### Journal of Mind and Medical Sciences

Volume 2 | Issue 2

Article 4

#### 2015

## Toxicological Analysis of Some Drugs of Abuse in Biological Samples

Anne Marie Ciobanu Carol Davila University of Medicine and Pharmacy

Daniela Luiza Baconi Carol Davila University of Medicine and Pharmacy, daniela baconi@yahoo.com

Cristian Bălălău Carol Davila University of Medicine and Pharmacy

Carolina Negrei Carol Davila University of Medicine and Pharmacy

Miriana Stan Carol Davila University of Medicine and Pharmacy

See next page for additional authors

Follow this and additional works at: http://scholar.valpo.edu/jmms Part of the <u>Medicine and Health Sciences Commons</u>

#### **Recommended** Citation

Ciobanu, Anne Marie; Baconi, Daniela Luiza; Bălălău, Cristian; Negrei, Carolina; Stan, Miriana; and Bârcă, Maria (2015) "Toxicological Analysis of Some Drugs of Abuse in Biological Samples," *Journal of Mind and Medical Sciences*: Vol. 2 : Iss. 2, Article 4. Available at: http://scholar.valpo.edu/jmms/vol2/iss2/4

This Review Article is brought to you for free and open access by ValpoScholar. It has been accepted for inclusion in Journal of Mind and Medical Sciences by an authorized administrator of ValpoScholar. For more information, please contact a ValpoScholar staff member at scholar@valpo.edu.

## Toxicological Analysis of Some Drugs of Abuse in Biological Samples

#### Authors

Anne Marie Ciobanu, Daniela Luiza Baconi, Cristian Bălălău, Carolina Negrei, Miriana Stan, and Maria Bârcă

# **Toxicological analysis of some drugs of abuse in biological samples**

# Anne Marie Ciobanu<sup>1</sup>, <u>Daniela Baconi<sup>2</sup></u>, Cristian Bălălău<sup>3</sup>, Carolina Negrei<sup>2</sup>, Miriana Stan<sup>2</sup>, Maria Bârcă<sup>1</sup>

 <sup>1</sup> Carol Davila University of Medicine and Pharmacy, Faculty of Pharmacy, Medicines Control
 <sup>2</sup> Carol Davila University of Medicine and Pharmacy, Faculty of Pharmacy, Department of Toxicology
 <sup>3</sup> Carol Davila University of Medicine and Pharmacy, Faculty of General Medicine, St. Pantelimon Hospital Corresponding author: Daniela Luiza Baconi, e-mail: <u>daniela\_baconi@yahoo.com</u>

**Running title:** Toxicological analysis for drugs abuse **Keywords**: toxicological analysis, drugs of abuse, screening test, GCMS, HPLC, LCMS

www.jmms.ro 2015, Vol. II (issue 2): 108- 127. Date of submission: 2015-03-11; Date of acceptance: 2015-06-17

#### Abstract

Consumption of drugs of abuse is a scourge of modern world. Abuse, drug addiction and their consequences are one of the major current problems of European society because of the significant repercussions in individual, family, social and economic level. In this context, toxicological analysis of the drugs of abuse in biological samples is a useful tool for: diagnosis of drug addiction, checking an auto-response, mandatory screening in some treatment programs, identification of a substance in the case of an overdose, determining compliance of the treatment.

The present paper aims to address the needs of healthcare professionals involved in drugs addiction treatment through systematic presentation of information regarding their toxicological analysis. Basically, it is a tool that help you to select the suitable biological sample and the right collecting time, as well as the proper analysis technique, depending on the purpose of analysis, pharmacokinetic characteristics of the drugs of abuse, available equipment and staff expertise.

#### Introduction

Consumption of drugs of abuse is a scourge of modern world, regardless the fact that we are talking about high-risk drugs or about the off label use of certain authorized medicinal products. It is a large-scale, multifactorial, dynamic phenomenon which affects all the age groups, but predominantly the one between 14 and 35 years old. Abuse and drug addiction, as well as their consequences are one of the major problems in the current European society (1- 3). This is due to their significant repercussions in individual, family and social level (crime, social marginalization, and death due to overdose or by suicide) as well as in the economic level: dependence treatment costs but also the costs of the therapy for viral and bacterial infections associated with the intravenous consumption (AIDS, HVC, or reappearance of TBC) (4, 5). Given the major risks associated with the drugs of abuse, their analysis in biological samples is a useful tool for:

- Initial diagnosis of drug addiction
- Checking an auto-response, a declaration
- Mandatory *screening* in some treatment programs
- Screening as a method of tracking drug effects over time
- Identification of the substance in case of an overdose
- Determination of treatment compliance

#### Discussion

*Toxicological analysis* represents the whole analytical processes through which the presence of a toxic substance in an analysed sample is determined. It includes the physicochemical methods for the isolation, identification and quantification of toxic substances in the air, water, soil, food, delict objects and organic products for the prevention or diagnosis of intoxications (6-9).

*Drugs of abuse* are those substances which, as a result of pleasant effects they produce, are used for other purposes than the ones they are intended to. For example, the therapeutic effect in the

benzodiazepines case or the industrial use in volatile solvents case. Drugs of abuse are those substances whose possession, transport or storage is restricted by law, due to potential harmful effect on the consumer and include materials manufactured under license, as well as illicit products manufactured in clandestine laboratories or natural products (10- 14).

The methodology of toxicological analyses of drugs of abuse is developed based on:

- Type of sample used
- Scope of analysis
- Pharmacokinetic features and biotransformation of the illicit substance
- Available equipment and reagents
- Staff expertise
- Costs.

*Biological samples.* Depending on the purpose of the analysis, the substances of abuse may be determined from different biological samples.

**Blood/plasma:** first choice for the quantitative determination of drugs; therapeutic levels in the blood are low but, when they are consumed abusively, the concentrations may be 2-3 times higher.

**Urine:** first choice for screening of drugs of abuse. It is available in sufficient quantity and substances or metabolites are present in relatively high concentrations.

**Hair:** it is used for the determination of the history of an abuse substance consumption. Detection is possible at10-14 days to 90 days after ingestion.

Saliva: it is used for the screening of drugs of abuse consumed within the last 24 hours.

**Meconium:** reveals maternal history of drugs of abuse consumption in the last 20 weeks of pregnancy and allow the choice of therapy for mother and new-born.

Breast milk: it is used for the determination of the exposure extent of the infant to drugs of abuse.

For example, in table no. 1 are presented the chromatographic techniques used for the analysis of methadone cited in the literature, grouped according to biological samples in which the determination is carried out (15- 16).

BIOLOGICAL SAMPLE	GC-MS	HPLC	LC-MS
Plasma		Schmidt N. et al.,1992 Foster D. et al, 2001 Foster D. et al, 2003 Hallinan R. et al, 2006	Liang H.R. et al., 2004 Whittington D. et al., 2004 Etter M.L. et al., 2005 Lehotay D.C. et al., 2005 Rook E.J. et al., 2005 Quintela O. et al., 2006 Jenkins R. et al., 2006 Shakleya D.M. et al, 2007
Urine	Larson M. et al, 2009 Moore C. et al., 2001	Cheng YF. et al., 1999	Dams R. et al, 2003 Shakleya D.M. et al, 2010
Breast milk			Choo R.E. et al, 2007 Jansson L.M. et al, 2007
Meconium			Choo R. E. et al, 2005
Umbilical cord	Nikolaou P.D. et al., 2008		de Castro A. et al., 2009
Placenta		Nanovskaya T.N. et al., 2004	
Saliva			Concheiro M. et al, 2010 Rodriguez Rosasa M.E. et al., 2003 Ortelli D. et al., 2000
Hair	Moeller M.R. et al., 1993 Lucas A.C.S. et al., 2000 Girod C. et al., 2001		Kintz P. et al., 2009
Sweating	Brunet B.R. et al., 2008		

Table 1. Chromatographic analysis of methadone according to the biological samples

The detection time of abuse substances is varying in different biological samples. For example, drugs of abuse are detected in saliva within minutes after consumption and in urine only after 4-8 hours (17-19).

Biological samples matrix is very complex and contains other endogenous or exogenous substances in addition to substances of interest. This is the reason that, in most cases, is necessary to use specific isolation procedures (20- 24).

Abuse substance	Saliva	Urine
Marijuana	12-24 hours	Days/wk. Depending on the frequency of use
Opioid	12-24 hours	2-4 days
Amphetamine	24-48 hours	1-2 days
Benzodiazepine	24-48 hours	1 week
Cocaine	12-24 hours	2-3 days

 Table 2. The detection time of certain substances of abuse in saliva and urine

**Procedures for extraction** of drugs of abuse from biological samples:

The *liquid - liquid extraction* (LLS): the method used for emergency analysis and for unknown analysis when substances with physico-chemical properties must be extracted. This process facilitates the extraction of a drug from aqueous solutions in organic solvents and involves a relatively high consumption of solvents and multiple operations of extraction and separation.

*Solid phase extraction* (SPE): the aim of this method is the extraction, purification, and, sometimes, the concentration of non-volatile or semi-volatile substances for analysis. It involves passing aqueous solution through a column with silica based desiccant, active carbon and resins. It is a more expensive process and often less sensitive (25-29).

#### Testing of substances of abuse extracted from biological samples

The tests for substances of abuse shall be sub-divided into two types of analytical procedures:

- Screening tests: are quick, simple and requires a minimum previous processing of the sample.
   Examples: immunoassays, Thin Layer Chromatography (TLC).
- *Confirmatory tests*: are performant, sensitive, selective methods that reduce the number of false-positive / false-negative results. Examples: Gas Chromatography-Mass Spectrometry (GCMS), High performance liquid chromatography (HPLC), Liquid chromatography-Mass Spectrometry (LCMS).

1. Screening methods for the determination of drugs of abuse: thin layer chromatography, immunoassay

For the toxicological screening of drugs of abuse simple, quick and inexpensive analytical methods are required. Screening methods plays an important role in the forensic medicine laboratories, both in the analysis of incriminated objects as well as in the analysis of biological samples. Due to the large diversity of samples it is practically impossible to use extraction methods and sophisticated and time-consuming instrumental techniques for analysis of all samples. Therefore, it is absolutely necessary to use simple screening tests to restrict further research area (30- 33).

The conditions that have to be met by a method of analysis to be used as screening test:

- easy to performed
- quick
- not require a complicated and unaffordable equipment
- require few usual reagents
- not require highly qualified personnel
- inexpensive
- able to be performed also outside a lab
- require a minimum processing of the samples.

The interpretation of screening tests results is a complex process, which requires an overview on limitations raised by the analysis method, by pharmacokinetic and biotransformation characteristics of the incriminated substance, but also by psychological, physiological and pathological pattern of the patient (including history of drug dependence) (34). A negative result does not necessarily indicate the absence of the substance, that can be present but at a level below the detection limit of the method. A true-positive result from a screening test will not indicate the dose, the time or the route of administration, and it doesn't make the difference between an occasional or a chronic administration. That's why, it is recommended to use more specific and performant analytical methods for confirming the screening test results (35- 38).

Screening tests are used in several purposes: forensic (analysis of incriminated samples), clinical or medical care (admission in substitution treatment, compliance of treatment, testing abstinence during therapy), occupational medicine, doping tests. The most commonly used screening tests at the present time are *thin-layer chromatography (TLC) and immunoassays*.

*Thin-layer chromatography* (TLC) is a wide spread technique used for the separation and identification of substances. It is used to analyse bulk active substances, pharmaceutical products, but also illicit substances or biological samples. Conventional TLC is a fast and low-cost method for qualitative analysis. Requires a minimum and readily available equipment, and experimental techniques are easily acquired. These determinations are not expensive, and can be carried out in laboratories with limited facilities (39- 42).

The Committee of Systematic Toxicological Examination of the International Association of Forensic Toxicologists (TIAFT), recommends 10 separation systems to identify medicinal substances and drugs of abuse, depending on their acid-base character. The correspondence between the different psycho-active substances and TIAFT recommended systems is shown in table number 5.

Drug of abuse	TA	TB	TC	TE	TL	TAE	TAF	TAJ	TAK	TAL
5-methyltryptamine	56	-	-	-	-	-	-	-	-	-
amphetamine	43	20	9	43	18	12	75	-	-	-
benzphetamine	73	67	70	87	70	60	-	-	-	-
benzoylecgonine	21	0	1	-	-	-	-	-	-	-
cannabidiol	94	-	-	95	-	-	-	88	76	97
cannabinol	94	-	-	95	-	-	-	90	77	97
cocaine	65	45	47	77	54	35	30	13	0	2
Δ <sup>9</sup> - THC	11	-	-	31	-	-	-	0	1	31
diamorphine	47	15	38	49	4	26	33	25	5	64
dimethyltryptamine	46	15	10	63	11	14	56	2	3	41

Table 5. TLC methods for the analysis of psycho-active substances according to TIAFT

dimethyltryptamine	40	9	9	50	6	14	39	-	-	-
DOM	51	15	17	41	16	9	76	-	-	-
ketamine	63	37	63	79	64	68	72	47	4	43
lysergic acid	58	0	0	0	0	70	16	48	7	79
mescaline	20	3	10	24	12	6	63	2	9	51
methamphetamine	31	28	13	42	5	9	63	0	3	45
methadone	48	59	20	77	27	16	60	8	0	45
morphine	37	0	9	20	1	18	23	0	0	15
psilocin	39	5	9	47	9	14	48	-	-	-
psilocybin	5	0	-	0	0	80	1	-	-	-

**Immunoassays** are commonly used as screening tests for testing drugs of abuse. Often, they are not making any discrimination between the related compounds, so the results obtained are likely to be cross-reactive. For this reason, such methods are followed by confirmation using performant separation technique such as GC-MS for qualitative analysis and HPLC for quantitative analysis.

Used on a larger scale, the immunoassays methods are based on the antigen - antibody reaction. The quality of antibody is critical for the sensitivity, precision and accuracy of the determination. In order to generate a measurable signal, the immunoassay technique uses a specific antibody for the identified compound or class of compounds and a labelled form of the same compound or the antibody. The labelling may be done with a radioisotope in case of radioimmunoassay (RIA), an active enzyme in case of enzyme-linked immunosorbent assay (ELISA) or a fluorescent compound in case of fluorescence immunoassay (FIAS) (43- 46). Polarization Immunoassay (FPIA) use fluorescein attached to one compound (antigen) as marker. When it is bound to antibody, fluorescein molecular rotation slows down and leads to changes in the polarization of fluorescent emission. The polarization p is inversely proportional to the concentration of the unbound compound. The main advantage is the exceptional stability of the FPIA reagents, which enable the tracing of the calibration curve valid for longer time and the automation of the determination (47).

In the immunoassay methods, the biological sample requires a minimum previous preparation (e.g. simple centrifugation in the case of urine). After the initial immunoassay test of the screening program, usually we proceed to identify the particular compound involved using performant separation methods from complex matrices as biological samples (48- 50).

#### 2. Toxicological examination of the drugs of abuse - confirmatory methods

In order to eliminate false positive or false negative screening tests, toxicological analysis is continued with confirmatory tests.

Confirmatory tests:

- Are effective methods, sensitive, selective, accurate, reproducible;
- Are performant column chromatographic methods: GC-MS, HPLC, LC-MS;
- Requires a laborious sample preparation stage;
- Requires expensive equipment and highly qualified personnel;
- Are analysed with higher costs.

Chromatography is a method of separating components of a mixture on the basis of their different distribution between two phases, one of which is stationary - generally fixed on a support (glass or aluminium plate, paper sheet, steel column, etc.) and other, mobile, which moves in relation to the fixed phase. This conducted to a different migration of the components leading to their separation. The mobile phase is gas in GC-MS methods and liquid in HPLC and LC-MS methods. The chromatography is used both for qualitative and quantitative determination of the chemical substances. The identification is based on the time required for the migration of the substance into the separation system. The assay is based on the proportionality of the amount to the peak area (51).

#### **Confirmatory Tests - gas chromatography coupled with mass spectrometry (GC-MS)**

The GC-MS is the gold standard for a reliable identification of the drugs of abuse in all kinds of samples. It combines the advantages of gas chromatography with those of mass-spectrometry. Mass

spectrometry is an analytical technique used to identify organic substances, based on pattern recognition's of fragments resulting from the ionization.

References	United Nations International Drug Control Programme	Goldberger B. A. et al.	De Giovanni N. et al.	Wang W.L. et al.
Detected compounds	Drugs of abuse	Heroin, 6-acetyl morphine	Heroin, cocaine and metabolites	Cocaine, morphine, codeine, heroin and metabolites
Column	Diffrent capillary colum 12 m x 0,2 mm d.i.	Rtx-5, 15m x 25 mm d.i.	HP-1, 12 m x 0,2 mm d.i.	HP-1, 12 m x 0,2 mm d.i., film de 0,33 μm
Carrier gas	Не	He ultrapur		Не
Flow rate	1,9 mL/min	1,2 mL/min		1 mL/min
Injection type		splitless		splitless
Injector temperature	250°C	250 °C	250 °C	250 °C
Oven temperature	150 – 300 °C, with gradient of 12 °C/min	1 min. to 150 °C, followed by increasing to 200 °C with a gradient of 12.5 °C / min, maintained at 200 °C for 15 s, increasing to 290 °C with 30 °C/min, held for 4 min	120 °C for 1 min, increasing to 220 °C with a gradient of 20 °C / min, then to 260 °C with 5 °C / min and finally to 280 °C with a 20 °C / min, held for 2 min	70 ° for 1 min, increasing to 220 °C with a gradient of 35 °C / min, constant at 220 °C 0.25 min, increasing to 250 °C at 10 °C / min, held at 250 ° C for 3 min
Derivatization	SIM or TFA	-	BSTFA -1%TMCS	-
Observation	abuse	6-acetyl morphine: LOD 1,0 μg/L, range 1,0 – 500 μg/L, r >0,995	LOD 50 ng/mL; range 50-500 ng/mL	Hair: range 0,1 – 10,0 ng/mg Urine, saliva, plasma: range 1 – 100 ng/mL
Extraction	Liquid-liquid extraction or SPE with C18 cartridge	SPE with ZSDAU020 cartridge	SPE with C18 cartridge	SPE
Biological samples	Blood, saliva and urine	Blood, plasma, saliva and urine	Urine	Blood, saliva, urine and hair

Table 6. GC-MS methods for heroin and its metabolites analysis in biological samples

The mass spectrum recorded is compared with libraries of mass spectrum. In table number 6 are

listed a few GC-MS methods for heroin and its metabolites analysis in biological samples.

Confirmatory Tests – High Performance Liquid Chromatography (HPLC)

HPLC method is the first choise for the quantitative determination of drugs of abuse in all kinds of samples. Quantitative determination is based on proportionality between peak area and amount of the analyte in the sample. In Table number 7 several HPLC methods for analysis of the heroin and its metabolites in biological samples are presented (52).

	United Nations International Drug Control Programme	United Nations International Drug Control Programme	Katagi M. et al.	Bourquin D. et al.	Low A.S. et al.	Umans J. G. et al.
Detected compound s			Heroin and metabolites (monoacetylmorphinae, diacetylmorphine)	Heroin, morphine, codeine and their metabolites	Heroin, 6- monoacetyl- morphine, codeine, pholcodine, dihydrocodeine, morphine	Heroin, 6- monoacetyl -morphine, morphine
Detection	UV, λ=218nm	Electrochemical	Mass spectrometry	DAD, $\lambda$ =210 nm	UV, λ=280 nm	UV, λ=218 nm
Column	LiChrosorb 60, 5 µm, 30cm x 4 mm d.i.	ODS, 5µm, 25 cm x 4,6 mm d.i.	Capcell Pak SCX 1,5 mm d.i. x 150 mm, 5 µm	C18, 125 x 2 mm d.i., 3 µm	Hypersil 3µm, 200 x 2 mm d.i.	LiChrosorb Si 60 5µm, 30 x 4 mm d.i.,
Faza mobilă	acetonitril e: ammonia: methanol: acetic glacial acid soluția B	acetonitrile - 0.2M sodium perchlorate buffer / 0.005 M sodium citrate (1: 9, v / v)	10 mM ammonium acetate (pH 6.0) - acetonitrile (30:70, v / v)	o-phosphoric acid, dicyclohexylamine, acetonitrile, water,	dichloromethane, pentane, methanol, diethylamine	acetonitrile, ethanol concentrate d ammonia, methanol, glacial acetic acid
Elution	Izocratic	Izocratic	Izocratic	Gradient	Izocratic	Izocratic
Flow rate	1,3 mL/min	1,9 mL/min		0.2 mL/min	0,4 mL/min	80 mL/ora (1.33 mL/min)
Extracție	LLE or SPE with C-18 cartridge	LLE or SPE with C-18 cartridge	SPE	ex SPE with C-18 cartridge	SPE with C-18 cartridge	LLE
Biological samples	Blood, saliva and urine	Blood, saliva and urine	Urine	PLasma	Urine	Blood

Table 7. HPLC methods for analysis of the heroin and its metabolites in biological samples

Confirmation Tests - liquid chromatography coupled with mass spectrometry (LC-MS)

Liquid chromatography coupled with mass spectrometry is a modern hyphenated technique which combines the advantages of HPLC with those of mass-spectrometry. The mass spectrum recorded is compared with libraries of mass spectrum. It is used for both qualitative and quantitative determination,

having as advantages the selectivity and the increased sensitivity (53).

In table number 8 a few LC-MS methods for analysis of the methadone and its metabolites in biological samples are listed (54).

References	Vlase L. et al.	Dams R. et al.	Danielson T.J. et al.	Widschwendter C. G. et al.	Kelly T. et al.	Rosas M.E. et al.
Detected compounds	Methadone	Opioids, cocaine and metabolites	Methadone and her metabolites (EDDP, EMDP)	Methadone and quetiapine	Enantiomers of the methadone and of EDDP, EMDP	Enantiomers of the methadone and of EDDP
Detection	$\begin{array}{c} \text{MRM} \\ \rightarrow 265 \end{array} (310$	SRM	MRM	MRM (m/z 310 → 265; 384 → 253)	MRM	SIM (m/z 310, 278, 313, 281)
Ionization	ESI	API	?	API	API	ESI
Column	Zorbax SB- C18 (100 x 3.0 mm, 3.5 µm I.D.)	Synergi Polar RP (150 x 2 mm, 4 µm)	?	Waters Acquity, C18 (50 x 2,1 mm, 1,7 μm)	AGP alpha- glicopro-tein	AGP alpha- glicopro-tein
Internal standard	-	Deuterated isotopes of the analysed compounds	Deuterated isotopes of the analysed compounds	D3-methadone	-	D3-methadone and D3-EDDP
Mobile phase	Acetonitrile: 0.2% formic acid 45:55 (v/v)	Mixture of 10 mM ammonium formate or 0.001% formic acid and acetonitrilein various proportions		Mixture of acetonitrile and 5 mM formic acid	20 mM acetic acid: isopropanol 93: 7 (v/v)	Acetonitrile: ammonium acetate buffer 18:82 (v/v)
Elution	isochratic	gradient		gradient	isochratic	isochratic
Flow rate Column temperature	1 mL/min 45°C	300 μL/min 25°C	-	0,25 mL/min -	0,9 mL/min -	0,9 mL/min 25°C
Extraction	De- proteinized plasma with methanol Urine diluted with water	Without	?	With acetonitrile and centrifugation	?	?
Biological samples	Plasma Urine	Urine	Blood Liver	Urine	Hair	Saliva

*Table 8. LC-MS methods for methadone and its metabolites determination in biological samples* 

#### Conclusions

The methodology of a toxicological analysis of the substances of abuse shall be developed on the basis of: test sample type, analysis purpose, pharmacokinetic and biotransformation particularities of substance, equipment and reagents available, stuff expertise, cost.

Immunoassays offers a flexible approach of the analyses of drugs of abuse from different biological samples and represents a convenient method and a quick screening test for a large number of samples, with different matrixes. A true-positive result of an initial screening test, will not indicate on its own the dose, the time or the route of administration and it will not make the difference between an occasional administration or a chronic one.

*Screening* tests require subsequently performing confirmation tests for removing false positiveâ/ false negative results. Confirmation tests are modern chromatographic techniques (GC-MS, HPLC, LC-MS), high-performance, sensitive, selective, accurate. Confirmation tests have as disadvantages: timeconsuming step for the processing of the samples, expensive equipment, highly qualified staff and high cost.

The identification and the assay of drugs of abuse and their metabolites in biological samples provides to the specialists (doctors, authorities in the field of health, representatives of law) an objective tool for the diagnostic of abuse or for the monitoring of addictions treatment. Interpretation of the results is a complex process and requires an overview of: the analysis method, pharmacokinetic and biotransformation particularities of the substance, clinical pattern of the patient (including history of drug dependence).

**ACKNOWLEDGEMENT**: This paper is supported by Sectoral Operational Programme Human Resources Development (SOP HRD), financed from the European Social Fund and by the Romanian Government under the contract number POSDRU/159/1.5/S/132395

#### References

- Concheiro M, Gray TR, Shakleya DM, Huestis MA. High-throughput simultaneous analysis of buprenorphine, methadone, cocaine, opiates, nicotine, and metabolites in oral fluid by liquid chromatography tandem mass spectrometry. *Anal Bioanal Chem. 2010*; 398(2): 915–924.
- Hoja H, Marquet P, Verneuil B, Lotfi H, Penicaut B, Lachatre G. Applications of liquid chromatography-mass spectrometry in analytical toxicology: a review., *J. Anal. Toxicol. 1997*; 21(2): 116-126.
- Brunet BR, Allan JB, Karl BS, Patrick M, Marilyn AH. Development and validation of a solidphase extraction gas chromatography-mass spectrometry method for the simultaneous quantification of methadone, heroin, cocaine and metabolites in sweat. *Anal Bioanal Chem.* 2008; 392(2): 115–127.
- Dams R., Murphy CM, Lambert WE, Huestis MA. Urine drug testing for opioids, cocaine, and metabolites by direct injection liquid chromatography/tandem mass spectrometry, *Rapid Commun. Mass Spectrom.* 2003; 17: 1665–1670.
- 5. Cheng Y, Neue UD, Woods LL. Novel high-performance liquid chromatographic and solid-phase extraction methods for quantitating methadone and its metabolite in spiked human urine, *Journal of Chromatography B. 1999*, 729: 19–31.
- De Giovanni N, Rossi SS. Simultaneous detection of cocaine and heroin metabolites in urine by solid-phase extraction and gas chromatography-mass spectrometry *J. Chromatography B 1994*, 658: 69-73.
- 7. \*\*\* http://what-when-how.com/forensic-sciences/presumptive-chemical-tests/
- \*\*\* Recommended Methods for the Detection and Assay of heroin, Cannabinoids, Cocaine, Amphetamine, Metamphetamine and Ring-Substituted Amphetamine Derivatives in Biological Specimens – United Nations International Drug Control Programme (UNIDCDP), New York, 1995.
- 9. Aitken CG. Sampling How big a sample?, J. Forensic Sci. 1999, 44: 750-760.

- 10. Baconi D. Toxicomanii Note de curs, Ed. Tehnoplast Company SRL, București, 2005.
- Baconi D, Bălălău C. Toxicologia substanțelor de abuz, Ed. Universitară Carol Davila, București, 2013.
- 12. Bourquin D, Lehman T, Hämmig R, Bührer M, Brenneisen R. High-performance liquid chromatographic monitoring of intravenously administered diacetylmorphine and morphine and their metabolites in human plasma. *J. Chromatography B* 1997, 694:233-238.
- Choo RE, Lauren M. Jansson KS, Marilyn AH. A Validated Liquid Chromatography–Atmospheric Pressure Chemical Ionization-Tandem Mass Spectrometric Method for the Quantification of Methadone, 2-Ethylidene-1,5-dimethyl-3,3- diphenylpyrrolidine (EDDP), and 2-Ethyl-5-methyl-3,3-diphenylpyroline (EMDP) in Human Breast Milk. *J Anal Toxicol. 2007*; 31(5): 265–269.
- Choo RE, Constance MM, Hendree EJ, Marilyn AH. Determination of methadone, 2-ethylidene-1,5-dimethyl-3,3- diphenylpyrrolidine, 2-ethyl-5-methyl-3,3-diphenylpyraline and methadol in meconium by liquid chromatography atmospheric pressure chemical ionization tandem mass spectrometry. J. Chromatogr. B 2005; 814: 369–373.
- 15. Danielson TJ, Mozayani A, Sanchez LA. Methadone and methadone metabolites in postmortem specimens *Forensic Sci Med Pathol 2008;* 4: 170-174.
- 16. de Castroa A, Marta C, Diaa MS, Marilyn AH. Development and validation of a liquid chromatography mass spectrometry assay for the simultaneous quantification of methadone, cocaine, opiates and metabolites in human umbilical cord. *J. Chromatogr. B* 2009; 877: 3065–3071.
- Etter ML, George KG, Eichhorst J, Lehotay DC. Determination of free and protein-bound methadone and its major metabolite EDDP: Enantiomeric separation and quantitation by LC/MS/MS. *Clinical Biochemistry 2005*; 38: 1095–1102.
- Flanagan RJ, Braithwaite RA, Brown SS, Widdop B, de Wolff FA. Basic Analytical Toxicology -World Health Organization, Geneva, 1995.

- 19. Foster DJ, Andrew AS, Jason MW, Felix B. Population pharmacokinetics of (R)-, (S)- and racmethadone in methadone maintenance patients. *Br J Clin Pharmacol*, 2004; 57(6): 742–755.
- 20. Foster DJ, Somogyi AA, Bochner F, Stereoselective quantification of methadone and its major oxidative metabolite, 2-ethylidene-1,5-dimethyl-3,3-diphenylpyrrolidine, in human urine using high-performance liquid chromatography, *Journal of Chromatography B 2000*; 744: 165–176.
- 21. Fríguls B, Joya X, García-Algar O, Pallás CR, Vall O, Pichini S. A comprehensive review of assay methods to determine drugs in breast milk and the safety of breastfeeding when taking drugs. *Anal Bioanal Chem. 2010*; 397(3): 1157-1179.
- 22. Gergov M, Nokua P, Viori E, Ojanpero I. Simultaneous screening and quantification of opioid drugs in post-mortem blood and urine by LCMS *Forensic Science International 2009*; 36-43.
- 23. Gheorghe M, Bălălău D, Ilie M, Baconi DL, Ciobanu AM. Component analysis of illicit heroin samples by GC-MS method *Farmacia 2008*, LVI (5): 577-582.
- 24. Gheorghe M, Bălălău D, Ilie M, Baconi DL, Ciobanu AM. Qualitative analysis of confiscated illegal drugs by thin-layer chromatography, *Farmacia 2008*, LVI (5): 541-546
- 25. Girod C, Staub C. Methadone and EDDP in hair from human subjects following a maintenance program: results of a pilot study. *Forensic Science International 2001*; 117(3): 175-184.
- Goldberger BA, Darwin WD, Grant TM, Allen AC, Caplan YH, Cone EJ. Measurement of Heroin and Its Metabolites by Isotope-Dilution Electron-Impact Mass Spectrometry. *Clin. Chem. 1993*; 39(4): 670-675.
- 27. Hallinan R, Raya J, Byrne A, Kingsley A, Attia J. Therapeutic thresholds in methadone maintenance treatment: A receiver operating characteristic analysis. *Drug and Alcohol Dependence* 2006; 81: 129–136.
- Jansson LM, Robin EC, Harrow C, Martha V, Schroeder JR, Lowe R, Huestis MA. Concentrations of Methadone in Breast Milk and Plasma in the Immediate Perinatal Period. *J Hum Lact.* 2007; 23(2): 184–190.

- 29. Katagi M, Nishikawa M, Tatsuno M, Miki A, Tsuchihashi H. Column-switching high-performance liquid chromatography–electrospray ionization mass spectrometry for identification of heroin metabolites in human urine. *Journal of Chromatography B 2001*; 751: 177–185.
- 30. Kelly T, Doble P, Dawson M. Chiral analysis of methadone and its major metabolites (EDDP and EMDP) by liquid cromatography-mass spectrometry. *J Chromatogr B Analyt Technol Biomed Life Sci.* 2005; 814(2): 315-323.
- 31. Kintz P, Julie E, Marion V, Vincent C. Interpretation of hair findings in children after methadone poisoning. *Forensic Science International 2009*; 196(1-3): 51-54.
- 32. Larson ME, Thomas MR. Quantification of a Methadone Metabolite (EDDP) in Urine: Assessment of Compliance. *Clinical Medicine & Research 2009*; 7(4): 134-141.
- 33. Lehotay DC, George S, Etter ML, Graybiel K, Eichhorst JC, Fern B, Wildenboer W, Selby P, Kapur B. Free and bound enantiomers of methadone and its metabolite, EDDP in methadone maintenance treatment: Relationship to dosage? *Clinical Biochemistry* 2005; 38: 1088–1094.
- 34. Liang HR, Foltz RL, Meng M, Bennett P. Method development and validation for quantitative determination of methadone enantiomers in human plasma by liquid chromatography/tandem mass spectrometry. J. Chromatogr. B. 2004; 806(2): 191-198.
- 35. Low AS, Taylor RB. Analysis of common opiates and heroin metabolites in urine by highperformance liquid chromatography. *J. Chromatography B. 1995*; 663: 225-233.
- 36. Lucas CS, Bermejob AM, Tabernerob MJ, Fernándezb P, Strano-Rossic S. Use of solid-phase microextraction (SPME) for the determination of methadone and EDDP in human hair by GC–MS. *Forensic Science International 2000*; 107(1-3): 225-232.
- 37. Moeller MR, Feya P, Wennig R. Simultaneous determination of drugs of abuse (opiates, cocaine and amphetamine) in human hair by and its application to a methadone treatment program. *Forensic Science International 1993*; 63(1-3): 185-206.

- Moffat AC, Osselton MD, Widdop B. (eds) Clarke's analysis of drugs and poisons, third edition,
   2004, Pharmaceutical Press
- 39. Moore C, Guzaldo F, Hussain MJ, Lewis D. Determination of methadone in urine using ion trap GC/MS in positive ion chemical ionization mode. *Forensic Science International 2001*; 119(2): 155-160.
- 40. Nanovskaya TN, Sujal VD, Ilona AN, Zharikova OL, Gary DVH, Mahmoud SA. Methadone metabolism by human placenta. *Biochemical Pharmacology* 2004; 68: 583–591.
- 41. Nikolaou Panagiota D., Ioannis I. Papoutsis, Julia Atta-Politou, Sotiris A. Athanaselisa, Chara A. Spiliopouloa, Antony C. Calokerinos, Constantinos P. Maravelias Validated method for the simultaneous determination of methadone and its main metabolites (EDDP and EMDP) in plasma of umbilical cord blood by gas chromatography–mass spectrometry. *J. Chromatogr. B 2008*; 867: 219–225.
- 42. Ortelli Didier, Serge Rudaz, Anne-Françoise Chevalley, Annie Mino, Jean-Jacques Deglon, Luc Baland, Jean-Luc Veuthey Enantioselective analysis of methadone in saliva by liquid chromatography–mass spectrometry. *J. Chromatogr. A 2000*, 871(1-2): 163-172.
- 43. Perrigo BJ, Joynt BP. Use of ELISA for the detection of common drugs of abuse in forensic whole blood samples, *Can. Soc. Forensic Sci. J. 1995*; 28: 261-269.
- 44. Quintela O, Lopez P, Bermejo AM, M. Lopez-Rivadulla. Determination of methadone, 2ethylidene-1,5-dimethyl-3,3-diphenylpyrrolidine and alprazolam in human plasma by liquid chromatography–electrospray ionization mass spectrometry. *J. Chromatogr. B* 2006; 834: 188–194.
- 45. Rodriguez Rosas M.E., Kenzie L. Preston, David H. Epstein, Eric T. Moolchan, Irving W. Wainer -Quantitative determination of the enantiomers of methadone and its metabolite (EDDP) in human saliva by enantioselective liquid chromatography with mass spectrometric detection. *J. Chromatogr. B 2003*; 796(2): 355-370.

- 46. Rook Elisabeth J., Michel J.X. Hillebrand, Hilde Rosing, Jan M. van Reeb, Jos H. Beijnen The quantitative analysis of heroin, methadone and their metabolites and the simultaneous detection of cocaine, acetylcodeine and their metabolites in human plasma by high-performance liquid chromatography coupled with tandem mass spectrometry. J. Chromatogr. B 2005; 824: 213–221.
- 47. Rosas ME, Preston KL, Epstein DH, Moolchan ET, Wainer IM. Quantitative determination of the enantiomers of methadone and its metabolite (EDDP) in human saliva by enantioselective liquid chromatography with mass spectrometric detection. *J Chromatogr B Analyt Technol Biomed Life Sci. 2003*; 796(2): 355-370.
- 48. Schmidt N, Brune K, Geisslinger G. Stereoselective determination of the enantiomers of methadone in plasma using high-performance liquid chromatography, *J.Chromatogr. B Biomed Sci. Appl. 1992*; 583(2): 195-200.
- 49. Shakleya D.M., Riet Dams, Robin E. Choo, Hendree Jones, Marilyn A. Huestis Simultaneous Liquid Chromatography–Mass Spectrometry Quantification of Urinary Opiates, Cocaine, and Metabolites in Opiate-Dependent Pregnant Women in Methadone-Maintenance Treatment. J Anal Toxicol. 2010; 34(1): 17–25.
- 50. Shakleya Diaa M., Lauren M. Jansson, Marilyn A. Huestis Validation of a LC–APCI-MS/MS method for quantification of methadone, 2-ethylidene-1,5-dimethyl-3,3-diphenylpyrrolidine (EDDP) and 2-ethyl-5-methyl-3,3-diphenylpyraline (EMDP) in infant plasma following protein precipitation. *J. Chromatogr. B. 2007*; 856: 267–272.
- 51. Umans JG, Chiu TS, Lipman RA, Schultz MF, Shin SU, Inturrjsl CE. Determination of heroin and its metabolites by high performance liquid chromatography J. Chromat. 1982; 233: 213-225.
- 52. lase L, Popa DS, Leucuța SE, Loghin F. Bioanalysis of methadone in human plasma and urine by LC/MS/MS *Revue Roumaine de Chimie 2008*, 53(12): 1157–1164.

- 53. Wang WL, Darwin WD, Cone EJ. Simultaneous assay of cocaine, heroin and metabolites in hair,plasma, saliva and urine by gas chromatography-mass spectrometry. *J. Chromat. B 1994*, 660 : 279-290.
- 54. Widschwendter CG, Zernig G, Hofer A. Quetiapine cross reactivity with urine methadone immunoassays. *Int Clin Psychopharmacol* 2006; 21: 81-85.