

Biofloc technology application in aquaculture to support sustainable development goals

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Summary

Biofloc technology (BFT) application offers benefits in improving aquaculture production that could contribute to the achievement of sustainable development goals. This technology could result in higher productivity with less impact to the environment. Furthermore, biofloc systems may be developed and performed in integration with other food production, thus promoting productive integrated systems, aiming at producing more food and feed from the same area of land with fewer input. The biofloc technology is still in its infant stage. A lot more research is needed to optimise the system (in relation to operational parameters) e.g. in relation to nutrient recycling, MAMP production, immunological effects. In addition research findings will need to be communicated to farmers as the implementation of biofloc technology will require upgrading their skills.

Aquaculture as a food-producing sector offers ample opportunities to alleviate poverty, hunger and malnutrition, generates economic growth and ensures better use of natural resources (Food and Agriculture Organization, 2017). Aquaculture production is projected to rise from 40 million tonnes by 2008 to 82 million tonnes in 2050 (FAO, 2010). The necessity to increase aquaculture production has been triggered by the increasing demand per capita in parallel to the increase of global population. However, the development of a sustainable aquaculture industry is particularly challenged by the limited availability of natural resources as well as the impact of the

industry on the environment (Costa-Pierce *et al.*, 2012; Verdegem, 2013). With these limitations in mind, the development of sustainable aquaculture industry should focus on the conceptualization of systems that despite their high productivity and profitability, utilize fewer resources including water, space, energy and eventually capital, and at the same time has lower impact on the environment (Asche *et al.*, 2008; FAO, 2017). Along with SDG 14 targets, sustainable aquaculture development could contribute to multiple objectives including ending poverty (SDG 1), ending hunger, achieving food security and improved nutrition (SDG 2) and promoting sustained, inclusive and sustainable economic growth (SDG 8) (Food and Agriculture Organization, 2017).

One of the strategies to improve aquaculture production and sustainability should focus on enhancing feed nutrient utilization. This can be developed by two different approaches, i.e. (i) by increasing the feed quality and feeding strategy in a way that the nutrients can be efficiently delivered and finally utilized and (ii) by re-utilizing the nutrient waste through modifications in the culture system. In an aquatic system, nutrients can be removed by various natural biogeochemical processes involving mostly microorganisms with various functions in nutrient cycles. The nutrient waste in an aquaculture system is mostly generated from unconsumed feed and the digestion and metabolic processes of feed. Nutrient waste in an aquaculture system may be re-utilized directly by other organisms at lower trophic levels, which utilize feed particles as their food source, or indirectly by the conversion of the nutrients into microbial biomass that may eventually be consumed by the cultured animal itself or other animal as their food source.

Biofloc technology is mainly based on the principle of waste nutrients recycling, in particular nitrogen, into microbial biomass that can be used *in situ* by the cultured animals or be harvested and processed into feed ingredients (Avnimelech, 2009; Kuhn *et al.*, 2010). Heterotrophic microbiota is stimulated to grow by steering the C/N ratio in the water through the modification of the carbohydrate content in the feed or by the addition of an external carbon source in the water (Avnimelech, 1999), so that the bacteria can assimilate the waste ammonium for new biomass production. Hence, ammonium/ammonia can be maintained at a low and non-toxic concentration so that water replacement is no longer required.

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Biofloc technology enhances the production and productivity by its contribution to the supply of good quality fish juveniles, the latter being one of the most important inputs in the production. In addition, it contributes to the improvement of the fish production. In relation to the former, biofloc technology could support the supply of good quality seeds by improving the reproductive performance of aquaculture animals and by enhancing the larvae immunity and robustness (Ekasari *et al.*, 2015; Ekasari *et al.*, 2016; Emerenciano *et al.*, 2013). In relation to the latter, the application of biofloc technology in grow out systems of some aquaculture species could improve net productivity by 8–43%, relative to the non-biofloc control (traditional with water exchange, clear water system or recirculating aquaculture system) (Ekasari, 2014).

Biofloc systems provide a nutritious food source and can improve feed utilization efficiency

In situ utilization of microbial flocs generated in biofloc systems by some aquaculture organisms as well as the utilization of processed bioflocs as a feed ingredient has been well documented (Kuhn *et al.*, 2009, 2010; Anand *et al.*, 2014). Ju *et al.* (2008) demonstrated that the concentrations of free amino acids such as alanine, glutamate, arginine and glycine, which are known attractants in shrimp diet (Nunes *et al.*, 2006), are present in bioflocs. Levels in bioflocs were found to be comparable to that of the shrimp commercial diet suggesting that bioflocs are likely to be recognized as food particles by some aquaculture organisms. Furthermore, biofloc technology application in larviculture (at least to some species which can handle particles in suspension) may provide easily accessible food source for the larvae outside the regular feeding moments, thus minimizing possible negative social interaction during feeding (Ekasari *et al.*, 2015).

Studies have demonstrated a more efficient dietary nutrient assimilation in this system. Da Silva *et al.* (2013) reported that the application of biofloc technology on Pacific white shrimp super intensive culture considerably enhanced N and P utilization efficiency up to 70% and 66%, respectively, relative to conventional intensive culture systems with regular water exchange. Another report by Avnimelech (2007) noted that applying biofloc technology in tilapia intensive cultures increased nitrogen recovery from 23% to 43%. Essentially, biofloc studies with Pacific white shrimp (Xu and Pan, 2012), tilapia (Azim and Little, 2008) and green tiger shrimp (Megahed, 2010) clearly showed the possibility to reduce protein content in the feed. Moreover, Ray *et al.* (2010a) pointed out that the use of plant-based diet (96% protein obtained from plant-based ingredients) is favourable in a biofloc system. The reduction of protein content of the

feed and the use of plant-based protein sources in the feed are considered to be more sustainable and eco-friendly because of the reduced production of nitrogenous and phosphorous waste. It also reduces the dependency on overexploited marine resources.

Bioflocs may contribute to the supply of essential nutrients and digestive enzymes either through the stimulation of endogenous production or microbial secretion (Xu and Pan, 2012; Anand *et al.*, 2014), and the enhancement of nutrient bioavailability that facilitates higher nutrient assimilation. As a protein source, bioflocs could be considered as a good protein source for shrimp and a useful protein source for tilapia and mussel (Ekasari *et al.*, 2014a,b). Bioflocs also contain various bioactive compounds including essential fatty acids, carotenoids, free amino acids and chlorophylls (Ju *et al.*, 2008), trace minerals (Tacon *et al.*, 2002) and vitamin C (Crab *et al.*, 2012) which are known to have positive effects on aquaculture animals including the enhancement of antioxidant status, growth, reproduction and immune response.

Bioflocs also offers a lot of MAMPs (microbial associated molecular patterns), which may be recognized as immunostimulants, resulting in higher resistance to diseases (Ekasari *et al.*, 2014a,b). Interestingly, when biofloc technology was applied in tilapia broodstock culture system, it enhanced the immunological status contributing to the improvement of the larvae robustness against diseases and environmental stress test (Ekasari *et al.*, 2015; Ekasari *et al.*, 2016). In biofloc systems, aquaculture animals may also benefit from reduced pathogen pressure. Some studies demonstrated that the presence of potentially pathogenic bacteria might be reduced in biofloc systems (Crab *et al.*, 2010b; Zhao *et al.*, 2012). It has been suggested that the reduction of *V. harveyi* population in biofloc environment might be related to the disruption of *V. harveyi* cell-to-cell communication also known as an important factor in determining the pathogenicity of this particular bacterium (Crab *et al.*, 2010b).

Biofloc systems reduce water utilization and waste generation

Equally important as target species production enhancement, the application of biofloc technology may significantly reduce the quantity of water used, a main resource in aquaculture. To illustrate, an intensive zero exchange lined shrimp pond only required 1–2.26 m³ kg⁻¹ shrimp, whereas a conventional system with regular water exchange may require water up to 80 m³ kg⁻¹ (Hargreaves, 2006). In addition, Luo *et al.* (2014) noted that water consumption of biofloc-based tilapia culture system was 40% lower than that of recirculating aquaculture system (RAS).

Most of the studies applying biofloc technology confirmed that the N and P waste in this system could be reduced, corroborating the role of this system on the improvement of aquaculture productivity and the reduction of environmental impact from aquaculture unit (among others, Pérez-Fuentes *et al.*, 2013; Luo *et al.*, 2014). Although heterotrophic bacteria are the main nitrogen conversion agent, biofloc system also facilitate other nitrogen conversion mechanisms including nitrification (Ekasari, 2014), phototrophic N uptake (Emerenciano *et al.*, 2013c) and denitrification (Hu *et al.*, 2014) (all dependent on the prevailing environmental conditions). The nutrient recycling by the microbial loop involves the uptake of inorganic phosphorus by heterotrophic bacteria (Kirchman, 1994), which is not only reducing the discharged P, but also enhancing the bioavailability of this nutrient for the cultivated animals. The level of P assimilation efficiency of fishmeal and plant-based ingredients by fish has been perceived to be limited by the high level of indigestible bone-P and phytate-P; therefore, it is likely that this nutrient will be egested in the faeces rather than utilized by the cultivated animals. The consumption of the microbial biomass in the biofloc might therefore facilitate P assimilation, in particular the indigestible one, from the feed to the cultivated organisms thus reducing the nutrient waste (Luo *et al.*, 2014, Da Silva *et al.*, 2013).

Biofloc-based integrated aquaculture system for higher productivity, higher nutrient utilization and lower aquaculture pollution

A possible modification in biofloc-based aquaculture to maximize nutrient utilization efficiency is by the applying nutrient recycle principle in an integrated aquaculture system. The faster conversion of nutrient by the microbes associated in bioflocs or periphyton may provide more digestible and nutritious additional food source for both main cultured organism and other species added into the system. In this way, utilization of the wasted nutrients is expected to be more efficient and less pollution is generated. The recent study by Liu *et al.* (2014) showed that the addition of maize to stimulate bioflocs grown in an integrated culture of shrimp, spotted scat and water spinach significantly increased shrimp total yield, reduced total food conversion ratio (FCR) and lowered total P and total N in the cultured water. Interestingly, combining biofloc system with integrated multi trophic culture system may also enhance nutrient utilization efficiency. Ekasari (2014) demonstrated that combining biofloc-based shrimp culture system with tilapia, mussel and seaweed resulted in higher production, higher feed N and P recovery by the shrimp and the entire culture system, and simultaneously resulted in reduced waste nutrient and microbial biomass. Furthermore, the addition of seaweed or macrophytes

(Brito *et al.*, 2014; Liu *et al.*, 2014; Pinho *et al.*, 2017) in a biofloc-based integrated aquaculture system may also bring about the possibility to capture the excess CO₂, which may result in an increase in C utilization efficiency and a reduction in the emission of GHG. This additional benefit in nutrient utilization efficiency should stimulate further research on the possibility of incorporating biofloc system into an integrated multitrophic culture system to mitigate negative environmental impact of aquaculture nutrient wastes.

Conclusion

Biofloc technology application offers benefits in improving aquaculture production that could contribute to the achievement of sustainable development goals. This technology could result in higher productivity with less impact to the environment. Furthermore, biofloc systems may be developed and performed in integration with other food production, thus promoting productive integrated systems, aiming at producing more food and feed from the same area of land with fewer input. The biofloc technology is still in its infant stage. A lot more research is needed to optimize the system (in relation to operational parameters) e.g. in relation to nutrient recycling, MAMP production and immunological effects. In addition, research findings will need to be communicated to farmers as the implementation of biofloc technology will require upgrading their skills.

Conflicts of Interest

None declared.

References

- Anand, P.S.S., Kohli, M.P.S., Kumar, S., Sundaray, J.K., Roy, S.D., Venkateshwarlu, G., *et al.* (2014) Effect of dietary supplementation of biofloc on growth performance and digestive enzyme activities in *Penaeus monodon*. *Aquaculture* **418**: 108–115.
- Asche, F., Roll, K.H. and Tveterås, S. (2008). Future trends in aquaculture: productivity growth and increased production. In *Aquaculture in the Ecosystem*. Holmer, M., Black, K., Duarte, C.M., Marbà, N., Karakassis, I., (eds). Dordrecht, The Netherlands: Springer Science + Business Media B.V., pp. 271–292.
- Avnimelech, Y. (1999) Carbon/nitrogen ratio as a control element in aquaculture systems. *Aquaculture* **176**: 227–235.
- Avnimelech, Y. (2007) Feeding with microbial flocs by tilapia in minimal discharge bio-flocs technology ponds. *Aquaculture* **264**: 140–147.
- Avnimelech, Y. (2009) *Biofloc Technology — A Practical Guide Book*. Baton Rouge, LA: The World Aquaculture Society, p. 182.

- Azim, M.E., and Little, D.C. (2008) The bioflocs technology (BFT) in indoor tanks: water quality, bioflocs composition, and growth and welfare of Nile tilapia (*Oreochromis niloticus*). *Aquaculture* **283**: 29–35.
- Brito, L.O., Arana, L.A.V., Soares, R.B., Severi, W., Miranda, R.H., da Silva, S.M.B.C., et al. (2014) Water quality, phytoplankton composition and growth of *Litopenaeus vannamei* (Boone) in an integrated biofloc system with Gracilariabirdiae (Greville) and *Gracilaria domingensis* (Kützinger). *Aquac Int* **22**: 1649–1664.
- Costa-Pierce, B.A., Bartley, D.M., Hasan, M., Yusoff, F., Kaushik, S.J., Rana, K., et al. (2012) Responsible use of resources for sustainable aquaculture. In *Proceedings of the Global Conference on Aquaculture 2010: Farming the Waters for People and Food*. Subasinghe, R.P., Arthur, J.R., Bartley, D.M., De Silva, S.S., Halwart, M., Hishamunda, N., Mohan, C.V., Sorgeloos, P., (eds). Rome, Italy: Food and Agriculture Organization of the United Nation, pp. 113–436.
- Crab, R., Lambert, A., Defoirdt, T., Bossier, P., and Verstraete, W. (2010b) The application of bioflocs technology to protect brine shrimp (*Artemia franciscana*) from pathogenic *Vibrio harveyi*. *J Appl Microbiol* **109**: 1643–1649.
- Crab, R., Defoirdt, T., Bossier, P., and Verstraete, W. (2012) Biofloc technology in aquaculture: beneficial effects and future challenges. *Aquaculture* **356**: 351–356.
- Da Silva, K.R., Wasielesky, W., and Abreu, P.C. (2013) Nitrogen and phosphorus dynamics in the biofloc production of the pacific white shrimp, *Litopenaeus vannamei*. *J World Aquac Soc* **44**: 30–41.
- Ekasari, J. (2014) Biofloc technology as an integral approach to enhance production and ecological performance of aquaculture. Dissertation. Ghent University
- Ekasari, J., Azhar, M.H., Surawidjaja, E.H., Nuryati, S., De Schryver, P., and Bossier, P. (2014) Immune response and disease resistance of shrimp fed biofloc grown on different carbon sources. *Fish Shellfish Immunol* **41**: 332–339.
- Ekasari, J., Rivandi, D. R., Firdausi, A. P., Surawidjaja, E. H., Zairin, M., Bossier, P., et al. (2015) Biofloc technology positively affects Nile tilapia (*Oreochromis niloticus*) larvae performance. *Aquaculture* **441**: 72–77.
- Ekasari, J., Suprayudi, M.A., Wiyoto, W., Hazanah, R.F., Lenggara, G.S., Sulistiani, R., et al. (2016) Biofloc technology application in African catfish fingerling production: the effects on the reproductive performance of broodstock and the quality of eggs and larvae. *Aquaculture* **464**: 349–356.
- Ekasari, J., Zairin, M., Putri, D. U., Sari, N. P., Surawidjaja, E. H., and Bossier, P. (2015) Biofloc-based reproductive performance of Nile tilapia *Oreochromis niloticus* L. broodstock. *Aquac Res* **46**: 509–512.
- Emerenciano, M., Cuzon, G., Paredes, A., and Gaxiola, G. (2013) Evaluation of biofloc technology in pink shrimp *Farfantepenaeus duorarum* culture: growth performance, water quality, microorganisms profile and proximate analysis of biofloc. *Aquacult Int* **21**: 1381–1394.
- Food and Agriculture Organization (2010) *The State of World Fisheries and Aquaculture 2010*. Rome: Food and Agriculture Organization, 179 pp.
- Food and Agriculture Organization (2017) *FAO and the SDGs. Indicators: Measuring up to the 2030 Agenda for Sustainable Development*. Rome: FAO, 39 pp. <http://www.fao.org/3/a-i6919e.pdf>
- Hargreaves, J. A. (2006) Photosynthetic suspended-growth systems in aquaculture. *Aquacult Eng* **34**: 344–363.
- Hu, Z., Lee, J.W., Chandran, K., Kim, S., Sharma, K., and Khanal, S.K. (2014) Influence of carbohydrate addition on nitrogen transformations and greenhouse gas emissions of intensive aquaculture system. *Sci Total Environ* **470**: 193–200.
- Ju, Z.Y., Forster, I., Conquest, L., Dominy, W., Kuo, W.C., and Horgen, F.D. (2008) Determination of microbial community structures of shrimp floc cultures by biomarkers and analysis of floc amino acid profiles. *Aquac Res* **39**: 118–133.
- Kirchman, D. L. (1994) The uptake of inorganic nutrients by heterotrophic bacteria. *Microb Ecol* **28**: 255–271.
- Kuhn, D.D., Boardman, G.D., Lawrence, A.L., Marsh, L., and Flick, G.J. (2009) Microbial floc meals as a replacement ingredient for fish meal and soybean protein in shrimp feed. *Aquaculture* **296**: 51–57.
- Kuhn, D.D., Lawrence, A.L., Boardman, G.D., Patnaik, S., Marsh, L., and Flick, G.J. (2010) Evaluation of two types of bioflocs derived from biological treatment of fish effluent as feed ingredients for Pacific white shrimp, *Litopenaeus vannamei*. *Aquaculture* **303**: 28–33.
- Liu, L., Hu, Z., Dai, X., and Avnimelech, Y. (2014) Effects of addition of maize starch on the yield, water quality and formation of bioflocs in an integrated shrimp culture system. *Aquaculture* **418**: 70–86.
- Luo, G., Gao, Q., Wang, C., Liu, W., Sun, D., Li, L., et al. (2014) Growth, digestive activity, welfare, and partial cost-effectiveness of genetically improved farmed tilapia (*Oreochromis niloticus*) cultured in a recirculating aquaculture system and an indoor biofloc system. *Aquaculture* **422**: 1–7.
- Megahed, M. (2010) The effect of microbial biofloc on water quality, survival and growth of the green tiger shrimp (*Penaeus semisulcatus*) fed with different crude protein levels. *J Arab Aquacult Soc* **5**: 119–142.
- Nunes, A.J.P., Sa, M.V.C., Andriola-Neto, F.F., and Lemos, D. (2006) Behavioral response to selected feed attractants and stimulants in Pacific white shrimp, *Litopenaeus vannamei*. *Aquaculture* **260**: 244–254.
- Pérez-Fuentes, A., Pérez-Rostro, C.I., and Hernández-Vergara, M. (2013) Pond-reared Malaysian prawn *Macrobrachium rosenbergii* with the biofloc system. *Aquaculture* **400**: 105–110.
- Pinho, S.M., Molinari, D., de Mello, G.L., Fitzsimmons, K.M., and Emerenciano, M.G.C. (2017) Effluent from a biofloc technology (BFT) tilapia culture on the aquaponics production of different lettuce varieties. *Ecol Eng* **103**: 146–153.
- Ray, A.J., Lewis, B.L., Browdy, C.L., and Leffler, J.W. (2010) Suspended solids removal to improve shrimp (*Litopenaeus vannamei*) production and an evaluation of a plant-based feed in minimal-exchange, superintensive culture systems. *Aquaculture* **299**: 89–98.
- Tacon, A.G.J., Cody, J.J., Conquest, L.D., Divakaran, S., Forster, I.P., and Decamp, O.E. (2002) Effects of culture system on the nutrition and growth performance of Pacific

- white shrimp *Litopenaeus vannamei* (Boone) fed different diets. *Aquacult Nutr* **8**: 121–137.
- Verdegem, M.C.J. (2013) Nutrient discharge from aquaculture operations in function of system design and production environment. *Rev Aquacult* **4**: 1–14.
- Xu, W.J., and Pan, L.Q. (2012) Effects of bioflocs on growth performance, digestive enzyme activity and body composition of juvenile *Litopenaeus vannamei* in zero-water exchange tanks manipulating C/N ratio in feed. *Aquaculture* **356**: 147–152.
- Xu, W.J., and Pan, L.Q. (2013) Enhancement of immune response and antioxidant status of *Litopenaeus vannamei* juvenile in biofloc-based culture tanks manipulating high C/N ratio of feed input. *Aquaculture* **412**: 117–124.
- Zhao, P., Huang, J., Wang, X.H., Song, X.L., Yang, C.H., Zhang, X.G., and Wang, G.C. (2012) The application of bioflocs technology in high-intensive, zero exchange farming systems of *Marsupenaeus japonicus*. *Aquaculture* **354**: 97–106.