

PRELIMINARY RESULTS ON THE INVERTEBRATE FAUNA (ARANEAE, ORTHOPTERA, HETEROPTERA AND HYMENOPTERA: FORMICIDAE) OF ALKALINE GRASSLANDS OF THE HUNGARIAN-ROMANIAN BORDER

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Introduction

Alkaline grasslands, which have been present in the Pannonian Basin since the Pleistocene (Kun 1998, Molnár and Borhidi 2003), belong to the most typical communities in this region (Illyés et al. 2007). In Hungary, alkaline grasslands form the third part of grassland habitats. Only a small proportion of these are of ancient origin, most of them are secondary, and originated mainly as the result of river regulations and drainages in the 19th and 20th century (Kun 1998). According to Molnár and Borhidi (2003) about 40 percent of alkaline grasslands can be considered as natural or semi-natural habitats from the point of view of nature conservation.

In Hungary, a total of 54 plant communities are known in saline habitats, which are though relatively species-poor but have characteristic and manifold species composition (Tóth and Szendrei 2006). The vegetation pattern of alkaline grasslands strongly depends on soil salinity, salt quality, depth of maximum salt content and water availability. The typical zonations of saline vegetation are *Artemisia* salt steppe, alkaline berm ("szikpadka"), Pannonic *Camphorosma* hollow, dense and tall *Puccinellia* sward, alkaline vein ("szikér"), salt meadow and salt marsh (Molnár and Borhidi 2003).

Due to the exceptionally rich fauna and mosaic flora with several endemics and subendemics (Kelemen, 1997), alkaline grasslands are valuable from a nature conservation perspective.

In 2010 a faunistic survey was carried out in order to compare the invertebrate fauna of two neighbouring alkaline grasslands separated by the Hungarian-Romanian border. Preliminary results of the study are presented in the following.

Materials and methods

The study was carried out in the border region of Gyula, Hungary and Vărşand, Romania. Data were collected with a variety of collecting methods in the following habitat types:

Gyula I: (1) loess steppe and salt meadow; (2) salt meadow; (3) salt meadow and *Artemisia* salt steppe; (4) loess steppe; (5) Pannonic *Camphorosma* hollow and dense and tall *Puccinellia* sward with salt meadow; (6) salt meadow; (7) *Artemisia* salt steppe with dense and tall *Puccinellia* sward patches and Pannonic *Camphorosma* hollow; (8) loess steppe patches; (9) transition from *Artemisia* salt steppe to dense and tall *Puccinellia* sward; (10) salt meadow.

Sampling was performed with pitfall trapping and sweep netting during summer 2010. At each habitat 5-5 pitfall traps (500 ml plastic jars filled with ethylene-glycol) were set at intervals of approximately 4 m. Trapping was continuous from 1 June to 22 September 2010. 5 sweep net samples (each of them consisted of 50 sweeps) were taken at each habitat in 1 June, 5 July, 4 August and 22 September. Net contents were emptied into sealable plastic bags filled with some ethyl alcohol.

Sweep netting (5 × 50 sweeps) was also employed in the following sites and habitat types:

Gyula II: (1) salt meadow with loess steppe and *Artemisia* salt steppe; (2) salt meadow; (3) new abandonment on arable lands and salt meadow.

Gyula III: (1) salt meadow and *Artemisia* salt steppe; (2) *Artemisia* salt steppe with salt meadow patches.

Szabadkigyós: (1) *Artemisia* salt steppe; (2) salt marsh.

Elek: (1) loess steppe; (2) salt meadow with Pannonic *Camphorosma* hollow, dense and tall *Puccinellia* sward and *Artemisia* salt steppe patches; (3) loess steppe; (4) *Artemisia* salt steppe.

Kétegyháza: (1) salt meadow and salt marsh; (2) salt meadow.

Vărşand I: (1) uncharacteristic grassland; (2) *Achillea* salt steppe with loess steppe patches; (3) loess steppe; (4) *Artemisia* salt steppe with salt meadow patches and salt meadow with *Artemisia* salt steppe patches.

Pilu: (1) uncharacteristic grassland (or degraded loess steppe); (2) *Achillea* salt steppe with *Artemisia* salt steppe patches; (3) degraded loess steppe; (4) *Artemisia* salt steppe with salt meadow patches and Pannonic *Camphorosma* hollow; (5) *Artemisia* salt steppe and salt meadow with loess steppe patches; (6) salt meadow with uncharacteristic grassland.

Pitfall trapping and D-vac sampling (using a Stihl® BG56 Leaf Blower/VAC vacuum sampling device) were conducted in the following sites and habitat types:

Gyula IV: (1) salt meadow; (2) salt steppe with uncharacteristic vegetation.

Várşand II: (1) degraded loess steppe; (2) *Artemisia* salt steppe; (3) hygrophile meadow close to an irrigation canal; (4) salt meadow with ruderal plants.

D-vac sampling consisted of 25 sample units of 1 m² with a ca. 5-minutes vacuum time per habitat. Sampling was performed in 4 July, 7 August, 22 August and 27 September.

Spiders were determined according to Heimer and Nentwig (1991), Loksa (1969, 1972), Roberts (1985, 1987), Nentwig et al. (2003), Fuhn and Niculescu-Burlacu (1971) and Sterghiu (1985). Orthopteran specimens were identified using the keys of Kis (1976, 1978), Harz (1957), Móczár (1969) and Knechtel and Popovici-Biznosanu (1959). True bugs were identified according to Benedek (1969), Halászfy (1959), Kis (1984, 2001), Kis and Kondorosy (1999), Vásárhelyi (1978, 1983) and Wagner (1952, 1966, 1967). Ant specimens were determined using the keys of Seifert (1988, 1997, 2007), Czechowski et al. (2002), Csósz (1999) and Kutter (1977).

Results and discussion

Araneae

A total number of 1541 spider individuals (1344 adult and 197 juvenile) of 97 species were identified from pitfall trap and D-vac samples (Table 1). Among the rare species we can mention *Urocoras longispinus* (Kulczynski, 1897).

We collected several agrobiont and agrophile species. *Pardosa agrestis* (Westring, 1862), *Meioneta rurestris* (C.L. Koch, 1836), *Alopecosa pulverulenta* (Clerck, 1757), *Trochosa ruricola* (De Geer, 1778) and *Erigone dentipalpis* (Wider, 1834) are known to occur at agroecosystems and disturbed habitat types (Hänggi et al. 1995, Bogya and Markó 1999, Kiss and Samu 2000, Samu and Szinetár 2002). The occurrence and the high numbers of collected individuals of these species are presumably brought about not the influence of the fauna of the surrounding arable fields. According to Wissinger (1997) the agrobiont fauna consists of species adapted to predictably ephemeral habitats, they evolved the "cyclic colonization" strategy from natural or semi-natural refuges. According to this theory the agrobiont fauna possibly originated from regularly disturbed habitat types such as the annually inundated alkaline grasslands (Szita et al 1998, 2002, Samu and Szinetár 2000).

There are several data on the spider fauna of the alkaline grasslands of this region (Szita et al. 1998, 1999, 2000). The previously little known gnaphosid spider was recently found also at saline steppes and salt marsh meadows (Dudás 2001, Szita et al. 2000).

Table 1. List of spider species collected from the study sites.

Family: Uloboridae	
<i>Uloborus walckenaerius</i> (Latreille, 1806)	6
Family: Theridiidae	
<i>Episimus truncatus</i> (Latreille, 1809)	12
<i>Euryopsis quinqueguttata</i> Thorell, 1875	6
<i>Neottiura suaveolens</i> (Simon, 1879)	2
<i>Phylloneta impressa</i> (L.Koch, 1881)	1
<i>Simitidion simile</i> (C.L.Koch, 1836)	1
<i>Steatoda phalerata</i> (Panzcr, 1801)	3
Family: Linyphiidae	
<i>Agyneta</i> sp.	1
<i>Ceratinella brevis</i> (Wider, 1834)	4
<i>Cresmatoneta mutinensis</i> (Canestrini, 1868)	5
<i>Diplostyla concolor</i> (Wider, 1834)	16
<i>Erigone dentipalpis</i> (Wider, 1834)	2
<i>Goniatium rubens</i> (Blackwall, 1833)	1
<i>Linyphia hortensis</i> (Sundevall, 1830)	12
<i>Linyphia triangularis</i> (Clerck, 1757)	4
<i>Meioneta rurestris</i> (C.L.Koch, 1836)	74
<i>Micrargus apertus</i> (O.P. –Cambridge, 1881)	1
<i>Neriere peltata</i> (Wider, 1834)	3
<i>Trichoncus hackmani</i> Millidge, 1956	2
<i>Walckenaeria capito</i> (Westring, 1861)	6
<i>Walckenaeria</i> sp.	1
Family: Tetragnathidae	
<i>Pachygnatha clercki</i> (Sundevall, 1823)	1
<i>Pachygnatha degeeri</i> Sundevall, 1830	3
<i>Tetragnatha extensa</i> (Linnaeus, 1758)	5
Family: Araneidae	
<i>Araneus quadratus</i> (Clerck, 1757)	3
<i>Araneus diadematus</i> Clerck, 1757	24
<i>Argiope bruennichi</i> (Scopoli, 1772)	38
<i>Mangora acalypha</i> (Walckenaer, 1802)	2
Family: Lycosidae	
<i>Alopecosa accentuata</i> (Latreille, 1817)	1
<i>Alopecosa cuneata</i> (Clerck, 1757)	3
<i>Alopecosa pulverulenta</i> (Clerck, 1757)	15
<i>Arctosa leopardus</i> (Sundevall, 1833)	1
<i>Aulonia albimana</i> (Walckenaer, 1805)	285
<i>Hogna radiata</i> (Latreille, 1817)	6

<i>Pardosa agrestis</i> (Westring, 1862)	31
<i>Pardosa amentata</i> (Clerck, 1757)	3
<i>Pardosa hortensis</i> (Thorell, 1872)	1
<i>Pardosa prativaga</i> (L. Koch, 1870)	27
<i>Pirata latitans</i> (Blackwall, 1841)	1
<i>Pirata uliginosus</i> (Thorell, 1856)	5
<i>Trochosa robusta</i> (Simon, 1876)	99
<i>Trochosa ruricola</i> (De Geer, 1778)	9
<i>Trochosa terricola</i> Thorell, 1856	22
<i>Xerolycosa miniata</i> (C.L. Koch, 1834)	2
Family: Pisauridae	
<i>Pisaura mirabilis</i> (Clerck, 1757)	36
Family: Oxyopidae	
<i>Oxyopes heterophthalmus</i> (Latreille, 1804)	1
Family: Titanoecidae	
<i>Titanoeca veteranica</i> Herman, 1879	1
Family: Liocranidae	
<i>Agroeca lusatica</i> (L. Koch, 1875)	1
<i>Liocranoeca striata</i> (Kulczynski, 1882)	2
Family: Corinnidae	
<i>Phrurolithus festivus</i> (C.L. Koch, 1835)	107
<i>Phrurolithus minimus</i> C.L. Koch, 1839	35
Family: Agelenidae	
<i>Agelena labyrinthica</i> (Clerck, 1757)	3
Family: Dictynidae	
<i>Dictyna arundinacea</i> (Linnaeus, 1758)	11
<i>Dictyna latens</i> (Fabricius, 1775)	3
Family: Miturgidae	
<i>Cheiracanthium punctorium</i> (Villers, 1789)	25
Family: Gnaphosidae	
<i>Drassodes pubescens</i> (Thorell, 1856)	9
<i>Drassyllus praeficus</i> (L. Koch, 1866)	38
<i>Drassyllus pusillus</i> (C.L. Koch, 1833)	4
<i>Gnaphosa lucifuga</i> (Walckenaer, 1802)	2
<i>Gnaphosa rufula</i> (L. Koch, 1866)	5
<i>Haplodrassus minor</i> (O.P.-Cambridge, 1879)	38
<i>Haplodrassus signifer</i> (C.L. Koch, 1839)	11
<i>Micaria formicaria</i> (Sundevall, 1832)	15
<i>Micaria pulicaria</i> (Sundevall, 1832)	1
<i>Micaria</i> sp.	1
<i>Trachyzelotes pedestris</i> (C.L. Koch, 1837)	56
<i>Zelotes electus</i> (C.L. Koch, 1839)	20

<i>Zelotes gracilis</i> Canestrini, 1868	3
<i>Zelotes hermani</i> (Chyzer, 1878)	5
<i>Zelotes latreillei</i> (Simon, 1878)	47
<i>Zelotes</i> sp.	1
Family: Sparassidae	
<i>Micrommata virescens</i> (Clerck, 1757)	1
Family: Zoridae	
<i>Zora spinimana</i> (Sundevall, 1833)	16
Family: Thomisidae	
<i>Misumena vatia</i> (Clerck, 1757)	2
<i>Ozyptila pullata</i> (Thorell, 1875)	3
<i>Ozyptila simplex</i> (O.P.-Cambridge, 1862)	3
<i>Synema globosum</i> (Fabricius, 1775)	1
<i>Thomisus onustus</i> (Walckenaer, 1806)	3
<i>Xysticus cristatus</i> (Clerck, 1757)	1
<i>Xysticus erraticus</i> (Blackwall, 1834)	1
<i>Xysticus kochi</i> (Thorell, 1872)	2
Family: Amaurobiidae	
<i>Urocoras longispinus</i> (Kulczynski, 1897)	1
Family: Philodromidae	
<i>Philodromus aureolus</i> (Clerck, 1757)	3
<i>Philodromus fuscomarginatus</i> (De Geer, 1778)	1
<i>Philodromus margaritatus</i> (Clerck, 1757)	1
<i>Thanatus arenarius</i> Thorell, 1872	23
<i>Thanatus formicinus</i> (Clerck, 1757)	1
<i>Tibellus oblongus</i> (Walckenaer, 1802)	13
Family: Salticidae	
<i>Euophrys frontalis</i> (Walckenaer, 1802)	3
<i>Heliophanus cupreus</i> (Walckenaer, 1802)	1
<i>Heliophanus flavipes</i> (Hahn, 1832)	1
<i>Leptorchestes berolinensis</i> (C.L.Koch, 1846)	1
<i>Macaroeris nidicolens</i> (Walckenaer, 1802)	1
<i>Mithion canestrini</i> (Ninni, 1868)	10
<i>Pellenes nigrociliatus</i> (Simon, 1875)	2
<i>Phlegra fasciata</i> (Hahn, 1826)	17
<i>Talavera aequipes</i> (O.P.-Cambridge, 1871)	1
Total:	1344

A total of 7750 orthopteran specimens (5142 adult and 2608 juvenile) were identified from the pitfall trap, sweep netting and D-vac samples, which represent 42 species (Table 2).

The most abundant species was clearly *Melanogryllus desertus*, followed by *Euchorthippus declivus* and *Tartarogryllus burdigalensis*.

The number of species was the highest in the mosaics of loess steppes (21 species), saline meadows (22 species) and saline meadows with *Artemisia* salt steppe patches (18 species). We found two species, *Acrida hungarica* and *Epacromius coerulipes*, which are protected in Hungary. In addition, we collected two sporadic and six rare species.

Table 2. List of orthopteran species occurring in the study sites. ^P: protected species in Hungary, *: sporadic species in Hungary, **: rare species in Hungary.

Order: Ensifera	
Superfamily: Tettigonioidea	
<i>Conocephalus discolor</i> Thunberg, 1815	36
<i>Decticus verrucivorus</i> (Linnaeus, 1758)	2
<i>Leptophyes albobittata</i> (Kollar, 1833)	13
<i>Leptophyes discoidalis</i> (Frivaldsky, 1867)	1
<i>Metrioptera bicolor</i> (Philippi, 1830)	25
<i>Metrioptera roeseltii</i> (Hagenbach, 1822)	10
<i>Phaneroptera falcata</i> (Poda, 1761)	2
<i>Platycleis affinis</i> Fieber, 1853	20
<i>Platycleis grisea</i> (Fabricius, 1781)	11
<i>Platycleis intermedia</i> (Serville, 1838)	1
<i>Platycleis vittata</i> (Charpentier, 1825)	89
<i>Ruspolia nitidula</i> (Scopoli, 1786) *	9
<i>Tettigonia caudata</i> (Charpentier, 1845)	3
<i>Tettigonia viridissima</i> (Linnaeus, 1758)	1
Superfamily: Grylloidea	
<i>Gryllus campestris</i> Linnaeus, 1758	64
<i>Melanogryllus desertus</i> (Pallas, 1771) **	2238
<i>Modicogryllus frontalis</i> (Fieber, 1844) **	9
<i>Oecanthus pellucens</i> (Scopoli, 1763)	25
<i>Tartarogryllus burdigalensis</i> (Latreille, 1804) **	463
Order: Caelifera	
Superfamily: Acridoidea	
<i>Acrida hungarica</i> (Herbst, 1786) ^P	66
<i>Aiolopus thalassinus</i> (Fabricius, 1781)	185
<i>Calliptamus italicus</i> (Linnaeus, 1758)	1

<i>Chorthippus brunneus</i> (Thunberg, 1815)	18
<i>Chorthippus dichrous</i> (Eversmann, 1895) **	17
<i>Chorthippus dorsatus</i> (Zetterstedt, 1821)	27
<i>Chorthippus mollis</i> (Charpentier, 1825)	5
<i>Chorthippus oschei</i> (Helversen, 1986)	275
<i>Chorthippus paralellus</i> (Zetterstedt, 1821)	293
<i>Chorthippus vagans</i> (Eversmann, 1848) **	18
<i>Chrysochraon dispar</i> (Germar, 1831)	3
<i>Épacromius coerulipes</i> (Ivanov, 1887) ^{P.**}	6
<i>Euchorthippus declivus</i> (Brisout de Barneville, 1848)	865
<i>Euchorthippus pulvinatus</i> (Fischer de Waldheim, 1846) *	11
<i>Euthystira brachyptera</i> (Ocskay, 1826)	12
<i>Omocestus haemorrhoidalis</i> (Charpentier, 1825)	133
<i>Omocestus petraeus</i> (Brisout de Barneville, 1855)	8
<i>Omocestus rufipes</i> (Zetterstedt, 1821)	5
<i>Pezotettix giornae</i> (Rossi, 1794)	53
<i>Stenobothrus crassipes</i> (Charpentier, 1825)	113
<i>Stenobothrus lineatus</i> (Panzer, 1796)	1
Superfamily: Tetrigoidea	
<i>Tetrix subulata</i> (Linnaeu, 1758)	2
<i>Tetratetrix tenuicornis</i> (Shalberg, 1893)	3
Total:	5142

Heteroptera

A total number of 505 adult individuals of 54 species were collected by pitfall traps and D-vac sampling (Table 3).

Sweep netting or suction sampling are generally used to sample Heteroptera assemblages in grasslands (Standen 2000, Coscaron et al. 2009), pitfall trapping is not necessary (Standen 2000). Collecting true bugs from the ground-level generally needs great effort and has trifling result compared with collecting from the vegetation (Rédei et al. 2003). However, the sampling of epigeic true bugs may result important and valuable faunistical data. Several rare and new species for the Hungarian fauna were collected exclusively from the ground surface (e.g. Torma 2005). In the alkaline grasslands of Gyula, several rare true bug species were collected by pitfall traps, too. These species were mainly predaceous true bugs, e.g. *Prostemma sanguinea* (Rossi, 1790), *Alloeorhynchus flavipes* Fieber, 1836, *Himacerus (Stalia) boops* (Schiodte, 1870) and *Pirates hybridus* (Scopoli, 1763). The phytophagous species were mainly polyphagous bugs, but some specialist herbivorous true bugs were also collected, e.g. *Piesma kochiae* (Beckegur, 1867), *Sciocoris sulcatus* Fieber, 1851 and *Vilpianus galii* (Wolff,

1802). Several species preferred alkaline grassland habitats, e.g. *Lygaeosoma anatomicum* Seidenstücker, 1960 and *Henestaris halophilus* (Burmeister, 1835).

The number of collected species was low comparing with both Hungarian (e.g. Vászárhelyi 1985, Kondorosy 2000, 2003, Kondorosy and Harmat 1998) and Romanian (e.g. Kis 1972, 1976, Torma 2009a, 2009b) Heteroptera faunistical studies, however it presumably change when processing of sweep netting material will be finished.

The nomenclature of true bugs followed the work of Kondorosy (1999).

Table 3. List of true bug species collected from the study sites.

Family: Tingidae	
<i>Acalypta marginata</i> (Wolff, 1804)	26
<i>Kalama tricornis</i> (Schränk, 1801)	4
<i>Lasiacantha gracilis</i> (Herrich-Schäffer, 1830)	1
Family: Miridae	
<i>Acetropis carinata</i> (Herrich-Schaeffer, 1842)	5
<i>Adelphocoris lineolatus</i> (Goeze, 1778)	2
<i>Adelphocoris ticinensis</i> (Mayer-Dur, 1843)	1
<i>Amblytylus concolor</i> Jakovlev, 1877	1
<i>Charagochilus weberi</i> E. Wagner, 1953	1
<i>Hallodapus rufescens</i> (Burmeister, 1835)	1
<i>Lygus pratensis</i> (Linnaeus, 1758)	2
<i>Notostira erratica</i> (Linnaeus, 1758)	7
<i>Omphallonotus quadriguttatus</i> (Kirschbaum, 1856)	3
<i>Polymerus vulneratus</i> (Panzer, 1806)	1
Family: Nabidae	
<i>Alloeorhynchus flavipes</i> Fieber, 1836	1
<i>Himacerus (Stalia) boops</i> (Schiodte, 1870)	1
<i>Nabis (s. str.) pseudoferus</i> Remane, 1949	1
<i>Nabis (s. str.) punctatus</i> Costa, 1847	1
<i>Nabis pseudoferus / punctatus</i> ♂♀	3
<i>Prostemma aeneicolle</i> Stein, 1857	4
<i>Prostemma g. guttula</i> (Fabricius, 1787)	1
<i>Prostemma sanguinea</i> (Rossi, 1790)	28
Family: Reduviidae	
<i>Pirates hybridus</i> (Scopoli, 1763)	3
Family: Piesmatidae	
<i>Piesma kochiae</i> (Beckegur, 1867)	6
Family: Lygaeidae sensu lato	
<i>Beosus quadripunctatus</i> (Müller, 1766)	3
<i>Dimorphopterus doriae</i> (Ferrari, 1874)	177

<i>Emblethis griseus</i> (Wolff, 1802)	2
<i>Graptopeltus lynceus</i> (Fabricius, 1775)	2
<i>Henestaris halophilus</i> (Burmester, 1835)	2
<i>Lygaeosoma anatolicum</i> Seidenstücker, 1960	41
<i>Megalonotus chiragra</i> (Fabricius, 1787)	1
<i>Megalonotus sabulicola</i> (Thomson, 1870)	1
<i>Metopoplax origani</i> (Kolenati, 1845)	4
<i>Microplax interrupta</i> (Fieber, 1837)	2
<i>Ortholomus punctipennis</i> (Herrich-Schäffer, 1839)	2
<i>Peritrechus gracilicornis</i> (Puton, 1877)	3
<i>Peritrechus nubilus</i> (Fallén, 1807)	2
<i>Plinthisus</i> (<i>Plinthisomus</i>) <i>pusillus</i> (Scholtz, 1846)	2
<i>Tropisteithus holosericeus</i> (Scholtz, 1846)	4
<i>Xanthochilus quadratus</i> (Fabricius, 1798)	4
Family: Pyrrhocoridae	
<i>Pyrrhocoris apterus</i> (Linnaeus, 1758)	12
<i>Pyrrhocoris marginatus</i> (Kolenati, 1845)	6
Family: Rhopalidae	
<i>Chorosoma schillingi</i> (Schummel, 1829)	3
<i>Myrmus miriformis</i> (Fallén, 1807)	2
<i>Stictopleurus crassicornis</i> (Linnaeus, 1758)	17
Family: Cydnidae	
<i>Geotomus punctulatus</i> (Costa, 1847)	17
<i>Legnotus picipes</i> (Fallén, 1807)	30
Family: Scutellaridae	
<i>Eurygaster maura</i> (Linnaeus, 1758)	35
Family: Pentatomidae	
<i>Aelia acuminata</i> (Linnaeus, 1758)	1
<i>Dolycoris baccarum</i> (Linnaeus, 1758)	5
<i>Podops inuncta</i> (Fabricius, 1775)	4
<i>Sciocoris cursitans</i> (Fabricius, 1794)	7
<i>Sciocoris distinctus</i> Fieber, 1851	2
<i>Sciocoris sulcatus</i> Fieber, 1851	5
<i>Vilpianus galii</i> (Wolff, 1802)	3
Total:	505

Hymenoptera: Formicidae

A total of 9110 ant individuals (8925 workers, 128 queens and 57 males) were identified from the pitfall trap samples of July-September, which represent 25 species (Table 4), among which *Lasius nitidigaster* Seifert, 1996 is a new species for the Hungarian myrmecofauna (Lőrinczi, submitted manuscript).

The most abundant species was *Lasius paralienus*, followed by *Tetramorium cf. caespitum*, *Tapinoma erraticum* and *Myrmica slovacica*, the latter of which is a characteristic species of saline grasslands. These species, together with *Formica rufibarbis* and *F. cunicularia*, were also those that occurred almost in all habitat types. The number of species was the highest in the mosaics of loess steppes and saline meadows (up to 18 species), while the lowest in habitats consisting of Pannonic *Camphorosma* hollows and dense and tall *Puccinellia* swards (less than 8 species).

The presence of *Ponera testacea* in the collected material is worth noticing, since it has not been found in the Great Hungarian Plain so far, and only one syntopic occurrence of the two *Ponera* species has been recorded in Hungary (Csósz and Scifert 2003). *P. testacea* is widely distributed in Southern and Central Europe, where it associates with open and xerothermous grasslands, particularly those on sand, rocky limestone, dolomite and siliceous rock (Csósz and Scifert 2003, Czechowski and Radchenko 2010). The two specimens of *P. testacea* collected in Gyula were found in a habitat of loess steppe patches.

Table 4. List of ant species identified from the alkaline grasslands of Gyula.

	workers	queens	males
Subfamily: Ponerinae			
<i>Ponera coarctata</i> (Latreille, 1802)	-	2	-
<i>Ponera testacea</i> Emery, 1895	2	-	-
Subfamily: Myrmicinae			
<i>Anergates atratulus</i> (Schenck, 1952)	-	1	-
<i>Myrmica gallienii</i> Bondroit, 1920	97	23	1
<i>Myrmica sabuleti</i> Meinert, 1861	281	-	1
<i>Myrmica scabrinodis</i> Nylander, 1846	-	2	-
<i>Myrmica slovacica</i> Sadil, 1952	1013	37	-
<i>Myrmica specioides</i> Bondroit, 1918	138	2	-
<i>Myrmica</i> sp.	-	-	24
<i>Solenopsis fugax</i> (Latreille, 1798)	47	23	23
<i>Tetramorium cf. caespitum</i>	1470	4	-
Subfamily: Dolichoderinae			
<i>Tapinoma erraticum</i> (Latreille, 1798)	1158	-	-
<i>Tapinoma madeirense</i> Forel, 1895	146	-	-
Subfamily: Formicinae			
<i>Camponotus atricolor</i> (Nylander, 1849)	27	-	-
<i>Formica cunicularia</i> Latreille, 1798	68	-	-
<i>Formica rufibarbis</i> Fabricius, 1793	391	-	-
<i>Formica sanguinea</i> (Latreille, 1798)	15	-	-

<i>Lasius carniolicus</i> Mayr, 1861	1	-	-
<i>Lasius distinguendus</i> (Emery, 1916)	1	2	-
<i>Lasius flavus</i> (Fabricius, 1782)	-	2	-
<i>Lasius fuliginosus</i> (Latreille, 1798)	2	-	-
<i>Lasius niger</i> (Linnaeus, 1758)	709	1	-
<i>Lasius nitidigaster</i> Seifert, 1996	-	3	-
<i>Lasius paraliemus</i> Seifert, 1992	2382	25	-
<i>Lasius</i> sp.	-	-	8
<i>Plagiolepis pygmaea</i> (Latreille, 1798)	56	1	-
<i>Polyergus rufescens</i> (Latreille, 1798)	921	-	-
Total:	8925	128	57

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