BOREAL ENVIRONMENT RESEARCH 18: 269–279 ISSN 1239-6095 (print) ISSN 1797-2469 (online) © 2013 Helsinki 28 June 2013

Effect of microtopography on isotopic composition of methane in porewater and efflux at a boreal peatland

Maxim Dorodnikov^{1)2)*}, Maija Marushchak²⁾, Christina Biasi²⁾ and Martin Wilmking¹⁾

¹⁾ Institute of Botany and Landscape Ecology, University of Greifswald, Grimmer Str. 88, D-17487 Greifswald, Germany

Received 11 Mar. 2011, final version received 8 June 2012, accepted 30 May 2012

Dorodnikov, M., Marushchak, M., Biasi, Ch. & Wilmking, M. 2013: Effect of microtopography on isotopic composition of methane in porewater and efflux at a boreal peatland. *Boreal Env. Res.* 18: 269–279.

The application of stable isotopes is an approach to identify pathways of methanogenesis, methane (CH_{4}) oxidation and transport in peatlands. We measured the stable C isotopic characteristics (δ^{13} C) of CH₄ in peat profiles below hummocks, lawns and hollows of a Finnish mire to study the patterns of CH_4 turnover. Porewater CH_4 concentrations ([CH_4]; at 0.5–2 m) increased with depth below all microforms. Emissions of CH_4 from hummocks were the lowest, and increased with the increasing water-saturated zone, being ~10 times higher from hollows. Thus, the microtopography of the peatland did not affect the porewater $[CH_4]$ in the water-saturated part of the peat profile, but the CH_4 emissions were affected due to differences in the oxidative potential of the microforms. There was a decrease in δ^{13} C-CH₄ with depth below all microforms indicating dominance of CO₂reduction over acetate cleavage pathway of methanogenesis at deep peat layers. However, estimated potential portions of transported CH₄ comprised 50%–70% of the δ^{13} C-CH₄ enrichment on microforms at the 0.5-m depth, hereby masking the acetate cleavage pathway of methanogenesis. Stable C composition (δ^{13} C) of CH₄ proved to be a suitable (but not sufficient) tool to differentiate between types of methanogenesis in continuously watersaturated layers below microforms of a peatland. Combined flux-based and multi-isotopic approaches are needed to better understand the CH₄ turnover process.

Introduction

Boreal peatlands represent about 15% of the total storage of terrestrial carbon (C) (Turunen *et al.* 2002) and are substantial contributor (30%) of methane (CH₄) — an important greenhouse gas (IPCC 2007) — to the atmosphere (Reeburgh *et al.* 1998). Understanding the processes of C cycling in boreal peatlands is thus critical

for estimating current and future global CH_4 budgets.

Generally, C cycling in peatlands is controlled by a number of natural parameters, which results in high heterogeneity of CH_4 fluxes. In addition to the well-studied controls of CH_4 fluxes in peatlands, such as water-table position, peat temperature and substrate quality (reviewed in Lai 2009), identification of the C substrates

²⁾ Department of Environmental Science, Bioteknia 2, University of Eastern Finland, FI-70211 Kuopio, Finland (*corresponding author's e-mail: maxim.dorodnikov@uef.fi)

for CH_4 production also plays a key role for understanding spatial and temporal variations of CH_4 fluxes.

Application of stable isotopes is an approach to identify the pathway by which CH₄ is formed (Whiticar 1999, Conrad 2005). Methane produced by acetate cleavage (acetoclastic pathway) is not as depleted in ¹³C as CH₄ produced from CO₂ reduction with H₂ (hydrogenotrophic pathway) (Whiticar et al. 1986). Based on vertical profiles of CH₄ stable isotope ratios in peat, it was shown that the upper peat profile of wetlands was dominated by acetoclastic and the lower profile by hydrogenotrophic methanogenesis (Hornibrook et al. 1997, Popp et al. 1999). Knowledge about the contribution of different methanogenic pathways to the total CH₄ production within a peat profile will help to identify the pattern of decomposition of fresh vs. old C, thus, the fate of C pools with rapid turnover time vs. long term C pools (Beer & Blodau 2007). Enrichment of ¹³C in CH₄ in the upper peat profile is also caused by methanotrophic activity, as methanotrophic microbes discriminate strongly against ¹³C (Whiticar 1999, De Visscher et al. 2004). Transport of CH_4 either mediated by plants or due to simple diffusion through the peat profile also preferentially removes ¹²C-CH₄ from the soil. This fractionation depends on a transport mechanism, water-table level, time of day, and season (Popp et al. 1999, De Visscher et al. 2004, Chanton et al. 2005).

Whereas some information exists about seasonal and vertical changes in isotopic composition of CH_{A} in peat profiles (Avery *et al.*) 1999, Steinmann et al. 2008), there is a lack of information about the effect of the peatland microtopography on the patterns of CH₄ isotopic signatures. The surface of a (boreal) peatland can be differentiated into microscale subunits, so called microforms (hummocks, lawns, hollows), according to hydrological characteristics (water table level) and main vegetation communities (Becker et al. 2008). In turn, plant communities, especially bryophytes, are good predictors of CH₄ flux, and vascular vegetation may play an active and passive role in promoting CH₄ emission (Bubier et al. 1995). Carbon compounds exuded from plant roots can act as labile substrates which enhance methanogenesis, and vascular plants may act as a conduit for CH_4 from anaerobic zone of a wetland to the atmosphere bypassing oxidation in the aerobic zone (reviewed by Lai 2009). Water-table levels vary between microforms increasing in the order hummocks–lawns–hollows, thus resulting in differences in thickness of the oxidative zone and, hence, in CH_4 fluxes. Studies utilizing chamber technique to measure CH_4 emissions generally show the lowest CH_4 fluxes at hummocks and the highest at hollows (Dalva *et al.* 2001, Johansson *et al.* 2006, Forbrich *et al.* 2010). However, the processes involved in methanogenesis in deeper layers below different microforms have not been well studied.

In the current study, we aimed to identify the CH₄ production pathways at hummocks, lawns and hollows of a minerogenic, oligotrophic low-sedge pine peatland in Finland. We used stable C isotopic characteristics (δ^{13} C) of CH₄ to differentiate between types of methanogenesis in continuously water-saturated layers below microforms of the peatland and attempted to follow CH, throughout the peat profile up to the atmosphere. The research questions of the study were: (i) how do $[CH_{4}]$ and $\delta^{13}C$ -CH₄ at the three microform types change from the below-ground water-saturated peat layers to CH₄ emitted to the atmosphere and (ii) how does microtopography of the peatland affect the CH₄ turnover processes? Based on the research questions we put forward the following hypothesis: contribution of hydrogenotrophic vs. acetoclastic type of methanogenesis should increase with depth and differ between microforms due to differences in plant communities and water table depth at hummocks, lawns and hollows.

Material and methods

Experimental site

The study was conducted on a natural minerogenic, oligotrophic low-sedge pine fen Salmisuo in eastern Finland, located in the North Karelian Biosphere Reserve (62°47′N, 30°56′E). The site is described in detail elsewhere (Saarnio *et al.* 1997, Alm *et al.* 1999, Becker *et al.* 2008, Jager *et al.* 2009). The surface of the peatland was subdivided into three main microform types according to vegetation communities and moisture conditions: dry and elevated hummocks (dominating plants *Eriophorum vaginatum*, *Pinus sylvesteris*, *Andromeda polifolia*, *Sphagnum fuscum*), intermediate lawns (*Eriophorum vaginatum*, *Sphagnum balticum*, *Sphagnum papillosum*), and wet hollows (*Scheuchzeria palustris*, *Sphagnum balticum*). During the study period, the depth of the water table was -23 ± 5 cm from the surface of hummocks, -5 ± 2 cm from the surface of lawns and 0 ± 2 cm on hollows.

Porewater CH₄ collection and CH₄ flux measurements

The CH₄ sampling campaign was carried out between 1 and 20 July 2009. During that time span weather conditions were moderately humid with 26 mm of precipitation and the average daily temperature of 18.1 °C. Because of these weather conditions, no substantial water-tablelevel fluctuations were observed at the experimental site.

Porewater CH₄ was sampled in situ using modified diffusion chambers ("peepers"; Steinmann and Shotyk 1996). A diffusion chamber consisted of a polypropylene centrifuge tube (Rotilabo Eco 50 ml, 30×115 mm) with a cutout window $(20 \times 65 \text{ mm})$ and a polyethersulfone membrane filter (Sterlitech Corp., WA, USA) tightly sealed over the window by melted polyethylene (Steinel Vertrieb GmbH, Germany). Chosen materials of the chamber were inert to chemical composition of peatland water and resistant to microbial activity, thus minimizing potential influence on porewater [CH₄] and its isotopic composition. Pore diameter of the membrane filter (0.2 μ m) allowed ions and dissolved gases to enter the inner volume of a chamber, but prevented penetration of microorganisms and fine roots (Steinmann and Shotyk 1996). Prior to installation on the peatland, each chamber with closed cap was tested for watertightness. Diffusion chambers (76 units) pre-filled with deionized water were inserted below the microforms (8 hummocks, 7 lawns, 5 hollows) at depths of 0.5, 1.0, 1.5 and 2.0 m. All installed chambers were allowed to equilibrate for 20 days. Preliminary tests showed that this amount of time was sufficient for equilibration of CH₄ in diffusion chambers with the surrounding environment (data not shown). To install chambers into the peat below the microforms, polypropylene tubes (diameter 40 mm, wall thickness 2 mm) were used. Separate tubes were used for each of the four depths studied. Each installation tube was closed with a cap from the bottom preventing peat from filling the tube during installation. A side of a tube was perforated about 15 cm from the bottom in order to allow free movement of water through the tube. Tubes were vertically inserted into the peat down to the depths studied and left for two days prior to installation of the diffusion chambers in order to allow the peat to recover from the disturbance. After 20 days of porewater CH₄ equilibration, ca. 30 ml aliquot of the water in diffusion chambers was transferred with a syringe to glass bottles (100 ml; flushed with N₂ and prevacuated) at the site, and transported to the laboratory for subsequent measurements. All the chambers including those at the shallowest 0.5-m depth under hummocks were permanently under water during the equilibration period.

Measurements of CH_4 efflux to the atmosphere were performed using the closed chamber technique (Forbrich et al. 2010). Namely, aluminium chambers with the size $600 \times 600 \times$ 320 mm were employed. The chambers were equipped with a vent tube and a fan to allow for air mixing inside the chamber. Chamber fluxes were measured from previously installed frames at the same sampling plots, where porewater CH₄ was collected. Five air samples were taken with 60-ml syringes for determination of CH, flux at even intervals during closure time of 20 min. The 5th sample was taken in duplicate for ¹³C-CH₄ measurements. During the 20 days porewater sampling period, chamber CH₄ fluxes were measured four times at each sampling plot.

Methane concentrations in chamber flux samples were analyzed within one day from sampling with a gas chromatograph (Shimadzu 14-A) equipped with a flame ionisation detector. Two repeated measurements were made from each gas sample. Porewater [CH₄] was measured from the headspace of 100 ml sample bottles (3 replicates of 1 ml) on the same chromatograph but using a calibration standard with a higher CH_4 concentration (50 ppm instead of 3 ppm for chamber flux samples).

Stable isotopic analysis of C-CH₄

The samples of porewater and emitted CH₄ were injected with over pressure to 35 ml Wheaton glass vials equipped with rubber septa. The vials were further sealed with hot-melt glue for storage until the stable isotopes of C-CH₄ were analyzed at the Department of Environmental Science, University of Eastern Finland, Kuopio. Porewater samples with high [CH₄] were diluted with 99.999% N₂ to reach concentrations suitable for the analysis (no such dilution was needed for chamber CH₄ flux samples). The ¹³C/¹²C ratios were determined with an isotope ratio mass spectrometer (Delta plus XP; Thermo, Bremen, Germany) interfaced with a gas chromatograph (Trace GC Ultra, Finnigan) by a continuous flow system (Conflo III; Thermo Finnigan Germany; GC/C/IRMS) as described in Kankaala et al. (2007).

Calculations and statistics

Porewater [CH₄] were recalculated into μ mol l⁻¹, and above-ground CH₄ flux is given in mg m⁻² h⁻¹. The natural ¹³C/¹²C ratio in CH₄ was expressed in δ ¹³C per mil PDB (‰):

$$\delta^{13}$$
C (‰) = [(R_{sample}/R_{PDB}) - 1] × 1000, (1)

where R_{sample} is the isotopic ratio ${}^{13}\text{C}/{}^{12}\text{C}$ of CH₄ in the sample, and R_{PDB} is the isotopic ratio of Pee Dee Belemnite as the standard for C.

Stable C isotopic composition (δ^{13} C) of emitted CH₄ was calculated according to Krüger *et al.* (2002) by applying a correction for the contribution of the isotopic composition of atmospheric CH₄ present at the time of the chamber closure. An initial [CH₄] of 2.06 ± 0.2 ppm (atmospheric value from eddy measurements at the same site; I. Forbrich, unpubl. data) and an initial δ^{13} C-CH₄ of -44.93‰ ± 1.98‰ (atmospheric value from a littoral wetland in the same region; N. Welti unpubl. data) were used in the calculations. Because of very low emission rates of CH_4 at hummocks, it was not possible to estimate the respective $\delta^{13}C-CH_4$ values in the fluxes above hummocks.

The differences in porewater $[CH_4]$, CH_4 fluxes and $\delta^{13}C$ - CH_4 values between microforms and depths were evaluated with two-way ANOVA and Fischer's LSD test using STA-TISTICA 7.0 (StatSoft, USA). Prior to testing, all the data were checked for normality (Kholmogorov-Smirnov test) and homogeneity (Levene's test). The variables were treated as independent for all depths below a microform type and a certain depth between microforms.

A simple model for isotopic fractionation was used to assess the potential effect of CH_4 oxidation and transport on shifts in $\delta^{13}C-CH_4$ across the peat profile for current experimental data (adapted from Liptay *et al.* 1998). The larger the estimated portion of CH_4 transported and/or oxidized, the weaker evidence for methanogenic pathway is provided by measured $\delta^{13}C-CH_4$ values.

The portion of CH_4 transported in a peat profile was calculated using the following equation:

$$|f_{\rm tr}| = (\delta_{n+1} - \delta_n) / [(\alpha_{\rm tr} - 1) \times 10], \qquad (2)$$

where $f_{\rm tr}$ is the portion (%) of transported CH₄, δ_n and δ_{n+1} are the δ^{13} C values of CH₄ in lower- and upper-laying peat horizons, respectively, and $\alpha_{\rm tr}$ is the isotopic fractionation associated with gas transport ($\alpha_{tr} = 1.0178$ from De Visscher *et al*. 2004). It has to be noted, that the gas diffusion, in theory, should result in the enrichment of CH₄ of the *n* layer as compared with the n + 1 layer (towards which the "lighter" CH_4 is diffused). Therefore, negative $|f_{\mu}|$ values are acceptable, which, in turn, may indicate the direction of gas diffusion. For example, f_{tr} between the depths of 2.0 and 1.5 m under hollows is calculated as follows: $\delta_{2.0 \text{ m}} = -69.2\%$ and $\delta_{1.5 \text{ m}} = -70.1\%$, hence $|f_{tr}| = [(-70.1) - (-69.2)]/[(1.0178 - 1) \times$ 10] = 5%.

The portion of CH_4 oxidized in the aerobic surface layer (within 0.5–0 m) and in the watersaturated rooted zone of aerenchymatic plants (0.5–1.0 m) of peatland was calculated using the following equation:

$$f_{ox} = (\delta_n - \delta_{n+1})/[(\alpha_{ox} - 1) \times 10],$$
 (3)

where f_{ox} is the portion (%) of oxidized CH₄, δ_n and δ_{n+1} are the δ^{13} C values of CH₄ from the porewater horizon 0.5 m and CH₄ emission, and 0.5 m and 1.0 m peat horizons, respectively; α_{ox} is the fractionation factor accounting for the preference of methanotrophic microbes for CH₄ containing the lighter C isotope ($\alpha_{ox} = 1.022$ from Coleman *et al.* 1981, Liptay *et al.* 1998).

For the $f_{\rm tr}$ calculations we assumed that CH₄ transport but not oxidation ($a_{\rm ox} = 1$) had a predominant effect on δ^{13} C-CH₄ in the deep watersaturated horizons (from 2 m to 1.0 m depth). Oxidation mostly affected δ^{13} C in the surface peat (0–0.5 m) and, hence, the emitted δ^{13} C-CH₄ above the peat surface ($a_{\rm tr} = 1$), whereas between the depths of 0.5 and 1.0 m, both CH₄ transport and oxidation in a rooted zone of aerenchymatic plants are equally important for the δ^{13} C-CH₄ values ($f_{\rm tr} + f_{\rm ox}$). The δ^{13} C-CH₄ at 2-m depth was assumed to be unaffected by diffusion, since lateral advection was reported to have a negligible effect on δ^{13} C-CH₄ (Chanton *et al.* 2002).

Results

Porewater [CH₄] and above-ground CH₄ fluxes

Porewater $[CH_4]$ increased with depth down to 1.5 m below all microforms (Fig. 1). At the depth of 2.0 m, however, porewater $[CH_4]$ did not differ below hummocks and lawns, but was significantly lower below hollows and as compared with the 1.5-m depth (Fig. 1 and Appendix 1). Type of microform (hummocks *vs.* lawns *vs.* hollows) had no statistically significant effect on porewater $[CH_4]$ at any of the depths studied (Fig. 1 and Appendix 3).

Fluxes of CH₄ decreased in the order hollows \geq lawns > hummocks and did not differ significantly among measurement days (Table 1). The lowest fluxes were 0.4 mg CH₄ m⁻² h⁻¹ at hummocks and the highest 6.0 mg CH₄ m⁻² h⁻¹ at hollows (Table 1). Because there were no sig-



Fig. 1. Porewater CH₄ concentrations (μ mol I⁻¹) at different depths below hummocks (n = 8), lawns (n = 7) and hollows (n = 5). Error bars show standard errors. Values followed by the same letters are not significantly different (at $p \le 0.05$ according to two-way ANOVA and Fischer's LSD test) between depths below each type of a microform. There were no significant differences between types of a microform for any depth horizon.

nificant differences between measurement days, the average of four CH_4 flux values could be related to porewater $[CH_4]$ equilibrated during 20 days of the field campaign. This was especially important for comparison of stable ¹³C/¹²C isotope ratio in porewater and emitted CH_4 .

Stable ¹³C/¹²C isotope ratios in porewater and emitted CH₄

Generally, there was an overall decrease of porewater δ^{13} C-CH₄ values with depth below all microforms (Fig. 2). However, no significant differences in δ^{13} C-CH₄ were found below 1 m under any microform (Fig. 2). Porewater at the shallowest depth (0.5 m) was enriched in δ^{13} C-CH₄ (-62.5‰ to -64‰) as compared



Fig. 2. Porewater δ^{13} C-CH₄ values ± SE (‰) at different depths below hummocks (n = 8), lawns (n = 7), hollows (n = 5) and δ^{13} C-CH₄ values (‰) in emission from the surface of lawns (n = 7) and hollows (n = 5). The δ^{13} C-CH₄ values in emission from hummocks could not be determined because of low CH₄ efflux. Values followed by the same letters are not significantly (at $p \le 0.05$ according to two-way ANOVA and Fischer's LSD test) different between depths below each type of a microform and emission (where possible). There were no significant differences between types of a microform for a depth horizon and in emission.

with porewater the deeper peat layers (up to -71%). There were no significant differences in $\delta^{13}\text{C-CH}_4$ among microforms at any of the studied depths (Fig. 2 and Appendix 2)

CH₄ emitted from the lawns and hollows was significantly more depleted in ¹³C (-68‰ to -69‰) than porewater CH₄ at the 0.5 m depth (Appendix 2) but the values were in the same range as δ^{13} C-CH₄ values at the other depths (Fig. 2). For hummocks, δ^{13} C-CH₄ values in CH₄ emission are not shown due to low CH₄ fluxes, which made it impossible to differentiate between δ^{13} C in CH₄ efflux and ambient atmospheric δ^{13} C-CH₄. Where measurable, δ^{13} C values in emitted CH₄ were not significantly affected by the type of microforms (Fig. 2).

Assessed portions of transported (f_{tr}) and oxidized (f_{ox}) CH₄

The calculated f_{tr} was the smallest at the deep peat horizons (1.0–2.0 m) comprising 3%–5% (Fig. 3). The intermediate peat horizon (0.5– 1.0 m) was the most affected by the processes of CH₄ transport and oxidation, since $f_{tr} + f_{ox}$ ranged between 51% and 68% under hummocks and lawns-hollows, respectively (Fig. 3 and Appendix 4). In contrast to porewater CH₄, the estimation of f_{ox} in CH₄ emitted from lawns and hollows provided negative (unreliable) values due to higher depletion of δ^{13} C-CH₄ in efflux as compared with δ^{13} C-CH₄ in porewater δ^{13} C-CH₄ at 0.5 m (Fig. 3). At hummocks, it was not possible to estimate f_{ox} because of unreliable δ^{13} C values measured in emitted CH₄.

Discussion

Porewater [CH₄] and above-ground CH₄ fluxes: effect of microtopography

The overall increase in porewater CH_4 with depth at the studied boreal mire complex is in agreement with the results of many other stud-

Table 1. Above-ground CH_4 fluxes from hummocks (n = 8), lawns (n = 7) and hollows (n = 5) during 20 days of the field campaign (1–20 July 2009). Same letters indicate no significant differences (at $p \le 0.05$ according to two-way ANOVA and Fischer's LSD test) between the types of microsites on one sampling date. There were no significant differences among dates of sampling for each type of microsite.

Microsite type	Day of sampling	Flux ± SE (mg m ⁻² h ⁻¹)
Hummock	5	0.40 ± 0.09^{a}
	9	0.45 ± 0.15 ^a
	13	0.40 ± 0.08^{a}
	17	0.42 ± 0.07^{a}
Lawn	5	3.69 ± 0.62^{b}
	9	3.43 ± 0.59 ^b
	13	3.79 ± 0.41 ^b
	17	4.53 ± 0.64 ^b
Hollow	5	5.10 ± 0.69 ^b
	9	4.95 ± 0.87 ^b
	13	5.05 ± 0.65 ^b
	17	5.97 ± 0.87^{b}





ies (Hornibrook et al. 1997, Chasar et al. 2000, Steinmann et al. 2008). Along with this, a threshold of 1.0 m existed, below which $[CH_{4}]$ was not significantly different (Fig. 1). Since the deeper vs. upper peat layers have lower hydraulic conductivity and less temperature fluctuations, and typically remain anoxic year round (Hornibrook et al. 1997), the CH_4 production there is sustained. Seemingly, such homogeneity of deeper peat layers revealed no effect of the type of a microform (hummocks, lawns, hollows) on the [CH₄] (Fig. 1 and Appendix 1). However, the current results are preliminary and longer observations are required to reveal the possible effect of microtopography of a peatland on the belowground $[CH_{4}]$ dynamics.

In contrast to porewater $[CH_4]$, CH_4 emitted from the surface was affected by microtopography: the lowest fluxes were found at the elevated hummocks and the highest at the water-saturated hollows, while intermediate fluxes of CH_4 were measured at the lawns (Table 1 and Appendix 3). The current results are consistent with those reported earlier for the same site (Saarnio *et al.* 1997, Forbrich *et al.* 2010) and those from other

observations (Dalva *et al.* 2001, Johansson *et al.* 2006) and can be explained by the increase of the oxidation layer in the order hollows < lawns < hummocks.

Isotopic evidence on CH₄ production, transport and oxidation

Porewater CH₄ below 1-m depth was significantly more depleted in ¹³C as compared with the shallowest 0.5-m depth (Fig. 2 and Appendix 1). Still, similarly to $[CH_4]$, $\delta^{13}C$ -CH₄ values did not differ significantly among microforms (and Appendix 2). Although rather few studies exist on $\delta^{13}C$ -CH₄ in peat layers below 1 m depth, our data are in a good agreement with most of the reported results from *Typha*-dominated fen in Canada (Hornibrook *et al.* 1997), littoral wetlands in the United States (Chasar *et al.* 2000) and acidic *Sphagnum* bog in Switzerland (Steinmann *et al.* 2008).

Overall depletion of ${}^{13}C$ in CH₄ with depth suggests increased contribution of hydrogenotrophic or CO₂ reduction pathway to the total methanogenesis (Whiticar *et al.* 1986, Hornibrook *et al.* 1997, Conrad 2005), as the discrimination of methanogenic microbes against heavier ¹³C is stronger during the hydrogenotrophic pathway as compared to the acetoclastic pathway (methanogenesis due to splitting of fermentation-derived acetic acid/acetates). Thus, the hydrogenotrophic pathway of methanogenesis at the depths below 1.0 m was sustained for the entire experimental site and was not affected — at least in the short-term — by the microtopography of the studied peatland.

It is important to note, that δ^{13} C-CH₄ values of the upper peat horizon (above 1.0 m) of the studied peatland could have been substantially affected by the processes of CH₄ transport and oxidation (Popp et al. 1999, Whiticar 1999, Chanton et al. 2005). This limits our possibilities to draw conclusions about the importance of the different methanogenic pathways for the total CH₄ production. In this study, we assumed that molecular diffusion, driven by the [CH₄] gradient between the anaerobic peat layers and the atmosphere, was the dominant transport mechanism by which CH₄ was released from the peatland. Although ebullition is an important transport mechanism in some wetlands (Glaser et al. 2004, Lai 2009), it was not considered in this study, since no events of ebullition were detected during the 20-day field campaign (neither by chamber measurements, nor by eddy covariance). Molecular diffusion affects isotopic composition of CH₄ differently than the other transport mechanisms, plant-mediated CH₄ transport and ebullition (Chanton et al. 1992, 2005, Liptay et al. 1998, De Visscher et al. 2004). When CH_4 is released by ebullition or through plants, it bypasses oxidation in the aerobic zone of peatland and, thus, remains unaffected by the substantial isotopic fractionation that occurs when CH₄ is oxidized. Both microbial-culture studies (Coleman et al. 1981, Liptay et al. 1998) and field studies (Tyler et al. 1994, Chanton et al. 2005) have shown that methanotrophic organisms preferentially consume lighter isotopes, leaving residual CH₄ enriched in ¹³C. Based on differences in δ^{13} C-CH₄ among peat layers, and CH₄ efflux and isotopic fractionation factors associated with gas transport and oxidation, we could assess the potential portions of transported/oxidized CH₄. This allowed understanding to which extent our data of δ^{13} C-CH₄ could be affected by the mentioned processes. The larger the portion of CH₄ transported or oxidized, the less certainly we can determine the methanogenic pathway based on measured δ^{13} C-CH₄ values, since changes in δ^{13} C-CH₄ induced by the oxidation and/or diffusive transport would mask the isotopic signal from methanogenesis.

Our data suggest that CH₄ transport by diffusion had a relatively small effect on δ^{13} C-CH₄ in deep peat horizons (> 1 m), as about 3% to 5% of CH₄ was potentially transported from 2-m through 1.5-m to 1-m depth during the 20-day measurement period. This was probably due to a relatively small [CH₄] gradient between 1-m and 2-m depths, and is generally in agreement with the concept of CH₄ dynamic in deeper peat horizons discussed above. Hence, δ^{13} C-CH₄ in deep peat layers below microforms were predominately affected by the methanogenic pathway (hydrogenotrophy) and to a lesser extent by CH₄ transport.

In contrast to the depths below 1 m, δ^{13} C-CH₄ between 1 m and 0.5 m was much more affected by CH₄ transport (Appendix 4). Moreover, CH₄ oxidation processes related to the supply of oxygen by aerenchymatic plants to roots in water saturated peat horizons (Joabsson and Christensen 2001) may have additionally affected δ^{13} C-CH₄ values measured at the 0.5–1.0-m depths. Thus, estimated portions of both transported and oxidized CH₄ at these depths reached about 70% under lawns and hollows. Thus, the relative enrichment of porewater CH₄ in ¹³C at the shallowest depth (0.5 m) of the studied peatland as compared with the deeper peat horizons cannot be attributed solely to the production by acetoclastic methanogenesis but to a large extent to CH₄ transport (diffusion and/or plant-mediated transport) and oxidation in the rhizosphere of aerenchymatic plants.

The calculated portions of oxidized CH₄ (f_{ox}) within the top 0.5 m of peat showed a negligible effect of oxidation on δ^{13} C of CH₄ emission from the lawns and hollows, which is in agreement with higher water table at these microforms. For the hummocks with thicker aerobic peat layer, oxidation was probably substantial, but it was not possible to reliably calculate δ^{13} C values in CH₄ due to low rates of efflux and small [CH₄]. The relative depletion of ¹³C in CH₄ emission from the lawns and hollows as compared with δ^{13} C-CH₄ at the 0.5-m depth could be attributed to the dilution with more ¹³C-depleted CH₄ from water-saturated peat horizons (deeper than 0.5 m) by means of plant-mediated transport (Chanton *et al.* 1992, Whalen 2005, Lai 2009).

Outlook

As discussed above, the effect of CH_4 transport and oxidation is especially important for the upper peat layers, where these processes can obscure the isotopic signature of acetoclastic methanogenesis and probably also level down the effect of microtopography on δ^{13} C-CH₄ values. Additional isotopic characteristics of CH₄ (D) and CO₂ (13 C) would help to reveal patterns of CH₄ turnover in peatlands, including the processes of CH₄ formation, transport and consumption (Bellisario et al. 1999, Clymo and Bryant 2008, Steinmann et al. 2008). Further, more information about $[CH_{4}]$ and $\delta^{13}C$ -CH₄ in the uppermost peat horizon (0-0.5 m) would be required. On the other hand, the lack of effect of peatland microtopography on porewater [CH₄] and its isotopic composition may be attributed to the relatively short time-span of the current study. Hence, studies extended in time and in space (including other peatlands with similar biogeochemical characteristics) may provide insights into the effects of microtopography onto the processes of CH_4 turnover in peatlands.

Conclusions

Stable C isotopic composition of porewater and emitted CH_4 proved to be a suitable (but not sufficient) tool to differentiate between types of methanogenesis in continuously water-saturated layers under microforms of a peatland. Combined flux-based and multi-isotopic approaches are needed to better understand the CH_4 turnover process. Based on $[CH_4]$ in porewater, CH_4 fluxes to the atmosphere and δ^{13} C-CH₄ values we conclude:

 The CO₂ reduction pathway contributed more than the acetate cleavage to total methanogenesis *in situ* in deep peat layers (> 1 m), whereas in the upper peat horizons (< 1 m) CH₄ transport and oxidation may substantially enrich ¹³C-CH₄ hence masking the ¹³C-CH₄ enrichment due to acetoclastic pathway of methanogenesis.

 The microtopography of the studied peatland had an effect on CH₄ emission but not on [CH₄] in the water-saturated peat layers. The above-ground CH₄ fluxes increased in the order hummocks < lawns ≤ hollows. This trend was most probably caused by the oxidative potential of the studied microforms.

Acknowledgements: Authors would like to greatly acknowledge the staff of the Laboratory for Experimental Ecology, Mekrijärvi research station, University of Joensuu, Finland and personally Prof. Taneli Kolström, Matti Lemettinen, Teijo Kortevaara, Eine Ihanus, Risto Ikonen for providing necessary conditions for work and accommodation. The study was supported by DFG Emmy Noether Programm (Wi 2680/2-1) and a Sofja Kovalevskaja Award (M. Wilmking) of the Alexander von Humboldt Foundation.

References

- Alm J., Saarnio S., Nykänen H., Silvola J. & Martikainen P. 1999. Winter CO₂, CH₄ and N₂O fluxes on some natural and drained boreal peatlands. *Biogeochem*. 44: 163–186.
- Avery G.B.Jr., Shannon R.D., White J.R., Martens C.S. & Alperin M.J. 1999. Effect of seasonal changes in the pathways of methane production on the δ^{13} C values of pore water methane in a Michigan peatland. *Global Biogeochem. Cycles* 13: 475–484.
- Becker T., Kutzbach L., Forbrich I., Schneider J., Jager D., Thees B. & Wilmking M. 2008. Do we miss the hot spots? The use of very high-resolution aerial photographs to quantify carbon fluxes in peatlands. *Biogeosciences* 5: 1387–1393.
- Beer J. & Blodau C. 2007. Transport and thermodynamics constrain belowground carbon turnover in a northern peatland. *Geochim. Cosmochim. Acta* 71: 2989–3002.
- Bellisario L.M., Bubier J.L., Moore T.R. & Chanton J.B. 1999. Controls on CH₄ emissions from a northern peatland. *Glob. Biogeochem. Cycles* 13: 81–91.
- Bubier J., Moore T.R. & Juggins S. 1995. Predicting methane emission from bryophyte distribution in northern peatlands. *Ecology* 76: 677–693.
- Chanton J.P., Arkebauer T.J., Harden H. & Verma S.B. 2002. Diel variation in lacunal CH₄ and CO₂ concentration and δ^{13} C in *Phragmites australis*. *Biogeochem*. 59: 287–301.
- Chanton J.P., Chasar L.S., Glaser P. & Siegel D.I. 2005. Carbon and hydrogen isotopic effects in microbial methane from terrestrial environments. In: Flanagan L.B., Ehleringer J.R. & Pataki D.E. (eds.), *Stable isotopes and biosphere–atmosphere interactions*, Physiological Ecol-

ogy, Elsevier, Amsterdam, pp. 85-105.

- Chanton J.P., Whiting G.J., Showers W.J. & Crill P.M. 1992. Methane flux from *Peltandra virginica*: stable isotope tracing and chamber effects. *Global Biochem. Cycles* 6: 15–31.
- Chasar L.S., Chanton J.P., Glaser P.H. & Siegel D.I. 2000. Methane concentration and stable isotope distribution as evidence of rhizospheric processes: comparison of a fen and bog in the glacial lake Agassiz peatland complex. *Ann. Bot.* 86: 655–663.
- Clymo R.S. & Bryant C.L. 2008. Diffusion and mass flow of dissolved carbon dioxide, methane, and dissolved organic carbon in a 7-m deep raised peat bog. *Geochim. Cosmochim. Acta* 72: 2048–2066.
- Coleman D.D., Risatti J.B. & Schoell M. 1981. Fractionation of carbon and hydrogen isotopes by methane-oxidizing bacteria. *Geochim. Cosmochim. Acta* 45: 1033–1037.
- Conrad R. 2005. Quantification of methanogenic pathways using stable carbon isotopic signatures: a review and proposal. Org. Geochem. 36: 739–752.
- Dalva M., Moore T.R., Arp P. & Clair T.A. 2001. Methane and soil and plant community respiration from wetlands, Kejimkujik National Park, Nova Scotia: Measurements, predictions, and climatic change. J. Geophys. Res. 106(D3): 2955–2962.
- De Visscher A., De Pourcq I. & Chanton J. 2004. Isotope fractionation effects by diffusion and methane oxidation in landfill cover soils. J. Geophys. Res. 109, D18111, doi:10.1029/2004JD004857.
- Forbrich I., Kutzbach L., Hormann A. & Wilmking M. 2010. A comparison of linear and exponential regression for estimating diffusive CH₄ fluxes by closed-chambers in peatlands. *Soil Biol. Biochem.* 42: 507–515.
- Glaser P.H., Chanton J.P., Morin P., Rosenberry D.O., Siegel D.I., Ruud O., Chasar L.I. & Reeve A.S. 2004. Surface deformations as indicators of deep ebullition fluxes in a large northern peatland. *Global Biogeochem. Cycles* 18:GB1003, doi:10.1029/2003GB002069.
- Hornibrook E.R.C., Longstaffe F.J. & Fyfe W.S. 1997. Spatial distribution of microbial methane production pathways in temperate zone wetland soils: stable carbon and hydrogen isotope evidence. *Geochim. Cosmochim. Acta* 61: 745–753.
- IPCC 2007. Climate change 2007: synthesis report. Intergovernmental Panel on Climate Change Fourth Assessment Report (AR4), IPCC, Geneva, Switzerland.
- Jager D., Wilmking M. & Kukkonen J. 2009. The influence of summer seasonal extremes on dissolved organic carbon export from a boreal peatland catchment: Evidence from one dry and one wet growing season. *Sci. Total Environ.* 407: 1373–1382.
- Joabsson A. & Christensen T.R. 2001. Methane emissions from wetlands and their relationship with vascular plants: an Arctic example. *Global Change Biol.* 7: 919– 932.
- Johansson T., Malmer N., Crill P.M., Friborg T., Akerman

J.H., Mastepanov M. & Christensen T.R. 2006. Decadal vegetation changes in a northern peatland, greenhouse gas fluxes and net radiative forcing. *Global Change Biol*. 12: 2352–2369.

- Kankaala P., Taipale S., Nykänen H. & Jones R.I. 2007. Oxidation, efflux, and isotopic fractionation of methane during autumnal turnover in a polyhumic, boreal lake. J. Geophys. Res.-Biogeo. 112, G02003, doi:10.1029/2006JG000336.
- Krüger M., Eller G., Conrad R. & Frenzel P. 2002. Seasonal variation in pathways of CH₄ production and in CH₄ oxidation in rice fields determined by stable carbon isotopes and specific inhibitors. *Global Change Biol.* 8: 265–280.
- Lai D.Y.F. 2009. Methane dynamics in northern peatlands: A review. *Pedosphere* 19: 409–421.
- Liptay K., Chanton J., Czepiel P. & Mosher P. 1998. Use of stable isotopes to determine methane oxidation in landfill cover soils. J. Geophys. Res. 103: 8243–8250.
- Popp T.J., Chanton J.P., Whiting G.J. & Grant N.1999. Methane stable isotope distribution at a *Carex* dominated fen in north central Alberta. *Global Biogeochem. Cycles* 13: 1063–1077.
- Reeburgh W.S., King J.Y., Regli S.K., Kling G.W., Auerbach N.A. & Walker D.A. 1998. A CH₄ emission estimate for the Kupurak River basin. *Alaska J. Geophys. Res.* 103(D22): 29005–29013.
- Saarnio S., Alm J., Silvola J., Lohila A., Nykänen H. & Martikainen P. 1997. Seasonal variation in CH₄ emissions and production and oxidation potentials at microsites on an oligotrophic pine fen. *Oecologia* 110: 414–422.
- Steinmann P., Eilrich B., Leuenberger M. & Burns S.J. 2008. Stable carbon isotope composition and concentrations of CO_2 and CH_4 in deep catotelm of a peat bog. *Geochim. Cosmochim. Acta* 72: 6015–6026.
- Steinmann P. & Shotyk W. 1997. Chemical composition, pH, and redox state of sulphur and iron in complete vertical porewater profiles from tow *Sphagnum* peat bogs, Jura Mountains, Switzerland. *Geochim. Cosmochim. Acta* 61: 1143–1163.
- Turunen J., Tomppo E., Tolonen K. & Reinikainen A. 2002. Estimating carbon accumulation rates of undrained mires in Finland — application to boreal and subarctic regions. *Holocene* 12: 69–80.
- Tyler S.C., Crill P.M. & Brailsford G.W. 1994. ¹³C/¹²C fractionation of methane during oxidation in a temperate forested soil. *Geochim. Cosmochim. Acta* 58: 1625–1633.
- Whalen S.C. 2005. Biogeochemistry of methane exchange between natural wetlands and the atmosphere. *Environ*. *Eng. Sci.* 22: 73–94.
- Whiticar M.J., Faber E. & Schoell M. 1986. Biogenic methane formation in marine and freshwater environments: CO₂ reduction vs. acetate fermentation — isotope evidence. *Geochim. Cosmochim. Acta* 50: 693–709.
- Whiticar M.J. 1999. Carbon and hydrogen isotope systematics of bacterial formation and oxidation of methane. *Chem. Geol.* 161: 291–314.

	SS	df	MS	F	p
[CH,] among depths below					
hummocks	7662878	3,22	2554293	2.8	0.063
lawns	5219294	3,20	1739765	2.0	0.152
hollows	6760264	3,20	2253421	3.2	0.047
[CH,] among microforms at					
0.5 m	506960	2,16	253480	0.3	0.726
1.0 m	2192443	2,15	1096221	1.7	0.221
1.5 m	1017656	2,16	508828	0.7	0.494
2.0 m	1381486	2,15	690743	0.6	0.586

Appendix 1. Comparisons (ANOVA) of CH₄ concentrations [CH₄] among microsites and depths.

Appendix 2. Comparisons (ANOVA) of δ^{13} C-CH₄ among microsites and depths.

	SS	df	MS	F	р
$\delta^{_{13}}$ C-CH, among depths below					
hummocks	150	3,22	50	9.9	< 0.001
lawns	262	3,20	87	19.2	< 0.001
hollows	218	3,20	73	23.1	< 0.001
δ^{13} C-CH, among microforms at					
0.5 m	1.2	2,16	0.6	0.1	0.865
1.0 m	6.0	2,15	3.0	1.0	0.402
1.5 m	12.8	2,16	6.4	3.8	0.046
2.0 m	8.3	2,15	4.1	0.5	0.619

Appendix 3. Comparisons (ANOVA) of above-ground CH₄ fluxes from peatland's microforms during the field campaign.

	SS	df	MS	F	р
CH, fluxes among sampling points from					
hummocks	0.1	3,28	0.04	0.1	0.959
lawns	15.7	3,24	5.2	2.2	0.110
hollows	23.3	3,16	7.8	0.9	0.451
CH, fluxes among different microforms of					
sampling pont 1	47.2	2,17	23.6	12.8	< 0.001
sampling pont 2	98.6	2,17	49.3	7.7	0.004
sampling pont 3	66.9	2,17	33.5	32	< 0.001
sampling pont 4	133.5	2,17	66.8	26.3	< 0.001

Appendix 4. Comparisons (ANOVA) of estimated portions of CH_4 oxidized (f_{ox}) and transported (f_{tr}) below peatland's microforms and among depths.

	SS	df	MS	F	р
f_{tat} f among depths below					
hummocks	1.1	2,16	0.5	12	0.001
lawns	1.8	2,15	0.9	60.6	< 0.001
hollows	1.9	2,15	0.9	25.4	< 0.001
f_{μ}, f_{μ} among microforms at					
0.5–1.0 m	0.118	2,16	0.059	0.9	0.443
1.0–1.5 m	0.007	2,15	0.004	0.6	0.554
1.5–2.0 m	0.004	2,15	0.002	0.1	0.916