



Immunobiological aspects of acute discogenic pain of low back pain

BRANKO UHODA

Specialist Hospital for Orthopaedics
Zadarska 62
23210 Biograd na moru
Croatia
E-mail: ortrav@email.t-com.hr

Key words: cytokine, disc, back pain, immunology

INTRODUCTION

Low back pain is the most common health problem for individuals between the ages of 20 and 50 years. In most cases, the origin of low back pain remains unsolved. Many of established anatomic causes of low back pain are believed to arise from damage of the intervertebral disc (IVD) indirectly through degeneration. Although causal relationship between disc degeneration and low back pain is not yet definitive, in some cases, pathogenetic mechanisms have been elucidated. For example, Moran and King (1) showed that spinal instability was one of the most common causes of low back pain, and other studies have shown that the degenerative process in discs is characterized by instability, especially in traumatic pathology.

Although disc degeneration is of great clinical importance, much remains unknown about its etiology and pathogenesis. Conditions that cause degradation of nuclear and annular material properties and hence the load bearing properties of the disc may lead to damage of other spinal structures, resulting in disc collapse, herniation or spondylosis. Among the many suspected conditions such as mechanical factors, genetics, and systemic factors, insufficient nutrition of the disc has been suspected as the underlying factor in disc degeneration (2, 8).

Nutrition and metabolism of intervertebral disc

The adult human disc is the largest avascular structure in the body. In adult discs, some cells may be as much as 6 to 8 mm from the nearest blood supply, which resides in the osseous endplate of the adjacent vertebral bodies. The vertebral endplate is covered by a thin layer of hyaline cartilage, the cartilaginous endplate, the deep calcified layer, and the underlying subchondral bone. The mineralized portion of endplate is penetrated by marrow contact channels (MCC), through which capillary buds emerge. These capillary buds connect the trabecular spaces to the cartilaginous endplate, but do not penetrate into it. In addition to the endplate, there is also a vascular network in the outer annulus. Several studies have demonstrated that the central region of the endplate is the predominant route of transport for metabolic processes of the disc (2, 39, 41).

The actual complex transport mechanisms of disc nutrition and metabolism have also been studied. For small solutes, diffusion is the predominant molecular transport mechanism from the endplate to the nucleus. It is sufficient for transport rates consistent with the metabolic needs of the disc. Because of the poroelastic nature of the disc matrix, fluid flow is inextricably coupled to the diurnal volume change in the

disc. This cyclic flow of fluid, exudation and imbibition, occurs mainly through the MCCs of endplates. Hence, for small solutes, diffusion may still predominate, but transport of larger solutes such as proteins are believed to be significantly facilitated by the bulk flow of fluid.

Nevertheless, whether the diffusion alone or in combination with convection, the predominant transport route for nutrients and metabolites in the disc remains through the endplates. Interestingly, endplate blood vessels have been observed to diminish after the first decade of life when the first signs of disc degeneration are evident. Also, reduced permeability of endplates is generally seen in association with disc degeneration and age-related changes. Both of these may be due to calcification of the endplates and occlusion of the MMCs observed with disease and age.

The intervertebral disc is avascular, and there are thus steep gradients in concentration of nutrients and metabolites from the blood supply at the discs' margins and to the cells in the center of the disc. Even in normal healthy animal discs, low oxygen and high lactic acid concentration (and thus acidic pH levels) compared to levels in the plasma have been measured. These gradients form when the cells use nutrients and produce metabolites at rates that are high compared to rates of transport into the disc and diffusion through the tissue. The actual concentrations of glucose, oxygen, and lactic acid in the disc will thus depend both on parameters affecting nutrient transport and on cellular metabolic rates.

Local concentrations of extracellular nutrients and metabolites are known to have a significant effect on the activity and survival of disc cells *in vitro*. However, using currently available techniques, measurement of these concentrations is invasive. Although measurements have been made in animal discs and in pathologic human discs during surgical procedures, such measurements cannot be carried out *in vivo* in normal human discs. Information on actual concentrations of nutrients and metabolites in disc can thus only be obtained from information on factor(s) regulating their concentrations gradients, namely transport into the disc and cellular metabolism (38, 39, 40).

There have been a number of studies on factors that affect transport into the disc and through the disc matrix. From these studies, it is clear that nutrient supply to the disc can be compromised by a decrease in the blood supply to the disc, by osteonecrosis, and by calcification of the cartilaginous endplate. Such changes can produce a measurable loss in nutrient supply and hence are strongly implicated in the etiology of disc degeneration. However, local nutrient concentrations are also strongly regulated by cellular demand: in a „dead« disc, there are no nutrient gradients, and the concentrations across the disc reach those of surrounding plasma (2,4, 39).

Far less is known about nutrient demand and metabolite production than about nutrient supply. There are only two studies to our knowledge that have investigated the rates of oxygen and glucose consumption and lactic

acid production by the disc. Both of these studies were on disc explants and examined changes in oxygen consumption and lactic acid production as a function of oxygen tension at constant glucose and pH levels. However, such studies do not give information about physiologic conditions in the disc, because changes in concentration cannot occur independently *in vivo*. Disc cells use glucose to produce adenosine triphosphate (ATP), with lactic acid as the resulting metabolite; the concentrations of glucose and lactic acid are thus necessarily coupled, as are oxygen and lactic acid. The center of the disc thus experiences low oxygen concentrations together with low glucose and high lactic acid concentrations (and hence acidic levels of pH). At present, it is not known how such a microenvironment will affect cellular metabolism; but the center of the disc appears most vulnerable to changes in nutrient concentrations and experiences the first signs of degeneration and death. It is thus of importance to understand disc cell metabolism and activity under such conditions (39, 40).

Pathophysiological mechanisms of chronic spinal pain syndromes

The initial mechanical injury caused by abnormal motions or loads to neural tissue initiate a complicated cascade of interrelated events inducing structural tissue changes such as swelling, oedema and local cellular infiltration.

Local inflammation may induce neural structural changes that may in turn be directly responsible for modulating local electrical changes in the neural tissue. Furthermore, a neuroimmune response is mounted in

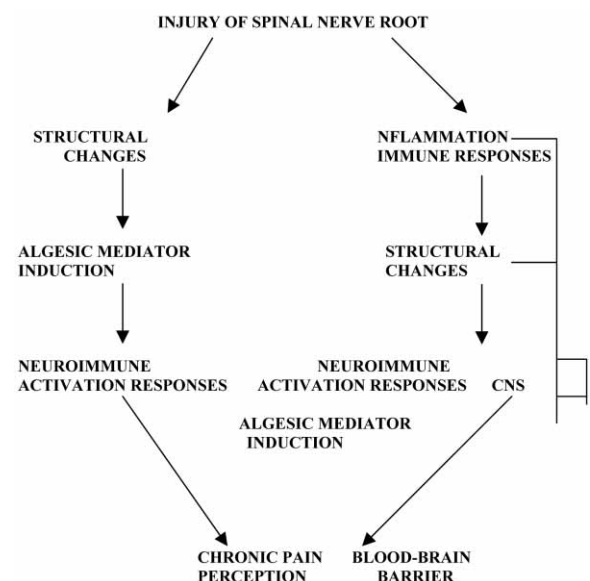


Figure 1. Schematic diagram of the conceptual model of events at injury and afterward that lead to chronic pain. Biomechanical and physiologic responses, as important contributors in this complex cascade of events, have complicated relations with each other and final pathway that elicits pain.

the spinal cord, where neurons and glia cells may produce a number of proinflammatory cytokines, which can in turn induce the expression of numerous algescic mediators that lead to enhanced nociceptive activity, and thus pain. In conjunction with the activation of these cells, cellular adhesion molecules are upregulated and peripheral T cells and macrophages infiltrate across the blood-brain barrier into parenchyma of the spinal cord. This neuro-inflammatory response can itself also enhance the neuro-immune activation response of the CNS and provide enhancement of algescic mediator production. It is important to recognize also that these CNS changes are transmitted to the brain. Affecting supraspinal pain mechanisms, the perception of chronic pain itself may play role in feeding back on the nociceptive responses (26, 27, 28, 33).

Immunologically mediated pathology of intervertebral disc tissue spinal pathology

The intervertebral disc has traditionally been regarded as biomechanically important structure in the spine, with characteristic biomechanical properties related to both the annulus fibrosus and the nucleus pulposus. However, research performed mainly during the last 15 years has revealed that the intervertebral disc cells is also biologically active, and that disc degeneration and herniation is associated with the production of such as pro-inflammatory cytokines (32, 33).

In the disc tissue several inflammatory mediators are produced such as phospholipase, A-2, cyclooxygenase (COX-2), prostaglandine E-2, Tumor, necrosis factor-alpha, nitric oxide, interleukine-1 alpha, interleukine-1 beta, IL-6, IL-8 and the macrophage infiltration into protruded disc material can be the trigger for cytokine production and for continuation of neurological symptoms (19, 25, 34, 35, 36).

However, certain degrees and certain phase(s) modulating immunologically the process of inflammatory cascade of spinal nerves are accompanied by various degrees of vascularisation conditioned by toxic content of degenerative disc tissue in epidural space, and by hyperalgesia (37).

Role of cytokines

It has been shown that herniated cervical and lumbar disc specimens spontaneously produce increased amounts of various substances, such as nitric oxide, interleukin-6, prostaglandin E2 and certain matrix metalloproteinases. Furthermore, the cells of intervertebral discs increase their production in vitro in these substances even more when stimulated by interleukin-1 beta. Increased levels of interleukin-6 and interleukin-8 have been found in disc specimens from patients with discogenic low back pain, indicating a possible role of such substances in the pathogenesis of low back pain. Increased levels of pro-inflammatory cytokines, such as interleukin-1 beta, have been found in fact joint tissues

in patients undergoing surgery for lumbar spinal stenosis and disc herniation. These observations suggest the involvement of pro-inflammatory cytokines in degenerated lumbar facet joints regarding the genesis of pain production.

In addition, intervertebral discs may injure spinal nerve roots not only by mechanical deformation but also by the presence of nucleus pulposus tissue in the epidural space.

Olmarker and colleagues 1993 (20) demonstrated that autologous nucleus pulposus induced histologic and functional changes in spinal nerve roots with no mechanical-compressive components (21). These nonmechanical effects have recently been attributed to cytokines related to intervertebral disc cells. One cytokine in particular that has been suggested to be of major importance in this context is tumor necrosis factor-alpha (TNF-alpha) (42).

TNF-alpha as key player in the initiation processes of the nucleus pulposus induced nerve root injury has been suggested to play a key role in mediating sensitization, as well as other nucleus pulposus-induced effects, such as intravascular coagulation of spinal nerve spinal root and dorsal ganglion oedema (DRG) blood flow reduction through spinal nerves and damage of myelin capsule (13). TNF-alpha is known to be produced and released from the chondrocyte-like cells of the nucleus pulposus (13, 42).

However, other cytokines such as interleukin-1 beta and interferon-gamma have also been suggested to be involved because of their known neurotoxic properties and their presence in the disc cells. Moreover, local application of nucleus pulposus in experimental animals may also induce a characteristic »Inflammatory crescent«, providing histologic confirmation of inflammatory reaction at the surface of dorsal root ganglion (DRG) (13, 20, 21, 22, 36).

Finally, it has been suggested that intervertebral disc may be invaded by newly formed blood vessels and nerve fibers following injury of the annulus fibrosus. The nerve fibers have been considered to induce low back pain. It seems that bioactive substances within the nucleus pulposus may be involved in the induction of a neovascularisation and reinnervation. Such ingrowth seems to be reduced by Doxycycline, Infliximab and Dexamethasone, cytokine inhibitors (21, 22, 23, 25).

Findings have shown that intervertebral disc degeneration also relates to a multitude of factors, among them cytokine-induced increased production of matrix metalloproteinases, a mechanism supported by an observed correlation between the levels of matrix metalloproteinases 2 and 9 and the grade of disc degeneration. initiate intervertebral disc degeneration. In addition, recently, genetic mechanisms underlying disc degeneration have been also identified (3, 4).

To conclude, the pathophysiology of sciatic pain in association with disc herniation has been shown to be related to a combination of biochemical nerve root irritat-

tion via pro-inflammatory cytokines, produced by nucleus pulposus cells and mechanical nerve root compression. The cytokine mediated nerve root inflammation induces a sensitization of nerve root so that mechanical nerve root compression leads to radiating sciatic pain. Animal experimental studies have shown that anti-TNF substances, e.g. infliximab and etanercept, can reduce or prevent TNF induced nerve root changes as well as pain behavioural reactions. Recent clinical studies have also shown positive results, indicating a possible role of anti-TNF treatment in patients with lumbar disc herniation and sciatica (19, 20, 21). Evidently, there is a large body of evidence indicating that pro-inflammatory cytokines and other related substances play important roles in the pathophysiology of various spinal pain conditions.

Role of complement membrane attack complexes

At the tissue level, apoptotic disc cells were identified in samples from anulus fibrosus, particularly in discs from older subjects. Moreover, Lotz *et al.* noted an increasing number of such apoptotic disc cells with increasing compressive stress in mouse tail discs (2).

Several recent studies have supported the concept of antibody-mediated mechanisms in disc pathophysiology, as suggested by the biochemical demonstration of immunoglobulins, and immunoglobulin complexes have been demonstrated by immunohistochemical methods in disc herniation (DH) tissues. Notably, such immune complexes have been never observed in control disc tissue samples. Immunoglobulin-antigen complexes have been observed around nucleus pulposus cells, and possibly in newly formed blood vessels within DH tissue. The role of such immune complexes in disc tissue pathophysiology remained unclear, however (8, 9, 10).

Among the three pathways of complement activation, the so-called classic pathway is activated by antigen-antibody complexes, whereas complement can be also activated alternatively by either microbial cell walls or the lectin pathway, in which microbial carbohydrates interact with mannose-binding protein in the plasma.

As final step in the classic pathway activation, several complement components (C5b, C6, C7, C8, C9) form a membrane attack complex (MAC) that can destroy cells by perforating their membranes. There currently is no conclusive evidence of microbial involvement in intervertebral disc degeneration or herniated disc, but there is indirect evidence suggesting a possibility of complement activation through the classic pathway mediated by immune (antigen-antibody) complexes. In particular, immune complexes juxtaposed with disc cells have been reported. Thus it was pertinent to look for the possibility of a complement-mediated pathway of disc cell destruction, and to determine whether such complement activation, if demonstrated, applies only to pathologic disc tissues (10, 11, 12, 14, 15, 16).

Pharmacotherapy of acute discogenic pain

Nonspecific cytokine inhibition

Anti-inflammatory cytokine therapy may become an effective treatment of sciatica due to disc herniation. Major advancement in the management of „discogenic« back pain will depend upon an appreciation of the importance of controlling neural inflammation in the early phases of the disease rather than developing new techniques of managing irreversible neural lesions and their iatrogenetic sequelae. The pain may occur as a result of compression of adjacent neural structures as well as by release of irritating chemicals.

In selected cases immediate relief from pain occurs after administration of anti-TNF-alpha medication. The important point is that with development of intraneural fibrosis the abnormalities become irreversible and at our present knowledge untreatable.

Thus, the early treatment of low back pain might prevent future development of irreversible local changes.

In approach to the problem of a syndrome acute low back pain due to disc pathology an attempt was made to combine therapeutical protocols for „Acute disc syndrome« and in combination with a cocktail of **Corticosteroids** (by direct inhibition of the expression of leukocyte adhesive molecules and suppression of synthesis of cytokines and blocking the initial steps in receptive transduction of signals into cell), and indirectly-induction of synthesis of macrocortine (**Lipomoduline**) which inhibits activation of phospholipase A2 and creating eicosanide by blocking the initial steps in receptive transduction of signals into cell. **Diuretics**, (by preventing i.e. reduce spinal root and dorsal ganglion oedema), **Doxycycline**, (blocking the activity of TNF-alpha) (42) **Indomethacin** i.e. **Pyroxicame** as equivalent to Indomethacin (by inhibition to neutrofile activation).

Although in recent literature there have been quoted about the attempts of treatment of acute ischiatic pain by use of selective monoclonal inhibition (Infliximab), we consider that it is still not applicable in clinical praxis because of complexity of immunological reaction, pleiotropism of cytokine actions and so far with its of toxicity and side-effects (34).

In conclusion, injury to nerve roots causes profound changes in input from primary afferents innervating the dorsal horn.

Early and timely applicated pharmacotherapy in acute phase of discogenic pain of spine, from a few hours to few days after the injury will prevent the development of syndrome of chronic spinal pain, which may end up with invalidity and suffer for millions in the world.

REFERENCES

1. MORAN F P, KING T 1957 Primary instability of lumbar vertebrae as a common cause of low back pain. *J Bone Joint Surg* 39B: 6–22
2. LOTZ J C, COLLIUO O K, CHIN J R *et al.* 1998 Compression – induced degeneration of the intervertebral disc: An *in vivo* mouse model and finite – element study. *Spine* 23: 2493–506
3. ESSER A F 1994 The membrane attack complex of complement: Assembly, structure and cytotoxic activity. *Toxicology* 87: 229–47
4. HABTEMARIAM A, GRONBLAD M, VIRRI J *et al.* 1996 Immunocytochemical localization of immunoglobulins in disc herniations. *Spine* 16: 1864–9
5. HELM K F, PETERS M S 1993 Deposition of membrane attack complex in cutaneous lesions of lupus erythematosus. *J Am Acad Dermatol* 28: 687–91
6. IGARASHI T, KIKUCHI S, SHUBAYEV V *et al.* 2000 Volvo Award Winner in Basic Science Studies. Exogenous tumor necrosis factor- α mimics nucleus pulposus-induced neuropathology: Molecular, histologic, and behavioral comparisons in rats. *Spine* 25: 2975–80
7. ITAGAKI S, AKIYAMA H, SAITO H *et al.* 1994 Ultrastructural localization of complement membrane attack complex (MAC) like immunoreactivity in brains of patients with Alzheimer's disease. *Brain Res* 645: 78–84
8. KANG J D, STEFANOVIĆ-RAČIĆ M, MCINTYRE L A *et al.* 1997 Toward a biochemical understanding of human intervertebral disc degeneration and herniation: contribution of nitric oxide, interleukins, prostaglandin E₂ and matrix metalloproteinases. *Spine* 22: 1065–73
9. KAWANA S, SHEN G H, KOBAYASHI Y *et al.* 1990 Membrane attack complex of complement in Henoch-Schönlein purpura skin and nephritis. *Arch Dermatol Res* 282: 183–7
10. KILGORE K S, SHEN J P, MILLER B F *et al.* 1995 Enhancement by the complement membrane attack complex of tumor necrosis factor- α -induced endothelial expression of E-selectin and ICAM-1. *J Immunol* 155: 1434–41
11. PHAM B N, MOSNIER J F, DURAND F *et al.* 1995 Immunostaining for membrane attack complex of complement is related to cell necrosis in fulminant and acute hepatitis. *Gastroenterology* 108: 495–504
12. SATOH K, KONNO S, NISHIYAMA K *et al.* 1999 Presence and distribution of antigen-antibody complex in the herniated nucleus pulposus. *Spine* 24: 1980–4.
13. SPILIOPOULOU I, KORVESSIS P, KONSTATINOU D *et al.* 1994 IgG, IgM concentration in the prolapsed human intervertebral disc and sciatica etiology. *Spine* 19: 1320–3
14. TOLONEN J, GRONBLAD M, VIRRI J *et al.* 1995 Basic fibroblast growth factor immunoreactivity in blood vessels and cells of disc herniations. *Spine* 20: 271–6
15. YOUNG J D, YOUNG T M 1990 Channel fluctuations induced by membrane attack complex C5b-9. *Mol Immunol* 27: 1001–7
16. BRISBY H, OLMARKER K, LARSSON K *et al.* 2002 Proinflammatory cytokines cerebrospinal fluid and serum in patients with herniation and sciatica. *Eur Spine J* 11: 62–6
17. OLMARKER K 2001 Radicular pain-recent pathophysiologic concept and therapeutic indication. *Schmerz* 15: 425–9
18. OLMARKER K, RYDEVIK B, NORDBORG C 1993 Autologous nucleus pulposus induces neurophysiologic and histologic changes in porcine cauda equina nerve roots. *Spine* 18: 1425–32
19. ANZAI H, HAMBAM M, ONDA A *et al.* 1996 Epidural application of disc-related cytokines on spinal nerve roots; a morphological and neurophysiological study. *Eur Spine J* 5: 187–92
20. OLMARKER K, STORKSON R, BERGE O G 2002 Pathogenesis of sciatic pain: a study of spontaneous behavior in rats exposed to experimental disc herniation. *Spine* 27: 1312–7
21. AHN S H, CHO Y W, AHN M W *et al.* 2002 mRNA expression of cytokines and chemokines in herniated lumbar intervertebral discs. *Spine* 27: 911–7
22. BURKE J G, WATSON R W, MCCORMACK D *et al.* 2002 Intervertebral disc which cause low back pain secrete high levels of proinflammatory mediators. *J Bone Joint Surg Br* 84: 196–201
23. CHAO C, H Y S 1994 Tumor necrosis factor- α potentiates glutamate neurotoxicity in human fetal brain cell cultures. *Dev neurosci* 16: 172–9
24. COLBURN R, RICKMAN A, DE LEO J 1999 The effect of site and Type of Nerve injury on spinal glial activation and neuropathic pain behaviour *Ep Neurol* 157: 289–304
25. DE LEO J A, COLBURN R W 1996 The Role of Cytokines in Nociception and Chronic Pain. In: Weinstein Jn, Gordon S L (eds.) *Low Back Pain: A Scientific and Clinical Overview*. AAOS Publishers, Rosemont, IL, p 163–85
26. DE LEO J A, YEZIERSKI R P 2001 The Role of Neuroinflammation and neuro Immune Activation in Persistent pain. *Pain* 91: 1–6
27. DUBNER R, HARGREAVES K M 1989 The Neurobiology of Pain and Its Modulation Clin. *J Pain* 2(Suppl): s1–6
28. HASHIZUME H, DE LEO J A, COLBURN R W *et al.* 2000 Spinal Glial Activation and Cytokine Expression after Lumbar Root Injury in the Rat. *Spine* 25: 1206–17
29. ZIMMERMAN M 2001 Pathology of Neuropathic Pain. *Eur J Pharmacol* 429: 23–37
30. WOOLF C J 1983 Evidence for a central Component of Post Injury pain Hypersensitivity. *Nature* 306: 686–8
31. JARO KARPINEN, TIMO KORTONEN, ANTTI MALMI-VAARA, LEENA PAIMELA, SEPPÖ SEITSAALO, HEIKKI HURRI 2003 Treatment of sciatica with infliximab, a monoclonal humanised chimeric antibody against TNF- α ; Abstract book 6th Congress of the European Federation of National Associations of Orthopaedics and Traumatology. Efort 2003 Helsinki, Finland, p 158 03057
32. ABE Y, AKEDA K, AN H S *et al.* 2007 Proinflammatory cytokines stimulate the expression of nerve growth factor by human intervertebral disc cell. *Spine* 32: 635–42
33. AOKI Y, AN H S, TAKAHASHI K *et al.* 2007 Axonal growth potential of lumbar dorsal root ganglion neurons in an organ culture system: response of nerve growth factor-sensitive neurons to neuronal injury and an inflammatory cytokine. *Spine* 32: 857–63
34. HAYASHI S, TAIRA A, INOUE G, OHTORI S *et al.* 2008 TNF- α in nucleus pulposus induces sensory nerve growth. *Spine* 33: 1542–6
35. YAMASHITA M, OHTORI S, KOSHIT T *et al.* 2008 Tumor necrosis factor- α in the nucleus pulposus mediates radicular pain, but not increase of inflammatory peptide, associated with nerve damage in mice. *Spine* 33: 1836–42
36. OHTORI S, INOUE G, KOSHIT T *et al.* 2007 Characteristics of sensory dorsal root ganglia neurons innervating the lumbar vertebral body in rats. *J Pain* 8: 483–8
37. KWON S M, EGUCHI M, WADA M *et al.* 2008 Specific Jagged-1 signal from bone marrow microenvironment is required for endothelial progenitor cell development for neovascularization. *Circulation* 118: 157–65
38. HOMBACH-KLONISCH S, PANIGRAHI S, RASHEDI I *et al.* 2008 Adult stem cells and their trans-differentiation potential-perspectives and therapeutic applications. *J Mol Med* 86: 1301–14
39. LEUNG V Y, HUNG S C, LILIC *et al.* 2008 Age-related degeneration of lumbar intervertebral discs in rabbits revealed by deuterium oxide-assisted MRI. *Osteoarthritis Cartilage* 16: 1312–8
40. ZHAO C Q, WANG L M, JIANG L S *et al.* 2007 The cell biology of intervertebral disc aging and degeneration. *Ageing Res Rev* 6: 247–61
41. TSCHOEKE S K, HELLMUTH M, HOSTMANN A *et al.* 2008 Apoptosis of human intervertebral disc after trauma compares to degenerated disc involving both receptor-mediated and mitochondrial-dependent pathways. *J Orthop Res* 26: 999–1006
42. OLMARKER K, LARSSON K 1998 Tumor necrosis factor alpha and nucleus pulposus induced nerve root injury. *Spine* 23: 2538–44
43. SCUDERI G J, CUELLEAR J, CUELLEAR V 2009 Epidural Interferon Gamma-Immunoreactivity. *Spine* 21: 2311–2317