

Tendinous tissue properties after short and long-term functional overload: Differences between controls, 12 weeks and 4 years of resistance training

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Tendinous tissue properties after short and long-term functional overload: Differences between controls, 12 weeks and 4 years of resistance training

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3 1 **Tendinous tissue properties after short and long-term functional overload:**
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5 2 **Differences between controls, 12 weeks and 4 years of resistance training**
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35 15 **Short Title:**

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37 16 Tendon adaptation to functional overload
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3 26 **Abstract**
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5 27 **Aim:** The potential for tendinous tissues to adapt to functional overload, especially after
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7 28 several years of exposure to heavy resistance training is largely unexplored. This study
8
9 29 compared the morphological and mechanical characteristics of the patellar tendon and knee-
10
11 30 extensor tendon-aponeurosis complex between young men exposed to long-term (4 years;
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13 31 n=16), short-term (12 weeks; n=15) and no (untrained controls; n=39) functional overload in
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15 32 the form of heavy resistance training. **Methods:** Patellar tendon cross-sectional area, vastus-
16
17 33 lateralis aponeurosis area and quadriceps femoris volume, plus patellar tendon stiffness and
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19 34 Young's modulus, and tendon-aponeurosis complex stiffness, were quantified with MRI,
20
21 35 dynamometry and ultrasonography. **Results:** As expected long-term trained had greater
22
23 36 muscle strength and volume (+58% and +56% vs untrained, both $P<0.001$), as well as a
24
25 37 greater aponeurosis area (+17% vs untrained, $P<0.01$), but tendon cross-sectional area (mean
26
27 38 and regional) was not different between groups. Only long-term trained had reduced patellar
28
29 39 tendon elongation/strain over the whole force/stress range, whilst both short-term and long-
30
31 40 term overload groups had similarly greater stiffness/Young's modulus at high force/stress
32
33 41 (short-term +25/22%, and long-term +17/23% vs untrained; all $P<0.05$). Tendon-aponeurosis
34
35 42 complex stiffness was not different between groups (ANOVA, $P = 0.149$). **Conclusion:**
36
37 43 Despite large differences in muscle strength and size, years of resistance training did not
38
39 44 induce tendon hypertrophy. Both short-term and long-term overload, demonstrated similar
40
41 45 increases in high force mechanical and material stiffness, but reduced elongation/strain over
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43 46 the whole force/stress range occurred only after years of overload, indicating a force/strain
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45 47 specific time-course to these adaptations.
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54 49 **Key Words:** Aponeurosis, Hypertrophy, Muscle, Resistance Training, Stiffness, Tendon
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51 Introduction

52 Tendons are integral to *in vivo* neuromechanical function transmitting skeletal muscle
53 contractile force to the skeleton whilst also optimising the contractile conditions via their
54 viscoelastic properties.^{1,2} The response of tendinous tissue to mechanical loading is of great
55 interest, since it may influence function^{3,4,5,6} and be related to injury incidence.⁷⁻⁹ Impaired
56 tendinous tissues properties are evident in older adults and patient groups¹⁰⁻¹⁴ and are
57 associated with reduced muscle-tendon unit functional capacity.^{3,5,11,15} Functional overload,
58 in the form of resistance exercise is widely recommended for improving musculo-skeletal
59 function of all adults,^{16,17} including older individuals and patients (e.g. osteoarthritis;¹⁸).
60 However our understanding of how tendons alter their properties in response to short-term
61 (weeks-months) and especially long-term (years) loading is limited. Tendinous tissue may
62 exhibit morphological (cross-sectional area), mechanical (stiffness), and material (Young's
63 modulus) adaptations to functional overload,^{19,20} however the magnitude and time-course of
64 these adaptations has not been clearly elucidated.

66 Whilst skeletal muscle tissue has been widely documented to undergo hypertrophy in
67 response to functional overload with resistance training,²¹ the evidence for tendon
68 hypertrophy is equivocal; short-term resistance training studies have reported region specific
69 increases in tendon cross-sectional area²²⁻²⁴ or no change.²⁵⁻²⁷ Explanations for this
70 controversy could be the relatively slow turnover of collagenous tissues,^{28,29} and thus changes
71 in tendon size within the first 14 weeks of resistance training that are on the threshold of what
72 can be accurately detected. A substantially longer exposure to functional overload may
73 provide sufficient time for the accumulation of new tissue and thus demonstrable tendon
74 hypertrophy. However, preliminary cross-sectional studies of long-term functional overload
75 vs untrained controls have used insufficient methods to dispel this conflict, reporting tendon

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3 76 hypertrophy (low resolution ultrasound³⁰) or no hypertrophy for 4 out of 5 tendon sites
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5 77 (limited locations along the tendon with MRI³¹). The potential for aponeurosis hypertrophy in
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7 78 response to resistance training has also had limited research attention.^{32,33}
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12 80 Short-term functional overload with resistance training (up to 14 weeks) utilising high load
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14 81 contractions consistently increases 'free' tendon^{22,24,34,35} and tendon-aponeurosis complex
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16 82 ^{23,25,26,36-38} stiffness. The increased tendon stiffness after short-term resistance training is
17
18 83 typically ascribed to the approximately parallel increases in tendon Young's modulus
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20 84 (material stiffness^{24,35}) rather than substantive changes in tendon size, as mentioned above.
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23 85 However, the potential for further changes in tissue mechanical and material stiffness after
24
25 86 long-term resistance training remains largely unexplored. Preliminary reports include no
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27 87 difference in patellar tendon material stiffness between long-term resistance trained and
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29 88 untrained men³⁰ and no additional changes in Achilles tendon-aponeurosis stiffness from 14
30
31 89 weeks to 18 months of resistance training in elderly women.³⁹ However, both these studies
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33 90 assessed stiffness at different forces/stresses, which confounds comparable measurements of
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35 91 the curvi-linear *in vivo* stress-strain relationship.^{40,41}
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41 93 Whilst existing data have been insufficient to confirm if functional overload results in
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43 94 hypertrophy of tendinous tissues, and whether mechanical and material stiffness continues to
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45 95 adapt with prolonged resistance training, we theorised that (i) the high tendinous tissue loads
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47 96 consequent to the known adaptations of large increases in muscle strength and size after long-
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49 97 term loading²¹, and (ii) prolonged exposure to these high loads would trigger substantial
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51 98 adaptive responses in the tendinous tissues, in order to constrain peak tissue strain within sub-
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53 99 failure physiological limits^{42,43}. The purpose of this study therefore was to compare the
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56 100 morphological and mechanical properties of the patellar tendon (stiffness, Young's modulus,
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3 101 CSA [mean and regional]) and quadriceps femoris tendon-aponeurosis complex (stiffness,
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5 102 muscle volume, vastus lateralis aponeurosis area), between participants exposed to long-term
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7 103 (4 years [LTT]) and short-term (12 weeks [STT]) resistance training and no (untrained
8
9 104 controls [UC]) functional overload. Specific hypotheses were that tendon characteristics
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11 105 would be progressive according to the duration of overload exposure (controls<short-
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13 106 term<long-term), and specifically that long-term overload (resistance training) would be
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15 107 characterised by not only greater muscle size and strength, but also a larger tendon and
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17 108 aponeurosis, higher tendon Young's modulus, as well as greater free tendon and tendon-
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19 109 aponeurosis stiffness.
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111 **Results**

112 **Group Characteristics**

113 Age, height and body mass were similar between UC and STT groups ($P = 0.262$, $P = 0.488$
114 and $P = 0.465$ respectively; Table 1), while LTT were younger, taller and had a larger body
115 mass than UC and STT (all $P \leq 0.003$). Tendon-aponeurosis complex and patellar tendon
116 length were similar between UC and STT ($P = 0.114$ and $P = 0.195$), although LTT had
117 longer tissue lengths than both UC and STT (tendon-aponeurosis complex LTT +6.5% vs UC
118 and +9.2% vs STT; both $P < 0.001$; patellar tendon length LTT +9.8% vs UC $P = 0.006$, and
119 +15.5% vs STT $P = 0.022$).
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120

121 **Muscle-tendon unit size and strength**

122 Maximal voluntary torque (Figure 4a) differed between all three groups, being considerably
123 greater in LTT than UC (+58.1%, $P < 0.001$, ES = 2.90 "very large") and STT (+34.4%, $P <$
124 0.001, ES = 1.66 "large"). STT was also stronger than UC (+17.6%, $P = 0.001$, ES = 1.04
125 "moderate"). QUADSVol (Figure 4b) was considerably larger in LTT than UC (+55.7%, $P <$

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3 126 0.001, ES = 3.55 “very large”) and STT (+46.2%, P < 0.001, ES = 2.83 “very large”),
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5 127 although SST was similar to UC (+7%, P = 0.179, ES = 0.42 “small”). Vastus lateralis
6
7 128 aponeurosis area (Figure 4c) was also larger in LTT than UC (+17.3%, P < 0.001, ES = 1.41
8
9 129 “large”) and STT (+13.5%, P = 0.006, ES = 1.09 “moderate”), but STT was not different to
10
11 130 UC (+3.3%, P = 0.331, ES = “small”). In contrast, patellar tendon mean CSA (Figure 4d) was
12
13 131 similar between groups (ANOVA P = 0.169), and this was also the case for regional patellar
14
15 132 tendon CSA (proximal, middle, distal; ANOVA P > 0.141; Table 2), demonstrating no
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17 133 overall or region specific hypertrophy.
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22 23 135 **Patellar tendon mechanical properties (Table 3)**

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25 136 The patellar tendon force-elongation relationships (Figure 2a) indicated that patellar tendon
26
27 137 elongation at the highest common force level (4200 N, Figure 2b) of LTT was 13.5% less
28
29 138 than UC (2.6 ± 0.5 vs 3.0 ± 0.6 mm, P = 0.063, ES = 0.75 “moderate”) and 15.4% lower than
30
31 139 STT (3.1 ± 0.6 mm, P = 0.048, ES = 0.86 “moderate”), indicating greater stiffness over the
32
33 140 whole force range up to 4200 N for LTT only, whereas STT and UC were similar (P = 0.698,
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35 141 ES = 0.10 “trivial”). However, patellar tendon stiffness measured over a high force range
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37 142 (3360-4200N; Figure 2c) was greater for SST (+24.5%, P = 0.0004, ES = 1.27 “large”) and
38
39 143 LTT (+16.7%, P = 0.021, ES = 0.85 “moderate”) than UC, though similar for LTT and STT
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41 144 (P = 0.287, ES = 0.35 “small”).
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146 Patellar tendon stress-strain relationships (Figure 3a) revealed that at the common stress level
147 of 40 MPa (Figure 3b), patellar tendon strain of LTT ($5.1 \pm 1.0\%$) was 24.4% lower than UC
148 ($6.4 \pm 1.4\%$, P = 0.008, ES = 1.30 “large”) and 19.9% less than STT ($6.7 \pm 1.7\%$, P = 0.006,
149 ES = 0.97 “moderate”), indicating greater material stiffness over the whole stress range up to
150 40 MPa for LTT only, whereas STT and UC were very similar (P = 0.369, ES = 0.17

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3 151 “trivial”). However, patellar tendon Young’s modulus (Figure 3c) derived over a common
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5 152 stress range (32-40 MPa) was greater for both SST (+21.9%, $P = 0.003$, $ES = 1.00$
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7 153 “moderate”) and LTT (+23.3%, $P = 0.002$, $ES = 1.13$ “moderate”) than UC, but was very
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9 154 similar for LTT and STT ($P = 0.855$, $ES = 0.06$ “trivial”).
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156 **Tendon-aponeurosis complex mechanical properties (Table 3)**

157 Force-elongation relationships for the tendon-aponeurosis complex (Figure 4a) showed that
158 at the common force level of 4200 N (Figure 4b), tendon-aponeurosis complex elongation
159 exhibited no main group effect (ANOVA, $P = 0.375$), indicating similar overall tendon-
160 aponeurosis complex elongation in both resistance trained and the untrained group. Likewise,
161 tendon-aponeurosis complex stiffness (Figure 4c) was not statistically different between
162 groups (ANOVA, $P = 0.149$).
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164 **Discussion**

165 The present study compared the morphological, mechanical and material properties of the
166 patellar tendon and knee extensor tendon-aponeurosis complex between young men exposed
167 to long-term (4 years), short-term (12 weeks) and no (untrained controls) functional overload.
168 The main findings were that despite large differences in muscle strength and size, there were
169 modest differences in aponeurosis size, greater in LTT only, and no differences in patellar
170 tendon CSA. Only LTT had reduced elongation/strain over the whole force/stress range up to
171 4200 N/40 MPa, whilst both overload groups had greater patellar tendon stiffness/Young’s
172 modulus at high force/stress than UC. Therefore short-term overload appears sufficient to
173 produce changes in high force mechanical and material stiffness, with no further adaptation
174 with prolonged exposure, but changes in elongation/strain over the whole force range
175 occurred only after years of overload (LTT). Contrary to these differences in tendon

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3 176 mechanics, tendon-aponeurosis complex stiffness and strain were similar between all three
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5 177 groups.
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10 179 **Tendon and aponeurosis size**

11 180 The duration of overload showed progressive differences in muscle strength between the
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13 181 groups (STT +18% and LTT +58% vs UC) and muscle size was also substantially greater
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15 182 after regular long-term loading (LTT +56% vs UC). Despite these substantial differences, and
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17 183 contrary to our hypothesis there were no group differences in patellar tendon mean or
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19 184 regional CSA. The similar patellar tendon CSA of STT than UC might have been expected as
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21 185 there is contrasting evidence for tendon hypertrophy after short-term resistance training (8-14
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23 186 weeks), that has been attributed to limited region-specific hypertrophy and/or slow tendon
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25 187 collagen turnover.^{28,29} Nonetheless we hypothesised that LTT would exhibit greater tendon
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27 188 hypertrophy, due to a combination of (i) their higher loading and stress/strain as a
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29 189 consequence of their substantially greater strength, and (ii) prolonged regular exposure to 4
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31 190 years of these high loads, that might provide sufficient stimulus for the disruption of tissue
32
33 191 homeostasis and time for an accumulation of tendon collagen.⁴⁴ However there were no
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35 192 differences in tendon CSA between LTT and UC despite the substantial differences in muscle
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37 193 strength and size, indicating that tendon size does not adapt in proportion to either muscle
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39 194 size or strength/loading. Our study **utilised** MRI (regarded as the most accurate method^{45,46})
40
41 195 **to assess CSA** along the full tendon length (typically 20 slices; capturing region specific
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43 196 CSA), and images were acquired with sensitive spatial resolution (2 mm thick images, 0 mm
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45 197 gap, pixel size 0.313 x 0.313 mm), as well as careful tendon segmentation performed on each
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47 198 image by the same-blinded investigator. Moreover, with this procedure tendon CSA measures
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49 199 demonstrated very good reliability ($CV \leq 3.5\%$), which provides confidence in the validity of
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51 200 this data. Hitherto, cross-sectional studies of tendon size in long-term vs untrained
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3 201 individuals present conflicting evidence from unconvincing methodologies (low resolution
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5 202 ultrasound³⁰; limited locations along the tendon with MRI³¹). A sole longitudinal study used
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7 203 mixed low and high load training of older females, reporting short-term tendon hypertrophy
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9 204 after 14 weeks, but no further long-term (1.5 years) changes, perhaps due to the surprising
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11 205 lack of long-term strength improvements and thus limited overload.³⁹ In contrast our results
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13 206 provides convincing evidence that long-term (4 years) exposure to high loads (+58% greater
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15 207 strength and +56% greater size) is not a stimulus for tendon hypertrophy.
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21 209 In support of our findings, there is evidence that functional overload via resistance training
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23 210 does not stimulate *in vivo* tendon collagen synthesis,⁴⁷ nor increased concentration of
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25 211 procollagen type 1 N-propeptide, a biomarker of collagen synthesis, in the patellar tendon
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27 212 peritendinous tissue, concomitant with no change in tendon CSA after short-term resistance
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29 213 training.²⁷ In contrast, lower intensity higher volume loading, equivalent to endurance
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31 214 training, might induce *in vivo* tendon collagen synthesis; increased peritendinous tissue
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33 215 procollagen peptide levels,⁴⁸ and uptake of radio-labelled amino acids,⁴⁹ although this is not a
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35 216 consistent finding.^{50,51} Furthermore long term habitual exposure to endurance training or high
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37 217 volumes of low-moderate loading have been found to induce greater tendon size: larger
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39 218 tendon CSA in distance runners vs non-runners^{52,53} and in dominant vs non-dominant limbs
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41 219 after asymmetrical loading⁵⁴. Therefore it may be that high volumes of low/moderate loading
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43 220 may be the important stimulus for tendon hypertrophy, as chronic exposure to high load does
44
45 221 not appear to be a key stimulus.
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51 223 In contrast to tendon size, LTT, but not STT, had a much larger VL aponeurosis area than UC
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53 224 (+17%), demonstrating that aponeurosis size is responsive to the long-term functional
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55 225 overload of the muscle-tendon unit via resistance training. This is coherent with the limited
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3 226 previous reports of greater aponeurosis size post short-term resistance training and in well-
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5 227 trained weightlifters vs untrained.^{32,33} The greater aponeurosis area for LTT vs UC was
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7 228 however substantially smaller than the difference in muscle size (+17 vs +56%), which may
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9 229 be attributable to the greater rate of myofibrillar than connective tissue collagen synthesis in
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11 230 response to resistance exercise.^{28,55}
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15 16 232 **Patellar Tendon Stiffness**

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18 233 In the absence of group differences in patellar tendon CSA, our results for patellar tendon
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20 234 stiffness were attributable to parallel changes in material stiffness, which is in accordance
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22 235 with extensive literature indicating that enhanced material properties are the primary cause of
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24 236 increased mechanical properties.^{19,20} As expected, STT and LTT possessed greater high force
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26 237 patellar tendon stiffness/Young's modulus than UC, which is in accordance with previous
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28 238 short-term resistance training studies^{22,24,30,34,35}; mean change after resistance training
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30 239 ~27/22% for tendon stiffness/modulus.²⁰ Interestingly though LTT had no greater patellar
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32 240 tendon stiffness/Young's modulus than STT.
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38 242 However LTT did demonstrate lesser overall patellar tendon elongation/strain at a common
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40 243 force/stress than STT and UC, and thus greater absolute and material stiffness over the whole
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42 244 force range. Consequently short-term overload was sufficient to produce changes in high
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44 245 force stiffness/Young's modulus, with no further adaptation after prolonged exposure, but
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46 246 changes in elongation/strain over the whole force range occurred only after years of overload
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48 247 (LTT). Given the identical elongation/strain at high forces/stress after STT and LTT, the
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50 248 effect of lower elongation/strain over the whole force range after LTT could only be due to
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52 249 greater stiffness/young's modulus at lower forces/stresses. Qualitatively this difference
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54 250 appears to be due to greater resistance to strain at low stress levels (<10MPa), as the gradients
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3 251 of the stress-strain relationships after the initial most compliant region of tendon deformation
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5 252 were equivalent in STT and LTT. Overall, our data imply that the potential mechanisms
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7 253 (changes to internal structure and/or composition^{56,57}) underpinning an increased high stress
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10 254 tendon modulus, after short-term loading, are likely saturated after 12 weeks leading to a
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12 255 plateau in adaptation, where as low stress-specific material adaptations appeared to continue
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14 256 as these were most pronounced after long-term functional overload. This stress specific time
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16 257 course of tendon adaptation, with decreased strain at high stresses occurring first after short-
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18 258 term loading, followed later by decreased strain at low-stresses after continued long-term
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20 259 loading is a novel finding. More detailed longitudinal investigations are required to verify
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22 260 these results and the mechanisms for this apparent stress specific time-course of adaptation.
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27 262 The validity our findings are reinforced by the thorough measurements of elongation,
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29 263 stiffness and strain; e.g. multiple contractions at a standardised loading rate, duplicate
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31 264 measurement sessions, measurements at identical absolute forces. Representative data were
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33 265 derived across two sessions to yield good inter-test reliability for all stiffness, elongation and
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35 266 stress measurements (CV <10%). In particular our methods avoided the likely bias of higher
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37 267 stiffness measurements for stronger individuals due to: contracting at a higher loading rate
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39 268 during fixed duration ramp contractions; or measuring stiffness/young's modulus at different
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41 269 absolute forces (i.e. relative forces) on the curvilinear force-elongation/stress-strain
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43 270 relationship.
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49 272 **Tendon-Aponeurosis Stiffness**

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52 273 Surprisingly, there were no group differences in tendon-aponeurosis complex mechanical
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54 274 properties despite the presence of much larger muscle and aponeurosis size for LTT. This
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56 275 finding is in contrast to previous reports of greater knee-extensor tendon-aponeurosis
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3 276 complex stiffness assessed with repeated measures pre and post short-term resistance
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5 277 training.^{26,36,37} It is possible that a cross-sectional design lacks the sensitivity to detect
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7 278 relatively modest differences in tendon aponeurosis stiffness. Alternatively, previous studies
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9 279 have commonly measured stiffness at different absolute forces pre and post training, which
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11 280 may have accentuated the scale of this training adaptation. The measurement of tendon-
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13 281 aponeurosis complex stiffness is also not considered to be as robust as that of 'free tendon'.⁵⁸
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16 282 This is because it may reflect not only tendon-aponeurosis deformation, but also the active
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18 283 state of the muscle fibres parallel to the aponeurosis,⁵⁹ as well as the fact that single site
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20 284 measures of aponeurosis deformation with 2-D imaging may provide only a crude index of
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22 285 the stiffness of a 3-D structure. Further investigations could incorporate three-dimensional
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24 286 imaging techniques (ultrasound^{60,61} or MRI^{62,63}) that can capture the complex bi-axial
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27 287 deformation of the muscle and aponeurosis along the length of the tendon-aponeurosis
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29 288 complex.
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34 290 In summary, the greater **quadriceps femoris** strength and volume in LTT (+58 and +56% vs
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36 291 UC) was associated with modest increases in **vastus lateralis** aponeurosis area (+17% vs UC),
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38 292 but not matched by a larger **patellar tendon** cross-sectional area, indicating that long-term
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40 293 functional overload via high force resistance training does not lead to extramuscular tendon
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42 294 hypertrophy. Short- and long-term overload groups had similar, but greater patellar tendon
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44 295 stiffness/Young's modulus at high force/stress than UC, but only LTT had reduced
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46 296 elongation/strain over the whole force/stress range up to 4200 N/ 40 MPa that was
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48 297 attributable to changes at lower force/stress. Therefore we found evidence for a stress specific
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50 298 time-course of adaptation in the patellar tendon with short-term overload sufficient to
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52 299 produce changes in high force mechanical and material stiffness, with no further adaptation
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55 300 to prolonged exposure, but increased low stress stiffness only occurring after years of
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3 301 overload (LTT).
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7 303 **Materials and Methods**
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10 304 **Participants**

11 305 Seventy young men provided written informed consent before completing this study, which
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14 306 was approved by the Loughborough University Ethical advisory committee, and was
15
16 307 conducted according to the principles expressed in the Declaration of Helsinki. All
17
18 308 participants were healthy and free from musculoskeletal injury with no previous history of
19
20 309 tendon pathology. The untrained control group (UC, n = 39) had no lower body resistance
21
22 310 training experience for >18 months. The short-term trained group (STT, n = 15) were
23
24 311 measured post 12-weeks of supervised resistance training. The long-term trained (LTT, n =
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26 312 16) group had 4.0 ± 0.8 (mean \pm SD) years of systematic heavy-resistance training
27
28 313 experience (~3 x wk of quadriceps sessions; typical exercises were squat, lunge, step-up, leg
29
30 314 press). LTT participants typically reported some nutritional supplement consumption
31
32 315 (predominantly whey protein and creatine), although none declared illegal performance-
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34 316 enhancing substance use.
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41 318 **Experimental Design**
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43 319 Participants visited the laboratory for a familiarisation session, (STT were familiarised pre-
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45 320 training) and two duplicate measurement sessions that were averaged to improve the
46
47 321 reliability of the measurements. Participants were seated in a custom-built isometric strength-
48
49 322 testing chair and completed a series of maximal voluntary contractions (MVCs) and ramp
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51 323 voluntary contractions of the knee extensors as well as knee flexor MVCs of the dominant leg
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53 324 (preferred kicking leg). MVCs established maximal voluntary torque (MVT) and ramp
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55 325 contractions were performed to permit tissue stiffness estimation. Knee joint torque was
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3 326 recorded throughout contractions. Knee flexor surface electromyography was recorded
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5 327 during knee flexor MVCs and knee extensor ramp contractions. All contractions were
6
7 328 performed with equivalent resting joint angle configurations. Ultrasound images of the vastus
8
9 329 lateralis and patellar tendon were recorded throughout the ramp contractions to assess tissue
10
11 330 elongation. Measurement sessions were performed at a consistent time of the day (± 2 hours),
12
13 331 separated by at least 2 days and started between 12:00–19:00 p.m. Participants were
14
15 332 instructed not to participate in strenuous physical activity, consume alcohol/refrain from
16
17 333 caffeine consumption in the 36/6 hours before measurement sessions. All participants were
18
19 334 instructed to maintain their habitual physical activity and diet throughout the study. For the
20
21 335 SST group, post-measurement sessions one and two took place 3-5 and 6-8 days following
22
23 336 the last training session. Magnetic resonance imaging (MRI) was performed to assess
24
25 337 quadriceps femoris muscle, vastus lateralis aponeurosis, and patellar tendon size. Participants
26
27 338 were instructed to refrain from strenuous physical activity in the 24 hours prior to the MRI
28
29 339 scan. For the STT group, MRI was conducted 2-3 days after the final training session and
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31 340 prior to post measurement sessions.
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342 **Short-term trained (STT) Group: Training Intervention**

343 Training sessions were completed three times per week on the same apparatus and with
344 equivalent joint angles as used for measurement sessions. After a brief warm-up of sub-
345 maximal contractions of both legs, participants completed four sets of ten unilateral isometric
346 knee-extensor contractions of each leg, with sets alternating between dominant and non-
347 dominant legs until 4 sets per leg had been completed. Contractions were sustained at
348 75%MVT, with 2 s rest between each contraction. In order to control the torque rise and hold
349 times, participants were presented with a target torque trace 2 s before every contraction and
350 instructed to match this target, which increased torque linearly from rest to 75% MVT over 1

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3 351 s before holding a plateau at 75%MVT for a further 3 s. MVCs were performed at the start of
4
5 352 each training week to re-establish MVT and prescribe training torques.
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9 354 **Torque Measurement**

10 355 Participants were positioned in an isometric strength-testing chair with resting knee and hip
11
12 356 joint angles of $\sim 115^\circ$ and $\sim 126^\circ$ (180° = full extension), respectively. **The resting joint angle**
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15
16 357 **configurations were determined from digitisation of sagittal plane video during pilot work.**
17

18 358 Adjustable straps were tightly fastened across the pelvis and shoulders to prevent extraneous
19
20 359 movement. An ankle strap (35 mm width reinforced canvas webbing) was placed $\sim 15\%$ of
21
22 360 tibial length (distance from lateral malleolus to knee joint space) above the medial malleolus,
23
24 361 and positioned perpendicular to the tibia and in series with a calibrated S-Beam strain gauge
25
26 362 (Force Logic, Berkshire, UK). The analogue force signal was amplified ($\times 370$; A50
27
28 363 amplifier, Force Logic UK) and sampled at 2,000 Hz using an A/D converter (Micro 1401;
29
30 364 CED, Cambridge, UK) and recorded with Spike 2 computer software (CED). In offline
31
32 365 analysis, force signals were low-pass filtered at 500 Hz using a fourth order zero-lag
33
34 366 Butterworth filter, gravity corrected by subtracting baseline force, and multiplied by lever
35
36 367 length, the distance from the knee joint space to the centre of the ankle strap, to calculate
37
38 368 torque values.
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44 370 **Knee Flexor Electromyography (EMG)**

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46 371 Surface EMG recordings over the biceps femoris (BF) and semitendinosus (ST) were made
47
48 372 with a wireless EMG system (Trigno; Delsys Inc, Boston, MA) were made during knee
49
50 373 flexor MVCs and knee extensor ramp contractions. Following preparation of the skin
51
52 374 (shaving, abrading and cleansing with alcohol) single differential Trigno standard EMG
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54 375 sensors (1 cm inter electrode distance; Delsys Inc, Boston, MA) were attached over each
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3 376 muscle using adhesive interfaces. Sensors were positioned parallel to the presumed frontal
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5 377 plane orientation of the underlying muscle fibres at 45% of thigh length (distance from the
6
7 378 greater trochanter to the lateral knee joint space) measured from the popliteal crease. EMG
8
9
10 379 signals were amplified at source (x300; 20-450 Hz bandwidth) before further amplification
11
12 380 (overall effective gain x 909) and sampled at 2000 Hz via the same A/D converter and
13
14 381 computer software as the force signal, to enable data synchronization. In offline analysis,
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16 382 EMG signals were corrected for the 48 ms delay inherent to the Trigno EMG system.
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384 **Knee Extension and Flexion Maximal Voluntary Contractions**

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23 385 Following a brief warm-up (3 s contractions at 50% [x3], 75% [x3] and 90% [x1] of
24
25 386 perceived maximal), participants performed 3-4 MVCs and were instructed to either 'push as
26
27 387 hard as possible' (knee extension) or 'pull as hard as possible' (knee flexion) for 3-5 s and
28
29 388 rest \geq 30 s. A horizontal cursor indicating the greatest torque obtained within the session was
30
31 389 displayed for biofeedback and verbal encouragement was provided during all MVCs. The
32
33 390 highest instantaneous torque recorded during any MVC was defined as MVT. During knee
34
35 391 flexor MVCs EMG amplitude was calculated as the root mean square (RMS) of the filtered
36
37 392 EMG signal of the BF and ST over a 500 ms epoch at knee flexion MVT (250 ms either side)
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39 393 and averaged across the two muscles to give knee flexor EMG_{MAX}.
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395 **MRI measurement of Muscle Tendon Unit Morphology and Moment Arm**

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47 396 T1-weighted MR (1.5 T Signa HDxt, GE) images of the dominant leg (thigh and knee) were
48
49 397 acquired in the supine position at a knee angle of 163° (due to constraints in knee coil size)
50
51 398 and analysed using OsiriX software (Version 6.0, Pixmeo, Geneva, Switzerland). Using a
52
53 399 receiver 8-channel whole body coil, axial images (time of repetition/time to echo 550/14,
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55 400 image matrix 512 x 512, field of view 260 x 260 mm, pixel size 0.508 x 0.508 mm, slice
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3 401 thickness 5 mm, inter-slice gap 0 mm) were acquired from the anterior superior iliac spine to
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5 402 the knee joint space in two overlapping blocks. Oil filled capsules placed on the lateral side
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7 403 of the thigh allowed alignment of the blocks during analysis. The quadriceps femoris (QF)
8
9 404 muscles (vastus lateralis [VL] vastus intermedius [VI], vastus medialis, and rectus femoris)
10
11 405 were manually outlined in every third image (i.e. every 1.5 cm) starting from the most
12
13 406 proximal image in which the muscle appeared. The volume of each muscle was calculated
14
15 407 using cubic spline interpolation (GraphPad Prism 6, GraphPad Software, Inc.). Total QF
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17 408 volume (QUADSvol) was the sum of the individual muscle volumes.
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23 410 As previously described, the deep aponeurosis of the vastus lateralis muscle was defined as
24
25 411 the visible dark black segment between the VL and VI muscles in the thigh MRI images.³²
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27 412 VL aponeurosis width was defined as the **transverse length** (cm) of the **deep aponeurosis**
28
29 413 (distinct black segment) **between the vastus lateralis and vastus intermedius**, traced manually
30
31 414 on every third image (i.e. every 1.5 cm), starting in the most distal image where the
32
33 415 aponeurosis was visible. Aponeurosis width measures were plotted against the longitudinal
34
35 416 aponeurosis length (distance between most proximal and distal image where the aponeurosis
36
37 417 was visible [cm]). **The surface area of VL aponeurosis was calculated as** the area under a
38
39 418 spline curve fitted to the aponeurosis width **and length plot, and termed** VL aponeurosis area
40
41 419 **(Figure 5).**
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47 421 Immediately after thigh imaging, a lower extremity knee coil was used to acquire axial (time
48
49 422 of repetition/time to echo 510/14, image matrix 512 x 512, field of view 160 x 160 mm, pixel
50
51 423 size 0.313 x 0.313, slice thickness 2 mm, inter-slice gap 0 mm) and sagittal images (time of
52
53 424 repetition/time to echo 480/14, image matrix 512 x 512, field of view 160 x 160 mm, pixel
54
55 425 size 0.313 x 0.313, slice thickness 2 mm, inter-slice gap 0 mm) of the knee joint. Contiguous
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3 426 axial images spanned patellar tendon length, which during analysis, were reconstructed to be
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5 427 aligned perpendicular to the line of action of the patellar tendon: straight line from the
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7 428 tendons posterior fibres insertion at the patellar apex to the posterior fibres tibial insertion.
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9 429 Images spanned from 2 cm superior to the patellar apex to 2 cm inferior to the tendon tibial
10
11 430 insertion Patellar tendon CSA (mm^2) was measured on each contiguous image along the
12
13 431 tendons length (first image where the patellar was no longer visible to the last image before
14
15 432 the tibial insertion). Images, viewed in greyscale, were sharpened and the perimeter manually
16
17 433 outlined. A spline curve was fitted to the tendon CSA values from each image and the
18
19 434 average of the spline equated to mean patellar tendon CSA (patellar tendon mean CSA). The
20
21 435 average of the spline CSA's measured over proximal, middle and distal thirds was defined as
22
23 436 proximal, mid and distal patellar tendon region CSA. Sagittal plane images were used to
24
25 437 determined patellar tendon moment arm, the perpendicular distance from the patellar tendon
26
27 438 line of action to the tibio-femoral contact point, which was the midpoint of the distance
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29 439 between the tibio-femoral contact points of the medial and lateral femoral condyles.
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36 441 **Ramp Contractions for Determination of Tissue Stiffness**

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38 442 Tissue stiffness was derived from synchronous recordings of torque and tissue elongation
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40 443 (see below, corrected for passive tissue displacement via video recording of knee joint
41
42 444 changes) during isometric knee extension ramp contractions. Participants completed two sub-
43
44 445 maximal (~75% MVT) practice ramp contractions prior to five maximal attempts with 90 s
45
46 446 rest between contractions. Prior to each ramp contraction participants were shown a target
47
48 447 torque-time trace on a computer monitor that increased at a constant gradient ($50 \text{ Nm}\cdot\text{s}^{-1}$
49
50 448 loading rate) from zero up to MVT. They were instructed to match the target trace as closely
51
52 449 as possible for as long as possible (i.e. up to MVT), and real-time torque was displayed over
53
54 450 the target torque-time trace for feedback. The preceding knee extensor MVCs and sub-
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3 451 maximal contractions were considered sufficient to elicit tissue preconditioning. The three
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5 452 most suitable ramp contractions, according to highest peak torque, the closeness to the target
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7 453 loading rate and ultrasound image clarity, were analysed and measurements averaged across
8
9 454 these three contractions.
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14 456 **Measurement of Tissue Elongation**

16 457 Video images from two ultrasound machines and one video camera were captured to obtain
17
18 458 tissue and knee joint displacements during ramp contractions. An ultrasound probe (7.5 MHz
19
20 459 linear array transducer, B-mode, scanning width 60mm and depth 50 mm; Toshiba Power
21
22 460 Vision 6000, SSA-370A; Otawara-Shi, Japan) was fitted into a custom made high-density
23
24 461 foam cast that was strapped to the lateral aspect of the thigh with the mid-point of the probe
25
26 462 positioned at ~50 % thigh length. The probe was aligned so the fascicles inserting into the
27
28 463 vastus lateralis (VL) muscle deep aponeurosis could be visualized at rest and during
29
30 464 contraction. An echo-absorptive marker (multiple layers of transpore medical tape) was
31
32 465 placed beneath the ultrasound probe to provide a reference for any probe movement over the
33
34 466 skin. Another ultrasound probe (5-10 MHz linear array transducer, B-mode, scanning width
35
36 467 92 mm and depth 65 mm, EUP-L53L; Hitachi EUB-8500) was fitted into a custom made
37
38 468 high-density foam cast that was held firmly over the anterior aspect of the knee with the
39
40 469 probe aligned longitudinal to the patellar tendon such that the patellar apex and insertion of
41
42 470 the posterior tendon fibres at the tibia could be visualized at rest and throughout the
43
44 471 contraction. The ultrasound machines were interfaced with the computer collecting torque
45
46 472 data in Spike 2 and the video feeds were recorded synchronously with torque using Spike 2
47
48 473 video capture at 25 Hz. During off-line analysis tissue elongation was tracked frame-by-
49
50 474 frame using public-domain (www.cabrillo.edu/~dbrown/tracker) semi-automatic video
51
52 475 analysis software: Tracker, version 4.86. The distance measured over the surface of the skin
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3 476 between the echo-absorptive marker on the VL and the tibial tuberosity defined resting
4
5 477 tendon-aponeurosis complex length. VL fascicle deep aponeurosis cross point displacement
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7 478 relative to the skin marker provided a measure of tendon-aponeurosis elongation. Patellar
8
9 479 tendon elongation was determined by the longitudinal displacement of the patella apex and
10
11 480 the tendon tibial insertion. The distal insertion of the patellar tendon was not monitored for
12
13 481 the purpose of estimating overall tendon-aponeurosis displacement. To enable correction of
14
15 482 tissue displacement due to joint angle changes during ramp contractions individual ratios of
16
17 483 tissue displacement relative to joint angular displacement ($\text{mm}/^\circ$) were obtained from passive
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19 484 movements (i.e. plotting the tissue displacement-knee joint angle relationship). This ratio was
20
21 485 used to determine tissue displacement resulting from knee angle change during ramp
22
23 486 contractions, which was subsequently subtracted from total measured displacement.
24
25 487 Corrections were only applied to aponeurosis displacement. Tendon elongation under passive
26
27 488 conditions was deemed negligible. Passive movements were conducted prior to the ramp
28
29 489 contractions. Participants were instructed to completely relax as their knee was moved
30
31 490 through 90 to 130°. During passive movements and ramp contractions, knee joint angle
32
33 491 (angle between visible markers placed on the greater trochanter, lateral knee joint space and
34
35 492 lateral malleolus) was derived from sagittal plane video recorded using a camera mounted on
36
37 493 a tripod positioned (1.5 m) perpendicular to the strength-testing chair. The video camera was
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39 494 interfaced with a computer and recorded using spike 2 video capture at 25 Hz
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41 495 (simultaneously with force, EMG, and ultrasound images during the ramp contractions) and
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43 496 analysed via Tracker software.
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51 52 **Calculation of Tendon Force**

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54 499 Patellar tendon force was calculated by dividing external absolute knee extensor torque by
55
56 500 the patellar tendon moment arm length. Direct measures of moment arm were acquired at
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3 501 rest from MRI images as indicated above (*MRI measurement*). Due to constraints in the size
4
5 502 of the knee coil, sagittal images were acquired in an extended knee position (~163°).
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7 503 Moment arm length for any specific knee angle measured at rest or during ramp contraction
8
9 504 was estimated from previously published data fitted with a quadratic function⁶⁴ scaled to each
10
11 505 participant's measured moment arm length at 163°. Absolute internal knee extensor torque
12
13 506 was given by summing net knee extension torque and the estimated knee flexor co-
14
15 507 contraction torque. Antagonist knee flexor torque was estimated by expressing the average
16
17 508 knee flexor EMG amplitude (RMS 50 ms moving window) during ramp contractions relative
18
19 509 to the knee flexor EMG_{MAX} and multiplying by the knee flexor MVT (assuming a linear
20
21 510 relationship between EMG amplitude and torque). During analysis, torque and EMG
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23 511 amplitude were down-sampled to 25 Hz to match the ultrasound video frequency.
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513 **Calculation of Tissue Stiffness and Tendon Young's Modulus**

514 For each of the three best ramp contractions analysed, tendon-aponeurosis (corrected for
515 passive tissue displacement) and patellar tendon elongation was plotted against total tendon
516 force (corrected for antagonist force). Force-elongation plots were fitted with a second-order
517 polynomial. Tendon-aponeurosis and patellar tendon stiffness was calculated as the gradient
518 (Δ tendon force [N]/ Δ elongation [mm]; N.mm⁻¹) of the respective force-strain curve over 80-
519 100% (3360-4200N) of an absolute tendon force (4200N) that corresponded to the lowest
520 common force level attained by all participants during ramp contractions. Tendon stress was
521 obtained by dividing tendon force by mean tendon CSA. Tendon strain was the percentage
522 tendon displacement relative to the resting tendon length. Resting PT length was defined as
523 the distance between the patella apex and tibial insertion as measured prior to the ramp
524 contractions. A patellar tendon stress-strain curve was plotted and patellar tendon Young's
525 modulus (GPa) calculated as the slope (Δ tendon stress [MPa]/ Δ tendon strain [%]) of the

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3 526 stress-strain curve derived over 80-100% of an absolute common stress (40 MPa). The
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5 527 stiffness and Young's modulus measures derived from each of the three ramp contractions
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7 528 analysed was averaged to give each individuals representative values.
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11 530 **Reproducibility and Statistical Analysis**

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14 531 The reproducibility of tendinous tissue measurements over the duplicate test sessions was
15
16 532 calculated for the whole cohort (test 1 vs test 2) as within participant co-efficient of variation
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18 533 ($CV_w, \%; [SD/mean]*100$): elongation [0-4200 N] of the patellar tendon (7.9%) and tendon-
19
20 534 aponeurosis complex (9.6%); stiffness [3360-4200 N] of the patellar tendon (9.9%) and
21
22 535 tendon-aponeurosis complex (8.6%); patellar tendon strain (8.0%) and Young's modulus
23
24 536 (9.0%). Patellar tendon CSA measurements were highly reproducible, as indicated by the co-
25
26 537 efficient of variation (CV_w) of repeat measurements 12 weeks apart for a sub-sample of the
27
28 538 untrained control group (n=14): mean (2.7%), proximal (3.0%), mid (3.1%) and distal
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30 539 (3.5%).
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36 541 Muscle strength and tissue mechanical/material properties measured during the duplicate
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38 542 laboratory sessions were averaged to produce criterion values for statistical analysis. An a
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40 543 priori significance level of $P < 0.05$ was set for all statistical tests which were performed using
41
42 544 SPSS Version 20.0 (IBM Corp., Armonk, NY). Descriptive data are presented as mean \pm
43
44 545 standard deviation (SD) and percentage differences in the group means are given in the text.
45
46 546 The influence of group (UC, STT, LTT) on all muscle and tendinous tissue variables was
47
48 547 examined by univariate ANOVA. Main group effects were followed by least significant
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50 548 difference (LSD) post-hoc paired comparisons to delineate between group differences; Holm-
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52 549 Bonferroni corrections were applied to LSD P-values, and between group Hedges g effect
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3 550 size (ES) was calculated.⁶⁵ Effect size magnitude was classified as <0.2= “trivial”; 0.2-0.6 =
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5 551 “small”; >0.6-1.2 = “moderate”; >1.2-2.0 = “large”; >2.0 = “very large”.⁶⁶
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9 552

10 553 **Acknowledgements**

11 554 None to declare
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15 556 **Competing Interests**

16 557 None to declare
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22
23 559 Arthritis Research UK Centre for Sport, Exercise and Osteoarthritis.
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26 561 **References**

- 27
28
29 562 1. Wilson A, Litchwark G: The anatomical arrangement of muscle and tendon enhances
30 563 limb versatility and locomotor performance. *Philos Trans R Soc Lond B. Biol Sci*, 368:
31 564 1540-1553, 2011.
32
33
34 565 2. Roberts TJ: Contribution of elastic tissues to the mechanics and energetics of muscle
35 566 function during movement. *J Exp Biol*, 219: 266-275, 2017.
36
37
38 567 3. Onambélé GL, Narici MV, Maganaris CN: Calf muscle-tendon properties and postural
39 568 balance in old age. *J Appl Physiol*, 100: 2047-2056, 2006.
40
41
42 569 4. Stafilidis S, Arampatzis A: Muscle-tendon unit mechanical and morphological properties
43 570 and sprint performance. *J Sports Sci*, 25: 1035-1046, 2007.
44
45
46 571 5. Karamanidis K, Arampatzis A, Mademli L: Age-related deficit in dynamic stability
47 572 control after forward falls is affected by muscle strength and tendon stiffness. *J*
48 573 *Electromyogr and Kinesiol*, 18: 980-989, 2008.
49
50
51
52
53
54
55
56
57
58
59
60

- 574 6. Mayfield DL, Cresswell AG, Litchwark GA. Effects of series elastic compliance on
575 muscle force summation and the rate of force rise. *J. Exp. Biol*, 219: 3261-3270, 2016.
- 576 7. Kvist M: Achilles tendon injuries in athletes. *Sports Med*, 18: 173-201, 1994.
- 577 8. Archambault JM, Wiley JP, Bray RC: Exercise loading of tendons and the development
578 of overuse injuries. A review of current literature. *Sports Med* 20: 77-89, 1995.
- 579 9. Roberts TJ, Konow N: How tendons buffer energy dissipation in muscle. *Exerc Sport Sci*
580 *Rev*, 41: 186-193, 2013.
- 581 10. Arya S, Kulig K: Tendinopathy alters mechanical and material properties of the Achilles
582 tendon. *J Appl Physiol*, 108: 670-675, 2010.
- 583 11. Matschke V, Jones JG, Lemmey AB, Maddison PJ, Thom JM: Patellar tendon properties
584 and lower limb function in rheumatoid arthritis and ankylosing spondylitis versus healthy
585 controls: a cross-sectional study. *Scientific World Journal*, 2013:514743, 2013.
- 586 12. Stenroth L, Peltonen J, Cronin NJ, Sipilä S, Finni T: Age-related differences in Achilles
587 tendon properties and triceps surae muscle architecture in vivo. *J Appl Physiol*, 113:
588 1537-1544, 2012.
- 589 13. Nielsen RH, Couppé C, Jensen JK, Olsen MR, Heinemeier KM, Malfait F, Symoens S,
590 De Paepe A, Schjerling P, Magnusson SP, Remvig L, Kjaer M: Low tendon stiffness and
591 abnormal ultrastructure distinguish classic Ehlers-Danlos syndrome from benign joint
592 hypermobility syndrome in patients. *FASEB J*, 28: 4668-4676, 2014.
- 593 14. Couppé C, Svensson RB, Kongsgaard M, Kovanen V, Grosset JF, Snorgaard O, Bencke
594 J, Larsen JO, Bandholm T, Christensen TM, Boesen A, Helmark IC, Aagaard P, Kjaer M,
595 Magnusson SP. Human Achilles tendon glycation and function in diabetes. *J Appl*
596 *Physiol*, 120: 130-137, 2016.
- 597 15. Stenroth L, Sillanpää E, McPhee JS, Narici MV, Gapeyeva H, Pääsuke M, Barnouin Y,
598 Jean-Hogrel JY, Butler-Browne G, Bijlsma A, Meskers CG, Maier AB, Finni T, Sipilä S.

- 1
2
3 599 Plantar flexor muscle–tendon properties are associated with mobility in healthy older
4
5 600 adults. *J Gerontol A Biol Sci Med Sci*, 70: 996-1002, 2015.
6
7 601 16. Liu CJ, Latham NK: Progressive resistance strength training for improving physical
8
9 602 function in older adults. *Cochrane Database Sys Rev*, 3: 1-272, 2009.
10
11 603 17. Ratamess NA, Alvar B., Evetoch T., Housh TJ, Kibler W, Kraemer WJ, Triplett:
12
13 604 Progression Models in Resistance Training for Healthy Adults. *Med Sci Sports Exerc*, 41:
14
15 605 687-708, 2009.
16
17
18 606 18. Yanan L, Youxin S, Shaoqing C, Yingjie Z, Ziyi Z, Changyan L, Meili L, Feiwen L,
19
20 607 Shuzhen L, Zhen H, Yiru W, Lu S, Wenting W, Zhengxuan Z, Xu W, Naixi Z: The
21
22 608 effects of resistance exercise patients with knee osteoarthritis: a systematic review and
23
24 609 meta-analysis. *Clin Rehabil* 30: 947-959, 2016.
25
26
27 610 19. Bohm S, Mersmann F, Arampatzis A: Human tendon adaptation in response to
28
29 611 mechanical loading: a systematic review and meta-analysis of exercise intervention
30
31 612 studies on healthy adults. *Sports Med Open* 1: 7, 2015.
32
33
34 613 20. Wiesinger HP, Kösters A, Müller E, Seynnes OR: Effects of increased loading on in vivo
35
36 614 tendon properties: a systematic review. *Med Sci Sports Exerc*, 47: 1885-1895, 2015.
37
38
39 615 21. Folland JP, Williams AG: The adaptations to strength training: morphological and
40
41 616 neurological contributions to increased strength. *Sports Med*, 37:145-168. 2007
42
43 617 22. Kongsgaard M, Reitelseder S, Pedersen TG, Holm L, Aagaard P, Kjaer M, Magnusson
44
45 618 SP: Region specific patellar tendon hypertrophy in humans following resistance training.
46
47 619 *Acta Physiol*, 191: 111-121, 2007.
48
49 620 23. Arampatzis A, Karamanidis K, Albracht K: Adaptational responses of the human Achilles
50
51 621 tendon by modulation of the applied cyclic strain magnitude. *J Exp Biol*, 210: 2743-2753,
52
53 622 2007.
54
55
56
57
58
59
60

- 1
2
3 623 24. Seynnes OR, Erskine RM, Maganaris CN, Longo S, Simoneau EM, Grosset JF, Narici
4
5 624 MV: Training-induced changes in structural and mechanical properties of the patellar
6
7 625 tendon are related to muscle hypertrophy but not to strength gains. *J Appl Physiol* 107:
8
9 626 523-530, 2009.
- 11 627 25. Arampatzis A, Peper A, Bierbaum S, Albracht K: Plasticity of human Achilles tendon
12
13 628 mechanical and morphological properties in response to cyclic strain. *J Biomech*, 43:
14
15 629 3073-3079, 2010.
- 17 630 26. Kubo K, Ikebukuro T, Maki A, Yata H, Tsunoda N: Time course of changes in the human
18
19 631 Achilles tendon properties and metabolism during training and detraining in vivo. *Eur J*
20
21 632 *Appl Physiol*, 112: 2679-2691, 2012.
- 23 633 27. Bloomquist K, Langberg H, Karlsen S, Madsgaard S, Boesen M, Raastad T: Effect of
24
25 634 range of motion in heavy load squatting on muscle and tendon adaptations. *Eur J Appl*
26
27 635 *Physiol*, 113: 2133-2142, 2013.
- 29 636 28. Smith K, Rennie MJ: New approaches and recent results concerning human-tissue
30
31 637 collagen synthesis. *Curr Opin Clin Nutr Metab Care*, 10: 582-590, 2007.
- 33 638 29. Heinemeier KM, Schjerling P, Heinemeier J, Magnusson SP, Kjaer M: Lack of tissue
34
35 639 renewal in human adult Achilles tendon is revealed by nuclear bomb (14)C. *FASEB J*,
36
37 640 27: 2074-2079, 2013.
- 39 641 30. Seynnes OR, Kamandulis S, Kairaitis R, Helland C, Campbell, EL, Brazaitis M,
40
41 642 Skurvydas A, Narici MV: Effect of androgenic-anabolic steroids and heavy strength
42
43 643 training on patellar tendon morphological and mechanical properties. *J Appl Physiol*, 115:
44
45 644 84-89, 2013.
- 47 645 31. Fukutani A, Kurihara T: Tendon cross-sectional area is not associated with muscle
48
49 646 volume. *J Appl Biomech*, 31: 176-180, 2015.
- 51
52
53
54
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56
57
58
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- 1
2
3 647 32. Abe T, Kumagai K, Bemben MG: Muscle aponeurosis area is hypertrophied and normal
4
5 648 muscle. *J. Trainol*, 1: 23-27, 2012.
6
7 649 33. Wakahara T, Ema R, Miyamoto N, Kawakami Y: Increase in vastus lateralis aponeurosis
8
9 650 width induced by resistance training: implications for a hypertrophic model of pennate
10
11 651 muscle. *Eur J Appl Physiol*, 115: 309-316, 2015.
12
13 652 34. Malliaras P, Kamal B, Nowell A, Farley T, Dhamu H, Simpson V, Morrissey D,
14
15 653 Langberg H, Maffuli N, Reeves ND: Patellar tendon adaptation in relation to load-
16
17 654 intensity and contraction type. *J Biomech*, 46: 1893-1899, 2013.
18
19 655 35. McMahon GE, Morse CI, Burden A, Winwood K, Onambélé-Pearson GL: The
20
21 656 manipulation of strain, when stress is controlled, modulates in vivo tendon mechanical
22
23 657 properties but not systematic TGF-B1 levels. *Physiol Rep*, 1: e00091, 2013.
24
25 658 36. Kubo K, Kanehisa H, Fukunaga T: Effect of different duration isometric contractions on
26
27 659 tendon elasticity in human quadriceps muscles. *J Physiol*, 536: 639-655, 2001.
28
29 660 37. Kubo K, Ikebukuro T, Yaeshima K, Yata H, Tsunoda N, Kaneshisa H: Effects of static
30
31 661 and dynamic training in the stiffness and blood volume of tendon in vivo. *J Appl Physiol*
32
33 662 106: 412-417, 2009.
34
35 663 38. Bohm S, Mersmann F, Tettke M, Kraft M, Arampatzis A: Human Achilles tendon
36
37 664 plasticity in response to cyclic strain: effect of strain rate and duration. *J Exp Biol*, 217:
38
39 665 4010-4017, 2014.
40
41 666 39. Epro G, Mierau A, Doerner J, Luetkens JA, Scheef L, Kukuk GM, Boecker H, Maganaris
42
43 667 CN, Brüggemann GP, Karamanidis K: The Achilles tendon is mechanosensitive in older
44
45 668 adults: adaptations following 14 weeks versus 1.5 years of cyclic strain exercise. *J Exp*
46
47 669 *Biol*, 2220: 1008-1018, 2017.
48
49 670 40. Maganaris CN, Paul JP: Load-elongation characteristics of in vivo human tendon and
50
51 671 aponeurosis. *J Exp Biol*, 203: 751-756, 2000.
52
53
54
55
56
57
58
59
60

- 1
2
3 672 41. Maganaris CN, Paul JP: Tensile properties of the in vivo human gastrocnemius tendon. J
4
5 673 Biomech 35: 1639-1646, 2002.
6
7 674 42. Matson A, Konow N, Miller S, Konow PP, Roberts TJ: Tendon material properties vary
8
9 675 and are interdependent among turkey hindlimb muscles. J Exp Biol, 215: 3552-3558,
10
11 676 2012.
12
13 677 43. LaCroix AS, Duenwald-Kuehl SE, Lakes RS, Vanderby R Jr: Relationship between
14
15 678 tendon stiffness and failure: a metanalysis. J Appl Physiol, 115: 43-51, 2013.
16
17 679 44. Svensson RB, Heinemeier KM, Couppé C, Kjaer M, Magnusson SP: Effect of aging and
18
19 680 exercise on the tendon. J Appl Physiol, 121: 1237-1246, 2016.
20
21 681 45. Couppé C, Svensson RB, Sødring-Elbørnd V, Hansen P, Kjaer M, Magnusson SP.
22
23 682 Accuracy of MRI technique in measuring tendon cross-sectional area. Clin Physiol Funct
24
25 683 Imag, 34: 237-241, 2014.
26
27 684 46. Kruse A, Stafilidis S, Tilp M. Ultrasound and magnetic resonance imaging are not
28
29 685 interchangeable to assess the Achilles tendon cross sectional-area. Eur J Appl Physiol,
30
31 686 117: 77-82, 2017.
32
33 687 47. Sullivan BE, Carroll CC, Jemiolo B, Trappe SW, Magnusson SP, Døssing S, Kjaer M,
34
35 688 Trappe TA: Effect of acute resistance exercise and sex on human patellar tendon
36
37 689 structural and regulatory mRNA expression. J Appl Physiol, 106: 468-475, 2008.
38
39 690 48. Langberg H, Skovgaard D, Karamouzis M, Bulow J, Kjaer M: Metabolism and
40
41 691 inflammatory mediators in the peritendinous space measured by microdialysis during
42
43 692 intermittent isometric exercise in humans. J Physiol, 515: 919-927, 1999.
44
45 693 49. Miller BF, Olesen JL, Hansen M, Døssing S, Crameri RM, Welling RJ, Langberg H,
46
47 694 Fyvbjerg A, Kjaer M, Babraj JA, Smith K, Rennie MJ: Coordinated collagen and muscle
48
49 695 protein synthesis in human patella tendon and quadriceps muscle after exercise. J Physiol,
50
51 696 567: 1021-1033, 2005.
52
53
54
55
56
57
58
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- 1
2
3 697 50. Dideriksen K, Sindby AK, Krogsgaard M, Schjerling P, Holm L, Langberg H: Effect of
4
5 698 acute exercise on patella tendon protein synthesis and gene expression. Springerplus, 2:
6
7 699 109, 2013.
8
9
10 700 51. Heinemeier KM, Bjerrum SS, Schjerling P, Kjaer M: Expression of extracellular matrix
11
12 701 components and related growth factors in human tendon and muscle after acute exercise.
13
14 702 Scand J Med Sci Sports, 23: e150-e161, 2013.
15
16 703 52. Kongsgaard M, Aagaard P, Kjaer M, Magnusson SP: Structural Achilles tendon
17
18 704 properties in athletes subjected to different exercise modes and in Achilles tendon rupture
19
20 705 patients. J Appl Physiol, 99: 1965-1971, 2005.
21
22
23 706 53. Wiesinger HP, Rieder F, Kösters A, Müller E, & Seynnes OR: Are sport-specific profiles
24
25 707 of tendon stiffness and cross-sectional area determined by structural or functional
26
27 708 integrity? *PLoS One*, 11: e0158441, 2016.
28
29
30 709 54. Couppé C, Kongsgaard M, Aagaard P, Hansen P, Bojsen-Moller J, Kjaer M, Magnusson
31
32 710 SP: Habitual loading results in tendon hypertrophy and increased stiffness of the human
33
34 711 patellar tendon. J Appl Physiol, 105: 805-810, 2008.
35
36 712 55. Babraj JA, Cuthbertson DJ, Smith K, Langberg H, Miller B, Krogsgaard MR, Kjaer M,
37
38 713 Rennie MJ: Collagen synthesis in human musculoskeletal tissues and skin. Am J Physiol
39
40 714 Endocrinol Metab, 289: E864-869, 2005.
41
42
43 715 56. Kjaer M, Jørgensen NR, Heinemeier K, Magnusson SP: Exercise and regulation of bone
44
45 716 and collagen tissue biology. Prog Mol Biol Transl Sci, 135: 259-291, 2015.
46
47 717 57. Buchanan CI, Marsh R: Effects of exercise on the biomechanical, biochemical and
48
49 718 structural properties of tendons. Comp Biochem Physiol A Mol Integr Physiol, 133:
50
51 719 1101-1107, 2002.
52
53
54
55
56
57
58
59
60

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2
3 720 58. Seynnes OR, Bojsen-Møller J, Albracht K, Arndt A, Cronin NJ, Finni T, Magnusson SP:
4
5 721 Ultrasound-based testing of tendon mechanical properties: a critical evaluation. *J Appl*
6
7 722 *Physiol*, 118: 133-141, 2015.
8
9
10 723 59. Lieber R, Leonard M, Brown-Maupin C: Effects of muscle contraction on the load-strain
11
12 724 properties of frog aponeurosis and tendon. *Cells Tissues Organs*, 166: 48-54, 2000.
13
14 725 60. Farris DJ, Trewartha G, McGuigan MP, Litchwark GA: Differential strain patterns of
15
16 726 human Achilles tendon determined in vivo with freehand three-dimensional ultrasound
17
18 727 imaging. *J Exp Biol*, 216: 594-600, 2013.
19
20
21 728 61. Raiteri BJ, Cresswell AG, Lichtwark GA: Three-dimensional geometrical changes of the
22
23 729 human tibialis anterior muscle and its central aponeurosis measured with three-
24
25 730 dimensional ultrasound during isometric contractions. *PeerJ*, 4: e2260, 2016.
26
27
28 731 62. Iwanuma S, Akagi R, Kurihara T, Ikegawa S, Kanehisa H, Fukunaga T, Kawakami Y:
29
30 732 Longitudinal and transverse deformation of human Achilles tendon induced by isometric
31
32 733 plantar flexion at different intensities. *J Appl Physiol*, 6: 1615-1621, 2011.
33
34 734 63. Reeves ND, Cooper G: Is human Achilles tendon deformation greater in regions where
35
36 735 cross-sectional area is smaller? *J Exp Biol*, 220: 1634-1642, 2017.
37
38
39 736 64. Kellis E, Baltzopoulos V: In vivo determination of the patella tendon and hamstrings
40
41 737 moment arms in adult males using videofluoroscopy during submaximal knee extension.
42
43 738 *Clin Biomech (Bristol Avon)*, 14: 118-124, 1999.
44
45 739 65. Lakens D: Calculating and reporting effect sizes to facilitate cumulative science: a
46
47 740 practical primer for t-tests and ANOVAs. *Front Psychol*, 4: 86, 2013.
48
49
50 741 66. Hopkins WG, Marshall SW, Batterham AM, Hanin J: Progressive statistics for studies in
51
52 742 sports medicine and exercise science. *Med Sci Sports Exerc*, 41: 3-13, 2009.
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3 745 **Physiological Relevance**
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5 746 The potential capacity for tendinous tissue to adapt to long-term functional overload via
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7 747 heavy resistance training is unclear. Our cross-sectional data show that short-term (12 weeks)
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9 748 and long-term (~ 4 years) resistance trained males had similar patellar tendon mechanical and
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11 749 material (Young's modulus) stiffness, with both resistance-trained groups having stiffer
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13 750 tendons than untrained controls. In contrary, neither resistance-trained group had larger
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15 751 tendon size. Furthermore, no differences in tendon-aponeurosis complex stiffness were
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17 752 observed between groups, despite substantially greater muscle strength and size in the long-
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19 753 term resistance trained group, which was also accompanied by a larger muscle-aponeurosis
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21 754 size.
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Table 1. Descriptive characteristics of the participants.

	UC	STT	LTT	ANOVA (P)
Sample size, n =	39	15	16	
Age, years	25 ± 2	25 ± 2	22 ± 2	<0.001
Height, cm	176 ± 6	175 ± 8	183 ± 6	0.001
Body mass, kg	72 ± 9	70 ± 9	90 ± 10	<0.001
Tendon-aponeurosis complex length, mm	336 ± 16	328 ± 17	358 ± 18	<0.001
Patellar tendon length, mm	47.7 ± 5.5	45.1 ± 5.5	52.1 ± 5.9	0.005
Patellar tendon moment arm, mm	43.8 ± 2.7	44.8 ± 3.1	45.8 ± 2.5	0.054

770 Data are mean ± SD.

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Table 2. Regional patellar tendon cross-sectional area (mm^2).

Region	UC (n=39)	STT (n=15)	LTT (n=16)	ANOVA (P)
Proximal	93.0 \pm 9.9	92.0 \pm 13.5	98.4 \pm 13.1	0.216
Mid	104.4 \pm 12.6	97.0 \pm 14.3	104.9 \pm 13.3	0.146
Distal	110.4 \pm 17.9	101.6 \pm 14.3	112.1 \pm 15.7	0.141

Data are mean \pm SD.

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3 805 **Table 3.** Summary of group differences in strength (maximal voluntary torque, MVT), muscle tendon
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5 806 unit size and tissue (tendon-aponeurosis complex and patellar tendon [PT]) stiffness between long-
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7 807 term resistance trained (LTT), short-term resistance trained (STT) and untrained control (UC) groups.
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<i>Function</i>	
MVT, Nm	LTT > STT > UC
<i>Muscle-Tendon Unit size</i>	
QUADSvol, cm ³	LTT > STT & UC
VL Aponeurosis Area, cm ²	LTT > STT & UC
PT CSA, mm ²	-
<i>Indices of Tissue Stiffness</i>	
PT elongation (0-4200 N), mm	LTT < STT & UC
PT stiffness (3360-4200 N), N.mm ⁻¹	LTT & STT > UC
PT strain (0-40 MPa), %	LTT < STT & UC
PT Young's modulus (32-40 MPa), GPa	LTT & STT > UC
Tendon-aponeurosis elongation (0-4200 N), mm	-
Tendon-aponeurosis stiffness (3360-4200 N), N.mm ⁻¹	-

37 808 Significant group differences groups: greater (>) or less (<) than. No
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39 809 difference (-). PT CSA: both mean and regional CSA measures.
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3 819 **Figure Legends**

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5 820 **Figure 1.** Group comparisons: Isometric knee extension maximal voluntary torque (a),
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7 821 Quadriceps femoris muscle volume (QUADSvol, b), Vastus Lateralis (VL) aponeurosis area
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9 822 (c) and Patellar tendon mean cross-sectional area (CSA, d) for untrained control (UC, n =
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11 823 39), short-term resistance trained (STT, n = 15) and long-term resistance trained (LTT, n =
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13 824 16) groups. Data are mean \pm SD. Bold numbers are between groups hedges g effect size.
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15 825 Post-hoc tests: Least significant difference Holm-Bonferroni corrected P-values. *P<0.05,
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17 826 †P<0.01, ‡P<0.001.
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23 828 **Figure 2. (a)** Relationships between patellar tendon force (N) and elongation (mm) in the
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25 829 untrained control (UC, n = 37), short-term resistance trained (STT, n = 15) and long-term
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27 830 resistance trained (LTT, n = 15) groups. Curves show the group mean relationship. Data
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29 831 points correspond to within group average values for the elongation at 10% intervals of group
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31 832 mean maximal voluntary tendon force, plotted up to 80% (highest common level achieved
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33 833 during ramp contractions). Error bars indicate the within-group standard deviation for force
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35 834 (y-axis bar) and elongation (x-axis bar). **(b) And (c)** Group comparisons of the patellar
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37 835 tendon elongation at the common force level of 4200 N, and patellar tendon stiffness
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39 836 (gradient of curves in **a** over 80-100% of the highest common force level [3360-4200 N]).
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41 837 Bars are mean \pm SD. Bold numbers are the between groups hedges g effect size. Post-hoc
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43 838 tests: Least significant difference Holm-Bonferroni corrected P-values. *P<0.05, ‡P<0.001.
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49 840 **Figure 3. (a)** Relationships between patellar tendon stress (MPa [N.mm²]) and strain (%) in
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51 841 the untrained control (UC, n = 37), short-term resistance trained (STT, n = 15) and long-term
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53 842 resistance trained (LTT, n = 15) groups. Curves show the group mean relationship. Data
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55 843 points correspond to within group average values for the strain at 10% intervals of group
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3 844 mean maximal voluntary tendon stress, plotted up to 80% (highest common level achieved
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5 845 during ramp contractions). Error bars indicate the within-group standard deviation for stress
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7 846 (y-axis bar) and strain (x-axis bar). **(b) And (c)** Group comparisons of the PT strain at the
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9 847 common stress level of 40 MPa, and PT Young's modulus (gradient of curves in **a** over 80-
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11 848 100% common stress level [32-40 MPa]). Bars are mean \pm SD. Bold numbers are the
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13 849 between groups hedges g effect size. Post-hoc tests: Least significant difference Holm-
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15 850 Bonferroni corrected P-values. † $P < 0.01$.

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21 852 **Figure 4.** **(a)** Relationships between patellar tendon force (N) and tendon-aponeurosis
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23 853 complex elongation (mm) in the untrained control (UC, $n = 37$), short-term resistance trained
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25 854 (STT, $n = 14$) and long-term resistance trained (LTT, $n = 16$) groups. Curves show the group
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27 855 mean relationship. Data points correspond to within group average values for the strain at
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29 856 10% intervals of group mean maximal voluntary tendon force, plotted up to 80% (highest
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31 857 common level achieved during ramp contractions). Error bars indicate the within-group
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33 858 standard deviation for force (y-axis bar) and strain (x-axis bar). **(b) And (c)** Group
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35 859 comparisons of the tendon-aponeurosis complex elongation at the common force level of
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37 860 4200 N, and tendon-aponeurosis complex stiffness (gradient of curves in **a** over 80-100%
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39 861 common force level [3360-4200 N]). Bars are mean \pm SD.

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45 863 **Figure 5.** Measurement of the surface area of the vastus lateralis deep aponeurosis. On axial
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47 864 magnetic resonance images at 1.5 cm intervals along the longitudinal aponeurosis length:
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49 865 distance from the most distal image to the most proximal where the aponeurosis was visible
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51 866 (a), the transverse length of the distinct visible black segment between the vastus lateralis
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53 867 (VL) and vastus intermedius (VI) muscles was traced manually and defined as aponeurosis
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55 868 width (b). The aponeurosis width measures on each image were plotted against aponeurosis
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869 length and the area under a cubic spline curve fitted through the data points defined the VL
870 aponeurosis area (c).

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Figure 1.

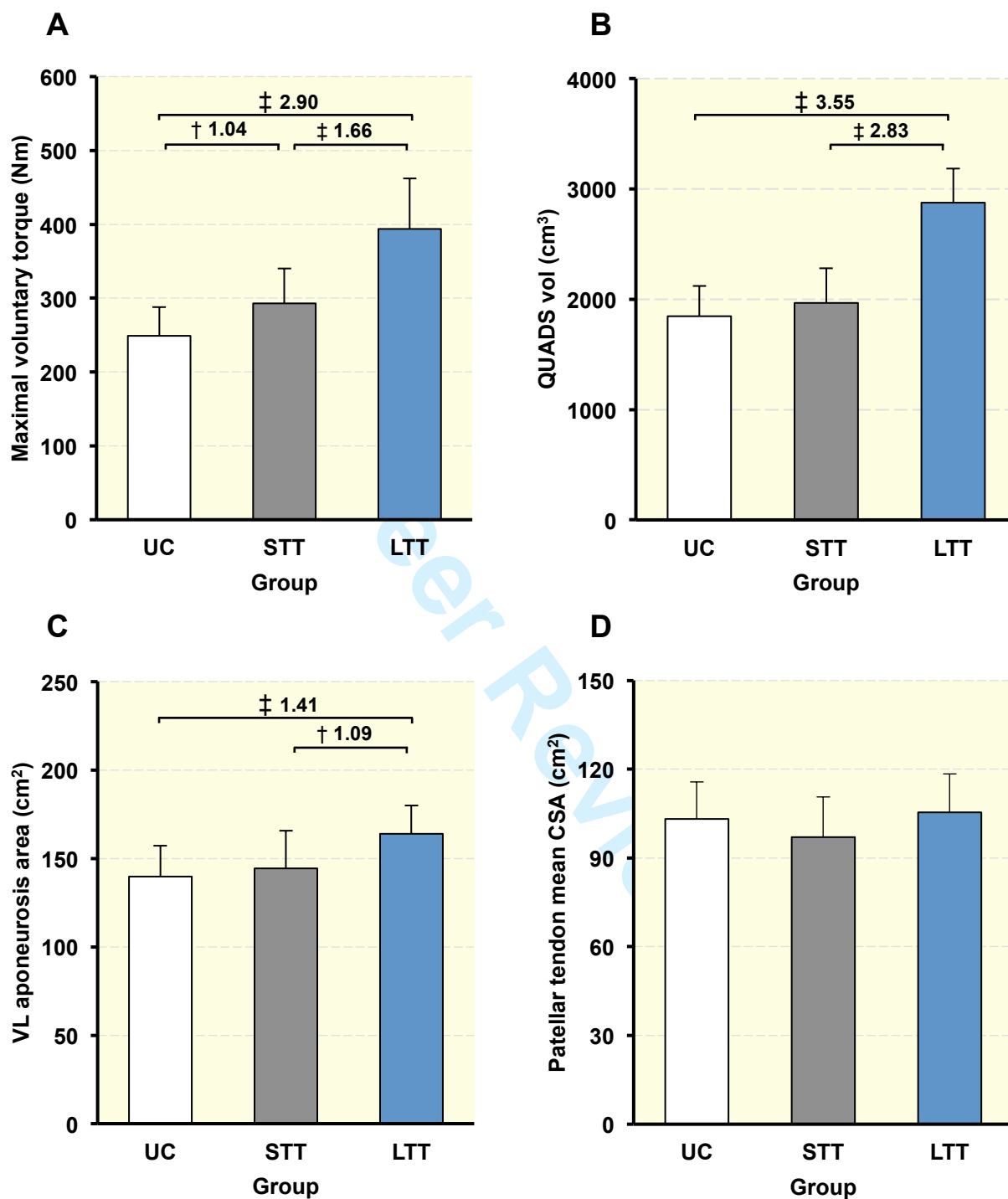
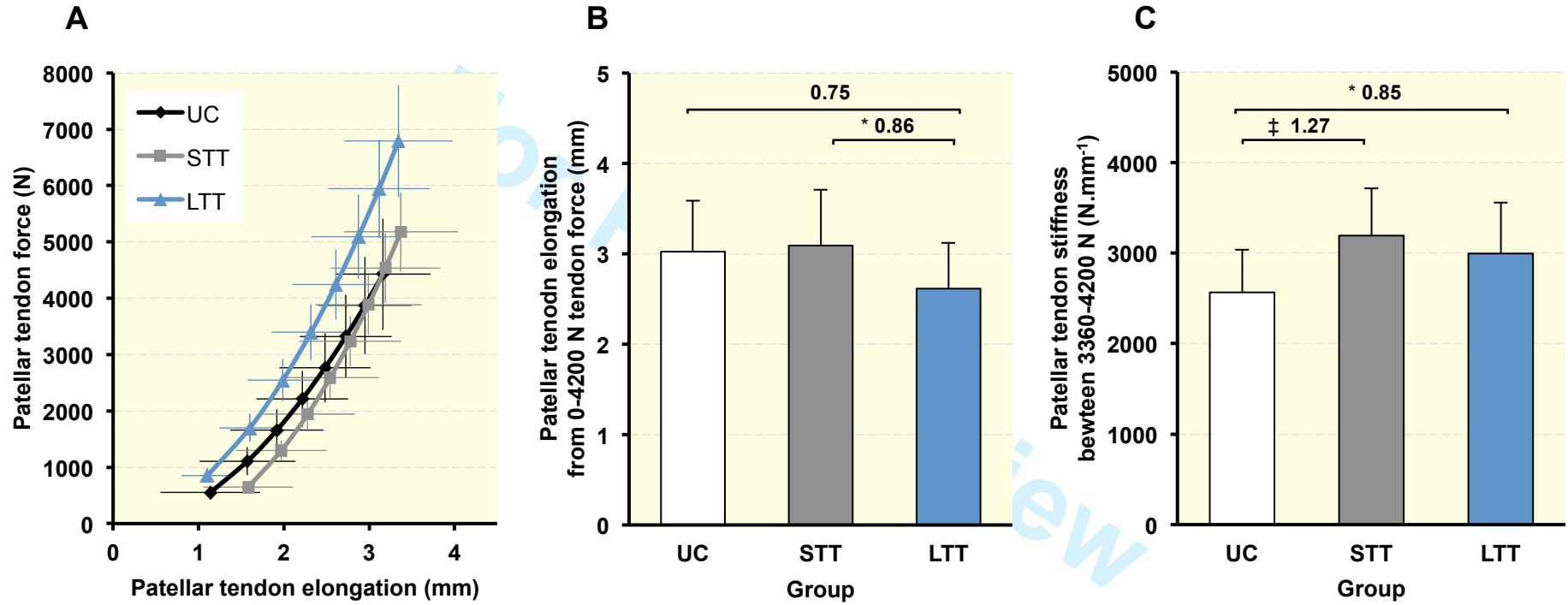


Figure 2.



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Figure 3.

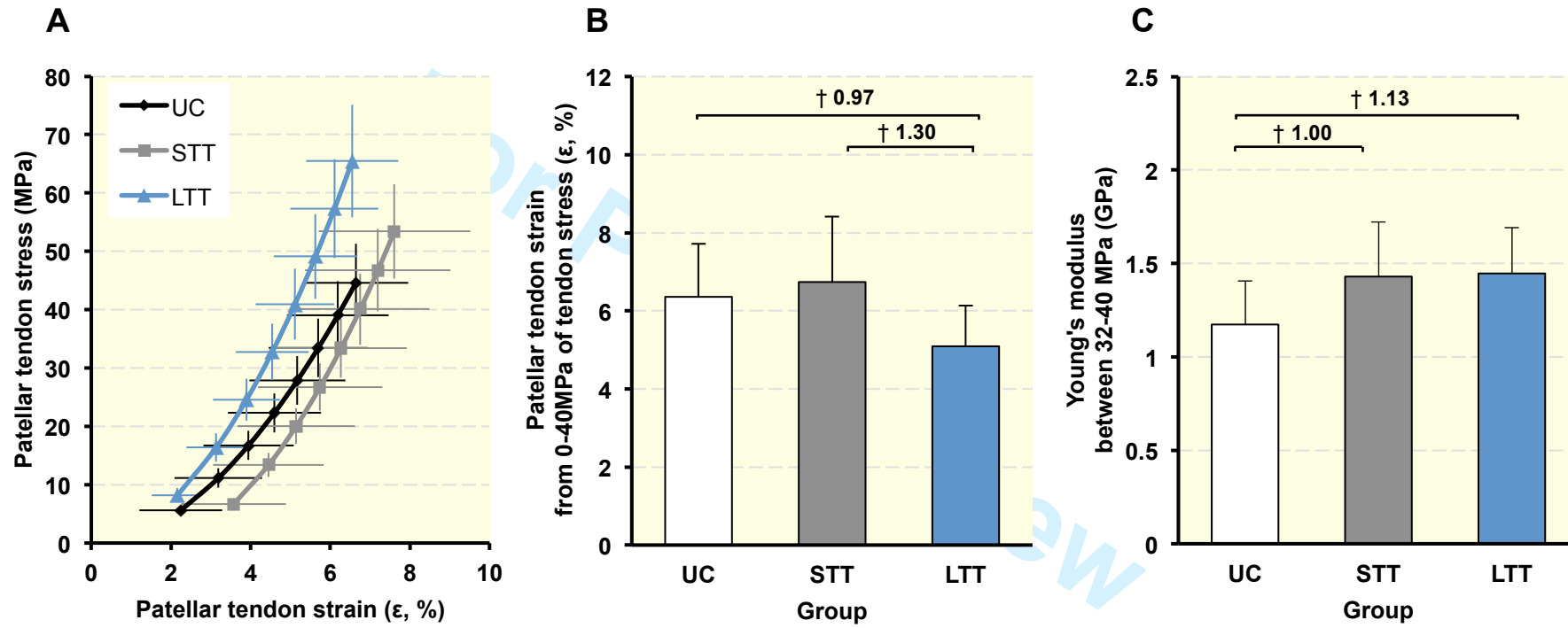
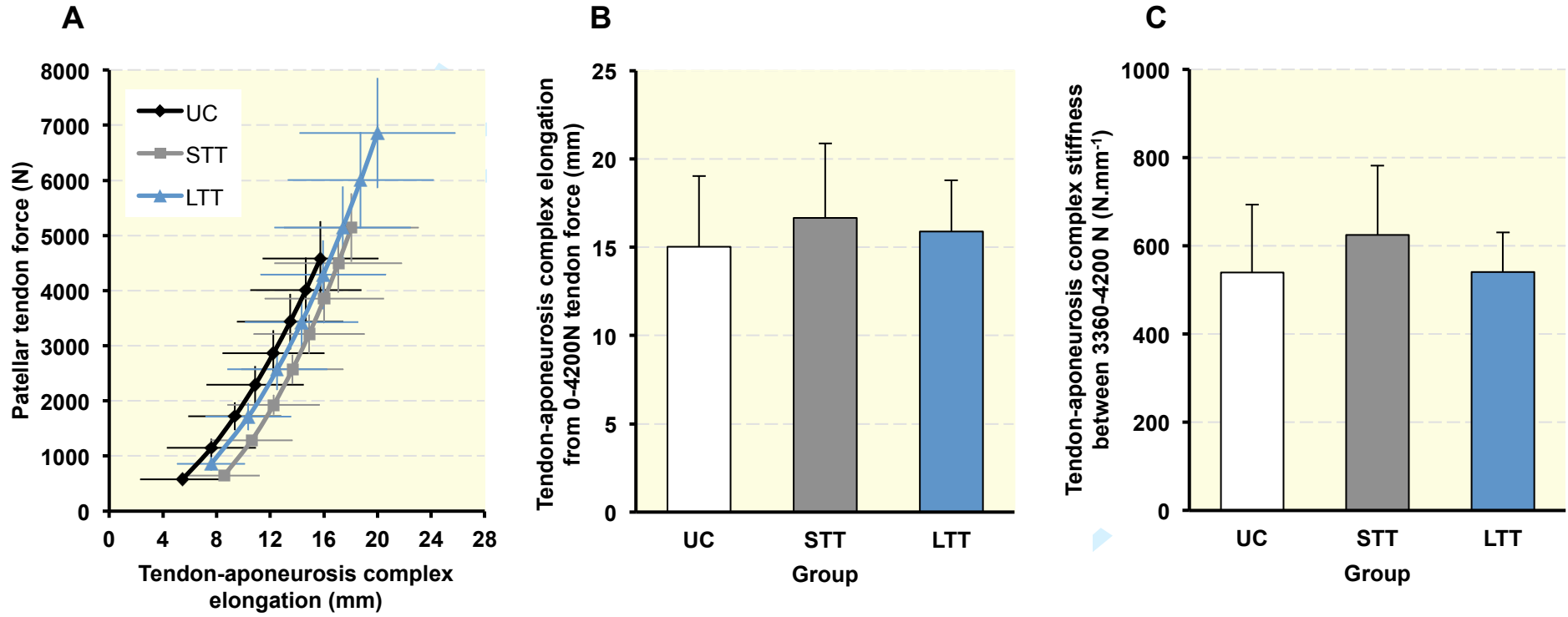
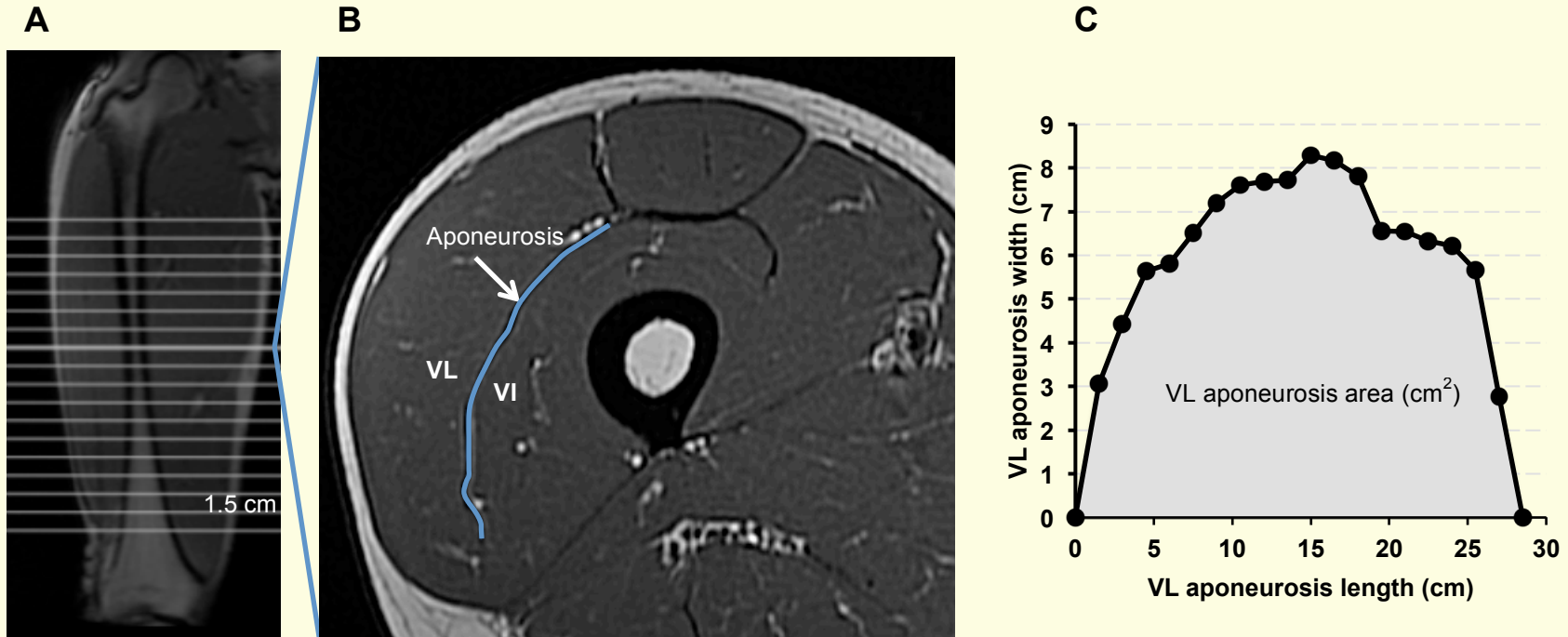


Figure 4.



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Figure 5.



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