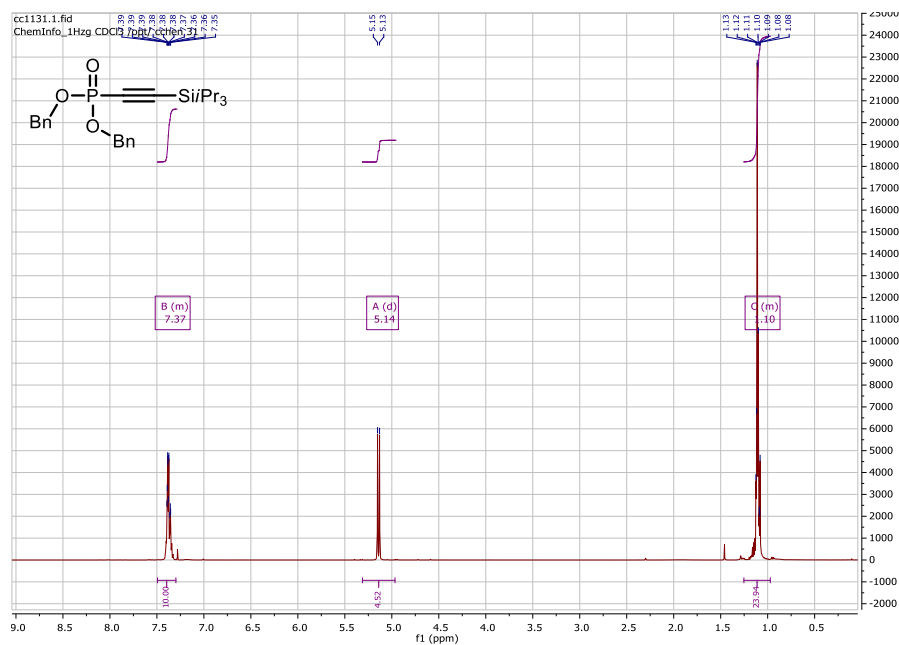
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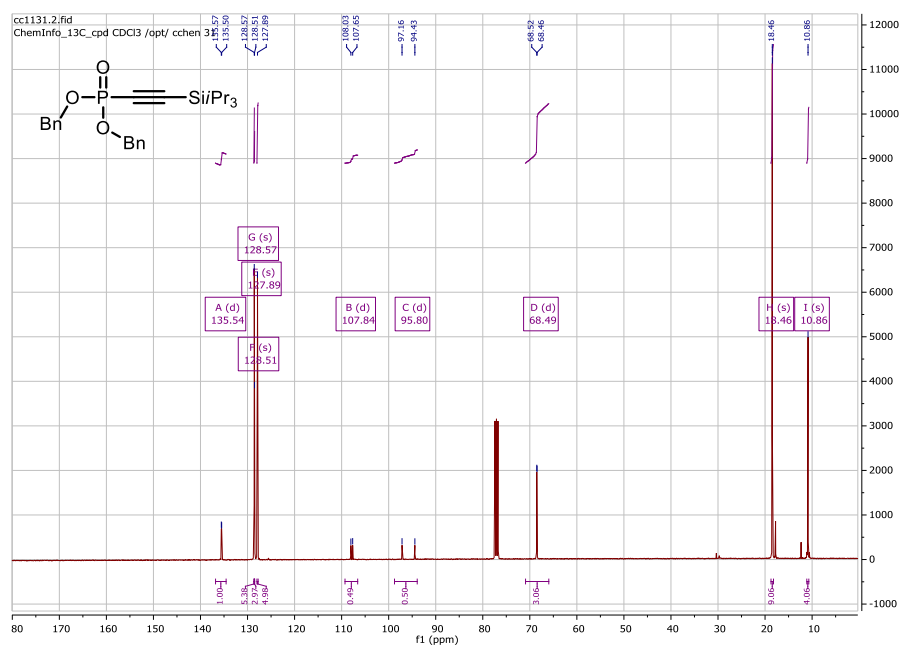
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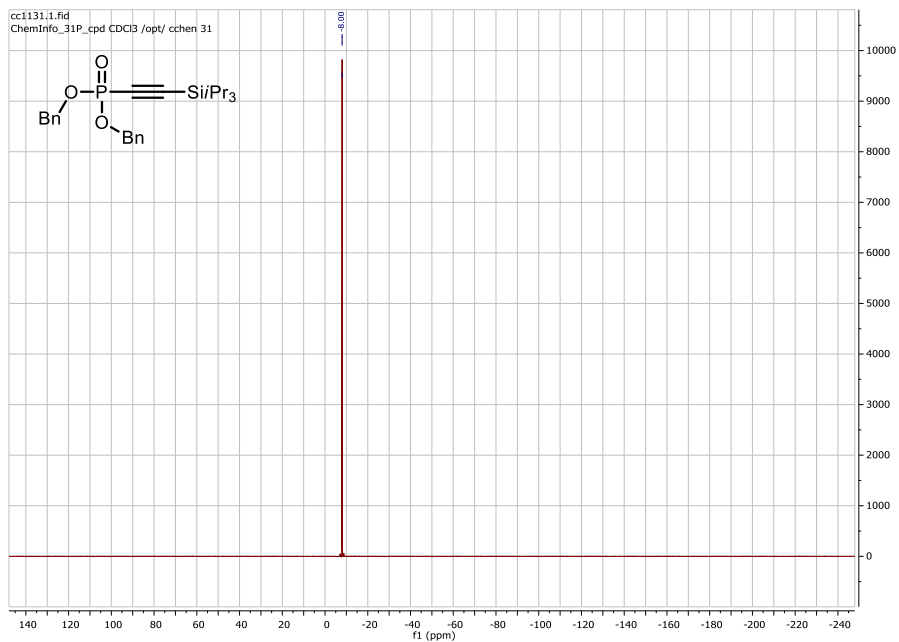
### $^1\text{H-NMR}$ (400 MHz, $\text{CDCl}_3$ ) of compound **7c**



### $^{13}\text{C-NMR}$ (101 MHz, $\text{CDCl}_3$ ) of compound **7c**



# <sup>31</sup>P-NMR (162 MHz, CDCl<sub>3</sub>) of compound **7c**

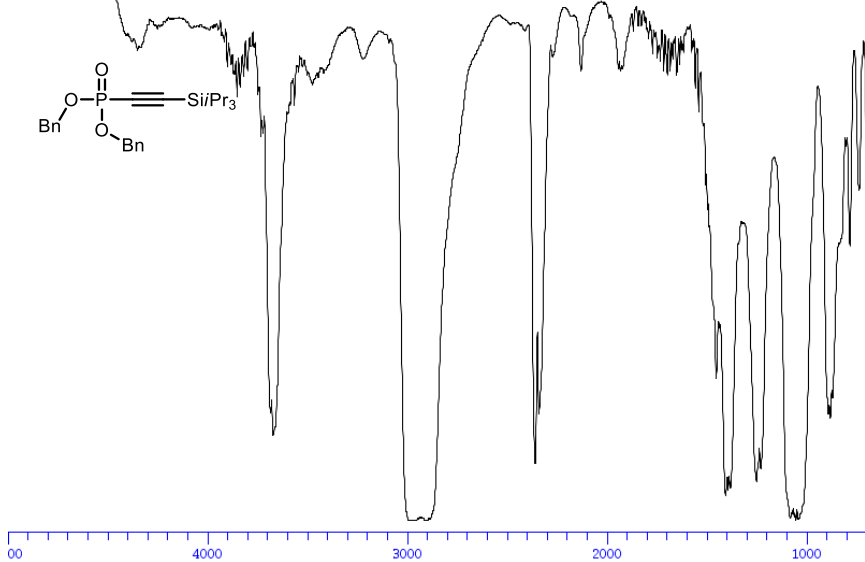


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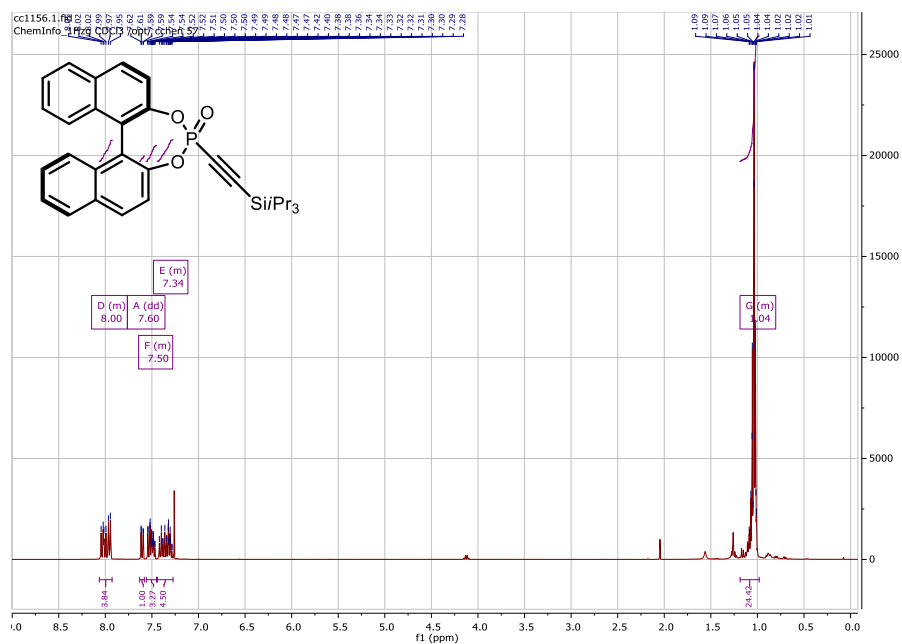
2

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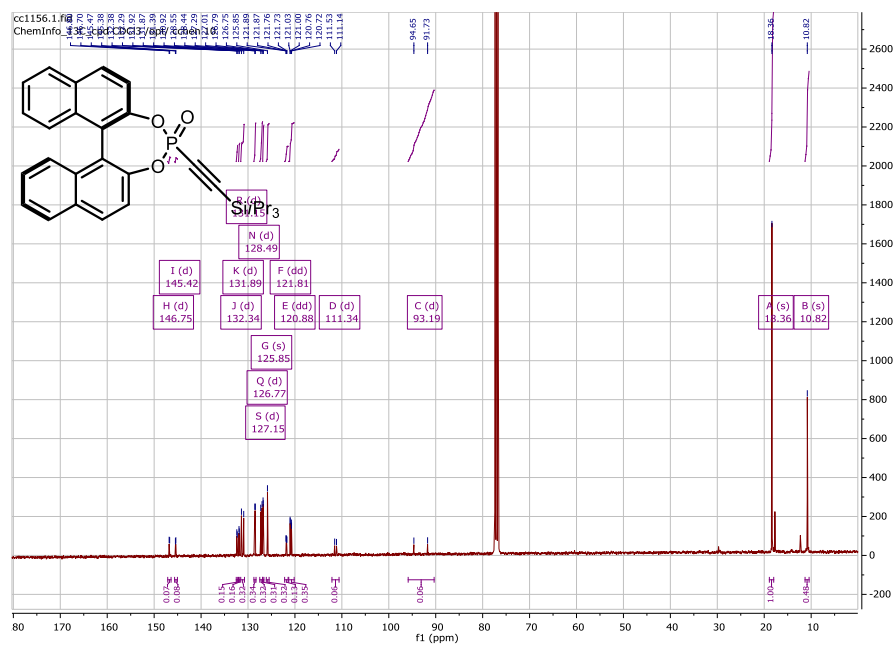
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**<sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) of compound 7d**

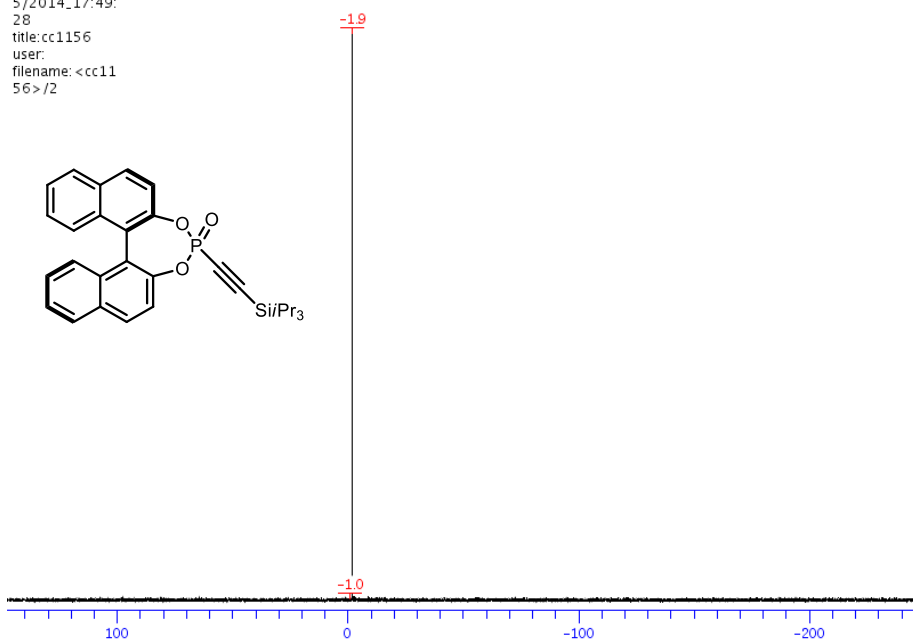


**<sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>) of compound 7d**

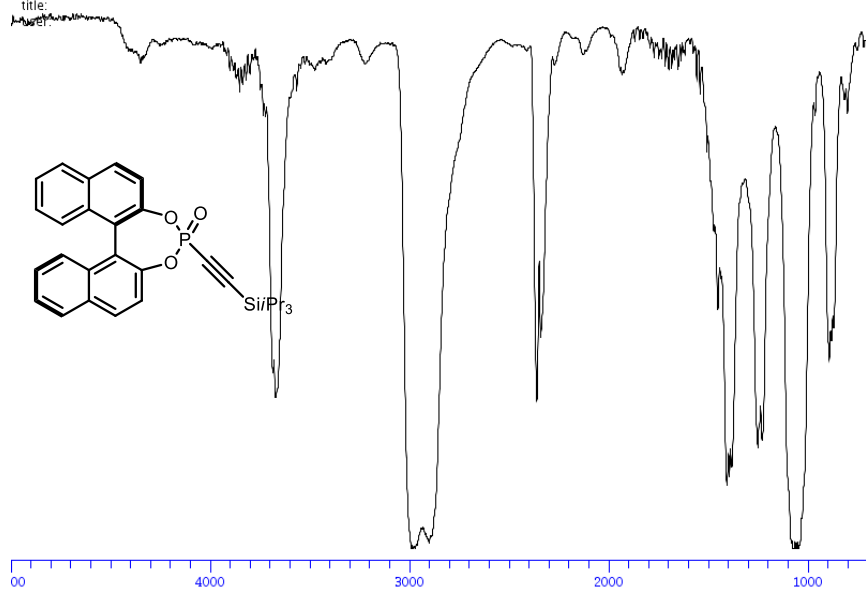


# <sup>31</sup>P-NMR (162 MHz, CDCl<sub>3</sub>) of compound 7d

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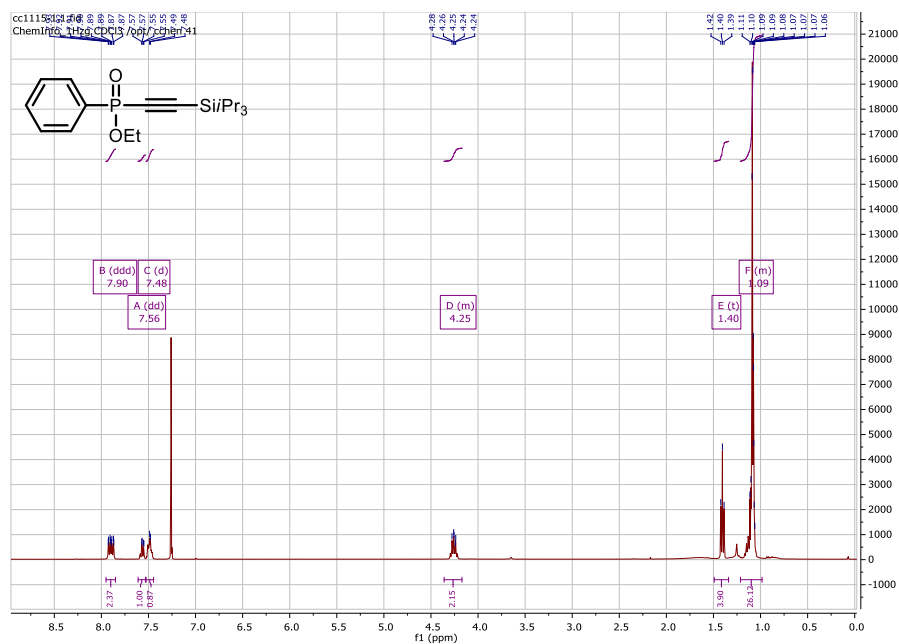


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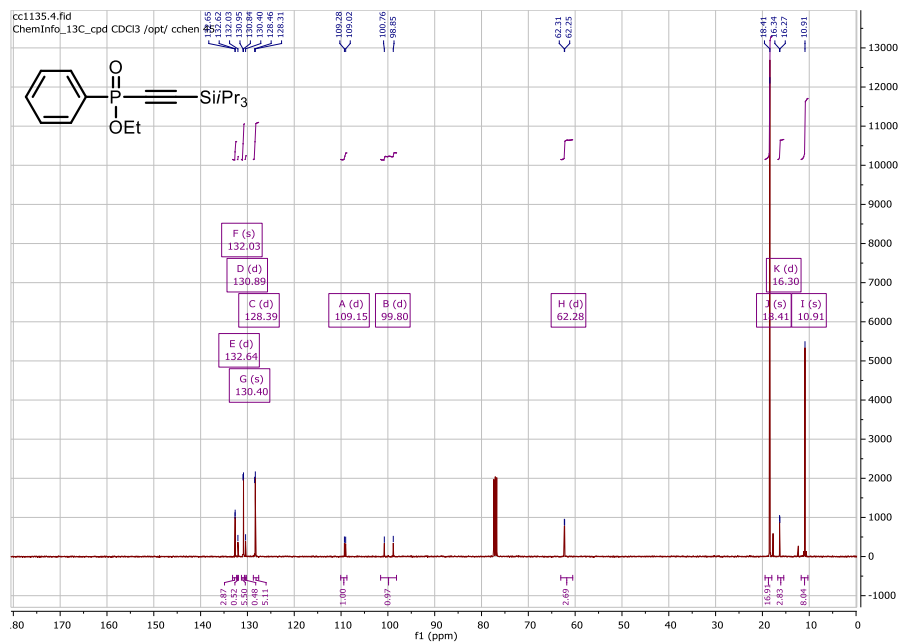




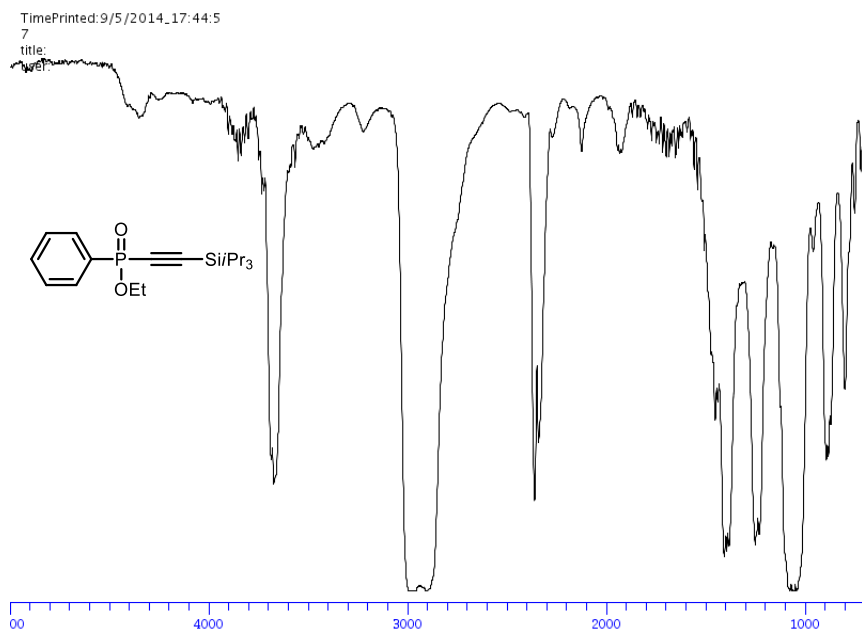
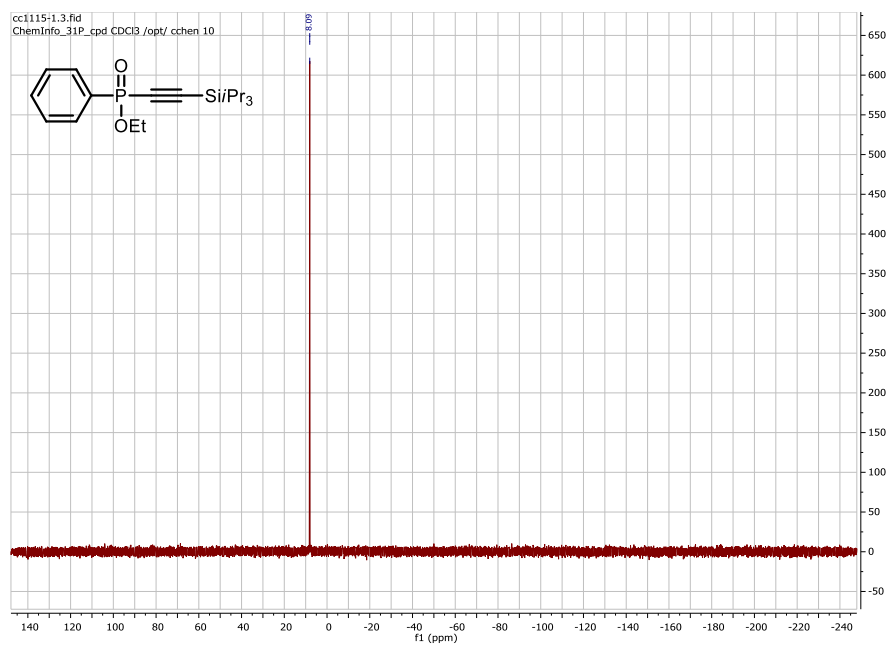
# <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) of compound 7e



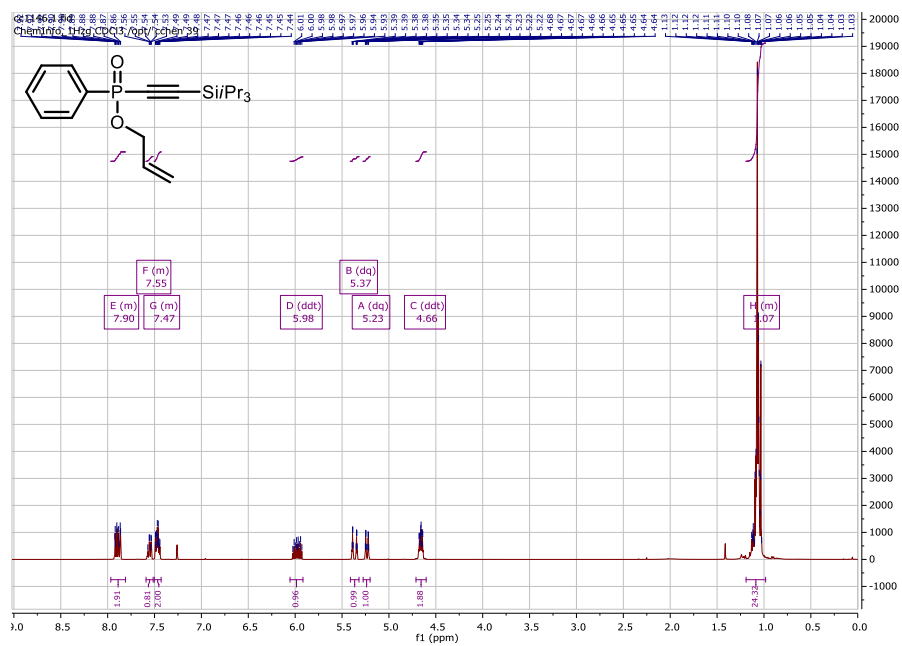
# <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>) of compound 7e



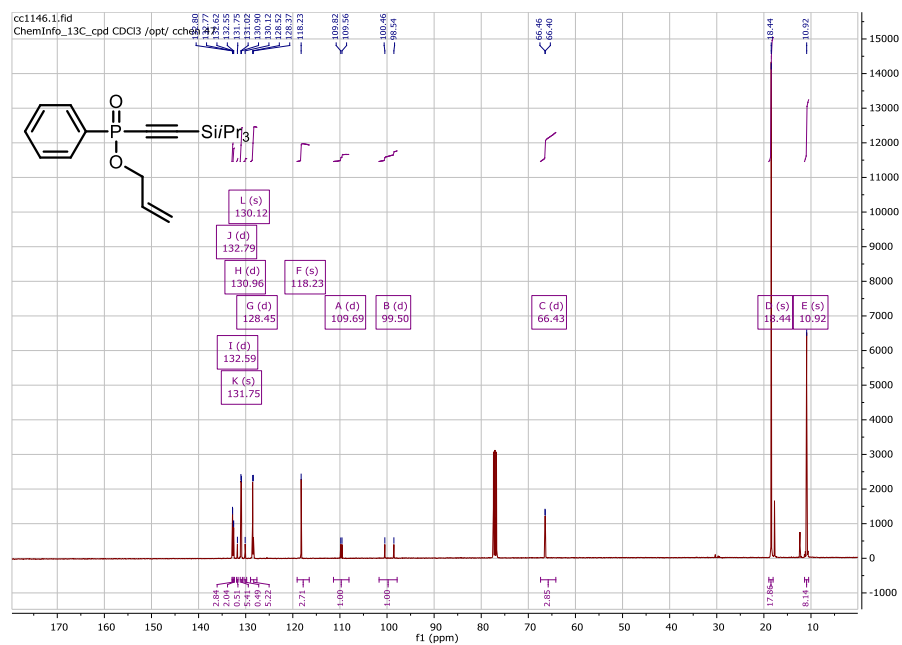
### $^{31}\text{P}$ -NMR (162 MHz, $\text{CDCl}_3$ ) of compound **7e**



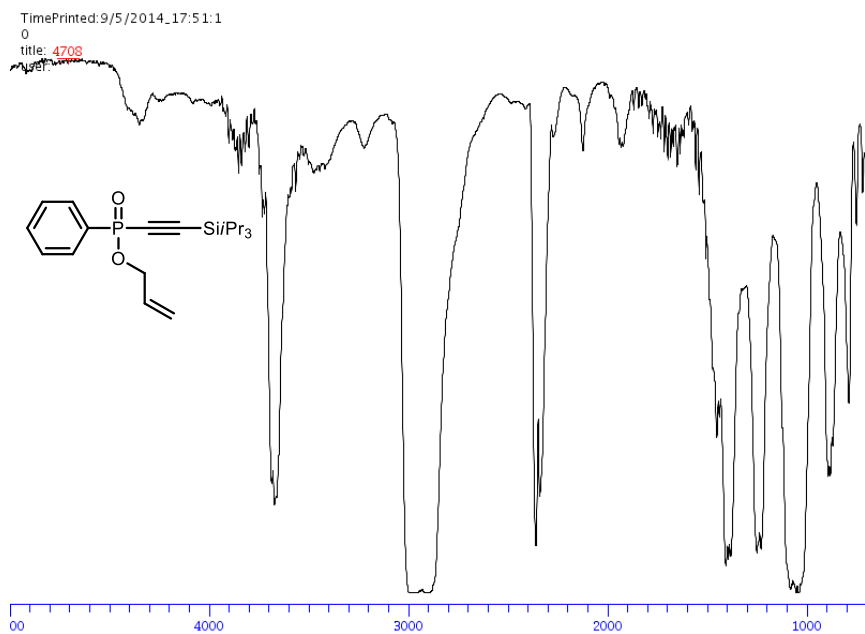
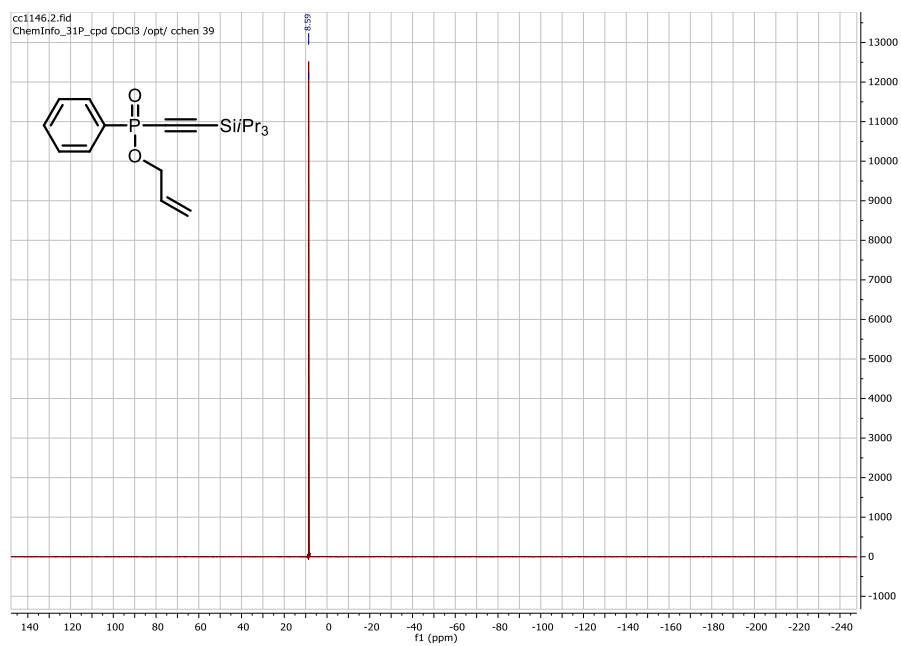
**<sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) of compound 7f**



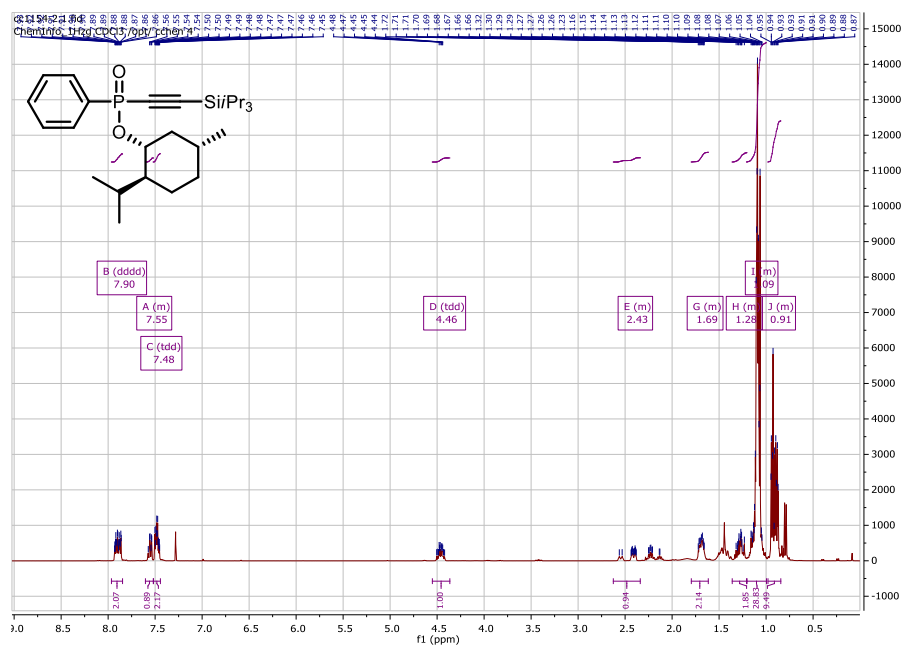
**<sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>) of compound 7f**



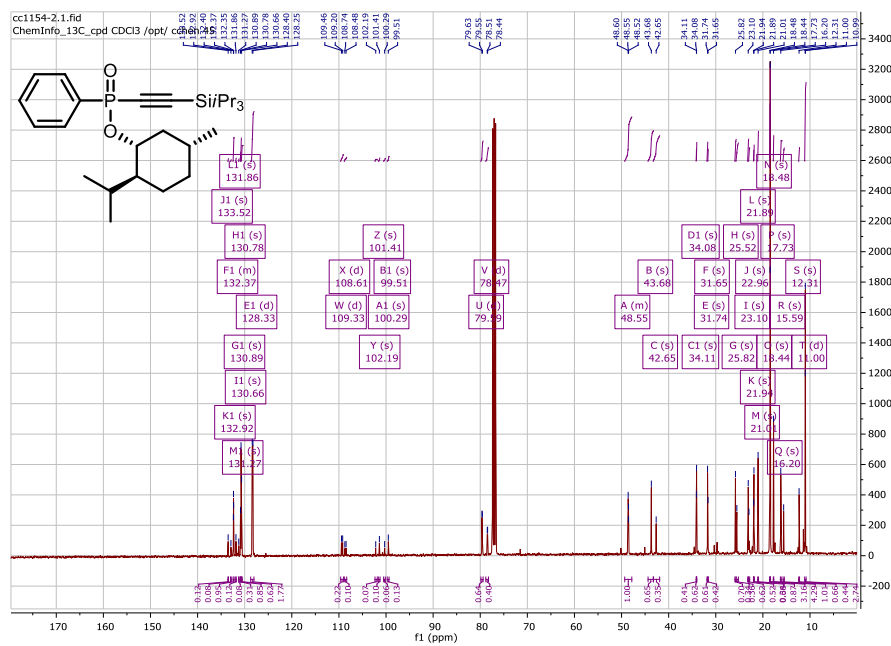
### $^{31}\text{P}$ -NMR (162 MHz, $\text{CDCl}_3$ ) of compound **7f**



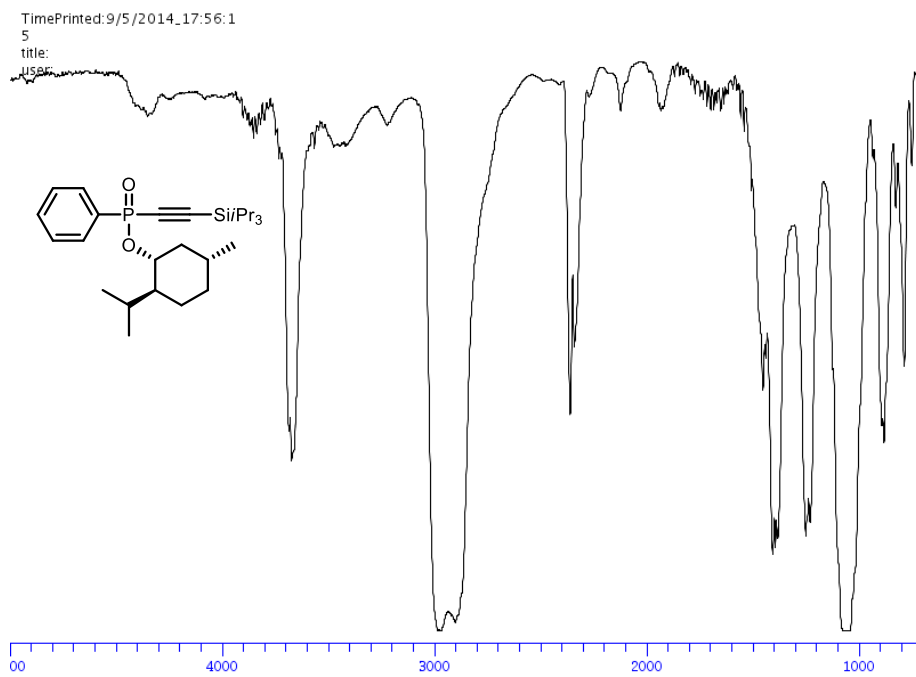
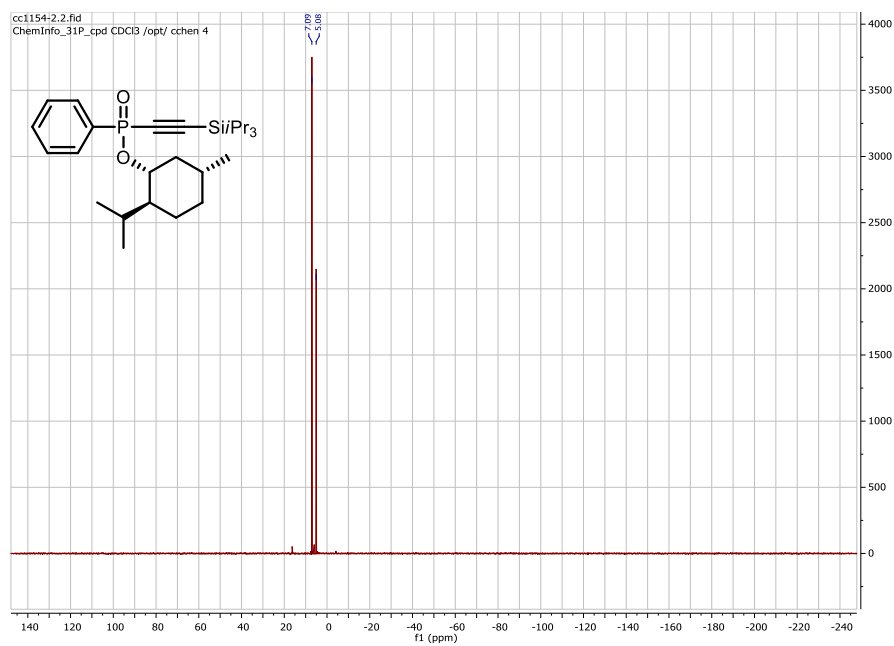
# <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) of compound 7g



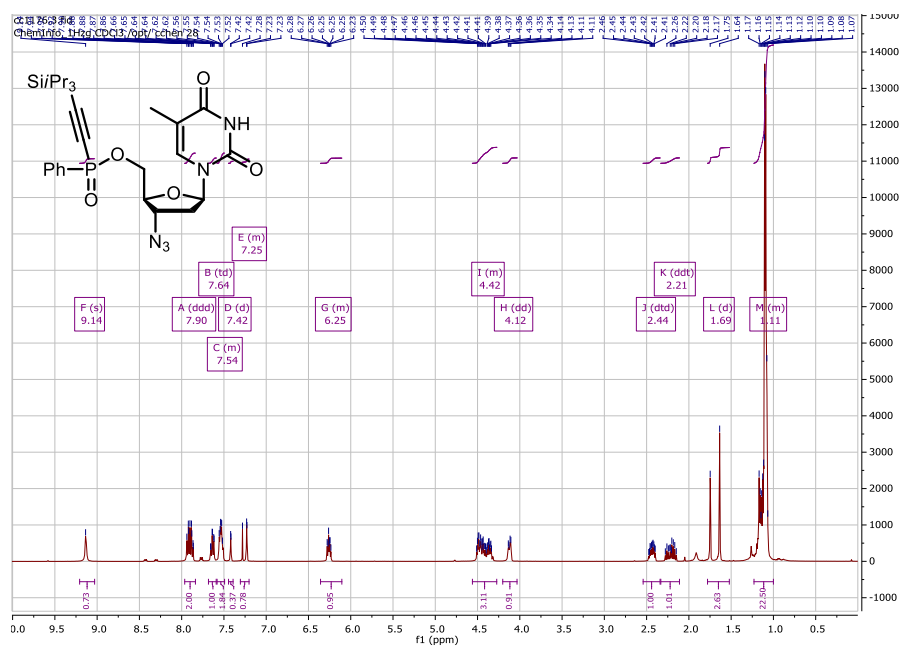
# <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>) of compound 7g



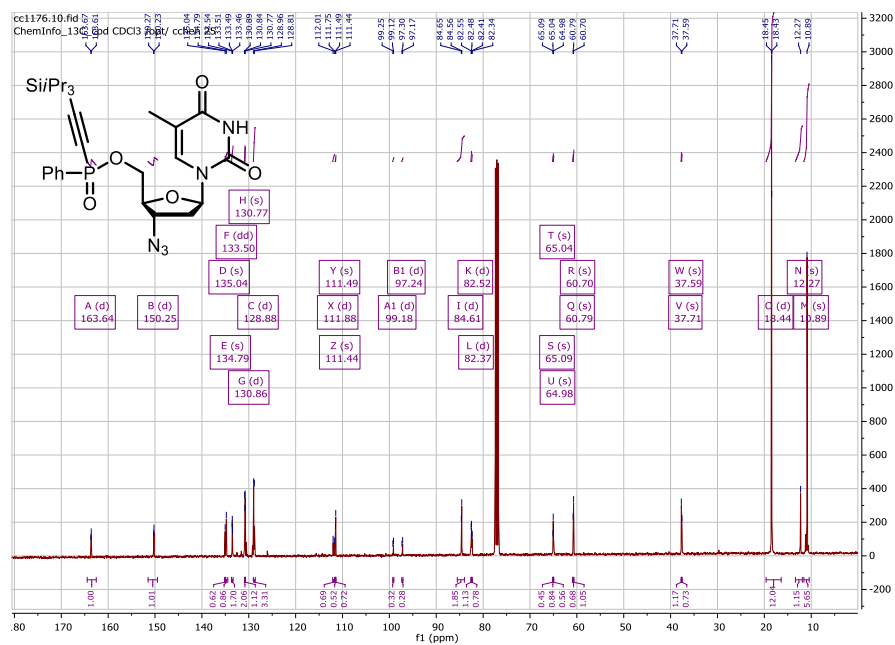
### $^{31}\text{P}$ -NMR (162 MHz, $\text{CDCl}_3$ ) of compound **7g**



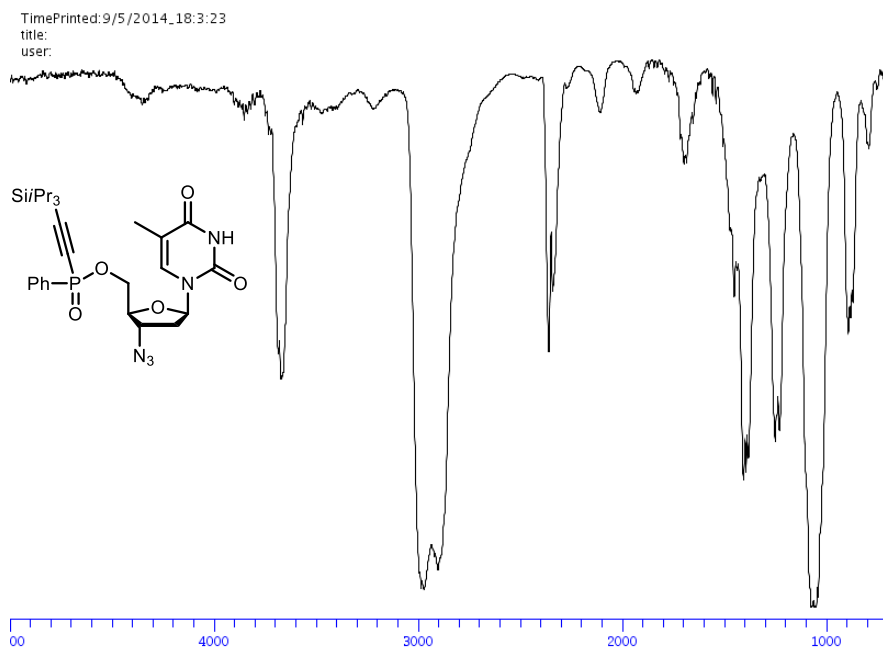
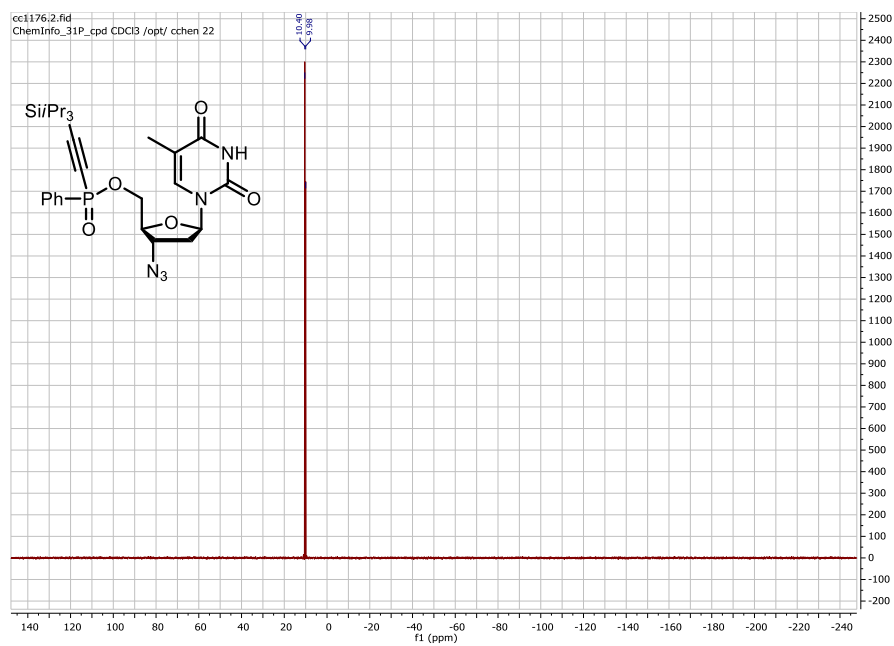
# <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) of compound 7h



# <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>) of compound 7h

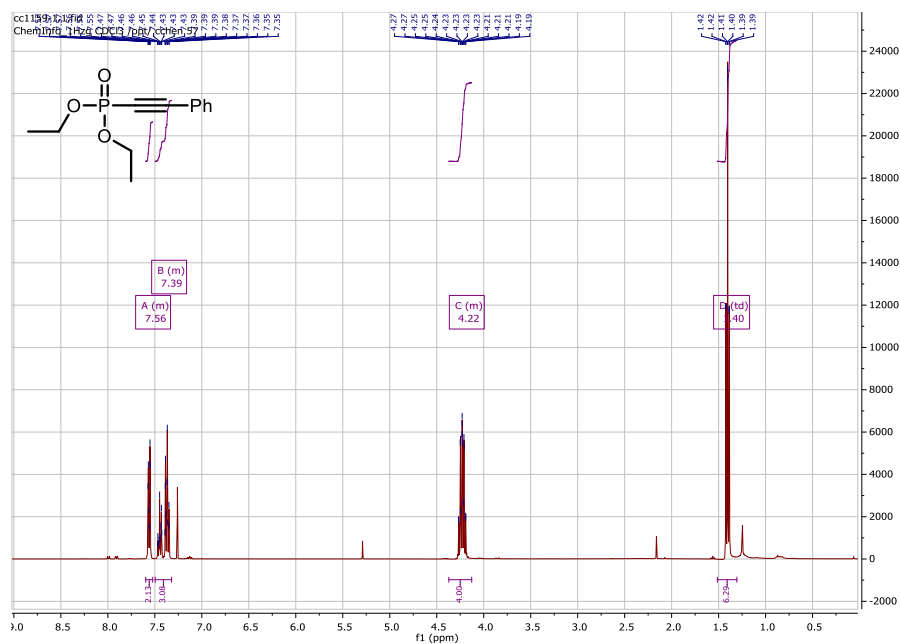


### <sup>31</sup>P-NMR (162 MHz, CDCl<sub>3</sub>) of compound **7h**

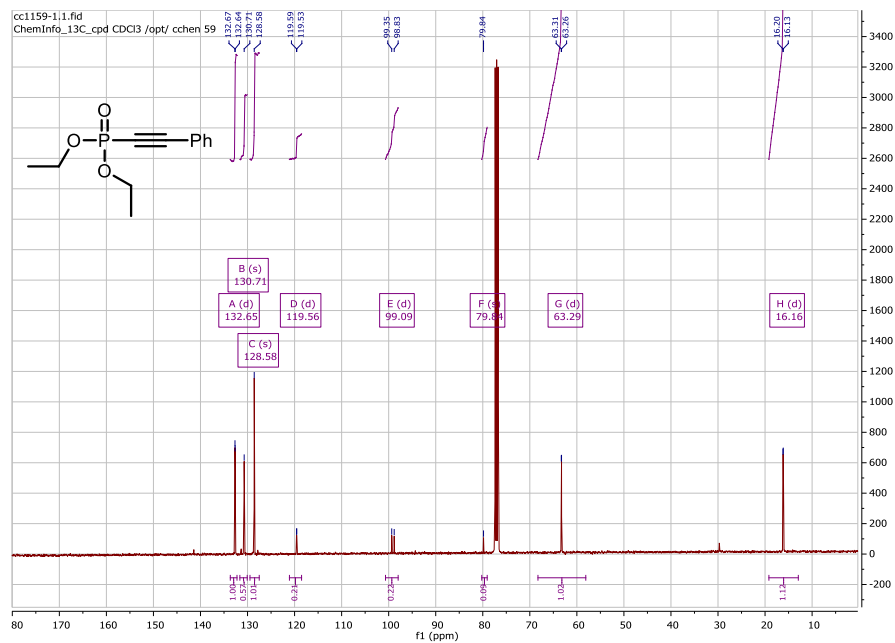




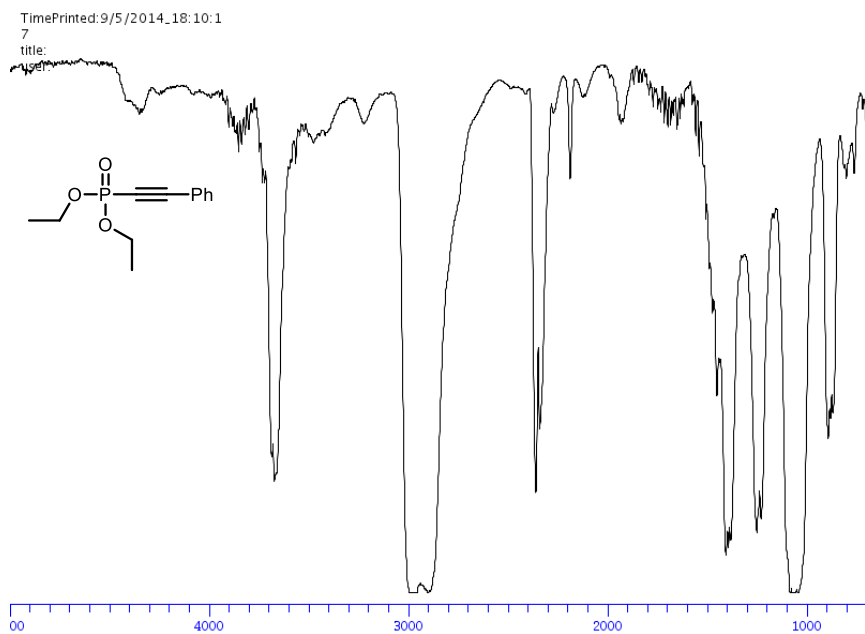
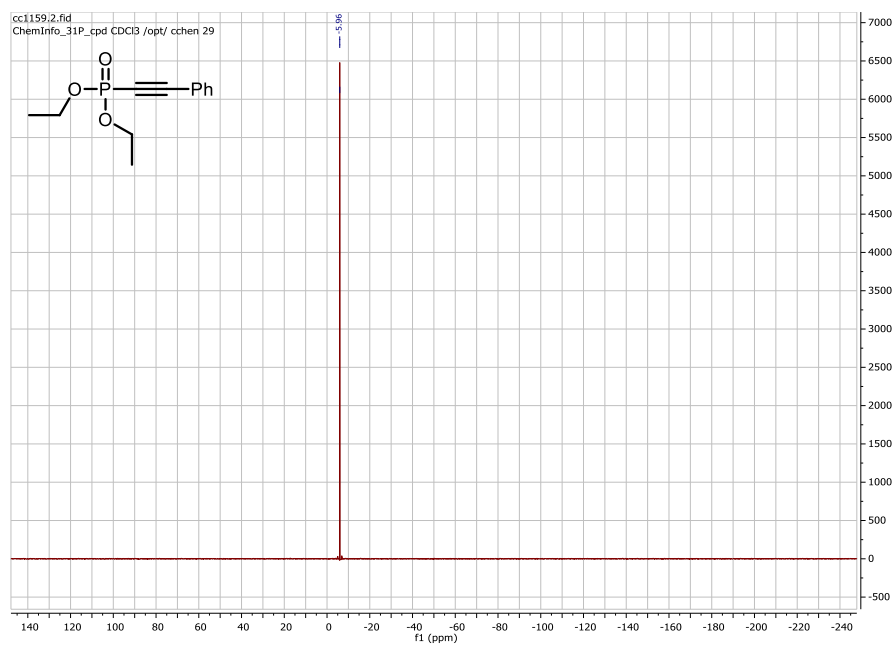
### <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) of compound 7j



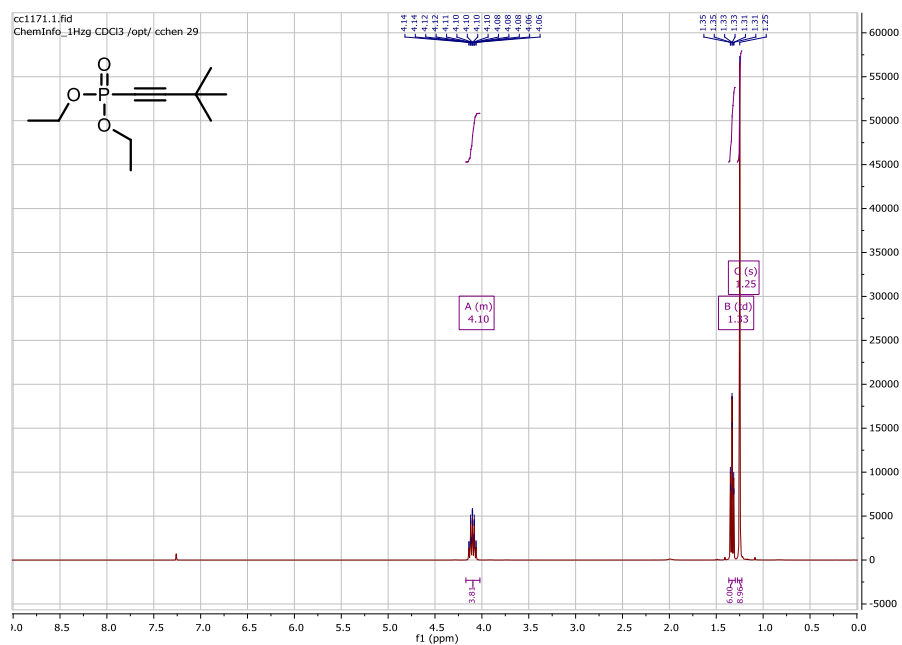
### <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>) of compound 7j



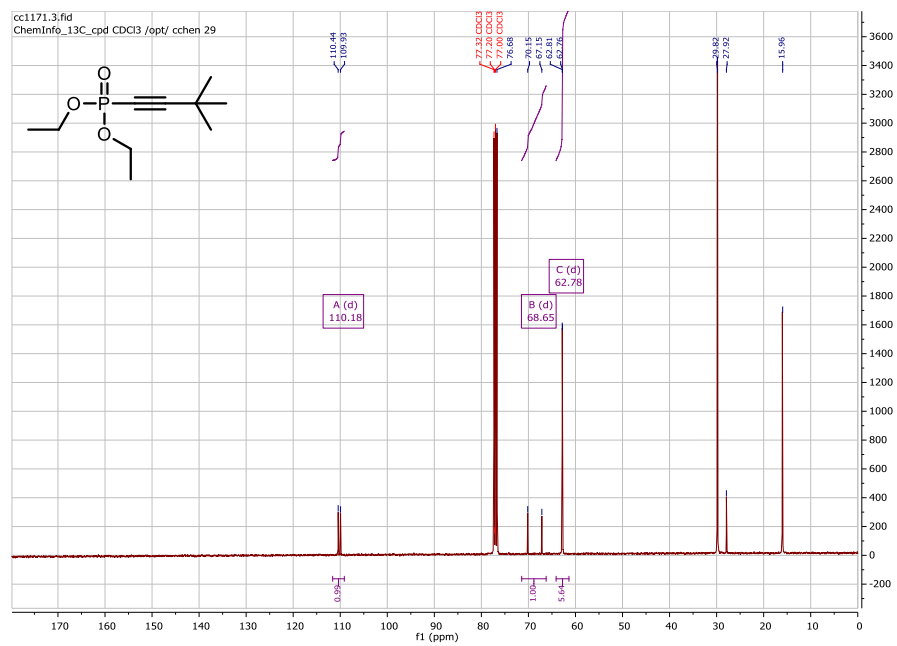
# <sup>31</sup>P-NMR (162 MHz, CDCl<sub>3</sub>) of compound **7j**



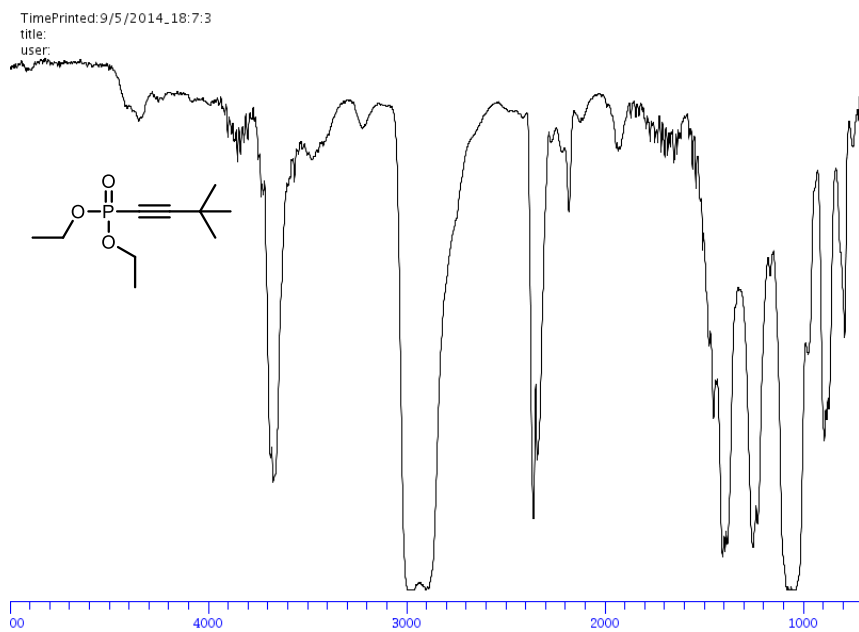
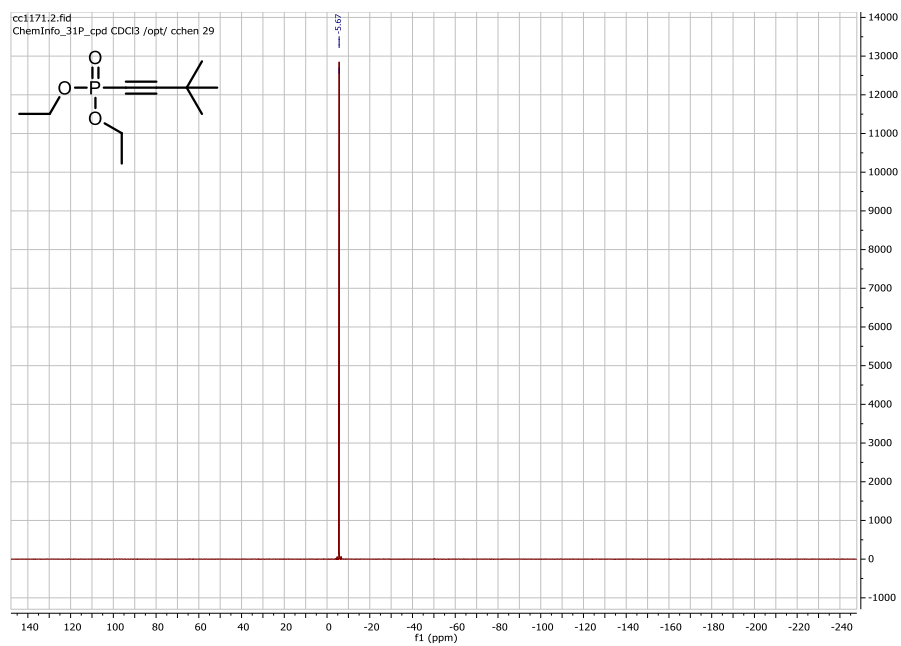
# <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) of compound 7k



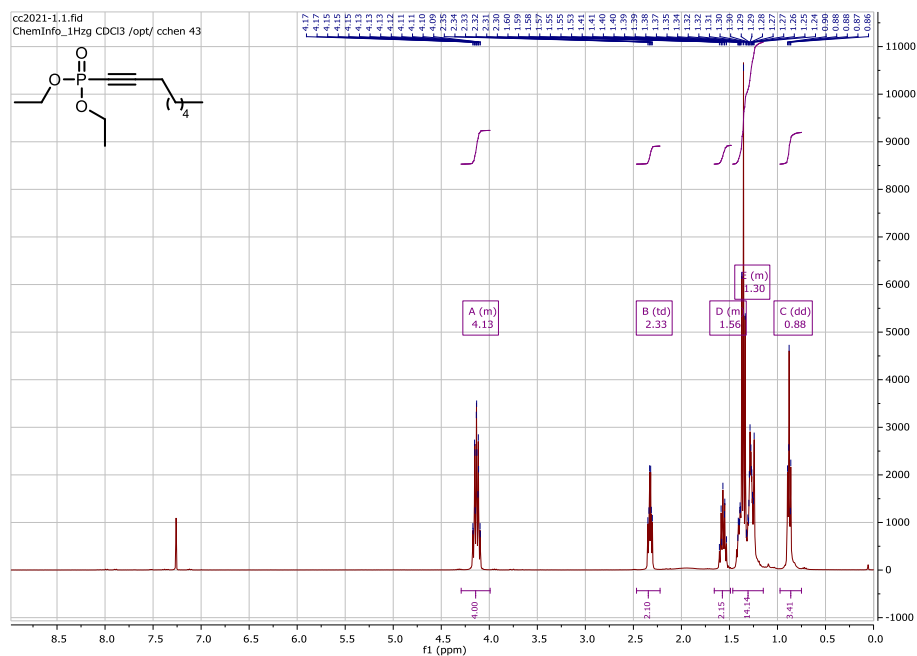
# <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>) of compound 7k



### $^{31}\text{P}$ -NMR (162 MHz, $\text{CDCl}_3$ ) of compound **7k**



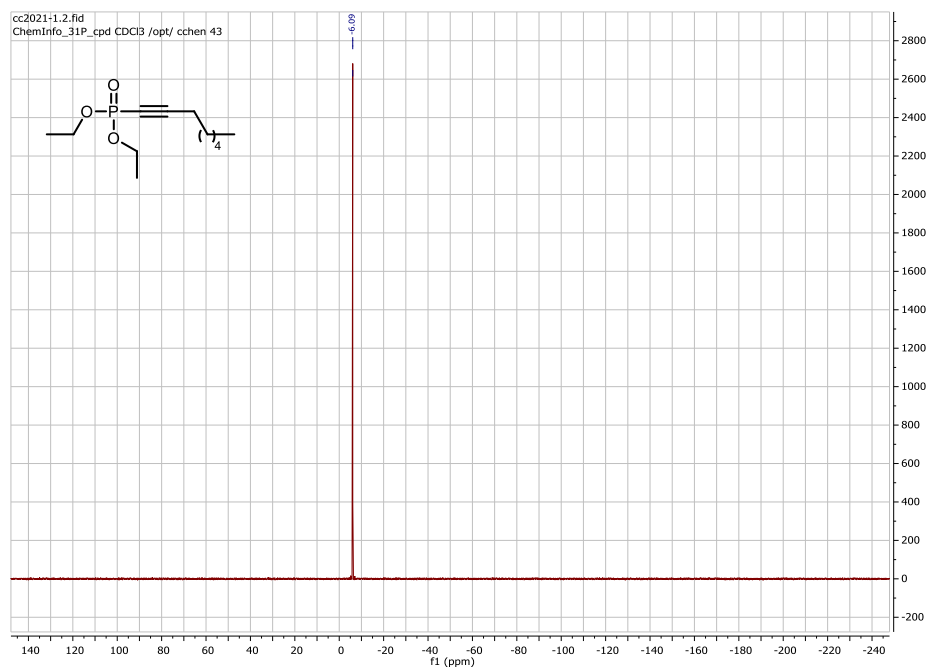
### <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) of compound **71**



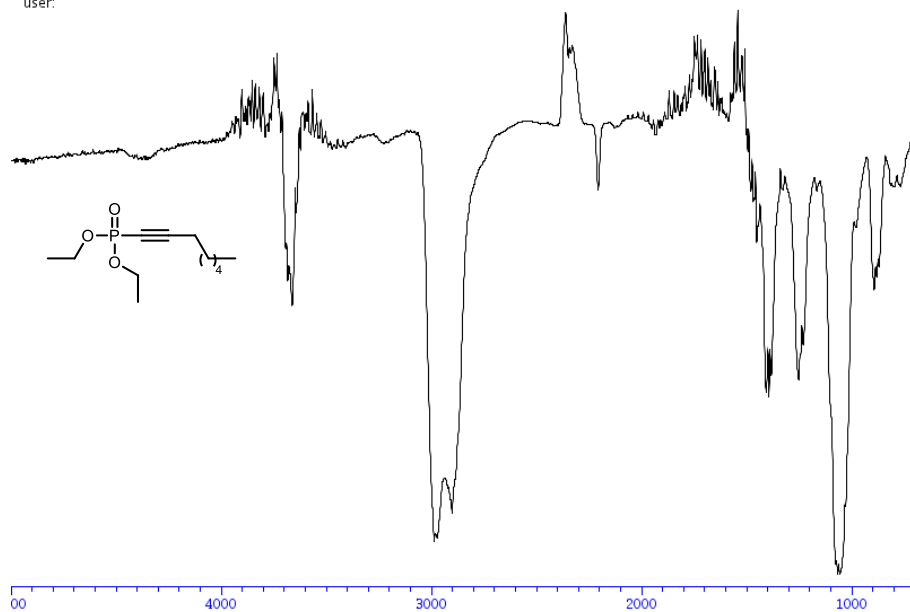
### <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>) of compound **71**



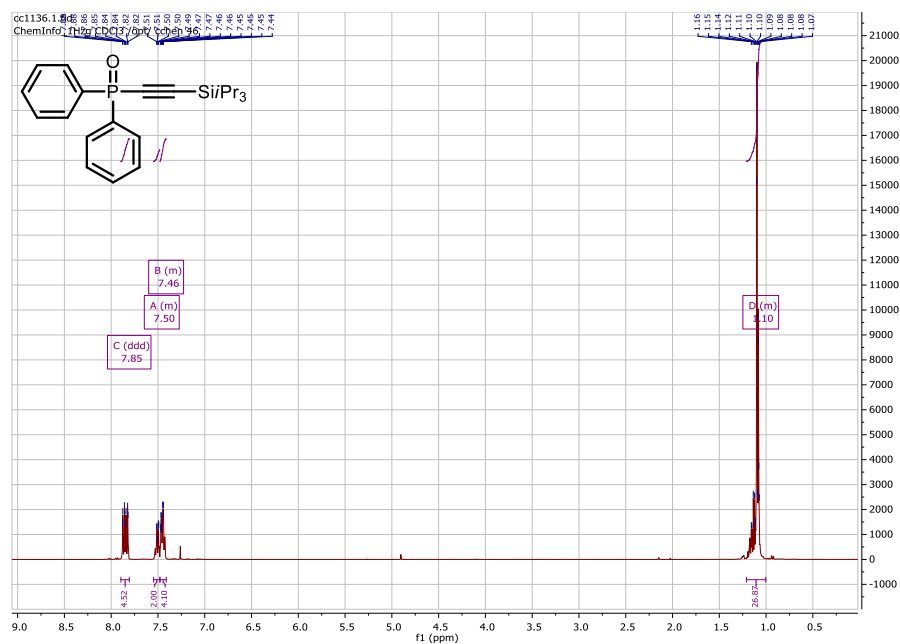
### <sup>31</sup>P-NMR (162 MHz, CDCl<sub>3</sub>) of compound **71**



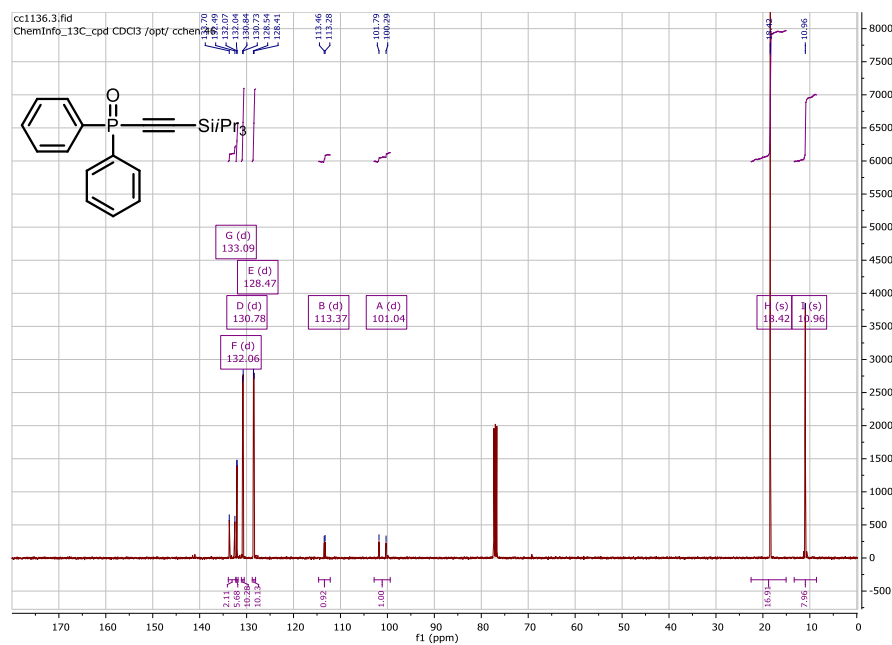
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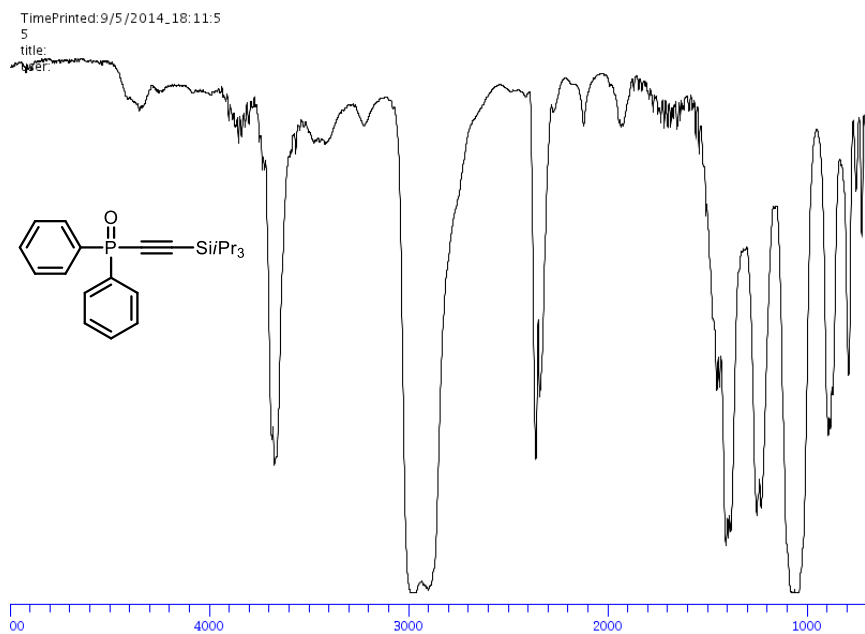
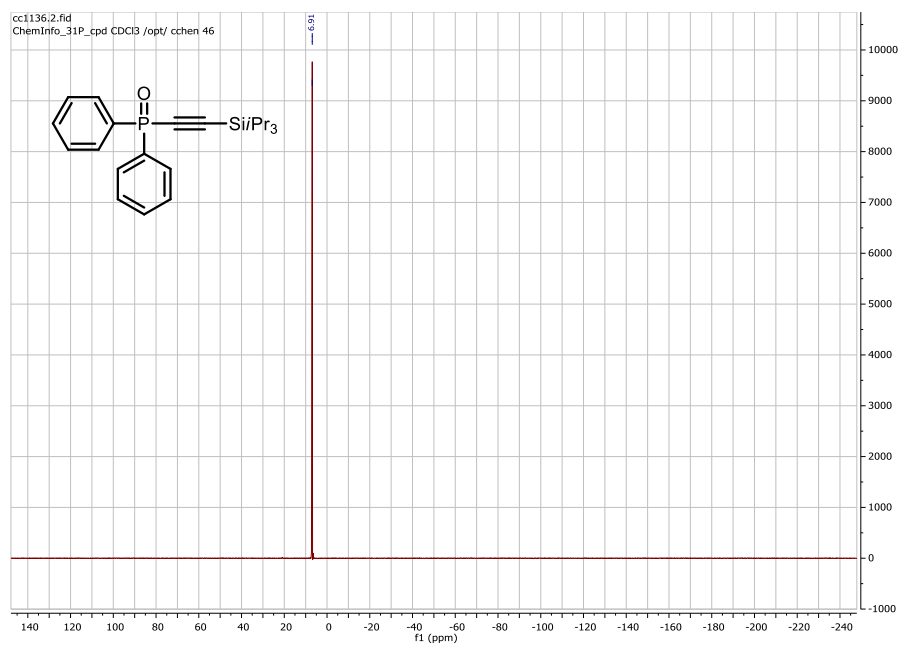
# <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) of compound 9a



# <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>) of compound 9a



### $^{31}\text{P}$ -NMR (162 MHz, $\text{CDCl}_3$ ) of compound **9a**

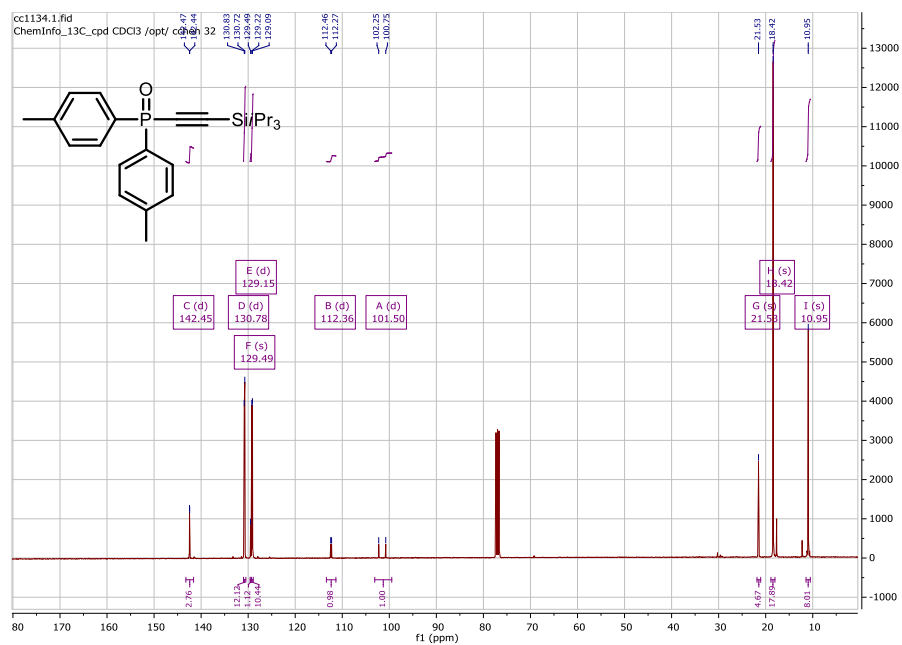




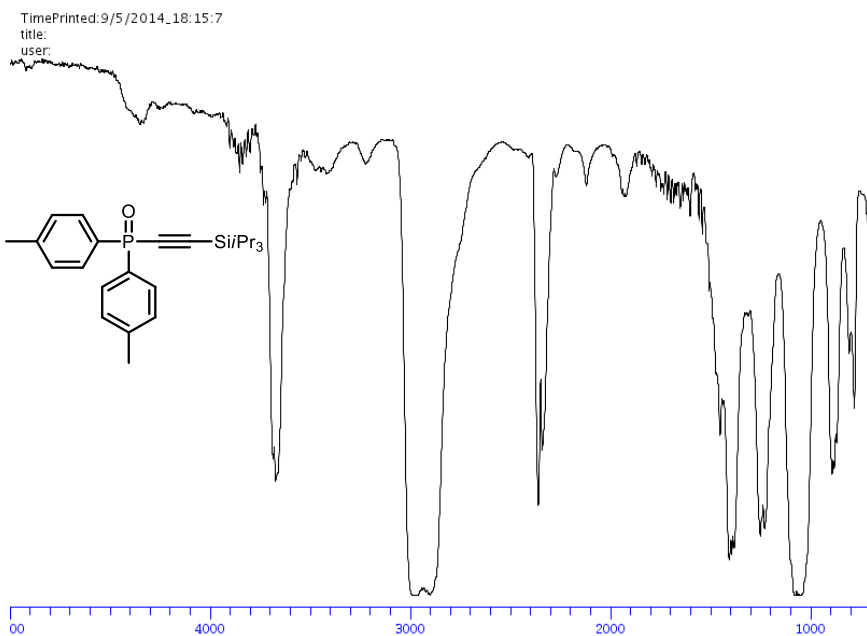
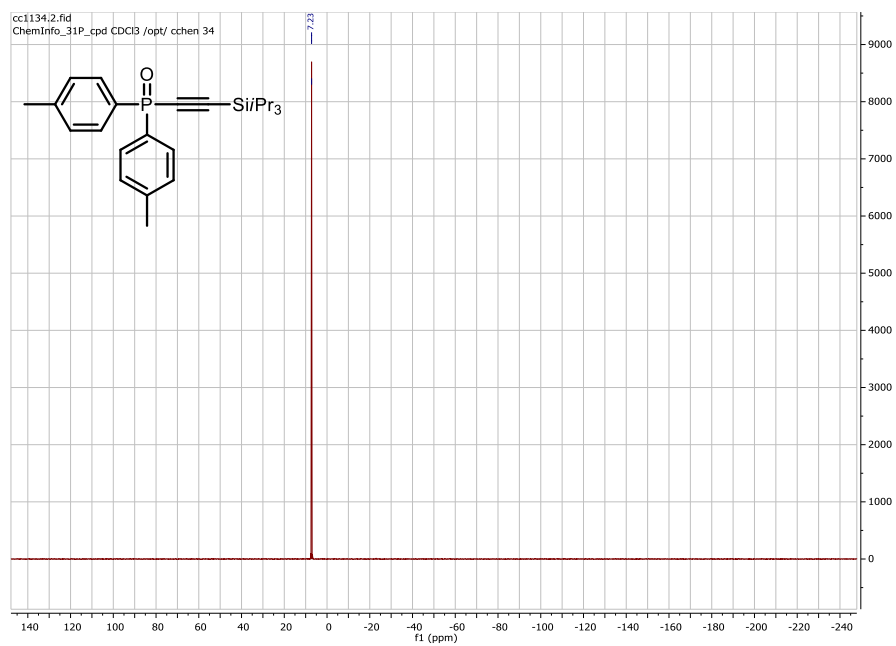
### $^1\text{H-NMR}$ (400 MHz, $\text{CDCl}_3$ ) of compound **9b**



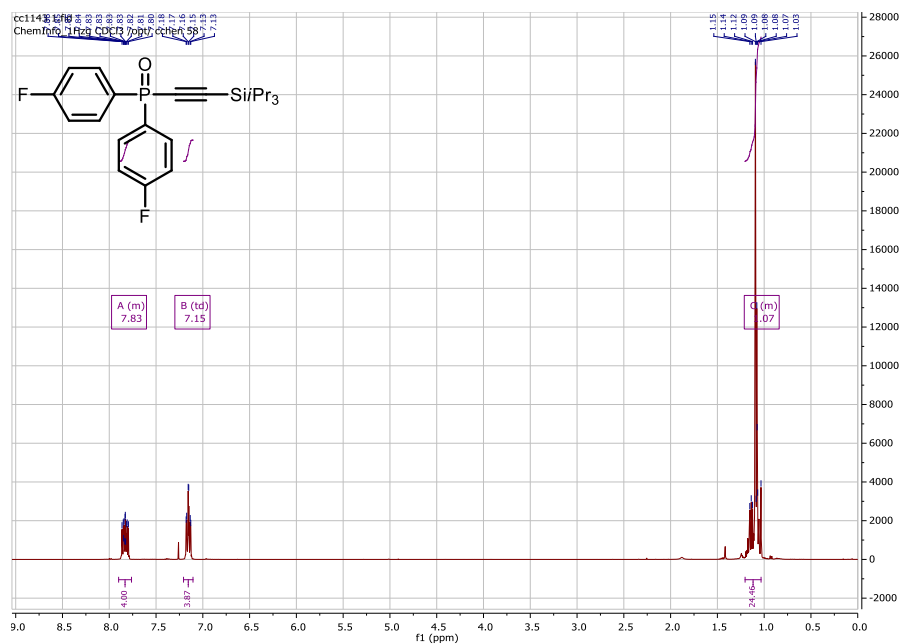
### $^{13}\text{C-NMR}$ (101 MHz, $\text{CDCl}_3$ ) of compound **9b**



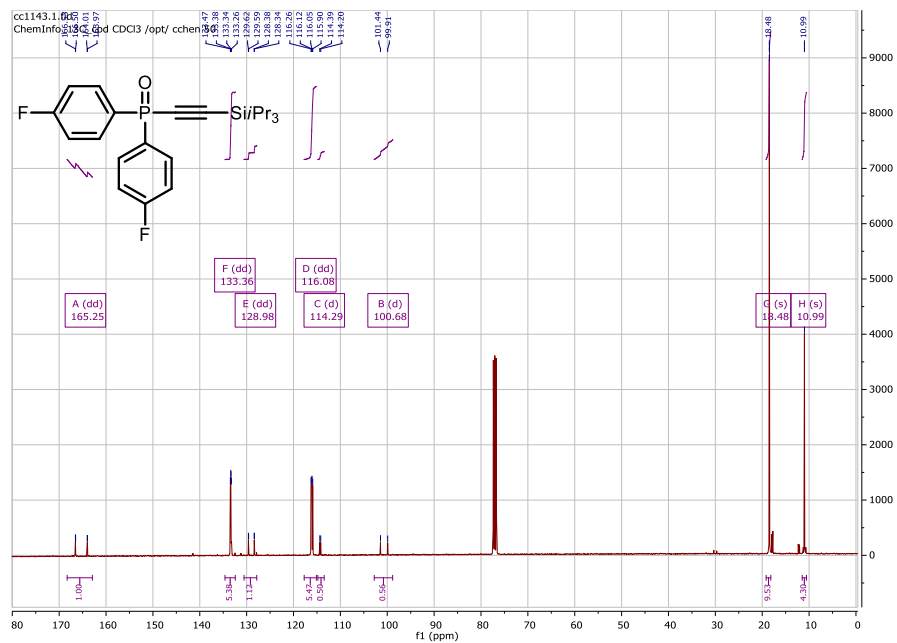
### $^{31}\text{P}$ -NMR (162 MHz, $\text{CDCl}_3$ ) of compound **9b**



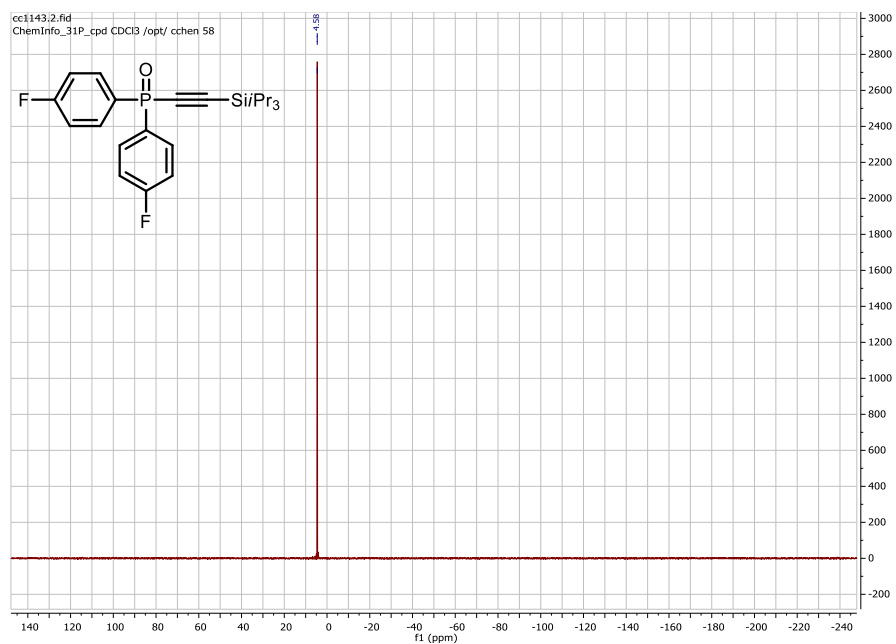
**<sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) of compound 9c**



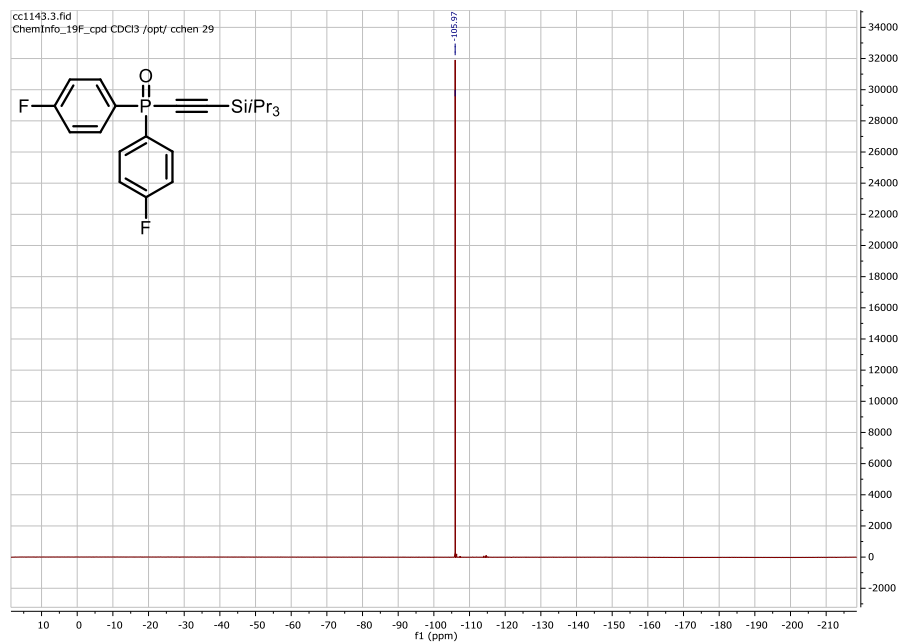
**<sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>) of compound 9c**



### $^{31}\text{P}$ -NMR (162 MHz, $\text{CDCl}_3$ ) of compound **9c**

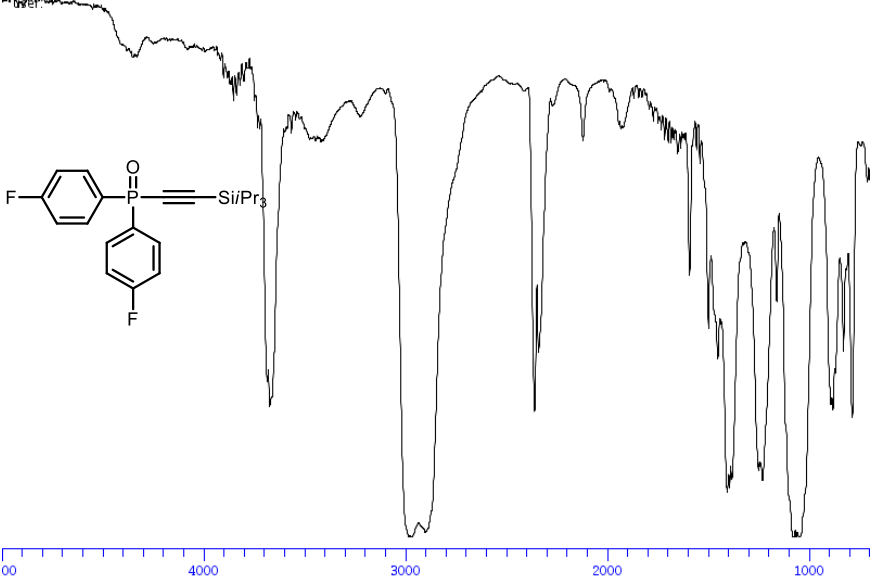


### $^{19}\text{F}$ -NMR (376 MHz, $\text{CDCl}_3$ ) of compound **9c**

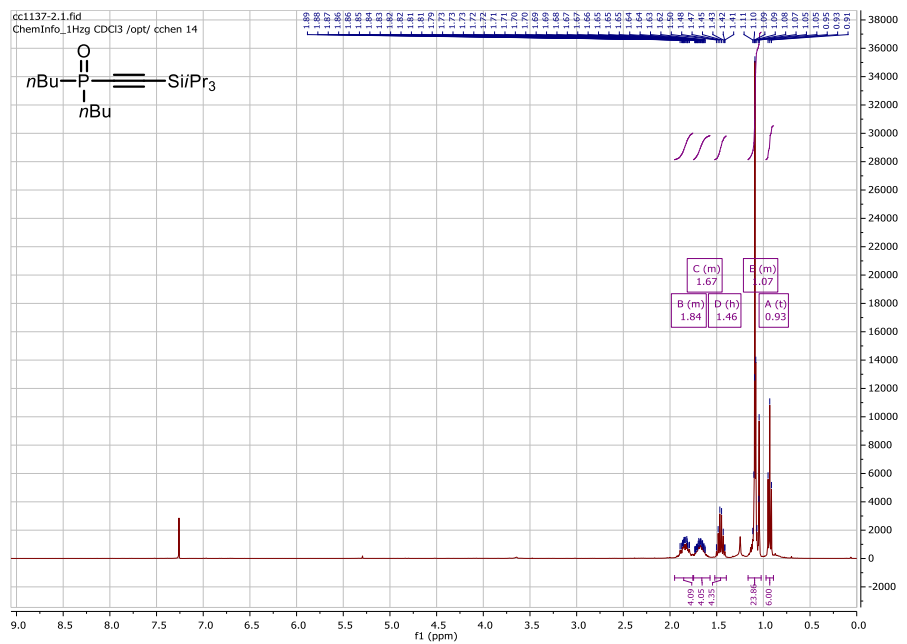


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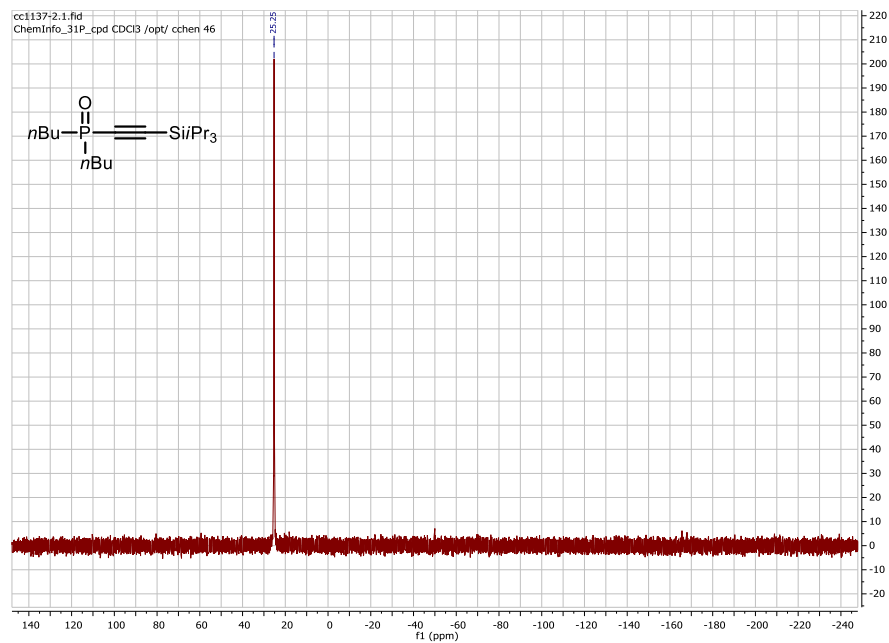
### <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) of compound 9d



### $^{13}\text{C}$ -NMR (101 MHz, $\text{CDCl}_3$ ) of compound **9d**

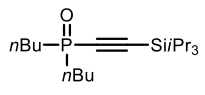
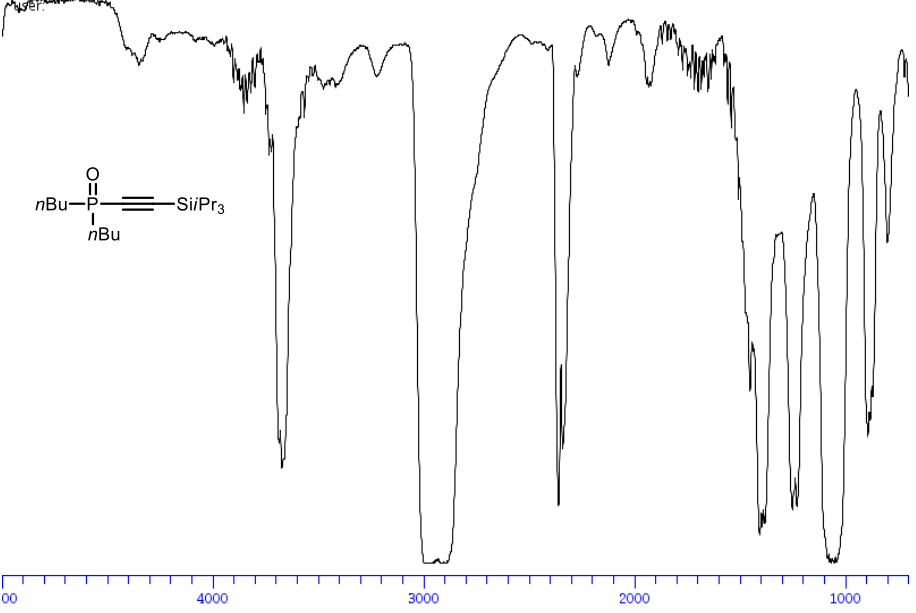


### $^{31}\text{P}$ -NMR (162 MHz, $\text{CDCl}_3$ ) of compound **9d**

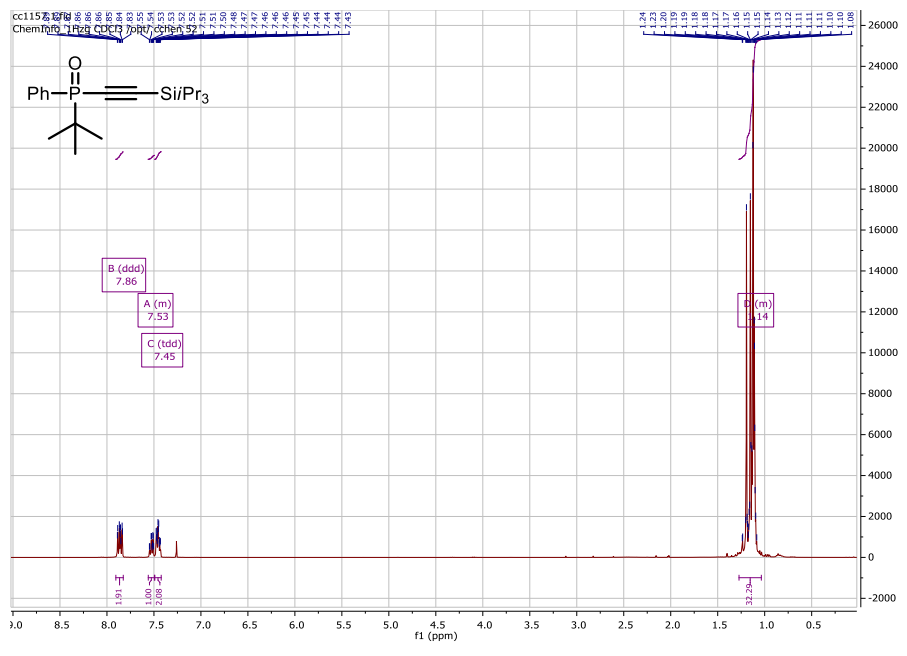


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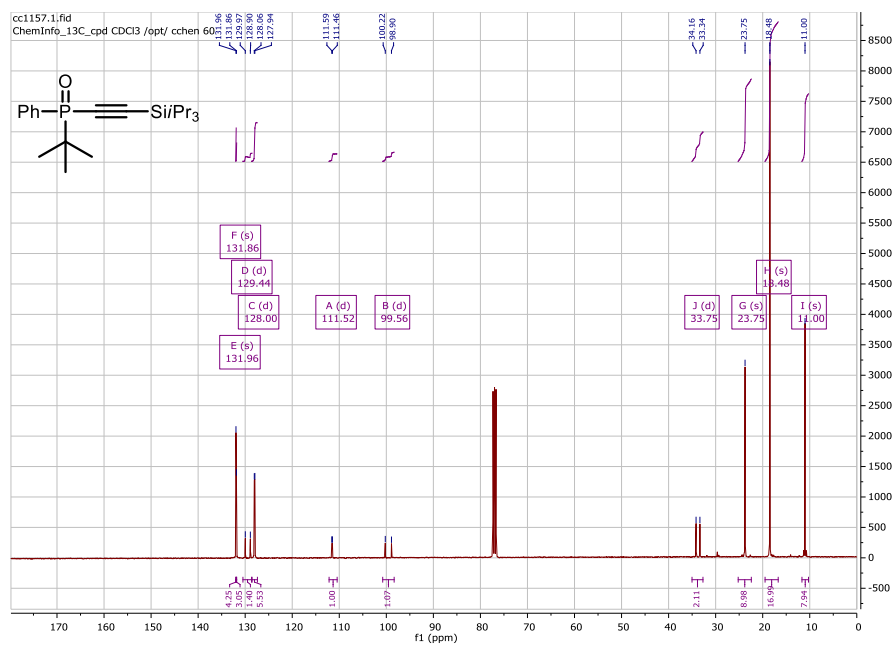
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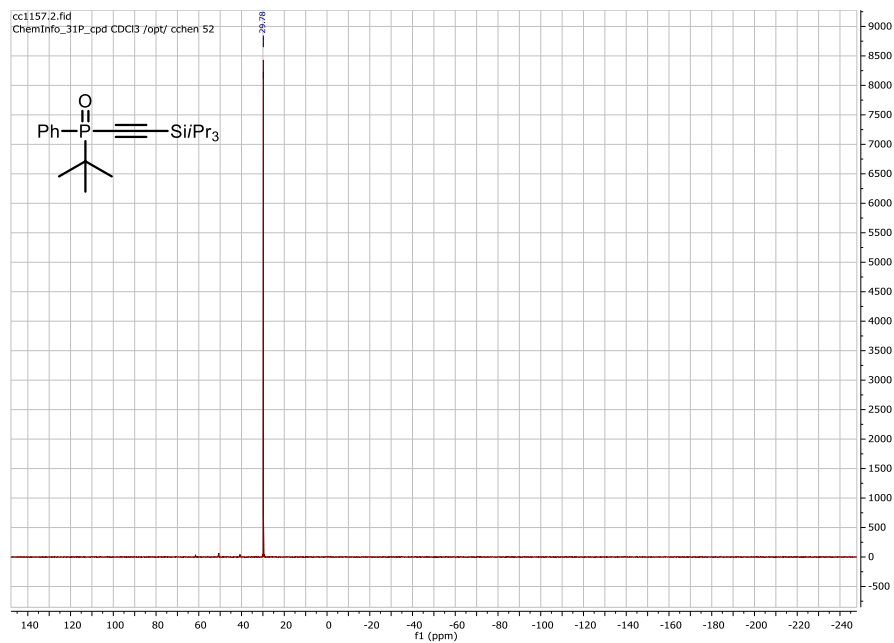
### <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) of compound 9e



### $^{13}\text{C}$ -NMR (101 MHz, $\text{CDCl}_3$ ) of compound **9e**



### $^{31}\text{P}$ -NMR (162 MHz, $\text{CDCl}_3$ ) of compound **9e**



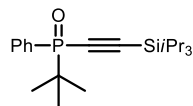
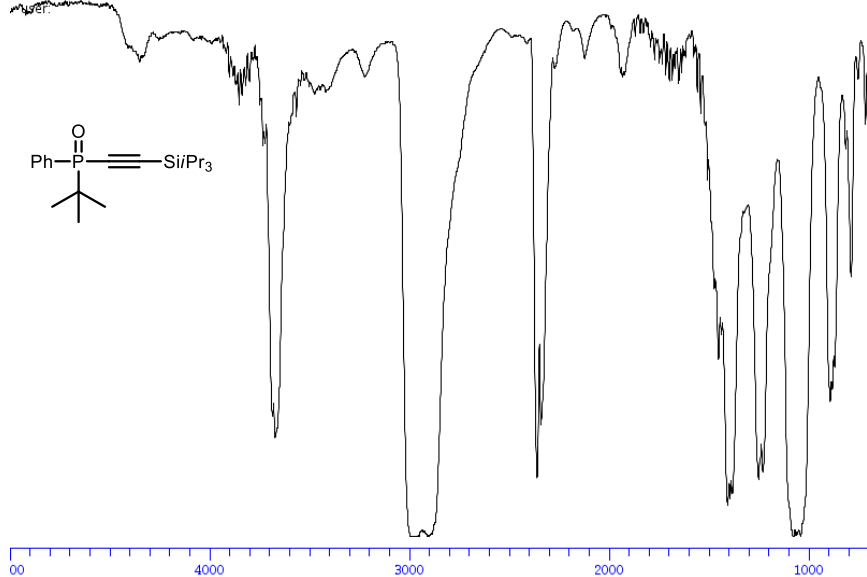


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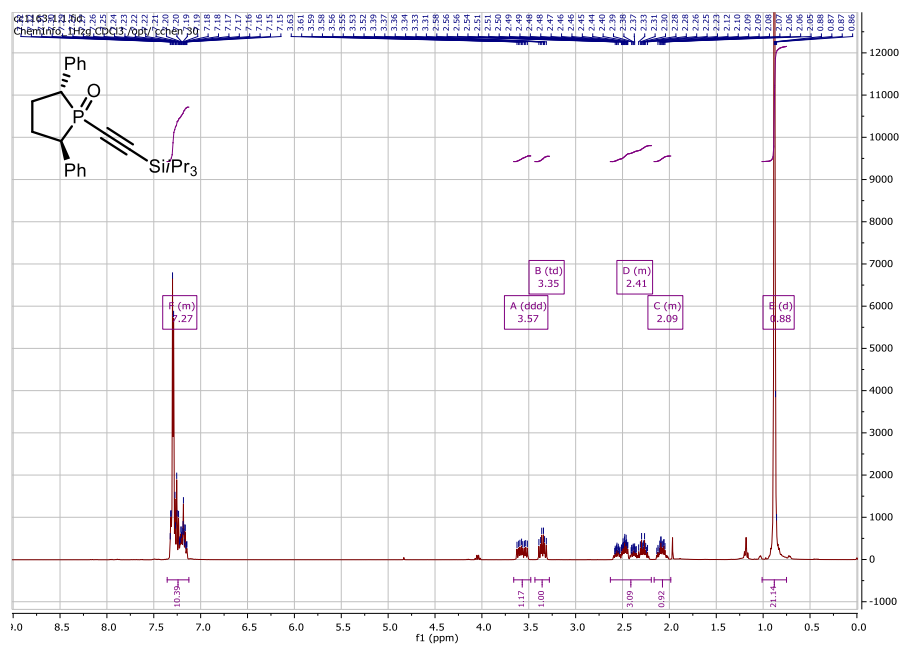
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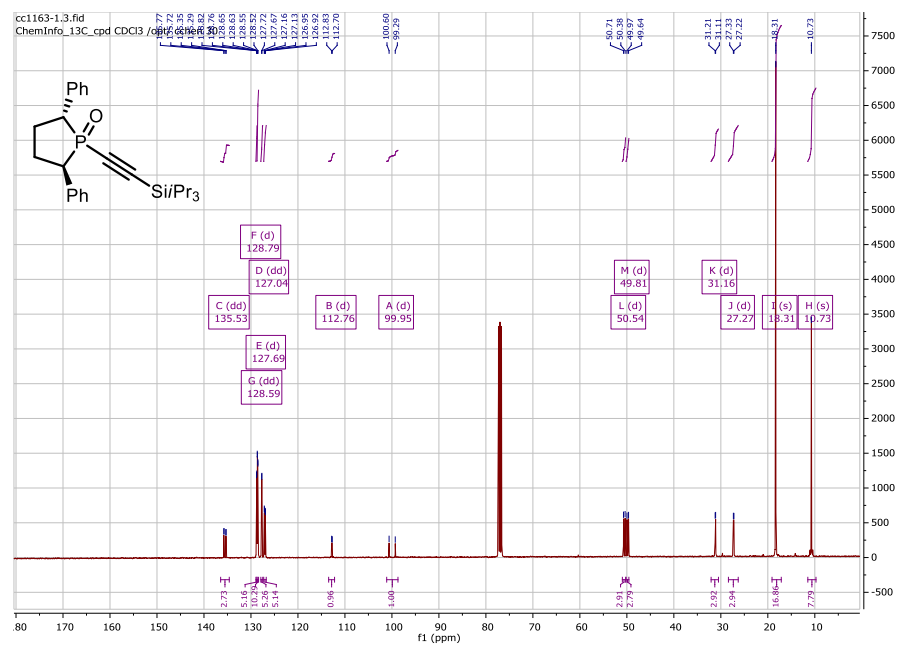
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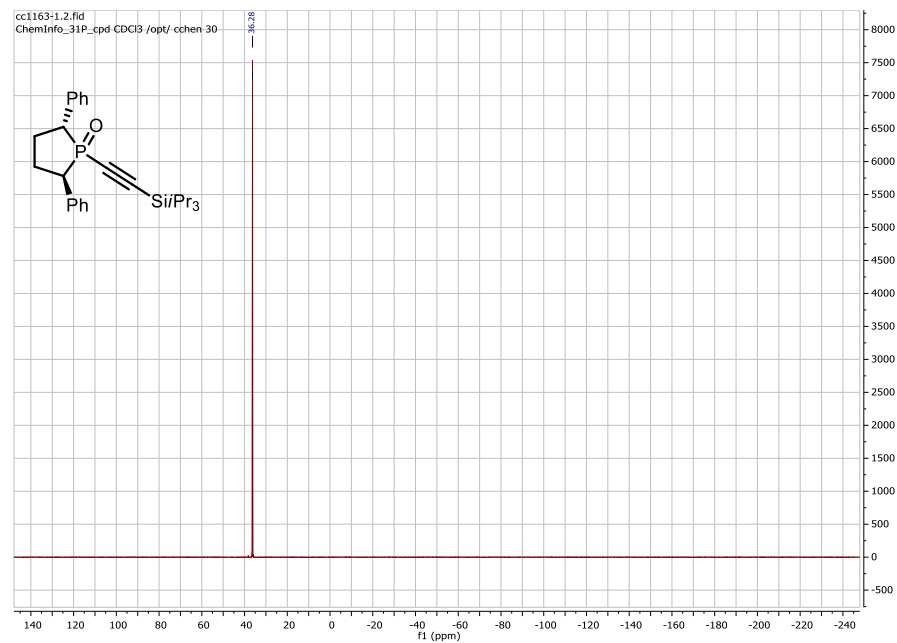
### <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) of compound 9f



### $^{13}\text{C}$ -NMR (101 MHz, $\text{CDCl}_3$ ) of compound **9f**



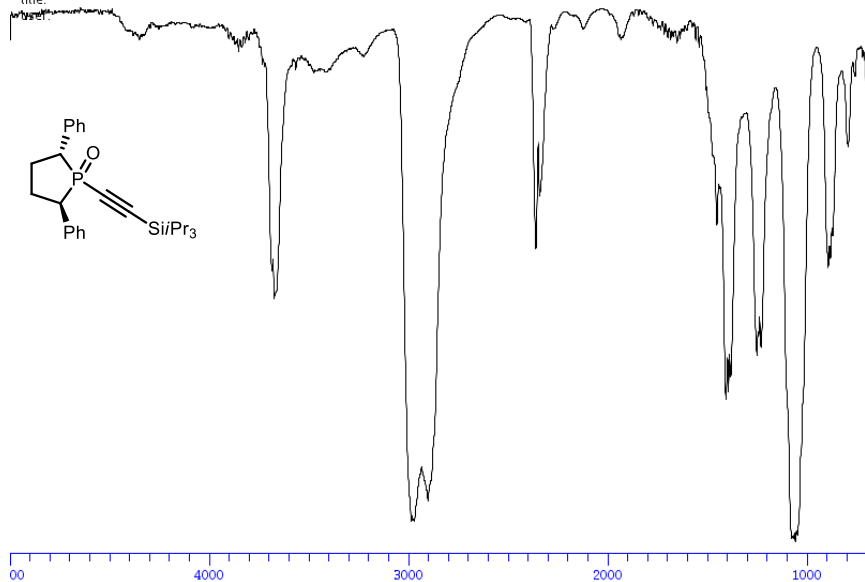
### $^{31}\text{P}$ -NMR (162 MHz, $\text{CDCl}_3$ ) of compound **9f**



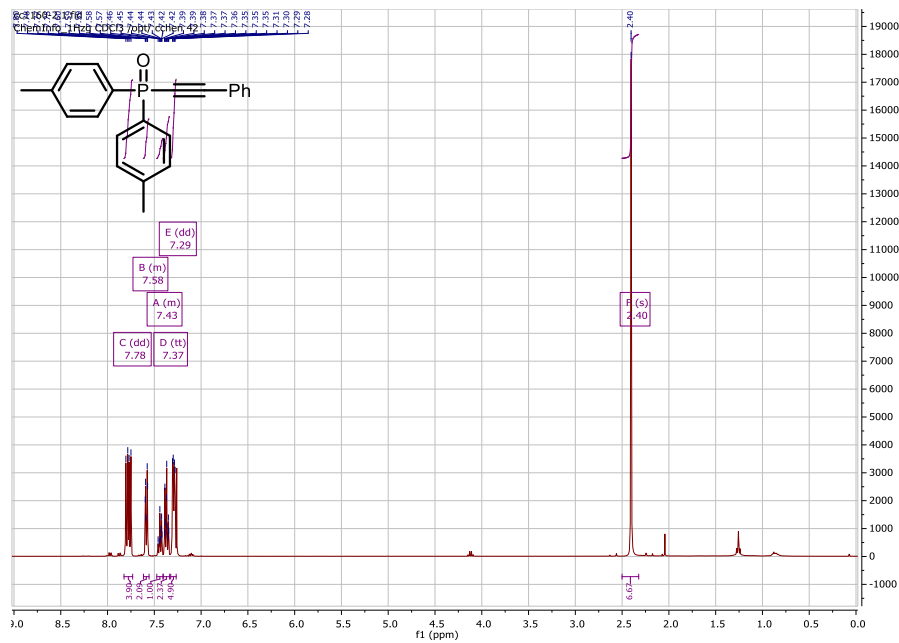
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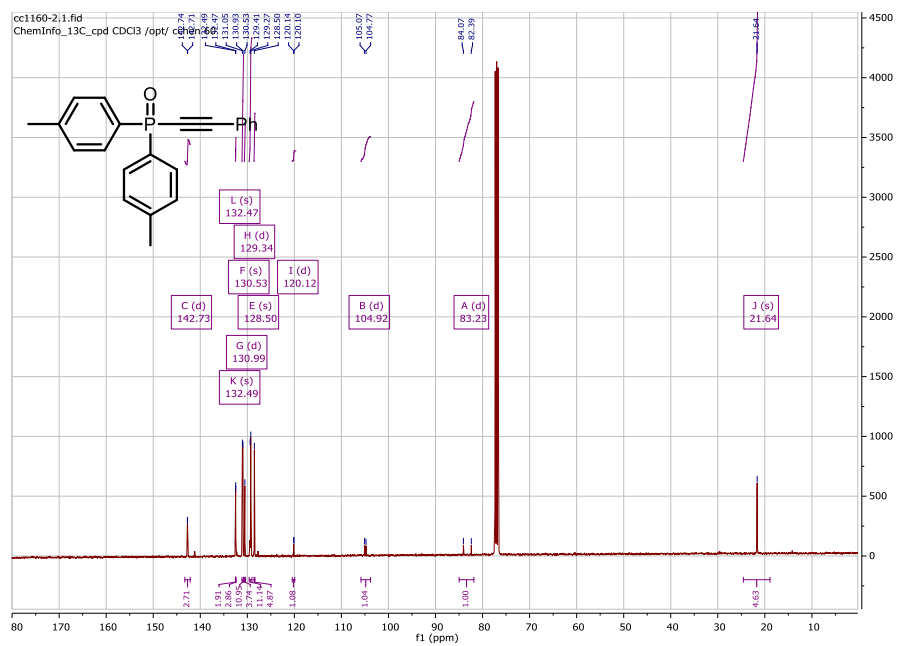
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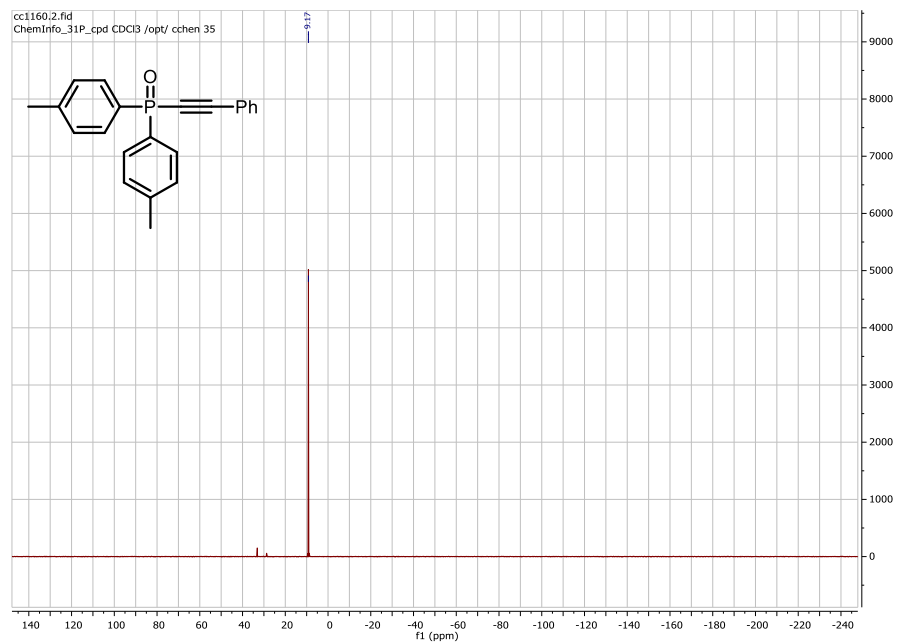
### <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) of compound 9i



### $^{13}\text{C}$ -NMR (101 MHz, $\text{CDCl}_3$ ) of compound **9i**



### $^{31}\text{P}$ -NMR (162 MHz, $\text{CDCl}_3$ ) of compound **9i**

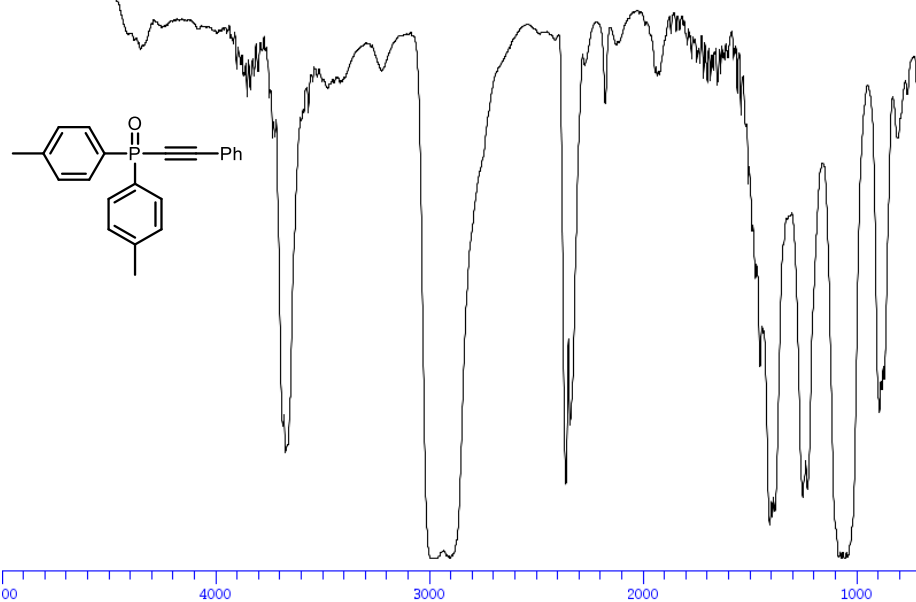


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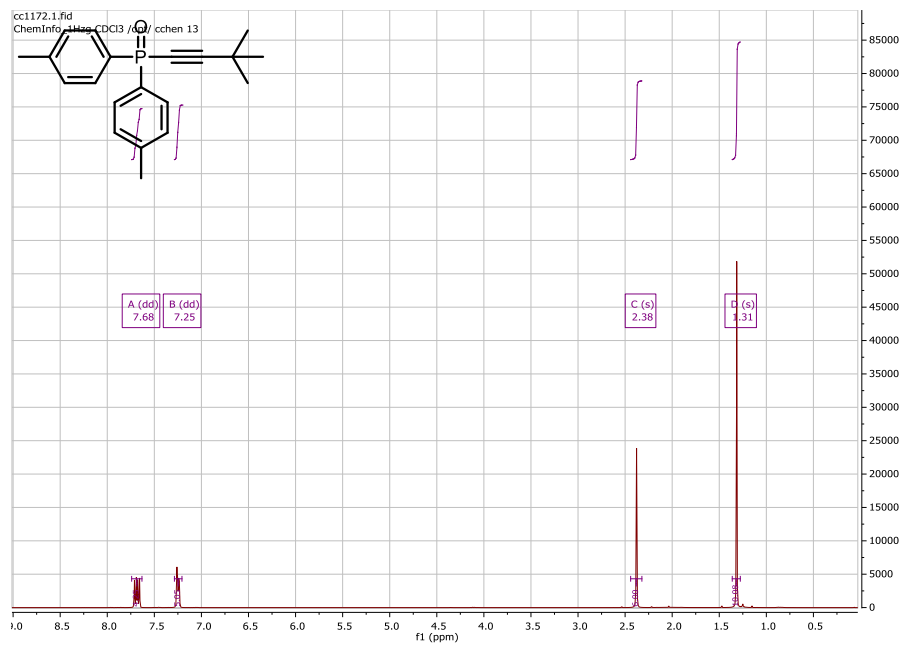
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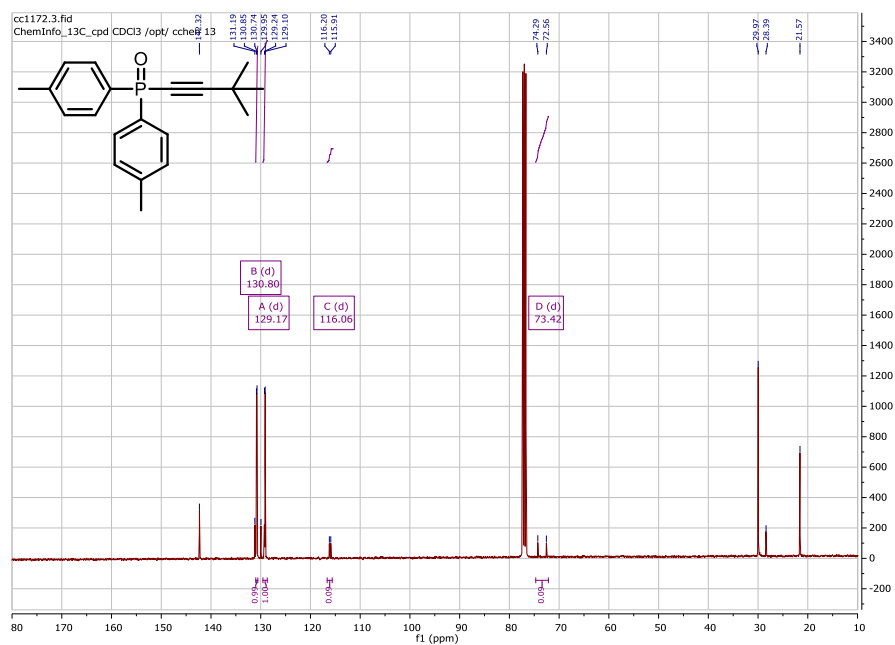
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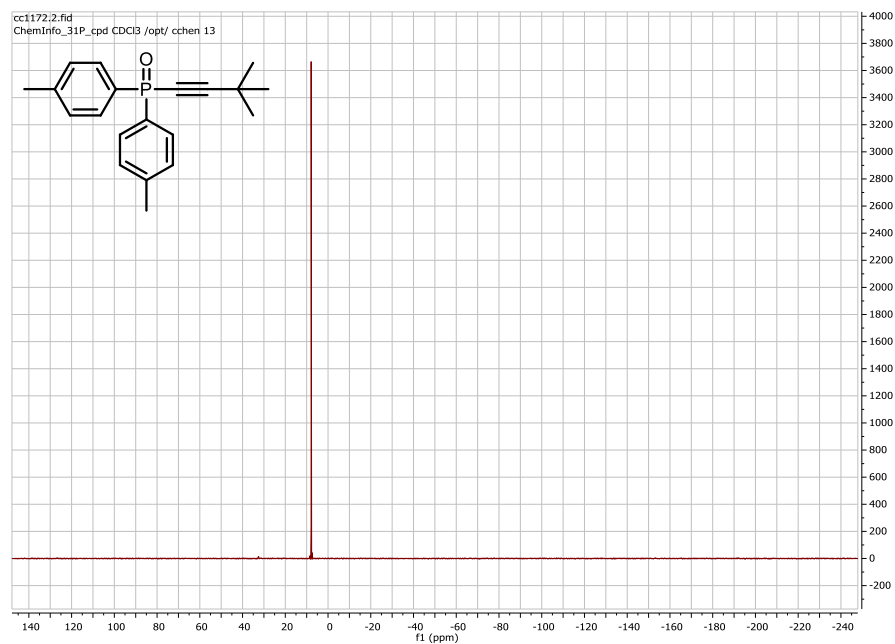
### <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) of compound 9j



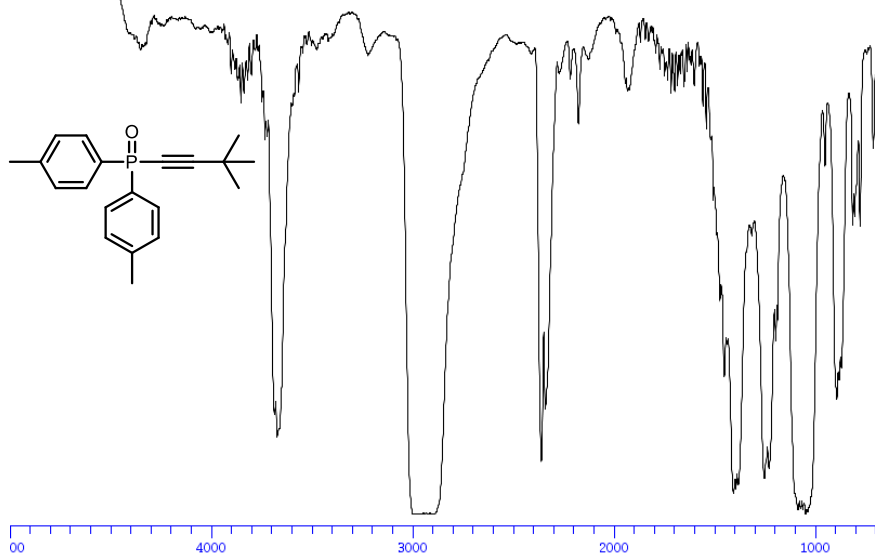
### $^{13}\text{C}$ -NMR (101 MHz, $\text{CDCl}_3$ ) of compound **9j**



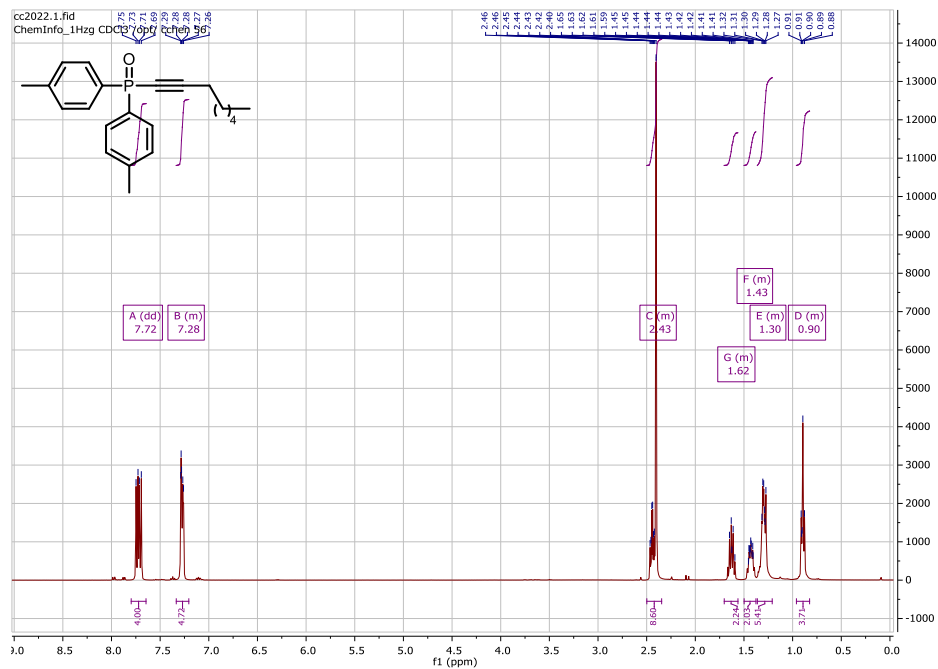
### $^{31}\text{P}$ -NMR (162 MHz, $\text{CDCl}_3$ ) of compound **9j**



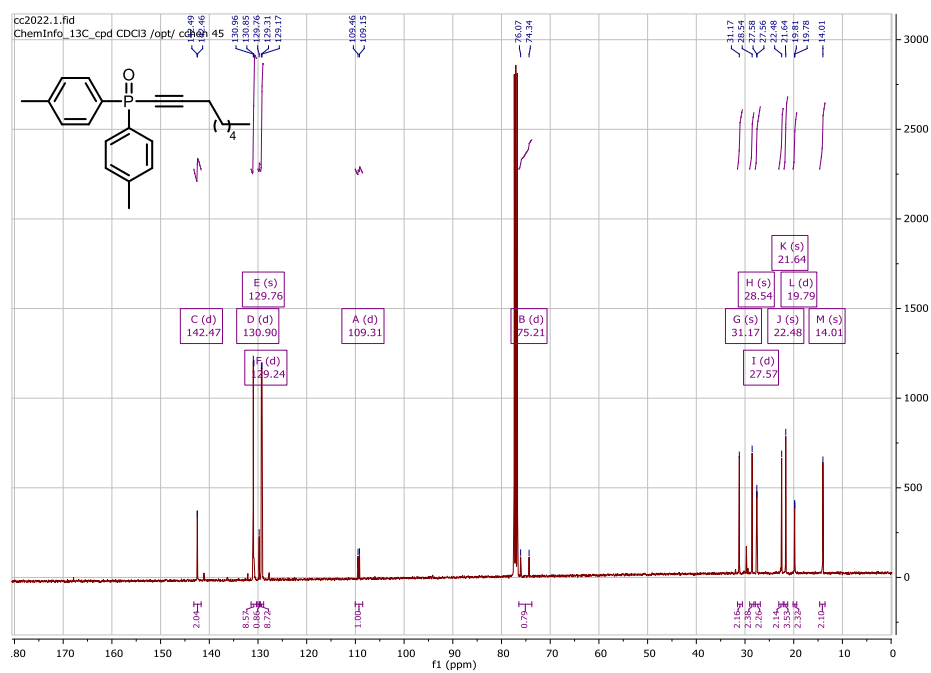
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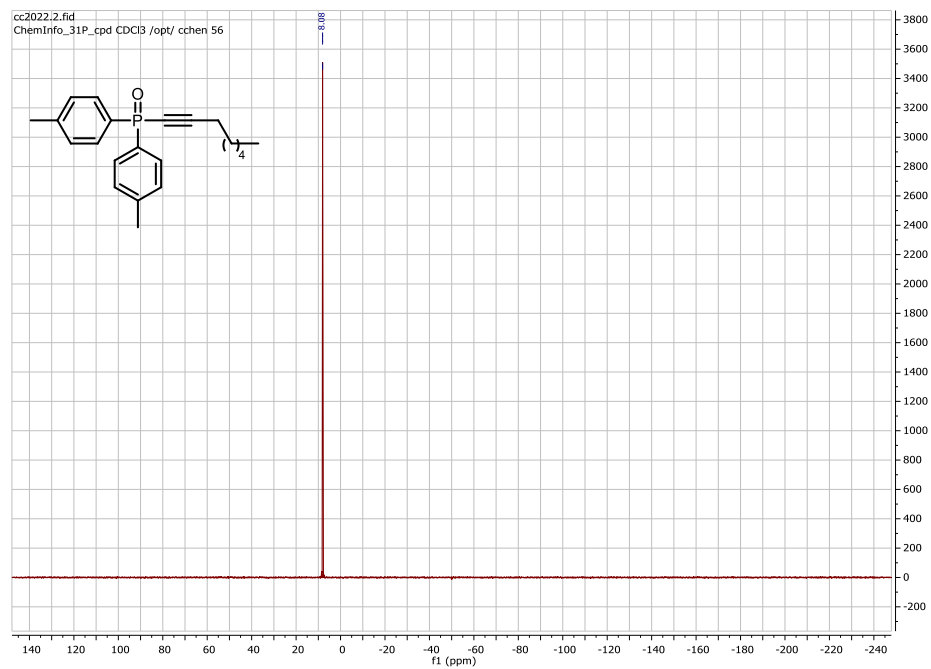
### <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) of compound 9k



### <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>) of compound **9k**



### <sup>31</sup>P-NMR (162 MHz, CDCl<sub>3</sub>) of compound **9k**





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