

# Evaluation of *Bacillus thuringiensis* Berliner as an alternative control of small hive beetles, *Aethina tumida* Murray (Coleoptera: Nitidulidae)

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**Abstract** Small hive beetles, *Aethina tumida* Murray, are parasites and scavengers of honeybee colonies, *Apis mellifera* L., and have become an invasive species that can cause considerable damage in its new distribution areas. An effective subspecies of *Bacillus thuringiensis* Berliner (=Bt) would provide an alternative to chemical control of this pest. Therefore, we tested three different Bt strains [*B. thuringiensis*, var. *aizawai* (B401<sup>®</sup>), *B. thuringiensis* var. *kurstaki* (Novodor<sup>®</sup>) and *B. thuringiensis* var. San Diego *tenebrionis* (Jackpot<sup>®</sup>)] and Perizin<sup>®</sup> (3.2% coumaphos), each applied on combs with a pollen diet fed to pairs of adult beetles. This evaluates the products for the suppression of successful small hive beetle reproduction. While none of the tested Bt strains showed a significant effect on the number of produced wandering larvae, we could confirm the efficacy of coumaphos for the control of small hive beetles. We further show that it is also efficient when applied with a lower concentration as a liquid on the combs. We suggest the continued search for efficient Bt strains naturally infesting small hive beetles in its endemic and new ranges, which may become a part of the integrated management of this pest.

## Introduction

The small hive beetle (=SHB), *Aethina tumida* Murray, is a parasite and scavenger of honeybee colonies, *Apis mellifera* L., native to Sub-Saharan Africa (Hepburn and Radloff [1998](#)). It has recently become an invasive species in North America (1996), Egypt (2000) and Australia (2002; cf. Neumann and Elzen [2004](#)). In North America, it continues to cause considerable damage in the populations of European-derived honeybees (Elzen et al. [1999](#); Hood [2004](#)). Adults and larvae of *A. tumida* can be successfully controlled in beehives with coumaphos as a contact agent using Check-Mite+<sup>TM</sup> strips (Bayer Healthcare AG, containing 10% coumaphos). The authors used the Check-Mite+<sup>TM</sup> plastic strips, cut in half and attached under bottom board trapping devices made of cardboard (Elzen et al. [1999](#)). Check-Mite+<sup>TM</sup> strips were initially applied for the control of the parasitic mite, *Varroa destructor* Anderson and Trueman. However, such conventional control carries the risk of pest resistance, as has already occurred in *V. destructor* (Pettis [2003](#)). Moreover, chemical residues may accumulate in the bee products (Neumann and Elzen [2004](#)).

*Bacillus thuringiensis* Berliner (=Bt) is efficiently used in a wide range of different integrated pest management programs. The Bt bacterium produces proteinaceous crystals encoded by cry genes during sporulation, which become toxic when digested by insects. Parts of the dissolved crystals bind to midgut glucoprotein receptors and subsequently toxic molecules are inserted into the plasma membrane, forming pores that cause changes in ion flux. The resulting osmotic swelling and lysis of midgut epithel cells eventually lead to the death of susceptible pest organisms (Krieg [1986](#); Gill et al. [1992](#)). Toxic crystals of Bt strains containing the cryII gene are successfully used in biological control systems, mostly against larvae of certain Lepidopteran and Dipteran pests. Some Bt strains are also efficacious against coleopteran larvae (Slaney et al. [1992](#)), which seem to be mostly susceptible to crystal proteins (cry) encoded by cryIII genes (Höfte and Whiteley [1989](#)). According to the considerable selectivity of Bt toxins (Höfte and Whiteley [1989](#); Schnepf et al. [1998](#)), even closely related species will not be affected (Slaney et al. [1992](#)), making Bt a suitable, as well as one of the most successful biological control agents. Aiming towards substitution of solely chemical control, we here evaluate the efficacy of commercially available products containing different Bt strains as an alternative control of SHB. Since the feeding larvae are the most destructive live stage of SHB, we concentrate on the ability of Bt to inhibit successful reproduction of SHB.

# Materials and methods

The following products containing different *Bt* strains were tested to determine their efficacy against SHB.

1. B401<sup>®</sup>, a biological larvicide used against wax moths, *Galleria mellonella* L., which contains delta-endotoxins of *B. thuringiensis* var. *aizawai*, serotype 7.
2. Jackpot<sup>®</sup>, containing *B. thuringiensis* var. *kurstaki*, which is used to control several Lepidopteran species,
3. Novodor<sup>®</sup>, based on *B. thuringiensis* var. San Diego *tenebrionis*, which is used against larvae of the Colorado potato beetle, *Leptinotarsa decemlineata* (Say) and of the Elm leaf beetle, *Pyrralta luteola* Muller.

We also tested Perizin<sup>®</sup> (3.2% coumaphos) in a liquid application form because Check-Mite+<sup>™</sup> strips containing coumaphos (10%) were successfully used as a contact treatment against SHB (Elzen et al. [1999](#)). Water was used as a negative control. As we did not assume an effect of *Bt* on adult SHB, we used the most practicable procedure to get 1st instar larvae on the food source by giving beetles the possibility to feed, mate and oviposit on treated combs. If the beekeeper would apply this method in the field, he would also spray the combs in the beehive with all stages of SHB present.

The experiments were conducted in February 2004 and 2005 in an environmental chamber [+30°C in permanent darkness] at Rhodes University (Grahamstown, Eastern Cape Province, South Africa). Adult SHB were reared on honeybee products using standard protocols (Neumann et al. [2001](#); Muerrle and Neumann [2004](#)). Newly emerging beetles were sexed (Schmolke [1974](#)) and kept in separate containers of their own sex with food ad lib (honey and water on cotton wool) to prevent mating prior to the experiments. Pieces of empty comb, filled with 5 g pollen as a protein source (Ellis et al. [2002](#)), were sprayed with the *Bt* products, Perizin<sup>®</sup> or water [0.5 ml H<sub>2</sub>O as control] according to the instructions of the respective supplier, using a manual plastic pump sprayer. The combs were placed in plastic containers [Ziplock, 946 ml] and five pairs of adult SHB were added to each container. Round holes [Ø 5.5 cm] covered with pieces of gauze [12 × 12 cm, mesh width ≤ 0.50 mm] provided air and prevented adult and larval escape.

For the evaluation of B401<sup>®</sup> (0.5 ml of 5% solution) and Perizin<sup>®</sup> [0.2 ml (Perizin<sup>®</sup> 3.2%: H<sub>2</sub>O, 1:50) + 0.3 ml H<sub>2</sub>O] in 2004, the adult SHB of both treatments and control (*N* = 5 replicates each) were given 5 days for feeding, mating and oviposition on the combs [8 × 5 cm] before their removal. When the emerging offspring have consumed the treated food, untreated pollen was provided ad lib until the larvae reached the wandering phase (Schmolke [1974](#)). Water was sprayed daily on the combs using a manual pump sprayer to ensure adequate humidity for larval development. The reproductive success in each treatment and in the controls was evaluated by counting all wandering larvae after 16 days.

As we did not intend to compare the results from experiments using different beetle populations, we modified the protocol of the experiments from 2004 as follows. In 2005 we observed that considerable oviposition by female SHB occurred already after 24 h. For testing *B. thuringiensis* var. *tenebrionis* and *B. thuringiensis* var. *kurstaki* adult SHB in both treatments (0.5 ml of 10% *Bt* solution each) and in the

control (0.5 ml H<sub>2</sub>O) ( $N = 8$  replicates each) 2 days were given for feeding, mating and oviposition on the combs [ $5 \times 5$  cm] instead of 5 days as in 2004, before they were removed. All wandering larvae were counted 15 days later, which is an adequate time window for reaching the wandering phase under the given environmental conditions.

Due to the different experimental designs in 2004 and 2005 the numbers of wandering larvae were compared exclusively within each experiment using Kruskal–Wallis ANOVAs, level of significance  $P = 0.05$ . Mann–Whitney  $U$  tests were used as post hoc tests with  $P = 0.0167$  as the Bonferroni–Fischer adjusted level of significance. All statistical analyses were conducted using the program STATISTICA<sup>®</sup>.

## Results

Significant differences were found between the number of wandering larvae after the application of Perizin<sup>®</sup> and B401<sup>®</sup> (Kruskal–Wallis ANOVA:  $H(2, N = 15) = 9.428675, P < 0.0089$ ; Fig. 1). While the number of offspring in the B401<sup>®</sup> treatment ( $204.4 \pm 59.7$ ) was not significantly different from the control ( $194.8 \pm 33.1$ ; Mann–Whitney  $U$  test  $Z = -0.209, P > 0.834$ ), the number of wandering larvae in the Perizin<sup>®</sup> treatment ( $11.4 \pm 17.2$ ) was significantly less compared to both the control and the B401<sup>®</sup> treatment (for both: Mann–Whitney  $U$  test  $Z = -2.611, P < 0.0089$ ). No significant differences were found between the number of wandering larvae from the *B. thuringiensis* var. *tenebrionis* ( $293.6 \pm 144.9$ ) and *B. thuringiensis* var. *kurstaki* ( $395.3 \pm 103.6$ ) treatments and the control ( $345.3 \pm 2.9$ ; Kruskal–Wallis ANOVA:  $H(2, N = 24) = 2.822, P > 0.243$ ).

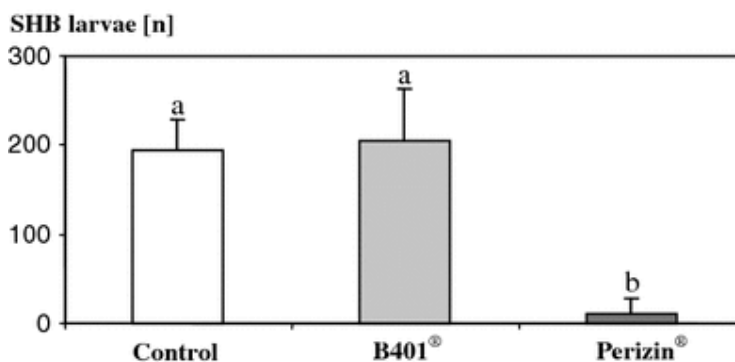


Fig. 1 Number of wandering SHB larvae (mean  $\pm$  SD) after 16 days in controls and in the treatments with Perizin<sup>®</sup> and B401<sup>®</sup>. Different letters indicate significant differences ( $P < 0.0089$ ) using Mann–Whitney  $U$  tests as post hoc tests

## Discussion

Our data give strong support to earlier studies (Elzen et al. 1999) that successful reproduction of SHB can be effectively reduced with coumaphos. While Elzen et al. (1999) used coumaphos as a contact treatment in bottom board trapping devices with Check-Mite+<sup>TM</sup> strips in higher concentrations (10%), we applied it in a lower concentration (3.2%) as a liquid directly on the combs (Perizin<sup>®</sup>). However, the mode of action of coumaphos in SHB remains to be investigated, as it is not clear, whether reproduction was reduced by oviposition failure of female SHB or by mortality of successfully hatched larvae. Regardless of the actual underlying effect and of the mode of application [ingested (this study) or as a contact agent (Elzen et al. 1999)], coumaphos seems to provide an efficient control suppressing SHB reproduction even at lower concentrations.

Despite their active toxin crystal type cryII, we tested Lepidopteran-specific *Bt* products (Jackpot<sup>®</sup> and B401<sup>®</sup>) because the efficiency of strains against coleopteran species cannot be predicted from the crystal morphology (Bernhard et al. [1997](#)). *Bt tenebrionis* and *Bt kurstaki*, which are used to control other beetle pests, were also tested. However, none of the tested *Bt* strains were efficient against SHB, supporting the notion of strain-dependant insect selectivity of *Bt* (Höfte and Whiteley [1989](#); Slaney et al. [1992](#)). Furthermore, it is safe to assume that the tested *Bt* products have not been applied before in South Africa against SHB. So we can also rule out that resistance against these *Bt* strains has been developed as known from other pest species (Tabashnik [1994](#)).

Further investigation would require to test more *Bt* strains, which are used to control other beetle pests. Another promising approach appears to be the development of new strains (Feitelson et al. [1992](#); Naimov et al. [2001](#)), which are specific to *A. tumida*. A further source for *Bt* toxins could be larvae, which died of bacterial infestations, e.g. in beehives and in laboratory rearing programs (Neumann et al. [2001](#); Muerrle and Neumann [2004](#)). According to the high probability to find *Bt* in the soil (Martin and Travers [1989](#)), we also propose to search for new strains, which are naturally infesting SHB in its endemic and new ranges. Such strains could provide an alternative approach for the control of this honeybee pest.

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