

# A Frequency and Molecular Typing Study of Methicillin-Resistant *Staphylococcus aureus* Isolates in Teaching Hospitals in Shahrekord, SouthWestern Iran

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## Abstract

**Background:** Methicillin-resistant *Staphylococcus aureus* (MRSA) remains a significant public health problem and treatment challenge.

**Objectives:** This study was conducted to determine the frequency, molecular types, and drug resistance of *S. aureus* isolated from nasal carriers in two teaching hospitals (Hajar and Kashani) in Shahrekord, southwestern Iran.

**Methods:** In this cross-sectional study, 262 nasal specimens were obtained from healthcare staff. The disk-diffusion method was used to detect MRSA. Nine antibiotic disks were used to determine the antibiotic susceptibility pattern. Staphylococcal cassette chromosome mec (SCCmec) types were identified by the multiplex polymerase chain reaction (PCR). The data analysis was performed using Fisher's exact test with SPSS software.

**Results:** Forty-eight (18.8%) specimens were identified as *S. aureus*, of which 30 (11.45%) specimens were methicillin resistant. The nasal colonization rate of the MRSA isolates was not associated with age or gender ( $P > 0.05$ ). The highest resistance (33%) recorded was to rifampin, and all the isolates were susceptible to quinupristin-dalfopristin, vancomycin, and linezolid. The SCCmec results showed that 16.7%, 6.7%, 20%, and 56.6% of MRSA isolates were types I, II, III, and IV, respectively.

**Conclusions:** Nasal isolates of MRSA were prevalent among hospital staff. The highest level of resistance was to rifampin, and all the isolates were susceptible to quinupristin-dalfopristin, vancomycin, and linezolid. SCCmec type 4 was the most frequent MRSA isolate.

**Keywords:** Methicillin-Resistant, Molecular Typing, Drug Resistance, *Staphylococcus aureus*

## 1. Background

Many species of pathogenic bacteria, including *Staphylococcus aureus*, have become antibiotic resistant. The potential of *S. aureus* to develop resistance rapidly to many antibiotics has led to the emergence of methicillin-resistant *S. aureus* (MRSA) (1, 2). Methicillin resistance is associated with the production of the PBP2a protein by the *mecA* gene. This gene is located on a 30 - 50 kb chromosomal DNA fragment, which is present in resistant strains but absent in susceptible ones (2, 3). The staphylococcal cassette chromosome mec (SCCmec) is a mobile genetic element that carries the *mecA* gene and other antibiotic-resistant genes in MRSA strains (4, 5). Staphylococcal cassette chromosome mec typing is performed to identify and differentiate community-associated MRSA infections from healthcare-associated ones. Staphylococcal cassette chro-

mosome mec typing has been widely used to identify several subtypes or variants of the main SCCmec types. Staphylococcal cassette chromosome mec typing using the multiplex polymerase chain reaction (PCR) is a simple method that can be applied in clinical microbiology laboratories (6, 7).

Approximately 44% of nosocomial infections worldwide are estimated to be due to MRSA (8). Moreover, MRSA is associated with excessively high healthcare costs in many countries. For example, annual costs to deal with the consequences of MRSA in the U.S. are estimated at over 13.8-billion dollars (9, 10). The prevalence of MRSA continues to increase among healthcare staff, with a high rate of nasal colonization by MRSA reported (11). Methicillin resistant *S. aureus* can be transmitted via the skin and hands of healthcare staff through the nostrils and lead to infection

and a variety of complications in patients (12). MRSA infection can result in severe complications, such as endocarditis, septicemia, pneumonia, and osteomyelitis (13-15).

Restrictions placed on drug treatment due to MRSA have made it more difficult to combat nosocomial and community-acquired infections (16-18). The transmission of MRSA to patients via the hands or nostrils of health-care staff can result in hospitalized patients experiencing major problems (19). Methicillin resistant *S. aureus* infections among healthcare staff remain high, and the treatment and hospitalization costs of MRSA-infected patients are enormous. Therefore, strategies to control the spread of MRSA are needed (20). Given the importance of the prevention and treatment of nosocomial infections, data are needed on the incidence rate, prevalence, antibiotic resistance, and bacterial typing of MRSA.

## 2. Objectives

This study was conducted to determine the frequency, molecular types, and drug resistance of *S. aureus* nasal isolates in two teaching hospitals in Iran using PCR and disk-diffusion methods.

## 3. Methods

### 3.1. Specimen Collection and Bacterial Identification

After obtaining ethical approval for the study from the ethics committee of Shahrekord University of Medical Sciences (Grant No. 1184), 262 people were enrolled in the study: 149 staff from Kashani hospital and 113 staff from Hajar hospital. The occupational categories consisted of physicians, nurses, health workers, technicians, administrative staff, and service personnel from the surgical ward, intensive care unit, kitchen, and laundry room.

To ensure qualitative and quantitative standardization in the study, one individual collected all the samples and recorded the data. The nasal specimens were collected from the anterior nares of the participants using labeled sterile cotton wool swabs. The specimens were immediately transferred to trypticase soy broth medium and incubated at 37°C for 24 hours. The specimens were cultured in blood agar and mannitol salt agar media (Hi Media, India) in the laboratory. If the colony became yellow in the mannitol salt agar medium, gram staining was conducted. To differentiate *Staphylococcus* spp. in gram-positive cluster-forming cocci, catalase and coagulase (with rabbit plasma) production tests were conducted, as well as DNase activity tests on DNase agar (Hi Media, India) and novobiocin susceptibility tests (21, 22). To detect MRSA isolates, oxacillin (1 µg) disk (Hi Media, India)

diffusion testing was performed. To determine the antibiotic susceptibility of the isolates, eight antibiotic disks (gentamicin, linezolid, vancomycin, rifampin, novobiocin, ticoplanin, tigecycline, and quinopristin-dalfopristin) obtained from Hi Media (India) were tested using the Kirby-Bauer disk-diffusion method (23).

### 3.2. DNA Extraction

Total DNA was extracted from the bacteria that grew on the culture media with a genomic DNA purification kit (CinnaGen Co., Iran, according to the manufacturer's instructions). The quality of the extracted DNA was measured at 260 nm wavelength according to Sambrook and Russell's method (24). The extracted DNA was stored at -20°C for later use.

### 3.3. Multiplex PCR Assay for Assignment of the *mec* Element Type

A multiplex PCR assay was used to identify the MRSA isolates. The multiplex PCR method is a rapid, accurate, and useful assay to detect the *mecA* gene in MRSA strains, particularly in a hospital setting (25).

The multiplex PCR included eight loci, A-H, selected based on *mec* element sequences (Table 1) (26). The *mecA* gene was also included in this protocol. The PCR reactions were conducted in a total volume of 25 µL containing the following: 2 µL of DNA sample, 2.5 µL of 10× PCR buffer, 2 µL of mixed dNTP, 2.5 µL of MgCl<sub>2</sub>, 0.5 µL of DNA Taq polymerase, and 0.5 µL of primers A and D for type I; primers B, C, D, and G for type II; primers C, E, F, and H for type III, and primers D and I for type IV. The PCR assay was performed in a DNA Thermal Cycler 480 (Applied Biosystems, U.S.) using the following parameters: denaturation for 5 min at 95°C; 35 cycles of 94°C for 45 seconds, 53°C for 40 seconds, and 72°C for 1 minutes, followed by a final extension for 7 minutes at 72°C. For type IV, the annealing temperature was 52°C for 50 seconds. The PCR products then underwent polyacrylamide gel (8%) electrophoresis and staining with silver nitrate.

### 3.4. Statistical Analysis

The data were analyzed using Fisher's exact test with SPSS, version 16 (SPSS Inc., U.S.). The level of significance was considered as 0.05.

## 4. Results

In total, the presence of MRSA in 262 samples was tested. Of the 148 samples from Kashani hospital, 76 (51%) were from males. Of the 114 samples from Hajar hospital, 48 (43%) were from males. Fisher's exact test indicated no

**Table 1.** Primers Used in the Multiplex Polymerase Chain Reaction<sup>a</sup>

Locus	Primer	Oligonucleotide Sequence (5' - 3')	Location	Amplicon Size, bp	Specificity, SCCmec Type
A	CIF2 F2	TTCGAGTTGCTGATGAAGAAGG	18398 - 18419 <sup>b</sup>	495	I
	CIF2 R2	ATTACCACAAGGACTACCAGC	18892 - 18871 <sup>b</sup>		
B	KDP F1	AATCATCTGCCATTGGTGATGC	10445 - 10467 <sup>c</sup>	284	II
	KDP R1	CGAATGAAGTAAAGAAAGTGG	10728 - 10707 <sup>c</sup>		
C	MEC1 P2	ATCAAGACTTGCATTCAGGC	42428 - 42447 <sup>c</sup>	209	II, III
	MEC1 P3	GCGGTTCAATTCATCTGTC	42636 - 42617 <sup>c</sup>		
D	DCS F2	CATCCTATGATAGCTTGGTC	38011 - 37992 <sup>b</sup>	342	I, II, IV
	DCS R1	CTAAATCATAGCCATGACCG	37670 - 37689 <sup>b</sup>		
E	RIF4 F3	GTGATTGTTCGAGATATGTGG	45587 - 45607 <sup>d</sup>	243	III
	RIF4 R9	CGCTTATCTGTATCTATCCG	45829 - 45809 <sup>d</sup>		
F	RIF5 F10	TTCTTAAGTACACGCTGAATCG	59573 - 59594 <sup>d</sup>	414	III
	RIF5 R13	GTCACAGTAATCCATCAATGC	59986 - 59965 <sup>d</sup>		
G	IS431 P4	CAGGTCTCTCAGATCTACG	49963 - 49982 <sup>c</sup>	381	II
	pUB110 R1	GAGCCATAAACCAATAGCC	50343 - 50323 <sup>c</sup>		
H	IS431 P4	CAGGTCTCTCAGATCTACG	29654 - 29673 <sup>d</sup>	303	III
	pT181 R1	GAAGAATGGGGAAAGCTTCAC	29976 - 29956 <sup>d</sup>		
mecA	MECA P4	TCCAGATTACAACCTCACCAGG	1190 - 1211 <sup>e</sup>	162	Internal control
	MECA P7	CCACTTCATATCTTGAACG	1351 - 1332 <sup>e</sup>		

<sup>a</sup>Loci G and H were included to distinguish variants IA from I and IIIA from III, respectively.

<sup>b</sup>Relative to accession No. AB033763, SCCmec type I

<sup>c</sup>Relative to accession No. D86934, SCCmec type II

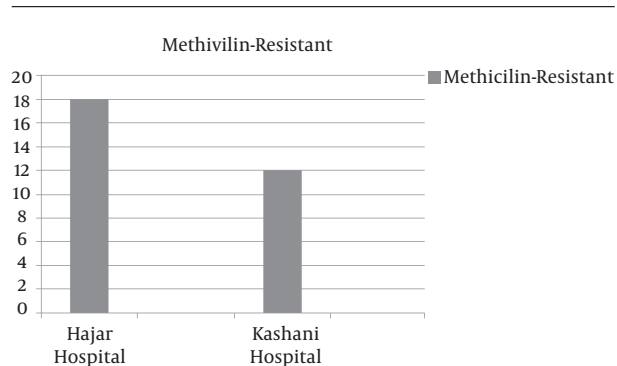
<sup>d</sup>Relative to accession No. AB037671, SCCmec type III

<sup>e</sup>Relative to accession No. Y00688, *mecA* gene

significant association between gender and the colonization rate of the MRSA isolates ( $P = 0.218$ ). The mean age of the participants from Hajar and Kashani hospitals was  $31.5 \pm 20$  and  $32 \pm 12$  (range: 21 - 51 and 21 - 45) years, respectively. In addition, Fisher's exact test indicated no significant association between age and the colonization rate of the MRSA isolates ( $P = 0.658$ ). The proportion, age, and gender of the *S. aureus*-positive participants are shown in Table 2.

Nineteen of the 148 (12.8%) isolates from Kashani hospital were identified as *S. aureus*, of which 12 (63% [8% of total isolates]) were MRSA. In Hajar hospital, 29 of the 114 isolates were identified as *S. aureus*, of which 18 (62% [15.7% of total isolates]) were MRSA (Figure 1). Based on a coagulase-positive test, *S. aureus* was detected in 48 samples (18%). In the oxacillin disk-diffusion test of the methicillin resistance of the coagulase-positive isolates, 30 of the 48 *S. aureus* isolates were MRSA.

To determine the prevalence of MRSA carriers among staff with different occupations in the hospitals, the participants were divided into three groups (physicians, nurses, and nontreatment). The latter consisted of service personnel, kitchen staff, office staff (secretaries and administrators), and guardians. In Hajar hospital, nurses (50%) ac-



**Figure 1.** Prevalence of Methicillin-Resistant *Staphylococcus aureus* in Each Hospital

counted for the highest proportion of carriers, and nontreatment staff accounted for the lowest proportion (22%). In Kashani hospital, nurses also comprised the highest proportion (41%) of carriers, and nontreatment staff comprised the lowest proportion (8%).

According to the antibiotic susceptibility analysis, the highest resistance obtained was to rifampin (33%), and all the isolates were susceptible to quinupristin-dalfopristin vancomycin, and linezolid. Table 3 summarizes the an-

**Table 2.** Demographic Characteristics of the Methicillin-Resistant *Staphylococcus aureus*-Positive Participants

Variable	Gender		Age	
	Female	Male	≤ 50	> 50
Group				
Frequency	18	30	42	6
%	37.5	62.5	87.5	12.5

tibiotic susceptibility pattern of the MRSA isolates. The *mecA* gene was detected in all the MRSA isolates by the PCR. The results of SCCmec typing based on the multiplex PCR method showed that of 30 tested isolates, five (16.7%) were type I, two (6.7%) were type II, six (20%) were type III, and seventeen (56.6%) were type IV MRSA (Figure 2). Figure 3 shows the SCCmec types of methicillin-resistant isolates detected in the two hospitals.

## 5. Discussion

The present study was conducted to determine the frequency and molecular types of MRSA isolated from nasal carriers in teaching hospitals in Shahrekord. In this study, 48 of 262 (18%) samples were identified as *S. aureus*, of which 30 (11.45%) were MRSA. In different studies of nasal samples obtained from healthcare staff, the prevalence of MRSA was reported to be 6.2%, 12.7%, 13.95%, 14.3%, 6.7%, and 18% in France, Ethiopia, Pakistan, India, and Saudi Arabia, respectively (27-32). Various prevalence rates (5.3 - 53.8%) of MRSA have been reported for *Staphylococcus* species isolated from individuals in different regions of Iran and from individuals in different occupations (33, 34). The different prevalence rates of this pathogen can be attributed to dissimilarities in healthcare policies (pattern of medication use), sample collection, nosocomial infection control, and the performance, education, and adherence of healthcare staff to hygiene-related recommendations, all of which potentially contribute to the distribution of resistant strains, including MRSA (30, 35-39).

In the present study, the colonization rate of MRSA isolates was not associated with age or gender. A previous study of the prevalence and antibiotic susceptibility pattern of nasal samples obtained from hospital staff and analyzed using the PCR method also found no significant difference in the carriage of this pathogen according to age and gender (33). However, Diawara et al. reported that age was significantly associated with being a carrier of this pathogen (40). Gebreyesus et al. demonstrated that women were more frequent carriers of the pathogen than men and that this finding was statistically significant (41). Several factors, such as the methodology of the study and occupational setting, may have contributed to these associations.

Nurses comprised 50% of the MRSA carriers in the present study. In other studies, nurses were also reported to be the most frequent carriers of MRSA (30, 32, 41, 42). The fact that nurses have more frequent contact with patients than other healthcare staff do and that they provide care to patients in different wards throughout the hospital likely explains this finding. As a result, nurses may have a greater risk than other staff of acquiring community-acquired MRSA. Therefore, the higher rate of MRSA acquisition among the nurses in the present study is not surprising.

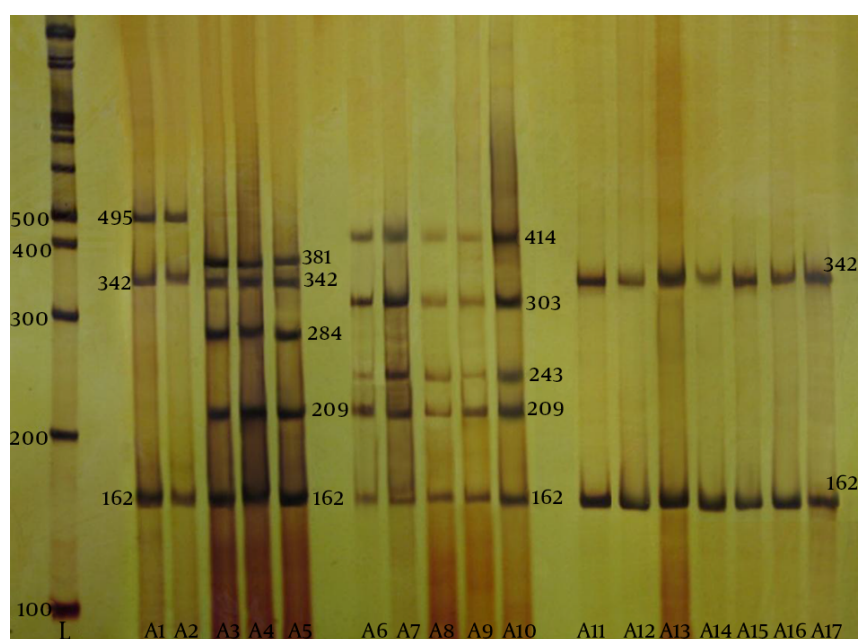
Among methicillin-resistant isolates, the highest resistance was to rifampin, and all the isolates were susceptible to quinipristin-dalfupristine, vancomycin, and linezolid. In a previous study, 64.1% of MRSA isolates were resistant to amikacin, and 76.92%, 51.28%, 87.18%, 71.8%, 10.26%, 5.13%, 89.74%, and 61.54% were resistant to ceftriaxone, ciprofloxacin, erythromycin, gentamicin, mupirocin, rifampin, tetracycline, and tobramycin, respectively (43). In the same study, all the MRSA and methicillin-susceptible *S. aureus* isolates were susceptible to fusidic acid, linezolid, teicoplanin, tigecycline, and vancomycin (43). In another study, MRSA isolates also appeared to exhibit high sensitivity to vancomycin, teicoplanin, and trimethoprim/sulfamethoxazole (44). Various factors, such as age, duration of treatment, and geographical region, have been shown to contribute to MRSA drug resistance (36, 45, 46). The results of the SCCmec typing in the present study demonstrated that 16.7% of MRSA isolates were type I, 6.7% were type II, 20% were type III, and 56.6% were type IV.

In a study of specimens obtained in a hospital setting, a PCR analysis revealed that SCCmec type I was the most frequent type (58.9%), followed by SCCmec type II (19.9%), type III (11.0%), and type IV (8.2%) (35). In another study, most isolates were SCCmec types II and IV (47). In a study of isolates obtained from healthcare staff, approximately half the specimens were types IVa and V, and no specimens were type I (48).

The prevalence of MRSA among healthcare staff in the two hospitals in southwestern Iran varied, and the estimated prevalence was lower than average. The findings indicated that the MRSA isolates showed the highest resistance to rifampin and that all the isolates were susceptible to quinipristin-dalfupristine, vancomycin, and lin-

**Table 3.** Antibiotic Susceptibility Pattern of Methicillin-Resistant *Staphylococcus aureus* Isolates

Antibiotic	Susceptible, No. (%)	Intermediate, No. (%)	Resistant, No. (%)
Bacitracin, 10 µg	24 (80)	0	6 (20)
Gentamycin, 10 µg	26 (86.7)	0	4 (13.3)
Novobiocin, 30 µg	25 (83)	1 (3)	4 (13.3)
Ticoplanin, 30 µg	18 (60)	5 (17)	7 (23)
Linezolid, 30 µg	30 (100)	0	0
Quinupristin-dalfopristin, 15 µg	30 (100)	0	0
Tigecycline, 15 µg	17 (56.7)	4 (13.3)	9 (30)
Rifampin, 5 µg	19 (63.4)	1 (3.3)	10 (33.3)
Vancomycin, 30 µg	30 (100)	0	0

**Figure 2.** View of the Stained Polyacrylamide Gel Showing Staphylococcal Cassette Chromosome mec (SCCmec) Types I-IV

From left to right, first lane, DNA marker; lane A1, standard SCCmec type 1; lane A2, SCCmec type 1; lane A3, standard SCCmec type 2; lanes A4 and A5, SCCmec type 2; lane A6, standard SCCmec type 3; lanes A7 to A10, SCCmec type 3; lane A11, standard SCCmec type IV; lanes A12 to A17, SCCmec type IV.

zolid. SCCmec type IV was the most prevalent MRSA isolate. Given the growth in the resistance of these bacteria to antibiotics, including methicillin, implementing plans to detect, control, and restrict carriers of MRSA, particularly healthcare staff who are in direct contact with patients, is vital to prevent the transmission of MRSA isolates to hospitalized patients.

### Acknowledgments

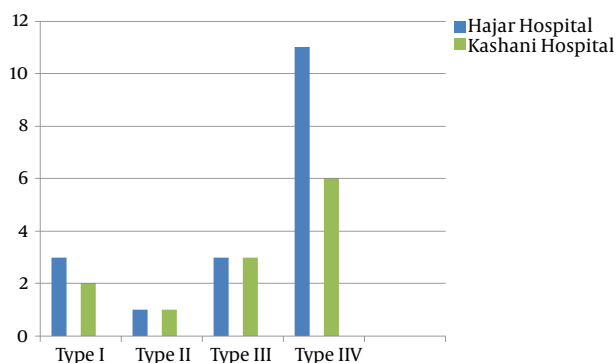
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### Footnotes

**Authors' Contribution:** Roohollah Taghaddosi contributed to drafting the manuscript and assisted in conducting the experiments. Maryam Safarpour Dehkordi and Marzieh Zeraatpisheh contributed to drafting the manuscript. Abolfazl Gholipour contributed to the design of the study and data analysis and revised the final version of the manuscript. Davood Darban Sarokhalil contributed to the data analysis and revised the final version of the manuscript. Fatemeh Heibati Goujani assisted in conducting the experiments.

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**Figure 3.** Staphylococcal Cassette Chromosome mec Typing Results for Methicillin-Resistant Packaging Samples in Each Hospital

Shahrekord University of Medical Sciences, Shahrekord, IR Iran (Grant No. 1184).

## References

- Chambers HF, Deleo FR. Waves of resistance: Staphylococcus aureus in the antibiotic era. *Nat Rev Microbiol.* 2009;7(9):629–41. doi: [10.1038/nrmicro2200](https://doi.org/10.1038/nrmicro2200). [PubMed: [19680247](https://pubmed.ncbi.nlm.nih.gov/19680247/)].
- Peacock SJ, Paterson GK. Mechanisms of Methicillin Resistance in Staphylococcus aureus. *Annu Rev Biochem.* 2015;84:577–601. doi: [10.1146/annurev-biochem-060614-034516](https://doi.org/10.1146/annurev-biochem-060614-034516). [PubMed: [26034890](https://pubmed.ncbi.nlm.nih.gov/26034890/)].
- Pantosti A, Sanchini A, Monaco M. Mechanisms of antibiotic resistance in Staphylococcus aureus. *Future Microbiol.* 2007;2(3):323–34. doi: [10.2217/17460913.2.3.323](https://doi.org/10.2217/17460913.2.3.323). [PubMed: [17661706](https://pubmed.ncbi.nlm.nih.gov/17661706/)].
- Zhang K, McClure JA, Elsayed S, Louie T, Conly JM. Novel multiplex PCR assay for characterization and concomitant subtyping of staphylococcal cassette chromosome mec types I to V in methicillin-resistant Staphylococcus aureus. *J Clin Microbiol.* 2005;43(10):5026–33. doi: [10.1128/JCM.43.10.5026-5033.2005](https://doi.org/10.1128/JCM.43.10.5026-5033.2005). [PubMed: [16207957](https://pubmed.ncbi.nlm.nih.gov/16207957/)].
- Hanssen AM, Ericson Sollid JU. SCCmec in staphylococci: genes on the move. *FEMS Immunol Med Microbiol.* 2006;46(1):8–20. doi: [10.1111/j.1574-695X.2005.00009.x](https://doi.org/10.1111/j.1574-695X.2005.00009.x). [PubMed: [16420592](https://pubmed.ncbi.nlm.nih.gov/16420592/)].
- Ghaznavi-Rad E, Nor Shamsudin M, Sekawi Z, van Belkum A, Neela V. A simplified multiplex PCR assay for fast and easy discrimination of globally distributed staphylococcal cassette chromosome mec types in methicillin-resistant Staphylococcus aureus. *J Med Microbiol.* 2010;59(Pt 10):1135–9. doi: [10.1099/jmm.0.021956-0](https://doi.org/10.1099/jmm.0.021956-0). [PubMed: [20616192](https://pubmed.ncbi.nlm.nih.gov/20616192/)].
- Milheirico C, Oliveira DC, de Lencastre H. Multiplex PCR strategy for subtyping the staphylococcal cassette chromosome mec type IV in methicillin-resistant Staphylococcus aureus: 'SCCmec IV multiplex'. *J Antimicrob Chemother.* 2007;60(1):42–8. doi: [10.1093/jac/dkm112](https://doi.org/10.1093/jac/dkm112). [PubMed: [17468509](https://pubmed.ncbi.nlm.nih.gov/17468509/)].
- Kock R, Becker K, Cookson B, van Gemert-Pijnen JE, Harbarth S, Kluytmans J, et al. Methicillin-resistant Staphylococcus aureus (MRSA): burden of disease and control challenges in Europe. *Euro Surveill.* 2010;15(41):19688. [PubMed: [20961515](https://pubmed.ncbi.nlm.nih.gov/20961515/)].
- Gould IM, Reilly J, Bunyan D, Walker A. Costs of healthcare-associated methicillin-resistant Staphylococcus aureus and its control. *Clin Microbiol Infect.* 2010;16(12):1721–8. doi: [10.1111/j.1469-0691.2010.03365.x](https://doi.org/10.1111/j.1469-0691.2010.03365.x). [PubMed: [20825434](https://pubmed.ncbi.nlm.nih.gov/20825434/)].
- Lee BY, Singh A, David MZ, Bartsch SM, Slayton RB, Huang SS, et al. The economic burden of community-associated methicillin-resistant Staphylococcus aureus (CA-MRSA). *Clin Microbiol Infect.* 2013;19(6):528–36. doi: [10.1111/j.1469-0691.2012.03914.x](https://doi.org/10.1111/j.1469-0691.2012.03914.x). [PubMed: [22712729](https://pubmed.ncbi.nlm.nih.gov/22712729/)].
- Elie-Turenne MC, Fernandes H, Mediavilla JR, Rosenthal M, Mathema B, Singh A, et al. Prevalence and characteristics of Staphylococcus aureus colonization among healthcare professionals in an urban teaching hospital. *Infect Control Hosp Epidemiol.* 2010;31(6):574–80. doi: [10.1086/652525](https://doi.org/10.1086/652525). [PubMed: [20426580](https://pubmed.ncbi.nlm.nih.gov/20426580/)].
- Aires De Sousa M, Santos Sanches I, Ferro ML, De Lencastre H. Epidemiological study of staphylococcal colonization and cross-infection in two West African Hospitals. *Microb Drug Resist.* 2000;6(2):133–41. doi: [10.1089/107662900419447](https://doi.org/10.1089/107662900419447). [PubMed: [10990268](https://pubmed.ncbi.nlm.nih.gov/10990268/)].
- Bassetti M, Treccarichi EM, Mesini A, Spanu T, Giacobbe DR, Rossi M, et al. Risk factors and mortality of healthcare-associated and community-acquired Staphylococcus aureus bacteraemia. *Clin Microbiol Infect.* 2012;18(9):862–9. doi: [10.1111/j.1469-0691.2011.03679.x](https://doi.org/10.1111/j.1469-0691.2011.03679.x). [PubMed: [21999245](https://pubmed.ncbi.nlm.nih.gov/21999245/)].
- Townell NJ, Munckhof WJ, Nimmo G, Bannan A, Holley A, Daniel A, et al. Community-associated methicillin-resistant Staphylococcus aureus endocarditis 'down under': case series and literature review. *Scand J Infect Dis.* 2012;44(7):536–40. doi: [10.3109/00365548.2012.664779](https://doi.org/10.3109/00365548.2012.664779). [PubMed: [22404422](https://pubmed.ncbi.nlm.nih.gov/22404422/)].
- Tattevin P, Schwartz BS, Graber CJ, Volinski J, Bhukhen A, Bhukhen A, et al. Concurrent epidemics of skin and soft tissue infection and bloodstream infection due to community-associated methicillin-resistant Staphylococcus aureus. *Clin Infect Dis.* 2012;55(6):781–8. doi: [10.1093/cid/cis527](https://doi.org/10.1093/cid/cis527). [PubMed: [22670044](https://pubmed.ncbi.nlm.nih.gov/22670044/)].
- Otter JA, French GL. Molecular epidemiology of community-associated methicillin-resistant Staphylococcus aureus in Europe. *Lancet Infect Dis.* 2010;10(4):227–39. doi: [10.1016/S1473-3099\(10\)70053-0](https://doi.org/10.1016/S1473-3099(10)70053-0). [PubMed: [20334846](https://pubmed.ncbi.nlm.nih.gov/20334846/)].
- Cimolai N. Methicillin-resistant Staphylococcus aureus in Canada: a historical perspective and lessons learned. *Can J Microbiol.* 2010;56(2):89–120. doi: [10.1139/w09-109](https://doi.org/10.1139/w09-109). [PubMed: [20237572](https://pubmed.ncbi.nlm.nih.gov/20237572/)].
- David MZ, Daum RS. Community-associated methicillin-resistant Staphylococcus aureus: epidemiology and clinical consequences of an emerging epidemic. *Clin Microbiol Rev.* 2010;23(3):616–87. doi: [10.1128/CMR.00081-09](https://doi.org/10.1128/CMR.00081-09). [PubMed: [20610826](https://pubmed.ncbi.nlm.nih.gov/20610826/)].
- Hosainzadegan H, Menati S, Tarahi M, Mohammadi F. Methicillin and vancomycin-resistant Staphylococcus aureus colonization frequency among hospital staff shohada Khorramabad. *J Med Lab Sci.* 2008;2(1):26–32.
- Farr BM, Jarvis WR. Would active surveillance cultures help control healthcare-related methicillin-resistant Staphylococcus aureus infections?. *Infect Control Hosp Epidemiol.* 2002;23(2):65–8. doi: [10.1086/502008](https://doi.org/10.1086/502008). [PubMed: [11893150](https://pubmed.ncbi.nlm.nih.gov/11893150/)].
- Reisner SB, Woods GL, Thomson RP. Specimen collection. In: Murray PR, Baron EJ, Pfaller MA, Tenoer FC, Tenover R. H. , editors. *Manual of Clinical Microbiology*. Washington: American Society for Microbiology; 1999. pp. 64–76.
- Pezlo MT, Amsterdam D, Anhalt JP, Lawrence T, Stratton NJ, Vetter EA, et al. Detection of bacteriuria and pyuria by URISCREEN a rapid enzymatic screening test. *J Clin Microbiol.* 1992;30(3):680–4. [PubMed: [1551986](https://pubmed.ncbi.nlm.nih.gov/1551986/)].
- Khan RA, Rahman AU, Ahmad A, Jaseem M, Jabbar A, Khan SA, et al. Prevalence and Antibiotic Susceptibility Profile of Methicillin-Resistant Staphylococcus aureus (MRSA) Isolated from Different Clinical Samples in District Peshawar. *J Appl Environ Biol Sci.* 2014;4(8S):40–6.
- Sambrook J, Russell DW. *Molecular cloning: a laboratory manual* 3rd edition. UK: ColdSpring-Harbour Laboratory Press; 2001.
- Pournajaf A, Ardebili A, Goudarzi L, Khodabandeh M, Narimani T, Abbaszadeh H. PCR-based identification of methicillin-resistant Staphylococcus aureus strains and their antibiotic resistance profiles. *Asian Pac J Trop Biomed.* 2014;4(Suppl 1):S293–7. doi: [10.12980/APJT.B.2014C423](https://doi.org/10.12980/APJT.B.2014C423). [PubMed: [25183100](https://pubmed.ncbi.nlm.nih.gov/25183100/)].

26. Oliveira DC, de Lencastre H. Multiplex PCR strategy for rapid identification of structural types and variants of the mec element in methicillin-resistant *Staphylococcus aureus*. *Antimicrob Agents Chemother*. 2002;**46**(7):2155–61. [PubMed: [12069968](#)].
27. Eveillard M, Martin Y, Hidri N, Boussougant Y, Joly-Guillou ML. Carriage of methicillin-resistant *Staphylococcus aureus* among hospital employees: prevalence, duration, and transmission to households. *Infect Control Hosp Epidemiol*. 2004;**25**(2):114–20. doi: [10.1086/502360](#). [PubMed: [14994935](#)].
28. Zakai SA. Prevalence of methicillin-resistant *Staphylococcus aureus* nasal colonization among medical students in Jeddah, Saudi Arabia. *Saudi Med J*. 2015;**36**(7):807–12. doi: [10.15537/smj.2015.7.11609](#). [PubMed: [26108584](#)].
29. Shibabaw A, Abebe T, Mihret A. Nasal carriage rate of methicillin resistant *Staphylococcus aureus* among Dessie Referral Hospital Health Care Workers; Dessie, Northeast Ethiopia. *Antimicrob Resist Infect Control*. 2013;**2**(1):25. doi: [10.1186/2047-2994-2-25](#). [PubMed: [24088259](#)].
30. Rashid Z, Farzana K, Sattar A, Murtaza G. Prevalence of nasal *Staphylococcus aureus* and methicillin-resistant *Staphylococcus aureus* in hospital personnel and associated risk factors. *Acta Pol Pharm*. 2012;**69**(5):985–91. [PubMed: [23061297](#)].
31. M R, D'Souza M, Kotigadde S, Saralaya KV, Kotian MS. Prevalence of Methicillin Resistant *Staphylococcus aureus* Carriage amongst Health Care Workers of Critical Care Units in Kasturba Medical College Hospital, Mangalore, India. *J Clin Diagn Res*. 2013;**7**(12):2697–700. doi: [10.7860/JCDR/2013/5160.3735](#). [PubMed: [24551616](#)].
32. Al-Humaidan OS, El-Kersh TA, Al-Akeel RA. Risk factors of nasal carriage of *Staphylococcus aureus* and methicillin-resistant *Staphylococcus aureus* among health care staff in a teaching hospital in central Saudi Arabia. *Saudi Med J*. 2015;**36**(9):1084–90. doi: [10.15537/smj.2015.9.12460](#). [PubMed: [26318466](#)].
33. Askarian M, Zeinalzadeh A, Japoni A, Alborzi A, Memish ZA. Prevalence of nasal carriage of methicillin-resistant *Staphylococcus aureus* and its antibiotic susceptibility pattern in healthcare workers at Namazi Hospital, Shiraz, Iran. *Int J Infect Dis*. 2009;**13**(5):241–7. doi: [10.1016/j.ijid.2008.11.026](#). [PubMed: [19269873](#)].
34. Taherikalani M, Mohammadzad MR, Soroush S, Maleki MH, Azizi-Jalilian F, Pakzad I, et al. Determining the prevalence of SCCmec polymorphism, virulence and antibiotic resistance genes among methicillin-resistant *Staphylococcus aureus* (MRSA) isolates collected from selected hospitals in west of Iran. *J Chemother*. 2016;**28**(2):104–9. doi: [10.1179/1973947815Y.0000000018](#). [PubMed: [25976554](#)].
35. Rhee Y, Aroutcheva A, Hota B, Weinstein RA, Popovich KJ. Evolving Epidemiology of *Staphylococcus aureus* Bacteremia. *Infect Control Hosp Epidemiol*. 2015;**36**(12):1417–22. doi: [10.1017/ice.2015.213](#). [PubMed: [26372679](#)].
36. Ebrahim-Saraie HS, Motamedifar M, Sarvari J, Hoseini Alfatemi SM. Emergence of SCCmec Type I Obtained From Clinical Samples in Shiraz Teaching Hospitals, South-West of Iran. *Jundishapur J Microbiol*. 2015;**8**(6):16998. doi: [10.5812/jjm.16998v2](#). [PubMed: [26322200](#)].
37. Akhtar N. Staphylococcal nasal carriage of health care workers. *J Coll Physicians Surg Pak*. 2010;**20**(7):439–43. doi: [07.2010/JCPS.439443](#). [PubMed: [20642942](#)].
38. Balbale SN, Hill JN, Guihan M, Hogan TP, Cameron KA, Goldstein B, et al. Evaluating implementation of methicillin-resistant *Staphylococcus aureus* (MRSA) prevention guidelines in spinal cord injury centers using the PARIHS framework: a mixed methods study. *Implement Sci*. 2015;**10**:130. doi: [10.1186/s13012-015-0318-x](#). [PubMed: [26353798](#)].
39. Kullar R, Vassallo A, Turkel S, Chopra T, Kaye KS, Dhar S. Degowning the controversies of contact precautions for methicillin-resistant *Staphylococcus aureus*: A review. *Am J Infect Control*. 2016;**44**(1):97–103. doi: [10.1016/j.ajic.2015.08.003](#). [PubMed: [26375351](#)].
40. Diawara I, Bekhti K, Elhabchi D, Saile R, Elmdaghri N, Timinouni M, et al. *Staphylococcus aureus* nasal carriage in hemodialysis centers of Fez, Morocco. *Iran J Microbiol*. 2014;**6**(3):175–83. [PubMed: [25870751](#)].
41. Gebreyesus A, Gebre-Selassie S, Mihert A. Nasal and hand carriage rate of methicillin resistant *Staphylococcus aureus* (MRSA) among health care workers in Mekelle Hospital, North Ethiopia. *Ethiop Med J*. 2013;**51**(1):41–7. [PubMed: [23930490](#)].
42. Ohadian Moghadam S, Pourmand MR, Davoodabadi A. The Detection of Mupirocin Resistance and Nasal Carriage of Methicillin Resistant *Staphylococcus aureus* among Healthcare Workers at University Hospitals of Tehran, Iran. *Iran J Public Health*. 2015;**44**(3):361–8. [PubMed: [25905079](#)].
43. Ohadian Moghadam S, Pourmand MR, Aminharati F. Biofilm formation and antimicrobial resistance in methicillin-resistant *Staphylococcus aureus* isolated from burn patients, Iran. *J Infect Dev Ctries*. 2014;**8**(12):1511–7. doi: [10.3855/jidc.5514](#). [PubMed: [25500648](#)].
44. Kim SH, Kim MG, Kim SS, Cha SH, Yeo SG. Change in Detection Rate of Methicillin-Resistant *Staphylococcus aureus* and *Pseudomonas aeruginosa* and Their Antibiotic Sensitivities in Patients with Chronic Suppurative Otitis Media. *J Int Adv Otol*. 2015;**11**(2):151–6. doi: [10.5152/iao.2015.1106](#). [PubMed: [26381007](#)].
45. Jung MY, Chung JY, Lee HY, Park J, Lee DY, Yang JM. Antibiotic Susceptibility of *Staphylococcus aureus* in Atopic Dermatitis: Current Prevalence of Methicillin-Resistant *Staphylococcus aureus* in Korea and Treatment Strategies. *Ann Dermatol*. 2015;**27**(4):398–403. doi: [10.5021/ad.2015.27.4.398](#). [PubMed: [26273155](#)].
46. Popovich KJ, Smith KY, Khawcharoenporn T, Thurlow CJ, Lough J, Thomas G, et al. Community-associated methicillin-resistant *Staphylococcus aureus* colonization in high-risk groups of HIV-infected patients. *Clin Infect Dis*. 2012;**54**(9):1296–303. doi: [10.1093/cid/cis030](#). [PubMed: [22354926](#)].
47. Lee CS, Montalmont B, O'Hara JA, Syed A, Chaussard C, McGaha TL, et al. Screening for methicillin-resistant *Staphylococcus aureus* colonization using sponges. *Infect Control Hosp Epidemiol*. 2015;**36**(1):28–33. doi: [10.1017/ice.2014.4](#). [PubMed: [25627758](#)].
48. Adwan K, Jarrar N, Abu-Hijleh A, Adwan G, Awwad E, Salameh Y. Molecular analysis and susceptibility patterns of methicillin-resistant *Staphylococcus aureus* strains causing community- and health care-associated infections in the northern region of Palestine. *Am J Infect Control*. 2013;**41**(3):195–8. doi: [10.1016/j.ajic.2012.03.040](#). [PubMed: [22998783](#)].