## 1 Thirsty tree roots exude more carbon

2

- 3 Catherine Preece\*<sup>1,2</sup>, Gerard Farré-Armengol<sup>1,2</sup>, Joan Llusià<sup>1,2</sup>, Josep Peñuelas<sup>1,2</sup>.
- 4 1 CREAF, Cerdanyola del Vallès, 08193, Spain.
- 5 2 CSIC Global Ecology Unit, CREAF-CSIC-UAB, Bellaterra, 08193, Spain.

6

7 Corresponding author: Catherine Preece, catherine.preece09@gmail.com, (+34) 935814677.

8 9

Post-print of: Preece, C. et al. "Thirsty tree roots exude more carbon" in Tree physiology (Ed. Oxford University Press), published online Jan. 2018. The final versión is available at DOI 10.1093/treephys/tpx163

#### **Abstract**

Root exudation is an important input of carbon into soils and affects plant and soil communities, but little is known about the effect of climatic factors such as drought on exudation and its ability to recover. We studied the impact of increasing drought on root exudation and its subsequent recovery in the Mediterranean tree species *Quercus ilex* in a greenhouse study by measuring the amount of total organic carbon in exudates. The amount of exudation per unit root area increased with drought duration and was 21% higher under the most extreme drought scenario compared with the non-droughted control. The amount of root exudation did not differ between the treatments following six weeks of re-watering, indicating a strong capacity for recovery in this species. We concluded that drought could affect the amount of root exudation which could in turn have a large impact on microbial activity in the rhizosphere, and alter these microbial communities, at least in the short term. This tree species may be able to return to normal levels of root exudation after a drought event, but long-term exudate-mediated impacts on Mediterranean forest soils may be an unforeseen effect of drought.

- Key words: Carbon; Drought; Mediterranean; Quercus ilex; Root Exudates;
- 27 Rhizodeposition.

#### Introduction

Drought intensity is increasing in many parts of the globe, through more frequent and longer periods of water stress (Dai 2011; Field et al. 2014; Touma et al. 2015), and this is likely to have negative impacts on plants and soils in both natural and agricultural habitats. Much remains to be understood about the impacts of water stress on plant-soil interactions and belowground processes. Rhizodeposition is the release of a wide range of compounds from roots into the soil (Bais et al. 2006), and the characterisation of rhizodeposits of tree species has become an important research focus (Prescott and Grayston 2013). Compounds released from roots as rhizodeposits can be divided into water-soluble exudates (e.g. sugars, amino acids, organic acids, and enzymes) and water-insoluble materials (e.g. mucilage, sloughed cells, and dying roots) (Merbach et al. 1999; Wichern et al. 2008).

Rhizodeposition can represent an important loss of carbon (*C*) from plants, estimated at 2-11% of the *C* fixed during photosynthesis (Jones et al. 2004; Jones et al. 2009; Pinton et al. 2007). However, the significance of rhizodeposition is not only the amount of *C* that is released, but the further impacts it has on microbial activity in the rhizosphere (Bais et al. 2006; Finzi et al. 2015; Huang et al. 2014). The majority of this *C* efflux is likely passively diffused, but roots may also increase *C* efflux under certain conditions, such as in response to toxic compounds (Badri and Vivanco 2009; Morel et al. 1986) or to increase nutrient mobilisation from the soil (Farrar et al. 2003). Root exudation is an important *C* source for soil microorganisms, and changes in the amount and composition of root exudates can have further effects on the characteristics of soil communities (Dennis et al. 2010; Haichar et al. 2008; Paterson et al. 2007), making it a key linkage between plants and soils. Overall, root exudation and other rhizosphere processes are increasingly being seen as important drivers of terrestrial *C* and nutrient cycling on an ecosystem scale (Bardgett et al. 2014; Finzi et al. 2015).

The effect of drought on root exudation is not well understood, with varied responses reported amongst studies, so effects may be difficult to predict (Preece and Peñuelas 2016). For example, C inputs (per individual and per gram of plant biomass) from rhizodeposits increased for *Lolium perenne*, *Festuca arundinacea*, and *Medicago sativa* in a 40-day drought pot experiment (Sanaullah et al. 2012) and for a mixture of perennial grasses and herbs water stressed for 56 days in a field experiment in a mountain meadow (Fuchslueger et al. 2014). In contrast, rhizodeposition decreased for *Calluna vulgaris* during drought in a 56-day field study with sites in the UK and Denmark (Gorissen et al. 2004). A recent review, summarising drought studies that measured root exudation, suggested that low-moderate drought increases exudation, but this effect is more variable under extreme water stress (Preece and Peñuelas 2016).

Periods of drought are predicted to increase in the Mediterranean region in the next decades (Field et al. 2014), but the effects of drought on root exudation in Mediterranean species have not been investigated. The holm oak, *Quercus ilex* L., is a key tree species throughout the Mediterranean Basin and has been well-studied, so it is a good candidate for investigating the impacts of drought on plant-soil interactions throughout this region. Both experimental and natural droughts have had negative impacts on survival and growth of this species, for example, a five-year experimental drought decreased stem diameter and increased stem mortality (Ogaya and Peñuelas 2007). Recurrent natural droughts have had a negative impact on survival and regeneration of the species, leading to changes in the forest canopy and loss of resilience (Lloret et al. 2004; Peñuelas et al. 2001), although there is evidence that a dampening of the drought effects occurs over longer periods of more than a decade (Barbeta et al. 2013).

We thus determined the effect of increasing drought intensities on *in situ* root exudation in a greenhouse experiment with three-year-old *Q. ilex* saplings. We hypothesised

that: (1) low-intensity (short duration) drought would increase exudation, whereas high-intensity (long-duration) drought might decrease it, based upon patterns found in the existing literature (Preece and Peñuelas 2016); (2) root exudation would be able to recover when the drought conditions ended, as this species is adapted to the drought-prone Mediterranean climate, but perhaps not following high intensity drought, which could lead to irreversible damage of the roots.

84

85

86

87

88

89

90

91

92

93

94

95

96

97

98

99

100

101

102

78

79

80

81

82

83

#### Materials and methods

Plant and soil material

A greenhouse experiment was established at the Autonomous University of Barcelona (Spain) in May 2015. The experiment comprised 180 three-year-old *Quercus ilex* L. (holm oak) saplings (provided by Forestal Catalana, Barcelona, Spain). Plants were re-potted in 3.5 1 pots, with a substrate consisting of 45% autoclaved peat, 45% sand, and 10% natural soil inoculum. The soil was collected from a natural holm oak forest on a south-facing slope (25%) slope) in the Prades Mountains in north-eastern Spain (41°13'N, 0°55'E; 930 m a.s.l.). The experiment was designed to include three soil types of control, droughted, and sterilised, with 60 plants in each soil. Therefore, the natural soil inoculum varied amongst the treatments: topsoil was collected from the control and drought plots of the long-term drought experiment for inoculating the corresponding control and drought soil treatments, respectively, in the current experiment. The sterilised soil treatment received autoclaved soil from the control plots. However, there was no difference in exudation between soil types, (data with points separated by soil is shown in Supplementary Figure S1) so data was pooled into one group for all remaining analyses. The roots of the Q. ilex saplings were carefully washed in water prior to replanting to remove all soil from the previous potting mix, so that the soil communities were representative of the new soil treatments. All plants were then given adequate water, to maintain soil moisture between 20-25%, for six weeks (until the end of June 2015) to allow them to adjust to the greenhouse environment.

## Experimental design

The drought treatment was applied by stopping water addition. Ten levels of drought were applied by varying the length of time without water - 0, 2, 4, 7, 9, 11, 14, 16, 18 and 21 days. Each drought level therefore had 18 pots, divided into six blocks. Samples of root exudates were collected (described below) from half of the pots at the end of each drought period, and the plants were harvested to obtain plant biomass and root:shoot ratio. The remaining pots were re-watered at optimal amounts (in order to achieve soil moisture of 20-25%) for six weeks, after each respective drought period, to represent a recovery phase, after which exudate and biomass measurements were collected (details given below). This amount of time for recovery was chosen as it should have been sufficient to allow the soil moisture return to normal and for roots to recover function, but still within the time-frame of the same summer period, allowing us to determine if recovery could happen in the same growing period.

Mean air temperature during the experiment (monitored using EL-USB-2 data logger, Lascar Electronics, Wiltshire, UK) was 26.7 °C. Soil temperature was monitored at a fine scale in five pots, across the different soil types (using a Decagon Em50 data logger with 5TM soil probes, Decagon Devices, Pullman, USA), and averaged 27.0 °C throughout the experiment (see Supplementary Figure S2). Soil moisture in each pot was measured at the start of the experiment and at the end of its drought period, and recovery period if relevant (using ML3 Theta Probe connected to a HH2 Moisture Meter from Delta-T Devices, Cambridge, UK). Mean soil moisture was 22.6% at the start of the experiment and decreased exponentially throughout the 21-day drought period to 0.3% at the end of the drought

treatment (see Supplementary Figure S3a). Soil moisture recovered quickly to about 20% within one week of re-watering and was successfully maintained at non-drought levels at a mean of 24.7% (see Supplementary Figure S3b).

131

132

133

134

135

136

137

138

139

140

141

142

143

144

145

146

147

148

149

150

151

152

130

128

129

#### Plant measurements

Root exudates were measured at the end of each drought period and again at the end of each six-week recovery period using an in situ measuring technique developed from that of Phillips et al. (2008). Briefly, a root was carefully excavated from the soil, cleaned, to remove any attached soil that could have affected the later measurement of carbon, placed in moist sand, and then wrapped in aluminium foil. This step allows the root to acclimate to being moved, and keeps it protected from physical damage and desiccation. The root was cleaned again after one day of acclimation and placed in a cuvette containing small glass beads (to apply physical pressure to the root to simulate soil) and a C-free nutrient solution (0.5 mm NH<sub>4</sub>NO<sub>3</sub>, 0.1 mm KH<sub>2</sub>PO<sub>4</sub>, 0.2 mm K<sub>2</sub>SO<sub>4</sub>, 0.4 mm CaCl<sub>2</sub>, 0.15 mm MgSO<sub>4</sub>) to prevent desiccation. The nutrient solution was replaced after two days with fresh solution (0.2) mm K<sub>2</sub>SO<sub>4</sub>, 0.4 mm CaCl<sub>2</sub>, 0.15 mm MgSO<sub>4</sub>), and the new solution was collected approximately 24 h later for the analysis of total organic C (non-purgeable organic C, using the Analytik Jena-Analyzer multi N/C 3100, Analytik Jena, Jena Germany). Three control cuvettes (without roots) were used for each drought level, and the mean C contents per hour of collection were subtracted from the C contents of the cuvettes containing roots. A few cuvettes with roots reported less C than the control cuvettes, implying that they were either subject to methodological problems or that there may have been re-uptake of the exuded C, and they were removed from further analysis.

The roots from the cuvettes were taken for measuring surface area (on fresh roots) using Image J software (Schneider *et al.*, 2012) and dry root biomass. The amount of C

released from the root was thus calculated per root area, as  $\mu g \ C \ cm^{-2} \ h^{-1}$ , and per root mass, as  $\mu g \ C \ g^{-1}$  dry mass  $h^{-1}$ . Total plant biomass and the root:shoot ratio did not differ by the length of the drought treatment or between drought and recovery. Mean plant biomass was  $50.1 \ g \ (\pm 1.95)$  and the mean root:shoot ratio was  $1.2 \ (\pm 0.03)$ .

#### Statistical analyses

The data for the root exudates were log-transformed to correct for positive skewness and to achieve normality of the residuals. The data were then analysed with linear mixed-effects models, to assess the effect of the duration of drought on the amount of C released. The difference between exudation during the drought and recovery treatments was also tested, as well as the interaction with drought duration. Block was included as a random factor, and analyses were performed using the nlme package in R (R Core Team, 2016).

## **Results**

Root exudation under drought

Mean root exudation per unit root area under control conditions (no drought) was 0.80  $\mu$ g C cm<sup>-2</sup> h<sup>-1</sup> (SE = 0.33), and the median value was 0.52  $\mu$ g C cm<sup>-2</sup> h<sup>-1</sup>. The raw data for the drought treatment varied greatly, with the first quartiles of 0.23  $\mu$ g C cm<sup>-2</sup> h<sup>-1</sup> and third quartile of 1.02  $\mu$ g C cm<sup>-2</sup> h<sup>-1</sup>. Exudation was positively linearly correlated with drought duration (P < 0.01) (Fig. 1). Mean exudation was 21% higher for the highest drought intensity (0.97  $\mu$ g C cm<sup>-2</sup> h<sup>-1</sup>) compared with the control, and median exudation was 38% higher for the highest drought level (0.72  $\mu$ g C cm<sup>-2</sup> h<sup>-1</sup>) compared with the control.

Exudation per unit of root biomass was not correlated with drought duration (see Supplementary Figure S4), and mean exudation for all drought levels was 56.7  $\mu$ g C g<sup>-1</sup> h<sup>-1</sup> (SE = 8.2) and the median value was 31.0  $\mu$ g C g<sup>-1</sup> h<sup>-1</sup>.

Recovery from drought

Root exudation following a six-week period of recovery was lower during the recovery than the drought per unit root area (P < 0.0001). Exudation was not correlated with drought duration, indicating that the plant roots were able to return to non-drought levels of exudation even after the most severe drought stress. Mean exuded C per unit root area (across all drought levels) was  $0.35~\mu g$  C cm<sup>-2</sup> h<sup>-1</sup> (SE = 0.05) and median was  $0.26~\mu g$  C cm<sup>-2</sup> h<sup>-1</sup> (see Supplementary Figure S4). There was an interaction between the duration of drought and type of experiment (drought versus recovery, P < 0.05) due to the drought and recovery exudation having different slopes in relation to duration of drought.

Exudation per unit root mass was also lower under recovery conditions compared with drought (P < 0.0001). It had a mean value of 25.2  $\mu$ g C g<sup>-1</sup> h<sup>-1</sup> (SE = 4.2) and median of 15.2  $\mu$ g C g<sup>-1</sup> h<sup>-1</sup>. There was no interaction between the duration of drought and type of experiment (drought or recovery experiment).

#### **Discussion**

Root exudation under drought

This result generally corroborates the few previous studies that have also measured rhizodeposition or root exudation under drought (Preece and Peñuelas 2016), which have reported generally higher C release under water stress. The amounts of C exuded in our study were similar to those in previous studies using a variety of methods (e.g. Brzostek et al., 2013; Meier et al., 2013; Baptist et al., 2015; Tückmantel et al., 2017), but were lower than those in other studies (e.g. Yin et al. 2013; Zhang et al. 2016).

Higher root exudation under drought could be an adaptation that helps the survival of roots, by creating better conditions for growth and survival. For example, increasing the

release of mucilage lubricates the roots so that they can pass through the soil more easily (Ahmed et al. 2014; Czarnes et al. 2000; Huang 2000) and have a higher chance of reaching areas with more water. Increased production of organic acids or extracellular enzymes involved in the release of soil nutrients may provide a short-term burst of available nutrients for plant uptake (Dakora and Phillips 2002; Jones 1998; Paterson 2003), which could also enable plants to increase root length and be more likely to arrive to soil that is less water stressed.

Alternatively, the increase in the amount of C detected in the root exudates with drought stress may have been due to increased cell damage and leakage of cell contents. A previous drought study of rhizodeposition also offered this suggestion (Henry et al. 2007), because water stress can induce root desiccation, mortality, and leakage of organic solutes (Huang and Gao 2000). Our experimental design did not allow us to conclusively determine the cause of the changes in root exudation, but the cause may differ between mild and extreme droughts. For example, seeking a better environment may be most likely at lower levels of water stress, and root cell damage may be most likely in extreme water stress. Our data tentatively support these different causes; plants in drought of less than 16 days maintained green leaves, but in the most extreme drought intensities (16-21 days without water) leaves were badly damaged or lost, suggesting root damage. Additionally, changes in root morphology may partly drive the exudation response to drought, if roots are desiccated, and therefore lead to a relative increase in root exudation per root area.

A previous literature review of the impacts of drought on rhizodeposition found that C release decreased at high levels of water stress (Preece and Peñuelas 2016), but overall our data did not provide supporting evidence. Although exudation seemed to drop at day 14, mean C exudation was highest in the three most intense drought levels (soil moisture <5%). Our results for *Q. ilex* highlight the importance of performing experiments at varying drought

intensities, because the response, even within a species, can vary considerably with the level of water stress.

## Recovery from drought

Exudation of C following a six-week period of recovery did not differ depending on the length of the drought treatment, indicating that the plant roots were able to return to non-drought levels of exudation even after the most severe drought stress. To the best of our knowledge, only one study about root exudation under drought has included a recovery or rewetting treatment. In that study, plant-derived extractable organic C in the soil in an Austrian mountain meadow with mostly perennial grasses and herbs was higher in a drought treatment and decreased to control levels after re-wetting (Fuchslueger et al. 2014). This result is in accordance with our findings, but the plants in the study by Fuchslueger *et al.* (2014) were mowed immediately before the re-wetting, so completely separating the effects of these two treatments is impossible.

#### *The future of root exudate studies*

This study is the first to use a relatively new and simple technique to assess the changes in the amount of root exudation over the course of increasing drought and subsequent recovery. It is also the first to measure C exudation from *Q. ilex* and thus provides novel information about the efflux of C from an important Mediterranean tree species. The method used allowed us to measure exudation *in situ* on many plants and without expensive equipment, and similar protocols have been used previously to measure the effect of water stress on exudates (Canarini et al. 2016; Karst et al. 2017), and give results that agree with findings using different methods (e.g. Fuchslueger et al. 2014; Sanaullah et al. 2012). For further explanation of the different methods available for measuring root exudation see

Kuzyakov (2006) which gives a good overview of the fate of C within soils and appropriate methods to measure this, and Oburger and Schmidt (2016) which is a more recent review of techniques, including an emphasis on imaging methods. The quantity of C in the root exudates returned to non-drought levels when re-watering commenced, but we do not know if the composition of exudates varied under water stress, and if so, how long the differences persisted. Exudate composition is therefore a very important area for future studies.

The general conclusion of this study is that the efflux of C from *Q. ilex* by root exudation increased during increasing drought, and C release was >20% higher than normal at the most extreme intensities of drought stress (soil moisture <5%). This increase in exudation may help the species to survive extreme droughts and represents an important change in the rhizosphere of this species, which could have further impacts on the soil microbial community. Moreover, we demonstrated that plants can have a large capacity for the recovery of root exudation, and this ability may be part of the strategy of drought tolerance in this key Mediterranean tree species.

## Acknowledgements

Funding was provided by the FP7 S-Clima project PIEF-GA-2013-626234, the European Research Council Synergy grant ERC-2013-726 SyG-610028 IMBALANCE-P, the Spanish Government project CGL2016-79835-P (FERTWARM) and the Catalan Government project SGR 2014-274. We would also like to thank the Unitat de Medi Ambient within the Centres Científics i Tecnològics (CCiTUB) of the University of Barcelona (UB) for assistance with the carbon analysis, and the technicians of the Facultat de Ciències at the Autonomous University of Barcelona (UAB) that assisted with the autoclaving of soil.

#### 278 References

- Ahmed MA, Kroener E, Holz M, Zarebanadkouki M, Carminati A (2014) Mucilage exudation facilitates root water uptake in dry soils. Funct Plant Biol. 41:1129-1137.
- Badri DV, Vivanco JM (2009) Regulation and function of root exudates. Plant, Cell & Environment. 32:666-681.
  - Bais HP, Weir TL, Perry LG, Gilroy S, Vivanco JM. 2006. The role of root exudates in rhizosphere interations with plants and other organisms. *In* Annual Review of Plant Biology, pp 233-266.
  - Baptist F, Aranjuelo I, Legay N, Lopez-Sangil L, Molero G, Rovira P, Nogues S (2015) Rhizodeposition of organic carbon by plants with contrasting traits for resource acquisition: responses to different fertility regimes. Plant Soil. 394:391-406.
  - Barbeta A, Ogaya R, Peñuelas J (2013) Dampening effects of long-term experimental drought on growth and mortality rates of a Holm oak forest. Glob Change Biol:doi: 10.1111/gcb.12269.
  - Bardgett RD, Mommer L, De Vries FT (2014) Going underground: root traits as drivers of ecosystem processes. Trends in Ecology & Evolution. 29:692–699.
  - Brzostek ER, Greco A, Drake JE, Finzi AC (2013) Root carbon inputs to the rhizosphere stimulate extracellular enzyme activity and increase nitrogen availability in temperate forest soils. Biogeochemistry. 115:65-76.
  - Canarini A, Merchant A, Dijkstra FA (2016) Drought effects on *Helianthus annuus* and *Glycine max* metabolites: from phloem to root exudates. Rhizosphere. 2:85-97.
  - Czarnes S, Hallett PD, Bengough AG, Young IM (2000) Root- and microbial-derived mucilages affect soil structure and water transport. European Journal of Soil Science. 51:435-443.
  - Dai A (2011) Drought under global warming: a review. Wiley Interdisciplinary Reviews: Climate Change. 2:45-65.
  - Dakora FD, Phillips DA (2002) Root exudates as mediators of mineral acquisition in low-nutrient environments. Plant Soil. 245:35-47.
  - Dennis PG, Miller AJ, Hirsch PR (2010) Are root exudates more important than other sources of rhizodeposits in structuring rhizosphere bacterial communities? FEMS Microbiology Ecology. 72:313-327.
  - Farrar J, Hawes M, Jones D, Lindow S (2003) How roots control the flux of carbon to the rhizosphere. Ecology. 84:827-837.
  - Field CB, Barros VR, Mach KJ, Mastrandrea MD, Aalst Mv, Adger WN, Arent DJ, Barnett J, Betts R, Bilir TE, Birkmann J, Carmin J, Chadee DD, Challinor AJ, Chatterjee M, Cramer W, Davidson DJ, Estrada YO, Gattuso JP, Hijioka Y, Hoegh-Guldberg O, Huang HQ, Insarov GE, Jones RN, Kovats RS, Lankao PR, Larsen JN, Losada IJ, Marengo JA, McLean RF, Mearns LO, Mechler R, Morton JF, Niang I, Oki T, Olwoch JM, Opondo M, Poloczanska ES, Pörtner HO, Redsteer MH, Reisinger A, Revi A, Schmidt DN, Shaw MR, Solecki W, Stone DA, Stone JMR, Strzepek KM, Suarez AG, Tschakert P, Valentini R, Vicuña S, Villamizar A, Vincent KE, Warren R, White LL, Wilbanks TJ, Wong PP, Yohe GW (2014) Technical Summary. In: Field CB, Barros VR, Dokken DJ, Mach KJ, Mastrandrea MD, Bilir TE, Chatterjee M, Ebi KL, Estrada YO, Genova RC, Girma B, Kissel ES, Levy AN, MacCracken S, Mastrandrea PR, White LL (eds) Climate Change 2014: Impacts, Adaptation, and Vulnerability Part A: Global and Sectoral Aspects Contribution of Working Group II to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA, pp 35-94.
  - Finzi AC, Abramoff RZ, Spiller KS, Brzostek ER, Darby BA, Kramer MA, Phillips RP (2015) Rhizosphere processes are quantitatively important components of terrestrial carbon and nutrient cycles. Glob Change Biol. 21:2082-2094.
  - Fuchslueger L, Bahn M, Fritz K, Hasibeder R, Richter A (2014) Experimental drought reduces the transfer of recently fixed plant carbon to soil microbes and alters the bacterial community composition in a mountain meadow. New Phytol. 201:916-927.
- Gorissen A, Tietema A, Joosten NN, Estiarte M, Peñuelas J, Sowerby A, Emmett BA, Beier C (2004)
  Climate change affects carbon allocation to the soil in shrublands. Ecosystems. 7:650-661.

- Haichar FeZ, Marol C, Berge O, Rangel-Castro JI, Prosser JI, Balesdent J, Heulin T, Achouak W (2008) Plant host habitat and root exudates shape soil bacterial community structure. ISME J. 2:1221-1230.
- Henry A, Doucette W, Norton J, Bugbee B (2007) Changes in crested wheatgrass root exudation caused by flood, drought, and nutrient stress. Journal of Environmental Quality. 36:904-912.
- Huang B (2000) Role of root morphological and physiological characteristics in drought resistance of plants. Plant–environment interactions Marcel Dekker Inc, New York:39-64.
- Huang B, Gao H (2000) Root physiological characteristics associated with drought resistance in tall fescue cultivars. Crop Science. 40:196-203.
  - Huang X-F, Chaparro JM, Reardon KF, Zhang R, Shen Q, Vivanco JM (2014) Rhizosphere interactions: root exudates, microbes, and microbial communities. Botany. 92:267-275.
- Jones DL (1998) Organic acids in the rhizosphere a critical review. Plant Soil. 205:25-44.

340

344

345

346

347348

349

350

351 352

353

354

355

356

357

358

359 360

361 362

363

364

365 366

367

368

369 370

374

377

- Jones DL, Hodge A, Kuzyakov Y (2004) Plant and mycorrhizal regulation of rhizodeposition. New Phytol. 163:459-480.
  - Jones DL, Nguyen C, Finlay RD (2009) Carbon flow in the rhizosphere: carbon trading at the soil—root interface. Plant Soil. 321:5-33.
  - Karst J, Gaster J, Wiley E, Landhäusser SM (2017) Stress differentially causes roots of tree seedlings to exude carbon. Tree Physiol. 37:154-164.
    - Kuzyakov Y (2006) Sources of CO2 efflux from soil and review of partitioning methods. Soil Biol Biochem. 38:425-448.
  - Lloret F, Siscart D, Dalmases C (2004) Canopy recovery after drought dieback in holm-oak Mediterranean forests of Catalonia (NE Spain). Glob Change Biol. 10:2092-2099.
    - Meier IC, Avis PG, Phillips RP (2013) Fungal communities influence root exudation rates in pine seedlings. FEMS Microbiology Ecology. 83:585-595.
    - Merbach W, Mirus E, Knof G, Remus R, Ruppel S, Russow R, Gransee A, Schulze J (1999) Release of carbon and nitrogen compounds by plant roots and their possible ecological importance. Journal of Plant Nutrition and Soil Science. 162:373-383.
  - Morel JL, Mench M, Guckert A (1986) Measurement of Pb<sup>2+</sup>, Cu<sup>2+</sup> and Cd<sup>2+</sup> binding with mucilage exudates from maize (*Zea mays* L.) roots. Biol Fertil Soils. 2:29-34.
  - Oburger E, Schmidt H (2016) New methods to unravel rhizosphere processes. Trends Plant Sci. 21:243-255.
  - Ogaya R, Peñuelas J (2007) Tree growth, mortality, and above-ground biomass accumulation in a holm oak forest under a five-year experimental field drought. Plant Ecol. 189:291-299.
  - Paterson E (2003) Importance of rhizodeposition in the coupling of plant and microbial productivity. European Journal of Soil Science. 54:741-750.
  - Paterson E, Gebbing T, Abel C, Sim A, Telfer G (2007) Rhizodeposition shapes rhizosphere microbial community structure in organic soil. New Phytol. 173:600-10.
  - Peñuelas J, Lloret F, Montoya R (2001) Severe drought effects on Mediterranean woody flora in Spain. Forest Science. 47:214-218.
  - Phillips RP, Erlitz Y, Bier R, Bernhardt ES (2008) New approach for capturing soluble root exudates in forest soils. Functional Ecology. 22:990-999.
- Pinton R, Varanini Z, Nannipieri P (2007) The rhizosphere: biochemistry and organic substances at the soil-plant interface. CRC press.

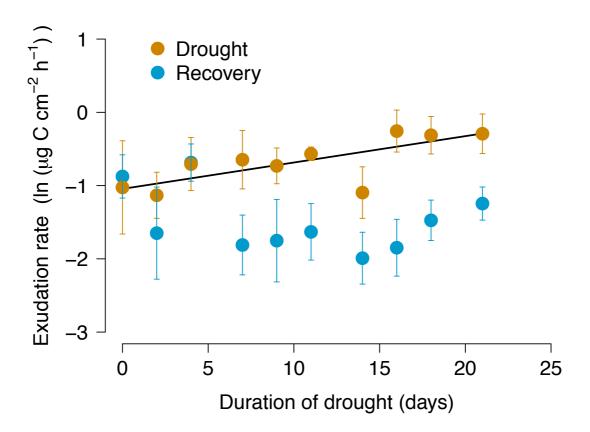
  Preece C, Peñuelas J (2016) Rhizodeposition under drought and consequences for soil communities
  - Preece C, Peñuelas J (2016) Rhizodeposition under drought and consequences for soil communities and ecosystem resilience. Plant Soil. 409:1-17.
- Prescott CE, Grayston SJ (2013) Tree species influence on microbial communities in litter and soil: Current knowledge and research needs. For Ecol Manage. 309:19-27.
  - R Core Team. 2016. R: A language and environment for statistical computing. *In* R Foundation for Statistical Computing, Vienna, Austria.
- Sanaullah M, Chabbi A, Rumpel C, Kuzyakov Y (2012) Carbon allocation in grassland communities under drought stress followed by C-14 pulse labeling. Soil Biol Biochem. 55:132-139.
- 381 Schneider CA, Rasband WS, Eliceiri KW (2012) NIH Image to ImageJ: 25 years of image analysis. 382 Nature methods. 9:671.
- Touma D, Ashfaq M, Nayak MA, Kao S-C, Diffenbaugh NS (2015) A multi-model and multi-index evaluation of drought characteristics in the 21st century. Journal of Hydrology. 526:196-207.

- Tückmantel T, Leuschner C, Preusser S, Kandeler E, Angst G, Mueller CW, Meier IC (2017) Root exudation patterns in a beech forest: Dependence on soil depth, root morphology, and environment. Soil Biology and Biochemistry. 107:188-197.
- Wichern F, Eberhardt E, Mayer J, Joergensen RG, Müller T (2008) Nitrogen rhizodeposition in agricultural crops: Methods, estimates and future prospects. Soil Biology and Biochemistry. 40:30-48.

- Yin HJ, Xiao J, Li YF, Chen Z, Cheng XY, Zhao CZ, Liu Q (2013) Warming effects on root morphological and physiological traits: The potential consequences on soil C dynamics as altered root exudation. Agric For Meteorol. 180:287-296.
- Zhang Z, Qiao M, Li D, Yin H, Liu Q (2016) Do warming-induced changes in quantity and stoichiometry of root exudation promote soil N transformations via stimulation of soil nitrifiers, denitrifiers and ammonifiers? European Journal of Soil Biology. 74:60-68.

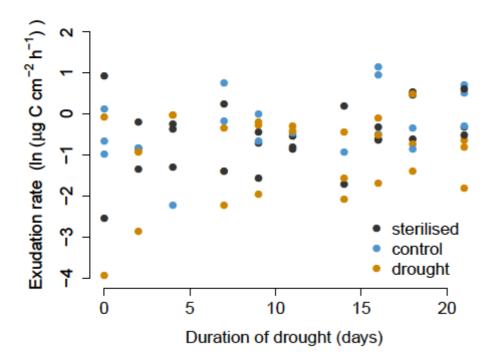
## **Figures**

Fig. 1. Exudation of carbon (natural logged) from plant roots under increasing drought duration, shown as carbon per cm<sup>2</sup> root area. Orange points are mean exudations at the end of the drought periods and blue points are mean exudations after six weeks of re-watering (recovery). Error bars are one standard error. Carbon exudation after drought increased with the duration of drought (P < 0.05, slope = 0.04). Carbon exudation after recovery was not affected by the duration of the previous drought. Exudation was higher during the drought than the recovery period (P < 0.0001), and there was a significant interaction between the duration of drought and whether plants were measured after drought or after recovery (P < 0.05).

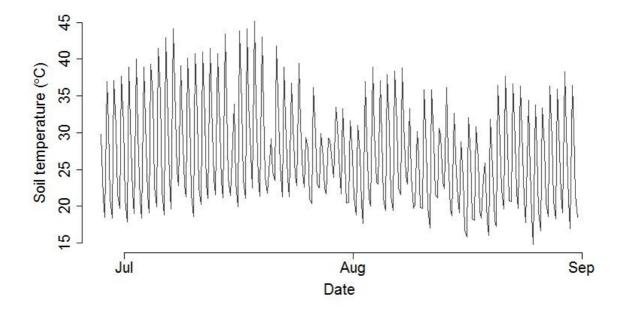


# **Electronic Supplementary Data**

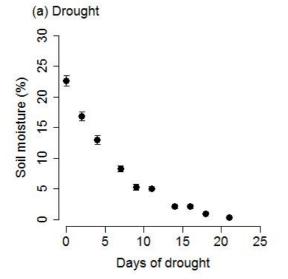
**Fig. S1.** Exudation of carbon (natural logged) from plant roots during the drought treatment, at different durations of drought, shown as carbon per cm<sup>2</sup> root area for individual plants. Data is colour-coded by soil type (sterilised, control or droughted) and there was no significant effect of soil type one exudation.

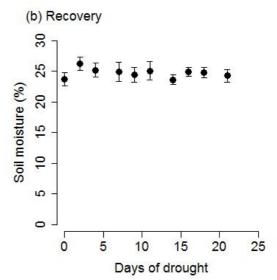


**Fig. S2.** Mean soil temperature throughout the experiment measured in five pots (due to availability of soil moisture probes).



**Fig. S3.** Percentage soil moisture for the drought levels (a) at the end of the drought treatment and (b) at the end of the recovery period. Points are means for each drought level and standard error bars are shown (n = 36 for drought measurements and n = 18 for recovery measurements).





**Fig. S4.** Exudation of carbon (natural logged) from roots under increasing drought duration, shown as carbon per gram of roots. Orange points are mean exudations at the end of the drought periods and blue points are mean exudations after six weeks of re-watering (recovery). Standard error bars are shown. There was no effect of drought duration on carbon exudation. Exudation was higher during the drought than the recovery period (P < 0.0001), but there was no significant interaction between the duration of drought and whether plants were measured after drought or after recovery).

