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*Multiple inert gases elimination technique (MIGET)  
and gas exchange in obstructive respiratory diseases.*

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## Summary

Among the four traditional causes of arterial hypoxemia (table 1) and hypercapnia (alveolar hypoventilation and  $V_A/Q$  heterogeneity)  $V_A/Q$  inequality is the most important mechanism of respiratory gas exchange impairment. In the mid 1970s a new technique has been developed able to study ventilation-perfusion ratios distributions named “multiple inert gases elimination technique”, *MIGET*.

The technique is based on the quantitative relation of the blood-gas partition coefficient ( $\lambda$ ) of a gas - the  $V_A/Q$  ratio - and the capacity of an alveolar unit to exchange that gas (Fahri, 1967). By using 6 different inert gases in trace concentrations, covering a wide spectrum of partition coefficients from 0.005 (SF<sub>6</sub>) to 300 (acetone), it is possible to characterize the distribution of the  $V_A/Q$  ratios within the whole lung. By using a lung model of 50 compartments, retentions and excretions of six gases give an estimation of a continuous distribution of the pulmonary blood flow and alveolar ventilation, respectively, against  $V_A/Q$  ratios on a logarithmic scale.

A complete  $V_A/Q$  study also analyses extra pulmonary factors determining gas exchange as cardiac output, total ventilation, oxygen consumption,  $FIO_2$ , type and concentration of Hb, pH and body temperature.

The two *major advantages* of this technique are:

- It gives an estimation of alveolar ventilation and pulmonary blood flow without disturbing either vascular or bronchomotor tone;

- It facilitates the understanding of the complex interplay of intra and extra pulmonary factors determining pulmonary gas exchange.

MIGET has been used to study the mechanisms of gas exchange impairment in main respiratory diseases, their eventual relationships with structural alterations, the effects of O<sub>2</sub> breathing and the effects of some of the drugs commonly used.

The first studies conducted between 70s and 80s on *asthmatic patients* described a variable degree of V<sub>A</sub>/Q inequalities (increased blood flow dispersion) beside a normal or mildly reduced PaO<sub>2</sub>.

Patients with stable chronic severe asthma showed modest V<sub>A</sub>/Q abnormalities maintaining a near normal PaO<sub>2</sub>, despite a severe degree of airway obstruction, likely because of less inflammatory changes at a peripheral level or because of a more active hypoxic pulmonary vasoconstriction. These data contrast with those of COPD patients showing, for a similar degree of bronchoconstriction, much greater V<sub>A</sub>/Q inequalities.

In acute severe asthma gross V<sub>A</sub>/Q abnormalities were observed, proportionally with asthma attack severity, with predominant bimodal blood flow distribution and a variable amount of cardiac output to areas with low V<sub>A</sub>/Q ratios.

No correlation has been found between respiratory and inert gases exchange and airways bronchoconstriction indices. This finding suggests that spirometric abnormalities reflect predominantly bronchoconstriction in larger and medium sized airways, while gas exchange and V<sub>A</sub>/Q abnormalities reflect more peripheral airways impairment.

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Summary.

In *COPD* patterns and severity of  $V_A/Q$  inequalities differ among patients and change with the evolution of the disease and with the clinical state of the patient.

In all cases  $V_A/Q$  inequalities appear to be the main cause of hypoxemia but no  $O_2$  diffusion limitation or true shunt have never been detected.

No correlation has been found between severity of airflow obstruction and  $V_A/Q$  inequalities as patients with mild to moderate *COPD* already show notable  $V_A/Q$  mismatch. However the degree of  $V_A/Q$  dispersion is usually higher in more severe patients.

Barberà and coworkers studied the mechanisms of gas exchange impairment during *COPD* exacerbation. They observed increases in Log SDQ, low  $V_A/Q$  ratio units, cardiac output and oxygen consumption. By using MIGET algorithm, they definitely demonstrated that gas exchange worsening during exacerbations is essentially due to  $V_A/Q$  mismatching and amplified by the decreased mixed venous  $PO_2$ , resulting from a greater  $VO_2$ .

The effects of bronchodilators in *COPD* have also been investigated since the beginning of 80s: an unexpected worsening of basal degree of  $V_A/Q$  mismatching was observed after  $\beta_2$ -agonists, likely as effect of hypoxic vasoconstriction release.

***“Adenosine 5'-monophosphate (AMP) challenge in mild asthma: cellular and gas exchange responses”***

*Abstract*

BACKGROUND: It has been suggested in asthma that bronchial challenge to indirect agents, such as 5'-adenosine monophosphate (AMP), offers a better estimate of airway inflammation than direct agents. We explored the effects of AMP challenge on lung function, including ventilation-perfusion ( $V_A/Q$ ) relationships as a marker of peripheral airway inflammation, and induced sputum, and compared with methacholine (MCh) bronchoprovocation. METHODS: A randomized, single-blinded, cross-over study using dose-response curves was designed, one week apart. Twelve non-smoking mild asthmatics (age,  $25 \pm (\text{SE})1$  yr;  $\text{FEV}_1$ ,  $92 \pm 4\%$  predicted) were studied at baseline and 5, 15 and 45 min after equivalent target responses to provoke a 30% fall in  $\text{FEV}_1$  (AMP, by  $35 \pm 2\%$ ; MCh, by  $37 \pm 2\%$ ). Sputum was collected before and 4 h after each challenge. RESULTS: Five min after challenge,  $\text{PaO}_2$  fell (AMP, by  $4.1 \pm 0.4$  kPa; MCh, by  $4.3 \pm 0.4$  kPa) due to  $V_A/Q$  mismatching as assessed by an increased overall index of  $V_A/Q$  heterogeneity (DISP R-E\*; normal values,  $<3.0$ ) (AMP, by  $3.8 \pm 0.7$ ; MCh, by  $4.6 \pm 0.7$ ), without differences between agents. Compared with MCh, there were increases in cardiac output (by  $20 \pm 8\%$ ) and oxygen consumption (by  $10 \pm 4\%$ ) ( $p < 0.05$  each) after AMP possibly related to an inotropic effect. Four h after challenge, sputum neutrophils increased after AMP (from  $48 \pm 7\%$  to  $62 \pm 6\%$ ,  $p < 0.05$ ) without associated changes in eosinophils. The number of neutrophils in the baseline sputum showed a significant correlation with the

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Summary.

concentration of IL-8 in the supernatant ( $r$ , 0.767;  $p < 0.01$ ) and with the increases in respiratory system resistances (Rrs;  $r$ , 0.803;  $p < 0.01$ ) and alveolar-arterial  $PO_2$  difference (Aa $PO_2$ ;  $r$ , 0.657;  $p < 0.05$ ) after AMP challenge. CONCLUSIONS: AMP challenge provoked an intense bronchoconstriction of similar magnitude to that of gas exchange abnormalities along with a neutrophilic response. MCh caused similar lung function changes without any sputum cellular effect.

**“Effects of nebulized salbutamol on pulmonary gas exchange during COPD exacerbations and in stable conditions”.**

*Abstract*

Short-acting  $\beta_2$ -agonists are commonly used in COPD, although their effects on pulmonary gas exchange are not fully understood.

We investigated the effects of nebulized salbutamol (5 mg) on ventilation-perfusion ( $V_A/Q$ ) inequalities in COPD patients during exacerbations (phase E) and in stable clinical condition (phase S). We studied 20 patients (1F:19M; 5 smokers, 15 ex-smokers;  $67 \pm 2$ [SEM] yrs;  $FEV_1$ ,  $37 \pm 4\%$  pred.) before and at 30 and 90 min after salbutamol. Nine patients completed both phases E and S (seven with  $V_A/Q$  measurements in both phases), 11 only phase E.

In phase E salbutamol significantly improved  $FEV_1$  and inspiratory capacity (by  $14 \pm 3\%$  and  $10 \pm 3\%$ ;  $p < 0.01$ ), increased cardiac output ( $Q_T$ , by  $13 \pm 3\%$ ,  $p < 0.01$ ) and decreased mean arterial pressure (MAP; by  $8 \pm 2\%$ ,  $p < 0.01$ ), while  $PaO_2$  showed a mild, not significant, decrease only at 30 min ( $PaO_2$  from  $61.1 \pm 2.0$  to  $59.1 \pm 2.0$ ) due to a mild increase in the dispersion of

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Summary.



pulmonary blood flow (Log SDQ; from  $1.17 \pm 0.07$  to  $1.23 \pm 0.06$ ,  $p < 0.01$ ). Patients of phase S, in comparison with phase E (paired analysis), showed a similar spirometric response to salbutamol but more marked and prolonged negative effects on gas exchange, as  $\text{PaO}_2$  passed from  $70.7 \pm 4.4$  to  $63.6 \pm 3.5$  (30 min) and to  $64.5 \pm 3.2$  mmHg (90 min) and LOG SDQ passed from  $0.96 \pm 0.09$  to  $1.12 \pm 0.13$  and to  $1.15 \pm 0.13$  (two-ways ANOVA,  $p < 0.01$ ;  $p < 0.02$ ).  $Q_T$  and MAP responses to salbutamol in phase S did not significantly differ from phase E.

The bronchodilator effect of salbutamol in COPD is associated with worsening of gas exchange and  $V_A/Q$  mismatching due to pulmonary vasodilatation. This effect is more evident in stable condition as pulmonary vasculature tone is more relaxed and liable to vasodilatation than during exacerbations (hypoxic pulmonary vasoconstriction).

*In conclusion*, the first study on AMP and MCh challenges in asthma was aimed at the investigation on the old hypothesis of dissociation between large and medium sized airways, accounting for the obstructive spirometric abnormalities of asthma, and small airways, where inflammatory and remodelling processes can affect gas exchange efficacy ( $\text{PaO}_2$ ,  $\text{AaPO}_2$ ,  $V_A/Q$  balance) independently by the bronchomotor tone. Unfortunately we could not find any differences in terms of  $V_A/Q$  imbalance between two challenges, as probably the impact of such an intense bronchoconstriction (PD30) is too high to let visible gas exchange differences specifically related to peripheral airways involvement. However the neutrofilic response we observed after

AMP is a confirmation that this model of airways challenge is more representative of natural broncho-constrictive and inflammatory processes of real asthma attacks.

By continuing on this line of investigation on small airways we intend to integrate the information deriving from MIGET with old and newer other techniques (closing volume, exhaled NO, ventilatory scan, etc.) looking for parameters providing some functional and morphological information on pathological processes of small airways. We aim to evaluate the presence of  $V_A/Q$  and gas exchange abnormalities, which remain commonly unidentified, beside mild spirometric abnormalities and to correlate them with the other parameters considered, hoping to achieve a new definition of small airways diseases in asthma.

The second work focuses on COPD by comparing the effects of inhaled salbutamol on gas exchange during exacerbations and under stable conditions.

In particular, this clinical research experience has provided a better knowledge of  $\beta_2$ -agonists vasoactive and inotropic effects, potentially influencing the general hemodynamic balance, and has underlined, therefore, the complexity of the interaction among intra pulmonary and extra pulmonary factors determining pulmonary gas exchange in COPD.

# Introduction

## Principles of physiology and assumptions.

The principal function of the lungs is to provide oxygen uptake ( $VO_2$ ) and carbon dioxide elimination ( $VCO_2$ ) adequate to satisfy the whole body metabolic request. Pulmonary gas exchange requires adequate levels of ventilation and perfusion in the alveoli. The old compartmental model of Fenn et al. and Riley and Cournard describes three ideal lung zones of ventilation-perfusion matching: 1. the *ideal lung*, where ventilation and blood flow are appropriately apportioned; 2. the *shunt fraction*, in which perfusion reaches unventilated areas; the *physiological dead space*, where the compartment is ventilated but not perfused [1-2]. Unfortunately, this historical model is not sufficient to explain  $V_A/Q$  relations inequalities that have been demonstrated to be the main cause of gas exchange abnormalities. Among the four traditional causes of arterial hypoxemia (table 1) and hypercapnia (alveolar hypoventilation and  $V_A/Q$  heterogeneity)  $V_A/Q$  inequality is the most important mechanism of respiratory gas exchange impairment [3-5].

*Table 1. Factors determining arterial hypoxemia.*

Intrapulmonary		Extrapulmonary
<u>Main factors</u>		
<ul style="list-style-type: none"> <li>• Alveolar hypoventilation</li> <li>• <math>V_A/Q</math> mismatching</li> <li>• Shunt</li> <li>• Alveolar-end capillary <math>O_2</math> diffusion limitation</li> </ul>		<ul style="list-style-type: none"> <li>• ↓Minute Ventilation</li> <li>• ↓Cardiac Output</li> <li>• ↓Inspired <math>PO_2</math></li> <li>• ↑<math>O_2</math> uptake</li> </ul>
<u>Secondary factors</u>		
		<ul style="list-style-type: none"> <li>• ↓<math>P_{50}</math></li> <li>• ↓ [Hb]</li> <li>• ↑pH</li> </ul>

$P_{50}$ =  $PO_2$  that corresponds to 50% oxyhaemoglobin saturation

Consequently, the most complete approach to investigate the pathophysiology of gas exchange impairment in respiratory diseases is the study of  $V_A/Q$  inequality.

In the mid 1970s a new technique has been developed able to study ventilation-perfusion ratios distributions named “multiple inert gases elimination technique”, *MIGET*.

This technique is based on the historical work of Fahri of the mid 1960s that demonstrated the quantitative relation of the blood-gas partition coefficient ( $\lambda$ ) of a gas - the  $V_A/Q$  ratio - and the capacity of an alveolar unit to exchange that gas [6].

The basic equation for a single lung unit gas exchange was established:

$$P_c P_A = P_v \cdot \lambda / (\lambda + V_A/Q) \quad \text{Equation 1}$$

This equation expresses mass balance during steady state elimination of the inert gas (gas not participating in metabolic processes) and indicates that both end capillary ( $P_c$ ) and alveolar ( $P_A$ ) partial pressures of an inert gas (assumed to be equal in a single lung unit) depend on the partial pressure of this gas in the venous side of the capillary ( $P_v$ ) and on the solubility of that gas, expressed as partition coefficient ( $\lambda$ ) and the  $V_A/Q$  ratio of the lung unit.

As  $P_c/P_v$  and  $P_A/P_v$  represent retention in the blood and excretion of the gas, respectively, the above equation can also be expressed as following:

$$R = E = \lambda / (\lambda + V_A/Q). \quad \text{Equation 2}$$

The retention and excretion of an inert gas depend on its partition coefficient and on  $V_A/Q$  ratio of that unit. By using 6 different inert gases in trace

concentrations, covering a wide spectrum of partition coefficients from 0.005 (SF<sub>6</sub>) to 300 (acetone), it is possible to characterize the distribution of the  $V_A/Q$  ratios within the whole lung within the widest possible range (from 0.05 to 100). The six gases generally used, in increasing order of  $\lambda$ , are: SF<sub>6</sub>, ethane, cyclopropane, enflurane or halothane, diethylether and acetone. By using a lung model of 50 compartments, the retentions of six gases give an estimation of a continuous distribution of the pulmonary blood flow against  $V_A/Q$  ratios on a logarithmic scale (fig 1). Analogously, the excretions of six inert gases provide an estimation of the distribution of the alveolar ventilation against  $V_A/Q$  ratios.

This technique, applicable both in health and disease, despite the classical division of lung in three functional department [1-2] permits to separate areas with low but finite  $V_A/Q$  ratios ( $V_A/Q$  ratio < 0.1) from areas whose  $V_A/Q$  ratios is zero (shunt), and regions with high  $V_A/Q$  ratios ( $V_A/Q$  ratio > 10) from regions that are unperfused (dead space). After reaching a steady state, the concentrations of each gas are measured in the mixed arterial blood and mixed expiratory gas. The curve relating arterial concentration and solubility is transformed into a virtually continuous distribution of blood flow against  $V_A/Q$ , using techniques of numerical analysis. The relation of between expired concentration and solubility is similarly converted into the distribution of ventilation.

Some important *assumptions* are at the base of MIGET:

- a. The existence of steady state conditions in all  $V_A/Q$  units that means the constancy of excretion and retention during the whole period of measurements.
- b. Diffusion equilibration between alveolar gas and end capillary blood for each gas.
- c. A lung model consisting in a number of homogenous compartments in parallel, each with constant and continuous ventilation and perfusion.
- d. All diffusive processes affecting pulmonary gas movements are sufficiently rapid not to affect gas exchange.

MIGET is able to provide then, a quantitative picture of lung units with particular  $V_A/Q$  ratios in a good graphical representation and computes the amount of blood flow and ventilation associated with these lung units (Figure 1). This image is useful in giving an overview of the distributions and suggesting patterns of distribution as unimodal (normal or broad), bimodal or trimodal.

The quantification of  $V_A/Q$  inequalities is better expressed through some numerical indices:

- *Mean V, Mean Q*, mean values of  $V_A/Q$  ratios of ventilation and perfusion curves.
- *Log SDV, Log SDQ*, the standard deviation of mean values in a logarithmic scale.

- Low  $V_A/Q$ , the percentage of blood flow in units whose  $V_A/Q$  ratio is lower than 0.01.
- *High*  $V_A/Q$ , the percentage of blood flow in units whose  $V_A/Q$  ratio is greater than 10.
- *Shunt*, blood flow percentage to lung units with  $V_A/Q$  ratio  $<0.05$ . No post-pulmonary shunt is detectable by MIGET.
- *Dead space*, ventilation in units of  $V_A/Q$  ratio  $>100$ .
- *DISP R-E\**, the root mean square value of retention minus excretion, an overall index of  $V_A/Q$  heterogeneity.

As the MIGET algorithm does not consider diffusion limitations, the expected value of  $PaO_2$  through this mathematical model does not always correspond to the real  $PaO_2$  measured in the blood; the positive *difference between expected and measured  $PaO_2$*  can likely be ascribed to an oxygen diffusion limitation.

Since the first works made in the 70s on the mathematical analysis of  $O_2$  and  $CO_2$  behaviour in blood, it was clear that variables derived from physiological gases ( $PaO_2$ ,  $PaCO_2$ , alveolar-arterial  $PO_2$  difference [ $AaPO_2$ ], venous admixture and physiological dead space) vary with  $V_A/Q$  matching, but also with changes in minute ventilation, cardiac output, inspired  $PO_2$ , etc. (table 1) [7].

A complete  $V_A/Q$  study also analyses extra pulmonary factors determining gas exchange as cardiac output, that can directly be measured (if inert gases levels are known in mixed venous blood, arterial blood and mixed expired gas) or using indocyanine green, total ventilation ( $V_E$ ), oxygen

consumption ( $\text{VO}_2$ ),  $\text{FIO}_2$ , type and concentration of Hb, pH and body temperature. MIGET permits to estimate the influence of each of extra pulmonary factors by introducing the observed values (singly or in any combination) of each factor in the algorithm that calculates  $V_A/Q$  inequality and the expected  $\text{PaO}_2$ .

This estimation is useful to understand the qualitative and quantitative role of each intra pulmonary and extra pulmonary factors in determining gas exchange.

The two *major advantages* of this technique are:

- It gives an estimation of alveolar ventilation and pulmonary blood flow without disturbing either vascular or bronchomotor tone;
- It facilitates the understanding of the complex interplay of intra and extra pulmonary factors determining pulmonary gas exchange.

### **Practical aspects**

Originally the technique was performed using inert gases levels determined from three different sites: mixed venous blood, arterial blood and mixed expiratory gas. The modality permits a direct measurement of all variables of equation 2 and a direct calculation of cardiac output. It implies the placement of a catheter in the pulmonary artery.

The version of MIGET most commonly used only requires mixed expiratory and peripheral venous sampling. In this case cardiac output is measured by indocyanine green (dye dilution) (Figure 2).



To inscribe the dye curve, 5 mg of dye in 1 ml of water are rapidly flushed into the venous catheter and arterial blood is withdrawn at a constant rate of 20 ml·min<sup>-1</sup>. Adequate curves are obtained prior to recirculation for the conventional Stewart-Hamilton analysis with a cardiac output computer (DC-410 Waters Instruments Inc. Rochester, MN).

The following equation is at the base of cardiac output measurement:

$$Q_T \cdot \lambda \cdot P_v = Q_T \cdot \lambda \cdot P_a + V_E \cdot P_E \quad \text{Equation 3}$$

Where  $P_E$  is mixed expiratory gas and  $P_v$  can be computed from mass balance:

$$P_v = P_a + (V_E \cdot P_E / \lambda \cdot Q_T). \quad \text{Equation 4}$$

Finally, a third modality of MIGET requiring only mixed expiratory and peripheral venous sampling is also available. This is obtained from a distally oriented canula inserted into a peripheral vein (usually in the forearm opposite to the side of the infusion of the inert gas solution). As inert gases are not metabolized by tissues after 90 min of infusion in a resting position virtual equilibration between blood and tissues is achieved and the peripheral venous blood reflects the inert gas concentration of the inflowing arterial blood.

## Instrumentation

Some fundamental instruments are necessary in MIGET:

- *Exhaled air mixing box* (heated), for the mixed expiratory gas sampling: an adapted circuit takes the expired air to a mixing box (10 L) that is heated at a temperature of about 40-45 °C , in order to avoid inert gases deposition on tube internal walls. Mixed expiratory samples are collected from the mixing box to measure inert and respiratory gases.
- *Respirometer*, Ventilatory recordings (minute ventilation, respiratory rate, tidal volume) are taken during the study. A Wright respirometer is commonly used.
- *Inert and respiratory gases analyser*. Inert gases detection is commonly made by a chromatograph accurately set-up to describe all gases peaks. Chromatograph is provided with a flame ionisation detector (FID) able to measure all gases excluding SF<sub>6</sub> that are hydrocarbons and with an electron capture detector (ECD) to measure SF<sub>6</sub> [8-9].
- *A cardiac output computer*, if a pulmonary catheterisation is not performed.

## **MIGET in healthy subjects**

The typical  $V_A/Q$  distribution of normal subjects aged less than 30 years at rest, breathing room air, is characterised by narrow perfusion and ventilation curves centred around 1  $V_A/Q$  ratio in the abscissa (figure 1).

Mean values of the second moment of the distribution (Log SDQ and Log SDV) range from 0.35 to 0.43 [10]. The upper 95% confidence limit for Log SDQ is 0.60 and for Log SDV is 0.65 [11-14].

No perfusion in lung units with  $V_A/Q$  ratios  $<0.005$  (shunt) is present [15]. The amount of ventilation to lung units with  $V_A/Q$  ratios  $>100$  (dead space) is approximately 30%, including anatomical, physiological and instrumental dead space.

Neither perfusion to lung units with  $V_A/Q$  ratio  $<0.10$  (low  $V_A/Q$ ) is observed, nor ventilation to lung units with  $V_A/Q$  ratio  $>10$  (high  $V_A/Q$ ).

Cardus et al. investigated changes of  $V_A/Q$  inequality in healthy subjects with aging [16]. By studying 64 individuals aged between 18 to 71 years they observed only a slight increase in Log SDQ and Log SDV in parallel with  $PaO_2$  mild decrease (by 6 mmHg). The upper 95% confidence limits in subjects aged 70 yrs were 0.70 for Log SDQ and 0.75 for Log SDV. They demonstrated that the observed decrease in arterial oxygenation (increased alveolar-arterial  $O_2$  gradient,  $AaPO_2$ ) with age is due to  $V_A/Q$  inequality.

## **MIGET in respiratory diseases**

In the last three decades enormous progress has been made in the study of pathophysiology of acute and chronic respiratory diseases with the contribution of MIGET.

After a wide investigation on MIGET in normal subjects under different conditions of exercise, altitude, O<sub>2</sub> inspiratory fractions (FIO<sub>2</sub>) and age [10-16], MIGET has been used to study the mechanisms of gas exchange impairment in main respiratory diseases, their eventual relationships with structural alterations, the effects of O<sub>2</sub> breathing and the effects of some of the drugs commonly used [17-18].

The milestone in the literature on the contribution of MIGET to pulmonary medicine is the series published on Thorax between 1994 and 1995 composed of six articles [19-24].

## **Bronchial asthma**

The first description of gas exchange impairment (mild to moderate hypoxemia) in asthmatic patients belongs to two Australian chest physicians in the 1967 [25]. McFadden et al confirmed the presence of hypoxemia and, in more severe cases, of hypercapnia in 101 patients suffering an acute exacerbation of asthma [26]. They postulated that obstructive changes in peripheral airways could explain the refractoriness to standard bronchodilators and relapses. Since MIGET was introduced in respiratory medicine in the 70s, a big amount of information has been collected on the mechanisms of gas exchange impairment in the different clinical forms of asthma.

The first study of Wagner on asymptomatic asthmatics in the 1978 showed that beside normal values of  $\text{PaO}_2$  (all but one patient had normal or slightly reduced  $\text{PaO}_2$ ) a bimodal blood flow distribution (increased Log SDQ) was described, with areas with low  $V_A/Q$  ratio [27]. Subsequent studies showed lower prevalence of  $V_A/Q$  inequalities, probably as a consequence of differences in patient selection, clinical management and treatment [28-31].

In patients with stable chronic severe asthma the  $V_A/Q$  abnormalities were quite modest (broad unimodal distribution, mildly increased Log SDQ and Log SDV) [32-33]. Likely, these patients maintain a near normal  $\text{PaO}_2$  despite a severe degree of airway obstruction because of less inflammatory changes at a peripheral level or because of a more active hypoxic pulmonary vasoconstriction. This findings contrast with those of COPD patients that,

with a similar degree of bronchoconstriction, show much greater  $V_A/Q$  inequalities and gas exchange impairment.

In acute severe asthma gross  $V_A/Q$  abnormalities were observed, proportionally with asthma attack severity, with predominant bimodal blood flow distribution and a variable amount of cardiac output to areas with low  $V_A/Q$  ratios [34-36]. Neither dead space nor high  $V_A/Q$  ratios were described. The increased concentration of inspired  $O_2$  and the administration of bronchodilators appeared to enhance the perfusion of low  $V_A/Q$  ratios.

All these studies have led to some general conclusions:

- Usually  $PaO_2$  is relatively preserved; this data could be explained with the contribution of increased ventilation and cardiac output (extra pulmonary factors);
- No correlation has been found between respiratory and inert gases exchange and airways bronchoconstriction indices [37-38]. This suggests that spirometric abnormalities reflect predominantly bronchoconstriction in larger and medium sized airways, while gas exchange and  $V_A/Q$  abnormalities reflect more peripheral airways impairment.

## **COPD**

Chronic obstructive pulmonary diseases are characterised by a wide spectrum of gas exchange abnormalities, ranging from mild hypoxemia to severe respiratory failure requiring ventilatory support. In all cases  $V_A/Q$  inequalities appear to be the main cause of hypoxemia [39]. By converse, no  $O_2$  diffusion limitation or true shunt have been demonstrated in COPD patients.

The patterns and the severity of  $V_A/Q$  inequalities differ among COPD patients and change with the evolution of the disease and with the clinical state of the patient.

An historical work of Wagner and coworkers described two different patterns of  $V_A/Q$  mismatch in a group of severe COPD patients [39]. Some patients showed an increased number of high  $V_A/Q$  ratio units (*high pattern*); other patients showed numerous areas with low  $V_A/Q$  ratios (*low pattern*) (Figure 3). While the *high pattern* was more frequent among emphysematous patients, no other consistent association between  $V_A/Q$  pattern and clinical pictures was found.

Other studies followed this one but no correlation was found between severity of airflow obstruction and  $V_A/Q$  inequalities as patients with mild to moderate COPD already showed notable  $V_A/Q$  mismatch [40-41]. However the degree of  $V_A/Q$  dispersion of these patients was lower than in more severe patients.

During exacerbations it has been observed worsening of  $V_A/Q$  distributions that are probably related to reversible functional abnormalities (mucus

impaction, bronchospasm, bronchial wall oedema) [42-43]. Barberà and coworkers studied the mechanisms of gas exchange impairment during COPD exacerbation [43]. Thirteen patients were studied during hospitalization and approximately 5 weeks after discharge. During exacerbations they observed increases in Log SDQ, due to a greater perfusion of low  $V_A/Q$  ratio units, in cardiac output and oxygen consumption ( $VO_2$ ). By analysing the specific effect of each intra and extra pulmonary factor determining arterial oxygenation through MIGET logarithm it was demonstrated that gas exchange worsening during exacerbations is essentially due to  $V_A/Q$  mismatching and amplified by the decreased mixed venous  $PO_2$ , resulting from a greater  $VO_2$ .

The effects of bronchodilators in COPD have been investigated since the beginning of 80s, as they have known spirometric and clinical effects but not so clear effects on gas exchange. The bronchodilation induced by these drugs was unexpectedly accompanied by a worsening of basal degree of  $V_A/Q$  mismatching, likely as effect of a possible interaction with the mechanism of hypoxic vasoconstriction [44-46]. This hypothesis is sustained by the work of Melot and coworkers that described a fall in pulmonary vascular resistance and  $PaO_2$  after the administration of nifedipine, a calcium channel blocker, able to suppress the beneficial effect of hypoxic vasoconstriction [47].

Other bronchodilators, like ipratropium bromure or aminophylline, acting through different mechanisms of action, did not change  $V_A/Q$  balance [45,48].



## References

1. Riley RL, Cournand A. "Ideal" alveolar air and the analysis of ventilation-perfusion relationships in the lung. *J Appl Physiol* 1949;1:825-47.
2. Fenn WO, Rahn H, Otis AB. A therapeutical analysis of the composition of alveolar air at altitude. *Am J Physiol* 1946;146:637-53.
3. Krogh A, Lindhard J. The volume of the dead space in breathing and the mixing of gases in the lung of man. *J Physiol (London)* 1917;51:59-90.
4. Haldane JS, *Respiration*. New Haven: Yale University Press, 1932.
5. West JB. Causes of carbon dioxide retention in lung disease. *N Engl J Med* 1971;264:1232-6.
6. Farhi LE. Elimination of inert gas by the lung. *Respir Physiol*. 1967 Aug;3(1):1-11.
7. Olszowka AJ, Farhi LE. A system of digital computer subroutines for blood gas calculations. *Respir Physiol*. 1968 Mar;4(2):270-80.].
8. Wagner PD, Saltzman HA, West JB. Measurements of continuous distributions of ventilation-perfusion ratios: theory. *J Appl Physiol* 1974;36:588-599.
9. Wagner PD, Lopez FA. Gas chromatography techniques in respiratory physiology. In: Otis AB, ed. *Techniques in life science*. Ireland:Elsevier, 1984:403/1-403/24.
10. Wagner PD, Evans JW. Conditions for equivalence of gas exchange in series and parallel models of the lung. *Respir Physiol* 1977;31:117-38.
11. Gale GE, TorreBueno J, Moon RE, Saltzman HA, Wagner PD. Ventilation perfusion inequality in normal humans during exercise. *J Appl Physiol* 1985;58: 976-88.
12. Wagner PD, Gale GE, Moon RE, TorreBueno J, Saltzman HA. Pulmonary gas exchange in humans exercising at sea level and simulated altitude. *J Appl Physiol* 1986;61:260-70.

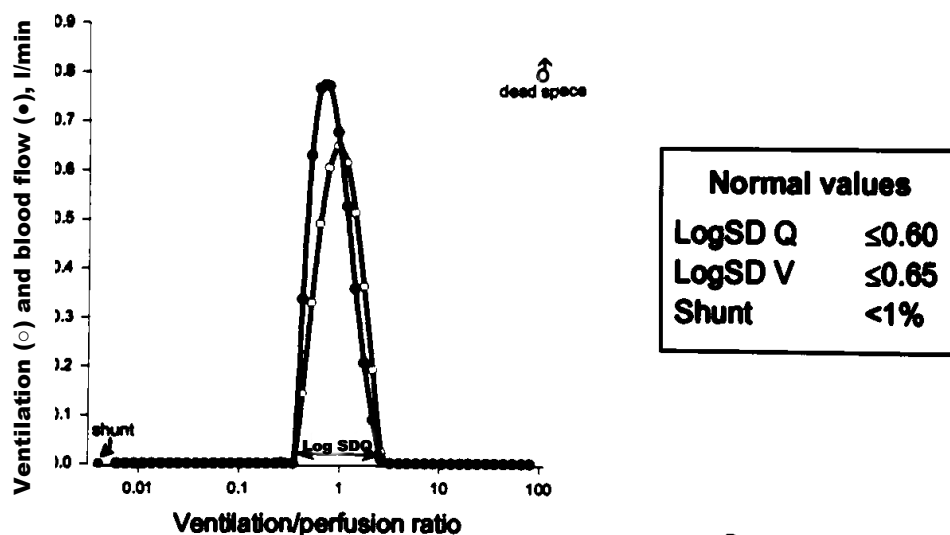
13. Hammond MD, Gale GE, Kapiran KS, Ries A, Wagner PD. Pulmonary gas exchange in humans during exercise at sea level. *J Appl Physiol* 1986;60:1590-8.
14. Hammond MD, Gale GE, Kapiran KS, Ries A, Wagner PD. Pulmonary gas exchange in humans during normobaric hypoxic exercise. *J Appl Physiol* 1985; 58:978-88.
15. Wagner PD, Laravuso RB, Uhl RR, West JB. Continuous distribution of ventilation perfusion ratios in normal subjects breathing air and 100% O<sub>2</sub>. *J Clin Invest* 1974; 54:54-68.
16. Cardus J, Burgos F, Diaz O, Roca J, Barbera JA, Marrades RM, Rodriguez-Roisin R, Wagner PD. Increase in pulmonary ventilation-perfusion inequality with age in healthy individuals. *Am J Respir Crit Care Med*. 1997 Aug;156(2 Pt 1):648-53.
17. R Rodriguez-Roisin and PD Wagner. Clinical relevance of ventilation-perfusion inequality determined by inert gas elimination. *Eur Respir J* 1990 3: 469-482.
18. Wagner PD, Rodriguez-Roisin R. Clinical Advances in Pulmonary Gas Exchange. State of the art/Conference Report. *Am Rev Respir Dis* 1991; 143:883-888.
19. Roca J, Wagner PD. Contribution of multiple inert gas elimination technique to pulmonary medicine. 1. Principles and information content of the multiple inert gas elimination technique. *Thorax*. 1994 Aug;49(8):815-24. Review.
20. Agusti AG, Barbera JA. Contribution of multiple inert gas elimination technique to pulmonary medicine. 2. Chronic pulmonary diseases: chronic obstructive pulmonary disease and idiopathic pulmonary fibrosis. *Thorax*. 1994 Sep;49(9):924-32.
21. Rodriguez-Roisin Roca J. Contribution of multiple inert gas elimination technique to pulmonary medicine. 3. Bronchial asthma. *Thorax* 1994 Oct 49(10):1027-33
22. Manier G, Castaing Y. Contribution of multiple inert gas elimination technique to pulmonary medicine 4. Gas exchange abnormalities in pulmonary vascular and cardiac disease. *Thorax*. 1994 Nov;49(11):1169-74.

23. Melot C. Contribution of multiple inert gas elimination technique to pulmonary medicine. 5. Ventilation-perfusion relationships in acute respiratory failure. *Thorax*. 1994 Dec;49(12):1251-8.
24. Hedenstierna G. Contribution of multiple inert gas elimination technique to pulmonary medicine. 6. Ventilation-perfusion relationships during anaesthesia. *Thorax*. 1995 Jan;50(1):85-91.;
25. Tai E, Read J. Blood-gas tensions in bronchial asthma. *Lancet*. 1967 Mar 25;1(7491):644-6.
26. McFadden ER Jr, Lyons HA. Arterial-blood gas tension in asthma. *N Engl J Med*. 1968 May 9;278(19):1027-32.
27. Wagner PD, Dantzker DR, Iacovoni VE, Tomlin WC, West JB. Ventilation-perfusion inequality in asymptomatic asthma. *Am Rev Respir Dis*. 1978 Sep;118(3):511-24.
28. Wagner PD, Hedenstierna G, Bylin G. Ventilation-perfusion inequality in chronic asthma. *Am Rev Respir Dis*. 1987 Sep;136(3):605-12.
29. Young IH, Corte P, Schoeffel RE. Pattern and time course of ventilation-perfusion inequality in exercise-induced asthma. *Am Rev Respir Dis*. 1982 Mar;125(3):304-11.
30. Rodriguez-Roisin R, Ferrer A, Navajas D, Agusti AG, Wagner PD, Roca J. Ventilation-perfusion mismatch after methacholine challenge in patients with mild bronchial asthma. *Am Rev Respir Dis*. 1991 Jul;144(1):88-94.
31. Lagerstrand L, Larsson K, Ihre E, Zetterstrom O, Hedenstierna G. Pulmonary gas exchange response following allergen challenge in patients with allergic asthma. *Eur Respir J*. 1992 Nov;5(10):1176-83.
32. Corte P, Young IH. Ventilation-perfusion relationships in symptomatic asthma. Response to oxygen and clemastine. *Chest*. 1985 Aug;88(2):167-75.

33. Ballester E, Roca J, Ramis L, Wagner PD, Rodriguez-Roisin R. Pulmonary gas exchange in severe chronic asthma. Response to 100% oxygen and salbutamol. *Am Rev Respir Dis.* 1990 Mar;141(3):558-62.
34. Roca J, Ramis L, Rodriguez-Roisin R, Ballester E, Montserrat JM, Wagner PD. Serial relationships between ventilation-perfusion inequality and spirometry in acute severe asthma requiring hospitalization. *Am Rev Respir Dis.* 1988 May;137(5):1055-61.
35. Ballester E, Reyes A, Roca J, Guitart R, Wagner PD, Rodriguez-Roisin R. Ventilation-perfusion mismatching in acute severe asthma: effects of salbutamol and 100% oxygen. *Thorax.* 1989 Apr;44(4):258-67.
36. Ferrer A, Roca J, Wagner PD, Lopez FA, Rodriguez-Roisin R. Airway obstruction and ventilation-perfusion relationships in acute severe asthma. *Am Rev Respir Dis.* 1993 Mar;147(3):579-84.
37. Wagner PD, Hedenstierna G, Rodriguez-Roisin R. Gas exchange, expiratory flow obstruction and the clinical spectrum of asthma. *Eur Respir J.* 1996 Jun;9(6):1278-82
38. Rodriguez-Roisin R. Acute severe asthma: pathophysiology and pathobiology of gas exchange abnormalities. *Eur Respir J.* 1997 Jun;10(6):1359-71.
39. Wagner PD, Dantzker DR, Dueck R, Clausen JL, West JB. Ventilation-perfusion inequality in chronic obstructive pulmonary disease. *J Clin Invest.* 1977 Feb;59(2):203-16.
40. Agustí AG, Barbera JA, Roca J, Wagner PD, Guitart R, Rodriguez-Roisin R. Hypoxic pulmonary vasoconstriction and gas exchange during exercise in chronic obstructive pulmonary disease. *Chest.* 1990 Feb;97(2):268-75.
41. Barbera JA, Ramirez J, Roca J, Wagner PD, Sanchez-Lloret J, Rodriguez-Roisin R. Lung structure and gas exchange in mild chronic obstructive pulmonary disease. *Am Rev Respir Dis.* 1990 Apr;141(4 Pt 1):895-901.

42. Ferrer A, Roca J, Barbera JA, Agusti AGN, Wagner PD, Rodriguez-Roisin R. Pattern and time course of ventilation-perfusion mismatch during exacerbation of chronic obstructive pulmonary disease. *Am Rev Respir Dis* 1990;141:A929
43. Barbera JA, Roca J, Ferrer A, Felez MA, Diaz O, Roger N, Rodriguez-Roisin R. Mechanisms of worsening gas exchange during acute exacerbations of chronic obstructive pulmonary disease. *Eur Respir J*. 1997 Jun;10(6):1285-91
44. Ringsted CV, Eliassen K, Andersen JB, Heslet L, Qvist J. Ventilation-perfusion distributions and central hemodynamics in chronic obstructive pulmonary disease. Effects of terbutaline administration. *Chest*. 1989 Nov;96(5):976-83.
45. Ferrer A, Viegas C, Montserrat JM, Roca J, Wagner PD, Rodriguez-Roisin R. Gas exchange responses to bronchodilators in chronic obstructive pulmonary disease. *Am Rev Respir Dis* 1991;143:A447.
46. Viegas CA, Ferrer A, Montserrat JM, Barbera JA, Roca J, Rodriguez-Roisin R. Ventilation-perfusion response after fenoterol in hypoxemic patients with stable COPD. *Chest*. 1996 Jul;110(1):71-7.
47. Melot C, Hallemans R, Naeije R, Mols P, Lejeune P. Deleterious effect of nifedipine on pulmonary gas exchange in chronic obstructive pulmonary disease. *Am Rev Respir Dis*. 1984 Oct;130(4):612-6.
48. Barbera JA, Reyes A, Roca J, Montserrat JM, Wagner PD, Rodriguez-Roisin R. Effect of intravenously administered aminophylline on ventilation/perfusion inequality during recovery from exacerbations of chronic obstructive pulmonary disease. *Am Rev Respir Dis*. 1992 Jun;145(6):1328-33.

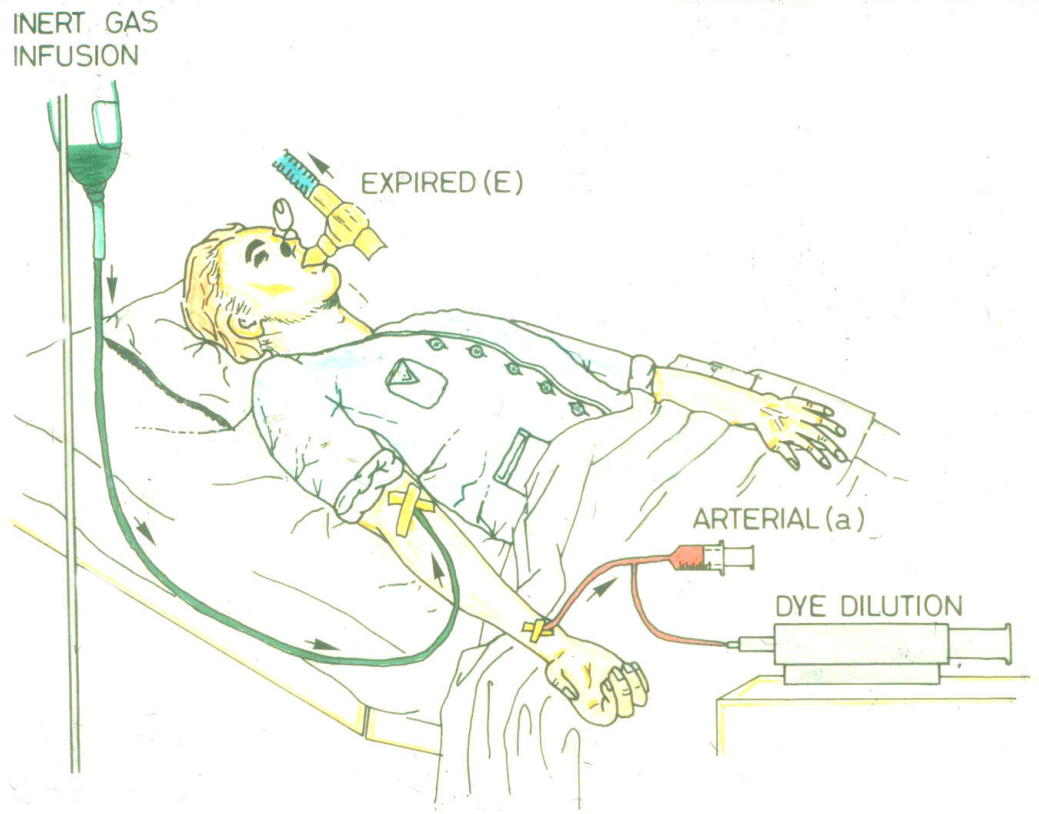
## Ventilation-perfusion distribution in healthy subjects



Data from J Cardús et al.  
AJRCCM 1997;156:648

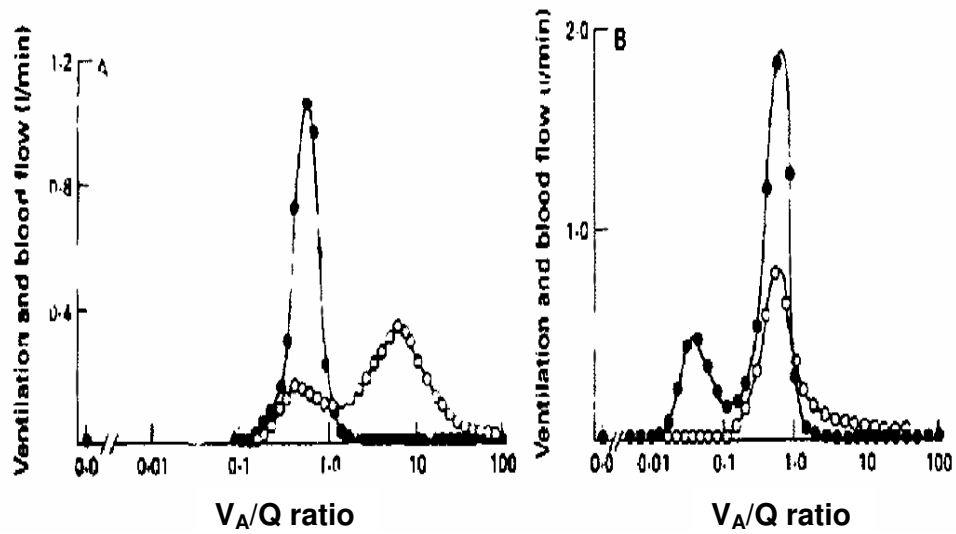
**Figure 1.**

Ventilation-perfusion distributions. Ventilation (○) and perfusion (●) are plotted against  $V_A/Q$  ratio on a logarithmic scale in a resting young healthy subject breathing room air. Both curves are centred (first moment) around a  $V_A/Q$  ratio of 1 and they are narrow (second moment). No perfusion to low  $V_A/Q$  units ( $V_A/Q$  ratio < 0.1) nor ventilation to high  $V_A/Q$  units ( $V_A/Q$  ratio > 10) are observed. Note also the absence of shunt. Each individual data point represents a particular amount of blood flow (●) or alveolar ventilation (○) to the corresponding pulmonary compartment ( $V_A/Q$  ratio). Total cardiac output corresponds to the sum of the 50 blood flow points and total alveolar ventilation is the sum of the 50 ventilation points.



*Figure 2.*

With this MIGET modality mixed expiratory gas and arterial blood samples are collected; the inert gases solution is infused into a peripheral superficial vein. Through the arterial catheter the dye dilution is detected after indocyanine green bolus (5 mg of dye in 1 ml of water ) is rapidly flushed into the venous catheter and arterial blood is withdrawn at a constant rate of 20 ml·min<sup>-1</sup>.



**Figure 3.**

A) Ventilation-perfusion distribution in patient with emphysema-type COPD. Note the bimodal pattern of ventilation distribution ( $\circ$ ) with areas with high  $V_A/Q$  ratio. B) Ventilation-perfusion ratio distribution in a patient with bronchitis-type COPD. The blood flow distribution ( $\bullet$ ) is bimodally shaped due to the presence of alveolar units with low  $V/Q$  ratio.



# ADENOSINE 5'-MONOPHOSPHATE (AMP) CHALLENGE IN MILD ASTHMA: CELLULAR AND GAS EXCHANGE RESPONSES

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Università di Pisa.

**Running head:** *AMP challenge and asthma*

**Key Words:** Bronchial Challenge, Induced Sputum, Multiple Inert Gas  
Elimination Technique, Obstructive Airway Diseases.

## INTRODUCTION

Adenosine 5'-monophosphate (AMP) is a natural nucleoside and mediator of airway inflammation increasingly used in inhalational challenge tests even though its pathophysiology remains insufficiently understood. It is a potent indirect agent inducing bronchoprovocation through the activation of inflammatory pathways at the level of bronchial surface and through local or central neuronal reflexes.[1-3]

It has been suggested that AMP is a more potent inflammatory mediator than methacholine (MCh), a well-known bronchoconstrictive agent directly acting on airway tone. Compared to direct agents such as MCh and histamine, an AMP challenge could reproduce the lung function abnormalities spontaneously occurring during asthma attacks and better detect the inflammatory events of the disease at both central and peripheral airways.[4-8]

Accordingly, we hypothesized that AMP challenge in asthmatic patients could evoke greater gas exchange abnormalities and a different cellular response to that induced by MCh. There is evidence for a partial dissociation between expiratory airflow obstruction and gas exchange disturbances in acute asthma, since spirometric abnormalities, that mainly reflect narrowing in large and middle-sized airways, do not run in parallel to gas exchange abnormalities, better correlated to more peripheral airways dysfunction. [9-11]

On this basis we used the multiple inert gas elimination technique (MIGET) to comprehensively assess pulmonary gas exchange and induced sputum cellular composition after AMP and MCh challenges producing an equivalent degree of bronchoconstriction. The comparison of the results obtained by the two challenges could facilitate a better insight into the pathophysiology of AMP and asthma.

## MATERIAL AND METHODS

**Study population.** Twelve non-smokers patients (5 females) with stable mild asthma were recruited for the study, as approved by the Ethical Research Committee at Hospital Clínic of Barcelona (age,  $25\pm 1$  yr; FEV<sub>1</sub>,  $3.7\pm 0.2$  L [ $92\pm 4\%$  predicted], FEV<sub>1</sub>/FVC,  $80\pm 3\%$ ; PD<sub>20</sub>,  $0.4\pm 0.2$   $\mu$ mol). All subjects gave informed written consent after the purpose, risks and potential benefits of the study were explained to them. Inclusion criteria were: age between 18 and 45 yr; diagnosis of bronchial asthma according to GINA criteria [12]; FEV<sub>1</sub>>70% predicted (>1.5 L); a positive MCh (PD<sub>20</sub><1.9  $\mu$ mol) bronchial challenge on their first visit; absence of respiratory co-morbidities or any systemic or cardiopulmonary disease other than asthma; no asthma exacerbation within the last 6 weeks before the study; no treatment with systemic glucocorticosteroids in the previous 3 months. One patient received regular inhaled glucocorticosteroids (budesonide, 400  $\mu$ g b.i.d.) and two patients fixed inhaled combination therapy (budesonide/formoterol, 320/9  $\mu$ g b.i.d.). All patients referred use of rescue short-acting  $\beta_2$ -agonists.

**Study design.** A randomized, double-blinded, cross-over design was used. After a first visit to evaluate the clinical and functional status, a sample of induced sputum was collected in a second visit, one week later. Then, after another week, the patients were challenged with AMP and MCh (Sigma-Aldrich Química SA, Madrid, Spain), using a nebuliser attached to a breath-activated dosimeter (Spira Elektro2, Respiratory Care Center, Finlandia), one

week apart. The nebuliser was set to nebulise for 0.6 s at a pressure of 200 kPa with a flow of 0.6 L/s. During the challenges the patients breathed room air and were seated in an armchair. All asthma medication was withheld for 48 h before arrival to the laboratory and patients were asked to refrain from caffeine-containing beverages/foods for at least 12 h before testing. After ensuring steady-state conditions, a set of duplicate measurements of all variables was obtained (baseline). Maintenance of steady-state conditions was demonstrated by stability ( $\pm 5\%$ ) of both ventilatory and haemodynamic variables, and by the close agreement between duplicate measurements of mixed expired and arterial  $O_2$  and  $CO_2$  (within  $\pm 5\%$ ). The patients were then challenged with AMP/MCh following the recommended standardized procedures. [13-15] Doubling concentrations of AMP/MCh (from 0.39 to 400 mg/dL) were administered by 5 consecutive dosimeter inhalations until the 30% fall in  $FEV_1$  was achieved; at this point, resistance of respiratory system (Rrs) was also measured. Duplicate measurements were repeated after 5, 15, and 45 min. All sets consisted in the following steps in sequence:  $FEV_1$ , Rrs and ventilatory recordings; inert and respiratory gas sampling; and, haemodynamic recordings. Four hours after the beginning of each AMP/MCh challenge, induced sputum was collected. No patient needed rescue medication with short-acting bronchodilators during or immediately after the end of the study.

**Measurements.** On the day of the study, forced spirometry (CPF-S; Medical Graphics Corporation, St. Paul, MN USA) was performed according

to ATS recommendations, using our own predicted equations.[16][17] Rrs, PaO<sub>2</sub>, PaCO<sub>2</sub> and pH, the alveolar-arterial PO<sub>2</sub> difference (AaPO<sub>2</sub>), oxygen uptake (VO<sub>2</sub>) and carbon dioxide production (VCO<sub>2</sub>), minute ventilation (V<sub>E</sub>) and respiratory rate (RR) were measured, or calculated, as previously described. [18] MIGET was used to estimate the distributions of V<sub>A</sub>/Q ratios without sampling mixed venous inert gases. Cardiac output was directly measured by dye dilution technique (indocyanine green), in the customary manner.[18-19] A three-lead electrocardiogram, heart rate (HR) and systemic arterial pressure (Ps) and arterial oxygen saturation (SaO<sub>2</sub>) through a pulseoximeter (HP M1020A, Hewlett-Packard, Boblingen, Germany) were continuously recorded throughout the whole study (HP 1046A Monitor, Hewlett-Packard, Waltham, MA, USA).

**Induced sputum.** We performed the inhalation for the same period (7 min) of increasing concentrations of hypertonic saline (3%, 5%, 7%) following the technique described by Pizzichini et al.[20] The procedure was stopped when an adequate sputum sample was collected (approximately 1 g of plugs), or if troublesome symptoms were present. The sputum was processed within 30 and 120 min from the collection of the more viscid proportions of the sputum (plugs). The reproducibility of measurements was calculated by the intraclass correlation coefficient. In two patients, sputum data were not available due to their poor viability (<50%). Interleukin-8 (IL-8) in sputum supernatants was also assessed.

**Statistical analysis.** Results are expressed as mean±SE and 95% confidence interval (CI). The effects of AMP and MCh challenges on the different end-point variables were assessed by a two-way repeated analysis of variance (ANOVA). Whenever there was a significant difference, *post hoc* comparisons at each time point were performed using paired t-test. Sputum cell differences were assessed using Paired t-test. Pearson's rank test was used for correlations. All analyses were performed with SPSS version 10.1 (SPSS Inc, Chicago, IL, USA). Statistical significance was set at  $p < 0.05$  in all instances.

## RESULTS

**Baseline conditions.** There were no clinical or functional differences between MCh and AMP study days, all patients having normal  $V_A/Q$  distributions (Table 1).[21]

**Lung Function after AMP/MCh challenges.** Along with a comparable degree of bronchoconstriction ( $FEV_1$  fall: AMP,  $35\pm 2\%$ ; MCh,  $37\pm 2\%$ ; Rrs increase: AMP,  $114\pm 18\%$ ; MCh,  $92\pm 13\%$ ) both AMP and MCh challenges (cumulative doses,  $6.67\pm 13.33$  mg and  $0.15\pm 0.13$  mg) produced, at 5 min, similar gas exchange abnormalities, as shown by a decreased  $PaO_2$  (by  $31\pm 3\%$  and  $32\pm 3\%$ ) and an increased  $AaPO_2$ , respectively (Table 2). The arterial oxygenation defects observed after both challenges were essentially caused by mild-to-moderate  $V_A/Q$  imbalance, as reflected by increases in the dispersion of blood flow (Log SDQ, normal values,  $\leq 0.60$  [21]) and DISP R-E\* (normal values,  $\leq 3.0$ ) (Table 2).[22] Broadened unimodal  $V_A/Q$  patterns were observed in each participant. All these abnormalities showed at 15 and 45 min a similar trend to recover after both challenges, except for a mild, residual, increased Rrs and  $AaPO_2$  observed at the last time point. By contrast, compared with MCh, AMP induced at 5 min increases in  $VO_2$  (by  $10\pm 4\%$ ) and in  $Q_T$  (by  $20\pm 8\%$ ) ( $p < 0.05$  each);  $VO_2$  also resulted increased at 45 min and  $Q_T$  at 15 min (Table 2).



**Induced sputum.** Four hours after AMP challenge neutrophils increased from  $48\pm 7\%$  to  $62\pm 6\%$ , ( $p<0.05$ ) without other associated cellular changes (Table 3; Figure 1). By contrast, the cellular composition of induced sputum collected after MCh did not vary from baseline. The number of neutrophils in the baseline sputum showed a significant correlation with the concentration of IL-8 in the supernatant ( $r, 0.769$ ;  $p<0.01$ ) and with the increases in Rrs ( $r, 0.803$ ;  $p<0.01$ ) and AaPO<sub>2</sub> ( $r, 0.657$ ;  $p<0.05$ ) observed after AMP challenge (Figure 2). A significant correlation was also shown between the baseline concentration of IL-8 in the sputum supernatant and the increase in Rrs at 5 min on AMP study day ( $r, 0.726$ ;  $p<0.01$ ).

## DISCUSSION

There were two major novel findings in our study. Firstly, we demonstrated that AMP bronchoconstriction provoked, in patients with mild asthma, identical gas exchange abnormalities and  $V_A/Q$  imbalance than MCh while undergoing an equivalent degree of airflow limitation. Secondly, the AMP challenge caused mild neutrophilia in induced sputum 4 hours after challenge. In addition, we confirmed that AMP is an effective and clinically well-tolerated bronchoconstrictive agent that associates haemodynamic and metabolic effects.[6]

There is strong evidence in the literature that bronchial responsiveness to AMP correlates well with airways inflammatory markers (blood and sputum eosinophils) and that reflects acute changes in airways inflammation, as seen by therapeutical (corticosteroids) and/or environmental (allergens avoidance) interventions.[4][7][8] On this basis, we postulated that AMP challenge, by acting through inflammatory mechanisms, could be more influential in small, peripheral airways, thereby inducing greater gas exchange and  $V_A/Q$  abnormalities than MCh, a more centrally acting agent. Notwithstanding, our study failed to confirm our hypothesis, in that after both challenges patients showed a similar degree of hypoxaemia and increased  $AaPO_2$ , due to identical  $V_A/Q$  mismatching. It is generally held that gas exchange impairment in asthma is mainly induced by small airways, but experimental studies have shown that the  $V_A/Q$  imbalance becomes much more disrupted with larger airways narrowing. It is highly likely that the intense level of

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bronchoconstriction ( $\sim PD_{30}$ ) could hide any difference in  $V_A/Q$  mismatching due to the different bronchoconstrictive mechanisms of action of AMP and MCh.

Our study demonstrated that AMP is a well-tolerated effective bronchoconstrictive agent, with an average cumulative dose ratio to MCh of approximately 45. Furthermore, AMP challenge induced increases  $Q_T$  and  $VO_2$  at different points in time, possibly as a consequence of the inotropic effects of AMP and its metabolite adenosine, via the activation of specific receptors ( $A_2$ ).[23] These complementary effects need to be taken into account in the management of asthmatic patients with potential cardiovascular disturbances.

In conjunction with the functional abnormalities alluded to, there was a significant mild increase in neutrophils in induced sputum 4 hours after AMP inhalation. Although there is certain evidence of a correlation between cellular composition of the sputum and AMP responsiveness in asthmatic patients, there are few data regarding the effects of AMP challenge on induced sputum.[4] Van der Berge and coworkers showed an increase in eosinophils in the sputum collected 1 hour after AMP inhalation in patients with mild asthma, at variance with our finding.[24] Differences in patients' characteristics, inhalational procedures and, above all, in the time course of sputum collection after challenge might explain in part these discrepancies. Notwithstanding, it might be possible that our finding reflect a late, neutrophilic stage of the inflammatory process triggered by AMP challenge. By contrast, the study of Van der Berge may well reflect an earlier phase,

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mainly characterised by eosinophilic involvement. [24] In a previous work with mild asthmatic patients, we observed an increase in neutrophils in the sputum 4 hours after challenge with platelet-activating factor (PAF), a potent inflammatory mediator possibly involved, but never sufficiently proven, in the pathogenesis of asthma.[25] This late, neutrophilic response to PAF may thus support our hypothesis about the inflammatory response to AMP challenge. A different study in asthmatic patients challenged with inhaled leukotriene D4 (LTD4), demonstrated an increase of eosinophils in the sputum 4 hours after challenge, also shown by our group at a shorter time point (75 min) after LTD4. [26][27] The activation of a different pathway of bronchoconstriction (direct mechanisms of action) could explain such different findings. A confirmation of the validity of our “neutrophilic model of bronchoconstriction” comes from the interesting observation that a greater number of neutrophils in the baseline sputum and higher concentrations of IL-8 in the sputum supernatant, a chemotactic factor of neutrophils, correlated closely with the mechanic (Rrs) and gas exchange ( $AaPO_2$ ) abnormalities provoked by AMP. In summary, for the same degree of bronchoconstriction, AMP challenge did not induce more marked gas exchange disturbances than MCh, but caused mild neutrophilia in induced sputum at 4 hours after challenge.

## REFERENCES

1. Cushley MJ, Tattersfield AE, Holgate ST. Inhaled adenosine and guanosine on airway resistance in normal and asthmatic subjects. *Br J Clin Pharmacol* 1983; 15(2):161-165.
2. Van Schoor J, Joos GF, Pauwels RA. Indirect bronchial hyperresponsiveness in asthma: mechanisms, pharmacology and implications for clinical research. *Eur Respir J* 2000; 16(3):514-533.
3. Spicuzza L, Polosa R. The role of adenosine as a novel bronchoprovocant in asthma. *Curr Opin Allergy Clin Immunol* 2003; 3(1):65-69.
4. Van den Berge M, Meijer RJ, Kersjens HA. PC<sub>20</sub> adenosine 5'-monophosphate is more closely associated with airflow inflammation in asthma than PC<sub>20</sub> methacholine. *Am J Respir Crit Care Med* 2001; 163:1546-1550.
5. Joos GF. Bronchial hyperresponsiveness: too complex to be useful? *Curr Opin Pharmacol* 2003; 3(3):233-238.
6. Joos GF, O'Connor B, Anderson SD et al. Indirect airway challenges. *Eur Respir J* 2003; 21(6):1050-1068.
7. Van Den Berge M, Kerstjens HA, Meijer RJ et al. Corticosteroid-induced improvement in the PC<sub>20</sub> of adenosine monophosphate is more closely associated with reduction in airway inflammation than improvement in the PC<sub>20</sub> of methacholine. *Am J Respir Crit Care Med* 2001; 164(7):1127-1132.

8. Benckhuijsen J, van den Bos JW, van Velzen E et al. Differences in the effect of allergen avoidance on bronchial hyperresponsiveness as measured by methacholine, adenosine 5'-monophosphate, and exercise in asthmatic children. *Pediatr Pulmonol* 1996; 22(3):147-153.
9. Rodriguez-Roisin R, Roca J. Contributions of multiple inert gas elimination technique to pulmonary medicine. 3. Bronchial asthma. *Thorax* 1994; 49(10):1027-1033.
10. Wagner PD, Hedenstierna G, Rodriguez-Roisin R. Gas exchange, expiratory flow obstruction and the clinical spectrum of asthma. *Eur Respir J* 1996; 9(6):1278-1282.
11. Rodriguez-Roisin R. Acute severe asthma: pathophysiology and pathobiology of gas exchange abnormalities. *Eur Respir J* 1997; 10(6):1359-1371.
12. Global Initiative for Asthma ([www.ginasthma.com](http://www.ginasthma.com)).
13. Crapo RO, Casaburi R, Coates AL et al. Guidelines for methacholine and exercise challenge testing-1999. *Am J Respir Crit Care Med* 2000; 161(1):309-329.
14. Sterk PJ, Fabbri LM, Quanjer PH et al. Airway responsiveness. Standardized challenge testing with pharmacological, physical and sensitizing stimuli in adults. *Eur Respir J Suppl* 1993; 16:53-83.
15. Cain H. Bronchoprovocation testing. *Clin Chest Med* 2001; 22(4):651-659.
16. Standardization of Spirometry, 1994 Update. American Thoracic Society. *Am J Respir Crit Care Med* 1995; 152(3):1107-1136.
17. Roca J, Sanchis J, Agusti-Vidal A et al. Spirometric reference values from a Mediterranean population. *Bull Eur Physiopathol Respir* 1986; 22(3):217-224.

18. Echazarreta AL, Gomez FP, Ribas J et al. Pulmonary gas exchange responses to histamine and methacholine challenges in mild asthma. *Eur Respir J* 2001; 17(4):609-614.
19. Roca J, Wagner PD. Contribution of multiple inert gas elimination technique to pulmonary medicine. 1. Principles and information content of the multiple inert gas elimination technique. *Thorax* 1994; 49(8):815-824.
20. Pizzichini E, Pizzichini MM, Efthimiadis A et al. Indices of airway inflammation in induced sputum: reproducibility and validity of cell and fluid-phase measurements. *Am J Respir Crit Care Med* 1996; 154(2 Pt 1):308-317.
21. Cardús J, Burgos F, Diaz O et al. Increase in pulmonary ventilation-perfusion inequality with age in healthy individuals. *Am J Respir Crit Care Med* 1997; 156:648-653.
22. Gale GE, Torre-Bueno JR, Moon RE et al. Ventilation-perfusion inequality in normal humans during exercise at sea level and simulated altitude. *J Appl Physiol*. 1985;58(3):978-88.
23. Tabrizchi R, Bedi S. Pharmacology of adenosine receptors in the vasculature. *Pharmacol Ther*. 2001;91(2):133-47.
24. Van den Berge M, Kerstjens HA, de Reus DM et al. Provocation with adenosine 5'-monophosphate, but not methacholine, induces sputum eosinophilia. *Clin Exp Allergy* 2004;34(1):71-6.
25. Gabrijelcic J, Acuña A, Profita M et al. Neutrophil airway influx by platelet-activating factor in asthma: role of adhesion molecules and LTD4 expression. *Eur Respir J* 2003 ;22 :290-297.

26. Diamant Z, Hilterman JT, van Rensen EL et al. The effect of inhaled leukotriene D4 and methacholine on sputum cell differentials in asthma. *Am J Respir Crit care Med.* 1997; 155(4):1247-53.
27. Echazarreta AL, Dahlen B, García G et al. Pulmonary gas exchange and sputum cellular responses to inhaled leukotriene D4 in asthma. *Am J Respir Crit Care Med.* 2001;164:202-206.



**Table 1. Basal conditions on AMP and MCh days.**

	<b>AMP</b>	<b><u>MCh</u></b>
<b>FEV<sub>1</sub>, L</b>	3.2 ± 0.2	3.2 ± 0.2
<b>FEV<sub>1</sub>, % predicted</b>	85 ± 6	83 ± 6
<b>FEV<sub>1</sub>/FVC %</b>	81 ± 3	81 ± 2
<b>Rrs, cm H<sub>2</sub>O·L<sup>-1</sup>·s</b>	4.6 ± 0.3	4.6 ± 0.4
<b>PaO<sub>2</sub>, KPa</b>	13.4 ± 0.3	13.5 ± 0.2
<b>AaPO<sub>2</sub>, KPa</b>	0.52 ± 0.16	0.27 ± 0.12
<b>VO<sub>2</sub>, mL·min<sup>-1</sup></b>	197 ± 9	186 ± 12
<b>Q<sub>T</sub>, L·min<sup>-1</sup></b>	5.7 ± 0.3	5.6 ± 0.3
<b>Log SDQ</b>	0.47 ± 0.04	0.44 ± 0.02
<b>DISP R-E*</b>	3.15 ± 0.49	2.38 ± 0.26

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Rrs: respiratory system resistance; PaO<sub>2</sub>: arterial oxygen partial pressure; AaPO<sub>2</sub>: alveolar-arterial oxygen partial pressure difference; VO<sub>2</sub>: oxygen uptake; Q<sub>T</sub>: cardiac output; Log SDQ: dispersion of pulmonary blood flow (dimensionless); DISP R-E\*: dispersion of retention minus excretion inert gases corrected for dead space (dimensionless).

**Table 2. Mean differences (and 95% IC) after AMP and MCh challenges.**

		<u>5 min</u>	<u>15 min</u>	<u>45 min</u>	<u>p value</u> <sup>†</sup>
<b>FEV<sub>1</sub>, L</b>	AMP	-1.1 (-1.3 to -1.0)	-0.6 (-0.8 to -0.5)	-0.3 (-0.4 to -0.1)	NS
	MCh	-1.2 (-1.4 to -1.0)	-0.7(-0.8 to -0.5)	-0.2 (-0.4 to -0.1)	
<b>Rrs, cm H<sub>2</sub>O·L<sup>-1</sup>·s</b>	AMP	4.9 (3.4 to 6.5)	3.7 (0.9 to 6.6)	1.6 (0.2 to 3.0)	NS
	MCh	4.3 (2.9 to 5.6)	2.7 (1.4 to 3.9)	1.9 (1.1 to 2.7)	
<b>PaO<sub>2</sub>, KPa</b>	AMP	-4.1 (-4.9 to -3.2)	-2.1 (-3.1 to -1.3)	-0.4 (-1.2 to 0.5)	NS
	MCh	-4.3 (-5.3 to -3.3)	-2.7 (-3.9 to -1.5)	-0.9 (-1.9 to 0.1)	
<b>AaPO<sub>2</sub>, KPa</b>	AMP	3.3 (2.7 to 4.0)	1.7 (0.9 to 2.5)	0.7 (0.4 to 1.6)	NS
	MCh	3.7 (2.8 to 4.5)	2.3 (1.6 to 3.1)	0.8 (0.3 to 1.3)	
<b>VO<sub>2</sub>, mL·min<sup>-1</sup></b>	AMP	21 (2 to 40) <sup>‡</sup>	10 (-6 to 26)	17 (0.0 to 34) <sup>‡</sup>	<0.05
	MCh	8 (-8 to 23)	1 (-12 to 15)	-3 (-22 to 16)	
<b>Q<sub>T</sub>, L·min<sup>-1</sup></b>	AMP	1.0 (0.2 to 1.8) <sup>‡</sup>	0.6 (0.1 to 1.1) <sup>‡</sup>	0 (-0.3 to 0.3)	<0.01
	MCh	0.2 (-0.2 to 0.5)	0 (-0.45 to 0.49)	-0.2 (-0.6 to 0.1)	
<b>Log SDQ</b>	AMP	0.27 (0.18 to 0.36)	0.13 (0.03 to 0.23)	0.05 (0.04 to 0.14)	NS
	MCh	0.38 (0.26 to 0.49)	0.27 (0.17 to 0.36)	0.12 (0.06 to 0.18)	
<b>DISP R-E*</b>	AMP	3.77 (2.25 to 5.28)	1.41 (0.22 to 2.60)	0.33 (-1.45 to 0.80)	NS
	MCh	4.59 (3.09 to 6.08)	3.26 (1.75 to 4.77)	1.65 (0.81 to 2.49)	

For abbreviations see Table 1.

<sup>†</sup> Significance of the interaction between the effects of the bronchoprovocative agent and time course (repeated measures ANOVA);

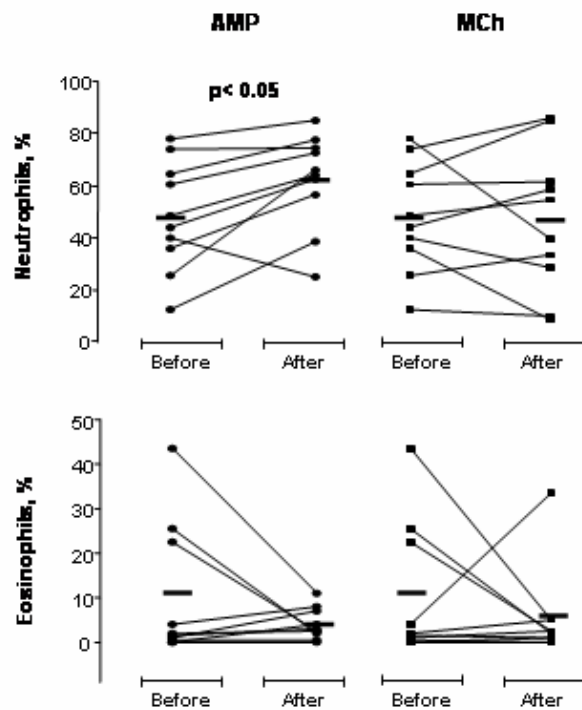
<sup>‡</sup> p<0.05 compared with MCh (paired t-test).

**Table 3. Induced sputum analysis before and after AMP and MCh challenges**

	<u>Baseline</u>	<u>AMP</u>	<u>MCh</u>
<b>Total Cell Count, x10<sup>4</sup>·g<sup>-1</sup></b>	1,124 ± 215	1,136 ± 278	985 ± 230
<b>Cell viability, %</b>	71 ± 3	71 ± 5	71 ± 3
<b>Squamous cells, %</b>	3 ± 1	4 ± 1	4 ± 1
<b>Macrophages, %</b>	36 ± 5	31 ± 5	43 ± 8
<b>Lymphocytes, %</b>	4 ± 0	2 ± 1	4 ± 1
<b>Neutrophils, %</b>	48 ± 7	62 ± 6*	47 ± 9
<b>Eosinophils, %</b>	10 ± 5	4 ± 1	5 ± 3
<b>IL-8, pg/mL</b>	2204 ± 702	1624 ± 522	1246 ± 206

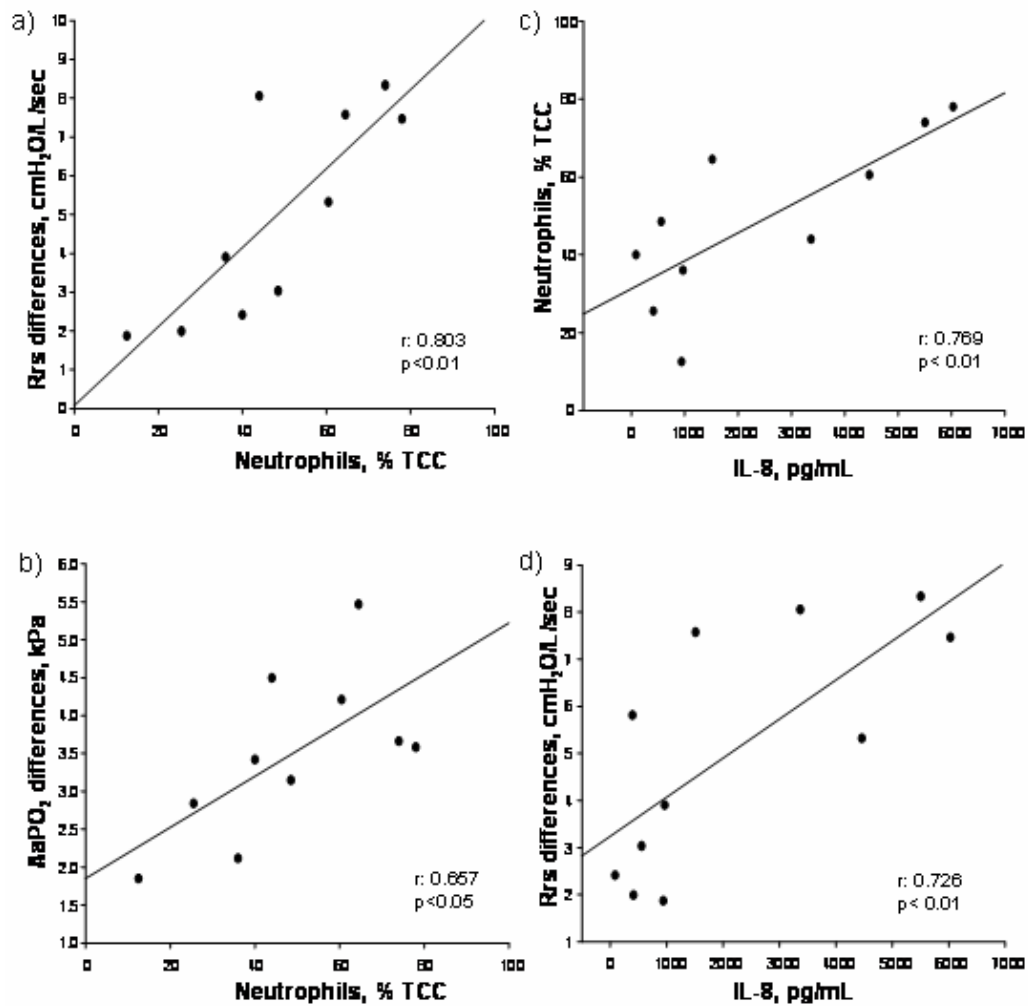
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Values are mean ± SE; \* p: 0.02 compared with baseline (paired t-test).



**Figure 1.**

Neutrophils and eosinophils in the induced sputum before and after each challenge, expressed as percentage of total cell count (TCC).



**Figure 2.**

Plots between changes in Rrs (Fig. 2a) and AaPO<sub>2</sub> (Fig. 2b) after AMP with the number of neutrophils in the baseline sputum; plots between neutrophils in the baseline sputum (Fig. 2c) and changes in Rrs after AMP (Fig. 2d) with IL-8 concentration in sputum supernatant. For abbreviations see Table 1 and Figure 1.

*EFFECTS OF NEBULIZED SALBUTAMOL ON  
PULMONARY GAS EXCHANGE DURING COPD  
EXACERBATIONS  
AND IN STABLE CONDITIONS*

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**Running head:** *COPD and salbutamol*

**Key Words:** COPD exacerbations, Multiple Inert Gas Elimination Technique,  
nebulized salbutamol, Pulmonary gas exchange.

## INTRODUCTION

Episodes of exacerbations are the most common complications in the natural history of chronic obstructive pulmonary diseases (COPD). These episodes are always characterized by a significant worsening in pulmonary gas exchange that results in severe hypoxemia with or without hypercapnia [1]. Both arterial hypoxemia and hypercapnia are modulated by a complex interplay between intrapulmonary (ventilation-perfusion [ $V_A/Q$ ] mismatching) and extrapulmonary (minute ventilation [ $V_E$ ], cardiac output [ $Q_T$ ], and oxygen consumption [ $VO_2$ ]) factors [2]. It has been demonstrated that the principal mechanism of arterial hypoxemia in patients with COPD is  $V_A/Q$  imbalance, both in stable conditions and during exacerbations [3].

Short-acting  $\beta_2$ -agonists [SABAs] are one of the mainstays of treatment of COPD exacerbations [4].

SABAs are potent bronchodilators but also vasoactive agents with inotropic and chronotropic effects, hence potentially influencing different aspects of cardiopulmonary function [5]. However, a comprehensive assessment of the effects of such bronchodilators on intrapulmonary and extrapulmonary determinants of gas exchange during COPD exacerbations remains unsettled. Accordingly, we investigated in patients with COPD exacerbations needing hospitalization the effects of a therapeutic dose of nebulized salbutamol (5.0 mg) on pulmonary gas exchange using the multiple inert gas elimination technique (MIGET).

The same patients were studied during stable conditions, 2-3 months after their episode of exacerbation in order to assess how the factors contributing to gas exchange abnormalities vary during recovery in response to nebulized salbutamol and to facilitate, therefore, a better insight into the pathophysiology of COPD.



## MATERIAL AND METHODS

**Study population.** We studied 20 patients (1 female; 5 smokers, 15 ex-smokers,  $58 \pm 8$  pack-years), mean ( $\pm$  SEM) age  $67 \pm 2$  yrs who fulfilled the criteria for the diagnosis of COPD [6] and were admitted to hospital because of an acute exacerbation of their disease, according to the decision of the attending physicians. At the inclusion in the study ten patients reported a history of more than 10 yrs of COPD, 6 patients between 5 and 10 yrs, 4 patients less than 5 yrs. Pulmonary function tests performed during stable conditions (before or after 2-5 months the exacerbation episode) showed a severe airflow obstruction (post-bronchodilator FEV<sub>1</sub>:  $39 \pm 4\%$  of pred.; FVC:  $2.8 \pm 0.2$  L; FEV<sub>1</sub>/FVC ratio:  $0.40 \pm 0.03$ ), a normal total lung capacity (TLC:  $108\% \pm 4$  of pred.) a severe lung hyperinflation (RV:  $187 \pm 13\%$  of pred.; IC/TLC:  $0.28 \pm 0.03$ ) and a moderate-to-severe reduction of carbon monoxide transfer factor (DL<sub>CO</sub>:  $53 \pm 4\%$  of pred.; K<sub>CO</sub>:  $65 \pm 5\%$  of pred.). Patients showed moderate hypoxemia and mild hypercapnia (PAO<sub>2</sub>:  $65.7 \pm 2.7$  mmHg; PaCO<sub>2</sub>:  $42.1 \pm 1.5$  mmHg). As two patients died for medical complications before completing the study (one for an acute intestinal occlusion, the other during an exacerbation of the respiratory disease) and other two were lost at follow-up (multiple re-exacerbations) data of lung volumes, DL<sub>CO</sub> and arterial blood gases in stable conditions are available only from 16 patients. All patients were in GOLD stages III or IV. On admission to the emergency room, patients showed moderate hypoxemia (PaO<sub>2</sub>  $62.2 \pm 2.2$  mmHg) and hypercapnia (PaCO<sub>2</sub>  $44.4 \pm 3.0$  mmHg) with a

normal arterial pH ( $7.42 \pm 0.01$ ) even considering that in 6 and 3 cases, respectively, the first measurement of arterial gases was performed with supplemental oxygen of 1 and 2 L/min. Patients with complications (pneumonia, pulmonary thromboembolism, pleural effusion) and with comorbidities, like bronchial asthma, lung cancer and cardiovascular diseases or requiring mechanical ventilation were excluded from the study. All patients received standard treatment with supplemental oxygen, inhaled bronchodilators (salbutamol and ipratropium bromure), intravenous corticosteroids and antibiotics. The mean length of hospital stay was of  $7 \pm 1$  days.

**Study design.** Patients were studied within the first 4 days (median, 3) of hospitalization, when they were able to breathe through the breathing circuit and 2-3 months after hospital discharge, once they were in stable condition and in absence of any other re-exacerbation of the respiratory disease from the first study. Medication as decided by the attending physician was maintained unmodified. Only one patient received supplemental oxygen therapy at 1.5 L/min (16 hours daily) after the exacerbation. During the studies all patients were afebrile, breathed room air and were seated in an armchair. Measurements were performed before (at least 6 hrs after the last dose of inhaled bronchodilators and intravenous corticosteroids) and at 30 and 90 min after nebulization of 5mg of salbutamol. After ensuring steady-state conditions, a set of duplicate measurements of all variables was obtained at each time point. Maintenance of steady-state conditions was demonstrated by stability ( $\pm 5\%$ ) of both ventilatory and

haemodynamic variables, and by the close agreement between duplicate measurements of mixed expired and arterial O<sub>2</sub> and CO<sub>2</sub> (within ±5%). All sets of measurements consisted in the following steps in sequence: FEV<sub>1</sub>, forced vital capacity (FVC), inspiratory capacity (IC) and ventilatory recordings; inert and respiratory gas sampling; haemodynamic recordings (systemic arterial pressure; cardiac output, Q<sub>T</sub>; heart rate, HR).

Spontaneous sputum was collected before and between 30 and 90 min after salbutamol.

**Measurements.** On the day of the study, forced spirometry and Inspiratory capacity (IC) (CPF-S; Medical Graphics Corporation, St. Paul, MN USA) were performed according to ATS/ERS recommendations, using our own predicted equations.[7][8] PaO<sub>2</sub>, PaCO<sub>2</sub> and pH, alveolar-arterial PO<sub>2</sub> difference (AaPO<sub>2</sub>), oxygen uptake (VO<sub>2</sub>) and carbon dioxide production (VCO<sub>2</sub>), minute ventilation (V<sub>E</sub>) and respiratory rate (f) were measured, or calculated, as previously described. [9] MIGET was used to estimate the distributions of V<sub>A</sub>/Q ratios without sampling mixed venous inert gases. Unfortunately, V<sub>A</sub>/Q data were lost for technical problems in 4 studies of the exacerbation phase and in one study of stable phase. Q<sub>T</sub> was directly measured by dye dilution technique (indocyanine green), in the customary manner.[9][10] A three-lead electrocardiogram, heart rate (HR) and systemic arterial pressure and arterial oxygen saturation (SaO<sub>2</sub>) through a pulseoximeter (HP M1020A, Hewlett-Packard, Boblingen, Germany) were continuously recorded throughout the whole study (HP 1046A Monitor,

Hewlett-Packard, Waltham, MA, USA). Exhaled breath condensate (EBC) was also collected before and at 90 min after salbutamol using the EcoScreen (Jaeger, Germany), according to ATS/ERS recommendations. [11]

**Statistical analysis.** Results are expressed as mean  $\pm$  SE. The effects of nebulized salbutamol on the different end-point variables in each phases of the study (exacerbation and stable condition) were assessed by a one-way and two-ways repeated analysis of variance (ANOVA). Whenever there was a significant difference, *post hoc* comparisons at each time point were performed using paired t-test. All analyses were performed with SPSS version 11.1 (SPSS Inc, Chicago, IL, USA). Statistical significance was set at  $p < 0.05$  in all instances.

## RESULTS

### Exacerbation.

At baseline, patients showed a severe degree of bronchoconstriction (FVC,  $36 \pm 2\%$  of predicted; FEV<sub>1</sub>,  $24 \pm 2\%$  of predicted), with high values of V<sub>E</sub> and f (Table 1). Mean PaO<sub>2</sub> was moderately to severely reduced (range, 45.7 - 76.0 mmHg) while PaCO<sub>2</sub> was moderately increased (range, 33.3 - 68.2 mmHg). Gas exchange abnormalities run in parallel with moderate-to-severe V<sub>A</sub>/Q imbalance: the dispersions of both blood flow (Log SDQ, range: 0.60 - 1.75) and ventilation (Log SDV, range: 0.70 - 1.26) distributions and the overall index of V<sub>A</sub>/Q heterogeneity (DISP R-E\*, range: 8.7 - 20.7) were increased (Table 1). Shunt (percentage of blood flow to units with V<sub>A</sub>/Q ratio < 0.005) and areas with high (10 - 100) V<sub>A</sub>/Q ratios were negligible, while areas with low (< 0.1, excluding shunt) V<sub>A</sub>/Q ratios and dead space (> 100 V<sub>A</sub>/Q ratios) were moderately increased. Patients showed increased Q<sub>T</sub> and HR with normal values of mean arterial pressure (Ps) and O<sub>2</sub> uptake (VO<sub>2</sub>). At 30 min, nebulized salbutamol significantly improved FEV<sub>1</sub> (by 14%) and IC (by 10%), while V<sub>E</sub> and f remained stable. PaO<sub>2</sub> and PaCO<sub>2</sub> slightly decreased along further significant increases in AaPO<sub>2</sub> (by 2.0 mmHg), in Log SDQ (by 7%) and in areas with low V<sub>A</sub>/Q ratios (by 27%). In addition, salbutamol increased Q<sub>T</sub> (by 13%) and HR (by 11%) and decreased Ps (by 8 mmHg). At 90 min, in comparison with baseline, FEV<sub>1</sub>, IC, were still increased; respiratory (PaO<sub>2</sub>, PaCO<sub>2</sub>, AaPO<sub>2</sub>) and inert (Log SDQ, Low V<sub>A</sub>/Q,

DISPR-E\*) gas exchange indices showed a trend toward baseline. Hemodynamic changes (increased HR and decreased Ps) were still present at 90 min, except for  $Q_T$  that showed a trend toward baseline.  $VO_2$  remained stable during all the study.

### **Stable conditions.**

Nine patients of the initial sample were excluded from the second phase of the study for multiple re-exacerbations and two individuals, as previously described, died. The remaining nine patients repeated the study  $14 \pm 3$  weeks after discharge, once they were in clinical stable conditions. At baseline, as compared to exacerbation, patients had lower  $V_E$  ( $p < 0.05$ ), a comparable  $f$  and a similar airflow obstruction but less hyperinflation, as shown by greater IC ( $p < 0.02$ ) (Table 2). Gas exchange impairment and  $V_A/Q$  inequality were significantly minor under stable condition as shown by  $PaO_2$ ,  $AaPO_2$  and Log SDQ ( $p < 0.05$  each). Cardiac Output and HR ( $p < 0.05$ ), as expected, were lower than during exacerbation. No significant differences were observed in Ps and  $VO_2$ . At 30 min, nebulized salbutamol induced significant increases in  $FEV_1$  (by 17%) and IC (by 17%) without any change in the ventilatory pattern ( $V_E$ ,  $f$ ) (Table 3). We also observed significant decrease in  $PaO_2$  (by 6.7 mmHg) and increase of  $AaPO_2$  (by 5.3 mmHg), with an unchanged  $PCO_2$ . Arterial deoxygenation was associated with a significant worsening in  $V_A/Q$  imbalance as shown by increases in Log SDQ (by 15%) and DISP R-E\* (by 33%). In addition, salbutamol induced

significant increases in  $Q_T$  (by 23%), HR (by 17%) and  $VO_2$  (by 24 mL/min) and decreased  $P_s$  (by 6 mmHg). At 90 min  $FEV_1$  and IC were still increased, the arterial deoxygenation ( $PaO_2$ ,  $AaPO_2$ ) was similar to 30 min, such as the increases in Log SDQ and DISPR-E\* (Table 3). Also the hemodynamic changes (increased  $Q_T$  and HR, decreased  $P_s$ ) were still present at 90 min, while  $VO_2$  returned to baseline.

### **Comparison between exacerbation and stable COPD.**

By comparing the effects of salbutamol observed during both phases of the study (9 patients, 7 with complete  $V_A/Q$  data) through a two-ways ANOVA analysis, we detected similar spirometric and hemodynamic responses, as shown by  $FEV_1$ , IC and  $Q_T$  time courses in Figure 1. By contrast, whereas arterial oxygenation showed only negligible changes during the exacerbation,  $PaO_2$  in stable COPD decreased by  $7 \pm 2$  mmHg at 30 min and by  $6 \pm 2$  mmHg at 90 min and  $AaPO_2$  increased by  $5 \pm 1$  mmHg (30 min) and  $6 \pm 1$  mmHg (90 min) ( $p < 0.01$  each). Similarly,  $V_A/Q$  inequality worsening after salbutamol resulted significantly greater and longer under stable conditions, as shown by DISPR-E\* time course (Figure 1). In fact, during the exacerbation DISPR-E\* increased by 6% at 30 min and returned to baseline at 90 min; by contrast under stable conditions we observed an increase of 31% up to 90 min ( $p < 0.05$ ).

## DISCUSSION

During COPD exacerbations nebulized salbutamol induced significant bronchodilator (increases in FEV<sub>1</sub> and IC) and hemodynamic (increased Q<sub>T</sub> and heart rate, decreased MAP) effects up to 90 min after nebulized salbutamol. We also observed a temporary, further, deterioration of V<sub>A</sub>/Q distribution accompanied by a mild gas exchange worsening 30 min after salbutamol.

The same patients in stable convalescence showed a similar response to salbutamol in terms of spirometric and hemodynamic changes, even though at baseline they were in better clinical and functional conditions. By contrast, worsening in gas exchange and V<sub>A</sub>/Q imbalance were much greater in stable conditions and lasted up to 90 min after salbutamol.

The strict relation between gas exchange and V<sub>A</sub>/Q imbalance has been widely investigated. Barberá and coworkers by studying COPD exacerbated patients, estimated that 46% of hypoxemia is attributable to V<sub>A</sub>/Q inequality, whether a 28% to the combined effect of increased VO<sub>2</sub> and Q<sub>T</sub>[3]. In our study VO<sub>2</sub> did not appear to play a significant role in both phases of the study while Q<sub>T</sub> was higher during the exacerbation. As Q<sub>T</sub> increase is able to augment the oxygen concentration of mixed venous blood and potentially counterbalance the effects of V<sub>A</sub>/Q imbalance on arterial oxygenation, it has often supposed an important contribution of Q<sub>T</sub> in determining PaO<sub>2</sub> in COPD exacerbations.



By contrast, our study shows similar  $Q_T$  responses after salbutamol in both COPD exacerbation and stable convalescence, but a significant deoxygenation only in stable COPD, suggesting only a limited role of  $Q_T$  in determining  $PaO_2$ .

A worsening of arterial oxygenation and  $V_A/Q$  balance after  $\beta_2$ -agonists administration has been previously described and ascribed to the release of hypoxic pulmonary vasoconstriction [12-16]. Ringsted and coworkers, in comparison with our study, observed an analogous response of  $V_A/Q$  distributions after the administration of intravenous terbutaline in stable COPD patients with a similar degree of bronchoconstriction[14]. Viegas et al. also described similar data with the administration of nebulized fenoterol in stable COPD patients [16]. Our data confirm this hypothesis as  $PaO_2$  fall in stable conditions runs in parallel with the increased dispersion of pulmonary blood flow (Log SDQ), suggesting pulmonary vasodilatation.

However, this phenomenon appears to be mild during COPD exacerbations as probably in presence of more hypoxemia, airflow obstruction, hyperinflation and higher hyperkinetic vascular state, the pulmonary vascular tone is more hypoxic and more constricted, hence less liable to vasodilatation. By contrast, under stable conditions gas exchange response to salbutamol is much greater as, in less hypoxemic conditions, the pulmonary vasculature is less constricted and more sensitive to vasodilatation i.e. hypoxic pulmonary vasoconstriction release and further  $V_A/Q$  worsening.

Similarly, the work of Ringsted showed minor  $V_A/Q$  and gas exchange abnormalities in those patients who presented very severe airways

obstruction, likely because pulmonary vascular tone and its flexibility are limited in more advanced stages of COPD such as during exacerbations [15]. A clinical implication of this finding is that particular attention is necessary when SABA are administered to COPD patients as arterial oxygenation can temporarily worsen notwithstanding the general improvement of dyspnoea that is likely more related, to the degree of hyperinflation than to PaO<sub>2</sub> [17].

In conclusion, this work by offering a complete and paired analysis of all parameters modulating respiratory gas exchange under different clinical conditions, clarifies effects and mechanisms of action of  $\beta_2$ -agonists in COPD and, contemporary, underlies the central role of hypoxic pulmonary vasoconstriction in regulating V<sub>A</sub>/Q and gas exchange efficacy in COPD.

## REFERENCES

1. Rodriguez-Roisin R, Barberà JA, Roca J. Pulmonary gas exchange. In Calverley PMA, McNee W, Pride NB, Rennard SI eds. Chronic obstructive pulmonary disease (2nd edition). Arnold, London 2003; 175-193.
2. Ferrer A, Rodriguez-Roisin R. Ventilation-Perfusion distributions in disease. In: Martin JG, ed. The physiological basis of respiratory disease. BC Decker Inc, Hamilton 2005; 185-202.
3. Barberà JA, Roca J, Ferrer A, Fèlez MA, Díaz O, Roger N, Rodriguez Roisin R. Mechanisms of worsening gas exchange during acute exacerbations of chronic obstructive pulmonary disease. *Eur Resp J* 1997; 10:1285-91.
4. Pauwels RA, Buist AS, Calverley PM, Jenkins CR, Hurd SS; GOLD Scientific Committee. Global strategy for the diagnosis, management, and prevention of chronic obstructive pulmonary disease. NHLBI/WHO Global Initiative for Chronic Obstructive Lung Disease (GOLD) Workshop summary. *Am J Respir Crit Care Med.* 2001 Apr;163(5):1256-76.
5. Crane J, Burgess C, Beasley R. Cardiovascular and hypokalaemic effects of inhaled salbutamol, fenoterol, and isoprenaline. *Thorax.* 1989 Feb;44(2):136-40.
6. Standards for the diagnosis and treatment of patients with COPD: a summary of the ATS/ERS position paper. *ERJ* 2004, 23:932-946.
7. M. R. Miller, J. Hankinson, V. Brusasco, F. Burgos, R. Casaburi, A. Coates, R. Crapo, P. Enright, C. P. M. van der Grinten, P. Gustafsson, R. Jensen, D. C. Johnson, N. MacIntyre, R. McKay, D. Navajas, O. F. Pedersen, R. Pellegrino, G. Viegi, and J. Wanger. Standardisation of spirometry. *Eur. Respir. J.*, Aug 2005; 26: 319 - 338.

8. Roca J, Sanchis J, Agusti-Vidal A et al. Spirometric reference values from a Mediterranean population. *Bull Eur Physiopathol Respir* 1986; 22(3):217-224.
9. Echazarreta AL, Gomez FP, Ribas J et al. Pulmonary gas exchange responses to histamine and methacholine challenges in mild asthma. *Eur Respir J* 2001; 17(4):609-614.
10. Roca J, Wagner PD. Contribution of multiple inert gas elimination technique to pulmonary medicine. 1. Principles and information content of the multiple inert gas elimination technique. *Thorax* 1994; 49(8):815-824.
11. Horváth I, Hunt J, Barnes PJ. On behalf of the ATS/ERS Task Force on Exhaled Breath Condensate Exhaled breath condensate: methodological recommendations and unresolved questions. *Eur. Respir. J., Sep 2005; 26: 523 - 548.*
12. Bernasconi M, Brandolese R, Poggi R, et al. Dose/response effects and time course of inhaled fenoterol on respiratory mechanisms and arterial oxygen tension in mechanically ventilated patients with chronic airflow obstruction. *Intensive Care Med* 1990;16:108-14.
13. Carlone S, Angelici E, Palange P, et al. Effects of fenoterol on oxygen transport in patients with chronic airflow obstruction. *Chest* 1988; 93:790-94.
14. Ringsted CV, Eliassen K, Andersen JB, Heslet L, Qvist J. Ventilation-perfusion distributions and central hemodynamics in chronic obstructive pulmonary disease. Effects of terbutaline administration. *Chest*. 1989 Nov;96(5):976-83.
15. Ferrer A, Viegas C, Montserrat JM, Roca J, Wagner PD, Rodriguez-Rosin R. Gas exchange responses to bronchodilators in chronic obstructive pulmonary disease. *Am Rev Respir Dis* 1991;143:A447.

16. Viegas CA, Ferrer A, Montserrat JM, Barberà JA, Roca J, Rodriguez-Roisin R. Ventilation-perfusion response after fenoterol in hypoxemic patients with stable COPD. *Chest* 1996;110(1):71-77.
17. Mahler DA. Mechanisms and measurement of dyspnea in chronic obstructive pulmonary disease. *Proc Am Thorac Soc*. 2006 May;3(3):234-8. Review.

**Table 1. Acute effects of salbutamol during COPD exacerbations (n= 20).**

	Baseline	30 min	90 min	p <sup>†</sup>
<b>FEV<sub>1</sub>, L</b>	0.73 ± 0.06	0.83 ± 0.07**	0.81 ± 0.06**	<0.01
<b>IC, L</b>	1.62 ± 0.13	1.76 ± 0.13**	1.77 ± 0.13**	<0.01
<b>V<sub>E</sub>, L/min</b>	8.6 ± 0.4	8.9 ± 0.4	8.6 ± 0.5	NS
<b>f, min<sup>-1</sup></b>	17 ± 1	18 ± 1	18 ± 1	NS
<b>PaO<sub>2</sub>, mmHg</b>	61.1 ± 2.0	59.1 ± 2.0	60.7 ± 2.0	NS
<b>PaCO<sub>2</sub>, mmHg</b>	46.2 ± 2.1	44.7 ± 1.9**	44.3 ± 1.6	<0.02
<b>AaPO<sub>2</sub>, mmHg</b>	36.5 ± 2.5	38.5 ± 2.3**	35.2 ± 2.5	<0.01
<b>HR, min<sup>-1</sup></b>	90 ± 3	99 ± 4**	95 ± 3**	<0.01
<b>Ps, mmHg.</b>	95 ± 3	87 ± 2**	89 ± 2**	<0.01
<b>Q<sub>T</sub>, L/min</b>	7.1 ± 0.4	8.0 ± 0.5**	7.3 ± 0.4	<0.01
<b>VO<sub>2</sub>, mL/min</b>	242 ± 7	249 ± 8	257 ± 8	NS
<b>Shunt, % of Q<sub>T</sub></b>	1.8 ± 0.7	2.0 ± 0.8	1.7 ± 0.7	NS
<b>Low V<sub>A</sub>/Q, % Q<sub>T</sub></b>	9.5 ± 2.4	12.6 ± 3.0**	8.7 ± 2.2	<0.05
<b>Log SDQ</b>	1.17 ± 0.07	1.23 ± 0.07**	1.13 ± 0.08	<0.01
<b>Log SDV<sub>E</sub></b>	1.00 ± 0.04	0.99 ± 0.04	0.98 ± 0.04	NS
<b>High V<sub>A</sub>/Q, % V<sub>E</sub></b>	1.3 ± 0.7	1.4 ± 0.7	1.1 ± 0.4	NS
<b>Dead space, % V<sub>E</sub></b>	34.5 ± 2.0	35.8 ± 1.9	35.2 ± 2.0	NS
<b>DISP R-E*</b>	14.9 ± 0.8	15.9 ± 0.9	14.0 ± 1.0**	<0.01

<sup>†</sup> Repeated measures ANOVA; \*\* p<0.05, post-hoc analysis (paired t-test) compared with baseline. Definition of the abbreviations. f: respiratory rate; AaPO<sub>2</sub>: alveolar-arterial oxygen difference; MAP: mean systemic arterial pressure; Shunt, % of Q<sub>T</sub>: % of Q<sub>T</sub> to lung units with V<sub>A</sub>/Q ratios<0.05. Low V<sub>A</sub>/Q, % Q<sub>T</sub>: % of Q<sub>T</sub> to lung units with V<sub>A</sub>/Q ratio< 0.1, excluding shunt; Log SDQ: dispersion of blood flow distribution (normal, 0.30-0.65); Log SDV: dispersion of the alveolar ventilation distribution (normal, 0.30-0.60); High V<sub>A</sub>/Q, % V<sub>E</sub>: % of V<sub>A</sub> to lung units with V<sub>A</sub>/Q ratio>10, excluding dead space; dead space: % of V<sub>A</sub> to lung units with V<sub>A</sub>/Q ratio> 100; DISPR-E\*: dispersion of retention minus excretion inert gases corrected for dead space (normal, ≤ 3).

**Table 2. Spirometric, hemodynamic and gas exchange data during exacerbations and under stable conditions (n= 9).**

	<b>Exacerbation</b>	<b>Stable Conditions</b>	<b>p<sup>†</sup></b>
<b>FEV<sub>1</sub>, L</b>	0.76 ± 0.08	0.97 ± 0.16	NS
<b>FEV<sub>1</sub>, % pred.</b>	24 ± 2	31 ± 4	NS
<b>FVC, L</b>	2.15 ± 0.18	2.42 ± 0.25	NS
<b>FEV<sub>1</sub>/FVC%</b>	51 ± 3	54 ± 5	NS
<b>IC, L</b>	1.56 ± 0.15	1.99 ± 0.26	<0.02
<b>V<sub>E</sub>, L/min</b>	8.9 ± 0.5	7.3 ± 0.5	<0.05
<b>f, min<sup>-1</sup></b>	17 ± 1	17 ± 2	NS
<b>PaO<sub>2</sub>, mmHg</b>	63.5 ± 3.0	70.7 ± 4.4	<0.05
<b>PaCO<sub>2</sub>, mmHg</b>	43.0 ± 1.9	43.4 ± 1.8	NS
<b>AaPO<sub>2</sub>, mmHg</b>	36.9 ± 4.1	26.5 ± 3.1	<0.01
<b>Q<sub>T</sub>, L/min</b>	6.5 ± 0.4	5.7 ± 0.5	NS
<b>HR, min<sup>-1</sup></b>	81 ± 4	73 ± 4	<0.05
<b>Ps, mmHg</b>	90 ± 4	96 ± 4	NS
<b>VO<sub>2</sub>, mL/min</b>	246 ± 11	211 ± 12	NS
<b>SHUNT % Q<sub>T</sub></b>	0.5 ± 0.4	0.9 ± 0.5	NS
<b>Low V<sub>A</sub>/Q % Q<sub>T</sub></b>	8.8 ± 3.6	4.5 ± 2.6	NS
<b>Log SD Q</b>	1.24 ± 0.11	0.98 ± 0.09	<0.05
<b>Log SD V<sub>E</sub></b>	0.93 ± 0.05	0.91 ± 0.07	NS
<b>High V<sub>A</sub>/Q % V<sub>E</sub></b>	0.37 ± 0.23	1.63 ± 0.82	NS
<b>Dead Space % V<sub>E</sub></b>	35 ± 3	33 ± 4	NS
<b>DISP R-E*</b>	14.58 ± 1.40	11.72 ± 1.39	NS

<sup>†</sup> Paired t-test analysis; (n= 7 for V<sub>A</sub>/Q variables). Definition of the abbreviations: see Table 2.

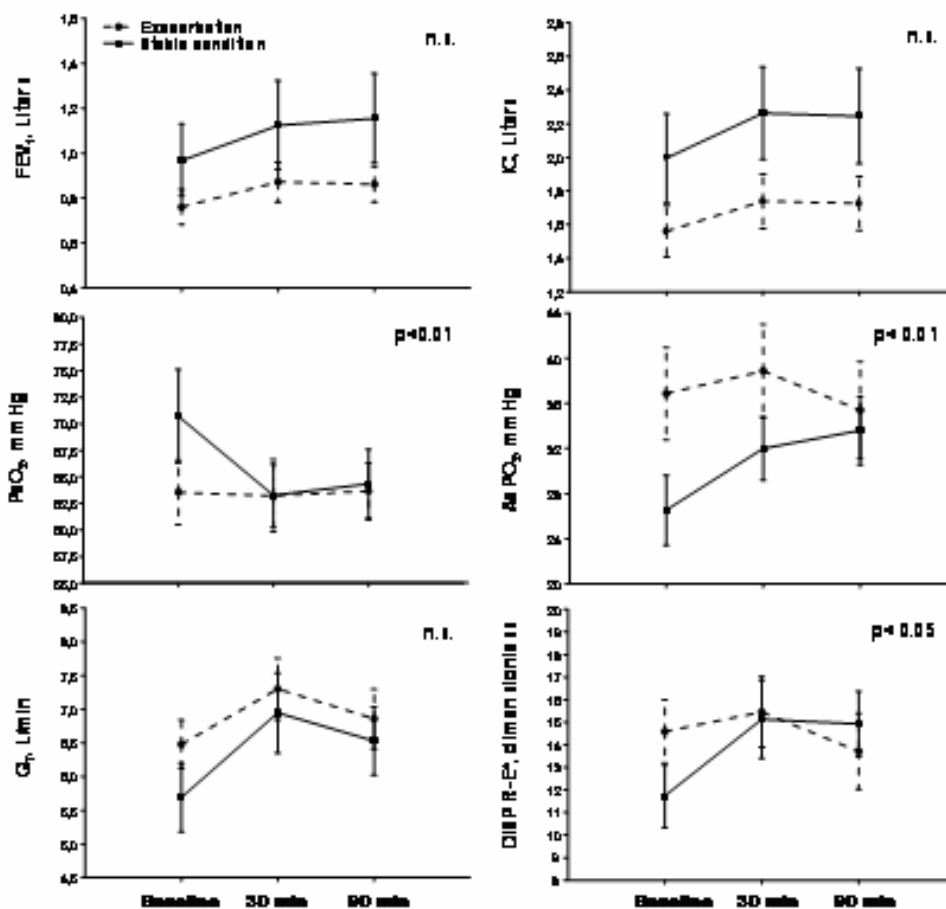
**Table 3. Effects of salbutamol during stable conditions (n= 9).**

	<b>Baseline</b>	<b>30 min</b>	<b>90 min</b>	<b>p<sup>†</sup></b>
<b>FEV<sub>1</sub>, L</b>	0.97 ± 0.16	1.13 ± 0.20**	1.16 ± 0.20**	<0.01
<b>FEV<sub>1</sub>, % pred.</b>	31 ± 4	36 ± 5**	36 ± 9**	<0.01
<b>FVC, L</b>	2.42 ± 0.25	2.57 ± 0.27**	2.62 ± 0.29**	<0.05
<b>FEV<sub>1</sub>/FVC%</b>	40 ± 5	45 ± 5**	44 ± 5**	<0.05
<b>IC, L</b>	1.99 ± 0.26	2.26 ± 0.28**	2.24 ± 0.28**	<0.01
<b>V<sub>E</sub>, L/min</b>	7.3 ± 0.5	7.9 ± 0.5	7.4 ± 0.4	NS
<b>f, min<sup>-1</sup></b>	16 ± 2	15 ± 1	16 ± 1	NS
<b>PaO<sub>2</sub>, mmHg</b>	70.7 ± 4.4	63.2 ± 3.4**	64.3 ± 3.2**	<0.05
<b>PaCO<sub>2</sub>, mmHg</b>	43.4 ± 1.8	43.0 ± 1.6	42.6 ± 1.7	NS
<b>AaPO<sub>2</sub>, mmHg</b>	26.5 ± 3.1	32.0 ± 2.8**	33.6 ± 3.0**	<0.01
<b>Q<sub>T</sub>, L/min</b>	5.7 ± 0.5	7.0 ± 0.6**	6.5 ± 0.5**	<0.01
<b>VO<sub>2</sub>, mL/min</b>	211 ± 12	232 ± 11**	213 ± 14	<0.01
<b>HR, min<sup>-1</sup></b>	73 ± 4	85 ± 4**	82 ± 3**	<0.01
<b>Ps, mmHg</b>	96 ± 4	91 ± 4**	89 ± 4**	<0.05
<b>SHUNT % Q<sub>T</sub></b>	0.84 ± 0.44	1.36 ± 0.50**	1.32 ± 0.54**	<0.05
<b>Low V<sub>A</sub>/Q % Q<sub>T</sub></b>	4.11 ± 2.43	8.06 ± 3.54	9.61 ± 3.92	NS
<b>Log SDQ</b>	0.96 ± 0.09	1.12 ± 0.13**	1.15 ± 0.13**	<0.05
<b>Log SDV</b>	0.91 ± 0.06	1.02 ± 0.07	0.96 ± 0.06	NS
<b>High V<sub>A</sub>/Q % V</b>	1.39 ± 0.73	0.98 ± 0.71	0.89 ± 0.68	NS
<b>Dead Space % V</b>	32.80 ± 4.10	34.21 ± 3.63	35.56 ± 2.89	NS
<b>DISP R-E*</b>	11.35 ± 1.31	14.68 ± 1.48	14.14 ± 1.47	<0.01

† Repeated measures ANOVA; \*\* p<0.05, post-hoc analysis (paired t-test) compared with baseline; (n=8 for V<sub>A</sub>Q data).  
 Definition of the abbreviations: see Table 1.



**Figure 1. Compared effects of salbutamol during exacerbations and under stable conditions.**



p-value, significance of two-ways ANOVA analysis

## Conclusions and future perspectives

The main research work has been focused on the basic pathophysiology of gas exchange of two very common respiratory diseases, asthma and chronic obstructive pulmonary disease.

Particularly, research on bronchial *asthma* has been conducted through a model of bronchial challenge with adenosine 5'monophosphate that is considered today innovative as it can simulate a real asthma attack through the activation of indirect inflammatory and bronchoconstrictive patterns. In particular we wanted to investigate the old hypothesis of dissociation between large and medium sized airways, accounting for the obstructive spirometric abnormalities of asthma, and small airways, where inflammatory and remodelling processes can affect gas exchange efficacy ( $PaO_2$ ,  $AaPO_2$ ,  $V_A/Q$  balance) independently by the bronchomotor tone [1-2]. The existence of "a small airways disease" in asthma is an issue giving rise to an increasing interest of chest physicians. Unfortunately only MIGET studies of Wagner PD and Rodriguez Roisin R could give some scientific support to this hypothesis that remains difficult to detect as large airways tone influences the insight of pathological events affecting the more peripheral part of the lung. In addition, if we exclude few invasive methodologies analysing directly lung tissue (for instance, pulmonary biopsy), there are few possibilities to investigate pathophysiology of peripheral airways. A big contribution has been given by MIGET providing a picture of ventilation and distributions in functional lung units without interfering with broncho-motor and vascular tones. Even being a

difficult and not practical technique it gives, with the necessary experience and practice, a fundamental contribution to clinical research. Today we consider of great interest the investigation on small airways through the integration of MIGET with old and newer other techniques in order to obtain a better control and follow-up of the disease and to verify new therapeutical approaches. On this base a new research protocol on mild-to-moderate asthma has been proposed in order to study patients with intermittent symptoms, treated with combined inhaled therapy ( $\beta_2$ -agonists - corticosteroids), by using different techniques able to provide some functional and morphological information on pathological processes of small airways. In particular we plan to study:

1. *Flow-volume curves*, with particular attention at medium and low volumes (MEF and FEF<sub>25-50-75</sub>];
2. *Closing volume*; an increased value of closing volume or closing capacity expresses the precocious collapse of lower parts of the lungs and seems to be a good marker of small airways obstruction even in presence of a normal FEV<sub>1</sub>;
3. *Bronchial and alveolar production of NO*; exhaled NO is considered a good marker of airways inflammatory disorders and a recent mathematical model is able to discriminate the bronchial and alveolar production by applying different resistances at mouth.
4. *Respiratory gases and AaPO<sub>2</sub>*, through the direct measurement of respiratory rate (VCO<sub>2</sub>/VO<sub>2</sub>);
5. *MIGET*;

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Conclusions and future perspectives

6. *Pulmonary ventilation scan*. It analyses pulmonary ventilation distribution through the inhalation of TC<sub>99</sub>-labelled micro-particles; the recent technique, developed in the Cardio-Thoracic Department of Pisa, avoids adequately spotting images due to large airways deposition of the particles and allows a good definition of more peripheral ventilated areas. The aim is to evaluate the presence of  $V_A/Q$  and gas exchange abnormalities, which remain commonly unidentified, beside mild spirometric abnormalities and to correlate them with the other parameters considered, hoping to find a new definition of small airways diseases in asthma through the combination of new and old techniques and new parameters specific for small airways able to open to new follow-up and therapeutical approaches.

The other respiratory disease object of the present investigation with the contribution of MIGET is that heterogeneous group of clinical, functional and biological conditions called "*chronic obstructive pulmonary diseases*", including, basically, chronic bronchitis, emphysema and, according to some authors, chronic asthma when flow obstruction reversibility is lost.

Particularly, this clinical research experience has underlined the complexity of the interaction among intra pulmonary and extra pulmonary factors determining pulmonary gas exchange in COPD. Few methodologies can probably provide so complete information about pathophysiological mechanisms of a respiratory disease as MIGET. In addition, the model of study adopted in this work has two major advantages:

- it provides a better knowledge of  $\beta_2$ -agonists that are not only bronchodilators but also vasoactive and inotropic agents profoundly influencing, therefore, the general hemodynamic balance;

- it underlines the central role of pulmonary circulation and vascular tone regulation in pathophysiology of COPD, whereas the “vascular side” of respiratory diseases is often forgotten and airways abnormalities are uniquely considered.

On the base of this research experience on COPD a new research protocol is actually been considering, aimed at the investigation of COPD heterogeneity that remains, despite the enormous progresses made in this field, an unsettled issue.

Despite a universal definition of COPD, including all diseases characterized by poorly reversible airflow limitation [3], huge clinical, functional and radiological differences are described among individuals with COPD label and, more important, prognosis is diverse [4] and response to therapy may be different. COPD includes different specific phenotypes, chronic bronchitis (CB) pulmonary emphysema (PE) and, according to some authors, chronic asthma, when airways obstruction reversibility has disappeared. Major differences exist between asthma and COPD (CB/PE): clinical history, bronchial responsiveness, DLCO, biological markers and structural alterations of the lung usually permit to differentiate asthma from CB/PE, even for a similar degree of airway obstruction. A recent work of Fabbri et al. [5] highlights the importance of comparing asthma and chronic bronchitis patients of the same age class and with a comparable degree of airflow

obstruction. By using imaging (HRTC), functional (including salbutamol reversibility and methacholine responsiveness) and pathological techniques (exhaled NO, induced sputum, BAL, bronchial biopsy) the authors defined different phenotypes: patients with a history of asthma had more eosinophils in blood, sputum, BAL and airway mucosa, and a higher CD4/CD8 ratio of T cells infiltrating the airway mucosa. As expected, they also had less residual volume, higher DLCO, exhaled NO and reversibility to bronchodilator and steroids, and a lower HRTC-emphysema-score.

It is clear that a large variability exists also within the big group of patients with chronic bronchitis with or without emphysema, in symptoms (perception of dyspnoea, presence of cough and phlegm, etc.), time course, radiological aspects and systemic effects of the disease. Important functional differences are also observed in DLCO, residual volume and hyperinflation.

In addition, beyond the old definition of COPD subtypes of “blue brothers” and “pink puffer” that correspond to important pathological and functional differences, today it has become common to differentiate patients by systemic effects of COPD, on the basis of exercise capacity, body mass index and the involvement of peripheral muscles.

At the origin of such variability is probably the existence of different pathogenetic mechanisms, different factor risks and predisposing factors that are not easy to unravel. The literature on MIGET in COPD has put into evidence the existence of different patterns of  $V_A/Q$  distributions and has showed the intent to find some correlation between functional data and structure even with some difficulties but the combination of MIGET with more

modern radiological and biological techniques could give us much more information [6-8]

The coexistence of chronic inflammation and oxidative stress alterations at level of small airways, of obstructive bronchiolitis and of parenchymal destruction in a variable proportion, makes COPD pathology extremely heterogeneous [9-11]. On this line, a new project of investigation has been planned on COPD heterogeneity (functional, biological and radiological features) that integrates the information deriving from HRCT and functional tests with ventilation - perfusion study (MIGET), biological measurements and markers of systemic involvement.

The model of  $V_A/Q$  study before and after the administration of inhaled salbutamol, a bronchodilator and vasoactive agent, could help to investigate the interplay of intra-pulmonary and extra-pulmonary factors modulating gas exchange in different pathogenetic conditions.

The information deriving from the analysis of cell composition, inflammatory and oxidative stress markers in sputum and BAL (IL-8, TNF- $\alpha$ , NE, VEGF, MMP, NCA) and exhaled breath condensate (EBC) (8-isoprostane, MDA, LTB<sub>4</sub>, cys-LT) and from exhaled NO (bronchial and alveolar fractions) is fundamental to investigate the chronic inflammatory process causing airway remodelling, narrowing of small airways and loss of lung elastic recoil, and could have important therapeutical implications.

Three classes of COPD patients could be selected, similar for age (55 - 65 years) and airway obstruction ( $FEV_1$ : 50 - 70% of predicted), by clinical history and radiological aspects: chronic asthma, chronic bronchitis with and

without emphysema. Possible targets of investigation are: the effects of inhaled salbutamol on gas exchange and ventilation/perfusion relationships; differences among the three classes of patients by the analysis of sputum, BAL, exhaled air and blood samples. The correlation among basal measurements of pulmonary function, gas exchange, inflammatory and oxidative stress markers can also be evaluated. The peripheral involvement could be evaluated through 6 min walking test and the evaluation of free fat mass (bio-electric impedance).



## REFERENCES

1. Wagner PD, Hedenstierna G, Rodriguez-Roisin R. Gas exchange, expiratory flow obstruction and the clinical spectrum of asthma. *Eur Respir J.* 1996 Jun;9(6):1278-82.
2. Rodriguez-Roisin R. Acute severe asthma: pathophysiology and pathobiology of gas exchange abnormalities. *Eur Respir J.* 1997 Jun;10(6):1359-71.
3. Pauwels RA, Buist AS, Calverley PM, Jenkins CR, Hurd SS; GOLD Scientific Committee. Global strategy for the diagnosis, management, and prevention of chronic obstructive pulmonary disease. NHLBI/WHO Global Initiative for Chronic Obstructive Lung Disease (GOLD) Workshop summary. *Am J Respir Crit Care Med.* 2001 Apr;163(5):1256-76.
4. Burrows B, Bloom JW, Traver GA, Cline MG. The course and prognosis of different of chronic airways obstruction in a sample from general population. *N Engl J Med* 1987;317:1309-1314.
5. Fabbri LM, Romagnoli M, Corbetta L et al. Differences in airway inflammation in patients with fixed airflow obstruction due to asthma or chronic obstructive pulmonary disease. *Am J Respir Crit Care Med.* 2003;167:418-24.
6. Wagner PD, Dantzker DR, Dueck R, Clausen JL, West JB. Ventilation-perfusion inequality in chronic obstructive pulmonary disease. *J Clin Invest.* 1977 Feb;59(2):203-16.
7. Agusti AG, Barbera JA, Roca J, Wagner PD, Guitart R, Rodriguez-Roisin R. Hypoxic pulmonary vasoconstriction and gas exchange during exercise in chronic obstructive pulmonary disease. *Chest.* 1990 Feb;97(2):268-75.

8. Barbera JA, Ramirez J, Roca J, Wagner PD, Sanchez-Lloret J, Rodriguez-Roisin R. Lung structure and gas exchange in mild chronic obstructive pulmonary disease. *Am Rev Respir Dis.* 1990 Apr;141(4 Pt 1):895-901.
9. Boschetto P, Miniati M, Miotto D et al. Predominant emphysema phenotype in chronic obstructive pulmonary disease patients. *Eur Respir J* 2003;21:450-54.
10. Turato G, Zuin R, Miniati M, et al. Airway inflammation in severe chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2002;166:105-10.
11. Saetta M, Turato G, Maestrelli P et al. Cellular and structural bases of chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2001;163:1304-09.