

International Science and Investigation Journal

ISSN: 2251-8576 **2015, 4(1)**

Journal homepage: www.isijournal.info

Received December 2014; Accepted January 2015

Effects of ultrasonic on microorganisms and enzymes

Bahram Hosseinzadeh Samani¹, Zahra Lorigooini²

¹Department of mechanics of biosystem, Shahrekord University, Shahrekord, Iran.

²Medical Plants Research Center, Shahrekord University of Medical Sciences,

Sharekord, Iran.

*Corresponding author's E-mail: <u>b.hosseinzadehsamani@gmail.com</u>

Abstract

Removing on pathogens is the key concern of non-thermal technology in food products. Recently, inclusion of ultrasound into the food industry has been a subject of research and development. The application of ultrasound in food industry is undoubtedly the notion of consideration and development and, the same as other cases in other areas, the sound ranges can would fall into the following categories of high frequency, low energy, diagnostic ultrasound in the MHz range and low frequency, high energy, power ultrasound in kHz range. The advantageous application of power ultrasound is reflected in its chemical, mechanical, or physical effects on the process or product. Microorganisms Inactivation mechanism, based on physical and chemical factors that ultrasonic was caused on liquid food. When it comes to physical and chemical effects, one would distinguish between them as thinning cell membrane, localized heat and pressure changes (5500°C, 50Mpa); hydroxyl radical and sonochemical reactions, respectively. Ultrasound-assisted inactivation of microorganisms and enzymes brings about the extension of shelf life of raw materials or prepared foods. The study focuses on the related literature having to do with the effect of ultrasonic on microorganism and enzymes.

Keywords: Ultrasonic, Innovation Technology, Pasteurization, Microorganism

Introduction

Ultrasonic is defined as the frequency above 20 kHz. It ascends to MHz range and finally, at around 1 GHz, progresses into what is conventionally called the hypersonic regime, where an average sound velocity is about 5000 m /s. In water, the most widely used liquid, the sound velocity is about 1500 m /s, having wavelengths of 3 mm to 30µm for the above frequency range.

The application of ultrasound in food industry is undoubtedly the notion of consideration and development and, the same as other cases in other areas, the sound

ranges can would fall into the following categories of high frequency, low energy, diagnostic ultrasound in the MHz range and low frequency, high energy, power ultrasound in kHz range. During the expansion cycle, high intensity ultrasonic waves make small bubbles grow in liquid. When such waves achieve a level where there is no energy absorption, violent implode occurs to be called cavitation. What happens during implosion is that very high temperatures (approximately 5000 K) and pressures (estimated at 50000 kPa) are included in these bubbles (Raso et al., 1998).

A number of changes are made by high-intensity acoustic radiation among which is the propagation occurring through a medium. Several mechanism are at work to demonstrate such changes, however, it is not to say that al mechanisms are known.

These changes can be explained by several mechanisms, but not all mechanisms involved are known or well understood. Following is the summary of the reported effects and proposed mechanisms by different scholars:

Heating (Lehmann and Krusen, 1954, Suslick et al., 1985, Fayyazi et al, 2012). Cavitation: (Suslick, 1988, Fayyazi et al, 2012, HosseinzadehSamani, 2014). Structural effects: (Ensminger, 1986).Compression and rarefaction: (Ensminger, 1986, Hosseinzadeh Samani, 2014).Turbulence: (Ensminger, 1988).

The use of ultrasound in food processing is embedded into two basic branches:

1. Monitoring a process or product

2. Affecting a process or product

Power ultrasound

The inclusion of ultrasound in line with improving processes and products is not welcomed by food manufacturing, however, it is considered in other industries. Based on the application, High-intensity sound is commonly used for the applications with frequency either in the sonic (<18 kHz) or ultrasonic (18 kHz) range. The advantageous application of sound is reflected in its chemical, mechanical, or physical effects on the process or product. Sonochemistry, which is a new term to be developed in chemistry, takes the advantage relevant to ultrasonic chemical effects (Suslick, 1986).

Common uses encompass acceleration of conventional and decomposition reactions, degradation of polymers, and polymerization reactions (HosseinzadehSamani, 2014). When sonication is run on particles of material in a liquid, different physical and mechanical effects are brought about.

Large particles are subject to surface erosion (via cavitation collapse in the surrounding liquid) or particle size reduction (due to fission through interparticle collision or the collapse of cavitation bubbles formed on the surface). The results of conducted researches in the field of the effect of sonication on suspended powders have manifested that the particles can be forced into violent collision that, in the case of metals, fusion can occur in some cases, the colliding powders are affected by chemical reaction. Thus, when copper and sulphur are sonicated together in hexane for 1 hour, 65% Cu2S is generated (Goh et al., 1994).

Effect of ultrasonic on microbial and enzyme

Removing on pathogens is the core concern of non-thermal technology in food products. Relying on the related literature one would claim that a number of non-thermal methods can reduce thermal effect and sometimes help destroying microorganisms cell membrane through remobing thermal effect (Piyasena et al., 2003). The use of ultrasonic in food products has been discussed in some review papers (Knorr et al., 2004, Pingret et al., 2013).

Microorganisms Inactivation mechanism, based on physical and chemical factors that ultrasonic was caused on liquid food. When it comes to physical and chemical effects,

one would distinguish between them as thinning cell membrane, localized heat and pressure changes (5500°C, 50Mpa); hydroxyl radical and sonochemical reactions, respectively(Butz and Tauscher, 2002, Piyasena et al., 2003, Valdramidis et al., 2010). The effectiveness and efficiency of ultrasonic in microorganism inactivation rely heavily on physical and chemical properties, the volume of food product, microorganism type, the processing temperature, ultrasound intensity and time exposed (Piyasena et al., 2003, Lee et al., 2009a). Ultrasonic which is known as a potential technology in lione with reducing the amount ofg microorganisms to 5 log cycles enjoying FDA standard has been developed in juices. When compared with conventional thermal method, ultrasonic is not able to remove microorganisms in food products. Attaining considerable inactivation effect necessitates that the process be integrated with other methods such as heat and pressure, called hybrid technology (Guerrero et al., 2005, Lee et al., 2009b).

Ultrasound-assisted inactivation of microorganisms and enzymes brings about the extension of shelf life of raw materials or prepared foods. One would demonstrate that the contribution of ultrasound to diverse microbial samples depends highly on the shape and size of the microorganisms (bigger cells being more sensitive than smaller ones, and coccal forms are more resistant than rod-shaped bacteria), type of cells (gram positive being more resistant than gram negative, aerobic being more resistant than anaerobic), and physiological state (younger cells being more sensitive than older ones, spores being much more resistant than vegetative cells) (Piyasena et al. 2003).

The findings of the studies conducted on the effect of ultrasonic on mircoorganisms and enzyme in different medium have been provided.

Ordonez et al. (1987) showed a decline in the effectiveness of ultrasound on bacteria inactivation at elevated temperatures (50°C-60°C). Garcia et al. (1989) concluded that this efficiency loss could be derived from the rise of vapor pressure in the sonicated medium that impaired or reduced the intensity of cavitational collapse, which was avoided by increasing the applied pressure of the sonicated medium. Monothermosonication (MTS) is a method in which ultraonication is combined with thermal and pressure inducing. Lopez et al. (1994) found that the monothermosonication treatment was much more efficient when compared with heat treatment alone aiming to inactivate enzymes such as *lipoxygenase*, POD, and PPO in buffer systems. The combination of ultrasound with chlorine treatment seemed to decline in 4-log of Salmonella on poultry surfaces (Lillard 1994). Proteases and lipases are found to be the affective factors for the shelf life of ultraheat-treated milk. These enzymes were inactivated up to 10 times faster employing monothermosonication treatment (Vercet et al. 1997). The capability of ultrasound to inactivatingobserved pathogenic microorganisms such as L. monocytogenes, a number of strains of Salmonella spp., E. coli, or S. aureus, which are increasingly found in outbreaks of food poisoning, has been noticed (Sala et al., 1995, Pagan et al. 1999). Pectin methylesterase from orange to be regarded as a thermostable enzymewas inactivated almost 500 times faster by monothermosonication treatment when compared with heat treatment at the same temperature (Vercet et al. 1999). The success of ultrasound application has been reported at ambient pressure in line with inactivatin food-quality-based enzymes. POD was inactivated by integrating heat with ultrasound at neutral (Gennaro et al. 1999) or low pH (Ku et al. 2000) and lipoxygenase was inactivated at low sonication intensities (Thakur and Nelson 1997).the combination of flow ultrasound treatment and mild heat (57 $^{\circ}$ C) for 18 min yielded 5-log reduction of *L. monocytogenes* in milk, a 5-log reduction in total aerobic bacteria in raw milk and a 6-log reduction in E. coli O157:H7 in pasteurized apple

cider Villamiel. Jong (2000) found no contribution of ultrasoundto endogenous milk enzymes such as alkaline phosphatase, g-glutamyltranspeptidase and lactoperoxidase at room temperature,however, synergistic inactivation was found to exist at higher temperatures ($60^{\circ}C-75^{\circ}C$).In another experiment, Seymour et al. (2002) reported microbial decontamination on the surface of minimally processed fruits and vegetables (lettuce, cucumber, carrots, parsley, and others) by ultrasound. In this research, the microorganisms were subsequently killed by inclusion of chemical sanitizers such as chlorine. Pectin methylesterase and polygalacturonase in tomato juice were also inactivated by monothermosonication treatment with higher efficiency (Vercet et al. 2002). Other researches such as Scouten and Beuchat(2002) indicated the decontamination of alfalfa seeds inoculated with *Salmonella* or *E. coli* O157 by combined treatments of ultrasound and Ca (OH) 2, which could be taken into account as an alternative to chlorine treatments so as to ignore contamination.

The activity of *pectinesterase* (PE) at themoderate pressure (100-300 kPa) of temperature below 100°C can be affected by Monothermosonication (Kuldiloke *et al.*, 2002).

A continuous ultrasound system accompanied by steam injection has emitted to fourfold higher inactivation rates of E. coli and Lactobacillus acidophilus in several liquid foods such as milk and fruit juices (Zenker et al. 2003). Guerrero et al. (2005) highlighted that the inactivation of S. cerevisiaecould be enhanced by incubating with low molecular weight chitosan prior to including ultrasound.Ugarte et al. (2006) investigated the application of acoustic energy to secure apple juice safety. Sonication was found to give rise to *E. coli* K12 cell destruction by 5.3-log, 5.0-log, and 0.1-log cycles at 40°C, 50°C, and 60°C, respectively. D'Amico et al. (2006) asserted that ultrasound treatments distinguished by inclusion or not including mild heating were effective in reducing L. monocytogenes in raw milk and E. coli O157:H7 in apple cider. Ferrante et al. (2007) reported that that the control of L. monocytogenesin orange juice could be developed by integrating high intensity ultrasound with mild heat treatment and natural antimicrobials.Salleh-Mack and Roberts, (2007) in their research studied the effects of temperature, sugar concentration (8, 12, and 16 g/100 ml), organic acids (citric and malic acids) and pH (2.5 and 4.0) on ultrasound pasteurization. In this research, E.coli was used as model organism. Based on the obtained results one would claim that ultrasound increased the sensitivity of E. coli to thermal inactivation. The presence of soluble solids had a protective effect where the sonication time requirement increased. Similar to heat sensitivity, the lower pH environment resulted in E. coli having less resistance to sonication (Salleh-Mack and Roberts, 2007). In the study conducted by Valero et al (2007) the influence of ultrasound and conventional heating under different processing conditions on the inactivation and potential subsequent growth of micro-organisms in orange juice was highlighted. This research gave its attention to ultrasonic parameter adjusting as frequency 500 kHz, power 240 W and exposed time 15 min. the findings revealed that the presence of pulp in the juice increased the resistance of micro-organisms to ultrasound. The combination of ultrasound and pressure and/or heat shows considerable promise for the inactivation of microorganisms and enzymes. Hence, techniques such as thermosonication, monosonication, and monothermosonication may be of more relevance in the future as an energy-efficient processing alternative for the food industry. Another research made use of thermosonication (TS) method categorized by frequency 24kH, power 400W, at amplitudes of 25, 50 and 75µm at 60, 65 and 70°C or heat only treatments on inactivation *pectinmethylesterase* in tomato juice. the obtained results indicated that the TS treatment at 60 $^{\circ}$ C, 65 $^{\circ}$ C and 70 $^{\circ}$ C

for 41.8, 11.7 and 4.3 min exposure, respectively reduced *pectinmethylesterase* (PME) activity by 90% (Wu *et al.*, 2008).

Another experiment relied on investigating the impact of thermosonication (TS) and pulsed electric field (PEF), individually and combined, on the survival of *Listeria innocua* 11288 (NCTC) in milk. TS (400 W, 160 s) without pre-heating declined*L. innocua* by 1.2 log10 cfu mL⁻¹, while shorter treatment times produced negligible inactivation, showing that TS was a hurdle rather than an effective standalone treatment (Noci et al, 2009).

Tiwari et al (2009) drew his attention to investigating the effect of sonication on pectinmethylesterase (PME) activity and cloud stability of orange juice .Ultrasonic acoustic energy density (AED) levels of 0.42, 0.47, 0.61, 0.79 and 1.05 W/mL and treatment times of 0(Control), 2, 4, 6, 8 and 10 min were investigated. The highest PME inactivation level noticed was 62% for sonication at the highest AED level and treatment time. These findings highlighted that the cloud stability of sonicated orange juice depends both on PME inactivation and particle size reduction. Industrial relevance: Power ultrasound is taken into account as a non-thermal pasteurization method that has been found to enjoy the US FDA requirement for a 5 log reduction in E. coli relevant to fruit juices. another research focused on the effect of Ultrasonic treatment 20 kHz, monosonication, thermosonication, monothermosonication at temperature of 40, 37, 54 and 61°C and pressure of 100, 300, 400 and 500 kPa on exposed time of 4 min and conventional thermal method on *E.coli* inactivation. Derived conclusions from the study revealed that the mixed method could decrease inactivation time significantly (Lee *et al.*, 2009a). Recently, the research conducted by Pulsed light (PL) and Thermosonication (TS) has been reported to be applied alone or in combination making use of a continuous system to investigate their effect on Escherichia coli inactivation in apple juice (Muñoz, et al, 2012).another in-field experiment relied itself on surveying possible effects of ultrasound probe diameter, reactor diameter, and juice level in the reactor upon effectiveness of ultrasound waves on decontamination of sour cherry juice. The obtained results demonstrated that the contribution of probe diameter, reactor diameter and reactor height to ultrasonic effect were significant t (P < 0.01) (Hosseinzadeh Samani et al. 2013). The impacts of temperature (20–52 °C), acoustic intensity (60–120 W/cm²) and treatment time (40– 240 s) at a constant pressure (225 kPa) was the focus of another study through employing monothermosonication processing on microbial inactivation. Escherichia coliand Pseudomonas fluorescens showed to have increased by c. 1.6 log CFU/ml. Staphylococcus aureuswas reported to have lower inactivation values (1.05 log CFU/ml). One would obtain such values on the condition of 36 °C, 90 W/cm2 and 240 s (Cregenzán-Alberti et al, 2014). Other relevant researches include Blume and Nies (2003) (Facal coliforms and streptococci in municipal wastewater), Duckhouse et al (2004) (E.coli in Saline), Furuta et al (2004) (E.coli in water), Tsukamoto et al (2004) (Cryptosporidium parvaum in Water), Lo"rincz (2004) and Tsukamoto et al (2004) (Saccharomyces cerevisiae in Water), Lo pez-Malo et al (2005)(Aspergillusflavus and *Penicilliumdigitatum* in Sabouraud growth medium), Raviyanet al (2005) (pectinmethylesterase in tomato), Zhang et al (2006) (Microcystisaeruginosain BG11 growth medium), Dadjour et al (2006) (Legionella pneumophilain diluted medium), Alvarez et al (2006) (Salmonella Senftenberg 775W in McIlvaine citrate-phosphate buffer or nutrient broth), Borthwick et al (2005), Lee at al (2009) (E.coliin phosphate buffer) Hosseinzadeh Samani (2014) (E.coli and Saccharomyces cerevisiae in Sour Cherry).

Conclusion and recommendations

Based on the obtained results one would assert that it is feasible to make reduction in thermal impact through employing ultrasonic method where its addition to thermal effect would bring about destroying in microorganisms cell membrane intensity. The required time to do the operations is limited in this method, however, it necessitates to be mixed with other existing methods so as to reduce the time of processing and to give rise to ultrasonic effect. Having provided the related review of literature, we have made a number of suggestions regarding how to use the ultrasonic method. When it comes to manipulate the pasteurization systems, it is necessary to mix ultrasonic with heat and pressure. In addition, making use of optimized ultrasonic device e.g. probe and reactor dimensions is further suggested to achieve optimum ultrasonic effect.

References

- Álvarez, I., et al. (2006). "Inactivation of Salmonella Senftenberg 775W by ultrasonic waves under pressure at different water activities." International journal of food microbiology 108(2): 218-225.
- Borthwick, K., et al. (2005). "Development of a novel compact sonicator for cell disruption." Journal of microbiological methods 60(2): 207-216.
- Butz, P. and B. Tauscher (2002). "*Emerging technologies: chemical aspects*." Food research international 35(2): 279-284.
- Cregenzán-Alberti, O., et al. (2014). "Suitability of ccRSM as a tool to predict inactivation and its kinetics for Escherichia coli, Staphylococcus aureus and Pseudomonas fluorescens in homogenized milk treated by manothermosonication (MTS)." Food Control 39: 41-48.
- D'amico, D. J., et al. (2006). "Inactivation of microorganisms in milk and apple cider treated with ultrasound." Journal of Food Protection® 69(3): 556-563.
- De Gennaro, L., et al. (1999). "The use of ultrasound in food technology I: inactivation of peroxidase by thermosonication." Journal of Food Engineering 39(4): 401-407.
- Duckhouse, H., et al. (2004). "*The effect of sonication on microbial disinfection using hypochlorite*." Ultrasonics sonochemistry 11(3): 173-176.
- Ensminger, D. and Battelle (1988). "Acoustic and electroacoustic methods of dewatering and drying." Drying Technology 6(3): 473-499.
- Farshbaf Dadjour, M., et al. (2006). "Disinfection of Legionella pneumophila by ultrasonic treatment with TiO₂." Water research 40(6): 1137-1142.
- Fayyazi, E., et al. (2012). "Modelling and evaluation of some effective parameters on reactor design for optimized utilization of ultrasonic waves." Power 1: 4.
- Ferrante, S., et al. (2007). "Combined use of ultrasound and natural antimicrobials to inactivate Listeria monocytogenes in orange juice." Journal of Food Protection® 70(8): 1850-1856.
- Furuta, M., et al. (2004). "Inactivation of Escherichia coli by ultrasonic irradiation." Ultrasonics sonochemistry 11(2): 57-60.
- Garcia, M. L., et al. (1989). "Effect of heat and ultrasonic waves on the survival of two strains of Bacillus subtilis." The Journal of applied bacteriology 67(6): 619-628.
- Goh, N. K., et al. (1994). "Investigations of the effects of ultrasound on some metal and nonmetal systems." Ultrasonics sonochemistry 1(1): S41-S44.
- Guerrero, S., et al. (2001). "Effect of ultrasound on the survival of < i> Saccharomyces cerevisiae: influence of temperature, pH and amplitude." Innovative Food Science & Emerging Technologies 2(1): 31-39.

- Hosseinzadeh Samani, B., Khoshtghaza, M. H., Minaee, S., Abbasi, S. 2013. "*Effect* of ultrasonic waves on pasteurization of sour cherry juice". International Journal of Bioscience, 3: 193-200.
- Hosseinzadeh Samani, B. 2014. "Design, Development and Testing of an Automatic Fruit-juice Pasturization System Using Microwave – Ultrasonic Waves." Ph.D thesis. Department of Mechnics of agricultural machinery, Tarbiat modares University, Iran.
- Knorr, D., et al. (2004). "Applications and potential of ultrasonics in food processing." Trends in Food Science & Technology 15(5): 261-266.
- Ku, J.-Y., et al. (2000). "*Inactivation of peroxidase by hurdle technology*." Food Science and Biotechnology 9(2): 124-129.
- Kuldiloke, J. (2002). "Effect of ultrasound, temperature and pressure treatments on enzyme activity and quality indicators of fruit and vegetable juices", M.Sc. Thesis, Institute of Food Technology Food Biotechnology and Process Technology the Technical University of Berlin.
- Lee, H., et al. (2009). "Effect of pH on inactivation of Escherichia coli K12 by sonication, manosonication, thermosonication, and manothermosonication." Journal of food science 74(4): E191-E198.
- Lee, H., et al. (2009). "Inactivation of Escherichia coli cells with sonication, manosonication, thermosonication, and manothermosonication: Microbial responses and kinetics modeling." Journal of Food Engineering 93(3): 354-364.
- Lehmann, J. and F. Krusen (1954). "Effect of pulsed and continuous application of ultrasound on transport of ions through biologic membranes." Archives of physical medicine and rehabilitation 35(1): 20-23.
- Lillard, H. (1994). "Decontamination of poultry skin by sonication: Ultrasonic applications in the food industry." Food technology 48(12): 72-73.
- Lo" rincz, A. (2004). "Ultrasonic cellular disruption of yeast in water-based suspensions." Biosystems engineering 89(3): 297-308.
- López-Malo, A., et al. (2005). "Multifactorial fungal inactivation combining thermosonication and antimicrobials." Journal of food engineering 67(1): 87-93.
- Lopez, P., et al. (1994). "Inactivation of peroxidase, lipoxygenase, and polyphenol oxidase by manothermosonication." Journal of Agricultural and Food Chemistry 42(2): 252-256.
- Muñoz, A., et al. (2012). "Effects on Escherichia coli inactivation and quality attributes in apple juice treated by combinations of pulsed light and thermosonication." Food Research International 45(1): 299-305.
- Neis, U. and T. Blume (2003). "Ultrasonic disinfection of wastewater effluents for high-quality reuse." Water Recycling in the Mediterranean Region 3(4): 261-267.
- Noci, F., et al. (2009). "Effect of thermosonication, pulsed electric field and their combination on inactivation of Listeria innocua in milk." International Dairy Journal 19(1): 30-35.
- Pagán, R., et al. (1999). "Bacterial resistance to ultrasonic waves under pressure at nonlethal (manosonication) and lethal (manothermosonication) temperatures." Applied and environmental microbiology 65(1): 297-300.
- Pingret, D., et al. (2013). "*Ultrasound assisted Extraction*." Natural Product Extraction: Principles and Applications 21: 89.
- Piyasena, P., et al. (2003). "Inactivation of microbes using ultrasound: a review." International journal of food microbiology 87(3): 207-216.
- Raso, J., et al. (1998). "Influence of temperature and pressure on the lethality of ultrasound." Applied and environmental microbiology 64(2): 465-471.

- Raviyan, P., et al. (2005). "Ultrasonication for tomato pectinmethylesterase inactivation: Effect of cavitation intensity and temperature on inactivation." Journal of food engineering 70(2): 189-196.
- Sala, F., et al. (1995). "Effect of heat and ultrasound on microorganisms and enzymes". New methods of food preservation, Springer: 176-204.
- Salleh-Mack, S. and J. Roberts (2007). "Ultrasound pasteurization: The effects of temperature, soluble solids, organic acids and pH on the inactivation of Escherichia coli ATCC 25922." Ultrasonics sonochemistry 14(3): 323-329.
- Samani, B. H., et al. (2013). "Effect of ultrasonic waves on pasteurization of sour cherry juice." International Journal of Biosciences (IJB) 3(12): 193-200.
- Scouten, A. and L. Beuchat (2002). "Combined effects of chemical, heat and ultrasound treatments to kill Salmonella and Escherichia coli O157: H7 on alfalfa seeds." Journal of Applied Microbiology 92(4): 668-674.
- Seymour, I., et al. (2002). "Ultrasound decontamination of minimally processed fruits and vegetables." International journal of food science & technology 37(5): 547-557.
- Suslick, K. S., et al. (1985). "Determination of local temperatures caused by acoustic cavitation." IEEE 1985 Ultrasonics Symposium, IEEE.
- Thakur, B. and P. Nelson (1997). "Inactivation of lipoxygenase in whole soy flour suspension by ultrasonic cavitation." Food/Nahrung 41(5): 299-301.
- Tiwari, B. K., et al. (2009). "Effect of sonication on orange juice quality parameters during storage." International Journal of Food Science & Technology 44(3): 586-595.
- Tsukamoto, I., et al. (2004). "Inactivation of Saccharomyces cerevisiae by ultrasonic irradiation." Ultrasonics sonochemistry 11(2): 61-65.
- Ugarte Romero, E., et al. (2006). "Inactivation of Escherichia coli with power ultrasound in apple cider." Journal of food science 71(2): E102-E108.
- Valdramidis, V., et al. (2010). "Quantitative modelling approaches for ascorbic acid degradation and non-enzymatic browning of orange juice during ultrasound processing." Journal of Food Engineering 96(3): 449-454.
- Valero, M., et al. (2007). "Effects of ultrasonic treatments in orange juice processing." Journal of Food Engineering 80(2): 509-516.
- Vercet, A., et al. (1999). "Inactivation of heat-resistant pectinmethylesterase from orange by manothermosonication." Journal of Agricultural and Food Chemistry 47(2): 432-437.
- Vercet, A., et al. (2002). "The effects of manothermosonication on tomato pectic enzymes and tomato paste rheological properties." Journal of Food Engineering 53(3): 273-278.
- Wu, J., et al. (2008). "Effect of thermosonication on quality improvement of tomato juice." Innovative Food Science & Emerging Technologies 9(2): 186-195.
- Zenker, M., et al. (2003). "Application of ultrasound-assisted thermal processing for preservation and quality retention of liquid foods." Journal of Food Protection® 66(9): 1642-1649.
- Zhang, G., et al. (2006). "Ultrasonic frequency effects on the removal of Microcystis aeruginosa." Ultrasonics sonochemistry 13(5): 446-450.