

# Mucosal interleukin-21 mRNA expression level is high in patients with *Helicobacter pylori* and is associated with the severity of gastritis

NADER BAGHERI<sup>1</sup>, FATEMEH AZADEGAN-DEHKORDI<sup>2</sup>, MAHSA SHIRZAD<sup>3</sup>,  
BENHAM ZAMANZAD<sup>2</sup>, GHORBANALI RAHIMIAN<sup>4</sup>, AFSHIN TAGHIKHANI<sup>4</sup>,  
MAHMOUD RAFIEIAN-KOPAEE<sup>5</sup>, HEDAYATOLLAH SHIRZAD<sup>2</sup>

<sup>1</sup>Department of Immunology, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran

<sup>2</sup>Cellular and Molecular Research Centre, Shahrekord University of Medical Sciences, Shahrekord, Iran

<sup>3</sup>Faculty of Dentistry, Isfahan University of Medical Sciences, Isfahan, Iran

<sup>4</sup>Department of Internal Medicine, Shahrekord University of Medical Sciences, Shahrekord, Iran

<sup>5</sup>Medical Plants Research Centre, Shahrekord University of Medical Sciences, Shahrekord, Iran

## Abstract

*Helicobacter pylori* (*H. pylori*) infection is associated with gastritis and marked infiltration of the gastric mucosa by several cytokines secreting inflammatory cells. Different clinical forms of the infection may reflect distinctive patterns of cytokine expression. Interleukin (IL)-17, IL-21, IL-22, and IL-23 have been reported to be involved in *H. pylori*-induced gastric mucosal inflammation, but the details and relationship to different patterns of inflammation and virulence factors remain unclear. The present study was launched to analyse IL-6 expression in *H. pylori*-infected and uninfected gastric patients and to investigate its correlation with chronic gastritis among *H. pylori*-infected patients. Total RNA was extracted from the gastric antrum biopsies of 48 *H. pylori*-infected patients and 38 *H. pylori* uninfected patients. Mucosal IL-21 mRNA expression level in *H. pylori*-infected and uninfected gastric biopsy was determined by real-time PCR. The presence of *vacA* (vacuolating cytotoxin A) and *cagA* (cytotoxin associated gene A) virulence factors were evaluated using PCR. Interleukin-21 mRNA expression was significantly high in biopsies of *H. pylori*-infected patients compared to *H. pylori* uninfected patients, and the mucosal IL-21 mRNA level was positively correlated with the grade of chronic inflammation. There was no association between virulence factors and IL-21 mRNA expression. We believe that IL-21 might be involved in the pathogenesis of *H. pylori* and might be an index of the severity of chronic gastritis.

**Key words:** *Helicobacter pylori*, gastritis, IL-21, virulence factors.

(*Centr Eur Immunol* 2015; 40 (1): 61-67)

## Introduction

Colonisation by *Helicobacter pylori* is known to cause chronic gastritis and leads to the development of severe gastroduodenal diseases such as peptic ulcer, lymphoma of the mucosa associated lymphoid tissue (MALT), or gastric adenocarcinoma [1]. *Helicobacter pylori* infection is associated with gastritis and considerable infiltration of neutrophils, monocytes, and lymphocytes into the gastric mucosa, which contribute to maintaining and expanding local inflammation. Activation and migration of these in-

flammatory cells into the gastric mucosa is related to increased production of pro-inflammatory cytokines, which are believed to contribute in maintaining the gastric inflammation and causing epithelial cell damage [2-4]. The risk of different clinical expression of *H. pylori* infection is thought to rely on interaction between the host genetic factors and bacterial factors. For instance, polymorphisms of pro-inflammatory cytokine genes such as interleukin (IL)-8, IL-10, IL-17, IL-1 $\alpha$ , and tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ), which enhance inflammatory response of gastric mucosa and have been correlated to an increased risk of

Correspondence: Prof. Hedayatollah Shirzad, Cellular and Molecular Research Centre, Shahrekord University of Medical Sciences, phone +98-9131859510, fax +98-3813330709, e-mail: shirzad1951@yahoo.com or shirzadeh@SKUMS.ac.ir

gastric cancer and peptic ulcer [5, 6]. It has been shown that the mucosal levels of several cytokines were significantly higher in *H. pylori*-infected patients with virulence factors in comparison to *H. pylori*-infected patients without virulence factors [7, 8]. It is suggested that the clinical outcome of *H. pylori* infection is linked to certain strains such as the cytotoxin-associated gene (*cagA*) and vacuolating cytotoxin (*vacA*) [9]. The *cagA* gene, which is a marker for the presence of a pathogenicity island (*cagPAI*), has been shown to be involved in induction of pro-inflammatory chemokine release, which is also associated with more severe gastritis and higher prevalence of peptic ulcer and gastric cancer [10-12]. Vacuolating cytotoxin A has two variable parts: the s (signal peptide) region exists as allele s1 or s2; and the m (middle) region occurs as m1 or m2 allelic types. The allele combination of s1 m1 confers high, the s1 m2 intermediate, and the s2 m2 low toxin activity. The *vacA* s1 strain is mostly *cagA*-positive, thus the two markers are closely related [13]. There are geographic differences between *vacA* status and *H. pylori*-related diseases. In Western countries infection with *vacA* s1 strain is more common in patients with peptic ulcer than in those with chronic gastritis. However, in Asian populations the association between *vacA* diversity and clinical outcome is not established [14]. Interleukin-21 (IL-21) is a four-helix bundle cytokine that is produced by activated CD4<sup>+</sup> T cells and natural killer T (NKT) cells [15, 16]. T follicular helper (T<sub>fh</sub>) cells also produce IL-21, which regulates generation of humoral responses and germinal centre formation [17]. Interleukin-21 can stimulate non-immune cells to synthesise various inflammatory molecules [15]. Helper T cells that produce IL-17 ('TH-17 cells') are distinct subsets of pro-inflammatory cells whose *in vivo* function requires IL-23 but whose *in vitro* differentiation requires only IL-6 and transforming growth factor- $\beta$ 1 (TGF- $\beta$ 1). Studies have demonstrated that IL-6 induces expression of IL-21 that amplifies an autocrine loop to induce more IL-21 and IL-23 receptors in naive CD4<sup>+</sup> T cells. Both IL-23 and IL-21, along with TGF- $\beta$ 1, induce IL-17 expression independently of IL-6. Interleukin-21 and IL-23 induce the orphan nuclear receptor ROR $\gamma$ t, which in synergy with STAT3 enhances IL-17 expression. 'TH-17' cells produce IL-17, IL-17F, IL-21, and IL-22, which are thought to be required for the control of a variety of bacterial and fungal infections at mucosal surfaces [18, 19]. High levels of IL-21 expression occur in human Th1-associated diseases, and administration of IL-21 to mice enhances the influx of immune cells into inflamed tissues and the severity of immune-mediated diseases [20, 21]. In the light of this study, it is conceivable that IL-21 may play an important role in the initiation and perpetuation of chronic inflammatory processes and may be associated with virulence factors. We have therefore sought to determine the role of IL-21 in *H. pylori*-associated gastritis. We firstly examined IL-21 in the gastric mucosa of patients with

*H. pylori* infection. Secondly, we evaluated the effect of virulence factors *cagA* and *vacA* allelic variants in *H. pylori*-infected on the mucosal IL-21 mRNA level in gastric mucosa. Finally, we determined the correlation between mucosal IL-21 mRNA levels and grade of gastritis.

## Material and methods

### Patients and sampling

A total of 86 subjects undergoing endoscopy at Hajar University Hospital, in Shahrekord, Iran, were included in the study. The study was approved by the Human Research Ethics Committee of Shahrekord University of Medical Sciences, and informed consent was obtained from each volunteer before participation. The groups were characterised as follows: the *H. pylori*-positive group ( $n = 48$ : 16 males, 32 females; mean age:  $42.4 \pm 14.82$  years); the *H. pylori*-negative group ( $n = 38$ : 18 males, 20 females; mean age:  $38.18 \pm 16.51$  years). *Helicobacter pylori* infection was determined by the rapid urease test, PCR (16S rRNA and *glmM*) and histological examination of biopsies taken from the antrum. Patients were classified as *H. pylori* infected only if the four tests were positive. Four biopsies were collected from *H. pylori*-infected patients, and used for rapid urease test, histological examination, assessment of bacterial virulence factors, detection of *H. pylori*, and cytokine RNA analysis. This study was approved by the Ethics Committee of Shahrekord University of Medical Sciences, Shahrekord, Iran.

### Histological examination

Sections of biopsy specimens were embedded in 10% buffered formalin and stained with Haematoxylin and Eosin to examine gastritis, and with Giemsa to detect *H. pylori*. The histological severity of gastritis (acute and chronic) was blindly graded from normal to severe based on the degree of mononuclear cell (MNC) and polymorphonuclear leukocyte (PMN) infiltration according to the Updated Sydney system [22] on a four-point scale: 0, no; 1, mild; 2, moderate; and 3, severe changes. The grade of chronic and active inflammation was investigated for involvement according to the density of neutrophils and mononuclear in gastric mucosal crypts, from one to all crypts.

### Polymerase chain reaction amplification

DNA for polymerase chain reaction (PCR) was extracted using the Biospin Tissue Genomic DNA Extraction Kit (BioFlux, Japan). Specific primers for PCR amplification of different genes are shown in Table 1. For *vacA* and *cagA* evaluation, the PCR program comprised 35 cycles of denaturation (at 94°C for 30 s), annealing (at 56°C for 30 s, extension at 72°C for 30 s), and one final extension (at 72°C for 5 min).

**Table 1.** Polymerase chain reaction primers for amplification of virulence factors

| Size of PCR product (bp)   | Primer sequence  | Primer designation |
|----------------------------|--|--------------------|
| 567 bp (m1)<br>642 bp (m2) | vacAmF: 5-CAATCTGTCCAATCAAGCGAG-3<br>vacAmR: 5-GCGTCTAAATAATCCAAGG-3 | vacA m1/m2         |
| 259 bp (s1)<br>286 bp (s2) | VA1-F: 5-ATGGAAATACAACAAACACAC-3<br>VA1-R: 5-CTGCTTGAATGCGCCAAAC-3   | vacA s1/s2         |
| 232 bp                     | cag1: 5-ATGACTAACGAACTATTGATC-3<br>cag2 5-CAGGATTTTTGATCGCTTTATT-3   | cagA               |

### Quantitative analysis for interleukin-21 mRNA in the gastric mucosa using real-time polymerase chain reaction

Total RNA was isolated from whole gastric biopsy specimens using total RNA extraction biozol (Bioflux, Japan). An aliquot containing 0.2 µg of total RNA was used for the reverse transcription reaction, which was conducted using the superscript first-strand cDNA synthesis system (Fermentas, Finland) according to the manufacturer's instructions. The sequences of oligonucleotide primer and probe are shown in Table 2. The quantification of IL-21 mRNA levels was performed using a Rotor-Gene 3000 (Corbett). Real-time PCR reactions were performed in a total volume of 25 µl, containing 3 µl of synthesised cDNA solution, 12.5 µl of 2x Rotor-Gene Probe PCR Master Mix (Qiagen, Germany), 500 nM of each primer, and 250 nM of the TaqMan probe. Amplification program included a pre-warming step (10 min at 94°C), denaturation step (94°C for 15 s), and an annealing/extension step (60°C for 60 s). The expression levels of cytokine mRNA were expressed as the ratio of cytokine mRNA to β-actin mRNA (cytokine mRNA/β-actin mRNA). Each assay was performed in duplicate, and each cytokine assay was performed testing all RNA samples in the same experiment. Relative quantification of cytokine to β-actin (cytokine mRNA/β-actin mRNA) was determined using the  $2^{-\Delta Ct} = 2^{-(Ct, IL-21 - Ct, \beta-actin)}$  method.

### Statistical analysis

Data were analysed using GraphPad Prism 5 Demo (GraphPad Software, San Diego, California, USA). Data are expressed as raw and mean if not stated otherwise. Age was analysed by unpaired Student's *t*-test. In statistics, normality tests were used to determine whether a data set was well modelled by a normal distribution or not, cytokine expression was presented as mean, and differences between infected and uninfected groups were analysed using Student's *t*-test. Also for gene expression values between *H. pylori*-infected patients and virulence factors the parametric Student's *t*-test was used, and for comparison of more than two groups one-way ANOVA test was used. Categorical data (between virulence factors) are presented as frequencies; comparisons were made using the  $\chi^2$  or

**Table 2.** Primer and probe sequences employed in this study

| Gene    | Primer and probe sequence  |
|---------|--|
| β-actin | Forward 5-AGCCTCGCCTTTGCCGA-3<br>Reverse 5-CTGGTGCCTGGGGCG-3<br>Probe FAM-CCGCCGCCG<br>TCCACACCGCC-TAMRA                           |
| IL-21   | Forward 5- TGTGAATGACTTGGTCCCTGAA-3<br>Reverse 5- AACAGGAAAAAGCTGACCACTCA-3<br>Probe FAM-TCTGCCAGCTCCAGAAGATG<br>TAGAGACAAAC-TAMRA |

Fisher exact test if < 5 samples were in one group. *P* values < 0.05 were considered to be statistically significant.

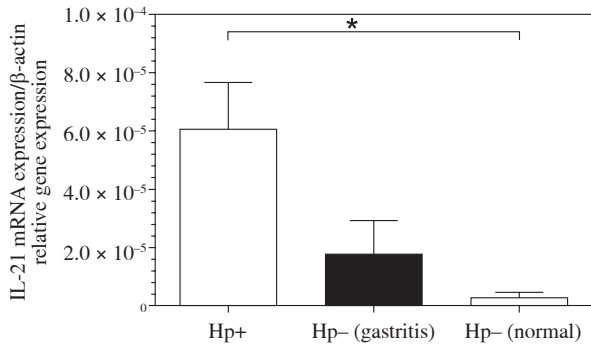
## Results

### Correlation of vacA alleles and presence of cagA gene

The correlation of s1 and s1m2 genotypes with cagA positive was statistically significant, and no correlation of m1 genotype was observed with cagA. The cagA gene was detected in 60.4% (29 strains) of *H. pylori*. The association between vacA alleles and the presence of virulence factors

**Table 3.** Correlation of vacA alleles with the cagA genotype of the samples studied

| vacA genotype  | cagA Positive | Negative |
|----------------|---------------|----------|
| s1m1           | 7             | 4        |
| s1m2           | 16            | 6        |
| s2m2           | 2             | 6        |
| <i>P</i> value |               | 0.06     |
| s1             | 26            | 11       |
| s2             | 2             | 6        |
| <i>P</i> value |               | 0.018    |
| m1             | 7             | 4        |
| m2             | 22            | 14       |
| <i>P</i> value |               | 0.881    |



**Fig. 1.** Relative expression level of interleukin (IL)-21 mRNA in gastric mucosa shown by *Helicobacter pylori* status. RNA was extracted from gastric biopsies of 48 *H. pylori*-infected patients, 26 *H. pylori*-negative patients with gastritis, 12 *H. pylori*-negative normal mucosa; and analysed for IL-21 by real-time polymerase chain reaction. Levels are normalised to β-actin. Statistical significance was evaluated using the one-way ANOVA test (\* $p < 0.05$ )

is described in Table 3. Out of 29 *H. pylori* subjects that were positive for cagA, 26 strains were associated with the toxin-producing vacA s1, and only 2 cagA-positive strains were vacA s2 ( $p < 0.018$ ). Sixteen strains were associated with the toxin-producing vacA s1m2, only 7 strains were vacA s1m2, and 2 strains were vacA s2m2 ( $p < 0.06$ ).

**Interleukin-21 expression in *Helicobacter pylori*-infected gastric mucosa**

Interleukin-21 expression was significantly greater in biopsy specimens of *H. pylori* infected patients compared with *H. pylori* negative (Fig. 1). If the mean in the *H. pylori*-positive group is 0.00006 and the mean in the

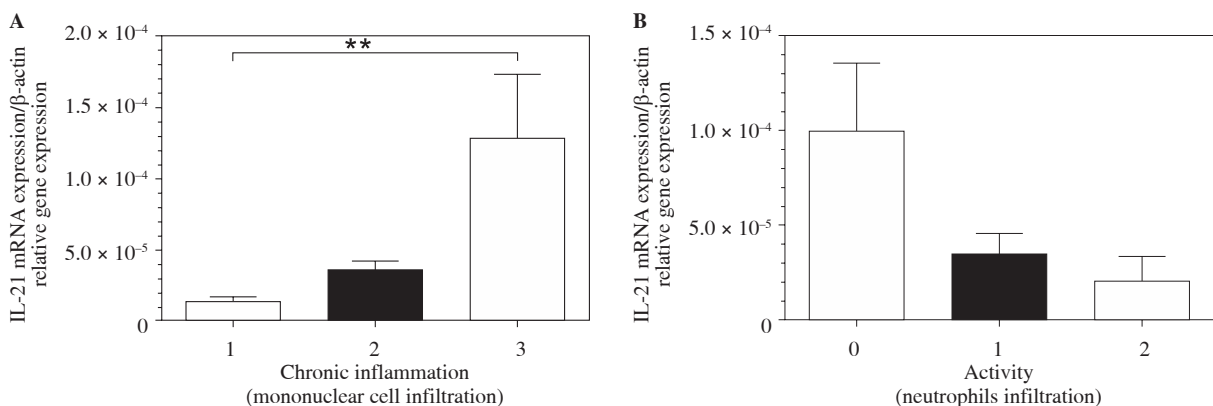
*H. pylori*-negative patients with gastritis is 0.000017, then the difference in the IL-21 mRNA expression in the *H. pylori*-positive patients compared with the *H. pylori*-negative patients with gastritis is 0.00006/0.000017, or 3.5 fold. Also, the difference in the IL-21 mRNA expression in the *H. pylori*-positive patients compared with the *H. pylori*-negative normal mucosa is 0.00006/0.0000027, or 22.22 fold, and *H. pylori*-negative patients with gastritis compared with the *H. pylori*-negative normal mucosa is 0.000017/0.0000027, or 6.3 fold.

**The effect of vacA allelic variants in *Helicobacter pylori* infected on the mucosal interleukin-21 mRNA level in gastric mucosa**

Our results showed that in *H. pylori*-infected patients, the mucosal IL-21 mRNA level was independent of the vacA status. The difference in the IL-21 mRNA expression in the vacA s1-positive *H. pylori* strains compared with the vacA s2-positive *H. pylori* strains was 0.00008/0.000041, or 1.95 fold. Mucosal IL-21 mRNA expression in *H. pylori*-infected patients with vacA m1-positive was not significantly higher than those observed in *H. pylori*-infected patients with vacA m2-positive (0.00008/0.00007, or 1.19 fold). Also, mucosal IL-21 mRNA expression in gastritis patients with vacA s1m1-positive was not significantly higher than those observed in gastritis patients with vacA s1m2-positive (0.000084/0.000089, or 0.94 fold) and vacA s2m2-positive (0.000084/0.000041, or 2.05 fold).

**The effect of cagA virulence factor in *Helicobacter pylori* infected on the mucosal interleukin-21 mRNA level in gastric mucosa**

Mucosal IL-21 mRNA level was independent of cagA virulence factor status. If the mean *H. pylori*-positive



**Fig. 2.** Relative expression levels of interleukin (IL)-21 mRNA in *Helicobacter pylori*-infected patients shown by grade of gastritis. The mucosal IL-21 mRNA level is positively correlated with the grade of chronic inflammation (A). Also, no correlation was seen between the mucosal IL-21 mRNA expression and the activity score (B). Statistical significance was evaluated using Student’s *t*-test (\*\* $p < 0.001$ ;  $p > 0.99$ )



group with *cagA* is 0.00007 and the mean *H. pylori*-positive group without *cagA* is 0.00003, then the difference in the IL-21 mRNA expression in the *cagA*-positive *H. pylori* strains compared with the *cagA*-negative *H. pylori* strains is 0.00007/0.00003, or 2.3 fold.

### Correlation between mucosal interleukin-21 mRNA levels and grade of gastritis

The degree of inflammatory activity was assessed and graded as follows: 54.1% (26) none, 39.6% (19) mild, and 6.3% (3) moderate. As far as chronic inflammation was concerned, there were 37.5% (18) mild, 33.3% (16) moderate, and 29.1% (14) severe in the studied population. Figure 2 shows the relationship between the mucosal IL-21 mRNA expression and the activity (neutrophil infiltration) and chronic inflammation (mononuclear cell infiltration) scores. There was a significant correlation between the mucosal IL-21 mRNA expression and the chronic inflammation scores. No correlation was observed between the mucosal IL-21 mRNA expression and the activity score.

### Discussion

Our findings showed that the mucosal IL-21 mRNA level increased in the *H. pylori*-infected patients compared with uninfected patients and correlated positively with the degree of chronic inflammation in patients infected with *H. pylori*. Mucosal IL-21 mRNA expression was also independent of the virulence factors of *H. pylori*. Similarly to other reports [23], we also found that the correlation of s1 and s1m2 genotypes with *cagA*-positive was statistically significant, but no correlation between m1 genotype and *cagA* was observed. We concluded that the dominant genotypes of *H. pylori* in Iranian patients with chronic gastritis might be *cagA*-positive s1/m2. Interleukin-21 is over-produced in other Th1-associated inflammatory diseases of the gastro-intestinal tract, such as *H. pylori*-related gastritis and celiac disease [24, 25]. The results of Caruso *et al.* indicate that IL-21 was over expressed in *H. pylori*-infected gastric mucosa and gastric epithelial AGS cells response to IL-21, increasing production of matrix metalloproteinase 2 (MMP-2) and MMP-9, (but not MMP-1, MMP-3, or MMP-7) which contribute to mucosal ulceration and epithelial damage [24]. High IL-21 production was also documented in autoimmune diseases such as psoriasis [26], atopic dermatitis (AD) [27], systemic lupus erythematosus (SLE) [28], graft-versus-host disease (GVHD) [29], and multiple sclerosis [30]. The data emerging from these studies show that IL-21 has broad effects on both innate and adaptive immune responses, and excessive production of IL-21 associates with destructive immune reactions in various organs. However, the basic mechanism underlying IL-21 synthesis in diseased organs remains to be clarified. The study by Sutherland *et al.* demonstrated that IL-21 levels were increased in the pancreas as NOD mice de-

veloped diabetes, and that CD4<sup>+</sup> and CD8<sup>+</sup> T-cells infiltrating the pancreas could respond to local IL-21 as they expressed IL-21R [31]. These data are consistent with recent studies from the laboratories of Leonard and Sarvetnick [32], showing that IL-21R<sup>-/-</sup> NOD mice are protected from insulinitis and type 1 diabetes. Interleukin-21 could also play a role in the pathogenesis of other immune-mediated pathologies as its levels were found to be up-regulated in synovial tissue samples of patients with rheumatoid arthritis (RA) and in labial salivary gland sections of patients with Sjögren's syndrome (SS), as well as in the plasma of patients with SS, patients with Behçet disease, and patients with Hashimoto's thyroiditis [33]. A previous published study showed that IL-21 was not able to trigger Th1 cell differentiation, but it could enhance ongoing Th1 cell responses. For example, Stregell *et al.* indicated that IL-21 enhanced interferon  $\gamma$  (IFN- $\gamma$ ) gene expression in activated human T cells and promoted the transcription of Th1-associated genes, including T-bet [34]. The same group also documented a synergistic effect of IL-21 and IL-15 or IL-21 and IL-18 on IFN- $\gamma$  mRNA synthesis and production [35], whereas Kasaian *et al.* demonstrated that IL-21 cooperates with IL-2 and IL-15 in promoting IFN- $\gamma$  synthesis [36]. Finally, a study previously showed that IL-21 enhanced T-bet and p-Stat4 expression and sustained IFN- $\gamma$  synthesis in gut mucosal lymphocytes of patients with Crohn's disease [20]. A recent study indicated an increase in Th17 in *H. pylori*-infected patients and in human gastric tumours, and further showed that *H. pylori*-primed GMF (gastric myofibroblasts/fibroblasts) promoted differentiation of TH-17. This process was dependent on IL-6, TGF- $\beta$ 1, and IL-21, which *H. pylori*-exposed and gastric tumour-derived MF (myofibroblast and fibroblast) produced at increased levels, consequently maintaining a stronger ability to induce TH-17 cells. Their findings suggest that the enhanced TH-17 promoting capacity of the GMF, derived from gastric tumours, might be among the key factors contributing to gastric tumour-promoting inflammatory milieu [37]. Another study indicated that IL-21 expression was markedly higher in human gastric cancer mucosa compared to normal [38]. Interleukin-21 can maintain chronic inflammation and/or favour tissue damage by promoting the recruitment of immune cells within the inflamed tissue [39], the expansion of autoreactive T cells [40], and the synthesis of extracellular matrix metalloproteinases [24].

---

*The authors declare no conflict of interest.*

*This study was derived from the MSc thesis of the first author and was financially supported by the research deputy of Shahrekord University of Medical Sciences. The authors are grateful to the staff of the Cellular & Molecular Research Centre, Shahrekord University of Medical Sciences, and the authorities of the endoscopy unit of Shahrekord Hajar Hospital for their valuable help.*

## References

1. Salimzadeh L, Bagheri N, Zamanzad B, et al. (2015) Frequency of virulence factors in *Helicobacter pylori*-infected patients with gastritis. *Microb Pathog* 80: 67-72.
2. Bagheri N, Azadegan-Dehkordi F, Shirzad H, et al. (2015) The biological functions of IL-17 in different clinical expressions of *Helicobacter pylori*-infection. *Microb Pathog* 81: 33-8.
3. Bagheri N, Taghikhani A, Rahimian G, et al. (2013): Association between virulence factors of *Helicobacter pylori* and gastric mucosal interleukin-18 mRNA expression in dyspeptic patients. *Microb Pathog* 65: 7-13.
4. Rahimian G, Sanei MH, Shirzad H, et al. (2014): Virulence factors of *Helicobacter pylori* vacA increase markedly gastric mucosal TGF- $\beta$ 1 mRNA expression in gastritis patients. *Microb Pathog* 67-68: 1-7.
5. Sugimoto M, Furuta T, Shirai N, et al. (2007): Different effects of polymorphisms of tumor necrosis factor- $\alpha$  and interleukin-1 beta on development of peptic ulcer and gastric cancer. *J Gastroenterol Hepatol* 22: 51-59.
6. Yuzhalin A (2011): The role of interleukin DNA polymorphisms in gastric cancer. *Hum Immunol* 72: 1128-1136.
7. Yamaoka Y, Kita M, Kodama T, et al. (1997): Induction of various cytokines and development of severe mucosal inflammation by cagA gene positive *Helicobacter pylori* strains. *Gut* 41: 442-451.
8. Bagheri N, Rahimian G, Salimzadeh L, et al. (2013): Association of the virulence factors of *Helicobacter pylori* and gastric mucosal interleukin-17/23 mRNA expression in dyspeptic patients. *EXCLI J* 12: 5-14.
9. Basso D, Plebani M (2004): *H. pylori* infection: bacterial virulence factors and cytokine gene polymorphisms as determinants of infection outcome. *Crit Rev Clin Lab Sci* 41: 313-337.
10. Brandt S, Kwok T, Hartig R, et al. (2005): NF- $\kappa$ B activation and potentiation of proinflammatory responses by the *Helicobacter pylori* CagA protein. *Proc Natl Acad Sci U S A* 102: 9300-9305.
11. Yamaoka Y, Soucek J, Odenbreit S, et al. (2002): Discrimination between cases of duodenal ulcer and gastritis on the basis of putative virulence factors of *Helicobacter pylori*. *J Clin Microbiol* 40: 2244-2246.
12. Bagheri N, Azadegan-Dehkordi F, Sanei H, et al. (2014): Associations of a TLR4 single-nucleotide polymorphism with *H. pylori* associated gastric diseases in Iranian patients. *Clin Res Hepatol Gastroenterol* 38: 366-371.
13. Atherton JC, Cao P, Peck RM Jr, et al. (1995): Mosaicism in vacuolating cytotoxin alleles of *Helicobacter pylori*. Association of specific vacA types with cytotoxin production and peptic ulceration. *J Biol Chem* 270: 17771-17777.
14. Yamaoka Y, Kodama T, Kita M, et al. (1998): Relationship of vacA genotypes of *Helicobacter pylori* to cagA status, cytotoxin production, and clinical outcome. *Helicobacter* 3: 241-253.
15. Parrish-Novak J, Dillon SR, Nelson A, et al. (2000): Interleukin 21 and its receptor are involved in NK cell expansion and regulation of lymphocyte function. *Nature* 408: 57-63.
16. Coquet JM, Kyparissoudis K, Pellicci DG, et al. (2007): IL-21 is produced by NKT cells and modulates NKT cell activation and cytokine production. *J Immunol* 178: 2827-2834.
17. Spolski R, Leonard WJ (2008): Interleukin-21: basic biology and implications for cancer and autoimmunity. *Annu Rev Immunol* 26: 57-79.
18. Zhou L, Ivanov II, Spolski R, et al. (2007): IL-6 programs T(H)-17 cell differentiation by promoting sequential engagement of the IL-21 and IL-23 pathways. *Nat Immunol* 8: 967-974.
19. Liang SC, Tan XY, Luxenberg DP, et al. (2006): Interleukin (IL)-22 and IL-17 are coexpressed by Th17 cells and cooperatively enhance expression of antimicrobial peptides. *J Exp Med* 203: 2271-2279.
20. Monteleone G, Monteleone I, Fina D, et al. (2005): Interleukin-21 enhances T-helper cell type I signaling and interferon- $\gamma$  production in Crohn's disease. *Gastroenterology* 128: 687-694.
21. Vollmer TL, Liu R, Price M, et al. (2005): Differential effects of IL-21 during initiation and progression of autoimmunity against neuroantigen. *J Immunol* 174: 2696-2701.
22. Manxhuka-Kerliu S, Telaku S, Devolli-Disha E, et al. (2009): *Helicobacter pylori* gastritis updated Sydney classification applied in our material. *Prilozi* 30: 45-60.
23. Warburton VJ, Everett S, Mapstone NP, et al. (1998): Clinical and histological associations of cagA and vacA genotypes in *Helicobacter pylori* gastritis. *J Clin Pathol* 51: 55-61.
24. Caruso R, Fina D, Peluso I, et al. (2007): IL-21 is highly produced in *Helicobacter pylori*-infected gastric mucosa and promotes gelatinases synthesis. *J Immunol* 178: 5957-5965.
25. Fina D, Sarra M, Caruso R, et al. (2008): Interleukin 21 contributes to the mucosal T helper cell type 1 response in coeliac disease. *Gut* 57: 887-892.
26. Caruso R, Botti E, Sarra M, et al. (2009): Involvement of interleukin-21 in the epidermal hyperplasia of psoriasis. *Nat Med* 15: 1013-1015.
27. Jin H, Oyoshi MK, Le Y, et al. (2009): IL-21R is essential for epicutaneous sensitization and allergic skin inflammation in humans and mice. *J Clin Invest* 119: 47-60.
28. Wong CK, Wong PT, Tam LS, et al. (2010): Elevated production of B cell chemokine CXCL13 is correlated with systemic lupus erythematosus disease activity. *J Clin Immunol* 30: 45-52.
29. Hippen KL, Bucher C, Schirm DK, et al. (2012): Blocking IL-21 signaling ameliorates xenogeneic GVHD induced by human lymphocytes. *Blood* 119: 619-628.
30. Tzartos JS, Craner MJ, Friese MA, et al. (2011): IL-21 and IL-21 receptor expression in lymphocytes and neurons in multiple sclerosis brain. *Am J Pathol* 178: 794-802.
31. Sutherland AP, Van Belle T, Wurster AL, et al. (2009): Interleukin-21 is required for the development of type 1 diabetes in NOD mice. *Diabetes* 58: 1144-1155.
32. Spolski R, Kashyap M, Robinson C, et al. (2008): IL-21 signaling is critical for the development of type I diabetes in the NOD mouse. *Proc Natl Acad Sci U S A* 105: 14028-14033.
33. Sarra M, Cupi ML, Pallone F, Monteleone G (2012): Interleukin-21 in immune and allergic diseases. *Inflamm Allergy Drug Targets* 11: 313-319.
34. Strengell M, Sareneva T, Foster D, et al. (2002): IL-21 up-regulates the expression of genes associated with innate immunity and Th1 response. *J Immunol* 169: 3600-3605.
35. Strengell M, Matikainen S, Siren J, et al. (2003): IL-21 in synergy with IL-15 or IL-18 enhances IFN- $\gamma$  production in human NK and T cells. *J Immunol* 170: 5464-5469.
36. Kasaian MT, Whitters MJ, Carter LL, et al. (2002): IL-21 limits NK cell responses and promotes antigen-specific T cell activation: a mediator of the transition from innate to adaptive immunity. *Immunity* 16: 559-569.
37. Pinchuk IV, Morris KT, Nofchissey RA, et al. (2013): Stromal cells induce Th17 during *Helicobacter pylori* infection and in the gastric tumor microenvironment. *PLoS One* 8: e53798.

38. Iida T, Iwahashi M, Katsuda M, et al. (2011): Tumor-infiltrating CD4+ Th17 cells produce IL-17 in tumor microenvironment and promote tumor progression in human gastric cancer. *Oncol Rep* 25: 1271-1277.
39. Caruso R, Fina D, Peluso I, et al. (2007): A functional role for interleukin-21 in promoting the synthesis of the T-cell chemoattractant, MIP-3alpha, by gut epithelial cells. *Gastroenterology* 132: 166-175.
40. King C, Ilic A, Koelsch K, Sarvetnick N. (2004): Homeostatic expansion of T cells during immune insufficiency generates autoimmunity. *Cell* 117: 265-277.