

RESEARCH ARTICLE

Relationship between Tobacco, *cagA* and *vacA i1* Virulence Factors and Bacterial Load in Patients Infected by *Helicobacter pylori*

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Data Availability Statement: The authors confirm that, for approved reasons, some access restrictions apply to the data underlying the findings. The data are available on request from the University of Cantabria Archive (<http://repositorio.unican.es/xmlui/handle/10902/5982>) for researchers who meet the criteria for access to confidential data. Requests may be sent to Professor Miguel Santibáñez (miguel.santibanezm@unican.es) or Juan Carlos Rodríguez (rodriguez_juadia@gva.es). This restriction is due to ethical compliance in order to not compromise study

Abstract

Background and Aim

Several biological and epidemiological studies support a relationship between smoking and *Helicobacter pylori* (*H. pylori*) to increase the risk of pathology. However, there have been few studies on the potential synergistic association between specific *cagA* and *vacA* virulence factors and smoking in patients infected by *Helicobacter pylori*. We studied the relationship between smoking and *cagA*, *vacA i1* virulence factors and bacterial load in *H. pylori* infected patients.

Methods

Biopsies of the gastric corpus and antrum from 155 consecutive patients in whom there was clinical suspicion of infection by *H. pylori* were processed. In 106 patients *H. pylori* infection was detected. Molecular methods were used to quantify the number of microorganisms and presence of *cagA* and *vacA i1* genes. A standardized questionnaire was used to obtain patients' clinical data and lifestyle variables, including tobacco and alcohol consumption. Adjusted Odds Ratios (OR_{adjusted}) were estimated by unconditional logistic regression.

Results

cagA was significantly associated with active-smoking at endoscope: OR_{adjusted} 4.52. Evidence of association was found for *vacA i1* (OR_{adjusted} 3.15). Bacterial load was higher in active-smokers, although these differences did not yield statistical significance (median of 262.2 versus 79.4 copies of *H. pylori* per cell).

participants' privacy, since the dataset is derived from clinical studies involving human participants.

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Conclusions

The association between smoking and a higher risk of being infected by a virulent bacterial population and with higher bacterial load, support a complex interaction between *H. pylori* infection and environmental factors.

Introduction

Interaction between *Helicobacter pylori* (*H. pylori*) and gastric mucosa is a complex phenomenon and as yet little understood. Though in most cases this microorganism does not cause any symptomatic pathology, in some cases it causes serious lesions in the mucosa, being the main risk factor for gastric cancer. This different patterns of evolution may be related to the host's characteristics (genetic, immunological), environmental exposures (socio-sanitary and life-style factors), and also to certain characteristics of the microorganism [1].

Many virulence genes have been described in *H. pylori*, and attempts have been made to associate them with different diseases. One of the most widely studied factors is the virulence *cagA* gene. This gene encodes the CagA protein, which can be directly injected by the bacterium into colonized epithelium via a type IV secretion system, leading to cellular morphological, anti-apoptotic and proinflammatory effects responsible in the long-term (years or decades) for ulcer and cancer [2].

Other virulence gene described in *H. pylori* is *vacA*, which encodes VacA, a vacuolating cytotoxin initially identified for its ability to induce vacuolation in epithelial cells [1,3–5]. Though all strains contain *vacA*, this gene contains three distinct segments that exhibit variation starting within the amino terminus. These areas of variation are broadly defined as the signal (*s*), intermediate (*i*), and middle (*m*) regions, and two or more primary variants have been described for each region: *s1* and *s2* for the signal region, *i1*, *i2*, and *i3* for the intermediate region, and *m1* and *m2* for the middle region [1,5–8]. Several studies have linked strains carrying the *s1-m1* allele of the toxin to more-severe disease outcomes, since these strains show the strongest vacuolating activity to the broadest range of cells [9]. The *i* region is positioned between the *s* and *m* regions and is the most recent region to be described. The *i1* variants of *vacA* have been shown to have stronger vacuolating activity than toxins containing the *i2* regions and recently, the *i* region has been suggested to be a better predictor of disease severity than either the *s* or *m* region, though the *i* region appears to covary with the *s* and *m* regions. This means that the more toxic *i1* region is often associated with *s1-m1* [7].

In addition, some studies have also described an association between the presence of a greater number of microorganisms and more serious gastric lesions with independence of these virulence factors [10,11].

Regarding the host's characteristics, several biological [12,13] and epidemiological studies support a relationship between smoking and *H. pylori* to increase the risk of pathology [14–17].

However, there have been few studies on the potential synergistic association between specific *cagA* and *vacA* virulence factors and tobacco use (chewing or smoking) [18,19,20] and to the best of our knowledge, there are not published studies on the relationship between tobacco use and *H. pylori* bacterial load.

Our aim was to study the relationship between smoking and *cagA*, *vacA i1* virulence factors and bacterial load in *H. pylori* infected patients.

Material and Methods

Ethics Statement

All subjects were informed of the study objectives and gave their informed consent prior to their inclusion in the study. The research protocol was approved by the local ethics and research committees of the participating hospital: "Comité de bioética asistencial (CBA) Dpto Salud 20-H. General" and "Comité ético de investigación clínica (CEIC) del Hospital General Universitario de Elche". Patient records/information was anonymized and de-identified prior to analysis."

Design and participants

Details of this study have been published elsewhere [11,21,22]. Biopsies of the gastric corpus and antrum from 155 consecutive patients in whom there was clinical suspicion of infection by *H. pylori* were processed: duodenal ulcer (n = 67), duodenitis (n = 19), gastric ulcer (n = 36), gastritis (n = 5), dyspepsia (n = 11), family history of gastric cancer (n = 3), and eradication failure (n = 14).

Prospective study including biopsies of the gastric corpus and antrum obtained from the first 155 symptomatic patients who consecutively attended the Endoscopy Unit of Elche University General Hospital for an upper digestive tract endoscopy between January 2009 and June 2010, in whom investigation of *H. pylori* was indicated

Data sources and variables

Measures. A standardized questionnaire was used to obtain patients' clinical data and life-style variables, including tobacco and alcohol consumption. Smoking habit was classified at the time of endoscopy as 'active smokers' versus 'non active smokers'. An 'active smoker' was defined as a smoker at the time the endoscopy was performed or up to a week previously, irrespective of the number of cigarettes smoked per day and for how many years. In a second step, a never smoker was defined as someone having smoked fewer than 100 cigarettes ever or less than one cigarette per day for one year. A former smoker was defined as someone having stopped smoking 1 or more years before the endoscopy.

[S1 Table](#) shows the age, gender, clinical characteristics and lifestyles of the 155 patients in relation to smoking status at endoscopy.

H. pylori infection was detected in 106 patients according to a validated real-time polymerase chain reaction (PCR) [21] that also allows quantifying the number of microorganisms per cell. In 96 and 73 out of these 106 patients, the remains of biopsies allowed us to determine *cagA*, and *vacA i1* genes respectively.

***cagA* determination and *vacA i1* region sequencing.** Detection of the *cagA* gene was done using a real-time PCR system designed in our laboratory [11,21]. After that, we also designed a nested PCR for the amplification of the intermediate region (i) the gene *vacA*, permitting the classification of the gene fragment at i1, i2 and i3 through three clusters (A, B and C) [5]. Primers used in the study of the virulence factors of *H. Pylori* are shown in [S2 Table](#).

Quantification of the number of microorganisms. The quantification of the microorganisms was carried out by amplifying a fragment of the urease gene; the results were expressed as the quotient of the number of microorganisms and the number of human cells in each gastric sample [21].

Statistical analyses

A non-parametric test (U Mann Whitney for independent samples) was used to compare the number of microorganisms in the biopsy and smoking status. In order to estimate the association between *cagA* and *vacA i1* virulence factors, and bacterial load and tobacco smoking, adjusted odds ratios (OR_{adjusted}) and 95% confidence intervals (95%CI) were calculated using unconditional logistic regression. All regression models included as covariates sex, age (< = 50 years, >50 years), average of pure ethanol (g/day) (never, moderate, high), consumption of PPI in the days prior to endoscopy (yes/no), consumption of NSAID (yes/no), and clinical presentation of UDTH (yes/no) entered as indicator variables.

Tests for trend in the ORs across exposure strata were calculated for tobacco smoking ordinal categorised (never, former, current smoker) by using logistic models that included categorical terms as continuous variables in a model with all the potential confounders. For trend-tests, we used the likelihood ratio test statistic with one degree of freedom.

The level of statistical significance was set at 0.05 and all tests were two-tailed. All analyses were performed with SPSS v.21.0.

Results

Table 1 presents the association between *cagA* and tobacco consumption. There was a statistically significant association between *cagA* positive status and active smoking at endoscope (OR_{adjusted} 4.52; 95%CI 1.28–15.98). Table 2 shows the associations between *vacA i1* region status and active smoking at endoscope. *vacA i1* positivity is associated with smoking, though this association does not attain statistical significance (crude OR 2.69; 95% CI 0.92–7.92).

Finally, bacterial load was very asymmetric and presented a high variability. The number of copies of *H. pylori* per cell in antrum according to real-time PCR was higher in active-smokers: Median = 262.2 [Interquartile Range = 2079.0] than in no active-smokers: Median = 79.4 [Interquartile Range = 890.8], although these differences were not statistically significant ($p = 0.35$). Positive associations without yielding statistical significance were found between active smoking and the number of microorganisms when the bacterial load was divided into above and below median: OR_{adjusted} 2.26 (95% CI 0.76–6.75) (see Table 3).

For both *cagA* and *vacA i1* virulence factors and bacterial load, a dose response pattern was obtained when non active smokers were divided into never and former smokers (current smokers were at higher risk than former smokers).

Table 1. Association between *cagA* status and tobacco smoking.

	N <i>cagA</i> neg	N <i>cagA</i> pos	OR	95%	CI	OR _{adjusted} ^a	95%	CI
Smoking Status at endoscope								
Non active smokers	34	31	1.00			1.00		
Active smokers	8	23	3.15	1.23	8.07	4.52	1.28	15.98
Smoking Status								
Never	22	23	1.00			1.00		
Former smoker	12	8	0.64	0.22	1.86	0.36	0.09	1.53
Current smoker	8	23	2.75	1.02	7.43	3.24	0.84	12.47
<i>p</i> -value for linear trend				0.067			0.123	

^aAdjusted for sex, age (< = 50 years, >50 years), average of pure ethanol (g/day) (Never, moderate, high), consumption of proton pump inhibitors in the days prior to endoscopy (Yes/No), consumption of non-steroid anti-inflammatory drugs (Yes/No), and clinical presentation of upper digestive tract haemorrhage (Yes/No).

Table 2. Association between *vacA i1* intermediate region status and tobacco smoking.

	N <i>vacA</i> Region_i1neg	N <i>vacA</i> Region_i1 pos	OR	95% CI	OR _{adjusted} ^a	95% CI
Smoking status at endoscope						
Non active smokers	21	26	1.00		1.00	
Active smokers	6	20	2.69	0.92–7.92	3.15	0.74–13.39
Smoking status						
Never	12	21	1.00		1.00	
Former smoker	9	5	0.32	0.09–1.17	0.38	0.07–2.13
Current smoker	6	20	1.91	0.60–6.05	2.20	0.46–10.58
<i>p</i> -value for linear trend				0.356		0.351

^aAdjusted for sex, age (< = 50 years, >50 years), average of pure ethanol (g/day) (Never, moderate, high), consumption of proton pump inhibitors in the days prior to endoscopy (Yes/No), consumption of non-steroid anti-inflammatory drugs (Yes/No), and clinical presentation of upper digestive tract haemorrhage (Yes/No).

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Discussion

Overall, in our study, when restricting to *H. pylori* infected patients, active smokers were at a higher risk of colonization by *cagA* positive virulent strains at the time of endoscope. Although results did not yield statistical significance, active smoking was also associated to *vacA i1* virulent strains and higher bacterial loads.

First contact with *H. pylori* is documented by a transmission of the bacteria within families, predominantly in the early childhood [23,24]. However, it has been found that the genome of *H. pylori* changes continuously during chronic colonization of an individual host with mixed strains by importing small pieces of foreign DNA from other strains during persistent or transient mixed infections [25–27].

The co-colonization of *H. pylori* could permit inter-strain DNA transfer, which could speed up the emergence of new strains/subpopulations that are better adapted or more virulent in a given human host than the initial infecting strains [25, 28–30].

The higher bile salt reflux and gastric bile salt and the lower concentrations of vitamin C in gastric juices [12,13] in smokers in comparison with non-smokers support a biological

Table 3. Association between bacterial load and tobacco smoking.

	<i>H. pylori</i> bacterial load		OR	95% CI	CI	OR _{adjusted} ^a	95% CI	CI
	≤ Median (N)	> Median (N)						
Smoking Status at endoscope								
Non active smokers	39	33	1.00			1.00		
Active smokers*	14	20	1.69	0.74–3.85		2.26	0.76–6.75	
Smoking status								
Never	28	21	1.00			1.00		
Formersmoker	11	12	1.45	0.54–3.93		1.29	0.34–4.90	
Current smoker	14	20	1.90	0.78–4.62		2.47	0.76–8.06	
<i>p</i> -value for linear trend				0.150			0.137	

^aAdjusted for sex, age (< = 50 years, >50 years), average of pure ethanol (g/day) (Never, moderate, high), consumption of proton pump inhibitors in the days prior to endoscopy (Yes/No), consumption of non-steroid anti-inflammatory drugs (Yes/No), and clinical presentation of upper digestive tract haemorrhage (Yes/No).

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plausibility on the relationship between smoking and a higher prevalence of new more virulent strains with higher bacterial loads.

A growing number of studies have begun to suggest that *vacA* and *cagA* interact in such a way as to affect disease severity [31–34]. In a previous work we found that the *cagA* gene was associated with high bacterial load with a clear, statistically significant dose-response relationship between load and presence of *cagA*. These data suggest that both parameters combine to produce clinically significant lesions, as compared with strains of low virulence that multiply very little in the gastric mucosa and so do not cause significant damage [11].

Our present data suggest that smoking could be a synergistic factor, interacting with both bacterial load and *H. pylori* virulence factors. Published studies would support an interaction between high virulence factors and smoking to increase the risk of intestinal metaplasia [14,18] and gastric cancer [15–17,19]. Recently, an association between *cagA* and other types of tobacco use such as tobacco-chewing has been also documented [20].

Since a synergistic effect seems plausible, the presence of high virulence strains of *H. pylori* should be included as a confusion variable and as a term of interaction in the associations traditionally established between smoking and gastric cancer [35,36].

If this hypothesised synergistic effect is firmly established, smoking cessation would be associated with a lower risk of high virulence strains infection, so the combined effect of smoking and *H. Pylori* trends on past and future gastric cancer incidence, could be higher than estimated by microsimulation models [37].

Up to our knowledge, this is the first study on the specific relationship between smoking and bacterial load. Since our results did not yield statistical significance, because of the asymmetry and high variability of bacterial load, further prospective studies with higher sample size and statistical power are necessary to corroborate these findings.

Number of CagA Glu-Pro-Ile-Tyr-Ala (EPIYA)-C segments (with multiple repeats increasing the virulence) may explain, to some extent, geographic differences in the incidence of gastric cancer in Western countries. It would be interesting to go in depth in relation to the association between smoking and these specific patterns of *cagA* gene repeat sequences. Further studies are also required to continue studying the relationship between tobacco and *vacA* alleles, as well as other more recently discovered virulence factors [38,39].

In conclusion, the associations between smoking and a higher prevalence of virulent bacterial population with higher bacterial loads, support a complex interaction between the microorganism, gastric mucosa and environmental factors such as tobacco smoking, in which synergistic effects could increase gastric pathology.

Supporting Information

S1 Table. Baseline characteristics of patients as a function of Smoking Status at endoscope.

^aSD denotes Standard Deviation ^bAlcoholism (high alcohol consumption) was defined as a consumption of more than 80g/day in men and 60 g/day in women ^cA Coffee/Tea Drinker at the endoscope was defined as someone having consumed at least two cups at week for at least one year. ^dComorbidity was defined as someone having high blood pressure, diabetes, cardiopathy, chronic obstructive pulmonary disease, or kidney failure.

(PDF)

S2 Table. Primers used in the study of the virulence factors of *H. pylori*.

(PDF)

Author Contributions

Conceived and designed the experiments: MS SB JCR GR. Performed the experiments: MS EA SB NA JS JCR AG JSV MRG MPZ RSL AB ELG EP CS GR. Analyzed the data: MS SB JCR GR. Contributed reagents/materials/analysis tools: MS SB JCR GR. Wrote the paper: MS EA SB NA JS JCR AG JSV MRG MPZ RSL AB ELG EP CS GR.

References

1. Polk DB, Peek RM Jr. *Helicobacter pylori*: gastric cancer and beyond. *Nat Rev Cancer*. 2010; 10:403–14. doi: [10.1038/nrc2857](https://doi.org/10.1038/nrc2857) PMID: [20495574](https://pubmed.ncbi.nlm.nih.gov/20495574/)
2. Oldani A, Cormont M, Hofman V, Chiozzi V, Oregioni O, Canonici A, et al. *Helicobacter pylori* counteracts the apoptotic action of its VacA toxin by injecting the CagA protein into gastric epithelial cells. *PLoS Pathog*. 2009; 5:e1000603. doi: [10.1371/journal.ppat.1000603](https://doi.org/10.1371/journal.ppat.1000603) PMID: [19798427](https://pubmed.ncbi.nlm.nih.gov/19798427/)
3. Atherton JC, Cover TL, Twells RJ, Morales MR, Hawkey CJ, Blaser MJ. Simple and accurate PCR-based system for typing vacuolating cytotoxin alleles of *Helicobacter pylori*. *J Clin Microbiol*. 1999; 37: 2979–2982. PMID: [10449485](https://pubmed.ncbi.nlm.nih.gov/10449485/)
4. Cover TL, Blanke SR. *Helicobacter pylori* VacA, a paradigm for toxin multifunctionality. *Nat Rev Microbiol*. 2005; 3: 320–332. PMID: [15759043](https://pubmed.ncbi.nlm.nih.gov/15759043/)
5. Jones KR, Jang S, Chang JY, Kim J, Chung IS, Olsen CH, et al. Polymorphisms in the intermediate region of VacA impact *Helicobacter pylori*-induced disease development. *J Clin Microbiol*. 2011; 49: 101–110. doi: [10.1128/JCM.01782-10](https://doi.org/10.1128/JCM.01782-10) PMID: [21084502](https://pubmed.ncbi.nlm.nih.gov/21084502/)
6. Atherton JC, Cao P, Peek RM Jr, Tummuru MK, Blaser MJ, Cover TL. Mosaicism in vacuolating cytotoxin alleles of *Helicobacter pylori*. Association of specific vacA types with cytotoxin production and peptic ulceration. *J Biol Chem*. 1995; 270: 17771–17777. PMID: [7629077](https://pubmed.ncbi.nlm.nih.gov/7629077/)
7. Rhead JL, Letley DP, Mohammadi M, Hussein N, Mohagheghi MA, Eshagh Hosseini M, et al. A new *Helicobacter pylori* vacuolating cytotoxin determinant, the intermediate region, is associated with gastric cancer. *Gastroenterology*. 2007; 133: 926–936. PMID: [17854597](https://pubmed.ncbi.nlm.nih.gov/17854597/)
8. Chung C, Olivares A, Torres E, Yilmaz O, Cohen H, Perez-Perez G. Diversity of VacA intermediate region among *Helicobacter pylori* strains from several regions of the world. *J Clin Microbiol*. 2010; 48: 690–696. doi: [10.1128/JCM.01815-09](https://doi.org/10.1128/JCM.01815-09) PMID: [20053862](https://pubmed.ncbi.nlm.nih.gov/20053862/)
9. Letley DP, Rhead JL, Twells RJ, Dove B, Atherton JC. Determinants of non-toxicity in the gastric pathogen *Helicobacter pylori*. *J Biol Chem*. 2003; 278: 26734–26741. PMID: [12738773](https://pubmed.ncbi.nlm.nih.gov/12738773/)
10. Lai YC, Wang TH, Huang SH, Yang SS, Wu CH, Chen TK, et al. Density of *Helicobacter pylori* may affect the efficacy of eradication therapy and ulcer healing in patients with active duodenal ulcers. *World J Gastroenterol*. 2003; 9: 1537–1540. PMID: [12854158](https://pubmed.ncbi.nlm.nih.gov/12854158/)
11. Belda S, Saez J, Santibáñez M, Rodríguez JC, Sola-Vera J, Ruiz-García M, et al. Relationship between bacterial load, morbidity and cagA gene in patients infected by *Helicobacter pylori*. *Clin Microbiol Infect*. 2012; 18: E251–3. doi: [10.1111/j.1469-0691.2012.03884.x](https://doi.org/10.1111/j.1469-0691.2012.03884.x) PMID: [22551001](https://pubmed.ncbi.nlm.nih.gov/22551001/)
12. Sobala GM, O'Connor HJ, Dewar EP, King RF, Axon AT, Dixon MF. Bile reflux and intestinal metaplasia in gastric mucosa. *J Clin Pathol*. 1993; 46: 235–40. PMID: [8463417](https://pubmed.ncbi.nlm.nih.gov/8463417/)
13. Jarosz M, Dzieniszewski J, Dabrowska-Ufniarz E, Wartanowicz M, Ziemiński S. Tobacco smoking and vitamin C concentration in gastric juice in healthy subjects and patients with *Helicobacter pylori* infection. *Eur J Cancer Prev*. 2000; 9: 423–428. PMID: [11201681](https://pubmed.ncbi.nlm.nih.gov/11201681/)
14. Russo A, Maconi G, Spinelli P, Felice GD, Eboli M, Andreola S, et al. Effect of lifestyle, smoking, and diet on development of intestinal metaplasia in *H. pylori*-positive subjects. *Am J Gastroenterol*. 2001; 96: 1402–1408. PMID: [11374674](https://pubmed.ncbi.nlm.nih.gov/11374674/)
15. Simán JH, Forsgren A, Berglund G, Florén CH. Tobacco smoking increases the risk for gastric adenocarcinoma among *Helicobacter pylori*-infected individuals. *Scand J Gastroenterol*. 2001; 36: 208–13. PMID: [11252415](https://pubmed.ncbi.nlm.nih.gov/11252415/)
16. Brenner H, Arndt V, Bode G, Stegmaier C, Ziegler H, Stümer T. Risk of gastric cancer among smokers infected with *Helicobacter pylori*. *Int J Cancer*. 2002; 98: 446–449. PMID: [11920598](https://pubmed.ncbi.nlm.nih.gov/11920598/)
17. Machida-Montani A, Sasazuki S, Inoue M, Natsukawa S, Shaura K, Koizumi Y, et al. Association of *Helicobacter pylori* infection and environmental factors in non-cardia gastric cancer in Japan. *Gastric Cancer*. 2004; 7: 46–53. PMID: [15052440](https://pubmed.ncbi.nlm.nih.gov/15052440/)
18. Peleteiro B, Lunet N, Figueiredo C, Carneiro F, David L, Barros H. Smoking, *Helicobacter pylori* virulence, and type of intestinal metaplasia in Portuguese males. *Cancer Epidemiol Biomarkers Prev*. 2007; 16: 322–6. PMID: [17301266](https://pubmed.ncbi.nlm.nih.gov/17301266/)

19. Wang XQ, Yan H, Terry PD, Wang JS, Cheng L, Wu WA, et al. Interactions between CagA and smoking in gastric cancer. *World J Gastroenterol*. 2011; 17: 3330–3334. doi: [10.3748/wjg.v17.i28.3330](https://doi.org/10.3748/wjg.v17.i28.3330) PMID: [21876621](https://pubmed.ncbi.nlm.nih.gov/21876621/)
20. Pandey A, Tripathi SC, Mahata S, Vishnoi K, Shukla S, Misra SP, et al. Carcinogenic Helicobacter pylori in gastric pre-cancer and cancer lesions: association with tobacco-chewing. *World J Gastroenterol*. 2014; 20: 6860–6968. doi: [10.3748/wjg.v20.i22.6860](https://doi.org/10.3748/wjg.v20.i22.6860) PMID: [24944476](https://pubmed.ncbi.nlm.nih.gov/24944476/)
21. Belda S, Saez J, Santibáñez M, Rodríguez JC, Galiana A, Sola-Vera J, et al. Quantification of Helicobacter pylori in gastric mucosa by real-time polymerase chain reaction: comparison with traditional diagnostic methods. *Diagn Microbiol Infect Dis*. 2012; 74: 248–252. doi: [10.1016/j.diagmicrobio.2012.07.007](https://doi.org/10.1016/j.diagmicrobio.2012.07.007) PMID: [22921814](https://pubmed.ncbi.nlm.nih.gov/22921814/)
22. Saez J, Belda S, Santibáñez M, Rodríguez JC, Sola-Vera J, Galiana A, et al. Real-time PCR for diagnosing Helicobacter pylori infection in patients with upper gastrointestinal bleeding: comparison with other classical diagnostic methods. *J Clin Microbiol*. 2012; 50: 3233–3237. doi: [10.1128/JCM.01205-12](https://doi.org/10.1128/JCM.01205-12) PMID: [22837325](https://pubmed.ncbi.nlm.nih.gov/22837325/)
23. Drumm B, Perez-Perez GI, Blaser MJ, Sherman PM. Intrafamilial clustering of Helicobacter pylori infection. *N Engl J Med*. 1990; 322: 359–363. PMID: [2300088](https://pubmed.ncbi.nlm.nih.gov/2300088/)
24. Kivi M, Johansson AL, Reilly M, Tindberg Y. Helicobacter pylori status in family members as risk factors for infection in children. *Epidemiol Infect*. 2005; 133: 645–652. PMID: [16050509](https://pubmed.ncbi.nlm.nih.gov/16050509/)
25. Suerbaum S, Smith JM, Bapumia K, Morelli G, Smith NH, Kunstmann E, et al. Free recombination within Helicobacter pylori. *Proc Natl Acad Sci U S A*. 1998; 95: 12619–12624. PMID: [9770535](https://pubmed.ncbi.nlm.nih.gov/9770535/)
26. Falush D, Kraft C, Taylor NS, Correa P, Fox JG, Achtman M, et al. Recombination and mutation during long-term gastric colonization by Helicobacter pylori: estimates of clock rates, recombination size, and minimal age. *Proc Natl Acad Sci U S A*. 2001; 98: 15056–15061. PMID: [11742075](https://pubmed.ncbi.nlm.nih.gov/11742075/)
27. Kennemann L, Didelot X, Aebischer T, Kuhn S, Drescher B, Droege M, et al. Helicobacter pylori genome evolution during human infection. *Proc Natl Acad Sci U S A*. 2011; 108: 5033–5038. doi: [10.1073/pnas.1018444108](https://doi.org/10.1073/pnas.1018444108) PMID: [21383187](https://pubmed.ncbi.nlm.nih.gov/21383187/)
28. Kersulyte D, Chalkauskas H, Berg DE. Emergence of recombinant strains of Helicobacter pylori during human infection. *Mol Microbiol*. 1999; 31: 31–43. PMID: [9987107](https://pubmed.ncbi.nlm.nih.gov/9987107/)
29. Cellini L, Di Campli E, Di Candia M, Marzio L. Molecular fingerprinting of Helicobacter pylori strains from duodenal ulcer patients. *Lett Appl Microbiol*. 2003; 36: 222–226. PMID: [12641715](https://pubmed.ncbi.nlm.nih.gov/12641715/)
30. Cellini L, Grande R, Di Campli E, Di Bartolomeo S, Capodicasa S, Marzio L. Analysis of genetic variability, antimicrobial susceptibility and virulence markers in Helicobacter pylori identified in Central Italy. *Scand J Gastroenterol*. 2006; 41: 280–287. PMID: [16497614](https://pubmed.ncbi.nlm.nih.gov/16497614/)
31. Van Doorn LJ, Figueiredo C, Mégraud F, Pena S, Midolo P, Queiroz DM, et al. Geographic distribution of vacA allelic types of Helicobacter pylori. *Gastroenterology*. 1999; 116: 823–830. PMID: [10092304](https://pubmed.ncbi.nlm.nih.gov/10092304/)
32. Yamazaki S, Yamakawa A, Okuda T, Ohtani M, Suto H, Ito Y, et al. Distinct diversity of vacA, cagA, and cagE genes of Helicobacter pylori associated with peptic ulcer in Japan. *J Clin Microbiol*. 2005; 43: 3906–3916. PMID: [16081930](https://pubmed.ncbi.nlm.nih.gov/16081930/)
33. Argent RH, Thomas RJ, Letley DP, Rittig MG, Hardie KR, Atherton JC. Functional association between the Helicobacter pylori virulence factors VacA and CagA. *J Med Microbiol*. 2008; 57: 145–150. doi: [10.1099/jmm.0.47465-0](https://doi.org/10.1099/jmm.0.47465-0) PMID: [18201978](https://pubmed.ncbi.nlm.nih.gov/18201978/)
34. Jang S, Jones KR, Olsen CH, Joo YM, Yoo YJ, Chung IS, et al. Epidemiological link between gastric disease and polymorphisms in VacA and CagA. *J Clin Microbiol*. 2010; 48: 559–567. doi: [10.1128/JCM.01501-09](https://doi.org/10.1128/JCM.01501-09) PMID: [19955279](https://pubmed.ncbi.nlm.nih.gov/19955279/)
35. Trédaniel J, Boffetta P, Buiatti E, Saracci R, Hirsch A. Tobacco smoking and gastric cancer: review and meta-analysis. *Int J Cancer*. 1997; 72: 565–73. PMID: [9259392](https://pubmed.ncbi.nlm.nih.gov/9259392/)
36. González CA, Pera G, Agudo A, Palli D, Krogh V, Vineis P, et al. Smoking and the risk of gastric cancer in the European Prospective Investigation Into Cancer and Nutrition (EPIC). *Int J Cancer*. 2003; 107: 629–634. PMID: [14520702](https://pubmed.ncbi.nlm.nih.gov/14520702/)
37. Yeh JM, Hur C, Schrag D, Kuntz KM, Ezzati M, Stout N, et al. Contribution of H. pylori and smoking trends to US incidence of intestinal-type noncardia gastric adenocarcinoma: a microsimulation model. *PLoS Med*. 2013; 10:e1001451. doi: [10.1371/journal.pmed.1001451](https://doi.org/10.1371/journal.pmed.1001451) PMID: [23700390](https://pubmed.ncbi.nlm.nih.gov/23700390/)
38. Vannini A, Roncarati D, Spinsanti M, Scarlato V, Danielli A. In depth analysis of the Helicobacter pylori cag pathogenicity island transcriptional responses. *PLoS One*. 2014; 9: e98416. doi: [10.1371/journal.pone.0098416](https://doi.org/10.1371/journal.pone.0098416) PMID: [24892739](https://pubmed.ncbi.nlm.nih.gov/24892739/)
39. Roesler BM, Rabelo-Gonçalves EM, Zeitune JM. Virulence Factors of Helicobacter pylori: A Review. *Clin Med Insights Gastroenterol*. 2014; 7: 9–17. doi: [10.4137/CGast.S13760](https://doi.org/10.4137/CGast.S13760) PMID: [24833944](https://pubmed.ncbi.nlm.nih.gov/24833944/)