



Journal of Global Pharma Technology

Available Online at www.jgpt.co.in

REVIEW PAPER

Use of Iranian Medicinal Plants Effective on Male Fertility Indices

Majid Shirani¹, Saeid Heidari-Soureshjani², Mahnaz Yavangi^{3*}

¹Cellular and Molecular Research Center, Shahrekord University of Medical Sciences, Shahrekord, Iran.

²Medical Plants Research Center, Shahrekord University of Medical Sciences, Shahrekord, Iran.

³Department of Gynecology, Endometrium and Endometriosis Research Center, Hamadan University of Medical Sciences, Hamadan, Iran.

*Corresponding Author: Mahnaz Yavangi

Abstract: This systematic review article was conducted to report medicinal plants that are effective on male fertility indices. To conduct this review, search terms fertility or pregnancy combined with medicinal plants, herb, and phyto were used to retrieve relevant publications indexed in ISI, PubMed, and Iranian scientific databases. Escanbil, *Fumaria parviflora* L., *Apium graveolens* L., *Achillea millefolium* inflorescence, *Urtica dioica* L., *Withania somnifera* L., *Phaleria macrocarpa*, *Satureja khuzestanica*, *Achillea millefolium*, *Malva sylvestris*, *Humulus lupulus* L., *Allium cepa*, *Petroselinum crispum*, *Carthamus tinctorius*, and *Zingiber officinale* were reported to be medicinal plants with fertility indices-promoting properties. Medicinal plants that are used in Iran affect hormonal compounds and other effective compounds on hormonal system as well as the proliferation and viability of sex cells due to their antioxidant properties, and therefore enhance fertility in human and animals.

Keywords: Physiotherapy, Infertility, Pregnancy, Iran.

Introduction

As one of the most common healthcare problems around the world, infertility imposes stupendous costs on families and challenges economic stability of healthcare system (1, 2). In Iran, infertility is considered one of the most important healthcare issues that affects the quality of life among affected people adversely and leads to psychosocial problems (3, 4). Although the reasons for infertility may be various and undetermined, oligozoospermia and asthenozoospermia are considered the most important causes of male infertility (5).

Treatment of infertility is one of the important challenges in the lives of infertile couples and the strategies of treating infertility should take into account certain issues such as age, side effects, treatment efficiency, and treatment costs in the patients (6), because treatments for infertility may be unsuccessful for several and therefore the patients' reasons. problems are likely to be doubled (7). © 2009-2016, JGPT. All Rights Reserved.

Therefore, efforts are being made to promote and find alternative treatments for infertility. Because of being economical and causing fewer side effects compared to synthetic drugs, medicinal plants can be potential suitable alternative therapies for several diseases (8-19). In this regard, different experimental and clinical studies have demonstrated the effects of these plants in addition to confirming their use in traditional medicine.

Studies indicate that in Iran, medicinal plants are much commonly used to treat different diseases (20-27) and also to the infertility (28). This systematic review was conducted to explain the status of Iran medicinal plants in preventing and treating male infertility via assessing publications on the effects of medicinal plants to enhance fertility indices (help treat infertility).

To conduct this review, search terms *fertility* or *pregnancy* combined with *medicinal plants*, *herb*, and *phyto* were used to retrieve

relevant publications indexed in ISI and PubMed. Then, the abstracts and data drawn from other resources were examined and only those studies that were conducted mainly to investigate the effects of the plants and their derivatives on male infertility in Iran and on medicinal plants native to Iran were included in analysis.

Results

There are several plants that have positive effects on fertility indices in laboratory animals and human, and there are different mechanisms for exertion of such properties.

Escanbil (Calligonum)

 \mathbf{is} plant Calligonum а from family Polygonaceae with antioxidant property due to the presence of quercetin and catechin. Therefore, this plant can play a role in sperm motility and viability. Administration with calligonum caused increase in the expression of the gene Catsper and subsequently improvement of sperm parameters in male mice (29).

Tribulus Terrestris (Bindii)

This plant is annual and occurs in different regions of the world. The effects of this plant were investigated on sex hormones and gonadotropins in laboratory mice with induced addiction to morphine. Administration with T. terrestris, as an antagonist, serves to reduce sex hormones except for follicle-stimulating hormone (FSH) in laboratory mice (30).

Fumaria parviflora L.

This plant has a special status in Iranian traditional medicine and contains antioxidant compounds that prevent the activity of oxidative substances, such that the ethanolic extract of this plant prevented reduction testicular weight. in serum level. diameter testosterone the of tubules. seminiferous the number of epididymal sperm, testicular content, dismutase, and glutathione superoxide peroxidase and therefore increased fertility in male mice with lead-induced testicular toxicity (31). F. parviflora leaf extract could increase fertility in male mice via enhancing androgenic activity and proliferation of sex cells as well as alkaloid compounds (with antioxidant property) (32).

Apium graveolens L. (Celery)

A. graveolens is from family Apiaceae. Administration with A. graveolens leaf extract after 30 days caused increase in testicular volume and seminiferous tubules diameter in mice. Besides that, the number spermatocytes, of spermatogonia, and spermatozoa increased, and therefore the process of spermatogenesis improved (33). In addition, oral administration of laboratory mice with A. graveolens leaf extract caused increase in the number of spermatocytes, Sertoli cells, and primary spermatocytes as well as the improvement of spermatogenesis (34). However, Modaresi et al. reported that doses of hydroalcoholic different Α. graveolens leaf extract displayed different activities such that 100 and 150 mg/kg doses led to decrease in FSH level and dosedependent adverse effects on pituitary in mice (35).

Achillea millefolium

A. millefolium is a flowering plant whose cyclophosplamide-induced effect in contraceptive side effects has been studied. Jalali et al. study showed that aqueous A. millefolium extract prevented decrease in testicular and epididymal weight in mice under treatment with cyclophosplamide and increased serum testosterone level, generally improving the process of spermatogenesis and fertility in Wistar rats (36). A study showed that degradation of sex cells and decrease in gonadotropins induced bv cyclophosplamide were neutralized by administration with A. millefolium extract in laboratory mice (37).

Urtica dioica L.

The effect of this plant, commonly known as common nettle, was investigated on laboratory male mice with induced addiction to nicotine by Jalili et al. Intraperitoneal administration of the mice with hydroalcoholic U. dioica extract caused increase in testicular weight, the number and motility of the sperm, normal sperm morphology, serum testosterone level, and the diameter of seminiferous tubules after 28 days. Therefore, administration with this plant plays a role in increasing the quality of spermatozoa and sperm parameters (38).

Ginseng

Korean red ginseng has antioxidant and antiapoptotic compounds and can decrease epididymo-orchitis in laboratory male mice such that it causes increase in sperm concentration and motility and the number of normal sperms and therefore improves fertility indices (39). Gavage administration of laboratory mice with ginseng and vitamin E caused decrease in the number of sperm including viable sperm as well as malformed and abnormal sperm. In addition, the rate of fertility increased in male mice administered with ginseng and vitamin E via inducing antioxidant property in sex cells (40).

Withania somnifera L.

W. somnifera is from family Solanaceae. A study demonstrated that oral administration with this plant caused increase in gonadotropin hormones in mice with already reduced fertility indices due to addiction to morphine, such that FSH and luteinizing hormone (LH) levels were modulated and estrogen and testosterone levels increased (41).

Phaleria macrocarpa

The effect of pulverized *P. macrocarpa* was investigated on laboratory mice fertility. This plant was reported to be one of the effective plants in treating infertility such that a study reported that the number of spermatogonial cells and the thickness of seminiferous tubules increased after the treatment and therefore it could be used as a complementary therapy (42).

Date palm pollen (*Phoenix dactylifera* L.)

P. dactylifera pollen is used to stimulate the libido and treat infertility. Treatment of infertile men with P. dactylifera pollen, as oral capsule, for two months was investigated by measuring sperm parameters. It was found that *P. dactylifera* pollen caused increase in the number and motility of sperm and improvement of sperm morphology. Therefore, P. dactylifera pollen can be used in the treatment of male infertility as a plant-based therapy (43).

Satureja khuzestanica

S. khuzestanica is from family Lamiaceae whose protective effect was investigated in preventing infertility in mice under treatment with cyclophosplamide. The essential oils of S. khuzestanica caused © 2009-2016, JGPT. All Rights Reserved. increase in plasma testosterone and sperm quality as well as decrease in DNA damage and oxidative stress (44).

Achillea millefolium

Akbarizadeh et al. investigated the protective effect of aqueous *A. millefolium* extract against cyclosporin on fertility indices in mice. *A. millefolium* neutralized the adverse effects of cyclosporin in reducing the number of sperm, sperm viability, laboratory fertility and led to significant increase in these parameters (45).

Malva sylvestris

M. sylvestris is a medicinal plant that has been used in Iranian traditional medicine for thousands of years and is from family Malvaceae. A study investigated the effect of this plant on sperm and soetmatogenesis phases in mice. According to the findings of of this study, the number primary spermatocytes and spermis increased significantly after treatment with hydroalcoholic *M. sylvestris* leaf extract (46).

Humulus lupulus L.

H. lupulus flower extract causes increase in estrogen and testosterone as well as the number of spermatogonial cells, spermatocytes, and spermatids by phytoestrogenic compounds and stimulating LH secretion (47).

Allium cepa

The effect of oral administration with A. cepa extract was investigated on testicular tissue in mice. The results demonstrated that oral administration with raw A. cepa extract affected cell proliferation in mice testicular tubules and accelerated the trend of spermatogenesis in addition to influencing the structure of seminiferous tract (48).

Petroselinum crispum

Bastampour et al. found that the studied concentrations of P. crispum leaf extract caused modulation of LH and FSH, while the minimum and moderate concentrations of this extract caused serum levels of testosterone to increase and maximum concentration of this extract caused significant decrease in the level of this hormone (49).

Carthamus tinctorius

In Iran, C. tinctorius flower has pharmaceutical uses. Administration with C.tinctorius extract caused increase in serum testosterone concentration and negative physical self-control of testosterone secretion in mice (50).

Zingiber officinale

Pulverized Z. officinale rhizome caused increase in the rate and percentage of viability and motility of sperm and therefore infertility in laboratory mice (51). Plant-based compounds

Rosmarinic Acid

Rosmarinic acid is a plant-based polyphenol whose effect was investigated on male mice exposed to electromagnetic field. The results demonstrated that serum testosterone level in mice under treatment with rosmarinic acid was higher than other groups, and this extract could be used to increase fertility with few side effects (52).

Traditional Prescriptions and Pharmaceutical Formulations

The effect of the combination of seven plants Allium cepa, Zingiber officinale, Ocimum basilicum, Cinnamomum verum, Citrus sinensis' peel, Citrullus lanatus' seeds and Daucus carota' seeds in a plant capsule was infertile investigated on men. After treatment with this capsule, the number and motility of the sperm as well as the number of normal sperm increased. Therefore, these plants caused improvement of sperm parameters through inducing antioxidant effect (53). A study investigated the effect of TOPALAF consisting of T. Terrestris fruit, almond kernel, watercress seeds, vegetable seeds, roots orchids, palm pollen, and fruit figs to enhance fertility in men. Treatment with this prescription caused improvement of sperm number and motility (54).

This review was conducted to report the plants and plant-based compounds that are

effective on male infertility. Most research conducted on this subject has been animal studies. In most studies, both in vivo and in vitro, oxidative stress has been considered to be a factor for declined fertility and the studied plants cause increase in fertility due antioxidant properties. Plant-based to antioxidant compounds have been investigated for neutralization of reactive oxygen species (ROS) and reduction in adverse effects of oxidative stress via increasing peroxidases concentrations in different cells of the body and their effects have been reported (55-60).

The effects of these antioxidant compounds have been also investigated on sex cells. Indeed, the final product of the process causes increase in the number of healthy sperm that has significant effects on fertility (61). Moreover, the presence of plant-based antioxidants causes certain changes in gene expression and therefore improves fertility indices (29). However, it should be taken into account that small amounts of ROS are needed for natural sperm function for male fertility, but excessively higher amounts of threshold amount of ROS and associated metabolites can attack DNA, lipids, and proteins and disturb enzymatic system. This process leads ultimately to irreversible changes and cell death that cause decline in sperm parameters and therefore fertility (62). Therefore, determining the threshold level of ROS or their metabolites and the administered doses of antioxidants are important issues (63-67).

According to reviewed studies in male and laboratory male animals, some plants exert their effects due in different mechanisms (Figure 1).



Figure 1: Several mechanisms herbal medicine to increase fertility

It should be taken into account that stress and social pressure in infertility can lead to excessive consumption of medicinal plants; therefore, necessary measures should be taken to prevent adverse side effects of using medicinal plants that are effective on infertility and associated toxicities (68).

Conclusion

Medicinal plants that are used in Iran can increase fertility due to antioxidant compounds, hormonal compounds, and other effective compounds on hormonal system, and improvement of sex cells proliferation

References

- 1. Dyer SJ, Patel M (2012) The economic impact of infertility on women in developing countries: a systematic review. Facts Views Vis Obgyn. 4(2):102-9.
- 2. Mascarenhas MN, Flaxman SR, Boerma T, Vanderpoel S, Stevens GA (2012) National, regional, and global trends in infertility prevalence since 1990: a systematic analysis of 277 health surveys. PLoS Med. 9(12):1001:356.
- Hasanpoor-Azghdy SB, Simbar M, Vedadhir A (2015) The social consequences of infertility among iranian women: a qualitative study. Int J Fertil Steril. 8(4):409-20.
- Hasanpoor-Azghdy SB, Simbar M, Vedadhir A(2014) The emotional-psychological consequences of infertility among infertile women seeking treatment: Results of a qualitative study. Iran J Reprod Med.12(2):131-8.
- Elussein EA, Magid YM, Omer MM, Adam I (2008) Clinical patterns and major causes of infertility among Sudanese couples. Trop Doct. 38(4):243-4.
- Quaas A, Dokras A (2008) Diagnosis and treatment of unexplained infertility. Rev Obstet Gynecol. 1(2):69-76.
- Simon A, Laufer N (2012) Assessment and treatment of repeated implantation failure (RIF). J Assist Reprod Genet. 29(11):1227-39.
- Kooti W, Hasanzadeh-Noohi Z, Sharafi-Ahvazi N, Asadi-Samani M, Ashtary-Larky D (2016) Phytochemistry, pharmacology, and therapeutic uses of black seed (Nigella sativa). Chin J Nat Med.14(10):732-45.
- Mahmoudian-Sani M, Luther T, Asadi-Samani M, Saeedi-Boroujeni A, Gholamian NA (2017) new approach for treatment of type 1 diabetes: Phytotherapy and phytopharmacology of regulatory T cells. J Ren Inj Prev.6(3):158-63.

and viability and are therefore used as complementary therapies to modern treatments in fertility centers. It should be also taken into consideration that one plant may increase fertility in one gender but exert opposite effects on another gender.

- 10. Parsaei P, Bahmani M, Naghdi N, Asadi-Samani M, Rafieian-Kopaei M (2016) The most important medicinal plants effective on constipation by the ethnobotanical documents in Iran: A review. Der Pharm Lett.8(2):188-94.
- 11. Sani MRM, Asadi-Samani M, Saeedi-Boroujeni A, Banitalebi-Dehkordi M, Bahmani M (2016) Suppressive effects of medicinal plants and their derivatives on inflammasome complex: A systematic review. Int J PharmTech Res.9(6):325-35.
- Parsaei P, Bahmani M, Karimi M, Naghdi N, Asadi-Samani M, Rafieian-Kopaei M. (2016) A review of analgesic medicinal plants in Iran. Der Pharm Lett. 8(2):43-51.
- 13. Nasri H (2016) Impact of garlic extract on platelet function and structure. Ann Res Antioxid.1(1):e01.
- 14. Nasri H, Abedi-Gheshlaghi Z, Rafieian-Kopaei M (2016) Curcumin and kidney protection; current findings and new concepts. Acta Persica Pathophysiol.1(1):e01.
- 15. Dehghan Shahreza F (2016) Hibiscus esculentus and diabetes mellitus. J Nephropharmacol. 5(2):104-5.
- 16. Kafeshani M (2015) Ginger, microinflammation and kidney disease. J Ren Endocrinol.1(1):e04.
- 17. Rafieian-Kopaei M (2013) Medicinal plants for renal injury prevention. J Ren Inj Prev.2(2):63-5.
- 18. Rafieian-Kopaei M (2015) Natural sources of vitamin D. J Parathyr Dis. 3(1):10-1.
- Raeisi E, Shahbazi-Gahrouei D, Heidarian E (2016) Pineapple extract as an efficient anticancer agent in treating human cancer cells. Front Cancers.1(1):e03.

- 20. Asadi-Samani M, Kooti W, Aslani E, Shirzad HA (2016) systematic review of Iran's medicinal plants with anticancer effects. J Evid Based Complementary Altern Med.21(2):143-53.
- Baharvand-Ahmadi B, Asadi-Samani M (2017) Medicinal plants and treatment of hypertension; evidence from Iran. J Nephropharmacol.6(1):3-8.
- 22. Bahmani M, Asadi-Samani M (2016) Native medicinal plants of Iran effective on peptic ulcer. J Inj Inflam.1(1):e05.
- 23. Bahmani M, Tajeddini P, Ezatpour B, Rafieian-Kopaei M, Naghdi N, Asadi-Samani M (2016) Ethenobothanical study of medicinal plants against parasites detected in Shiraz, southern part of Iran. Der Pharm Lett.8(1):153-60.
- 24. Jivad N, Asadi-Samani M, Moradi MT (2016) The most important medicinal plants effective on migraine: A review of ethnobotanical studies in Iran. Der Pharm Chem.8(2):462-6.
- 25. Jivad N, Bahmani M, Asadi-Samani M (2016) A review of the most important medicinal plants effective on wound healing on ethnobotany evidence of Iran. Der Pharm Lett.8(2):353-7.
- 26. Parsaei P, Bahmani M, Naghdi N, Asadi-Samani M, Rafieian-Kopaei M, Boroujeni S (2016) Shigellosis phytotherapy: A review of the most important native medicinal plants in Iran effective on Shigella. Der Pharm Lett.8(2):249-55.
- 27. Shabanian S, Bahmani M, Asadi-Samani M (2016) The medicinal plants effective on female hormones: A review of the native medicinal plants of Iran effective on estrogen, progesterone, and prolactin. J Chem Pharm Sci.9(3):1270-6.
- 28. Khadivzadeh T, Ghabel M (2012) Complementary and alternative medicine use in pregnancy in Mashhad, Iran, 2007-8. Iran J Nurs Midwifery Res.17(4):263-9.
- 29. Askari Jahromi M, Movahedin M, Mazaheri Z, Amanlu M, Mowla SJ, Batooli H(2014) Evaluating the effects of Escanbil (Calligonum) extract on the expression level of Catsper gene variants and sperm motility in aging male mice. Iran J Reprod Med.12(7):459-66.

- 30. Ghosian Moghaddam MH, Khalili M, Maleki M, Ahmad Abadi ME (2013)The Effect of Oral Feeding of Tribulus terrestris L. on Sex Hormone and Gonadotropin Levels in Addicted Male Rats. Int J Fertil Steril. 7(1):57-62.
- 31. Dorostghoal M, Seyyednejad SM, Jabari A (2014) Protective effects of Fumaria parviflora L. on lead-induced testicular toxicity in male rats. Andrologia.46(4):437-46.
- 32. Dorostghoal M, Seyyednejad SM, Khajehpour L, Jabari A (2013) Effects of Fumaria parviflora leaves extract on reproductive parameters in adult male rats. Iran J Reprod Med.11(11):891-8.
- 33. Hardani A, Afzalzadeh MR, Amirzargar A, Mansouri E, Meamar Z (2015) Effects of aqueous extract of celery (Apium graveolens L.) leaves on spermatogenesis in healthy male rats. Avicenna J Phytomed.5(2):113-9.
- 34. Kooti W, Mansouri E, Ghasemiboroon M, Harizi M, Ashtary-Larky D, Afrisham R (2014) The effects of hydroalcoholic extract of Apium graveolens leaf on the number of sexual cells and testicular structure in rat. Jundishapur J Nat Pharm Prod.9(4):e17532.
- 35. Modaresi M, Ghalamkari G, Jalalizand A (2012) The Effect of Celery (Apium graveolens) Extract on the Reproductive Hormones in Male Mice. Apcbee Proc.4:99-104.
- 36. Jalali AS, Hasanzadeh S, Malekinejad H (2012) Achillea millefolium inflorescence aqueous extract ameliorates cyclophosphamide-induced toxicity in rat testis: stereological evidences. Chin J Nat Med.10(4):247-54.
- 37. Shalizar-Jalali A, Hasanzadeh S, Malekinejad H (2013) Beneficial effects of Achillea millefolium aqueos extract against cyclophosphamide-induced reproductive toxicity. J Exp Integr Med.3(2): 113-9.
- 38. Jalili C, Salahshoor MR, Naseri A (2014) Protective effect of Urtica dioica L against nicotine-induced damage on sperm parameters, testosterone and testis tissue in mice. Iran J Reprod Med.12(6):401-8.

- 39. Eskandari M, Jani S, Kazemi M, Zeighami H, Yazdinezhad A, Mazloomi S, et al. (2016) Ameliorating effect of ginseng on epididymo-orchitis inducing alterations in sperm quality and spermatogenic cells apoptosis following infection by uropathogenic Escherichia coli in Rats. Cell J.18(3):446-57.
- 40. Hosseini A, Zare S, Ghaderipakdel F, Ahmadi A (2010) Evaluate the antioxidant effect of ginseng extract and vitamin E on fertility Laboratory male rats after long-term treatment with cyclophosphamide. Q Reprod Infertil.11(4):227-37.
- 41. Rahmati B, Moghaddam MHG, Khalili M, Enayati E, Maleki M, Rezaeei S (2016) Effect of Withania somnifera (L.) dunal on sex hormone and gonadotropin levels in addicted male rats. Int J Fertil Steril.10(2):239-44.
- 42. Parhizkar S, Zulkifli SB, Dollah MA (2014) Testicular morphology of male rats exposed to Phaleria macrocarpa (Mahkota dewa) aqueous extract. Iran J Basic Med Sci.17(5):384-90.
- 43. Rasekh A, Jashni HK, Rahmanian K, Jahromi AS (2015) Effect of palm pollen on sperm parameters of infertile man. Pak J Biol Sci.18(4):196-9.
- 44. Rezvanfar MA, Sadrkhanlou RA, Ahmadi A, Shojaei-Sadee H, Rezvanfar MA, Mohammadirad A, et al. (2008) Protection of cyclophosphamide-induced toxicity in reproductive tract histology, sperm characteristics, and DNA damage by an herbal source; evidence for role of freeradical Hum toxic stress. Exp Toxicol.27(12):901-10.
- 45. Akbarizadeh Z, Najafi G, Farokhi F (2012) Effect of aquatic extract of Achillea millefolium on sperm and in vitro fertilization in adult rats treated with Cyclosporine A. Rostamineh.4(4):9-18.
- 46. Nouhizadeh Z, Parivar K, Hayati-Roudbari N (2014) The effect sylvestris Malva leaves on the sperm and spermatogenesis in mice. J Anim Biol.7(2):81-8.
- 47. Tavakkoli Kazeroni H, Hosseini SE, Shariati M (2014) The effect of hops (Humulus lupulus L.) ethanol extracts on the sexual hormones levels and sexual

dynastic cells of Syrian adult male mice. J Sabzevar Univ Med Sci. 21(3):514-21.

- 48. Nikroush M, Jalali M, Mohammadi S (2009) Effect of crude extract of the onion on rat testis tissue. Reprod Infertil Q. 10(4):239-44.
- 49. Bastampoor F, Sadeghi H, Hosseini S (2014) The Petroselinum crispum L. hydroalcoholic extract effects on pituitary- gonad axis in adult Rats. Armaghane danesh.19(4):305-13.
- 50. Modaresi M (2005) The effect of Carthamus tinctorius plant extracts on the pituitary-gonad axis and testicular histology in mice. ZUMS J.13(53):1-7.
- 51. Khaki A, Noori M, Fathi Azad F, Khaki A (2008) Onion and ginger effects on spermatogenesis in rats. Med J Tabriz Univ Med Sci.30(2):53-8.
- 52. Khaki A, Imani SAM, Golzar FS (2012) Effects of rosmarinic acid on male sex hormones (testosterone-FSH-LH) and testis tissue apoptosis after exposure to electromagnetic field (EMF) in rats. Afr J Pharm Pharmacol.6(4):248-52.
- 53. Ouladsahebmadarek E, Giasi GS, Khaki A, Ahmadi Y, Farzadi L, Ghasemzadeh A,(2016) et al. The effect of compound herbal remedy used in male infertility on spermatogenesis and pregnancy rate. Int J Womens Health.4(4):185-8.
- 54. Khoradmehr A, Khalili MA, Ramezani M, Vahidi S, Moein MR (2014) Improvement of sperm physiological parameters in patients with fertility problems after taking the herbal medicine "TOPALAF". Iran J Med Arom Plant.30(2):275-82.
- 55. Baradaran A (2017) Herbal antioxidant to ameliorate vascular biology. Angiol Persica Acta. 2(1):e01.
- 56. Hajian S (2015) Positive effect of antioxidants on immune system. Immunopathol Persa.1(1):e02.
- 57. Mohammadparast V (2016) Antioxidant efficacy of Hibiscus esculentus. Front Biomed.1(1):e04.
- 58. Nasri P (2017) Cancers and herbal antioxidants. Fron Biomark. 2(1):e01.
- 59. Rafieian-Kopaei M, Baradaran A, Rafieian M (2013) Plants antioxidants: From laboratory to clinic. J Nephropathol.2(2):152-3.

- 60. Nasri H (2016) Improving the nephrotoxicity of cyclosporine; the role of herbal drugs. Toxicol Persa. 1(1):e05.
- 61. Fallahi S, Rajaei M, Malekzadeh K, Kalantar SM (2015) Would Phoenix Dactyflera Pollen (palm seed) be considered as a treatment agent against Males' infertility? A systematic review. Electron Physician. 7(8):1590-6.
- 62. Agarwal A, Virk G, Ong C, du Plessis SS (2014) Effect of oxidative stress on male reproduction. World J Men's Health.32(1):1-17.
- 63. Khodadadi S, Rafieian-Kopaei M (2016) Herbs, health and hazards; a nephrology viewpoint on current concepts and new trends. Ann Res Antioxid.1(1):e05.
- 64. Nasri H (2016) Herbal drugs and new concepts on its use. J Prev Epidemiol.1(1):e01.

- 65. Nasri H (2017) Herbs and hazards; administration of herbal drugs in maintenance hemodialysis patients. Ann Res Dial. 2(1):e01.
- 66. Nasri H (2017) Herbal drugs; from molecular studies to bedside investigations. Aria J Front Biochem. 2(1):e01.
- 67. Baradaran A (2017) Administration of herbal drugs in geriatric individuals; trends on its helps and hazards. Geriatr Persia.1(1):e01.
- 68. Kaadaaga HF, Ajeani J, Ononge S, Alele PE, Nakasujja N, Manabe YC, et al (2014) Prevalence and factors associated with use of herbal medicine among women attending an infertility clinic in Uganda. BMC Complement Altern Med.14:27.