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# Genetic variation in *CFH* predicts phenytoin-induced maculopapular exanthema in European-descent patients

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Neurology<sup>®</sup> 2018;90:e1-e10. doi:10.1212/WNL.00000000004853

## Abstract

## Objective

To characterize, among European and Han Chinese populations, the genetic predictors of maculopapular exanthema (MPE), a cutaneous adverse drug reaction common to antiepileptic drugs.

## Methods

We conducted a case-control genome-wide association study of autosomal genotypes, including Class I and II human leukocyte antigen (HLA) alleles, in 323 cases and 1,321 drugtolerant controls from epilepsy cohorts of northern European and Han Chinese descent. Results from each cohort were meta-analyzed.

#### **Results**

We report an association between a rare variant in the complement factor H–related 4 (*CFHR4*) gene and phenytoin-induced MPE in Europeans ( $p = 4.5 \times 10^{-11}$ ; odds ratio [95% confidence interval] 7 [3.2–16]). This variant is in complete linkage disequilibrium with a missense variant (N1050Y) in the complement factor H (*CFH*) gene. In addition, our results reinforce the association between *HLA-A\*31:01* and carbamazepine hypersensitivity. We did not identify significant genetic associations with MPE among Han Chinese patients.

### Conclusions

The identification of genetic predictors of MPE in CFHR4 and CFH, members of the complement factor H–related protein family, suggest a new link between regulation of the complement system alternative pathway and phenytoin-induced hypersensitivity in Europeanancestral patients. **Correspondence** Prof. Cavalleri gcavalleri@rcsi.ie

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The Article Processing Charge was funded by the European Commission OpenAIRE2020 project.

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# Glossary

ADR = adverse drug reaction; AED = antiepileptic drug; cADR = cutaneous adverse drug reaction; *CFH* = complement factor H; *CFHR4* = complement factor H–related 4 gene; CI = confidence interval; CPNDS = Canadian Pharmacogenomics Network for Drug Safety; GWAS = genome-wide association study; HLA = human leukocyte antigen; ILAE-CGC = International League Against Epilepsy Complex Genetics Consortium; MPE = maculopapular exanthema; OR = odds ratio; SJS/TEN = Stevens-Johnson syndrome and toxic epidermal necrolysis; SNP = single nucleotide polymorphism.

Idiosyncratic cutaneous adverse drug reactions (cADRs) can have a genetic predisposition. The HLA-B\*15:02 allele is a predictor of carbamazepine-induced Stevens-Johnson syndrome and toxic epidermal necrolysis (SJS/TEN) in individuals of Han Chinese and Southeast Asian descent, while a recent meta-analysis suggests that the allele is also a significant risk factor for oxcarbazepine-, phenytoin-, and lamotrigine-induced SJS/TEN.<sup>1,2</sup> However, the association with HLA-B\*15:02 does not extend to the milder but more common maculopapular exanthema (MPE) phenotype, and the allele is specific to individuals of Asian descent, limiting clinical utility across populations.<sup>3,4</sup> HLA-A\*31:01 has been confirmed as a transethnic risk factor for carbamazepineinduced cADRs, with the allele observed across populations of European, Japanese, and Korean descent.<sup>5-8</sup> Recently, HLA-A\*24:02 has been shown to associate with SJS in Han Chinese patients, irrespective of causal drug studied.<sup>9</sup> Genetic variation beyond the major histocompatibility locus has also been associated with cADRs. The CYP2C9\*3 allele correlates with phenytoin hypersensitivity in Han Chinese from Taiwan,<sup>10</sup> with a similar effect reported in a Thai population.<sup>11</sup> However, a genome-wide association study (GWAS) of lamotrigine and phenytoin-induced cADRs in Europeans did not detect significant predictors.<sup>12</sup> A summary of the associated genetic risk variants for cADRs in various populations is provided in table e-1 (links.lww.com/WNL/A56).

The EpiPGX Consortium was established to identify genetic markers of epilepsy treatment response. The International League Against Epilepsy Complex Genetics Consortium (ILAE-CGC) facilitates the discovery of genetic variants influencing epilepsy predisposition.<sup>13</sup> The EPIGEN consortium is a worldwide epilepsy genetics research framework and the Canadian Pharmacogenomics Network for Drug Safety (CPNDS) is an active surveillance network focused on identifying genomic markers of severe adverse drug reactions (ADRs) in children and adults.<sup>14</sup> Collaboration among these consortia has provided detailed phenotypes and genotypes for over 15,000 epilepsy cases, for the investigation of antiepileptic drug (AED)–induced MPE.

Availing of the joint resources of EpiPGX, ILAE-CGC, EPIGEN, and CPNDS, this study aimed to characterize, among European and Han Chinese populations, the genetic predictors of MPE, a cutaneous ADR common to particular AEDs. Specifically, we set out to test the following hypotheses: (1) whether population-specific genetic variants predict

MPE; (2) whether transethnic genetic variants predict MPE; (3) whether population-specific genetic variants predict AED-specific MPE; and (4) whether transethnic genetic variants predict AED-specific MPE.

## Methods

# Standard protocol approvals, registrations, and patient consents

All study participants provided written, informed consent for genetic analysis. Local institutional review boards approved study protocols at each contributing site.

## Study design

We conducted a retrospective case-control study in individuals of European and Han Chinese ethnicity. Participants were exposed to carbamazepine, lamotrigine, phenytoin, or oxcarbazepine. Our analyses were structured to test genetic variants for association with MPE within and across both of the broad ancestral groups, through logistic regression of genotype dosage and subsequent meta-analysis of regression coefficients. We tested for association with (1) aromatic AED-induced MPE vs controls tolerant to at least 3 aromatic AEDs, (2) carbamazepine-induced MPE vs carbamazepine-tolerant controls, (3) lamotrigine-induced MPE vs lamotrigine-tolerant controls, and (4) phenytoin-induced MPE vs phenytointolerant controls. Due to small sample size, oxcarbazepinerelated MPE was not analyzed as an individual case cohort.

## **Cohorts and phenotype definition**

Epilepsy cohorts from the ILAE-CGC, EpiPGX, and EPIGEN Consortia were included in the discovery GWAS metaanalysis (table 1). A European-descent replication cohort was assembled from sites in Brazil, Canada (via CPNDS), Liverpool, and other sites across the United Kingdom. Cases were defined as having MPE attributed to carbamazepine, lamotrigine, phenytoin, or oxcarbazepine as determined by their clinician, occurring within 3 months of initiation and resolving upon dose reduction or AED withdrawal. Control patients trialed carbamazepine, lamotrigine, phenytoin, or oxcarbazepine for at least 3 months without reporting a cADR. Epilepsyspecific patient demographics are presented in table e-2 (links. lww.com/WNL/A56).

## **Genotyping and imputation**

Genotyping of a subset of EpiPGX samples was performed at deCODE Genetics (Reykjavik, Iceland) using Illumina (San

Table 1	Breakdown of antiepileptic drug (AED)–induced maculopapular exanthema (MPE) cases and AED-tolerant
	controls in discovery dataset

	All aromatic AEDs		CBZ		LTG		РНТ	
Ethnicity	MPE <sup>a</sup>	Control <sup>b</sup>	MPE	Control	MPE	Control	MPE	Control
European	259	979	95	869	118	812	52	472
Han Chinese <sup>c</sup>	116	342	85	197	16	32	22	58
Subtotal	375	1,321	180	1,066	134	844	74	530

Abbreviations: CBZ = carbamazepine; ILAE = International League Against Epilepsy; LTG = lamotrigine; PHT = phenytoin.

<sup>a</sup> Individual participant counts only, despite 16 patients being cross-reactive to more than 1 AED.

<sup>b</sup> A total of 1,321 controls were tolerant to all 3 of CBZ, LTG, and PHT.

<sup>c</sup> Fifty-two carbamazepine-induced MPE cases from Guangzhou were available for analysis of human leukocyte antigen serotype data only.

Diego, CA) OmniExpress-12 v1.1 and OmniExpress-24 v1.1 single nucleotide polymorphism (SNP) arrays. The remainder of samples were genotyped locally using various Illumina beadchip SNP arrays, details of which are published elsewhere.<sup>13</sup> Genotyping and imputation quality control is described in appendix e-1 (links.lww.com/WNL/A57).

#### **Study power**

We estimated that the study had 80% power to detect a genetic predictor of relative risk (approximated to odds ratio)  $\geq$ 3 with an allele frequency  $\geq$ 2% and an  $\alpha$  level of  $1.25 \times 10^{-8}$ . Power for AED-specific and population-specific analyses are detailed in appendix e-1 (links.lww.com/WNL/A57).

#### Statistical analyses

Association analyses were conducted within the European and Asian subgroups using an additive logistic regression model. To account for genotype uncertainty, SNPTEST was used to apply a missing data likelihood score model that included sex, clinical site, and 5 principal components as covariates to control for bias and population stratification.<sup>15</sup> Fixed effects meta-analyses were conducted across the European and Asian subgroups using the software package METAL, applying genomic control correction within cohorts.<sup>16</sup> The threshold for statistical significance was set at  $1.25 \times 10^{-8}$ , reflecting an empirical Bonferroni correction for 4 tests, of the standard  $5 \times 10^{-8}$  genome-wide significance threshold. Conditional association analysis was performed on loci containing significant markers to establish whether other genetic variants in the region (1 Mb upstream and downstream) were independently associated with MPE. The conditional threshold for significance was set at  $5 \times 10^{-6}$ , based on a genome-wide estimation of 10,000 imputed variants per 2 MB region.<sup>13</sup> We applied the Stouffer z trend test to the combined results from the discovery and replication cohorts.

## **Confirmatory genotyping**

Where an association signal satisfied the threshold for significance, additional genotyping and resequencing were performed in a subset of patients and results were compared with imputation dosage files. The variant rs78239784 was confirmed by Sanger sequencing in 100 patients from the original discovery cohort. For the purpose of replication, we genotyped the rs78239784 variant in an independent cohort of 13 phenytoin-induced MPE cases and 88 phenytoin-tolerant controls.

## Results

#### **Cohort description**

In total, 375 MPE cases and 1,321 controls satisfied our criteria for inclusion in the discovery analyses (see Methods and table 1). There were 16 patients with cross-reactivity to 2 or more aromatic AEDs, 8 of whom were hypersensitive to carbamazepine and lamotrigine. Genome-wide array data for 323 cases and 1,321 controls were available for analysis. Broad European or Han Chinese ancestry was assigned to each participant according to principal components analysis (figure e-1, links.lww.com/WNL/A55).

# Genome-wide association analysis of broad aromatic AED–induced MPE

After quality control (see appendix e-1, figure e-2, links.lww. com/WNL/A55, for details), 3,693,290 variants remained for analysis in the European dataset and 4,402,554 variants in the Han Chinese dataset. We only considered autosomal SNPs in our analyses. To test hypothesis (1), that population-specific genetic markers predispose to MPE, a logistic regression analysis of all MPE cases was performed separately in the European and Han Chinese ancestral subgroups. We did not observe any genome-wide significant markers for MPE due to any aromatic AED in either Europeans or Han Chinese. The study was powered to detect an effect of relative risk >3.5 in the European cohort and >5 in the Han Chinese.

To test hypothesis (2), that transethnic genetic markers predispose to MPE, a fixed-effects meta-analysis of the association results for European and Han Chinese ancestral subgroups was performed. We did not observe any genome-wide significant markers for MPE shared among European or Han Chinese subgroups (figure 1A). This analysis was powered to detect an effect size >3.

# Genome-wide association analysis of specific aromatic AED–induced MPE

To test hypothesis (3), that genetic variants for MPE are AED-specific and population-specific, logistic regression analyses of AED-specific MPE was performed separately in the European and Han Chinese ancestral groups (figures e-3 and e-4, links.lww.com/WNL/A55). Within the European subgroup, *HLA-A\*31:01* was significantly associated with carbamazepine-induced MPE in Europeans ( $p = 1.47 \times 10^{-10}$ , odds ratio [OR] [95% confidence interval (CI)] 5.5 [3.0–10]). Conditioning on *HLA-A\*31:01* did not reveal additional variants within the human leukocyte antigen (HLA) region that were independently contributing to carbamazepine-induced MPE. No genome-wide significant signals for lamotrigine-induced MPE were observed in Europeans. This analysis was powered to detect an effect size >6.

For phenytoin-induced MPE, we identified a significant association with rs78239784, an intronic variant of the complement factor H–related 4 gene (CFHR4). The risk allele, G, had a minor allele frequency of 12% in our European phenytoin-induced MPE cases compared to 1.5% in European phenytoin-tolerant controls ( $p = 2.94 \times 10^{-10}$ , OR [95% CI] 8.8 [4.0-19]; figure 2). Conditioning on rs78239784 did not reveal additional variants in this locus that were independently contributing to phenytoin-induced MPE. Using 1000 Genomes Phase III European population data, rs78239784 was found to be in complete linkage disequilibrium with rs35274867 ( $r^2 = 1$  and D' = 1), a missense variant coding for an asparagine to tyrosine substitution at amino acid 1,050 of the complement factor H (CFH) gene. The missense variant was not present in our association results, as it was filtered during quality control because the imputation score was <0.95. The imputation accuracy of the top variant rs78239784 was confirmed in our cohort via Sanger sequencing and TaqMan approaches. Of 100 samples tested, a 100% concordance rate was found between imputed and resequenced genotypes. Within the Han Chinese subgroup, no significant associations were found between autosomal SNPs or HLA alleles and AED-specific MPE. Summary results for known risk loci in our dataset were scrutinized and are presented in table 2. None of these loci was even nominally significant (p > p)0.05) in the Han Chinese subgroup.

In order to test hypothesis (4), that transethnic genetic markers predispose to AED-specific MPE, we meta-analyzed p values from association results for carbamazepine, lamotrigine, and phenytoin individually, across European and Han Chinese ancestral subgroups. There were no shared genomewide significant markers among our meta-analyses of AED-specific MPE (figure 1, B–D).

## Replication of CFHR4 signal

To replicate the association with phenytoin-induced MPE in an independent cohort, the variant rs78239784 was genotyped in self-reported European-descent cases and controls recruited through centers in Liverpool (United Kingdom), Sao Paolo (Brazil), and across Canada (CPNDS). Two heterozygous carriers were identified among 13 phenytoininduced MPE cases while only a single carrier was observed among 88 phenytoin-tolerant controls yielding a 2-tailed Fisher exact *p* value of 0.044. Pooling all cases and controls together, we report an overall *p* value of  $4.5 \times 10^{-11}$  (with a combined OR [95% CI] 7 [3.2–16]) for the association between rs78239784 and phenytoin-induced MPE in Europeans (table 3).

## Discussion

We detected a strong association between variants in the complement factor H regulatory pathway and phenytoininduced MPE in a European-descent patient population. The presence of the associated genotype increases risk for MPE 6fold. Our results indicate that risk variants for MPE tend to be drug-specific and population-specific.

These results point to the regulators of complement activation gene cluster as a genetic locus contributing to the onset of hypersensitivity to phenytoin. The most significant variant in our European subgroup analysis, rs78239784 (c.59-2448T>G), tags the missense variant rs35274867 (p. N1050Y) in CFH, suggesting aberrant complement activation as a potential causal mechanism in a subset of phenytoinsensitive individuals. According to data from the Exome Aggregation Consortium, CFH N1050Y has an allele frequency of approximately 2% in Europeans, 3% in African subpopulations, less than 1% in South Asians, and is almost invariant in East Asians.<sup>17</sup> Given the absence of this allele in East Asian populations, the lack of an association between the CFH locus and MPE in our Han Chinese cohort is unsurprising. We propose that population-specific, independent rare variants of large effect may explain a proportion of MPE cases, a similar paradigm to the rare variant model demonstrated in Crohn disease and ulcerative colitis.<sup>18</sup> CFH N1050Y has previously been associated with type 2 diabetes-associated end-stage kidney disease in an African American cohort.<sup>19</sup> Defects in CFH-related proteins are also associated with overactivation of the complement immune system and can lead to atypical hemolytic-uremic syndrome, C3 glomerulopathy, basal laminar drusen, immunoglobulin A nephropathies, and systemic lupus erythematosus.<sup>20</sup> Further, genetic variants in CFHR4 and CFH are associated with risk for agerelated macular degeneration.<sup>21</sup> We did not detect these symptoms among N1050Y carriers. While it is unclear whether phenytoin directly interacts with circulating CFHrelated proteins, it does not specifically increase serum complement levels.<sup>22</sup> Our findings offer an expanded insight into the role of the complement alternative pathway in hypersensitivity to AEDs.

Phenytoin is still used as a first-line treatment for epilepsy in many settings and is listed on the WHO list of essential



Figure 1 Meta-analysis results across European and Han Chinese cohorts

Manhattan (a) and quantile–quantile (b) plots for the meta-analyses of maculopapular exanthema vs tolerant controls, for (A) any antiepileptic drug (genomic inflation factor [ $\lambda$ ] = 1.01), (B) carbamazepine ( $\lambda$  = 1.01), (C) lamotrigine ( $\lambda$  = 0.99), and (D) phenytoin ( $\lambda$  = 0.98).

medicines.<sup>23</sup> Epidemiologic data on prescriptions of AEDs for epilepsy in the United Kingdom show that, in 2008, phenytoin accounted for 18% of all treated person-years in epilepsy and was most frequently used in the elderly.<sup>24</sup> Therefore,

a clinically useful prognostic test for phenytoin-induced cutaneous ADRs in European-ancestral individuals would be welcome. The sensitivity of the *CFHR4* variant as a prognostic marker is 16% and the specificity is 97%, which corresponds to





(A) Manhattan and (B) quantile–quantile plot of phenytoin-induced MPE in the European subgroup ( $\lambda$  = 1.01). The LocusZoom plot (C) highlights the most significant single nucleotide polymorphism (SNP), rs78239784 (purple dot), is an intronic variant in *CFHR4*.

a positive likelihood ratio of 5.93 (95% CI 2.8–12.6) and a negative likelihood ratio of 0.86 (95% CI 0.8–0.9). Assuming the pretest probability of the ADR is 5%, a positive test for this marker increases the probability of MPE to phenytoin sixfold to 30%, while a negative test reduces the probability marginally to 4.3%. There are an estimated 6 million people with epilepsy in Europe, which means approximately 90,000 people are at-risk carriers of this mutation.<sup>25</sup> We estimate that 208 (95% CI 103–431) patients of European ancestry would need to be screened to prevent one case, based on a previously reported formula,<sup>26</sup> which corresponds to an absolute risk reduction of 0.005 (95% CI 0.002–0.009). As a comparison, it is estimated that 442 Han Chinese patients would be needed to screened for *HLA-B\*15*: 02 in order to prevent a single carbamazepine-induced SJS/ TEN case. We would suggest that the clinical utility and cost-effectiveness of implementing preemptive screening be evaluated through a prospective study.

We did not replicate the association between *CYP2C9\*3* (rs1057910) and MPE in our cohort, irrespective of ethnicity or AED. This is not surprising given that the original association with phenytoin in Han Chinese was largely driven by SJS/TEN cases, which were excluded from this analysis. We did, however, detect a nonsignificant enrichment of *CYP2C9\*3* (p = 0.08) among the European phenytoin-

			European				Han Chinese		
Drug	Marker	MPE Homozygosity/ heterozygosity/ reference (MAF)	Control Homozygosity/ heterozygosity/ reference (MAF)	OR (95% CI)	<i>p</i> Value	MPE Homozygosity/ heterozygosity/ reference (MAF)	Control Homozygosity/ heterozygosity/ reference (MAF)	OR (95% CI)	<i>p</i> Value
CBZ	HLA-B*15:02	0/0/62 (0)	0/0/869 (0)	I	1	0/2/21 (0.04)	3/22/162 (0.07)	0.6 (0.1–2.8)	0.53
CBZ	HLA-A*31:01	0/16/79 (0.08)	0/27/841 (0.02)	5.5 (3.0–10)	$1.47 \times 10^{-10}$	0/1/22 (0.02)	0/10/156 (0.03)	0.8 (0.1–6.8)	0.81
LTG	HLA-A*24:02	2/14/102 (0.08)	8/122/681 (0.09)	0.9 (0.5–1.5)	0.71	0/4/12 (0.12)	1/7/20 (0.16)	0.7 (0.1–3.5)	0.62
РНТ	CYP2C9*3	0/10/42 (0.1)	0/51/421 (0.05)	1.8 (0.9–3.8)	0.08	0/1/21 (0.02)	0/2/56 (0.02)	0.9 (0.1–12)	0.92
РНТ	rs78239784	2/8/42 (0.12)	0/14/458 (0.02)	8.8 (4.0–19)	$2.94 \times 10^{-10}$	0/0/22 (0)	0/0/58 (0)	I	I
Abbrevia Represer	itions: CBZ = carbamaz ntative risk alleles from	epine; CI = confidence interv previous genome-wide asso	val; LTG = lamotrigine; MAF <sup>:</sup> vciation study findings and th	= mean allele frequen is report for antiepilep	cy; OR = odds ratio; PH stic drug-induced skin	H = phenytoin. rash studies (CBZ, LTG, and	PHT) were assessed for ass	ociation in our study.	o Values fro

induced MPE cases, but the effect size we observe (OR 1.8) is smaller than previously reported for phenytoin-induced MPE in Han Chinese (OR 5.5).<sup>10</sup> Notably, 2 *CYP2C9\*3* carriers among the phenytoin-induced MPE cases were also heterozygous for the *CFHR4* variant while only 3 of 560 controls were jointly heterozygous. The frequency of *CYP2C9\*3* differs between controls from European and Han Chinese subgroups in our study, which is in accordance with background population frequency reported in the Exome Aggregation Consortium (European: 7%, East Asian: 3%). While our results do not support a significant effect of *CYP2C9\*3* in MPE, larger cohorts including severe cADR cases may resolve the extent of the association across populations.

*HLA-A\*31:01* was the most strongly associated marker with carbamazepine-induced MPE in Europeans in this study. Forty-three of the 95 cases studied here were also included in the discovery publication,<sup>8</sup> but the effect of the allele remains significant when we restrict to new cases only ( $p = 4 \times 10^{-7}$ ), thus providing an additional independent replication of the initial finding. We confirm that *HLA-B\*15:02* is not associated with carbamazepine-induced MPE in Han Chinese, and no novel signals emerged for carbamazepine-induced MPE in either population. No significant predictors of lamotrigine-induced MPE were observed in either population tested. *HLA-A\*24:02* was not significantly associated with lamotrigine-induced MPE in either of the European or Han Chinese ancestral subgroups; rather this allele was observed to be more frequent among our lamotrigine-tolerant controls.

Our meta-analyses did not reveal any significant transethnic genetic markers for MPE due to any AED. There were considerably more European-descent patients in this analysis than any other ethnicity, and we recognize this as a limitation of the study. Analysis of non-European cohorts is warranted. A second limitation of our study was the low number of Han Chinese lamotrigine-related MPE cases, relative to Europeandescent cases. Therefore we cannot conclusively rule out genetic predictors of modest effect size for MPE to lamotrigine in Han Chinese or other non-European descent populations. We recognize that our replication cohort for phenytoin is small and comprises self-reported ancestral Europeans. Since we did not have full genotype array data for these individuals, we relied on Fisher exact test for calculating significance rather than logistic regression with correction for principal components. Additional studies of larger sample size are required to further characterize the association, improve the estimation of the risk effect size, and determine the prognostic ability and economics of screening for this marker. Finally, as this study was not powered to investigate MPE attributed to oxcarbazepine due to low sample size, further investigation is warranted.

We have identified a genetic predictor for a common adverse reaction to phenytoin in European-descent patients, adding a new pharmacogenetic marker for potential use in the treatment of epilepsy. This finding adds to the list of genetic Table 3 rs78239784 associates with phenytoin-induced maculopapular exanthema in Europeans

rs78239784	Discovery GWAS	Replication	Combined	Control
N	52	13	63	560
Frequency (G)	0.12	0.08	0.11	0.01
p Value	2.9 × 10 <sup>-10a</sup>	0.044 <sup>b</sup>	$4.5 \times 10^{-11}$	_
OR (95% CI)	8.8 (4.0–19)	15 (1.3–17)	7.0 (3.2–16)	—

Abbreviations: CI = confidence interval; GWAS = genome-wide association study; OR = odds ratio.

Discovery, replication, and combined association test results of rs78239784 with phenytoin-induced cutaneous adverse drug reactions in Europeans.

<sup>a</sup> *p* Value from logistic regression model with sex and 5 principal components as covariates.

<sup>b</sup> p Value from Fisher exact test.

predictors of hypersensitivity to anticonvulsant therapy and opens up a new avenue for understanding the biology underlying cutaneous adverse reactions. This finding can advance genetic testing in the clinic as it expands the array of genetic tests available to aid clinicians in reducing overall rates of discontinuation due to adverse events and improving patient safety.

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#### **Author contributions**

M.M.C., H.G., L.B., S.C., P.K., and G.L.C. contributed to the conception and design of the study and the acquisition, analysis, and interpretation of data. M.M.C. and H.G. performed all statistical analysis. A.I., S.M.S., R.S., S.W., F.B., S.R., K.H., C.L., A.G.M., P.A., M.J.B., B.F., M.R.J., N.S., G.E.B.W., C.J.D., B.C.C., D.S., J.E.Z., M.K., M.P., A.A., C.D., G.J.S., B.P. C.K., P.S., F.Z., A.C., W.S.K., J.W.S., H.L., K.M.K., S.W., M.K.,

L.J.G., R.K., N.D., C.Y., F.C., I.L.-C., T.J.O., W.-p.L., L.J.G., and K.S. contributed to the acquisition, analysis, and interpretation of data. All authors contributed to the critical revision of the final version of the manuscript for important intellectual content.

### Acknowledgment

The authors thank the following coinvestigators: the members of the EpiPGX Consortium, the members of the Canadian Pharmacogenomics Network for Drug Safety (CPNDS) Consortium, the members of the International League Against Epilepsy Consortium on Complex Epilepsies (ILAE-CGC), and the members of the EPIGEN Consortium (a full list of coinvestigators is found in the e-Appendix); Ana Fulgenico-Maisch, Simona Donatello, Clare O'Kennedy, Lisa Slattery, Paula Corr, and Joanna Fay for clinical assessment of patients, technical assistance with DNA extraction, and sample management; and the patients for their participation in the study.

## **Study funding**

This study was not industry-sponsored. The work was supported by a grant from the European Commission (7th Framework Programme Grant 279062, EpiPGX). M.M.C. and G.L.C. are supported by Science Foundation Ireland, grant 13/CDA/2223, and an RCSI seed funding grant GA 14-1899. This project was supported by the General Research Funds (HKU7623/08M and HKU7747/07M to S.C., CUHK4466/06M to P.K.) and Health and Medical Research Fund (HMRF 01120086 to P.K.) from Hong Kong. Some results presented in this article were prepared using the HPC facilities of the University of Luxembourg. This work was partly undertaken at UCLH/UCL, which received a proportion of funding from the Department of Health's NIHR Biomedical Research Centres funding scheme (J.W.S., S.M.S.). The work was also supported by the Epilepsy Society, UK (J.W.S., S.M.S.), by the foundation "no epilep," the German Chapter of the ILAE (DGfE) (both to H.L.). F.C. and I.L.-C. are supported by Fundação de Amparo à Pesquisa do Estado de São Paulo, Brazil, through grant 2013/07559-3. J.E.Z. and M.P. thank the NHS Chair of Pharmacogenetics programme and MRC Centre for Drug Safety Science for support in Liverpool. B.C.C. and C.J.D.R. are supported by the Canadian Institutes of Health Research (CIHR) Drug Safety and Effectiveness Network (FRN-117588), the Canada Foundation for Innovation and the Canadian Dermatology Foundation. G.E.B.W. is supported by a CIHR Fellowship. The funders of the study had no role in the study design, data collection, data analysis, data interpretation, or writing of the report. M.M.C., H.G., and G.L.C. had full access to all the data in the study and the corresponding authors had final responsibility for the decision to submit for publication.

## Disclosure

M. McCormack and H. Gui report no disclosures relevant to the manuscript. A. Ingason is an employee of deCODE Genetics/Amgen. D. Speed, G. Wright, E. Zhang, R. Secolin, C. Yasuda, M. Kwok, S. Wolking, F. Becker, and S. Rau report no disclosures relevant to the manuscript. A. Avbersek is employed by UCB Pharma SPRL, Belgium, as Associate Director. K. Heggeli, C. Leu, C. Depondt, and G. Sills report no disclosures relevant to the manuscript. A. Marson was awarded grants from GSK, Eisai, and UCB Pharma, which funded the National Audit of Seizure Management in Hospitals. P. Auce, M. Brodie, B. Francis, M. Johnson, B. Koeleman, P. Striano, A. Coppola, F. Zara, and W. Kunz report no disclosures relevant to the manuscript. J. Sander has served on scientific advisory boards for UCB Pharma and Eisai; has served on speaker's bureaus for UCB Pharma, Eisai, Teva, and Lundbeck; has received research support from UCB Pharma, GSK, Eisai, The Marvin Weil Epilepsy Research Fund, and NL Nationaal Epilepsie Fonds; and his current position is endowed by the UK Epilepsy Society. H. Lerche has received speaker or consultancy fees or travel support from Bial, Desitin, Eisai, GlaxoSmithKline, Pfizer, UCB Pharma, or Valeant. K. Klein reports personal fees from UCB Pharma, Novartis Pharma AG, and Eisai, outside of the submitted work. AC was awarded a grant from EISAI and personal fees for speaking from Eisai, outside of the submitted work. S. Weckhuysen and M. Krenn report no disclosures relevant to the manuscript. L. Gudmundsson is an employee of de-CODE Genetics/Amgen. K. Stefánsson is an employee of deCODE Genetics/Amgen. R. Krause, N. Shear, C. Ross, and N. Delanty report no disclosures relevant to the manuscript. M. Pirmohamed reports grants from MRC and grants from UK Department of Health during the conduct of the study. B. Carleton, through the Pharmaceutical Outcomes Programme (POPi), has received financial support for its pharmacogenomics research from the Canada Foundation for Innovation (CFI), Canadian Institutes of Health Research, Genome Canada, Genome British Columbia, and the Provincial Health Services Authority. Prof. Carelton has also received support by the University of British Columbia, Child & Family Research Institute, Vancouver, and Pfizer. Prof. Carleton has a patent and applications pending for biomarkers of anthracycline-induced cardiotoxicity and cisplatin-induced ototoxicity but no relevant financial disclosures relevant to this study. F. Cendes reports speaking fees from UCB Pharma, outside of the current work. I. Cendes-Lopes and W. Liao report no disclosures relevant to the manuscript. T. O'Brien reports grants from The Royal Melbourne Foundation during the conduct of the study. S. Sisodiya and S. Cherny report no disclosures relevant to the manuscript. P. Kwan has received speaker or consultancy fees and/or research grants from Eisai, GlaxoSmithKline, Johnson & Johnson, Pfizer, and UCB Pharma. L. Baum and G. Cavalleri

Received February 27, 2017. Accepted in final form October 2, 2017.

report no disclosures relevant to the manuscript. Go to

#### References

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- Li X, Yu K, Mei S, et al. HLA-B\*1502 increases the risk of phenytoin or lamotrigine induced Stevens-Johnson syndrome/toxic epidermal necrolysis: evidence from a meta-analysis of nine case-control studies. Drug Res 2015;65:107–111.
- 2. Chung WH, Hung SI, Hong HS, et al. Medical genetics: a marker for Stevens-Johnson syndrome. Nature 2004;428:486.
- Man CB, Kwan P, Baum L, et al. Association between HLA-B\*1502 allele and antiepileptic drug-induced cutaneous reactions in Han Chinese. Epilepsia 2007;48:1015–1018.

- Lonjou C, Thomas L, Borot N, et al. A marker for Stevens-Johnson syndrome: ethnicity matters. Pharmacogenomics J 2006;6:265–268.
- Amstutz U, Ross CJ, Castro-Pastrana LI, et al. HLA-A 31:01 and HLA-B 15:02 as genetic markers for carbamazepine hypersensitivity in children. Clin Pharmacol Ther 2013;94:142–149.
- Kim SH, Lee KW, Song WJ, et al. Carbamazepine-induced severe cutaneous adverse reactions and HLA genotypes in Koreans. Epilepsy Res 2011;97:190–197.
- Ozeki T, Mushiroda T, Yowang A, et al. Genome-wide association study identifies HLA-A\*3101 allele as a genetic risk factor for carbamazepine-induced cutaneous adverse drug reactions in Japanese population. Hum Mol Genet 2011;20: 1034–1041.
- McCormack M, Alfirevic A, Bourgeois S, et al. HLA-A\*3101 and carbamazepineinduced hypersensitivity reactions in Europeans. N Engl J Med 2011;364: 1134–1143.
- Shi YW, Min FL, Zhou D, et al. HLA-A\*24:02 as a common risk factor for antiepileptic drug-induced cutaneous adverse reactions. Neurology 2017;88:2183–2191.
- Chung WH, Chang WC, Lee YS, et al. Genetic variants associated with phenytoinrelated severe cutaneous adverse reactions. JAMA 2014;312:525–534.
- Tassaneeyakul W, Prabmeechai N, Sukasem C, et al. Associations between HLA class I and cytochrome P450 2C9 genetic polymorphisms and phenytoin-related severe cutaneous adverse reactions in a Thai population. Pharmacogenet Genomics 2016;26: 225–234.
- McCormack M, Urban TJ, Shianna KV, et al. Genome-wide mapping for clinically relevant predictors of lamotrigine- and phenytoin-induced hypersensitivity reactions. Pharmacogenomics 2012;13:399–405.
- International League Against Epilepsy Consortium on Complex Epilepsies. Electronic address e-auea. Genetic determinants of common epilepsies: a meta-analysis of genome-wide association studies. Lancet Neurol 2014;13:893–903.
- Carleton B, Poole R, Smith M, et al. Adverse drug reaction active surveillance: developing a national network in Canada's children's hospitals. Pharmacoepidemiol Drug Saf 2009;18:713–721.

- Marchini J, Howie B, Myers S, McVean G, Donnelly P. A new multipoint method for genome-wide association studies by imputation of genotypes. Nat Genet 2007;39: 906–913.
- Willer CJ, Li Y, Abecasis GR. METAL: fast and efficient meta-analysis of genomewide association scans. Bioinformatics 2010;26:2190–2191.
- 17. Lek M, Karczewski KJ, Minikel EV, et al. Analysis of protein-coding genetic variation in 60,706 humans. Nature 2016;536:285–291.
- Beaudoin M, Goyette P, Boucher G, et al. Deep resequencing of GWAS loci identifies rare variants in CARD9, IL23R and RNF186 that are associated with ulcerative colitis. PLoS Genet 2013;9:e1003723.
- Bonomo JA, Palmer ND, Hicks PJ, et al. Complement factor H gene associations with end-stage kidney disease in African Americans. Nephrol Dial Transpl 2014;29: 1409–1414.
- Skerka C, Chen Q, Fremeaux-Bacchi V, Roumenina LT. Complement factor H related proteins (CFHRs). Mol Immunol 2013;56:170–180.
- Fritsche LG, Igl W, Bailey JN, et al. A large genome-wide association study of agerelated macular degeneration highlights contributions of rare and common variants. Nat Genet 2016;48:134–143.
- Basaran N, Kansu E, Hincal F. Serum immunoglobulins, complement levels and lymphocyte subpopulations in phenytoin-treated epileptic patients. Immunopharmacol Immunotoxicol 1989;11:335–346.
- The selection and use of essential medicines. World Health Organ Tech Rep Ser 2015: vii–xv:1–546.
- Nicholas JM, Ridsdale L, Richardson MP, Ashworth M, Gulliford MC. Trends in antiepileptic drug utilisation in UK primary care 1993-2008: cohort study using the General Practice Research Database. Seizure 2012;21:466–470.
- Baulac M, de Boer H, Elger C, et al. Epilepsy priorities in Europe: a report of the ILAE-IBE Epilepsy Advocacy Europe Task Force. Epilepsia 2015;56:1687–1695.
- Chen Z, Liew D, Kwan P. Real-world efficiency of pharmacogenetic screening for carbamazepine-induced severe cutaneous adverse reactions. PLoS One 2014;9: e96990.