

Cite this: DOI: 10.1039/c2ra20559a

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COMMUNICATION

Design, testing and characterization of innovative TiN–TiO₂ surfaces inactivating bacteria under low intensity visible lightS. Rtimi,^a O. Baghriche,^a C. Pulgarin,^{*a} R. Sanjines^b and J. Kiwi^{*c}

Received 27th March 2012, Accepted 21st July 2012

DOI: 10.1039/c2ra20559a

Ti was sputtered in a plasma chamber under a N₂ atmosphere, depositing TiN films on polyester fibers. These films show a significant adsorption in the visible spectral region. A TiN layer 50 nm thick sputtered for 3 min under low intensity/actinic visible light led to the fastest bacterial inactivation (120 min). These innovative TiN nanoparticulate films were characterized by XPS, DRS and TEM.

Introduction

Sputtered TiN films are widely used as protective layers with high chemical resistance to corrosion/oxidation in the electrical and machinery/tool industry.¹ P. Kelly has recently reported TiN and other composite nitride structures with Ag are able to inactivate Gram-negative and Gram-positive bacteria in the dark.² Ag and Cu have also been sputtered on TiN rig metals in an Ar–N₂ atmosphere.³ Our laboratory has very recently reported the antibacterial activity of sputtered ZrN nanoparticulate films on polyester in the dark. A 15 min sputtered ZrN-sample led to bacterial inactivation within ~8 h.⁴

In the last decade, there has been increased interest in innovative antibacterial coatings due to the increasing resistance of pathogenic bacteria to antibiotics. Ag-films present acceptable adhesion and adequate bacterial inactivation kinetics along with favourable Ag-nanoparticle cell compatibility.^{5,6} But when washing textiles, some Ag leaches out and becomes an environmental problem.^{7,8} To avoid this environmental problem we investigated the bacterial inactivation by TiN films in this study since they do not contain Ag. Different antibacterial surfaces have been reported on textiles, glass, prostheses and catheters.^{9–13} Our laboratory has achieved bacterial inactivation on TiO₂,^{14–16} Cu and Ag films.^{17–20}

The objectives of this study are: a) to present the first report on the bactericide action of TiN-nanoparticle films activated by light in the visible range b) to explain the unexpected finding related the light

induced activity of TiN films by XPS and c) to report on the dependence of the *E. coli* inactivation kinetics on the intensity/dose of the commercial actinic lights. TiN films on hospital textiles as suggested in this study should negate the spreading of toxic bacteria when irradiated under visible or actinic light.

Experimental section

TiN was deposited onto polyester at room temperature. The polyester samples were 2 × 2 cm² in size. Before the deposition of the films, the residual pressure P_r in the sputtering chamber was typically $P_r \leq 10^{-4}$ Pa. The substrate-to-target distance was fixed at 9.5 cm. The TiN thin films were deposited by reactive DC magnetron sputtering (DC) using a 5 cm diameter Ti target 99.99 atomic% (Kurt J. Lesker, East Sussex, UK) in an Ar + N₂ atmosphere. The total working pressure $P_T = (P_{Ar} + P_{N_2})$ was fixed at 0.5 Pa and the ratio $P_{N_2}/P_T = 4.5\%$. The applied sputtering current on the Ti target was fixed at 250 mA providing a power of 112 Watt ($U = -450$ V) and a current density of 12.7 mA cm⁻².

The TiN film thickness was determined with a profilometer (Alphastep500, TENCOR). A film thickness of 50 nm was found for TiN-polyester after 3 min. Knowing the atomic distances of ~0.3 nm we can estimate that one layer is 10¹⁵ atoms cm⁻². The thickness of each layer is ~0.2 nm thick,²¹ then it follows that for a layer 50 nm thick, the amount sputtered is 2.5 × 10¹⁷ atoms cm⁻².

The polyester used corresponds to the EMPA test cloth sample No 407 Dacron polyethylene-terephthalate, type 54 spun, plain weave ISO 105-F04 used for color fastness determinations. The Ti sputtered on the polyester was evaluated by X-ray fluorescence (RFX, PANalytical PW2400).

The samples of *Escherichia coli* (*E. coli* K12) were obtained from the Zellkulturen GmbH (DSMZ) Braunschweig, Germany. The bacterial survival determination method has been recently reported.^{18,19} To verify that no re-growth of *E. coli* occurs after total inactivation, the TiN film was incubated for 24 h at 37 °C. Then bacterial suspensions of 100 microliters were deposited on 3 Petri dishes to obtain replica samples and the samples were incubated at 37 °C for 24 h. No bacterial re-growth was observed.

The irradiation of the polyester samples was carried out in a cavity provided with tubular Osram Lumilux 18W/827 actinic lamps. These lamps have a visible emission spectrum between 400 and 700 nm with an integral output of 1.1 mW cm⁻² resembling the light

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distribution found in solar irradiation. The bacterial inactivation kinetics is reported for diverse light intensities for both lamps.

XPS was carried out on an AXIS NOVA photoelectron spectrometer (Kratos Analytical, Manchester, UK). The surface atomic concentration was determined from the peak areas using sensitivity factors.²² The spectrum background was subtracted according to Shirley.²³ The XPS peaks of the Ti-species were analyzed by spectra deconvolution software (CasaXPS-Vision 2, Kratos Analytical UK).

Diffuse reflectance spectroscopy was carried out using a Perkin Elmer Lambda 900 UV-VIS-NIR spectrometer provided for with a PELA-1000 accessory within the wavelength range of 200–800 nm and a resolution of one nm. The absorption of the samples was plotted in Kubelka-Munk (KM) arbitrary units vs. wavelength.

A Philips CM-12 (field emission gun, 300 kV, 0.17 nm resolution) microscope at 120 kV was used to measure the grain size of the TiN-films. The textiles were embedded in epoxy resin 45 359 Fluka and the fabrics were cross-sectioned with an ultramicrotome (Ultracut E) and at a knife angle at 35°.

Results and discussion

Fig. 1a shows the inactivation of *E. coli* in the dark and under light by TiN sputtered polyester samples. *E. coli* inactivation occurred within 120–140 min. The formation of TiO₂ can be understood in terms of: a) the partial oxidation of TiN takes place in the presence of an oxygen source due to residual H₂O vapor in the sputtering chamber at the residual pressure $P_r = 10^{-4}$ Pa. This pressure is representative of about 10^{15} molecules cm⁻², there are sufficient

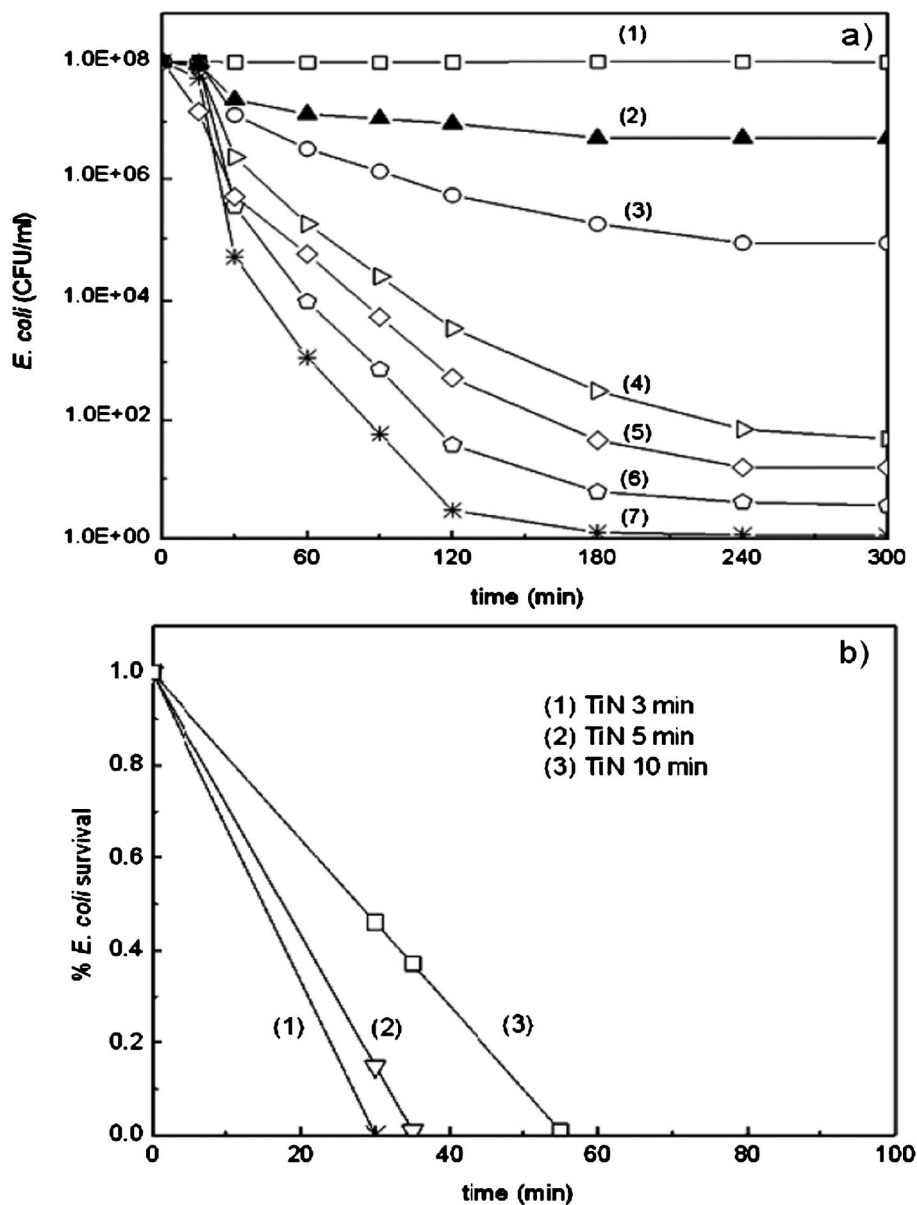


Fig. 1 *E. coli* survival on TiN-polyester sputtered for different times: (1) polyester alone, (2) TiN, 3 min in dark, (3) TiN, 1 min, (4) TiN, 1.5 min, (5) TiN, 10 min, (6) TiN, 5 min and (7) TiN, 3 min, irradiated with an Osram light (400–700 nm) L18W/827 (4 mW/cm²). Fig. 1b shows the *E. coli* survival percentage for TiN samples sputtered for different times, leading to a bacterial reduction of 3log₁₀.

O-radicals available to induce partial oxidation of TiN films^{19,21} and b) the films readily oxidize after deposition when exposed to air and during the sterilization process when autoclaving at 121 °C. If TiN films present an inherent photocatalytic activity, the photocatalysis by TiO₂ is not essential for bacterial inactivation.

Fig. 1a shows a progressively faster bacterial inactivation as the sputtering time increases from 1 to 3 min (traces 2, 3, 7). The amount of generated charges increases with a growing number of TiN layers up to a certain limit. At longer sputtering times, the bacterial inactivation in traces 5 and 6 (10 and 5 min respectively) slows down compared to the results presented in trace 7 (3 min). The increase in the TiN thickness hinders the free charge diffusion of carriers due to bulk inward diffusion.^{9,17} These charge carriers are responsible for electrostatic attraction with the bacteria. After 3 min sputtering time, the TiN coated polyester presents the highest amount of active sites/carriers in the TiN held in exposed positions leading to the shortest bacterial inactivation time. Fig. 1b shows the percentage decrease vs. time of the *E. coli* concentration for a bacterial reduction of 3log₁₀. Fig. 1b, trace 1 shows that a 3 min TiN sputtered sample is able to inactivate 99.9% *E. coli* within 30 min in a kinetically fast process.

Fig. 2 shows the bacterial inactivation kinetics mediated by TiN-polyester samples under light irradiation from a visible light source, Osram 18W/827. It is readily seen that the bacterial inactivation is strongly dependent on the applied light dose in the reactor cavity. The inset of Fig. 2 shows the spectral distribution of the Osram light source used.

Fig. 3a presents the evidence for TiO₂ (Ti 2p_{3/2} doublet) formation on the polyester when sputtering TiN for 3 min. The peaks in Fig. 3a assigned to TiN and Ti₂O₃ (Ti³⁺ in the net sense)²² have been deconvoluted by the program cited in the Experimental section. The TiN species shows a peak at 455.62 eV, the Ti³⁺ doublet is seen at 456.22 eV the Ti⁴⁺ doublet at 458.43 eV. Fig. 3b) presents the XPS deconvoluted spectra for the TiN sample at the end of the bacterial

inactivation process (120 min). The TiO₂ (Ti2p_{3/2} doublet) BE is seen to shift to 459.01 eV. Shifts ≥ 0.2 eV are due to changes in the oxidation states of the species.²³ This shift is due to a redox process taking place during bacterial inactivation involving the couple Ti⁴⁺/Ti³⁺ when oxidizing bacteria. The shift of the Ti₂O₃/Ti³⁺ doublet to 457.13 eV at 120 min reflects an increase in the reduced Ti³⁺-species at the end of the bacterial inactivation period. The surface atomic concentration of a TiN (3 min) sample at time zero was determined by XPS as: O1s 10.2%; Ti2p 44.7%; N1s 13.75% and C1s 22.31%.

Fig. 4 shows the diffuse reflection spectroscopy (DRS) of TiN sputtered for 1 and 3 min and for the polyester sample. The absorption of the TiN extends between 400 and 700 nm and a 0.34% Ti loading for the 3 min sputtered sample was determined by X-ray fluorescence spectroscopy (XRF).

Fig. 5 presents the TEM of: a) a polyester sample and b) a TiN sample sputtered on polyester for 3 min, presenting a width of 30–50 nm—equivalent to 150–250 TiN layers. By scanning transmission electron microscopy in high angular dark field imaging for TiN we found the N, Ti and O-atoms aligned in a continuous layer (data not shown). By X-ray diffraction (XRD) we attempted to determine the TiO₂ crystallographic phase sputtered on polyester. This was not possible, due to the small amount of TiO₂ sputtered on the polyester within 3 min.

Conclusions

This study presents the first report of TiN-surfaces as an effective bactericide photocatalyst when exposed to low intensity visible light. A 3 min TiN sputtered sample is able to inactivate 99.9% of *E. coli* in short times of up to 30 min. The magnitude of the optical absorption of TiN films runs parallel to the *E. coli* inactivation kinetics. Fig. 3 presents evidence for TiO₂ formation on the polyester when

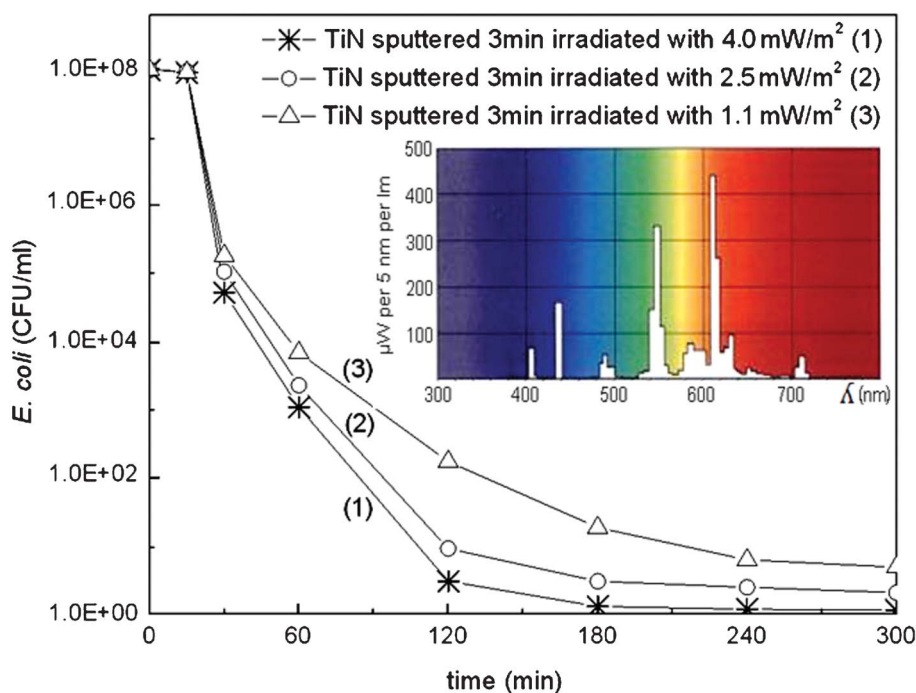


Fig. 2 *E. coli* survival on TiN polyester sputtered for 3 min and irradiated with a Lumilux Osram actinic light source (400–700 nm) 18W/827.

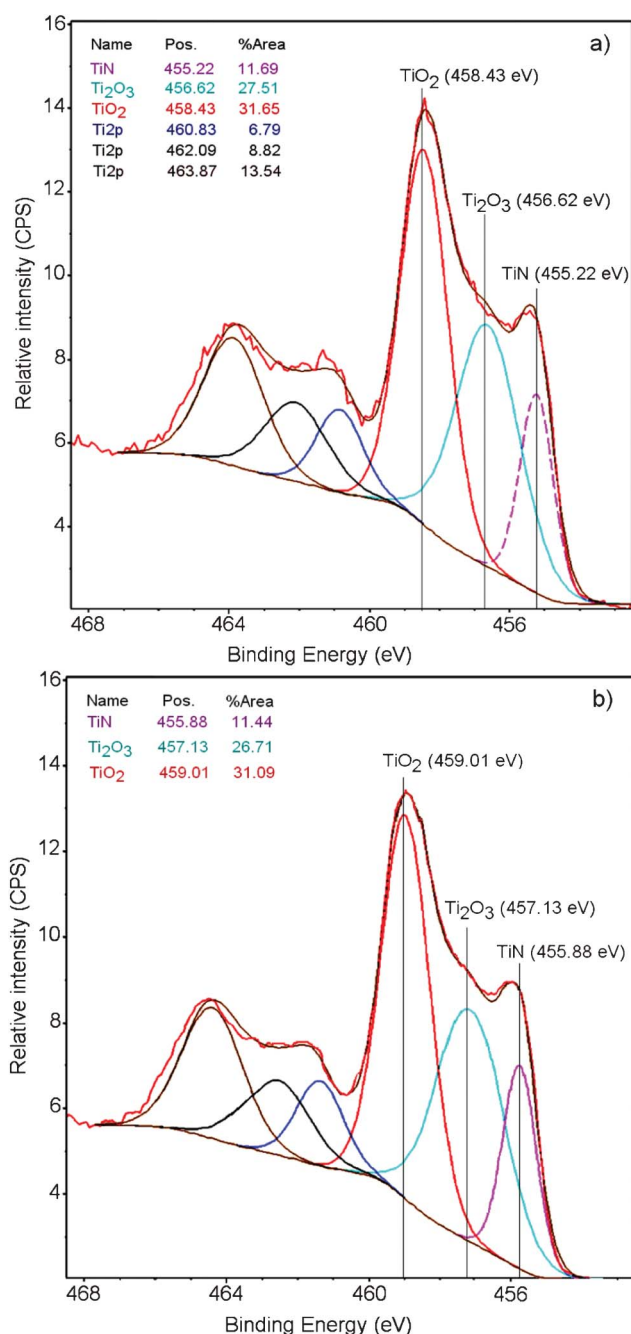


Fig. 3 X-ray photoelectron spectroscopy (XPS) of the TiN (3 min) in contact with bacteria for 3 s: a) at time = 0 min and b) at time = 120 min, showing the shift in the deconvoluted peaks after bacterial inactivation.

sputtering TiN on polyester and also for redox processes involving the Ti^{4+}/Ti^{3+} couple within the bacterial inactivation time.

Acknowledgements

We thank the COST Action MP0804 Highly Ionized Impulse Plasma Processes (HIPIMS) and the EPFL for support of this work.

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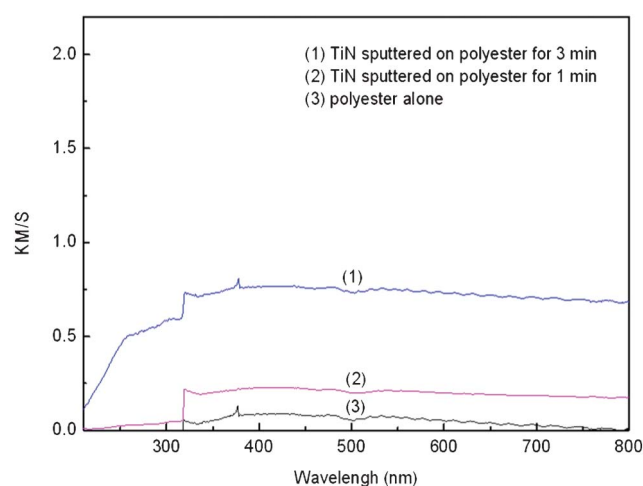


Fig. 4 DRS of the TiN sputtered on polyester for: (1) TiN 1 min; (2) 3 min, and (3) polyester alone.

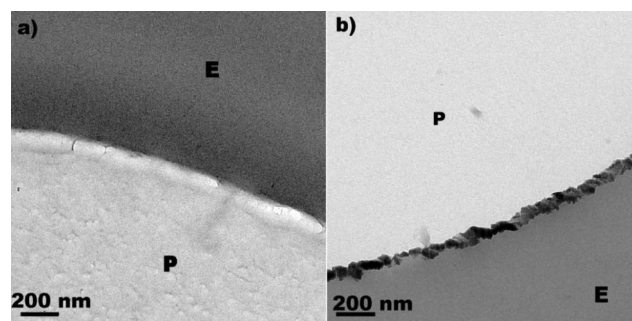


Fig. 5 Transmission electron microscopy (TEM) of a) polyester sample alone (E: epoxide, P: polyester, b) for a TiN (3 min) polyester sputtered sample.

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