

What is known about melatonin, chemotherapy and altered gene expression in breast cancer (Review)

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Abstract. Melatonin, synthesized in and released from the pineal gland, has been demonstrated by multiple *in vivo* and *in vitro* studies to have an oncostatic role in hormone-dependent tumors. Furthermore, several clinical trials point to melatonin as a promising adjuvant molecule to be considered for cancer treatment. In the past few years, evidence of a broader spectrum of action of melatonin as an antitumor agent has arisen; thus, melatonin appears to also have therapeutic effects in several types of hormone-independent cancer, including ovarian, leukemic, pancreatic, gastric and non-small cell lung carcinoma. In the present study, the latest findings regarding melatonin molecular actions when concomitantly administered with either radiotherapy or chemotherapy in cancer were reviewed, with a particular focus on hormone-dependent breast cancer. Finally, the present study discusses which direction should be followed in the next years to definitely clarify whether or not melatonin administration could protect against non-desirable effects (such as altered gene expression and post-translational protein modifications) caused by chemotherapy or radiotherapy treatments. As treatments move towards personalized medicine, comparative gene expression profiling with and without melatonin may be a powerful tool to better understand the antitumor effects of melatonin, the pineal gland hormone.

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1. Introduction: Chemotherapy for breast cancer

According to the World Cancer Research Fund International, breast cancer is the most frequent type of tumor suffered by women in the world, with ~1.7 million newly diagnosed cases in 2012 (1). Approximately 1 in 8 women will develop a mammary tumor during her lifetime. The American Cancer Society's report for the USA in 2015 informs of 231,840 new cases of invasive breast cancer, 60,290 women with carcinoma *in situ* and ~40,290 mortalities (2). In addition, breast cancer is the second cause of mortality by cancer in women, only exceeded by lung tumors (3). The mortality rates for breast cancer have been declining since 1989, particularly in premenopausal women, probably as a result of earlier detection as well as improved treatments (4). One fact that may also explain this decrease is the less frequent administration of hormone replacement therapy (HRT) following the publication of a report (the Women's Health Initiative) published in 2002, which suggests that HRT may be a risk factor that could explain the increase in the incidence of breast cancer (5).

Chemotherapy consists of treatment with cancer-killing drugs administered either intravenously or orally. Chemotherapy compounds are usually applied by intravenous infusion and, through the bloodstream, they reach growing cancer cells in almost all body tissues. Chemotherapy compounds work by targeting cells with a high rate of self-renewal, which is a hallmark of cancer cells (6). Chemotherapy is recommended following surgery (adjuvant chemotherapy): Surgery is performed to dissect the tumor, and adjuvant therapy is administered to try to eliminate any cancer cells that had not been removed by the surgery and may spread out later (7). The most frequently used adjuvant treatments to be administered subsequent to surgery are radiation, chemotherapy, targeted therapy and hormone therapy (8).

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1 Neoadjuvant therapy refers to treatments that are administered
2 prior to surgery instead of subsequent to surgery. The benefits
3 of neoadjuvant chemotherapy are that drugs may cause a
4 decrease in the size of the tumor, thus facilitating tumor
5 removal with a less extensive surgery (9). In addition, admin-
6 istering chemotherapy prior to the tumor being removed may
7 aid the subsequent monitoring of the disease, since in case the
8 first cocktail of drugs does not diminish the tumor size, other
9 compounds may be considered (10). Finally, chemotherapy
10 is also employed as a treatment strategy for patients whose
11 tumor has spread outside the mammary gland and underarm
12 area. Combinations of drugs are commonly used to treat
13 mammary tumors detected in the early stages of carcinogen-
14 esis, while advanced cancer is more commonly treated with
15 a unique chemotherapeutic molecule (11). Chemotherapy is
16 usually administered in cycles, with periods of administration
17 followed by resting periods to allow patient recovery and to
18 minimize the side effects of treatment (12).

19 Since microtubules participate in the migration of
20 chromosomes to opposite ends of mitotic cells during the
21 anaphase, microtubule inhibitors (MIs), also known as micro-
22 tubule-stabilizing agents, are molecules suitable to use in the
23 treatment of mammary tumors. MI agents include microtubule
24 depolymerizing compounds (*Vinca* alkaloids) and polymer-
25 izing agents (taxanes) (13). *Vinca* alkaloids derive from the
26 periwinkle plant *Catharanthus roseus* (14). The first clinical
27 trial demonstrating their efficacy in cancer was reported
28 in 1963 (15). Nowadays, these compounds are produced
29 synthetically and include vinblastine, vincristine, vindesine
30 and vinorelbine (14). The main mechanism that explains their
31 cytotoxicity is their capability to interfere with tubulin, with
32 subsequent microtubule function disruption (particularly
33 concerning microtubules implicated in the formation of
34 the mitotic spindle apparatus), leading to mitosis disruption
35 and finally resulting in metaphase arrest (16). These agents
36 interfere with the assembly of tubulin by introducing a wedge
37 between the contact surfaces of two tubulin molecules (17).

38 Taxanes are diterpenes obtained from *Taxus brevifolia* (18).
39 The first reported taxane, named taxol, was initially isolated in
40 1971 (19). Taxanes present difficulties in formulation because
41 they are poorly soluble in water, and for this reason, the first
42 clinical trial including taxanes was not reported until 1987 (20).
43 The other taxane currently in use is docetaxel, which is obtained
44 from *Taxus baccata* (21). Both paclitaxel and docetaxel act
45 as spindle poisons, stabilising the tubulin polymers against
46 depolymerisation. In addition, they also promote microtubulin
47 assembly. These two actions together block microtubule
48 dynamics and consequently lead to cell cycle arrest (22).
49 Taxanes induce changes in tubulin spatial conformation, which
50 interferes with the depolymerisation of microtubules in a
51 precise directional way, by binding a specific domain of tubulin
52 located in the internal surface of the microtubule (23,24).

53 Anthracyclines such as epirubicin and doxorubicin are
54 also commonly used in chemotherapy combined with other
55 chemotherapeutic drugs to treat breast cancer in patients who
56 have had surgery to remove the tumor (25). Their mechanism
57 of action is based on their ability of insertion between two
58 DNA strands, resulting in a DNA-anthracycline complex that
59 inhibits both DNA and RNA synthesis (26). This mechanism
60 also targets DNA for cleavage by topoisomerase II, which leads

to a cellular cascade that eventually results in cell death (27). 61
Epirubicin is frequently selected over doxorubicin in numerous 62
chemotherapy protocols, since it appears to have fewer side 63
effects (28). Eribulin is a new anthracycline approved by the 64
Food and Drug Administration of USA in 2010 to treat women 65
with metastatic tumors who had been previously treated with 66
at least two chemotherapeutic compounds indicated for the 67
treatment of metastatic breast cancer (29). This drug exerts 68
its anticancer effects by triggering a mitotic blockade, leading 69
cancer cells to enter apoptosis (30). 70

71 Other current approaches in the treatment of breast cancer
72 include compounds developed against specific identified
73 targets (molecular-targeted therapies) that contribute to tumor
74 growth (31). As an example, trastuzumab is a monoclonal antibody
75 used to treat patients with metastatic breast cancer. Trastuzumab
76 is indicated against tumors overexpressing the oncogene human
77 epidermal growth factor receptor 2 (HER2)/neu, since it targets
78 the membrane HER2/neu receptor, which normally promotes
79 normal cell growth and is also overexpressed in certain cancerous
80 breast tissues (32). Despite several randomized clinical trials with
81 promising results (33), other reports indicated that both *de novo*
82 and acquired resistance to trastuzumab could be developed (34).
83 Therefore, other recently described agents have been included
84 in the list of chemicals available to treat HER2-overexpressing
85 mammary tumors. One of them is lapatinib, a reversible inhibitor
86 of both epidermal growth factor receptor (EGFR) and HER2/neu
87 tyrosine protein-kinases (35), which was approved in 2007 for
88 women undergoing metastatic breast cancer with acquired
89 resistance to trastuzumab (36). Lapatinib was well tolerated and
90 displayed encouraging clinical results when used as a first-line
91 therapy agent in ErbB2-amplified tumors, either advanced local
92 tumors or metastatic breast cancer (37). Another molecule to be
93 considered is HKI-272, a (Her2)/neu receptor tyrosine kinase
94 inhibitor with an irreversible mechanism of inactivation and a
95 demonstrated clinical activity, which is well tolerated among
96 both high-dose trastuzumab pre-treated and non pre-treated
97 patients with advanced ErbB2-positive mammary tumors (38).
98 HKI-272 targets a cysteine residue located in the adenosine
99 triphosphate-binding pocket of the ERbB2 receptor, resulting
100 in the inhibition of the downstream signal transduction cascade
101 triggered, and consequently altering the cell cycle regulation (39).

102 Among the battery of promising new chemicals, drugs that
103 target heat shock protein (Hsp) 90 must be mentioned. Hsp90
104 belongs to the family of chaperones and establishes associations
105 with a set of different proteins that are known as 'Hsp90 client
106 proteins' (40). Multiple Hsp90 client proteins are implicated
107 in breast tumor progression and resistance to chemotherapy
108 treatments, including the receptor protein-tyrosine kinases of
109 the ErbB2 family, estrogen receptor (ER), Akt and mutated
110 versions of p53 (41). The efficacy of Hsp90 inhibitors has been
111 well documented in several preclinical cancer models. One
112 of these inhibitors is 17-AAG, which has completed phase I
113 testing (42,43).

114 Angiogenesis is a physiological process that consists
115 in new vessel formation. When a tumorous mass of cells is
116 growing, angiogenesis is crucial to maintain both tumor
117 growth and progression (44). Therefore, numerous drugs have
118 been tested during the past decades in the hope of identifying
119 specific inhibitors of the different pathways necessary for
120 angiogenesis (45). One of the key molecules in the formation

of new vessels is vascular endothelial growth factor (VEGF), probably the most studied angiogenic factor (46). VEGF is implicated in the progression of breast cancer and is also a potential prognosis biomarker (47). Bevacizumab (Avastin®) is a recombinant humanized monoclonal antibody that recognises all the known variants of VEGF-A (48). To date, bevacizumab is the unique anti-angiogenic chemical with clearly demonstrated benefits in metastatic breast cancer treatment clinical trials (49,50). Pazopanib is another inhibitor of VEGF receptor (VEGR) (51). Pazopanib also inhibits the signaling pathways downstream of the platelet-derived growth factor receptor and the mast/stem cell growth factor receptor c-KIT (52). Pazopanib treatment provides disease stability in patients with advanced breast cancer (53).

Estrogens are implicated in the development of the mammary gland, and are also known to be key stimulators of both the genesis and growth of mammary tumors (54). Therefore, one of the main strategies to fight breast cancer is neutralizing the stimulating actions of estrogens on the mammary gland (55). Several chemicals have been tested and commercialized, since they are selective inhibitors of the effects of estradiol on the breast. These include selective estrogen receptor modulators (SERMs), which are chemicals that directly bind to ER, thus inhibiting its actions by interfering with the binding of endogenous estrogens (56). Of these, tamoxifen (57,58) and a number of its derivatives (56) are the best known examples. Fulvestrant is an ER antagonist that has no agonist effect described and downregulates the protein levels of ER α (59). Fulvestrant is being currently administered to postmenopausal women with advanced breast cancer whose tumors are ER positive and have progressed upon receiving first-line endocrine therapy (60). Other compounds developed against estrogens are chemicals that prevent the production of steroids by downregulating the enzymes necessary for the conversion into estradiol from androgenic precursors. These drugs belong to the class known as selective estrogen enzyme modulators (SEEMs), which include both steroidal (such as formestane or exemestane) and non-steroidal (such as anastrozole and letrozole) compounds (61).

2. Melatonin and mammary cancer: *In vitro* and animal studies

Melatonin is an indolic hormone produced principally by the pineal gland. Melatonin is a ubiquitously distributed molecule with a variety of diverse functions (62). Melatonin employs a diverse set of mechanisms to regulate the physiology and molecular biology of cells (63). The majority of actions of melatonin are based on its ability to bind to melatonin membrane receptors, which are G-protein coupled receptors that trigger cellular signaling pathways (64). The pineal hormone also acts through orphan receptors or molecules such as calmodulin (65). Additionally, melatonin can detoxify free radicals and related oxygen derivatives via receptor-independent pathways (66).

Concerning tumorigenesis, numerous studies have been performed in animal models. Experimental approaches include increasing the activity of pinealocytes and administering exogenous melatonin, which cause a decrease in the number, incidence and development of chemically-induced mammary tumors (67). However, reduced levels of melatonin (for example, by removing the pineal gland) appear to stimulate breast

cancer progression (68). Several reports established a lower cancer risk among totally blind women (69-71). By contrary, a moderate but significant increase in the risk of developing breast cancer among women who have been working for long periods in rotating night shifts (which implicates that they were exposed to light during the night, and consequently, the nocturnal melatonin production was inhibited) has also been documented (72).

The antiproliferative effects of melatonin on the breast cancer cell line MCF-7 have been studied for more than two decades (73). The data available suggest that the inhibitory action of melatonin on mammary cancer estrogen-positive cell lines is based on its ability to regulate either the synthesis of estrogens or estrogen signaling pathways (74). Thus, the pineal hormone is capable of downregulating both the expression and activity of the enzymes necessary for the synthesis of estrogens from androgenic precursors, therefore acting as a SEEM. In the MCF-7 cell line, melatonin at physiological concentrations exhibits anti-aromatase properties (75). The pineal hormone is able to reduce the activity of aromatase, the principal enzyme in estrogen biosynthesis. Melatonin inhibits aromatase under basal conditions or when the enzyme activity is stimulated by cyclic adenosine monophosphate (cAMP) or cortisol (76). When gene expression was evaluated and the CYP19 gene (coding for aromatase) was examined, it was observed that melatonin downregulated its expression at the transcriptional level (76). The major bloodstream circulating form of physiologically inactive estrogen is estrone sulfate, which acts as an estrogen reserve (77). The enzyme steroid sulfatase (STS) converts inactive estrogen sulfates into estrone and estradiol. Estrone can be further transformed into physiologically active estrogen by the action of 17 β -hydroxysteroid dehydrogenase type 1 (17 β -HSD1). Finally, the enzyme estrogen sulfotransferase (EST) sulfonates estrogens to form biologically inactive estrogen sulfates. Both enzymes, STS and EST, serve a role in the modulation of the *in situ* levels of estradiol in hormone-dependent tumors (78). Melatonin modulates the expression and activity of aromatase, STS, 17 β -HSD1 and EST not only in tumor cells, but also in surrounding cells such as fibroblasts and endothelial cells (74,79-82).

Melatonin can also counteract the different actions of estrogens, thus functioning as a naturally occurring SERM (83). The mechanisms implicated in the antiestrogenic effects of melatonin are yet being elucidated. Unlike other antiestrogenic molecules such as tamoxifen, melatonin does not directly bind to ER (84). In estrogen-positive breast cancer cells, melatonin decrease the expression of ER α (85) and impairs the estrogen-mediated transcriptional activation of genes through destabilization of the estradiol-ER complex, preventing its binding to DNA in both estrogen response element (ERE)- and activator protein 1-containing promoters (86). This effect appears to be mediated by calmodulin, since melatonin behaves as a calmodulin antagonist. The pineal hormone promotes structural changes in the calmodulin-ER α protein complex, thus facilitating its binding to an ERE (87,88). Remarkably, melatonin does not alter the recruitment of co-activators triggered by ER α , suggesting that melatonin mechanisms of action diverge from those of other anti-estrogen chemicals used in breast cancer treatment (88). It is important to mention that only ER α , but not ER β , binds to calmodulin (84). The substitution for glycine of two lysine

residues located at positions 302 and 303 of the hinge domain of ER α generated a mutant version of ER α that was incapable of binding to calmodulin, and that therefore turned into a melatonin non-regulated receptor (88). The effects of melatonin may also be explained in terms of binding to its specific membrane receptors, such as melatonin receptor type 1 (MT1), resulting in an interplay with the ER signaling pathway (89). MT1 receptors are present in normal human breast tissues and in tumor tissues (90). Melatonin and estradiol signaling pathways converge and they have opposite effects over cAMP intracellular concentrations. In breast cancer cells, estrogens trigger adenylate cyclase activation, which results in increased cAMP cytoplasmic levels in a classical short-time second-messenger mechanism that is independent of transcription (91). The increase in cAMP levels cooperates with long-time genomic effects of estradiol, thus enhancing ER-mediated transcriptional activation (91). By contrary, melatonin, through its specific binding to its membrane receptor MT1, inactivates adenylate cyclase, resulting in decreased cAMP levels (92).

The fact that only those human mammary cancer cell lines that are ER α^+ are sensitive to the antimetabolic actions of melatonin supports the theory that the antitumor effects of this indolamine occur through its actions on breast cancer cells' estrogen-responsive pathways (93). Melatonin is able to block, under different culture conditions, the mitogenic effects of estradiol (94). The antiproliferative effect of this indolamine could be explained through the modification of the levels of estrogen-modulated proteins, several growth factors and proto-oncogenes such as cMYC, transforming growth factor (TGF) α , Trefol factor 1, also known as pS2, progesterone receptor (PGR), AP1 transcription factor subunit c-fos and TGF β in human estrogen-positive breast cancer cell lines (94,95). Estradiol enhances cell proliferation and provokes cell cycle progression (96). The inhibitory effect of melatonin (as occurs for tamoxifen) on cell proliferation is cell-cycle specific, causing the presence of melatonin a delay in the G₁-S transition (97). Changes in cell-cycle timing typically implicate modifications in various key proteins that regulate the process. The inhibitory action of the pineal hormone on cell cycle progression can be interpreted through its effects on the expression of certain proteins controlling the G₁-S cell cycle transition. Thus, several studies have demonstrated that melatonin increases the expression of p53 and p21^{Waf1} in experiments performed *in vitro* (98,99). The upregulation of these proteins may be a crucial mechanism explaining how melatonin impedes the progression through the cell cycle at the G₁-S transition. The accumulation of cells in G₁ forces them to enter G₀, causing the cancer cells to undergo a higher differentiation, since G₀ is characterized to be a quiescent state (100). This suggest that the anti-estrogenic, oncostatic and antiproliferative effects of melatonin on human mammary cancer estrogen-positive cell lines may be explained, at least in part, by the ability of the pineal hormone to inhibit cell proliferation at the same time that it enhances cell differentiation. Furthermore, several studies have demonstrated that melatonin inhibits human telomerase reverse transcriptase, which is the rate-limiting factor conditioning the telomerase activity in breast cancer cells (101,102).

Another effect of melatonin in MCF-7 cells is its ability to reduce their invasiveness (103,104). Melatonin

treatment decreases the attachment of the cells to the basement membrane (105). The pineal hormone reduces the chemotactic response of MCF-7 cells (105). Melatonin also blocks the cell migration and invasion that occurs in response to estradiol (105). Cancer cells' motility and invasion are known for being adhesion-dependent mechanisms that require the expression of cell-surface molecules necessary for adhesion (106). In tumor progression, downregulation or loss of expression of several of these surface-adhesion proteins frequently happens, which leads to the loss of cell-cell recognition and the acquisition of an invasive phenotype by the tumor cells; these events correlate with poor cell differentiation (107). All the factors mentioned above are associated with poor prognosis in cancer progression (108). Importantly, among the melatonin antitumor actions, it has been reported that melatonin induces the expression of both β_1 -integrin and E-cadherin, two main proteins essential for cell-cell and cell-matrix interactions; thus, melatonin changes estrogen-responsive tumor cells into a less invasive phenotype by inducing their differentiation (105).

Finally, melatonin exerts its modulatory effect in the tumor microenvironment by controlling the production and secretion of several cytokines. These cytokines are produced by breast cancer cells and regulate the differentiation of the fibroblasts located in close proximity to malignant epithelial cells. Additionally, it has been demonstrated that cytokines produced by malignant cells stimulate the aromatase expression and activity in these fibroblasts (109,110) and in proximal endothelial cells (111). VEGF is a growth factor that serves an essential role in angiogenesis. VEGF is produced and secreted by malignant epithelial cells and recognizes VEGFRs located in the cell surface of endothelial cells. The binding of VEGF to its receptor triggers a cascade of intracellular events that stimulate endothelial cells to undergo proliferation and migration (112,113). Therefore, pharmacological agents able to inhibit the production of this pro-angiogenic factor are of great interest, and could serve an essential role in impairing both tumor angiogenesis and tumor growth. Melatonin may be one molecule to consider, since it can regulate the paracrine mechanisms connecting tumor epithelial cells and the surrounding endothelial cells. One of the main actions of the pineal hormone is that melatonin treatment results in the downregulation of VEGF expression in estrogen-responsive breast cancer cells. As a result, the VEGF levels available for receptors expressed in endothelial cells are lower, and therefore, the number of cells producing estrogens in the proximity of the malignant cells is reduced. Reduced estrogen levels and a lower capability of formation of new vessels as a result of the presence of melatonin will diminish the tumor ability to spread and grow (114).

In summary, a unique molecule, melatonin, has anti-estrogenic properties: It selectively counterbalances the actions of estrogens in both normal and tumor breast tissues, and provides a novel strategy to reduce the local biosynthesis of estrogens from androgens (which in turn, is one of the principal objectives of antitumor pharmacological therapy) (115). These cumulative actions of the pineal hormone point to its potential application as an anticancer molecule in both the prevention and treatment of estrogen-positive tumors, since, as it has been pointed above, this molecule acts at different levels by

1 interfering with estradiol-dependent signaling pathways, both
2 in tumor cells and in the surrounding endothelial cells and
3 fibroblasts (116).

4 3. Melatonin and cancer: Clinical trials

5
6
7 As aforementioned, numerous experiments performed *in vitro*
8 (breast cancer cell lines such as MCF-7) and *in vivo* (animal
9 models) have well established the oncostatic properties of
10 melatonin (117). Since melatonin plasma levels are diminished
11 in estrogen-dependent breast cancer patients, various clinical
12 trials have been performed to evaluate the potential beneficial
13 effects of melatonin in human neoplasms. Following the pioneer
14 clinical work of Lissoni *et al* (118), who evaluated the effect of
15 melatonin in cancer patients bearing untreatable advanced solid
16 tumors, multiple studies have been performed and published to
17 date (119). However, the value of melatonin as an adjuvant agent
18 in cancer treatment is not totally clear, and consensus about
19 positive melatonin actions appears to be difficult to achieve.
20 A number of studies point to melatonin as a treatment with no
21 beneficial effects. In cachectic patients with gastrointestinal or
22 advanced lung cancer, including a dose of melatonin at night
23 did not improve parameters such as appetite, weight or quality
24 of life of patients, in comparison with patients who received
25 a placebo (120). In patients with brain metastases, high doses
26 of melatonin did not produce any beneficial effect (121). By
27 contrast, there are a large number of reports supporting the
28 potential benefits of melatonin if included in chemotherapy
29 protocols. In breast, lung and gastrointestinal cancer patients,
30 melatonin protected against thrombocytopenia, and stomatitis,
31 asthenia and neuropathy were less recurrent in the group of
32 melatonin-treated patients (122). It has also been reported that
33 melatonin offers certain protection to hematopoietic progeni-
34 tors from the toxic actions of anticancer chemotherapeutic
35 chemicals; thus, melatonin has been reported to attenuate the
36 damage to precursor blood cells caused by both radiotherapy
37 and chemotherapy treatments (123). It has also been suggested
38 that melatonin may protect patients against side effects such as
39 asthenia, cardiotoxicity and neurotoxicity caused by chemo-
40 therapy (124). Additionally, the pineal hormone increases the
41 1-year survival and tumor regression rates in cancer patients
42 with metastatic solid tumors with poor clinical status (125).
43 In metastatic non-small cell lung cancer patients treated either
44 with just chemotherapeutic agents or with chemotherapy plus
45 melatonin, both the overall tumor regression rate and the 5-year
46 survival rate of patients concomitantly receiving melatonin
47 were significantly higher as compared with those receiving
48 only chemotherapeutic agents. It appears that chemotherapy
49 was better tolerated in patients who also received the pineal
50 hormone (126). The study points to melatonin as an adjuvant
51 drug capable of improving the effectiveness of chemotherapy
52 in terms of both quality of life and survival of patients (126). A
53 recent report concludes that melatonin combined with chemo-
54 therapy did not significantly improved survival or ameliorated
55 various adverse side effects in patients with non-small cell lung
56 advanced cancer, although certain improvement in the quality
57 of life of these patients was observed (127). A systematic
58 review comprising data from 21 clinical trials (all the patients
59 enrolled in the studies were bearing solid tumors), in which the
60 effect of melatonin concomitantly added in conjunction with

61 chemotherapy or radiotherapy was evaluated, and supportive
62 care, partial response, complete response, 1-year survival
63 and chemotherapy-associated toxicities were assessed (128),
64 concluded that melatonin may serve a beneficial role in cancer
65 patients who are treated with chemotherapy. Patients who
66 received melatonin experienced substantial improvements,
67 particularly in terms of tumor remission and 1-year survival
68 rates and melatonin also ameliorated the side effects of
69 chemotherapy (127,128). Furthermore, another review summa-
70 rizing the data from eight eligible randomized controlled trials
71 (n=761) obtained similar conclusions (129).

72 4. Can melatonin enhance the beneficial and protect 73 against the deleterious effects of chemotherapy?

74
75
76 As aforementioned, it is well documented that melatonin
77 diminishes the incidence of chemically induced cancers and is
78 able to slow down the growth of certain hormone-responsive
79 cancers (67,117). The antitumor actions of this indolamine
80 have been described in breast cancer, both in *in vivo* animal
81 experiments (in 7,12-dimethylbenz[a]anthracene chemically
82 induced mammary tumors in rodents) and in *in vitro* assays
83 (in estrogen-positive human mammary cancer cell lines) (64).
84 Furthermore, there are numerous reports endorsing the
85 beneficial use of melatonin during chemotherapy in clinical
86 trials (118,122-124). Therefore, the ultimate goal of the
87 present review is to provide a compilation on the current
88 knowledge concerning the interplay of melatonin and chemo-
89 therapy agents at the molecular level. The PubMed database
90 (www.ncbi.nlm.nih.gov/pubmed) was interrogated for cita-
91 tions of 84 genes known to be commonly involved in the
92 dysregulation of several normal processes during breast
93 carcinogenesis, which are also present in breast cancer
94 cell lines. The list includes signal transduction genes and
95 other genes involved in usually altered pathways, including
96 cellular adhesion, angiogenesis, proteolytic activities, cell
97 cycle progression, cell cycle control and apoptosis (Table I).
98 Research into carcinogenic mechanisms (130) has identified
99 during the last decades numerous functional alterations due
100 to somatic mutations, gene expression alterations and altered
101 post-translational protein modifications (131). Therefore,
102 the database search was performed including as keywords
103 i) the name of each gene, ii) breast cancer, iii) melatonin, and
104 iv) chemotherapy.

105 For the majority of genes reviewed, there were thousands
106 of citations in the literature when the search included the name
107 of each gene and breast cancer as key words. When melatonin
108 was included in the selection criteria, the number of articles
109 was markedly reduced; thus, 53 out of the 84 genes surveyed
110 did not have a single report associated under these criteria.
111 Searching for ER together with melatonin and breast cancer
112 led to ~100 articles, whereas p53 and PGR searches produced
113 15 and 12 reports respectively, and the rest of the 28 genes
114 evaluated produced 1-9 articles.

115 The number of reports was further reduced if chemo-
116 therapy was also included in the selection criteria. Only 25 of
117 the 84 genes assessed are included in the list (Table I), indi-
118 cating that, for 59 out of 84 genes, there is not a single study
119 published including the gene name, melatonin, breast cancer
120 and chemotherapy as keywords.

In Table II, a search was performed including as keywords melatonin (20,724 articles) melatonin and breast cancer (524 articles), and 28 drugs commonly used as chemotherapy in breast cancer treatment (6,9). Apart from tamoxifen (23 articles fulfilled the criteria ‘melatonin, breast cancer and tamoxifen’), the results show that there is limited information at the molecular level concerning the implication of co-treatment with the pineal indolamine and chemotherapy agents in breast cancer targeted therapy, and no report was available for 13 of the 28 drugs searched. In conclusion, despite the fact that during decades numerous articles have reported experimental data from *in vitro* and *in vivo* experiments showing the oncostatic actions of melatonin, the present review demonstrated that there are limited reports studying the effects of melatonin and chemotherapy agents in combination in cancer treatments, and particularly, in breast cancer. Similarly, little is known about the role of melatonin regarding the expression and functionality of the genes reported to be altered in cancer, particularly in breast cancer. There is little information available nowadays about gene expression profiles in all types of cancer, and particularly in estrogen-responsive breast tumors. There is also limited information available on how the gene expression profile may be altered by treatment with different chemotherapeutic compounds and, remarkably, whether or not melatonin has protective effects when administered together with chemotherapeutic agents. The majority of information available about melatonin, cancer, chemotherapy, and altered gene expression and function has been published (132,133) in the last few years (Fig. 1). This indicates that nowadays there is a growing field of research about this topic. Several of the most relevant findings recently reported with regard to the modulatory role of melatonin in cancer at the molecular level include the circadian interruption of melatonin production by exposure to light at night, which results in the development of resistance to tamoxifen treatment in breast cancer patients (134). In this context, it has also been described how nocturnal disruption of melatonin plasma levels originates a complete loss of tumor response to the chemotherapeutic agent doxorubicin (135). The resistance of cancer cells to chemotherapy treatments such as doxorubicin usually implicates an upregulation of P-glycoprotein, which is responsible for drug efflux from cells (136). In this context, there is a report describing that melatonin treatment increases doxorubicin intracellular concentrations in cancer cells, suggesting that melatonin may inhibit P-glycoprotein (137). In breast cancer xenografts implanted in animal models (nude mice), treatment with the pineal hormone stopped the tumor progression by reducing tumor size and cell proliferation (Ki-67), as well as by inhibiting angiogenesis (138). Melatonin treatment results in increased expression of Bcl-2-like protein 11 (Bim) parallel to lower levels of cyclooxygenase (COX)-2, which in turns potentiate tunicamycin-induced apoptosis in mammary cancer cells (139). Regulation of the COX-2, Akt, p300 and apoptotic protease activating factor-1 signaling pathways by melatonin inhibits cell proliferation and triggers apoptosis in breast cancer cells in *in vitro* models (140). Melatonin also regulates mouse double minute 2 homolog (MDM2), since this indolamine strongly represses MDM2 gene expression and inhibits MDM2 translocation into the nucleus of the cells (141). This can be explained because melatonin stimulates ribosomal

protein L11 and inhibits the phosphorylation of MDM2 by Akt-phosphatidylinositol-4,5-bisphosphate 3-kinase (141). Melatonin downregulates sirtuin, which is a specific inhibitor of p300, and upregulates p300 and murine double minute X (MDMX) (141). As a consequence, cells exposed to melatonin exhibit significantly increased levels of p53 and of its acetylated form (141). Finally, there is a significant increase in p21 levels in melatonin-treated tumor cells (141). It has been reported that melatonin sensitizes non-small cell lung cancer cells harboring a mutated form of EGFR to gefitinib (a tyrosine kinase inhibitor) (142). In combination with cisplatin, melatonin enhances the cytotoxic effects of this chemotherapeutic agent and promotes the entry into apoptosis of lung cancer cells (143) and cervical cancer-derived HeLa cells (144). Consistently with these findings, co-treatment with melatonin and each of the following three chemotherapy agents: Cisplatin, 5-fluorouracil and doxorubicin, resulted in an enhancement of cytotoxicity and apoptosis triggered by chemotherapy in the cell line AR42J, which is derived from rat pancreatic tumors (145). There is only one report addressing the effect of melatonin combined with the purine nucleoside antimetabolite clofarabine, describing that melatonin use as co-treatment led to an enhanced cytotoxic effect of clofarabine in leukemic cell lines, which was associated with higher levels of acetylation (146). In ER⁺ breast cancer rat models treated with Adriamycin[®], melatonin co-treatment results in lighter tumor weights, increased tumor cell apoptosis, higher expression of E-cadherin and higher survival rate (147). In combination with the nucleoside analogue gemcitabine, recent reports demonstrate that melatonin inhibits both proliferation and invasion of pancreatic ductal adenocarcinoma cells through nuclear factor- κ B inhibition (148). Melatonin supports the effects of doxorubicin by activating transient receptor potential vanilloid 1 and apoptosis, thus inducing MCF-7 cell death (149). In a model of ovarian carcinoma, melatonin therapy promotes apoptosis along with the upregulation of p53, B-cell lymphoma (Bcl)-2-associated X protein (Bax) and cleaved caspase-3, suggesting that melatonin triggers apoptosis in ovarian cancer cells (150). In a gastric cancer cell line (AGS), p38, c-Jun N-terminal kinase and extracellular signal-regulated kinase were activated by melatonin, which also significantly increased caspase-3 activity, increased the expression of the pro-apoptotic gene Bax and decreased the expression of the anti-apoptotic gene Bcl-2 (151). Additionally, melatonin is able to strengthen the antitumor effects of cisplatin with low systemic toxicity (143).

5. Conclusions

Melatonin is a notable molecule to be considered in cancer treatment. A growing amount of evidence in the last few years has suggested that melatonin behaves as an oncostatic agent in a variety of cancer types in general, an in particularly, in hormone-dependent breast cancer, as documented from numerous studies performed either in animal models *in vivo* or in cell lines derived from human breast cancer *in vitro* (64,67,81). Clinical trials suggest that melatonin can have protective effects when administered along with chemotherapy in patients suffering from advanced solid tumors (129). The mechanisms underlying the oncostatic actions of the pineal hormone in

Table I. PubMed search results.^a

Gene name	Gene ^a	Gene AND breast cancer	Gene AND melatonin	Gene AND breast cancer AND melatonin	Gene AND breast cancer AND melatonin AND chemotherapy	Gene AND cancer AND melatonin AND chemotherapy
ER	68,885	31,082	189	96	31	52
p53	78,342	7,698	80	15	7	12
p21	31,647	2,174	35	9	5	12
VEGF	57,030	2,737	64	12	3	11
PGR	33,762	13,582	73	96	3	5
TGFB1	22,043	561	25	4	3	4
MYC	29,932	1,882	17	5	3	3
CDH1	23,363	2,200	13	5	3	3
IL6	96,419	1,106	162	3	2	10
GSTP1	35,897	901	91	2	2	7
c-JUN	102,575	2,952	106	6	2	6
RARB	11,545	651	76	9	2	5
CCND1	16,332	2,205	16	6	2	2
AR	22,741	1,690	64	3	1	11
AKT	54,018	3,992	92	6	1	10
Ki-67	20,827	3,129	18	5	1	5
ERK1	26,166	1,153	61	4	1	3
ERBB2	22,268	14,282	72	3	1	3
EGFR	35,864	3,550	7	3	1	3
IGFBP3	5,003	432	6	2	1	2
EGF	29,673	2,215	24	6	1	2
Rb	19,405	1,017	16	1	1	1
IGF1	2,700	160	15	1	1	1
CDK2	6,054	491	5	1	1	1
p73	2,089	113	1	1	1	1

^aA search in PubMed database was performed for 84 genes known to be altered in breast cancer. The numbers refer to the citation found when the keywords used were: i) 'Gene name'; ii) 'gene name' AND 'breast cancer'; iii) 'gene name' AND 'melatonin'; iv) 'gene name' AND 'breast cancer' AND 'melatonin'; v) 'gene name' AND 'breast cancer' AND 'melatonin' AND 'chemotherapy'; and vi) 'gene name' AND 'cancer' AND 'melatonin' AND 'chemotherapy'. In the table, only the 25 genes that appear at least in one publication with the criteria 'gene name' AND 'breast cancer' AND 'melatonin' AND 'chemotherapy' are shown. The genes that have appear in any publication under these criteria of search are gelatinase A, PTGS2, Bad, Bcl-2, BIRC5, gelatinase B, CTNBN1, APC, ASC, ATM, ABCB1, ABCG2, BRCA1, TFF3, cathepsin, μ -PA, SRC, PAI-1, serpine 1, JNK1, IGF1R, CDKN2A, ADAM23, PTEN, NOTCH1, THBS1, ID1, keratin 5, GATA3. ERK2, CCNE1, XBP1, NR3C1, BRCA2, MUC1, MLH1, keratin 19, NME1, TWIST1, FOXA1, RASFF1, HIC1, SFN, MGMT, CCND2, cystatin, GRB7, keratin8, GLI1, keratin18, SFRP1, SNAI2, p57, cyclin A1, CDH13, CSF1, SLIT2, SLC39A6 and PRM2.

estrogen-dependent breast tumors are based on its behaviour as a SERM and SEEM, as well as on the ability of melatonin to influence the communication among malignant epithelial cells, endothelial cells and fibroblasts in breast cancer (110,116). However, to date, there is limited knowledge about the interplays of melatonin and chemotherapy on molecular aspects such as gene expression profiles and gene post-translational modifications, which must be further addressed in the future.

6. Melatonin and cancer: What next?

Melatonin is a pleiotropic molecule that exerts numerous physiological functions and serves important roles in different

processes, including circadian rhythm, sleep and reproduction (152). A particular characteristic of the pineal hormone is the diversity of molecular mechanisms that act to regulate the above physiological processes. Melatonin reduces cell proliferation and growth of estrogen-positive breast tumors by interfering with estrogen signaling pathways. Data obtained from experiments performed in breast cancer cell lines (93,94) and animal models (81,116) have provided evidence that melatonin diminishes the incidence of mammary tumors and limits their growth *in vivo*, and inhibits the proliferation of human breast cancer cell lines and interferes with their metastatic behaviour *in vitro* (81,97). There are several proposed theories to explain the mechanisms by which melatonin reduces the growth and

1 Table II. Number of publications identified in the MEDLINE
 2 (<https://www.ncbi.nlm.nih.gov/pubmed>) journal citation
 3 database (accessed November 2015), when using as search
 4 term each of the 28 molecules currently used in breast cancer
 5 research and/or treatment, alone (second column), including
 6 melatonin (third column) or including melatonin plus breast
 7 cancer (fourth column) as searching criteria.

Drug name	Drug alone	Melatonin	Melatonin AND breast cancer
-	-	20,724	524
Vincristine	27,607	10	0
Eribulin	252	0	0
Paclitaxel	27,688	8	2
Docetaxel	11,691	3	1
Epirubicin	6,131	5	1
Lapatinib	1,807	1	0
Trastuzumab	7,382	2	0
Pazopanib	825	0	0
Bevacizumab	11,697	6	0
Fulvestrant	2,278	5	1
Anastrozole	1,761	1	0
Irosustat	46	0	0
Tanespicin	665	0	0
Cisplatin	59,192	50	4
Gemcitabine	12,035	6	1
Pitavastatin	658	3	1
Pravastatin	4,318	1	1
Vinblastine	1,582	14	0
Cyclophosphamide	64,488	63	8
Methotrexate	45,411	20	4
Fluorouracil	48,858	18	6
Adriamycin	60,505	74	8
Vinorelbine	3,472	2	0
Mitomycin	18,202	6	0
Capecitabine	4,746	1	0
Mitoxantrone	5,538	5	2
Carboplatin	13,873	4	0
Tamoxifen	25,107	44	23

47 development of tumors: i) Through an indirect mechanism, by
 48 downregulating the synthesis of estrogens via downregulation
 49 of the hypothalamic-pituitary-reproductive axis; ii) through a
 50 direct mechanism, by interfering with the activation of estradiol
 51 receptors at the cancer cell level, thus behaving as a SERM;
 52 and iii) melatonin can regulate the enzymes necessary for the
 53 synthesis of estrogens in other tissues, therefore behaving as a
 54 SEEM (79). It has been recently demonstrated that melatonin
 55 regulates the paracrine communication that occurs between
 56 malignant epithelial cancer cells, the surrounding adipose
 57 tissue (fibroblasts and adipocytes) and endothelial cells, mainly
 58 through the downregulation of the levels of growth factors and
 59 cytokines released by breast tumor cells (116). Thus, the effects
 60 of melatonin also include anti-angiogenic actions.

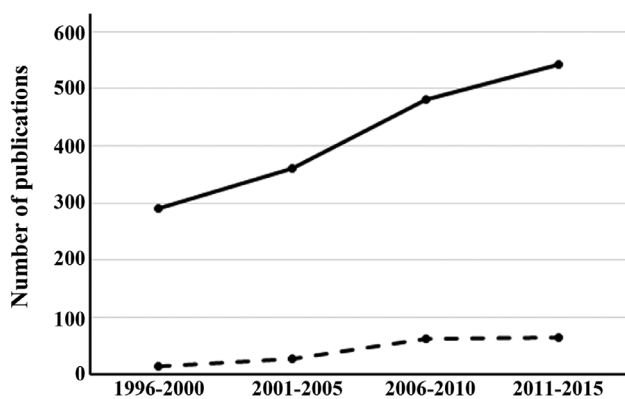


Figure 1. Number of publications regarding melatonin and cancer (solid line), and melatonin, cancer and gene expression (dotted line) published during the last two decades.

In summary, previous studies point to melatonin as a molecule that has a great potential to be useful as an anticancer chemical without producing adverse effects (128). Therefore, melatonin should be considered for both the prevention and therapy of estrogen-positive mammary tumors. There are, in our opinion, numerous noteworthy possibilities for future clinical applications of melatonin in several types of cancer, including breast cancer.

Recently, the inhibitory effects of melatonin have been described not only for estrogen-dependent breast tumors, but also for numerous different cancers, including gastric cancer, ovarian carcinoma, pancreatic ductal carcinoma, leukemic cell lines, cervical cancer and non-small lung carcinoma cells (25,137-139,142,143,153). The majority of the results are positive, and melatonin has been described as an inhibitor of tumor growth under both *in vitro* and *in vivo* experimental conditions (64,81,97). The results arisen in the past few years also suggest that melatonin, either alone or along with chemotherapy in cancer patients diagnosed with advanced solid tumors, helps to improve the outcomes of cancer regression and life expectancy of the patients (122,128). Additionally, chemotherapies are typically better tolerated by patients who are simultaneously treated with melatonin (125). Following the pioneer clinical study of melatonin potential positive effects in untreatable advanced cancer patients performed by Lissoni *et al* (118) several studies have been published (124-129). The main limitation is the requirement of further studies, including additional randomized double-blind controlled trials with much larger sample sizes and implicating several international hospital centres, since the data available nowadays derive from clinical trials including only a few hundred patients (128). Furthermore, it must be considered that not all the studies performed to date point to melatonin as a molecule that improves life expectancy and ameliorates the adverse effects of chemotherapy (120,121). Thus, in patients with advanced lung or gastrointestinal cancer, melatonin did not exhibit any beneficial effect, and as consequence, the value of melatonin as an adjuvant in the treatment of cancer remains unclear from these data (120).

One of the main objectives of the present review was to summarise the current knowledge regarding the interplay of melatonin and chemotherapy. Since the majority of the potentially beneficial effects of melatonin have been described in

1 estrogen-responsive breast cancer, the present study searched
2 information published on 84 genes known to be dysregulated
3 during breast carcinogenesis (corresponding to the genes
4 included in The Human Breast Cancer RT² Profiler PCR
5 array; Qiagen GmbH, Hilden, Germany). These genes encode
6 proteins implicated in signal transduction, angiogenesis,
7 proteolysis, cell cycle and apoptosis (154).

8 The present study also reviewed recently published
9 articles associating melatonin with chemotherapeutic
10 agents (135-137,142-149). The results indicate that, for 53 out
11 of the 84 genes evaluated, there are no current data available
12 regarding the effect of melatonin alone or in combination
13 with chemotherapy, either in *in vivo* or in *in vitro* studies. Our
14 findings also show that, apart from tamoxifen, there is limited
15 information from research performed at the molecular level
16 addressing the potential benefits of co-treatment of melatonin
17 with chemotherapeutic agents (155).

18 In summary, in our opinion, melatonin is an endogenous
19 produced hormone with a high potential of being included
20 as an effective anticancer molecule in the prevention and
21 treatment of, not only hormone-dependent cancers, but also,
22 other types of cancer, since its inhibitory effects have been
23 demonstrated in gastric, lung, pancreatic and hematopoietic
24 cancers (145,148,151,153). However, in the next years, addi-
25 tional research must be conducted to clarify if melatonin
26 administration in combination with chemotherapeutic agents
27 may constitute a novel anticancer treatment. In particular,
28 future research concerning the role of melatonin as a non-toxic
29 and low-cost drug to be considered in breast and other types of
30 tumors must be conducted, particularly at the molecular level.
31 Systematic screenings addressing the effects of chemotherapy
32 on genes known to be altered in different types of cancer, and
33 on how melatonin can modulate the expression and activity of
34 those genes, either when acting alone or in combination with
35 chemotherapy, should be performed. Once larger clinical trials
36 and additional molecular studies (including gene expression
37 profiles, post-translational modifications and individual gene
38 tests) have been conducted, it may be reasonable to recom-
39 mend melatonin as a potential drug to be considered in the
40 treatment of breast cancer.

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46 References

- 47 1. World Cancer Research Fund/American Institute for Cancer
48 Research: Second Expert Report. <http://www.wcrf.org/int/research-we-fund/continuous-update-project-cup/second-expert-report>. Accessed August 14, 2016.
- 49 2. American Cancer Society: Cancer facts and figures 2015.
50 <http://www.cancer.org/research/cancerfactsstatistics/cancer-factsfigures2015>. Accessed August 14, 2016.
- 51 3. American Cancer Society: What are the key statistics about
52 breast cancer? <http://www.cancer.org/cancer/breastcancer/detailedguide/breast-cancer-key-statistics>. Accessed August 14,
53 2016.
- 54 4. Reynolds T: Declining breast cancer mortality: What's behind it?
55 *J Natl Cancer Inst* 91: 750-753, 1999.
- 56 5. Rossouw JE, Anderson GL, Prentice RL, LaCroix AZ,
57 Kooperberg C, Stefanick ML, Jackson RD, Beresford SA,
58 Howard BV; Writing Group for the Women's Health Initiative
59 Investigators, *et al*: Risks and benefits of estrogens plus progestin
60 in healthy postmenopausal women. Principal results from the
61 Women's Health Initiative Randomized Controlled trial. *J Am
62 Med Assoc* 288: 321-333, 2002.
- 63 6. Greig RG and Trainer DL: Shaping future strategies for the phar-
64 macological control of tumor cell metastases. *Cancer Metastasis
65 Rev* 5: 3-14, 1986.
- 66 7. Perloff M and Holland JF: Surgical adjuvant chemotherapy. *Ann
67 Rev Med* 28: 475-488, 1977.
- 68 8. Maughan KL, Lutterbie MA and Ham PS: Treatment of breast
69 cancer. *Am Fam Physician* 81: 1339-1346, 2010.
- 70 9. Mamounas EP: Impact of neoadjuvant chemotherapy on locoregional
71 surgical treatment of breast cancer. *Ann Surg Oncol* 22:
72 1425-1433, 2015.
- 73 10. Masood S: Neoadjuvant chemotherapy in breast cancers.
74 *Womens Health (Lond)* 12: 480-491, 2016.
- 75 11. Suter R and Marcum JA: The molecular genetics of breast cancer
76 and targeted therapy. *Biologics* 1: 241-258, 2007.
- 77 12. BreastCancer.Org. How is Chemotherapy Given? <http://www.breastcancer.org/treatment/chemotherapy/process/how>. Accessed
78 August 14, 2016.
- 79 13. Jordan MA and Wilson L: Microtubules as a target for anticancer
80 drugs. *Nat Rev Cancer* 4: 253-265, 2004.
- 81 14. Moudi M, Go R, Yien CY and Nazre M: *Vinca* alkaloids. *Int J
82 Prev Med* 4: 1231-1235, 2013.
- 83 15. Johnson IS, Armstrong JG, Gorman M and Burnett JP Jr: The
84 *Vinca* Alkaloids: A new class of oncolytic agents. *Cancer Res* 23:
85 1390-1427, 1963.
- 86 16. Himes RH: Interactions of the catharanthus (*Vinca*) alkaloids
87 with tubulin and microtubules. *Pharmacol Ther* 51: 257-267, 1991.
- 88 17. Gigant B, Wang C, Ravelli RB, Roussi F, Steinmetz MO,
89 Curmi PA, Sobel A and Knossow M: Structural basis for the
90 regulation of tubulin by vinblastine. *Nature* 435: 519-522, 2005.
- 91 18. Graf E and Bertholdt H: Amorphous taxine & crystalline
92 taxine A; *Taxus* alkaloids. II. *Pharm Zentralhalle Dtschl* 96:
93 385-395, 1957 (In German).
- 94 19. Wani MC, Taylor HL, Wall ME, Coggon P and McPhail AT:
95 Plant antitumor agents. VI. The isolation and structure of taxol,
96 a novel antileukemic and antitumor agente from *Taxus brevifolia*.
97 *J Am Chem Soc* 93: 2325-2327, 1971.
- 98 20. Wiernik PH, Schwartz EL, Strauman JJ, Dutcher JP, Lipton RB
99 and Paietta E: Phase I clinical and pharmacokinetic study of
100 taxol. *Cancer Res* 47: 2486-2493, 1987.
- 101 21. Bissery MC, Guénard D, Guéritte-Voegelien F and Lavelle F:
102 Experimental antitumor activity of Taxotere (RP 56976,
103 NSC 628503), a taxol analogue. *Cancer Res* 51: 4845-4852, 1991.
- 104 22. Ringel I and Horwitz SB: Studies with RP 56976 (Taxotere): A
105 semisynthetic analogue of taxol. *J Natl Cancer Inst* 83: 288-291,
106 1991.
- 107 23. Xiao H, Verdier-Pinard P, Fernandez-Fuentes N, Burd B,
108 Angeletti R, Fiser A, Horwitz SB and Orr GA: Insights into the
109 mechanism of microtubule stabilization by Taxol. *Proc Natl
110 Acad Sci USA* 103: 10166-10173, 2006.
- 111 24. Snyder JP, Nettles JH, Cornet B, Downing KH and Nogales E:
112 The binding conformation of Taxol in beta-tubulin: A model
113 based on electron crystallographic density. *Proc Natl Acad Sci
114 USA* 98: 5312-5316, 2001.
- 115 25. Jain KK, Casper ES, Geller NL, Hakes TB, Kaufman RJ, Currie V,
116 Schwartz W, Cassidy C, Petroni GR, Young CW, *et al*: A prospective
117 randomized comparison of epirubicin and doxorubicin in patients
118 with advanced breast cancer. *J Clin Oncol* 3: 818-826, 1985.
- 119 26. Mizuno NS, Zakis B and Decker RW: Binding of daunomycin
120 to DNA and the inhibition of RNA and DNA synthesis. *Cancer
121 Res* 35: 1542-1546, 1975.
- 122 27. Andoh T: Inhibitors of DNA topoisomerases. *Gan To Kagaku
123 Ryocho* 24: 1526-1535, 1997 (In Japanese).
- 124 28. Khasraw M, Bell R and Dang C: Epirubicin: Is it like doxorubicin
125 in breast cancer? A clinical review. *Breast* 21: 142-149, 2012.
- 126 29. US Food and Drug Administration. Advisory Committees:
127 2010 Meeting Materials, Oncologic Drugs Advisory Committee.
128 <http://www.cancer.gov/about-cancer/treatment/drugs/fda-eribulinmesylate>. Accessed November 28, 2016
- 129 30. Towle MJ, Salvato KA, Wels BF, Aalfs KK, Zheng W, Seletsky BM,
130 Zhu X, Lewis BM, Kishi Y, Yu MJ and Littlefield BA: Eribulin
131 induces irreversible mitotic blockade: Implications of cell-based
132 pharmacodynamics for *in vivo* efficacy under intermittent dosing
133 conditions. *Cancer Res* 71: 496-505, 2011.

- 1 31. Marhold M, Bartsch R and Zielinski C: Recent developments and translational aspects in targeted therapy for metastatic breast cancer. *ESMO Open* 1: e000036, 2016.
- 2 32. Wong WM: Drug update: Trastuzumab: Anti-HER2 antibody for treatment of metastatic breast cancer. *Cancer Pract* 7: 48-50, 1999.
- 3 33. Pegram MD, Lipton A, Hayes DF, Weber BL, Baselga JM, Tripathy D, Baly D, Baughman SA, Twaddell T, Glaspy JA and Slamon DJ: Phase II study of receptor-enhanced chemosensitivity using recombinant humanized anti-p185HER2/neu monoclonal antibody plus cisplatin in patients with HER2/neu-overexpressing metastatic breast cancer refractory to chemotherapy treatment. *J Clin Oncol* 16: 2659-2671, 1998.
- 4 34. Mayer IA: Treatment of HER2-positive metastatic breast cancer following initial progression. *Clin Breast Cancer* 9 (Suppl 2): S50-S57, 2009.
- 5 35. Kaufman B, Trudeau M, Awada A, Blackwell K, Bachelot T, Salazar V, DeSilvio M, Westlund R, Zaks T, Spector N and Johnston S: Lapatinib monotherapy in patients with HER2-overexpressing relapsed or refractory inflammatory breast cancer: Final results and survival of the expanded HER2+ cohort in EGR103009, a phase II study. *Lancet Oncol* 10: 581-588, 2009.
- 6 36. Gomez HL, Doval DC, Chavez MA, Ang PC, Aziz Z, Nag S, Ng C, Franco SX, Chow LW, Arbushites MC, *et al*: Efficacy and safety of lapatinib as first-line therapy for ErbB2-amplified locally advanced or metastatic breast cancer. *J Clin Oncol* 26: 2999-3005, 2008.
- 7 37. Burris HA III: Dual kinase inhibition in the treatment of breast cancer: Initial experience with the EGFR/ErbB-2 inhibitor lapatinib. *Oncologist* 9 (Suppl 3): S10-S15, 2004.
- 8 38. Burstein HJ, Sun Y, Dirix LI, Jiang Z, Paridaens R, Tan AR, Awada A, Ranade A, Jiao S, Schwartz G, *et al*: Neratinib, an irreversible ErbB receptor tyrosine kinase inhibitor, in patients with advanced ErbB2-positive breast cancer. *J Clin Oncol* 28: 1301-1307, 2010.
- 9 39. Rabindran SK, Discafani CM, Rosfjord EC, Baxter M, Floyd MB, Golas J, Hallett WA, Johnson BD, Nilakantan R, Overbeek E, *et al*: Antitumor activity of HKI-272, an orally active, irreversible inhibitor of the HER-2 tyrosine kinase. *Cancer Res* 64: 3958-3965, 2004.
- 10 40. Darby JF and Workman P: Chemical biology: Many faces of a cancer-supporting protein. *Nature* 478: 334-335, 2011.
- 11 41. Hague A, Alam Q, Alam MZ, Azhar EI, Sait KH, Anfinan N, Mushtaq G, Kamal MA and Rasool M: Current understanding of HSP90 as a novel therapeutic target: An emerging approach for the treatment of cancer. *Curr Pharm Des* 22: 2947-2959, 2016.
- 12 42. Zsebk B, Citri A, Isola J, Yarden Y, Szöllosi J and Vereb G: Hsp90 inhibitor 17-AAG reduces ErbB2 levels and inhibits proliferation of the trastuzumab resistant breast tumor cell line JIMT-1. *Immunol Lett* 104: 146-155, 2006.
- 13 43. Beliakoff J and Whitesell L: Hsp90: An emerging target for breast cancer therapy. *Anticancer Drugs* 15: 651-662, 2004.
- 14 44. McDonald PC, Chafe SC and Dedhar S: Overcoming hypoxia-mediated tumor progression: Combinatorial approaches targeting pH regulation, angiogenesis and immune dysfunction. *Front Cell Dev Biol* 4: 27, 2016.
- 15 45. Rao N, Lee YE and Ge R: Novel endogenous angiogenesis inhibitors and their therapeutic potential. *Acta Pharmacol Sin* 36: 1177-1190, 2015.
- 16 46. Arjaans M, Schröder CP, Oosting SF, Dafni U, Kleibeuker JE and de Vries EG: VEGF pathway targeting agents, vessel normalization and tumor drug uptake: From bench to bedside. *Oncotarget* 7: 21247-21258, 2016.
- 17 47. Kerbel RS: Strategies for improving the clinical benefit of anti-angiogenic drug based therapies for breast cancer. *J Mammary Gland Biol Neoplasia* 17: 229-239, 2012.
- 18 48. Kerr DJ: Targeting angiogenesis in cancer: Clinical development of bevacizumab. *Nat Clin Pract Oncol* 1: 39-43, 2004.
- 19 49. Mackey JR, Kerbel RS, Gelmon KA, McLeod DM, Chia SK, Rayson D, Verma S, Collins LL, Paterson AH, Robidoux A and Pritchard KI: Controlling angiogenesis in breast cancer: A systematic review of anti-angiogenic trials. *Cancer Treat Rev* 38: 673-688, 2012.
- 20 50. Miller KD, Chap LI, Holmes FA, Cobleigh MA, Marcom PK, Fehrenbacher L, Dickler M, Overmoyer BA, Reimann JD, Sing AP, *et al*: Randomized phase III trial of capecitabine compared with bevacizumab plus capecitabine in patients with previously treated metastatic breast cancer. *J Clin Oncol* 23: 792-799, 2005.
- 21 51. Sloan B and Scheinfeld NS: Pazopanib, a VEGF receptor tyrosine kinase inhibitor for cancer therapy. *Curr Opin Investig Drugs* 9: 1324-1335, 2008.
- 22 52. Gril B, Palmieri D, Qian Y, Smart D, Ileva L, Liewehr DJ, Steinberg SM and Steeg PS: Pazopanib reveals a role for tumor cell B-Raf in the prevention of HER2+ breast cancer brain metastasis. *Clin Cancer Res* 17: 142-153, 2011.
- 23 53. Taylor SK, Chia S, Dent S, Clemons M, Agulnik M, Greci P, Wang L, Oza AM, Ivy P, Pritchard KI and Leighl NB: A phase II study of pazopanib in patients with recurrent or metastatic invasive breast carcinoma: A trial of the princess Margaret Hospital phase II consortium. *Oncologist* 15: 810-818, 2010.
- 24 54. Mauvais-Jarvis P, Kuttann F and Gompel A: Estradiol/progesterone interaction in normal and pathological breast cells. *Ann N Acad Sci* 464: 152-167, 1986.
- 25 55. Lumachi F, Santeufemia DA and Basso SM: Current medical treatment of estrogen receptor-positive breast-cancer. *World J Biol Chem* 6: 231-239, 2015.
- 26 56. Ellis AJ, Hendrick VM, Williams R and Komm BS: Selective estrogen receptor modulators in clinical practice: A safety overview. *Expert Opin Drug Saf* 14: 921-934, 2015.
- 27 57. Jordan VC: Tamoxifen as the first targeted long-term adjuvant therapy for breast cancer. *Endocr Relat Cancer* 21: R235-R246, 2014.
- 28 58. Dalmau E, Armengol-Alonso A, Muñoz M and Seguí-Palmer MA: Current status of hormone therapy in patients with hormone receptor positive (HR+) advanced breast cancer. *Breast* 23: 710-720, 2014.
- 29 59. Morris C and Wakeling A: Fulvestrant ('Faslodex')-a new treatment option for patients progressing on prior endocrine therapy. *Endocr Relat Cancer* 9: 267-276, 2002.
- 30 60. Johnston SJ and Cheung KL: Fulvestrant-a novel endocrine therapy for breast cancer. *Curr Med Chem* 17: 902-914, 2010.
- 31 61. Wong ZW and Ellis MJ: First-line endocrine treatment of breast cancer: Aromatase inhibitor or antioestrogen? *Br J Cancer* 90: 20-25, 2004.
- 32 62. Reiter RJ, Tan DX and Fuentes-Broto L: Melatonin: A multitasking molecule. *Prog Brain Res* 181: 127-151, 2010.
- 33 63. Reiter RJ, Tan DX and Galano A: Melatonin: Exceeding expectations. *Physiology (Bethesda)* 29: 325-333, 2014.
- 34 64. Hill SM, Belancio VP, Dauchy RT, Xiang S, Brimer S, Mao L, Hauch A, Lundberg PW, Summers W, Yuan L, *et al*: Melatonin: An inhibitor of breast cancer. *Endocr Relat Cancer* 22: R183-R204, 2015.
- 35 65. Dai J, Ram PT, Yuan L, Spriggs LL and Hill SM: Transcriptional repression of RORalpha activity in human breast cancer cells by melatonin. *Mol Cell Endocrinol* 176: 111-120, 2001.
- 36 66. Reiter RJ, Tan DX, Manchester LC, Lopez-Burillo S, Sainz RM and Mayo JC: Melatonin: Detoxification of oxygen and nitrogen-based toxic reactants. *Adv Exp Med Biol* 527: 539-548, 2003.
- 37 67. Blask DE, Pelletier DB, Hill SM, Lemus-Wilson A, Grosso DS, Wilson ST and Wise ME: Pineal melatonin inhibition of tumor promotion in the N-nitroso-N-methylurea model of mammary carcinogenesis: Potential involvement of antiestrogenic mechanisms in vivo. *J Cancer Res Clin Oncol* 117: 526-532, 1991.
- 38 68. Tamarkin L, Almeida OF and Danforth DN Jr: Melatonin and malignant disease. *Ciba Found Symp* 117: 284-299, 1985.
- 39 69. Coleman MP and Reiter RJ: Breast cancer, blindness and melatonin. *Eur J Cancer* 28: 501-503, 1992.
- 40 70. Kliukiene J, Tynes T and Andersen A: Risk of breast cancer among norwegian women with visual impairment. *Br J Cancer* 84: 397-399, 2001.
- 41 71. Flynn-Evans EE, Stevens RG, Tabandeth H, Schernhammer ES and Lockey SW: Total visual blindness is protective against breast cancer. *Cancer Causes Control* 20: 1753-1756, 2009.
- 42 72. Schernhammer ES, Laden F, Speizer FE, Willet WC, Hunter DJ, Kawachi I and Colditz GA: Rotating night shifts and risk of breast cancer in women participating in the nurses' health study. *J Natl Cancer Inst* 93: 1563-1568, 2001.
- 43 73. González A, Cos S, Martínez-Campa C, Alonso-González C, Sánchez-Mateos S, Mediavilla MD and Sánchez-Barceló EJ: Selective estrogen enzyme modulator actions of melatonin in human breast cancer cells. *J Pineal Res* 45: 86-92, 2008.
- 44 74. Cos S, González A, Martínez-Campa C, Mediavilla MD, Alonso-González C and Sánchez-Barceló EJ: Estrogen-signaling pathway: A link between breast cancer and melatonin oncostatic actions. *Cancer Detect Prev* 30: 118-128, 2006.
- 45 75. Martínez-Campa C, González A, Mediavilla MD, Alonso-González C, Sánchez-Barceló EJ and Cos S: Melatonin enhances the inhibitory effect of aminoglutethimide on aromatase activity in MCF-7 human breast cancer cells. *Breast Cancer Res Treat* 94: 249-254, 2005.

- 1 76. Cos S, Martínez-Campa C, Mediavilla MD and Sánchez-Barceló EJ: Melatonin modulates aromatase activity in MCF-7 human breast cancer cells. *J Pineal Res* 38: 136-142, 2005.
- 2
- 3 77. Purohit A, Woo LW and Potter BV: Steroid sulfatase: A pivotal player in estrogen synthesis and metabolism. *Mol Cell Endocrinol* 340: 154-160, 2011.
- 4
- 5 78. Sasano H, Suzuki T, Nakata T and Moriya T: New development in intracrinology of breast carcinoma. *Breast Cancer* 13: 129-136, 2006.
- 6
- 7 79. Cos S, González A, Martínez-Campa C, Mediavilla MD, Alonso-González C and Sánchez-Barceló EJ: Melatonin as a selective estrogen enzyme modulator. *Curr Cancer Drug Targets* 8: 691-702, 2008.
- 8
- 9 80. Álvarez-García V, González A, Martínez-Campa C, Alonso-González C and Cos S: Melatonin modulates aromatase activity and expression in endothelial cells. *Oncol Rep* 29: 2058-2064, 2013.
- 10
- 11 81. González A, Alvarez-García V, Martínez-Campa C, Mediavilla MD, Alonso-González C, Sánchez-Barceló EJ and Cos S: In vivo inhibition of the estrogen sulfatase enzyme and growth of DMBA-induced mammary tumors by melatonin. *Curr Cancer Drug Targets* 10: 279-286, 2010.
- 12
- 13 82. González A, Martínez-Campa C, Alonso-González C and Cos S: Melatonin affects the dynamic steady-state equilibrium of estrogen sulfates in human umbilical vein endothelial cells by regulating the balance between estrogen sulfatase and sulfotransferase. *Int J Mol Med* 36: 1671-1676, 2015.
- 14
- 15 83. Sánchez-Barceló EJ, Cos S, Mediavilla D, Martínez-Campa C, González A and Alonso-González C: Melatonin-estrogen interactions in breast cancer. *J Pineal Res* 38: 217-222, 2005.
- 16
- 17 84. García Pedrero JM, Del Río B, Martínez-Campa C, Muramatsu M, Lazo PS and Ramos S: Calmodulin is a selective modulator of estrogen receptors. *Mol Endocrinol* 16: 947-960, 2002.
- 18
- 19 85. Molis TM, Spriggs LL and Hill SM: Modulation of estrogen receptor mRNA expression by melatonin in MCF-7 human breast cancer cells. *Mol Endocrinol* 8: 1681-1690, 1994.
- 20
- 21 86. Rato AG, Pedrero JG, Martínez MA, del Río B, Lazo PS and Ramos S: Melatonin blocks the activation of estrogen receptor for DNA binding. *FASEB J* 13: 857-868, 1999.
- 22
- 23 87. Bouhoute A and Leclercq G: Calmodulin decreases the estrogen binding capacity of the estrogen receptor. *Biochem Biophys Res Commun* 227: 651-657, 1996.
- 24
- 25 88. del Río B, García Pedrero JM, Martínez-Campa C, Zuazua P, Lazo PS and Ramos S: Melatonin, an endogenous-specific inhibitor of estrogen receptor alpha via calmodulin. *J Biol Chem* 279: 38294-38302, 2004.
- 26
- 27 89. Ram PT, Dai J, Yuan L, Dong C, Kiefer TL, Lai L and Hill SM: Involvement of the mtl melatonin receptor in human breast cancer. *Cancer Lett* 179: 141-150, 2002.
- 28
- 29 90. Dillon DC, Easley SE, Asch BB, Cheney RT, Brydon L, Jockers R, Winston JS, Brooks JS, Hurd T and Asch HL: Differential expression of high-affinity melatonin receptors (MT1) in normal and malignant human breast tissue. *Am J Clin Pathol* 118: 451-458, 2002.
- 30
- 31 91. Aronica SM, Kraus WL and Katzenellenbogen BS: Estrogen action via the cAMP signaling pathway: Stimulation of adenylate cyclase and cAMP-regulated gene transcription. *Proc Natl Acad Sci USA* 91: 8517-8521, 1994.
- 32
- 33 92. Godson C and Reppert SM: The Mella melatonin receptor is coupled to parallel signal transduction pathways. *Endocrinology* 138: 397-404, 1997.
- 34
- 35 93. Hill SM, Spriggs LL, Simon MA, Muraoka H and Blask DE: The growth inhibitory action of melatonin on human breast cancer cells is linked to the estrogen response system. *Cancer Lett* 64: 249-256, 1992.
- 36
- 37 94. Cos S and Blask DE: Melatonin modulates growth factor activity in MCF-7 human breast cancer cells. *J Pineal Res* 17: 25-32, 1994.
- 38
- 39 95. Molis TM, Spriggs LL, Jupiter Y and Hill SM: Melatonin modulation of estrogen-regulated proteins, growth factors, and proto-oncogenes in human breast cancer. *J Pineal Res* 18: 93-103, 1995.
- 40
- 41 96. Hong J, Shah NN, Thomas TJ, Gallo MA, Yurkow EJ and Thomas T: Differential effects of estradiol and its analogs on cyclin D1 and CDK4 expression in estrogen receptor positive MCF-7 and estrogen receptor-transfected MCF-10Aewt5 cells. *Oncol Rep* 5: 1025-1033, 1998.
- 42
- 43 97. Cos S, Blask DE, Lemus-Wilson A and Hill AB: Effects of melatonin on the cell kinetics and 'estrogen-rescue' of MCF-7 human breast cancer cells in culture. *J Pineal Res* 10: 36-42, 1991.
- 44
- 45 98. Mediavilla MD, Cos S and Sánchez-Barceló EJ: Melatonin increases p53 and p21WAF1 expression in MCF-7 human breast cancer cells in vitro. *Life Sci* 65: 415-420, 1999.
- 46
- 47 99. Alonso-González C, González A, Martínez-Campa C, Menéndez-Menéndez J, Gómez-Arozamena J, García-Vidal A and Cos S: Melatonin enhancement of the radiosensitivity of human breast cancer cells is associated with the modulation of proteins involved in estrogen biosynthesis. *Cancer Lett* 370: 145-152, 2016.
- 48
- 49 100. Wang SC, Lin SH, Su LK and Hung MC: Changes in BRCA2 expression during progression of the cell cycle. *Biochem Biophys Res Commun* 234: 247-251, 1997.
- 50
- 51 101. Leon-Blanco MM, Guerrero JM, Reiter RJ, Calvo JR and Pozo D: Melatonin inhibits telomerase activity in the MCF-7 tumor cell line both in vivo and in vitro. *J Pineal Res* 35: 204-211, 2003.
- 52
- 53 102. Martínez-Campa CM, Alonso-González C, Mediavilla MD, Cos S, González A and Sánchez-Barceló EJ: Melatonin down-regulates hTERT expression induced by either natural estrogens (17beta-estradiol) or metalloestrogens (cadmium) in MCF-7 human breast cancer cells. *Cancer Lett* 268: 272-277, 2008.
- 54
- 55 103. Alonso-González C, Mediavilla D, Martínez-Campa C, González A, Cos S and Sánchez-Barceló EJ: Melatonin modulates the cadmium-induced expression of MT-2 and MT-1 metallothioneins in three lines of human tumor cells (MCF-7, MDA-MB-231 and HeLa). *Toxicol Lett* 181: 190-195, 2008.
- 56
- 57 104. Mao L, Yuan L, Slakey LM, Jones FE, Burow ME and Hill SM: Inhibition of breast cancer cell invasion by melatonin is mediated through regulation of the p38 mitogen-activated protein kinase signaling pathway. *Breast Cancer Res* 12: R107, 2010.
- 58
- 59 105. Cos S, Fernández R, Gúezmes A and Sánchez-Barceló EJ: Influence of melatonin on invasive and metastatic properties of MCF-7 human breast cancer cells. *Cancer Res* 58: 4383-4390, 1998.
- 60
- 61 106. Kawaguchi T: Cancer metastasis: Characterization and identification of the behavior of metastatic tumor cells and the cell adhesion molecules, including carbohydrates. *Curr Drug Targets Cardiovasc Haematol Disord* 5: 39-64, 2005.
- 62
- 63 107. Canel M, Serrels A, Frame MC and Brunton VG: E-cadherin-integrin crosstalk in cancer invasion and metastasis. *J Cell Sci* 126: 393-401, 2013.
- 64
- 65 108. Gui GP, Puddefoot JR, Vinson GP, Wells CA and Carpenter R: Altered cell-matrix contact: A prerequisite for breast cancer metastasis? *Br J Cancer* 75: 623-633, 1997.
- 66
- 67 109. González A, Alvarez-García V, Martínez-Campa C, Alonso-González C and Cos S: Melatonin promotes differentiation of 3T3-L1 fibroblasts. *J Pineal Res* 52: 12-20, 2012.
- 68
- 69 110. Alvarez-García V, González A, Alonso-González C, Martínez-Campa C and Cos S: Melatonin interferes in the desmoplastic reaction in breast cancer by regulating cytokine production. *J Pineal Res* 52: 282-290, 2012.
- 70
- 71 111. Alvarez-García V, González A, Alonso-González C, Martínez-Campa C and Cos S: Regulation of vascular endothelial growth factor by melatonin in human breast cancer cells. *J Pineal Res* 54: 373-380, 2013.
- 72
- 73 112. Gingis-Velitski S, Zetser A, Flugelman MY, Vladavsky I and Ilan N: Heparanase induces endothelial cell migration via protein kinase B/Akt activation. *J Biol Chem* 279: 23536-23541, 2004.
- 74
- 75 113. Gu Q, Wang D, Wang X, Peng R, Liu J, Jiang T, Wang Z, Wang S and Deng H: Basic fibroblast growth factor inhibits radiation-induced apoptosis of HUVECs. I. The PI3K/AKT pathway and induction of phosphorylation of BAD. *Radiat Res* 161: 692-702, 2004.
- 76
- 77 114. Alvarez-García V, González A, Alonso-González C, Martínez-Campa C and Cos S: Antiangiogenic effects of melatonin in endothelial cell cultures. *Microvasc Res* 87: 25-33, 2013.
- 78
- 79 115. Ahmad I and Shagufata: Recent developments in steroidal and nonsteroidal aromatase inhibitors for the chemoprevention of estrogen-dependent breast cancer. *Eur J Med Chem* 102: 375-386, 2015.
- 80
- 81 116. Cos S, Álvarez-García V, González A, Alonso-González C and Martínez-Campa C: Melatonin modulation of crosstalk among malignant epithelial, endothelial and adipose cells in breast cancer (Review). *Oncol Lett* 8: 487-492, 2014.
- 82
- 83 117. Proietti S, Cucina A, Reiter RJ and Bizarri M: Molecular mechanisms of melatonin's inhibitory actions on breast cancers. *Cell Mol Life Sci* 70: 2139-2157, 2013.
- 84
- 85
- 86
- 87
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- 120

118. Lissoni P, Barni S, Tancini G, Crispino S, Paolorossi F, Lucini V, Mariani M, Cattaneo G, Esposti D, Esposti G, *et al*: Clinical study of melatonin in untreatable advanced cancer patients. *Tumori* 73: 475-480, 1987.
119. Grant SG, Melan MA, Latimer JJ and Witt-Enderby PA: Melatonin and breast cancer: Cellular mechanisms, clinical studies and future perspectives. *Expert Rev Mol Med* 11: e5, 2009.
120. Del Fabbro E, Dev R, Hui D, Palmer L and Bruera E: Effects of melatonin on appetite and other symptoms in patients with advanced cancer and cachexia: A double-blind placebo controlled trial. *J Clin Oncol* 31: 1271-1276, 2013.
121. Berk L, Berkey B, Rich T, Hrushesky W, Blask D, Gallagher M, Kudrimoti M, McGarry RC, Suh J and Mehta M: Randomized phase II trial of high-dose melatonin and radiation therapy for RPA class 2 patients with brain metastases (RTOG 0119). *Int J Radiat Oncol Biol Phys* 68: 852-857, 2007.
122. Lissoni P, Tancini G, Barni S, Paolorossi F, Ardizzoia A, Conti A and Maestroni G: Treatment of cancer chemotherapy-induced toxicity with the pineal hormone melatonin. *Support Care Cancer* 5: 126-129, 1997.
123. Vijayalaxmi, Thomas CR Jr, Reiter RJ and Herman TS: Melatonin: From basic research to cancer treatment clinics. *J Clin Oncol* 20: 2575-2601, 2002.
124. Lissoni P: Is there a role for melatonin in supportive care? *Support Care Cancer* 10: 110-116, 2002.
125. Lissoni P, Barni S, Mandalà M, Ardizzoia A, Paolorossi F, Vaghi M, Longarini R, Malugani F and Tancini G: Decreased toxicity and increased efficacy of cancer chemotherapy using the pineal hormone melatonin in metastatic solid tumour patients with poor clinical status. *Eur J Cancer* 35: 1688-1692, 1999.
126. Lissoni P, Chillelli M, Villa S, Cerizza L and Tancini G: Five years survival in metastatic non-small cell lung cancer patients treated with chemotherapy alone or chemotherapy and melatonin: A randomized trial. *J Pineal Res* 35: 12-15, 2003.
127. Sookprasert A, Johns NP, Phunmanee A, Pongthai P, Cheawchanwattana A, Johns J, Konsil J, Plaimée P, Porasuphatana S and Jitpimolmard S: Melatonin in patients with cancer receiving chemotherapy: A randomized, double-blind, placebo-controlled trial. *Anticancer Res* 34: 7327-7337, 2014.
128. Seely D, Wu P, Fritz H, Kennedy DA, Tsui T, Seely AJ and Mills E: Melatonin as adjuvant cancer care with and without chemotherapy: A systematic review and meta-analysis of randomized trials. *Integr Cancer Ther* 11: 293-303, 2012.
129. Wang YM, Jin BZ, Ai F, Duan CH, Lu YZ, Dong TF and Fu QL: The efficacy and safety of melatonin in concurrent chemotherapy or radiotherapy for solid tumors: A meta-analysis of randomized controlled trials. *Cancer Chemother Pharmacol* 69: 1213-1220, 2012.
130. Russo J and Russo IH: The role of estrogen in the initiation of breast cancer. *J Steroid Biochem Mol Biol* 102: 89-96, 2006.
131. Chakravarthi BV, Nepal S and Varambally S: Genomic and epigenomic alterations in cancer. *Am J Pathol* 186: 1724-1735, 2016.
132. Lee SE, Kim SJ, Youn JP, Hwang SY, Park CS and Park YS: MicroRNA and gene expression analysis of melatonin-exposed human breast cancer cell lines indicating involvement of the anticancer effect. *J Pineal Res* 51: 345-352, 2011.
133. Lee SE, Kim SJ, Yoon HJ, Yu SY, Yang H, Jeong SI, Hwang SY, Park CS and Park YS: Genome-wide profiling in melatonin-exposed human breast cancer cell lines identifies differentially methylated genes involved in the anticancer effect of melatonin. *J Pineal Res* 54: 80-88, 2013.
134. Dauchy RT, Xiang S, Mao L, Brimer S, Wren MA, Yuan L, Anbalagan M, Hauch A, Frasc T, Rowan BG, *et al*: Circadian and melatonin disruption by exposure to light at night drives intrinsic resistance to tamoxifen therapy in breast cancer. *Cancer Res* 74: 4099-4110, 2014.
135. Xiang S, Dauchy RT, Hauch A, Mao L, Yuan L, Wren MA, Belancio VP, Mondal D, Frasc T, Blask DE and Hill SM: Doxorubicin resistance in breast cancer is driven by light at night-induced disruption of the circadian melatonin signal. *J Pineal Res* 59: 60-69, 2015.
136. He SM, Li R, Kanwar JR and Zhou SF: Structural and functional properties of human multidrug resistance protein 1 (MRP1/ABCC1). *Curr Med Chem* 18: 439-481, 2011.
137. Granzotto M, Rapozzi V, Decorti G and Giraldi T: Effects of melatonin on doxorubicin cytotoxicity in sensitive and pleiotropically resistant tumor cells. *J Pineal Res* 31: 206-213, 2001.
138. Jardim-Perassi BV, Arbab AS, Ferreira LC, Borin TF, Varma NR, Iskander AS, Shankar A, Ali MM and de Campos Zuccari DA: Effect of melatonin on tumor growth and angiogenesis in xenograft model of breast cancer. *PLoS One* 9: e85311, 2014.
139. Woo SM, Min KJ and Kwon TK: Melatonin-mediated Bim up-regulation and cyclooxygenase-2 (COX-2) down-regulation enhances tunicamycin-induced apoptosis in MDA-MDB-231 cells. *J Pineal Res* 58: 310-320, 2015.
140. Wang J, Xiao X, Zhang Y, Shi D, Chen W, Fu L, Liu L, Xie F, Kang T, Huang W and Deng W: Simultaneous modulation of COX-2, p300, Akt, and Apaf-1 signaling by melatonin to inhibit proliferation and induce apoptosis in breast cancer cells. *J Pineal Res* 53: 77-90, 2012.
141. Proietti S, Cucina A, Dobrowolny G, D'Anselmi F, Dinicola S, Masiello MG, Pasqualato A, Palombo A, Morini V, Reiter RJ and Bizarri M: Melatonin down-regulates MDM2 gene expression and enhances p53 acetylation in MCF-7 cells. *J Pineal Res* 57: 120-129, 2014.
142. Yun M, Kim EO, Lee D, Kim JH, Kim J, Lee H, Lee J and Kim SH: Melatonin sensitizes H1975 non-small-cell lung cancer cells harboring a T790M-targeted epidermal growth factor receptor mutation to the tyrosine kinase inhibitor gefitinib. *Cell Physiol Biochem* 34: 865-872, 2014.
143. Plaimée P, Weerapreeyakul N, Barusrux S and Johns NP: Melatonin potentiates cisplatin-induced apoptosis and cell cycle arrest in human lung adenocarcinoma cells. *Cell Prolif* 48: 67-77, 2015.
144. Pariente R, Pariente JA, Rodríguez AB and Espino J: Melatonin sensitizes human cervical cancer HeLa cells to cisplatin-induced cytotoxicity and apoptosis: Effects on oxidative stress and DNA fragmentation. *J Pineal Res* 60: 55-64, 2016.
145. Uguz AC, Cig B, Espino J, Bejarano I, Naziroglu M, Rodríguez AB and Pariente JA: Melatonin potentiates chemotherapy-induced cytotoxicity and apoptosis in rat pancreatic tumor cells. *J Pineal Res* 53: 91-98, 2012.
146. Yamanishi M, Narazaki H and Asano T: Melatonin overcomes resistance to clofarabine in two leukemic cell lines by increased expression of deoxycytidine kinase. *Exp Hematol* 43: 207-214, 2015.
147. Ma C, Li LX, Zhang Y, Xiang C, Ma T, Ma ZQ and Zhang ZP: Protective and sensitive effects of melatonin combined with Adriamycin on ER+ (estrogen receptor) breast cancer. *Eur J Gynaecol Oncol* 36: 197-202, 2015.
148. Ju HQ, Li H, Tian T, Lu YX, Bai L, Chen LZ, Sheng H, Mo HY, Zeng JB, Deng W, *et al*: Melatonin overcomes gemcitabine resistance in pancreatic ductal adenocarcinoma by abrogating nuclear factor- κ B activation. *J Pineal Res* 60: 27-38, 2016.
149. Kosar PA, Naziroglu M, Övey IS and Çiğ B: Synergic effects of doxorubicin and melatonin on apoptosis and mitochondrial oxidative stress in MCF-7 breast cancer cells: Involvement of TRPV1 channels. *J Membr Biol* 249: 129-140, 2016.
150. Chuffa LG, Alves MS, Martínez M, Camargo IC, Pinheiro PF, Domeniconi RF, Júnior LA and Martínez FE: Apoptosis is triggered by melatonin in an in vivo model of ovarian carcinoma. *Endocr Relat Cancer* 23: 65-76, 2016.
151. Li W, Fan M, Chen Y, Zhao Q, Song C, Yan Y, Jin Y, Huang Z, Lin C and Wu J: Melatonin induces cell apoptosis in AGS cells through the activation of JNK and P38 MAPK and the suppression of nuclear factor-kappa B: A novel therapeutic implication for gastric cancer. *Cell Physiol Biochem* 37: 2323-2338, 2015.
152. Reiter RJ, Tamura H, Tan DX and Xu XY: Melatonin and the circadian system: Contributions to successful female reproduction. *Fertil Steril* 102: 321-328, 2014.
153. Ma Z, Yang Y, Fan C, Han J, Wang D, Di S, Hu W, Liu D, Li X, Reiter RJ and Yan X: Melatonin as a potential anticarcinogen for non-small-cell lung cancer. *Oncotarget* 7: 46768-46784, 2016.
154. Stover DG and Wagle N: Precision medicine in breast cancer: Genes, genomes, and the future of genomically driven treatments. *Curr Oncol Rep* 17: 15, 2015.
155. Sanchez-Barcelo EJ, Mediavilla MD, Alonso-Gonzalez C and Reiter RJ: Melatonin uses in oncology: Breast cancer prevention and reduction of the side effects of chemotherapy and radiation. *Expert Opin Investig Drugs* 21: 819-831, 2012.