



Article

Sonication Effects on Atrazine Dissipation in Vadose Zone Sediment Slurries

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Abstract: Herbicide atrazine easily leaches to groundwater, where it is persistent. We studied whether sonication accelerates atrazine dissipation ($100 \text{ mg}\cdot\text{L}^{-1}$) in vadose zone sediment slurries. Sediments were from 11.3 to 14.6 m depths in an atrazine-contaminated groundwater area. The slurries and autoclave-sterilized slurries were sonicated (bath, 43 kHz, 320 W) for 0, 5, 10, 20, or 30 min once/twice a day, and atrazine concentrations were followed. Atrazine concentrations raised in the sterilized slurries sonicated twice a day for 10 min ($86.0 \pm 7.7 \text{ mg}\cdot\text{L}^{-1}$), while they remained low in the slurries ($56.6 \pm 10.9 \text{ mg}\cdot\text{L}^{-1}$) due to microbial degradation. Twice a day sonications for 20–30 min did not enhance microbial atrazine degradation. Chemical dissipation may have occurred in the sterilized slurries sonicated twice a day for 30 min. However, sonication did not decrease atrazine concentrations below those in the non-sonicated slurries ($55.1 \pm 7.8 \text{ mg}\cdot\text{L}^{-1}$) and sterilized slurries ($67.1 \pm 7.9 \text{ mg}\cdot\text{L}^{-1}$). Atrazine concentrations in the sterilized slurries were higher than in the slurries, indicating changes in sediment structure and adsorption due to autoclaving. Sonication parameters needed for releasing atrazine from interactions with particles may be close to those damaging microbial cells. This suggests difficulties in enhancing microbial atrazine degradation by sonication, though chemical degradation can be enhanced.

Keywords: atrazine; pesticide; sonication; autoclaving; dissipation; degradation; subsurface sediment slurry

1. Introduction

The herbicide atrazine (6-chloro-N-ethyl-N'-(1-methylethyl)-1,3,5-triazine-2,4-diamine) is used to control the growth of broad-leaved and grassy weeds. It and its degradation products are commonly found in the soil and surface water, and they can elute through the subsurface sediments to groundwater [1,2]. Atrazine concentrations can be particularly high in point source pollutions, e.g., in the vicinity of dealerships [3–5]. Atrazine has been reported to be an endocrine disrupting compound [6,7]. To protect the drinking water quality, the European Commission prohibited the sale of atrazine in 1992 [8].

Atrazine dissipation has been studied extensively due to its wide use, occurrence in environment as a contaminant, and possible adverse effects. Methods for the removal of atrazine from soil, subsurface sediments, and groundwater are needed. Microbes and microbial consortia can degrade and even mineralize atrazine [3,9], though the degradation rate is slow in subsurface sediments [5,10,11].

The chemical decomposition of atrazine by advanced oxidation methods like sonication has also been studied [12–18]. Mild sonication homogenizes suspensions, enhances extraction, and increases microbial productivity, while strong sonication conditions disrupt cells and cause the chemical degradation of xenobiotics [16,19–23]. Sonication separately and in combination with other advanced oxidation processes has induced chemical atrazine degradation in an aqueous solution [13–15,18].

Ultrasound generates hydroxyl radicals and other radical species at the extreme temperature and pressure conditions that occur when the cavities formed by sonication implode in water. The direct pyrolysis in the sonication cavities is regarded as negligible due to the low volatility of atrazine [14]. Atrazine has also been chemically degraded by ultrasound in a slurry, where the cavitation bubbles are preferentially formed at the interface of solid and liquid [16,17]. When the bubble collapses, a high-speed jet is created towards the surface of the particle. As a result, the adsorbed pesticides are exposed to high temperatures and may undergo pyrolysis. It is not known, however, whether microbial atrazine degradation can be enhanced by sonication. For instance, the microbial phenantrene degradation rate was increased fourfold by intermittent sonication in a solid-liquid system [24]. The enhanced degradation was attributed to the improved mass transport of phenantrene between the solid and liquid phase, which increased phenantrene bioavailability to microorganisms. However, the behavior of different contaminants in matrix-contaminant interactions differ [19].

To clarify the relationships between the sonication, and the microbial and chemical atrazine dissipation in subsurface sediments, we studied sonication effects on atrazine dissipation in the subsurface sediment slurries. The hypotheses of the study were that microbial (slurries) and chemical (sterilized slurries) atrazine dissipation can be enhanced by sonication (43 kHz, 320 W), while the null hypothesis was that dissipation cannot be improved. The high initial atrazine concentration of $100 \text{ mg}\cdot\text{L}^{-1}$ was chosen, to ensure that bioavailability does not limit microbial degradation, and to reveal atrazine behavior at concentrations above the water solubility limit of $33 \text{ mg}\cdot\text{L}^{-1}$ in slurry suspensions homogenized by sonication. High concentrations represent an example of point source pollution.

2. Materials and Methods

2.1. Sediments and Chemical Analyses

Sediments were collected in drillings next to the Lahti railway station (Finland), which is located in a pesticide-contaminated groundwater area. The drill diameter was 75 mm down to about 10 m, and below that 48 mm. Sediments from the depths of 11.3–14.6 m were collected in sealed plastic bags at the drilling site, where groundwater table was at about 15 m [25]. Sediment size fractions were separated by sieving for 30 min at the amplitude 50 (AS 200 basic, Retsch GmbH, Haan, Germany).

To determine their dry weight, 3–5 g of sediments in triplicate were dried for 16 h at $105 \text{ }^\circ\text{C}$, followed by heating at $550 \text{ }^\circ\text{C}$ for 4 h to measure the organic matter (SFS-EN 13039, SFS-EN 13040). Sediment total carbon (total-C) was analyzed in duplicate by LECO Model 2000 CNS analyzer (LECO Corporation, St. Joseph, MI, USA) according to its instructions. Sediment ammonium (the limit of detection, LOD, $0.039 \text{ }\mu\text{g}\cdot\text{g}^{-1}$), nitrate (LOD, $0.1 \text{ }\mu\text{g}\cdot\text{g}^{-1}$), nitrite (LOD, $0.1 \text{ }\mu\text{g}\cdot\text{g}^{-1}$), and elements (Cd, LOD $0.2 \text{ }\mu\text{g}\cdot\text{g}^{-1}$; Co; Cr; Cu; Pb; Mn; Ni; Fe; Zn) were determined as a purchased service from Ramboll Analytics Ltd. as presented in Talja et al. [10]. Microorganisms were cultivated from sediment slurries.

2.2. Sonication Experiments

Sonication experiments were done in 100 mL flasks with hole (diameter 5 mm) caps, which were covered with aluminum foil. The flasks contained 15.0 g (dry weight) of sediment, 50 mL of sterile distilled water, and $100 \text{ mg}\cdot\text{L}^{-1}$ of atrazine (0.5 mL from $10 \text{ g}\cdot\text{L}^{-1}$ stock in methanol; Dr. Ehrenstorfer GmbH, Augsburg, Germany). The sediment slurries were sterilized by autoclaving (Instru, Santasalo-Sohlberg, Helsinki, Finland) on three successive days for 1 h at $121 \text{ }^\circ\text{C}$ and 101 kPa pressure. The flasks were incubated at the room temperature of $21 \pm 2 \text{ }^\circ\text{C}$ in a shaker (120 rpm, Laboshake, Gerhardt, Konigswinter, Germany), all in triplicate.

In the 0, 5, and 10 min sonication experiment, the slurries amended with $100 \text{ mg}\cdot\text{L}^{-1}$ of atrazine were allowed to stabilize on the shaker (120 rpm) for five days, and then the effects of 0, 5, and 10 min sonications on atrazine concentrations were studied using Branson 8510 bath sonicator (43 kHz, RF-power 320 W; W.A. Brown Industrial Sales Inc., Richmond, VA, USA). The slurry and sterilized

slurry flasks were placed in a sonicator bath filled with water to the mark. The initial bath water temperature was at room temperature, and it increased approximately 1.3 °C/10 min sonication. The flasks were sonicated once a day for 0, 5, or 10 min on days 5, 6, 7, 8, 12, 13, 14, and 15, followed by a stabilization period. As the effects of the once a day sonications were minor, the sonications were continued twice a day for 0, 5, and 10 min on days 25, 26, 27, 28, 29, 32, 33, 34, 35, and 36. The 100 µL samples were taken on days 0, 1, and before the daily sonications. Prior to the samplings, the flasks were shaken to evenly distribute atrazine, and then large particles were allowed to settle for at least 15 min, while the suspended colloids remained in the water phase.

In the 0, 20, and 30 min sonication experiment, the slurries amended with 100 mg·L⁻¹ of atrazine were allowed to equilibrate on the shaker (120 rpm) for six days. Then the impacts of twice a day sonications for 0, 20, and 30 min on atrazine concentrations were studied in the slurries and sterilized slurries, as presented above. The flasks were sonicated twice a day for 0, 20, or 30 min on days 6, 7, 10, 11, 12, 13, and 14. The 100 µL samples were taken at the beginning of the experiment (day 0) and before the sonications on days 6, 10, 13, and 17. Samples were taken as in the 0, 5, and 10 min sonication experiment.

2.3. Pesticide Adsorption and Microbial Growth in the Sediment Slurries

To determine the quantity of atrazine remaining adsorbed in the sediments and microbial numbers in the slurries with and without atrazine addition, the following experiment was done. Sediments (15.0 g) and 50 mL of sterile distilled water were incubated with and without 100 mg·L⁻¹ of atrazine as presented above, all in duplicate. The slurries were stabilized for 10 days on a shaker as presented above (final atrazine concentration 61.9 ± 8.9 mg·L⁻¹ in atrazine-amended slurries) to allow adsorption to occur and reach the equilibrium stage [26]. The liquid phase was removed, ten-fold serial dilution was performed, and 100 µL was cultivated on mineral agar plates having 33 mg·L⁻¹ of atrazine as a nitrogen source, as presented in Liu et al. [27] and Pukkila et al. [28]. The colonies were counted after 5 days incubation at the temperature of 21 ± 2 °C. The sediment was extracted as presented below.

2.4. Pesticide Analyses

The 100 µL sample was supplemented with 400 µL of methanol:water (3:1, *v/v*), and 100 µL of the internal standard simazine in methanol: water (3:1, *v/v*) to gain the final concentration of 11.9–15.3 mg·L⁻¹. The simazine background level in the sediments was low, 11.4 µg·L⁻¹ (<0.1% of total). The 600 µL standards in methanol:water (3:1, *v/v*) contained 0.5, 2.5, 5, 10, 20 and 35 mg·L⁻¹ of atrazine (external standards), and the same quantity of simazine as in samples (internal standards). The responses were linear with the correlation coefficients of 0.997 ± 0.007 (R² ± S.D.). The quantification limit was about 0.56 mg·L⁻¹ for atrazine. The standards and samples were filtered through 0.45 µm GHP polypropylene membrane filters (Acrodisc®, Gelman, Pall Corporation Ltd., New York, NY, USA), and 20 µL was analyzed by high performance liquid chromatography (HPLC) equipped with Shimadzu Prominence SIL-20A auto sampler, LC-20AT solvent delivery module, DGU-20A5 on-line degasser, SPD-20A UV/VIS detector (225 nm; Shimadzu, Kyoto, Japan), SunFire column (C18, 3.5 µm, 3.0 mm × 150 mm, Waters, Milford, MA, USA) and Shimadzu LC Solution software control system (Shimadzu, Kyoto, Japan). The chromatographic separation was carried out using the following gradient profile: the initial acetonitrile concentration was held at 30% for 3.5 min, and then it was increased to 65%, which was held until 8.5 min. Finally, the acetonitrile concentration was decreased back to 30%, which was held until 14 min. The mobile phase flow rate was 0.4 mL·min⁻¹.

To extract atrazine adsorbed to 15 g of sediments, 20 µg of propazine (Dr. Ehrenstorfer GmbH, Augsburg, Germany) was added as an internal standard. The sediment was extracted three times with 18.0 mL of methanol:water (3:1, *v/v*) by shaking overnight at 200 rpm (Unimax 1010, Heidolph Instruments, Schwabach, Germany). Before the extractions, the sample was sonicated for 15 min at 20 °C (40 kHz, Everest, Istanbul, Turkey). The extracts were separated by centrifugation for

10 min at $1250\times g$ (Multifuge 15-R Heraeus, Kendro Laboratory Products, Osterode, Germany), and stored at $-20\text{ }^{\circ}\text{C}$. The combined extracts were evaporated to dryness, and the residue was extracted twice with $300\text{ }\mu\text{L}$ of acetone. The extraction solution was sonicated as above, followed by centrifugation ($13,000\times g$). The pooled acetone extracts were filtered, and pesticides were analyzed using Shimadzu GCMS-QP-2010Ultra GC-MS (Shimadzu, Kyoto, Japan), autosampler AOC-20i+s, ZB-5MS capillary column (29 m, 0.25 mm, 0.25 μm), carrier gas helium (1.29 mL/min), injector temperature $250\text{ }^{\circ}\text{C}$, and $2\text{ }\mu\text{L}$ splitless injection. The oven temperature was held at $120\text{ }^{\circ}\text{C}$ for 2 min, increased $20\text{ }^{\circ}\text{C}/\text{min}$ to $180\text{ }^{\circ}\text{C}$, held for 5 min, and then increased $20\text{ }^{\circ}\text{C}/\text{min}$ to $280\text{ }^{\circ}\text{C}$, held for 8 min. The mass spectra were recorded at an electron energy of 70 eV. The ion source temperature was $230\text{ }^{\circ}\text{C}$, and interface temperature was $250\text{ }^{\circ}\text{C}$. Five standards in acetone contained $0.133\text{--}3.435\text{ nmol}\cdot\text{L}^{-1}$ of atrazine, desethylatrazine (DEA), deisopropylatrazine (DIA), and desethyldeisopropylatrazine (DEDIA). The limit of quantification (LOQ) was about $0.1\text{ }\mu\text{g}/\text{sample}$. The ions followed were m/z 202 and 215 (quantification) for atrazine; 174 (quantification) and 187 for DEA, m/z 158 (quantification), 173 and 175 for DIA, m/z 110, 145 (quantification) and 147 for DEDIA, and m/z 172, 187, 214, and 229 (quantification) for propazine. The extraction efficiency of atrazine is $102.1\% \pm 14.0\%$. The quantity of atrazine, simazine and 2,6-dichlorobenzamide (BAM) in the sediments was determined as presented in Mattsson et al. [25].

2.5. Calculations

Results are presented on a dry weight basis. Atrazine concentrations are presented as mean \pm S.D. ($n \geq 3$). Results were analyzed using two-factor (sonication and autoclaving) repeated measures analysis of variance (RMA), followed by pairwise comparisons (PC) (IBM SPSS Statistics, version 23, IBM Corporation, New York, NY, USA). In the 0, 5, and 10 min sonications, the RMA was performed separately on days 5 to 15 (sonications once a day), and 25 to 36 (sonications twice a day). In the 0, 20, and 30 min sonications, the RMA was performed between days 6 to 17 (Table 1). The highest atrazine concentrations reached by sonications were compared using Anova, followed by Tukey honest significant difference (HSD) test. Mann-Whitney's test (M-W) was used for pairwise comparisons.

Table 1. The two factor (sonication, sterilization) repeated measures analysis of variance (RMA) of atrazine concentrations in the slurries and sterilized slurries, both sonicated for 0, 5, 10, 20, or 30 min ($n = 3$). Sonications were done once a day on days 5–15, and twice a day on days 25–36 and 6–17. The p -values of between subject effects are presented.

Sonication Time (min)	0, 5 and 10		0, 20 and 30
Time in experiment (day)	5–15	25–36	6–17
Slurries and sterilized slurries			
Sterilization	<0.001	<0.001	<0.001
Sonication	0.421	0.121	0.002
Sterilization * Sonication	0.433	0.007	0.050
Sterilized slurries			
Sonication	0.270	0.002	0.416
Slurries			
Sonication	0.554	0.296	0.001

* Interaction between sterilization and sonication.

3. Results

3.1. Sediments

Sediment cores were collected by drilling in a groundwater area contaminated with simazine, atrazine, and BAM in concentrations of 38, 13, and $5\text{ }\mu\text{g}\cdot\text{kg}^{-1}$ (dry weight), which are equivalent to 0.57, 0.195, and $0.075\text{ }\mu\text{g}$ in 15 g of sediments, respectively. The sediment size fractions were:

<45 μm , 0.9%; 45–100 μm , 2.6%; 100–125 μm , 3.4%; 125–250 μm , 33.7%; 250–500 μm , 38.6%; 0.5–1 mm, 16.7%; 1–2 mm, 3.5%; 2–4 mm, 0.2%; 4–8 mm, 0.2%; and >8 mm (diameter), 0.2%, i.e., the sediment was sandy. The fractions ≥ 4 mm were removed from the sonication experiments. Sediment organic matter ($4.3 \pm 0.2 \text{ mg}\cdot\text{g}^{-1}$) and total-C ($0.31 \text{ mg}\cdot\text{g}^{-1}$) concentrations were low, and ammonium, nitrate, nitrite, and Cd were below the detection limits. The sediments contained $3.4 \pm 1.0 \text{ }\mu\text{g}\cdot\text{g}^{-1}$ of cobalt, $12 \pm 4 \text{ }\mu\text{g}\cdot\text{g}^{-1}$ of chromium, $15 \pm 5 \text{ }\mu\text{g}\cdot\text{g}^{-1}$ of copper, $2.8 \pm 1.0 \text{ }\mu\text{g}\cdot\text{g}^{-1}$ of lead, $96 \pm 24 \text{ }\mu\text{g}\cdot\text{g}^{-1}$ of manganese, $6.1 \pm 2.0 \text{ }\mu\text{g}\cdot\text{g}^{-1}$ of nickel, $12 \pm 3 \text{ mg}\cdot\text{g}^{-1}$ of iron, and $27 \pm 9 \text{ }\mu\text{g}\cdot\text{g}^{-1}$ of zinc.

Sonication effects on atrazine dissipation were studied in the slurries and sterilized slurries, to separate microbial and chemical dissipation. Sonication times were once (days 5–15) and twice a day (days 25–36) for 0, 5 and 10 min in the first experiment, and twice a day (days 6–17) for 0, 20 and 30 min in the second experiment.

3.2. The 0, 5 and 10 min Sonication Experiment

Atrazine concentrations fell during the initial stabilization period from $100 \text{ mg}\cdot\text{L}^{-1}$ to $64.1 \pm 8.9 \text{ mg}\cdot\text{L}^{-1}$ in the slurries, and only to $79.5 \pm 12.3 \text{ mg}\cdot\text{L}^{-1}$ in the sterilized slurries, the difference of $15.4 \text{ mg}\cdot\text{L}^{-1}$ being statistically significant (M-W, $p = 0.007$; Figure 1). Atrazine concentrations in the slurries continued to be lower than in the sterilized slurries throughout the experiment (RMA, $p < 0.001$; Table 1), the difference between the non-sonicated slurries and the sterilized slurries being on the average $9.7 \pm 3.2 \text{ mg}\cdot\text{L}^{-1}$ on days 5–36.

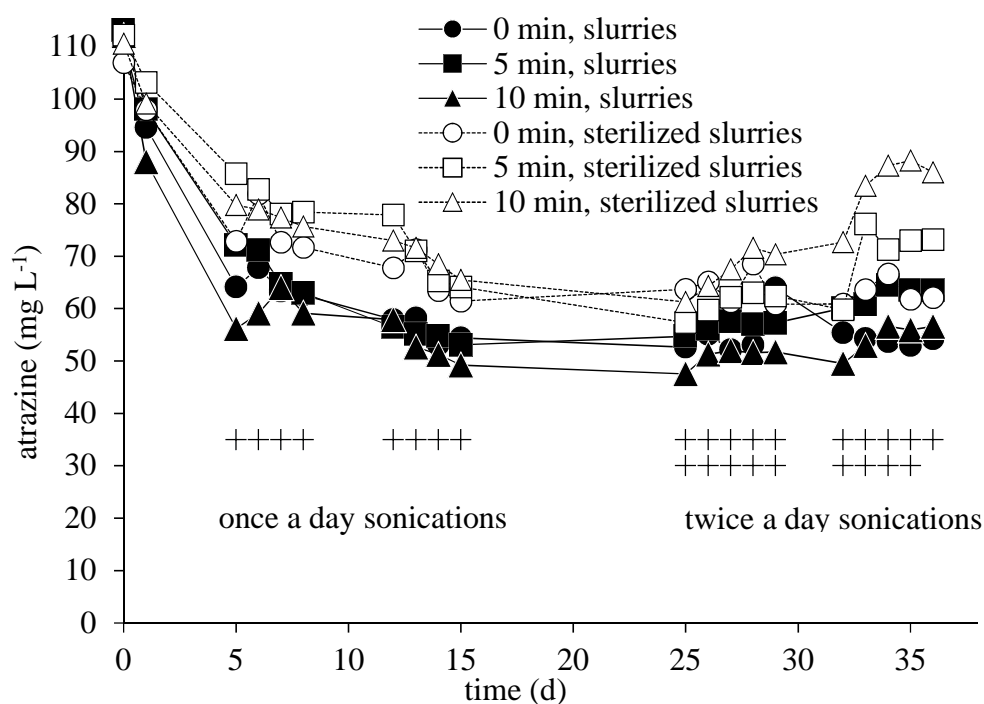


Figure 1. Atrazine concentrations ($n = 3$) in the slurries and sterilized slurries, all sonicated for 0, 5, or 10 min. One or two sonication times per day are marked with +. The average of standard deviations was $4.9 \pm 3.0 \text{ mg}\cdot\text{L}^{-1}$.

Once a day sonications for 5 and 10 min were not strong enough to affect atrazine concentrations in the slurries and sterilized slurries, compared to the non-sonicated slurries and sterilized slurries, as can be seen from two-factor RMA results on days 5–15 ($p = 0.421$ for sonication; Table 1, Figure 1). When the 5 and 10 min sonications were repeated twice a day between days 25–36, atrazine concentrations in the sterilized slurries sonicated for 10 min were higher than in those sonicated for 5 min, or in those without sonication (RMA, $p = 0.002$ for sonication; PC, $p \leq 0.002$; Figure 1, Table 1). Twice a

day sonications for 5 min were not strong enough to affect atrazine concentrations in the sterilized slurries, but sonications twice a day for 10 min increased atrazine concentrations to the highest measured values of $86.0 \pm 7.7 \text{ mg}\cdot\text{L}^{-1}$. In contrast, in the slurries twice a day sonications for 5 and 10 min did not affect significantly atrazine concentrations (RMA, $p = 0.296$ for sonication), which were $56.6 \pm 10.9 \text{ mg}\cdot\text{L}^{-1}$ on days 25–36. Indeed, an interaction between the sterilization and sonication was obtained in two-factor RMA on days 25–36 ($p = 0.007$), that is sonication effects on atrazine concentrations differed between the sterilized slurries and slurries. Twice a day sonications for 10 min increased atrazine concentrations in the sterilized slurries, but not in the slurries apparently due to microbial atrazine degradation.

3.3. The 0, 20 and 30 min Sonication Experiment

After the initial stabilization period, atrazine concentrations in the slurries ($53.8 \pm 5.4 \text{ mg}\cdot\text{L}^{-1}$) were again lower than in the sterilized slurries ($76.0 \pm 6.4 \text{ mg}\cdot\text{L}^{-1}$) (M-W, $p < 0.001$, Figure 2). The influence of sterilization continued to be significant also on days 6–17 (RMA, $p < 0.001$ for sterilization; Table 1, Figure 2). Atrazine concentrations in the non-sonicated slurries remained on average $23.5 \pm 1.4 \text{ mg}\cdot\text{L}^{-1}$ lower than in the non-sonicated sterilized slurries throughout the experiment on days 6–17. Decreased atrazine concentrations were also measured in the sonicated slurries, compared to the sonicated sterilized slurries.

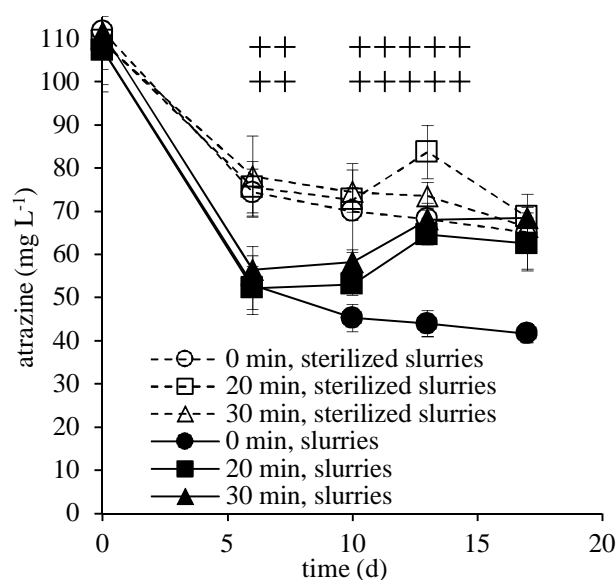


Figure 2. Atrazine concentrations ($n = 3$) in the slurries and sterilized slurries, all sonicated for 0, 20, or 30 min. Two sonications per day are marked with +.

Twice a day sonications for 20 and 30 min did not affect atrazine concentrations of $75.1 \pm 7.9 \text{ mg}\cdot\text{L}^{-1}$ in the sterilized slurries on days 6–17 (RMA, $p = 0.416$; Figure 2, Table 1), as the concentrations were still high immediately after the stabilization period. In contrast, twice a day sonications increased atrazine concentrations from $44.0 \pm 3.0 \text{ mg}\cdot\text{L}^{-1}$ of the non-sonicated slurries to $66.3 \pm 3.0 \text{ mg}\cdot\text{L}^{-1}$ of the slurries sonicated for 20 and 30 min, the difference of $22.3 \text{ mg}\cdot\text{L}^{-1}$ being statistically significant (RMA, $p = 0.001$; PC, $p \leq 0.002$). Thus, an interaction between the sterilization and sonication was obtained in two-factor RMA on days 6–17 ($p = 0.050$), which indicates that 20 and 30 min sonication effects on atrazine concentrations differed between the slurries and sterilized slurries. Twice a day sonications for 20 and 30 min did not affect atrazine concentrations in the sterilized slurries, but the sonications increased concentrations in the slurries. Microbes in the slurries did not have time to degrade more atrazine during the stabilization period, and thereafter the longest sonication times of 20 and 30 min may have disturbed the microbial degradation.

3.4. Summary of the Sonication Effects on Atrazine Concentrations

The highest atrazine concentrations, achieved in the slurries and sterilized slurries by sonication, were plotted as a function of twice a day sonication times of 5, 10, 20, and 30 min (Figure 3). Atrazine concentrations in the non-sonicated sterilized slurries were $65.8 \pm 4.5 \text{ mg}\cdot\text{L}^{-1}$. When the sterilized slurries were sonicated twice a day for 10–20 min, atrazine concentrations increased significantly by $19.6 \text{ mg}\cdot\text{L}^{-1}$ to $85.4 \pm 6.4 \text{ mg}\cdot\text{L}^{-1}$ (Anova and Tukey HSD, $p \leq 0.002$) (Figure 3). When the sonication time was further increased to twice a day for 30 min, atrazine concentrations in the sterilized slurries increased only by $7.6 \text{ mg}\cdot\text{L}^{-1}$ to $73.4 \pm 3.2 \text{ mg}\cdot\text{L}^{-1}$ compared to the non-sonicated sterilized slurries, which is not a statistically significant difference. Atrazine concentrations in the sterilized slurries sonicated twice a day for 10 min were higher than those in the sterilized slurries sonicated twice a day for 30 min (Tukey HSD, $p = 0.034$). In the latter, chemical atrazine dissipation may have occurred.

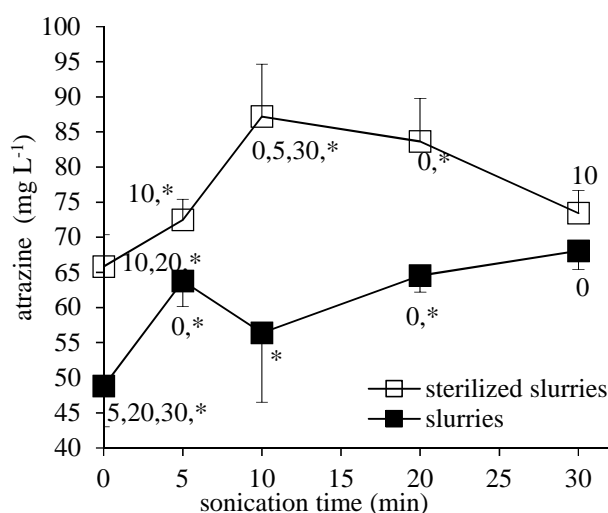


Figure 3. The highest atrazine concentrations in the slurries and sterilized slurries sonicated twice a day for 0 (days 34–36, $n = 9$, day 6, $n = 3$), 5 (days 34–36, $n = 9$), 10 (days 34–36, $n = 9$), 20 (day 13, $n = 3$), and 30 (day 13, $n = 3$) min. Numbers indicate statistically significant differences (Anova, Tukey HSD, $p < 0.050$) between sonication times, and asterisks indicate statistically significant differences (T-test, $p < 0.050$) between slurries and sterilized slurries.

Atrazine concentrations in the non-sonicated slurries were $48.8 \pm 5.8 \text{ mg}\cdot\text{L}^{-1}$ (Figure 3). The concentrations increased to $63.2 \pm 6.5 \text{ mg}\cdot\text{L}^{-1}$ due to the twice a day sonications for 5–30 min; on average, $14.4 \text{ mg}\cdot\text{L}^{-1}$ of atrazine was released by sonication compared to the non-sonicated slurries. Atrazine concentrations in the sonicated slurries were significantly higher than in the non-sonicated slurries (Anova, $p = 0.002$; Tukey HSD, $p \leq 0.017$), with the exception of 10 min sonications (Tukey HSD, $p = 0.360$). Atrazine concentrations increased least in the slurries sonicated twice a day for 10 min, while the atrazine quantity bound to the sediments in the non-sonicated slurries was only $4.6 \pm 3.9 \text{ mg}\cdot\text{kg}^{-1}$ ($1.4 \pm 1.2 \text{ mg}\cdot\text{L}^{-1}$). The DEA quantity in the sediment extracts of the non-sonicated slurries was below the LOQ, and DIA and DEDIA residues were not detected. In the experiment done to determine the pesticide adsorption and microbial growth, the microbial numbers increased from $7 \times 10^4 \text{ cfu}\cdot\text{mL}^{-1}$ of the non-atrazine amended slurries to $>10^8 \text{ cfu}\cdot\text{mL}^{-1}$ of the slurries amended with $100 \text{ mg}\cdot\text{L}^{-1}$ of atrazine. The microbial atrazine degradation seemed to be the best in the slurries sonicated twice a day for 10 min.

4. Discussion

Atrazine concentrations were the lowest in the slurries, and they could not be increased by sonication to the same levels as in the sterilized slurries, the difference being as much as $30.8 \text{ mg}\cdot\text{L}^{-1}$

between the slurries and sterilized slurries sonicated twice a day for 10 min, while only about $1.4 \pm 1.2 \text{ mg}\cdot\text{L}^{-1}$ of atrazine was adsorbed to the sediments. The sediments were collected from the depth of 11.3–14.6 m in groundwater area having atrazine in sediments and groundwater [10,25]. As the atrazine exposure increases the density or activity of atrazine-degrading microorganisms [5], a significant portion of atrazine in the slurries appeared to be degraded by inherent sediment microorganisms. Microbial growth increased by atrazine addition in the atrazine-amended sediment slurries compared to non-amended slurries, and ammonium, nitrate, and nitrogen concentrations in the sediments were below the detection limit. This suggests that the increased microbial growth could be related to the use of atrazine as a nitrogen source.

Microbial atrazine degradation ceased at high concentrations above the water solubility limit of about $33 \text{ mg}\cdot\text{L}^{-1}$ [29], at about $48.8 \text{ mg}\cdot\text{L}^{-1}$ in the non-sonicated slurries, and at about $63.2 \text{ mg}\cdot\text{L}^{-1}$ in the sonicated slurries. For comparison, pesticide dichlobenil dissipated in subsurface deposit slurries to the low concentration of $1.0\text{--}6.3 \text{ mg}\cdot\text{L}^{-1}$, and its degradation product BAM to $3.0\text{--}11.3 \text{ mg}\cdot\text{L}^{-1}$ [30]. Atrazine forms strong complexes in a solution with organic and inorganic sediment colloids [31,32]. Such interactions could prevent microbial atrazine degradation in the slurries, and also the atrazine release by sonication for microbial degradation.

Mild sonication could enhance microbial atrazine degradation, as 15 min sonication/240 min cycle has increased microbial ethanol production from cellulose (sonication horn, 36 kHz, 150 W) [33]. In this study, microbial atrazine degradation was the best in the sediment slurries sonicated twice a day for 10 min. However, sonication improved microbial degradation only slightly or not at all, as atrazine concentrations in the sonicated slurries did not fall below those in the non-sonicated slurries. In addition, the sonication time range that affected atrazine concentrations and allowed microbial atrazine degradation was narrow. Once a day sonications for 10 min, or twice a day sonications for 5 min did not affect atrazine concentrations, while the increase in the sonication time to twice a day for 20–30 min may already have inhibited microbial atrazine degradation. Similarly, 20 min sonications at the intervals of 120/240 min have enhanced microbial phenantrene degradation in a solid-liquid system, but not 25 min sonications at intervals of 205 min in two-liquid-phase systems (sonication horn, 42 kHz, 70 W) [24,34]. Biological activity started to decrease after 5 min sonication (40 kHz , $0.115 \text{ W}/\text{cm}^3$) in the regeneration of biological activated carbon [35]. Continuous sonication for 12 h prevented growth, cell division and sugar metabolism of *Klebsiella oxytoca* strain P2 (sonication horn, 36 kHz, 150 W) [33]. Microbes tolerate stronger sonication conditions in the slurries than in the pure liquid, and intermittent sonication better than continuous, while sonication time ranges improving microbial degradation may be quite narrow.

Microbial atrazine degradation in the slurries may have increased differences in atrazine concentrations between the slurries and sterilized slurries. Simultaneously autoclaving likely increased atrazine concentrations in the sterilized slurries compared to slurries (Figures 1 and 2, Table 1), similarly as anilines and trichlorophenoxyacetic acid (2,4,6-T) were less bound in autoclaved soils than in soils [36,37]. Autoclave sterilization decreases atrazine adsorption by altering the sediment structure and sorption capacity, such as dissolved organic carbon/matter in solution decreased atrazine adsorption [38,39]. Organic matter released in autoclaving could increase solubility, as humic substances in water increased chlordane solubility [40]. In the sediments of this study, organic matter and element concentrations were quite low, compared to the subsurface deposits [10,28] and materials like biochars [26], which resulted in low atrazine adsorption ($1.4 \pm 1.2 \text{ mg}\cdot\text{L}^{-1}$). It is difficult to estimate precisely how much of the difference in atrazine concentrations between the slurries and sterilized slurries was due to autoclaving and how much was due to microbial degradation. Part of atrazine could also be solid in the slurries of this study. However, atrazine was quite evenly distributed in the liquid phase, based on small deviations in the results between parallel experimental flasks under continuous shaking. Further, sonication suspended solid atrazine particles homogeneously in the slurries. Based on the highest measured concentrations in the sterilized slurries, the most appropriate

sonication time for dissolving atrazine and homogenizing atrazine suspensions was between 10 and 20 min.

Atrazine concentrations in the sterilized slurries sonicated twice a day for 30 min were lower than in those sonicated twice a day for 10 min (Figure 3). The twice a day sonications for 30 min may have been strong enough to cause chemical atrazine dissipation in the sterilized slurries, while atrazine concentrations in the slurries sonicated twice a day for 5–30 min were the same. The autoclave sterilization has disturbed the composition of subsurface deposits and enhanced chemical atrazine dissipation [10]. In this study, sonication conditions needed for chemical atrazine dissipation in the autoclave-sterilized slurries seemed to be milder than in the slurries.

According to the literature, chemical atrazine dissipation by sonication in aqueous solution has varied between 20% and 100%, depending on sonication powers, frequencies, and the type of the sonication equipment (Table 2). In studies using sonication horn, 20%–55% of atrazine was dissipated at the frequency of 20 kHz and powers of 18.5–22.8 W [14,15]. When the frequency increased to 500 kHz, the atrazine degradation rate increased to 100% [15]. When the power increased to 500 W, the atrazine degradation rate increased to >81% [13]. Generally, in the presence of the same sonication power and frequency, the chemical atrazine degradation decreases when the sonication horn is changed to the sonication bath, and aqueous solution is changed to the sediment slurry. Water in the sonication bath and soil/sediment in the slurry reduce the sonication efficiency for chemical atrazine degradation. In studies using the sonication horn in soil slurries and sonication powers between 150 and 1700 W (20 kHz), over 75% of atrazine was degraded [16,17]. Xu et al. [18] used the sonication bath with 400 kHz frequency and 120 W power to degrade 45% (100%) of atrazine in water using long sonication times of 60 (360) min. In this study, the sonication at the 43 kHz frequency and 320 W power twice a day for 30 min may have caused chemical atrazine dissipation of <26.6% of the initial quantity of 100 mg·L⁻¹ (day 13, Figure 2). However, concentrations did not decrease below those in the non-sonicated sterilized slurries. With the equipment used, longer sonication times are needed to increase chemical atrazine dissipation in the slurries.

Table 2. The percentages of atrazine degraded by sonication. The initial concentrations in experiments, and sonication frequency, power and time.

Sonication System	Sonication			Atrazine		Reference
	Frequency (kHz)	Power (W)	Time (min)	Concentration	Degradation%	
Sonication horn						
Water (pH 3)	20	500	150	0.193 mM	81	[13]
Water (pH 3)	20	500	60	0.057 mM	>90	[13]
Water	20	25	360	0.1 mM	20	[14]
Water (pH 7)	20	18.5	120	0.1 mM	55	[15]
Water (pH 7)	500	18.5	80	0.1 mM	100	[15]
Soil slurry (30 wt. %)	20	1700	2	14 µg/g	>94	[16]
Soil slurry (50 wt. %)	20	150	10	58 µg/g	>85	[17]
Soil slurry (16.6 wt. %)	20	150	10	58 µg/g	75	[17]
Sonication bath						
Slurry (23 wt. %)	43	320	5-30	0.46 mM *	0–27	This study
Water (pH 6.5)	400	120	60 (360)	0.02 mM	45 (>90)	[18]

* 333 µg/g.

Atrazine sale in Finland ended in 1992, but atrazine is still found in groundwater and subsurface sediments [10,25]. The results of this study showed that microbes able to degrade atrazine occur in the drilling site sediments collected from the groundwater recharge area, but the microbial atrazine degradation ceased at high concentrations. Therefore, microbial atrazine degradation may be limited during water run-off through the deep vadose zone sediments into the groundwater, though atrazine dissipation in flowing water has been greater than in stagnant water [41]. The interactions of atrazine and particles in the water phase appeared to be strong, in addition to the adsorption to sediments.

These interactions should be overcome for bioremediation of atrazine-contaminated groundwater and sediments. The sonication powers, frequencies and times able to release atrazine from the interactions were quite close to those damaging microorganisms. The enhancement of microbial atrazine degradation in environment by sonication may be difficult in sediments with variable composition. However, sonication can be used to enhance chemical atrazine degradation through direct pyrolysis on particle surfaces, and/or through formation of hydroxyl radicals and other radical species [17,18]. Xu et al. [18] found that atrazine sonodegradation proceeded to ammeline through the pathways of alkylic oxidation, dechlorination-hydroxylation, alkylic hydroxylation and dealkylation.

5. Conclusions

Atrazine appeared to be degraded in the slurries by indigenous sediment microbes, as atrazine concentrations could not be increased by sonications twice a day for 10 min, but they increased in the sterilized slurries. In addition, microbial numbers increased in the sediment slurries amended with $100 \text{ mg}\cdot\text{L}^{-1}$ of atrazine, but not in the slurries without atrazine addition, suggesting that atrazine was used as a nitrogen source in the slurries exhausted of nitrogen. However, microbial atrazine degradation was inhibited at high concentrations above the water solubility limit of $33 \text{ mg}\cdot\text{L}^{-1}$, at about $48.8 \text{ mg}\cdot\text{L}^{-1}$ in the non-sonicated slurries, and at about $63.2 \text{ mg}\cdot\text{L}^{-1}$ in the sonicated slurries. All water-soluble and also part of atrazine above the solubility limit (up to $48.8\text{--}63.2 \text{ mg}\cdot\text{L}^{-1}$) seemed not to be bioavailable, apparently due to the interactions with the slurry particles and sediments. Sonication times needed to release atrazine from such interactions for microbial degradation appeared to be close to those damaging microbes. Autoclave sterilization affected sediment structure, decreased atrazine adsorption, and increased its concentration in the sterilized slurries. Atrazine seemed to dissipate chemically in the sterilized slurries under milder sonication conditions than in the slurries. The concentrations in the sterilized slurries did not increase by twice a day sonications for 30 min, as they did in the slurries where microbial degradation may have impaired. However, atrazine concentrations in the slurries and sterilized slurries did not fall due to twice a day sonications for 5–30 min (43 kHz, 320 W, sonication bath) below those in the non-sonicated slurries and sterilized slurries. Although indigenous microbial atrazine degradation was observed in the heterogeneous sediments at high concentrations, it may be difficult to improve by sonication, opposite to the hypothesis. However, sonication can be used for enhanced chemical atrazine dissipation. In addition, the results revealed that microbial atrazine degradation ceased in the slurries at high concentrations above the water solubility limit.

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References

1. Meffe, R.; de Bustamante, I. Emerging organic contaminants in surface water and groundwater: A first overview of the situation in Italy. *Sci. Total Environ.* **2014**, *481*, 280–295. [[CrossRef](#)] [[PubMed](#)]
2. Lazorko-Connon, S.; Achari, G. Atrazine: Its occurrence and treatment in water. *Environ. Rev.* **2009**, *17*, 199–214. [[CrossRef](#)]
3. Mandelbaum, R.T.; Allan, D.L.; Wackett, L.P. Isolation and characterization of a *Pseudomonas* sp. that mineralizes the s-triazine herbicide atrazine. *Appl. Environ. Microbiol.* **1995**, *61*, 1451–1457. [[PubMed](#)]

4. Kruger, E.L. Enhanced Degradation of Pesticide Wastes in Soil: Implications for Bioremediation of Agrochemical Dealer Sites. Ph.D. Thesis, Iowa State University, Ames, IA, USA, 1996, 1996. Paper 11378.
5. Krutz, L.J.; Shaner, D.L.; Weaver, M.A.; Webb, R.M.T.; Zablutowicz, R.M.; Reddy, K.N.; Huang, Y.; Thomson, S.J. Agronomic and environmental implications of enhanced s-triazine degradation. *Pest. Manag. Sci.* **2010**, *66*, 461–481. [[CrossRef](#)] [[PubMed](#)]
6. Hayes, T.B.; Collins, A.; Lee, M.; Mendoza, M.; Noriega, N.; Stuart, A.A.; Vonk, A. Hermaphroditic, demasculinized frogs after exposure to the herbicide atrazine at low ecologically relevant doses. *Proc. Natl. Acad. Sci. USA.* **2002**, *99*, 5476–5480. [[CrossRef](#)] [[PubMed](#)]
7. Omran, N.E.; Salama, W.M. The endocrine disruptor effect of the herbicides atrazine and glyphosate on *Biomphalaria alexandrina* snails. *Toxicol. Ind. Health* **2016**, *32*, 656–665. [[CrossRef](#)] [[PubMed](#)]
8. European Union. *Council Directive 91/414/EEC: Concerning the Placing of Plant Protection Products on the Market*; Council of the European Union: Brussels, Belgium, 1991.
9. Wang, J.; Zhu, L.; Wang, J.; Xie, H. Isolation and characterization of atrazine mineralizing *Bacillus subtilis* strain HB-6. *PLoS ONE* **2014**, *9*, e107270. [[CrossRef](#)] [[PubMed](#)]
10. Talja, M.K.; Kaukonen, S.; Kilpi-Koski, J.; Malin, I.; Kairesalo, T.; Romantschuk, M.; Tuominen, J.; Kontro, M.H. Atrazine and terbutryn degradation in deposits from groundwater environment within the boreal region in Lahti, Finland. *J. Agric. Food Chem.* **2008**, *56*, 11962–11968. [[CrossRef](#)] [[PubMed](#)]
11. Issa, S.; Wood, M. Degradation of atrazine and isoproturon in the unsaturated zone: A study from Southern England. *Pestic. Sci.* **1999**, *55*, 539–545. [[CrossRef](#)]
12. Chiron, S.; Fernandez-Alba, A.; Rodriguez, A.; Garcia-Calvo, E. Pesticide chemical oxidation: State-of-the-art. *Water Res.* **2000**, *34*, 366–377. [[CrossRef](#)]
13. Bahena, C.L.; Martínez, S.S.; Guzmán, D.M.; del Refugio Trejo Hernández, M. Sonophotocatalytic degradation of alazine and gesaprim commercial herbicides in TiO₂ slurry. *Chemosphere* **2008**, *71*, 982–989. [[CrossRef](#)] [[PubMed](#)]
14. Bianchi, C.L.; Pirola, C.; Ragaini, V.; Selli, E. Mechanism and efficiency of atrazine degradation under combined oxidation processes. *Appl. Catal. B* **2006**, *64*, 131–138. [[CrossRef](#)]
15. Petrier, P.; David, B.; Laguian, S. Ultrasonic degradation at 20 kHz and 500 kHz of atrazine and pentachlorophenol in aqueous solution: Preliminary results. *Chemosphere* **1996**, *32*, 1709–1718. [[CrossRef](#)]
16. Collings, A.F.; Gwan, P.B.; Sosa Pintos, A.P. Soil remediation using high-power ultrasonics. *Sep. Sci. Technol.* **2007**, *42*, 1565–1574.
17. Collings, A.F.; Gwan, P.B. Ultrasonic destruction of pesticide contaminants in slurries. *Ultrason. Sonochem.* **2010**, *17*, 1–3. [[CrossRef](#)] [[PubMed](#)]
18. Xu, L.J.; Chu, W.; Graham, N. Atrazine degradation using chemical-free process of USUV: Analysis of the micro-heterogenous environments and the degradation mechanisms. *J. Hazard. Mater.* **2014**, *275*, 166–174. [[CrossRef](#)] [[PubMed](#)]
19. Tadeo, J.L.; Sánchez-Brunete, C.; Albero, B.; García-Valcárcel, A.I. Application of ultrasound-assisted extraction to the determination of contaminants in food and soil samples. *J. Chromatogr. A* **2010**, *1217*, 2415–2440. [[CrossRef](#)] [[PubMed](#)]
20. Chisti, Y. Sonobioreactors: Using ultrasound for enhanced microbial productivity. *Trends Biotechnol.* **2003**, *21*, 89–93. [[CrossRef](#)]
21. Rokhina, E.V.; Lens, P.; Virkutyte, J. Low-frequency ultrasound in biotechnology: State of the art. *Trends Biotechnol.* **2009**, *27*, 298–306. [[CrossRef](#)] [[PubMed](#)]
22. Ojha, K.S.; Mason, T.J.; O'Donnell, C.P.; Kerry, J.P.; Tiwari, B.K. Ultrasound technology for food fermentation applications. *Ultrason. Sonochem.* **2017**, *34*, 410–417. [[CrossRef](#)] [[PubMed](#)]
23. Huang, G.; Chen, S.; Dai, C.; Sun, L.; Sun, W.; Tang, Y.; Xiong, F. Effects of ultrasound on microbial growth and enzyme activity. *Ultrason. Sonochem.* **2017**, *37*, 144–149. [[CrossRef](#)]
24. Isaza, P.A.; Daugulis, A.J. Ultrasonically enhanced delivery and degradation of PAHs in a polymer-liquid partitioning system by a microbial consortium. *Biotechnol. Bioeng.* **2009**, *104*, 91–101. [[CrossRef](#)] [[PubMed](#)]
25. Mattsson, M.K.; Liu, X.; Yu, D.; Kontro, M.H. Depth, soil type, water table, and site effects on microbial community composition in sediments of pesticide-contaminated aquifer. *Environ. Sci. Pollut. Res.* **2015**, *22*, 10263–10279. [[CrossRef](#)] [[PubMed](#)]

26. Liu, N.; Charrua, A.B.; Weng, C.-H.; Yuan, X.; Ding, F. Characterization of biochars derived from agriculture wastes and their adsorptive removal of atrazine from aqueous solution: A comparative study. *Bioresour. Technol.* **2015**, *198*, 55–62. [[CrossRef](#)] [[PubMed](#)]
27. Liu, X.; Ecartot, M.; Kontro, M.H. The physicochemical conditions of isolation source determine the occurrence of *Pseudomonas fluorescens* group species. *Ann. Microbiol.* **2015**, *65*, 2363–2377. [[CrossRef](#)]
28. Pukkila, V.; Gustafsson, J.; Tuominen, J.; Aallonen, A.; Kontro, M.H. The most-probable-number enumeration of dichlobenil and 2,6-dichlorobenzamide (BAM) degrading microbes in Finnish aquifers. *Biodegradation* **2009**, *20*, 679–686. [[CrossRef](#)] [[PubMed](#)]
29. Tomlin, C.D.S. *The Pesticide Manual*, 12th ed.; The British Crop Protection Council: Omega Park Alton, UK, 2000; pp. 42–43.
30. Pukkila, V.; Kontro, M.H. Dichlobenil and 2,6-dichlorobenzamide (BAM) dissipation in topsoil and deposits from groundwater environment within the boreal region in southern Finland. *Environ. Sci. Pollut. Res.* **2014**, *21*, 2289–2297. [[CrossRef](#)] [[PubMed](#)]
31. de Jonge, L.W.; Kjaergaard, C.; Moldrup, P. Colloids and colloid-facilitated transport of contaminants in soils: An introduction. *Vadose Zone J.* **2004**, *3*, 321–325. [[CrossRef](#)]
32. Meng, Z.; Carper, R. Effects of hydration on the molecular structure of metal ion-atrazine dimer complexes: A MOPAC (PM3) study. *J. Mol. Struct.: THEOCHEM* **2000**, *531*, 89–98. [[CrossRef](#)]
33. Wood, B.E.; Aldrich, H.C.; Ingram, L.O. Ultrasound stimulates ethanol production during the simultaneous saccharification and fermentation of mixed waste office paper. *Biotechnol. Prog.* **1997**, *13*, 232–237. [[CrossRef](#)] [[PubMed](#)]
34. Isaza, P.A.; Daugulis, A.J. Enhanced degradation of phenantrene in a solid-liquid two-phase partitioning bioreactor via sonication. *Biotechnol. Bioeng.* **2010**, *105*, 997–1001. [[PubMed](#)]
35. Liu, C.; Sun, Y.; Wang, D.; Sun, Z.; Chen, M.; Zhou, Z.; Chen, W. Performance and mechanism of low-frequency ultrasound to regenerate the biological activated carbon. *Ultrason. Sonochem.* **2017**, *34*, 142–153. [[CrossRef](#)] [[PubMed](#)]
36. Bollag, J.-M.; Blattmann, P.; Laanio, T. Adsorption and transformation of four substituted anilines in soil. *J. Agric. Food Chem.* **1978**, *26*, 1302–1306. [[CrossRef](#)]
37. Koskinen, W.C.; Cheng, H.H. Elimination of Aerobic Degradation During Characterization of Pesticide Adsorption—Desorption in Soil. *Soil Sci. Soc. Am. J.* **1982**, *46*, 256–259. [[CrossRef](#)]
38. Baskaran, S.; Bolan, N.S.; Rahman, A.; Tillman, R.W. Effect of exogenous carbon on the sorption and movement of atrazine and 2,4-D by soils. *Aust. J. Soil Res.* **1996**, *34*, 609–622. [[CrossRef](#)]
39. Ben-Hur, M.; Letey, J.; Farmer, W.J.; Williams, C.F.; Nelson, S.D. Soluble and solid organic matter effects on atrazine adsorption in cultivated soils. *Soil Sci. Soc. Am. J.* **2003**, *67*, 1140–1146. [[CrossRef](#)]
40. Johnson-Logan, L.R.; Brohears, R.E.; Klaine, S.J. Partitioning behavior and the mobility of chlordane in groundwater. *Environ. Sci. Technol.* **1992**, *26*, 2234–2239. [[CrossRef](#)]
41. Schwab, A.P.; Splichal, P.A.; Banks, M.K. Persistence of atrazine and alachlor in ground water aquifers and soil. *Water, Air, Soil Pollut.* **2006**, *171*, 203–235. [[CrossRef](#)]

