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# MicroRNAs–Based Imaging Techniques in Cancer Diagnosis and Therapy

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## ABSTRACT

Cancer is one of the most serious global health concerns in different populations. Several studies indicated that there are many potentially promising cellular and molecular targets for cancer therapy within cancer cells and their microenvironment. Among different cellular and molecular targets involved in cancer pathogenesis, microRNAs (miRNAs) are well known as key targets for cancer therapy. miRNAs are one of main classes of non-coding RNAs. These molecules play important roles in different critical processes of cancer pathogenesis. Hence, this makes miRNAs as a suitable tool for cancer diagnosis and therapy. There are different approaches for monitoring miRNAs in cancer patients. Some conventional approaches including next-generation sequencing, real-time polymerase chain reaction (PCR), northern blotting, and microarrays could be used for assessment of miRNAs expression. Some studies revealed that the utilization of these approaches associated with various limitations. Recently, it has been revealed that molecular imaging techniques are powerful tools for monitoring of different cellular and molecular targets involved in various diseases such as cancer. These techniques help investigators to investigate and monitor miRNAs functions through assessing different targets by fluorescent proteins, bioluminescent enzymes, molecular beacons, as well as various nanoparticles. Therefore, utilization of molecular imaging techniques could assist investigators to better monitor and more effectively treat patients during different phases of malignancy. Here, we give a review on the current state of miRNAs-based imaging techniques in cancer diagnosis and therapy. *J. Cell. Biochem.* 9999: 1–8, 2017. © 2017 Wiley Periodicals, Inc.

**KEY WORDS:** MOLECULAR IMAGING; MicroRNA; CANCER; DIAGNOSIS; THERAPY

To date, cancer has been emerged as one of main health problems in worldwide [Faghihloo et al., 2016; Mirzaei et al., 2016a–c]. Despite of recent advances in cancer therapy (such as gene therapy, cell therapy, and molecularly-targeted therapies), this disease has remained as one of the major public health issues worldwide [Mirzaei et al., 2016d–f, 2017a; Mohammadi et al., 2016a]. Hence, the identification of new targets and molecules involved in cancer, from initiation to

progression and treatment, could contribute in understanding of the pathways involved in cancer pathogenesis [Arabpour et al., 2016; Mirzaei et al., 2016; Simonian et al., 2016]. These finding could lead to development of new and effective treatments in this field. One of the most important mediators in cancer pathogenesis is microRNAs (miRNAs) [Fathollahzadeh et al., 2016; Mohammadi et al., 2016b; Gholamin et al., 2017; Mirzaei et al., 2017b; Moridikia et al., 2017].

Conflicts of interest: None.

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MiRNAs are well known as a class of non-coding RNAs which play key roles in a wide range of biological processes including development, cellular differentiation, proliferation, and apoptosis in living organisms [Ryoo et al., 2013; Gholamin et al., 2016; Rashidi et al., 2016]. The lengths of these molecules are almost ~22 nucleotides [Mirzaei et al., 2016b]. Several studies indicated that miRNAs have a central role in pathogenesis of various diseases such as cancer [Gholamin et al., 2017; Mirzaei et al., 2017a; Moridikia et al., 2017]. Hence, detection and profiling of these molecules could contribute to the better understanding of cancer pathogenesis and, thereby, developing more precise strategies for early detection and/or treatment of cancer [Zubakov et al., 2010; Salarini et al., 2015; Gholamin et al., 2016; Mirzaei et al., 2016c]. So far, various conventional approaches such as microarray and RT-PCR have been used for miRNAs detection in different living systems [Li and Ruan, 2009; Momin et al., 2009; Hernandez et al., 2013]. These approaches are associated with some limitations. For example, these approaches are time-consuming and laborious and required to fix or lyse the cells and thus cannot monitor dynamic functions of miRNAs in living cells and organisms (In vivo) [Fang et al., 2006; Lu and Tsourkas, 2009; Ryoo et al., 2013]. To overcome the limitations, developing new and more efficient, particularly noninvasive repeated quantitative, methods are the most highly demanded. Recently, some imaging techniques have been emerged as effective tools for monitoring of miRNAs [Wang et al., 2009; Hernandez et al., 2013]. Imaging techniques are well known as powerful tools in monitoring of various targets and genes in many diseases such as cancer [Wang et al., 2009; Hernandez et al., 2013]. These techniques could provide new avenue in diagnosis and treatment of cancer. Various imaging techniques such as magnetic resonance imaging (MRI), florescence imaging, and bioluminescence imaging (BLI) could be used for monitoring and detection of miRNAs in patients with cancer [Wang et al., 2009; Hernandez et al., 2013]. In the present review, we will not only focus on a variety of miRNAs involved in cancer pathogenesis but also highlight some recently developed

imaging methods which assist investigators to detect miRNAs in a high throughput profiling and noninvasive repeated quantitative manner in preclinical and clinical cancer studies.

## MicroRNA AND CANCER

MiRNAs are one of main class of non-coding short RNAs that are highly conserved [Mirzaei et al., 2016e,b; Mohammadi et al., 2016b]. These RNAs regulate gene expression at different levels (protein and RNA) [Rashidi et al., 2017; Salarini et al., 2015]. MiRNAs have multiple biological roles in different processes within living organisms [Mirzaei et al., 2017a; Rashidi et al., 2017]. Figure 1 illustrate a scheme of miRNA biogenesis. Multiple lines of evidence indicated that miRNAs play important roles in initiation and progression of various cancers [Mirzaei et al., 2016f,g; Mohammadi et al., 2016b]. These molecules regulate different cellular and molecular pathways including Wnt, Notch, TGF- $\beta$  and play putative roles in epithelial-mesenchymal transition (EMT). It is plausible that some expression aberrations of miRNAs might lead to initiation and/or development of a variety of cancers [Mirzaei et al., 2016g,h; Mohammadi et al., 2016b] (Table I). Various studies have been revealed that miRNAs could be used as diagnostic, prognostic and therapeutic biomarkers for various cancers. Therefore, monitoring of miRNA expression patterns can assist the elucidation of the biogenesis and biological function of miRNAs in different phases of cancers.

## MicroRNA AND MOLECULAR IMAGING IN CANCER

The miRNAs are known to regulate the expression of genes involved in many cellular/and molecular pathways. The aberration of these

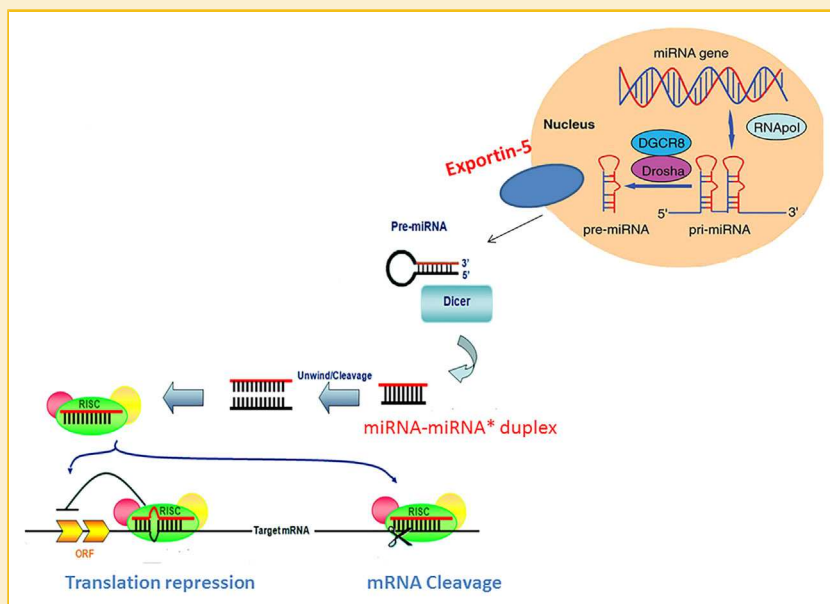


Fig. 1. A scheme of miRNA biogenesis.

TABLE I. Various MiRNAs Involved in Cancer Pathogenesis

| Type of cancer           | miRNA             | Expression in cancer | Target gene                   | Stage/patient sample                                  | Citation   |
|--------------------------|-------------------|----------------------|-------------------------------|---|--|
| Melanoma                 | miR-200c          | Down regulation      | ZEB1, DEF1, Nil-2-A           | I, II, III, IV/41                                     | Xu et al. [2012]   |
|                          | miR-211           | Down regulation      | MITF, AP1S2, SOX11, IGFBP5    | I, II, III, IV/41                                     | Xu et al. [2012]   |
|                          | miR-205           | Down regulation      | E2F1, E2F5                    | I, II, III, IV/41                                     | Xu et al. [2012]   |
|                          | miR-203           | Down regulation      | E2F3                          | I, II, III, IV/41                                     | Xu et al. [2012]   |
|                          | miR-33a           | Up regulation        | Pim-1, CDK6, cyclin D1        | I-III/140   | Friedman et al. [2012]   |
| Lung                     | miR-33a           | Up regulation        | p53, c-Myc                    | IIIB, IIIC, IV/59                                     | Segura et al. [2010]   |
|                          | miR-193b          | Up regulation        | MAPK, PI3K-AKT, p53, ErbB     | I, II/107   | Nadal et al. [2015]  |
|                          | miR-301           | Up regulation        | MAPK, PI3K-AKT, p53, ErbB     | I, II/107   | Nadal et al. [2015]  |
|                          | miR-141           | Up regulation        | MAPK, PI3K-AKT, p53, ErbB     | I, II/107   | Nadal et al. [2015]  |
|                          | miR-200b          | Up regulation        | MAPK, PI3K-AKT, p53, ErbB     | I, II/107   | Nadal et al. [2015]  |
| Breast                   | miR-21            | Up regulation        | PDCD4, HIF1A                  | 17  | Qi et al. [2009]   |
|                          | miR-10b           | Up regulation        | HOXD10                        | 23  | Ma et al. [2007]   |
|                          | miR-155           | Up regulation        | SOCS1, FOXO3                  | 15  | Jiang et al. [2010]  |
|                          | miR-373           | Up regulation        | CD44                          | 11  | Huang et al. [2008]  |
|                          | miR-520c          | Up regulation        | CD44                          | 11  | Huang et al. [2008]  |
| miR-125b                 | Up regulation     | EPO, EPOR            | 42                            | Ferracin et al. [2013]; van Schooneveld et al. [2015] |  |
| Liver                    | miR-18            | Up regulation        | -                             | 22  | Murakami et al. [2006]   |
|                          | miR-20            | Up regulation        | -                             | 22  | Murakami et al. [2006]   |
|                          | miR-195           | Down regulation      | -                             | 22  | Murakami et al. [2006]   |
|                          | miR-21            | Up regulation        | -                             | 20  | Li et al. [2009]   |
|                          | miR-101           | Down regulation      | -                             | 20  | Li et al. [2009]   |
| Ovarian                  | miR-92            | Up regulation        | -                             | 4   | Connolly et al. [2008]   |
|                          | miR-21            | Up regulation        | -                             | 38  | Resnick et al. [2009]  |
|                          | miR-155           | Down regulation      | -                             | 38  | Resnick et al. [2009]  |
|                          | miR-30c-1-3p      | Up regulation        | -                             | 24  | Häusler et al. [2010]  |
|                          | miR-342-3p        | Down regulation      | -                             | 24  | Häusler et al. [2010]  |
| Oral                     | miR-16            | Up regulation        | -                             | 35  | Suryawanshi et al. [2013]  |
|                          | let-7f            | Down regulation      | -                             | 360   | Zheng et al. [2013]  |
|                          | miR-21            | Up regulation        | -                             | 60  | Zahran et al. [2015]   |
|                          | miR-184           | Up regulation        | -                             | 60  | Zahran et al. [2015]   |
|                          | miR-145           | Down regulation      | -                             | 60  | Zahran et al. [2015]   |
|                          | miR-26a           | Down regulation      | <i>TMEM184B</i>               | 36  | Fukumoto et al. [2015]   |
|                          | miR-26b           | Down regulation      | <i>TMEM184B</i>               | 36  | Fukumoto et al. [2015]   |
|                          | miR-375           | Down regulation      | -                             | 51  | Lajer et al. [2011]  |
|                          | miR-31            | Up regulation        | -                             | 51  | Lajer et al. [2011]  |
|                          | miR-23            | Up regulation        | Mdm2, TSC1                    | Cell line   | Tang et al. [2011]   |
| Glioblastoma             | miR-10b           | Up regulation        | HOXD10, RhoC                  | 20  | Guessous et al. [2013]   |
|                          | miR-25            | Up regulation        | Mdm2, TSC1                    | 9/I, III  | Ciafre et al. [2005]   |
|                          | miR-16            | Up regulation        | BCL2                          | Cell line   | Chaudhry et al. [2010]   |
|                          | miR-19a           | Up regulation        | -                             | 118   | Jia et al. [2013]  |
|                          | miR-451           | Down regulation      | PI3K/AKT                      | Cell line   | Gal et al. [2008]  |
|                          | miR-145           | Down regulation      | Oct4, SOX2                    | Cell line   | Koo et al. [2012]  |
|                          | miR-373           | Up regulation        | -                             | 3   | Yang and Mei, 2015]  |
|                          | miR-181a          | Down regulation      | <i>CDKN1B</i>                 | 3   | Yang and Mei, 2015]  |
|                          | miR-125b          | Down regulation      | <i>CDK6, CDC25A</i>           | 3   | Yang and Mei, 2015]  |
|                          | let-7b            | Down regulation      | <i>CDK6, CDC25A</i>           | 3   | Yang and Mei, 2015]  |
| Prostate                 | miR-25            | Up regulation        | <i>BCL2L1</i>                 | 3   | Yang and Mei, 2015]  |
|                          | miR-18a           | Up regulation        | <i>BCL2L1</i>                 | 3   | Yang and Mei, 2015]  |
|                          | miR-20a           | Up regulation        | <i>BCL2L1</i>                 | 3   | Yang and Mei, 2015]  |
|                          | miR-141           | Up regulation        | -                             | 102   | Kelly et al. [2015]  |
|                          | miR-145           | Up regulation        | -                             | 102   | Kelly et al. [2015]  |
|                          | miR-155           | Up regulation        | -                             | 102   | Kelly et al. [2015]  |
|                          | let7a             | Down regulation      | -                             | 102   | Kelly et al. [2015]  |
|                          | miR-375           | Up regulation        | -                             | 102   | Kelly et al. [2015]  |
|                          | let-7             | Down regulation      | <i>KRAS</i>                   | Cell line   | Graziano et al. [2010]   |
|                          | miR-29            | Down regulation      | <i>MMP2, DNMT3A/B</i>         | Cell line   | Ding et al. [2011]   |
| Colon                    | miR-30a-5P        | Down regulation      | <i>DTL</i>                    | Cell line   | Baraniskin et al. [2012]   |
|                          | miR-34a           | Down regulation      | <i>FRA1, SIRT1, MYC, BCL2</i> | Cell line   | Schetter et al. [2012]   |
|                          | miR-17-92 cluster | Up regulation        | <i>E2F1</i>                   | Cell line   | Yu et al. [2012]   |
|                          | miR-95            | Up regulation        | <i>SNX1</i>                   | Cell line   | Huang et al. [2011]  |
|                          | miR-135a/b        | Up regulation        | <i>APC</i>                    | Cell line   | Luo et al. [2011]  |
| Gastric                  | miR-21            | Up regulation        | RECK                          | 59/I, II, III, IV                                     | Zhang et al. [2008]; Tsujiura et al. [2010]; Zheng et al. [2010] |
|                          | miR-17-5p         | Up regulation        | -                             | 87/I, II, III, IV                                     | Tsujiura et al. [2010]; Wang et al. [2012]                       |
|                          | miR-1             | Up regulation        | MET                           | 116/I, II, III, IV                                    | Liu et al. [2011]  |
|                          | miR-421           | Up regulation        | -                             | 141/I, II, III, IV                                    | Zhou et al. [2012]; Wu et al. [2014]                             |
|                          | miR-34            | Up regulation        | MET                           | 141/I, II, III, IV                                    | Zhou et al. [2012]; Wu et al. [2014]                             |
|                          | miR-195-5p        | Down regulation      | -                             | In vivo   | Gorur et al. [2013]  |
|                          | miR-196a          | Down regulation      | Annexin A1, HMGA2, HOXA8      | In vivo   | Tsai et al. [2012]   |
|                          | miR-203           | Down regulation      | EMT activators                | 130/I, II, III, IV                                    | Imaoka et al. [2015]   |
|                          | let-7a            | Down regulation      | -                             | 69/I, II, III, IV                                     | Arabpour et al. [2016]   |
|                          | miR-373           | Up regulation        | PPP6C                         | 110   | Wu et al. [2011]   |
| Hepatocellular carcinoma | miR-381           | Up regulation        | -                             | 110   | Murakami et al. [2013]   |

(Continued)

TABLE I. (Continued)

| Type of cancer | miRNA    | Expression in cancer | Target gene                       | Stage/patient sample | Citation   |
|----------------|----------|----------------------|-----------------------------------|----------------------|--|
|                | miR-130b | Up regulation        | TP53INP1                          | 57                   | Liu et al. [2012]; Wei et al. [2013]                                       |
|                | miR-20a  | Up regulation        |                                   | 110/I, II, III       | Fan et al. [2013]; Wei et al. [2013]                                       |
|                | miR-21   | Up regulation        | C/EBPb, RhoB, PDCD4, PTEN         | 137/I, II, III       | Ura et al. [2009]; Xu et al. [2011]; Wei et al. [2013]; Zhou et al. [2011] |
|                | miR-122  | Down regulation      | c-Myc, Bcl-w, ADAM-1, Wnt-1, MTTP | 90/B, C, D, A        | El-Garem et al. [2014]   |
|                | miR-223  | Down regulation      | STMN1                             | 110/I, II, III       | Ura et al. [2009]; Xu et al. [2011]; Wei et al. [2013]; Zhou et al. [2011] |

genes could contribute to cancer initiation and progression. In respect to the magnitude of miRNA genes in the pathogenesis and progression of cancer, the utilization of suitable methods for assessing miRNAs provide insight into new opportunities for cancer

treatment by modulating miRNA pathways and activities [Lee et al., 2008; Hernandez et al., 2013].

There are various conventional miRNA detection strategies including microarray, RT-PCR, and Northern blotting. Conventional

TABLE II. Various Techniques for Detecting MiRNAs in Cancer

| Technique  | miRNA        | Type of cancer              | Expression in cancer | Citation                   |                        |
|------------|--------------|-----------------------------|----------------------|----------------------------|------------------------|
| BLI        | miR-21       | Breast                      | Up regulation        | Hernandez et al. [2013]    |                        |
|            | miR-221      | Papillary thyroid carcinoma | Up regulation        | Kim et al. [2008]          |                        |
|            | miR-9        | Embryonic carcinoma         | Down regulation      | Ko et al. [2008]           |                        |
|            | miR-9        | Embryonic carcinoma         | Down regulation      | Ko et al. [2008]           |                        |
|            | miR-124a     | Embryonic carcinoma         | Up regulation        | Ko et al. [2009a]          |                        |
|            | miR-155      | Lung                        | Up regulation        | Yao et al. [2012]          |                        |
|            | miR-10b      | Breast                      | Up regulation        | Yigit et al. [2013]        |                        |
|            | miR-10b      | Adenocarcinomas             | Up regulation        | Yigit et al. [2013]        |                        |
|            | miR-1        | -                           | Up regulation        | Kang et al. [2015]         |                        |
|            | miR-26a      | -                           | Down regulation      | Kang et al. [2015]         |                        |
| Microarray | miR-124a     | -                           | Up regulation        | Kang et al. [2015]         |                        |
|            | miR-126      | -                           | Up regulation        | Kang et al. [2015]         |                        |
|            | miR-206      | -                           | Up regulation        | Kang et al. [2015]         |                        |
|            | miR-221      | -                           | Up regulation        | Kang et al. [2015]         |                        |
|            | miR-9        | -                           | Down regulation      | Kang et al. [2015]         |                        |
|            | miR-136      | Oral cancer                 | Down regulation      | Momen-Heravi et al. [2014] |                        |
|            | miR-147      | Oral cancer                 | Down regulation      | Momen-Heravi et al. [2014] |                        |
|            | miR-1250     | Oral cancer                 | Down regulation      | Momen-Heravi et al. [2014] |                        |
|            | miR-148a     | Oral cancer                 | Down regulation      | Momen-Heravi et al. [2014] |                        |
|            | miR-632      | Oral cancer                 | Down regulation      | Momen-Heravi et al. [2014] |                        |
|            | miR-646      | Oral cancer                 | Down regulation      | Momen-Heravi et al. [2014] |                        |
|            | miR-668      | Oral cancer                 | Down regulation      | Momen-Heravi et al. [2014] |                        |
|            | miR-877      | Oral cancer                 | Down regulation      | Momen-Heravi et al. [2014] |                        |
|            | miR-503      | Oral cancer                 | Down regulation      | Momen-Heravi et al. [2014] |                        |
|            | miR-220a     | Oral cancer                 | Down regulation      | Momen-Heravi et al. [2014] |                        |
|            | miR-323-5p   | Oral cancer                 | Down regulation      | Momen-Heravi et al. [2014] |                        |
|            | miR-24       | Oral cancer                 | Up regulation        | Momen-Heravi et al. [2014] |                        |
|            | miR-27b      | Oral cancer                 | Up regulation        | Momen-Heravi et al. [2014] |                        |
|            | let-7 family | Breast                      | Down regulation      | Litman et al. [2007]       |                        |
|            | miR-21       | Breast                      | Up regulation        | Litman et al. [2007]       |                        |
|            | miR-505-5p   | Breast                      | Down regulation      | Matamala et al. [2015]     |                        |
|            | miR-125b-5p  | Breast                      | Down regulation      | Matamala et al. [2015]     |                        |
|            | miR-21-5p    | Breast                      | Up regulation        | Matamala et al. [2015]     |                        |
|            | miR-96-5p    | Breast                      | Up regulation        | Matamala et al. [2015]     |                        |
|            | miR-576-5p,  | Glioblastoma                | Up regulation        | Dong et al. [2014]         |                        |
|            | miR-340      | Glioblastoma                | Up regulation        | Dong et al. [2014]         |                        |
|            | miR-626      | Glioblastoma                | Up regulation        | Dong et al. [2014]         |                        |
|            | miR-320      | Glioblastoma                | Down regulation      | Dong et al. [2014]         |                        |
|            | let-7g-5p    | Glioblastoma                | Down regulation      | Dong et al. [2014]         |                        |
|            | miR-7-5P     | Glioblastoma                | Down regulation      | Dong et al. [2014]         |                        |
| RT-PCR     | miR-223      | Gastric                     | Up regulation        | Gorur et al. [2013]        |                        |
|            | miR-106b     | Gastric                     | Up regulation        | Gorur et al. [2013]        |                        |
|            | miR-147      | Gastric                     | Up regulation        | Gorur et al. [2013]        |                        |
|            | miR-34a      | Gastric                     | Up regulation        | Gorur et al. [2013]        |                        |
|            | miR-130b     | Gastric                     | Up regulation        | Gorur et al. [2013]        |                        |
|            | miR-638      | Gastric                     | Down regulation      | Gorur et al. [2013]        |                        |
|            | miR-37       | Gastric                     | Down regulation      | Gorur et al. [2013]        |                        |
|            | miR18a       | Colon                       | Up regulation        | Giráldez et al. [2013]     |                        |
|            | miR19a       | Colon                       | Up regulation        | Giráldez et al. [2013]     |                        |
|            | miR15b,      | Colon                       | Up regulation        | Giráldez et al. [2013]     |                        |
|            | miR29a       | Colon                       | Up regulation        | Giráldez et al. [2013]     |                        |
|            | NGS          | miR-574-3p                  | Breast               | Down regulation            | Krishnan et al. [2015] |
|            |              | miR-660-5p                  | Breast               | Down regulation            | Krishnan et al. [2015] |

BLI: bioluminescence imaging, MB: molecular beacon, NGS: Next Generation Sequencing, RT-PCR: Reverse transcription polymerase chain reaction.

detection methods are associated with some limitations which indicate a necessity for devising and deploying high-throughput noninvasive repetitive and real-time imaging systems for the detection of miRNAs in preclinical and clinical settings [German et al., 2008; Ko et al., 2009a; Sun et al., 2010]. In the following section, we will describe recently developed miRNA imaging strategies such as the various bioluminescence systems, fluorescent imaging approaches, as well as magnetic resonance imaging. In addition, both the advantages and inherent inefficiencies of various imaging systems are also described.

Recent significant advancement in reporter-based optical imaging systems has provided the opportunities of noninvasive and repeated real-time analysis of the miRNA gene expression in living cells. These miRNA imaging approaches offer a better elucidation of the biogenesis and biological function of miRNAs in vivo as well as miRNAs expression profile in human diseases [Gambhir et al., 1999; Blasberg, 2003; Wang et al., 2003]. These imaging techniques could, for example, provide better data on intact biological context than the “snapshots” provided by in vitro assays [Lee et al., 2008; Hernandez et al., 2013].

Several studies indicated that there are different categories for miRNA imaging. One of the main categories is based on nanoparticles, Bioluminescent imaging (BLI), fluorescent proteins (FPs), and molecular beacon (MB) imaging.

Despite of much advancement in the field of imaging for detecting of miRNAs, this field is still in its infancy. Modern and new imaging techniques provide a new horizon for the study of various targets and molecules in different levels in living cells [Ottobri et al., 2006; Lee et al., 2008; Hernandez et al., 2013].

Some studies have been used bioluminescent reporter proteins such as Firefly luciferase (Fluc) and Gaussia luciferase (Gluc) for the imaging of various miRNAs in living cells [Ottobri et al., 2006]. Gaussia luciferase utilizes coelenterazine as a substrate and emits light with a peak at 480 nm with a broad spectrum extending to 600 nm. Firefly luciferase emits light with a peak at 562 nm. D-luciferin serves as a good substrate for Firefly luciferase [Gould and Subramani, 1988; Tannous et al., 2005].

Other systems are reporter-based miRNA detection imaging systems. In the presence of miRNA, these systems demonstrate a dropping in reporter signals which is correlated with translational repression of its target mRNA [Ko et al., 2008, 2009b; Kim et al., 2009]. These imaging systems could facilitate potential applications for assessing of miRNA levels during biological processes such as cell growth and differentiation and the cell cycle in living animals [Ko et al., 2009b]. For instance, these techniques could be utilized to detect and monitor differentiation patterns of stem cells by miRNA (cell-specific) expression in vivo or to detect miRNAs involved in cancer progression, for example, miR-221 and miR-21 [Ko et al., 2009b]. Table II represents a variety of miRNAs which are detected by various techniques.

## CONCLUDING REMARKS

MicroRNAs are a class of small non-coding RNA species, known as miRNAs, which control gene expression across various physiological and pathological processes. Their aberrant expression may be involved

in human diseases. Among human diseases, it has been shown that miRNAs are aberrantly expressed or mutated in cancer, indicating that they may play a role as a new class of oncogenes or tumor suppressor genes. As the miRNA field continues to grow and evolve, it is an important step to develop efficient tools for rapid, specific, sensitive, and noninvasive imaging detection of miRNAs toward understanding the functions of miRNAs in various regulatory pathways in vivo, which consequently effect on the development of miRNA-based diagnostic and therapeutic assays at molecular level and new targets in drug discovery. In contrast to conventional imaging systems, new molecular imaging systems such as reported-based optical imaging systems are multiplex and have high specificity against other family RNAs and minimum sample manipulation and could be utilized for studying living systems. However, once new generations of imaging systems (i.e., reporter-based imaging) involve genetic modification in studied subjects; there are still some concerns regarding their translation into clinical practice. In conclusion, molecular imaging techniques are robust tools for high-throughput noninvasive repetitive and real-time monitoring of biogenesis, localization patterns, and biological function of miRNAs in vivo as well as miRNAs expression profile in human diseases particularly cancer. Such imaging systems will deepen our knowledge of miRNAs expression patterns and their biological functions in various tumorigenic regulatory networks in vivo, which eventually effect on the development of miRNA-based diagnostic and therapeutic assays and new targets in cancer drug discovery.

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