Professional Paper

DETERMINATION OF ARTIFICIAL SWEETENERS IN BEVERAGES AND SPECIAL NUTRITIONAL PRODUCTS USING HIGH PERFORMANCE LIQUID CHROMATOGRAPHY

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This paper presents two high performance liquid chromatographic (HPLC) methods used for the separation and determination of artificial sweeteners aspartame, acesulphame K, sodium saccharin, and sodium cyclamate in beverages and special nutritional products (special food intended for specific population groups). All four compounds are soluble in aqueous solutions and can easily be separated and determined by HPLC with a diode array detector (DAD). The first method involved separation of aspartame, acesulphame K, and sodium saccharin on a C_{18} column with an isocratic elution of phosphate buffer and acetonitrile as mobile phase. The second method was used to separate sodium cyclamate on a C_{18} column with methanol and water as mobile phase. Under optimum conditions, both methods showed good analytical performance, such as linearity, precision, and recovery. The methods were successfully applied for the analysis of real samples of soft drinks and special nutritional products.

KEY WORDS: acesulphame K, aspartame, sodium cyclamate, sodium saccharin, validation

Artificial sweeteners are common in food, drinks, and pharmaceutical products (1). They are often used to control calorie intake and in certain medical conditions such as diabetes and hyperglycaemia (2). The most common are aspartame, sodium cyclamate, acesulphame K, and sodium saccharin, which are marketed in about 90 countries of the world (3). Aspartame (N-L- α -aspartyl-L-phenylalanine-methyl ester) contains two amino acids, aspartic and phenylalanine, and is between 160 and 220 times sweeter than sugar (4, 5). Sodium and calcium cyclamates are 30 to 40 times as sweet as sucrose (6). In combination with other low-calorie sweeteners, they are sweeter than the sum of individual sweeteners (7). Acesulphame K is a strong non-caloric sweetener which does not metabolise in the body, and is excreted unchanged (5). Sodium saccharin has the longest history of use, and has provoked strong controversy over its possible carcinogenic effects (3). The EU and Croatian regulations have established maximum permitted concentrations of artificial sweeteners in food (8-12). Their content in food, drinks, and dietary products is being determined using a variety of methods such as UV spectroscopy (1, 5), capillary electrophoresis (1, 4, 6), high performance liquid chromatography (1, 2, 5, 7), and ion chromatography (1, 3).

The aim of this study was to develop and test two methods for routine laboratory analysis of aspartame, sodium cyclamate, acesulphame K, and sodium saccharin in soft drinks and special nutritional products.

MATERIAL AND METHODS

Chemicals

Reference compounds (acesulphame K, sodium cyclamate, and sodium saccharin) were purchased from Sigma-Aldrich (Steinheim, Germany), and aspartame from Fluka (Buchs, Switzerland). All solid standards were of ≥98 % purity. HPLC-grade methanol and acetonitrile were obtained from Merck (Darmstadt, Germany). Water was purified using a Milli-Q Ultrapure water purification system (Millipore, Bedford, MA, USA). Potassium dihydrogen phosphate, phosphoric acid, sulphuric acid, heptane, sodium hypoclorite, sodium carbonate, and sodium sulphate were obtained from Kemika (Zagreb, Croatia).

Stock solutions were prepared separately by dissolving reference compounds in deionised water. Working standard solutions of aspartame, acesulphame K, and sodium saccharin were prepared by mixing and diluting aliquots of stock solutions. Sodium cyclamate in 30 mL stock solution was derivatised into cyclohexylsulphamic acid because of its poor absorbance.

Samples

For the analysis, we collected 104 products marketed in Croatia. Seventy-eight were soft drinks (41 from plant extracts, 11 fruit juice samples, and 26 artificially and flavoured drinks) and 26 were special nutritional products, such as baby food, gluten-free

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Figure 1 Chromatograms of a standard mixture of artificial sweeteners: 1 -acesulphame K, 2 - sodium saccharin, 3 - aspartame

food, food for specific medical purposes, food intended for people suffering of diabetes, food with a reduced or increased energy value, and nutritional supplements (15 in water soluble tablets, and 11 in powders).

Sample preparation

Samples of soft drinks were degassed for 10 minutes in an ultrasonic bath before dilution with water. To determine aspartame, acesulphame K, and sodium saccharin in soft drinks, we diluted 10-mL samples with deionised water in 100 mL volumetric flasks. Samples of special nutritional products were weighted to 5 g, placed into 100 mL glass flasks, diluted to 100 mL with deionised water, and degassed. Soft drinks and special nutritional product solutions were then filtered into vials through a 0.45 μ m membrane filter (Whatman, UK) and injected into the HPLC system (13).

To determine sodium cyclamate, 30 mL stock solutions were derivatised into cyclohexylsulphamic acid, as follows: the liquid sample, 1 mL of sulphuric acid, 10 mL of heptanes, and 2.5 mL of sodium hypoclorite solution were added into a separating funnel. After separating the phases, the aqueous phase was discarded and heptane layer washed with 25 mL of sodium carbonate solution (50 g L⁻¹). The lower phase was discarded and heptane phase dried with 1 g of sodium sulphate and filtered through a fluted filter paper and then through 0.45 μ m membrane filter into vials (14).

HPLC conditions

For chromatography we used Shimadzu LC Prominence 20 HPLC (Kyoto city, Japan) with a diode

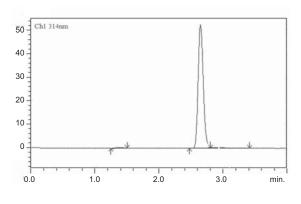


Figure 2 Characteristic chromatogram of cyclohexylsulphamic acid

		Name of artificial sweetener					
		Acesulfame K	Aspartame	Sodium saccharin	Sodium cyclamate		
Λ _{max} / nm		226	226 193		314		
Linearity range / mg L ⁻¹		0.3 to 24.3	0.5 to 40.5	0.4 to 32.4	0.5 to 40.5		
Correlation coefficient / r²		1	1	1	0.9999		
Linear equation		y=2128x+6696	y=6425x-291.9	y=4351x+7243	y=2334.5x-7599		
LOQ / mg L ⁻¹		0.3	0.5	0.4	0.5		
Precision / RSD, %	Injection repeatability	0.15	0.15	0.15	0.3		
	Analytical repeatability	0.5	0.7	0.5	0.9		
Recovery / %	2 x LOQ	101	97.5	99	101		
	10 x LOQ	98.4	96.2	100.6	99.5		

Table 1 Analytical performance of the methods used for determination of artificial sweeteners

Table 2 Concentration ranges of artificial sweeteners in samples of soft drinks and special nutritional products

	Soft drinks			Special nutritional products		Maximum
Analyte	Soft drinks from plant extracts	Fruit juices	Artificially and flavored drinks	Soluble tablets	Powders	permitted level / mg L ⁻¹
A acculations V	116.75 to	90.45 to	206.25 to	102.07 to	114.20 to	350
Acesulphame K	399.13	420.60	451.47	269.64	304.85	
Acceptone	153.69 to	80.29 to	198.22 to	143.97 to	156.97 to	600
Aspartame	876.42	435.05	709.36	338.75	554.35	
Sodium saccharin	35.11 to	40.01 to	65.77 to	21.67 to	39.86 to	80
Sodium saccharin	134.59	55.24	76.91	66.59	75.14	
Cadimus analamata	80.13 to	70.10 to	203.68 to	42.36 to	87.72 to	400
Sodium cyclamate	627.38	583.94	621.75	240.90	315.66	

array detector, and a C_{18} column (15 cm x 4.6 mm, 5 μ m) manufactured from Supelco (Texas city, USA).

The isocratic elute used for the separation of aspartame, acesulphame K, and sodium saccharin contained 15 % acetonitrile, and 75 % phosphate buffer (0.0125 mol L⁻¹ KH₂PO₄ at pH 3.5). Elution flow rate was 1.5 mL min⁻¹. Total running time was five minutes with injection volume of 10 μ L. Cyclohexylsulphamic acid was separated using isocratic elute consisting of 85 % methanol and 15 % water. The injection volume was 10 μ L and chromatograms were recorded for four minutes.

Standards were analysed on a separate chromatogram and quantification was based on a comparison between the peak areas of the sample and of the reference standard. The presence of an analyte in the sample was confirmed by a diode array detector. Peaks were confirmed by comparing their spectra with those of the aqueous solutions of the standards.

RESULTS

The described analytical methods effectively separated the artificial sweeteners of all analysed products. Figures 1 and 2 show the chromatograms of standard solutions.

Method validation data are shown in Table 1. Limits of quantifications (LOQs) were estimated with concentrations giving a signal-to-noise ratio of about 10. LOQ is defined as the lowest concentration on the calibration curve.

Linearity measurement was based on five concentration points with three replicates of standard solution for each concentration level.

The precision of our two methods was evaluated as injection repeatability and analytical repeatability based on five replicates of standard solutions. Precision is expressed as the relative standard deviation (RSD) of the replicates. Recovery was determined by spiking selected samples of each matrix with standards at concentrations twice the LOQ and 10 times the LOQ.

Application with real samples

Artificial sweeteners were found in all analysed products. All products truthfully declared to have "artificial sweeteners" or "no added sugars" on their labels. Table 2 shows concentration ranges and relative maximum levels for each analyte set by Croatian law (13). Twenty-five soft drink samples exceeded the maximum levels, as follows: seven made of plant extracts, two fruit juices, and sixteen artificially flavoured drinks. None of the samples of special nutritional products exceeded maximum limits.

CONCLUSION

We have found the two described methods suitable for routine analysis of artificial sweeteners in beverages and special nutritional products. Limits of quantification, precision, and recovery were satisfactory.

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Sažetak

ODREĐIVANJE UMJETNIH SLADILA U OSVJEŽAVAJUĆIM BEZALKOHOLNIM PIĆIMA I POSEBNIM PREHRAMBENIM PROIZVODIMA METODOM TEKUĆINSKE KROMATOGRAFIJE VISOKE DJELOTVORNOSTI

Ovaj rad predstavlja dvije metode za separaciju i određivanje umjetnih sladila aspartama, acesulfama K, natrijeva saharina i natrijeva ciklamata u osvježavajućim bezalkoholnim pićima i posebnim prehrambenim proizvodima tehnikom tekućinske kromatografije visoke djelotvornosti (HPLC). Sva četiri analita topljiva su u vodenim otopinama i lako se mogu razdvojiti i odrediti tekućinskom kromatografijom uz primjenu detektora s nizom dioda (DAD). Prva je metoda kromatografsko razdvajanje aspartama, acesulfama K i natrij saharina na C_{18} koloni izokratnim ispiranjem fosfatnog pufera i acetonitrila kao mobilne faze. Druga metoda omogućava određivanje natrijeva ciklamata na C_{18} koloni sa metanolom i vodom kao mobilnom fazom. Pod najpovoljnijim eksperimentalnim uvjetima obadvije metode pokazale su dobre analitičke karakteristike, kao što su linearnost, preciznost i analitički povrat. Navedene su metode primijenjene za analize realnih uzoraka osvježavajućih bezalkoholnih pića i hrane za posebne prehrambene potrebe.

KLJUČNE RIJEČI: acesulfam K, aspartam, natrijev ciklamat, natrijev saharin, validacija

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