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1 **Jet dispensing of multi-layered films for the co-delivery of three antihypertensive agents**

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15 **Abstract**

16 Three-layer thin films comprising of two polymers as substrate (ethyl cellulose and,
17 copovidone K28) and three antihypertensive agents (hydrochlorothiazide, amiloride HCl and
18 carvedilol) were printed using jet-dispensing technology. Two film formulations with different
19 ethyl cellulose to copovidone K28 ratio (i.e. 90/10 and 50/50 w/w) were prepared using a three-
20 course dispensing. The films were characterized regarding surface morphology, solid-state
21 properties, polymer-drug interactions, drug distribution in each layer and their *in-vitro* drug
22 release. All the components of the films were found to be in the amorphous state apart from
23 hydrochlorothiazide which retained its crystallinity. FT-IR spectroscopy revealed hydrogen
24 bond interactions between carvedilol and copovidone K28. Combinations of ethyl cellulose
25 and copovidone K28 provide suitable polymeric film substrates with the ability to modify drug
26 release. Particularly, decreased ethyl cellulose to copovidone K28 weight ratio was found to
27 suppress the crystallization of hydrochlorothiazide and to increase the release rate of the
28 dispensed drugs. Jet dispensing was found to be a rapid technology for the preparation of multi-
29 layered films that can be used as personalized formulations for the delivery of combinations of
30 drugs.

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32 **Keywords:** jet dispensing, films, combination therapy, antihypertensive drugs

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34 1. Introduction

35 Combination treatment has now become the standard line of treatment for cardiovascular
36 diseases reducing the burden of morbidity and mortality associated with these pathologies [1].
37 Especially, for the management of hypertension, monotherapy is effective in achieving the
38 target goal in only about 50% of patients and treatment with two or more agents from classes
39 with different pharmacological action is often necessary to achieve adequate blood pressure
40 control [2]. Moreover, combination therapy offers several advantages over monotherapy such
41 as superior antihypertensive efficacy, dose strength and fewer adverse effects. For that purpose,
42 the combination of multiple therapeutic agents into a single formulation with appropriate
43 release profiles and doses (potentially optimized for individuals) has been suggested as an
44 alternative approach [3].

45 The most recent methods for the manufacturing of multi-dosage formulations with well-defined
46 release profiles make use of 3D printing technologies. Khaled *et al.* applied 3D extrusion-based
47 printing for the manufacturing of multi-active solid dosage forms containing three [4] and five
48 compartmentalized drugs [5] with independently controlled and well-defined release profiles
49 indicating that printing technologies hold potential for the tailored manufacturing of medicines
50 paving the way towards personalized care and treatment.

51 Jet dispensing is a technology that can continuously dispense liquids with a wide range of
52 viscosities by moving the nozzle at high speeds across the x-axis and the stage across y-axis to
53 cover all coordinates and jetting precise volume of dots. It is a highly accurate technique and
54 can deliver nanoquantities of drugs which cannot be achieved by the current 3D printing
55 technology. According to this technology, the jetting device operates in a continuous mode by
56 using a pneumatic piston with a ball tip end to push fluid through a narrow orifice at the jet
57 nozzle tip. Air pressure is used to apply force and lifts ball-needle from its seat, thereby fluid

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58 is allowed to flow down and around the ball-needle tip. Subsequently, the air pressure is
59 released allowing the force from the compressed spring to slam the ball-needle tip back down
60 into its seat, separating and ejecting a dot from the fluid. As the ball returns, the force due to
61 acceleration breaks the stream of the drug–polymer solution, which is jetted through the nozzle.
62 The broken stream of the solution strikes the substrate from 1.0 mm to 3.5 mm above the board
63 forming the dot. In the field of pharmaceuticals, Scoutaris *et al.* [6] reported the use of jet
64 dispensing as a rapid and reproducible technology for the preparation of taste-masked mucosal
65 thin films with drug loadings ranging from 20 to 40% and excellent content uniformity.
66 Recently, Scoutaris *et al.* [7] investigated the use of jet dispensing as a high-throughput
67 screening technology for pharmaceutical cocrystals.

68 Hydrochlorothiazide (HCTZ) is a thiazide diuretic that works by increasing the amount of salt
69 and water that the kidney removes from the blood causing a decrease in blood volume. It exerts
70 its mechanism of action by inhibiting the re-absorption of salts from the renal tubules leading
71 to increased excretion of sodium and chloride ions and consequently of water. HCTZ is
72 employed for the treatment of oedema and hypertension. Its usual adult dose in the case of
73 hypertension is 25 mg orally once daily that can be increased to 50 mg daily as a single or two
74 divided doses [8]. Amiloride HCl (AMI) is a potassium-sparing diuretic which promotes
75 excretion of sodium and chloride while retaining potassium. AMI is prescribed as adjunct
76 therapy to thiazide or loop diuretics for the treatment of hypertension to achieve potassium
77 conservation and thus avoid hypokalaemia. Co-administration of AMI/HCTZ 2.5/25 mg is a
78 common fixed dose combination for the treatment of hypertension known with the non-
79 proprietary name co-amilozide. Carvedilol (CARV) is a non-selective β -blocking agent that
80 has been approved for the treatment of heart failure, essential hypertension and coronary artery
81 diseases. Controlled-release once-daily formulations of CARV are commercially available and
82 considered as a more convenient option conferring an adherence benefit over immediate-

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83 release formulations in conditions traditionally prone to noncompliance such as hypertension
84 [9]. CARV and HCTZ are often administered together as a fixed dose combination for the
85 treatment of essential hypertension particularly in cases where monotherapy cannot provide
86 sufficient lowering of the blood pressure [10]. A combination of more than two drugs may be
87 required for the adequate management of blood pressure. In such cases the third agent added
88 to the medication should belong to a different pharmacological class than the initial two drugs
89 [11].

90 The aim of this work is to use jet dispensing technology for the preparation of films which
91 combine multiple drugs with different dose and independent release profiles. Two polymers
92 (ethyl cellulose and copovidone K28) were used as substrate and hydrochlorothiazide,
93 amiloride HCl and carvedilol as the antihypertensive agents. The films produced were
94 characterized regarding their morphology, solid-state properties, drug-polymer interactions and
95 distribution in each layer. The effect of the ratio of the two polymers on the *in-vitro* drug
96 dissolution was also investigated. To the best of our knowledge this is the first study where the
97 use of jet dispensing technology is extended to the preparation of films containing multiple
98 drugs with independent release profiles.

100 2. Materials and methods

101 2.1. Materials

102 Hydrochlorothiazide (HCTZ), carvedilol (CARV) and amiloride hydrochloride hydrate (AMI)
103 were purchased from TCI (Tokyo Chemical Industry Ltd, UK). Ethyl cellulose (EC N10
104 Pharm) and Kollidon® VA 64 (copovidone K28) were purchased from Hercules GmbH (USA)
105 and BASF (Germany), respectively. The chemical structures of the antihypertensive agents and
106 polymers used for the preparation of the films are depicted in Fig. 1. Ethanol and methanol were
107 purchased from Sigma-Aldrich (UK). All the solvent used were of analytical grade.

108 2.2. Methods

109 2.2.1. Preparation of films by jet dispensing

110 According to the dosage design, the multi-layered structure of the film and the presence of the
111 polymers will allow controlling the release of each drug. Specifically, CARV and AMI are
112 included in the top layers of the film (layers 2 and 3) in order to achieve immediate release
113 which will be followed by controlled release of HCTZ and CARV embedded in the polymeric
114 matrix of the bottom layer (layer 1). A schematic diagram of the film is illustrated in Fig. 2a.

115 Jet dispensing was carried out using a laboratory DispenseMate 583 dispenser (Nordson-
116 Asymtek, Netherlands). The layers of the films were prepared by jetting solution of the drugs
117 and polymers in several parallel lines. For the purposes of this study, the nozzle speed and
118 jetting rate were set at 9 mm s^{-1} and 33 drops s^{-1} , respectively. The fluid pressure used was 11.4
119 bar and the nozzle $100 \mu\text{m}$. (Fig. 2b). The size of the ball tip and the seat was 2.4 mm and 0.32
120 μm , respectively. For the first layer, HCTZ (7.0 mg ml^{-1}), CARV (0.7 mg ml^{-1}), EC (13.87 mg
121 ml^{-1}) and copovidone K28 (1.54 mg ml^{-1}) were dissolved in ethanol. The same method was
122 applied to deposit pure AMI on the surface of the first films. AMI (10 mg ml^{-1}) was dissolved
123 in a mixture of ethanol/water (60/40 v/v). The precursor solution leading to the formation of

124 the third layer consisted of CARV (30.3 mg ml⁻¹) and copovidone K28 (60.6 mg ml⁻¹) dissolved
125 in methanol. Two types of formulations were prepared which differ in the ratio of
126 EC/copovidone K28 incorporated in the first layer of the film (i.e. 90/10 and 50/50) while layer
127 2 and 3 were common for both types of films. Table 1 summarizes the nominal composition of
128 each layer for the two formulations of jet-dispensed films prepared in this study and the volume
129 of the precursor solutions printed per film.

130 **2.2.2. Characterization of the films**

131 **Scanning electron microscopy**

132 Scanning electron microscopy (SEM) was used to study the surface morphology of the films.
133 Samples were placed on the holder with double-sided carbon tape and sputter-coated with gold
134 (Edwards 188, sputter coater S1508) under argon atmosphere. SEM micrographs were taken
135 using a Hitachi SU 8030 (Japan), at an accelerating voltage of 1.0 kV.

136 **X-ray diffraction**

137 X-ray diffraction (XRD) patterns were obtained with a Bruker D8 Advance (Germany). Cu K α
138 radiation at 40 mA and 40 kV with a step size of 0.02 2 theta and a speed of 0.3 s per step were
139 used, covering a 2 theta of 5-40°. The samples were rotated at 15 rpm. For the raw materials,
140 which were in the form of powders an exit slit of 0.2 mm was used. XRD of the films was
141 carried out using the transmission mode with a 0.6 mm exit slit. DIFFRACplus XRD
142 Commander was the analysis software.

143 **Differential scanning calorimetry**

144 Differential scanning calorimetry (DSC) was performed using a Mettler-Toledo 823e
145 (Switzerland) previously calibrated with indium. Weighed powder samples (2-5 mg) were
146 sealed in aluminium pans with a pierced lid and heated under nitrogen flow (50 ml min⁻¹) at a
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148 heating rate of 10 °C min⁻¹. The heating range for the polymers and the films was from 25 to
149 300 °C while for the raw drugs from 25 to 30 °C above the melting point.

150 **Fourier-transform infrared spectroscopy**

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152 Spectra of the raw drugs, polymers and jet-dispensed (JD) films were recorded using a
153 Spectrum100 FT-IR spectrometer (PerkinElmer, Inc., USA) equipped with the attenuated total
154 reflection sampling accessory. Samples were scanned at room temperature, at the transmission
155 mode, over a wavenumber range of 4000–450 cm⁻¹, 16 accumulations and 1 cm⁻¹ resolution.
156 Before each measurement, a background spectrum of air was acquired under the same
157 instrumental conditions. The acquired spectra were processed using the PerkinElmer Spectrum
158 Express software.

159 160 **Confocal Raman spectroscopy**

161 Raman mapping was performed in the JD films 50/50 EC/copovidone K28 using a Jobin-Yvon
162 LabRam 320 instrument equipped with an Olympus microscope (Horiba, Japan) by means of
163 a He–Ne ion laser ($\lambda = 632.8$ nm) and 1800 nm⁻¹. The experimental conditions were 100 nm
164 slit width, a 50 × Microsoft objective and 0.2 s acquisition times. Each spectrum was the mean
165 of two. The sample profiling was performed at step increments of 8 μ m in the X–Y direction
166 over an area of 2000 × 600 μ m². Principal component analysis (PCA) was used for analysis of
167 the data. Prior to the analysis, all spectra were baseline corrected and normalized using the
168 standard normal variate method (SNV) to avoid intensity deviation among the Raman spectra.
169 The Raman chemical maps were constructed by using Solo + Mia software (Eigenvector,
170 Research, Inc., USA).

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2.2.3 Drug content and *in-vitro* dissolution studies

Drug content analysis

Jet-dispensed films were dissolved in a solution of 20% methanol in water. Solutions were filtered through a 0.45 µm PTFE syringe filters. The filtrate was analysed by HPLC-UV (Agilent 1200 Series, Agilent technologies, Germany). The stationary phase was a Waters Spherisorb® C-18 column (250 mm x 4.6 mm x 5 µm; Waters Corporation, USA). The column was maintained at 40 °C and the injection volume was 20 µl. The monitored wavelength was 240 nm. For the simultaneous quantification of the three drugs, a gradient method was developed. The mobile phase consisted of the polar solvent A which was water with 1% triethanolamine (pH:2.2 adjusted by addition of orthophosphoric acid) and the organic solvent B which was acetonitrile. The flow rate was 1 ml min⁻¹ and the gradient schedule was as follows: 0-6 min isocratic at 20% B, 6-10 min 20 → 70% B, 10-20 min isocratic at 70% B, followed by a linear change to initial conditions in 5 min of column re-equilibration. For each drug, the correlation coefficient of the calibration curve was R² > 0.999 for a concentration range of 5-100 µg ml⁻¹, indicating acceptable linearity. Drug content analysis was carried out in triplicate for each formulation (n=3).

In-vitro dissolution studies

The *in-vitro* dissolution study of the jet-dispensed films was carried out using the USP apparatus type II (Varian 705 DS, Varian Inc., USA) at 37 °C and 100 rpm stirring speed. To evaluate the dissolution of the formulations under conditions mimicking the gastrointestinal environment, jet-dispensed films were subjected to dissolution testing for 2 h in hydrochloric acid media (900 mL) at pH:1.2 and then were transferred for 3 h in phosphate buffer solution at pH:6.8. At specific time intervals up to 5 h, 5 ml of dissolution medium was withdrawn, filtered through 0.45 mm PTFE syringe filter and placed in HPLC vials for assay while

197 immediately replaced with 5 ml of fresh medium. The HPLC conditions for the assay were
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2 198 identical to those for drug content determination. Dissolution tests were made in triplicate for
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5 199 each formulation (n = 3).
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201 3. Results and discussion

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3 202 Films of 1 cm² were produced by jet dispensing (Fig. 3). The size of the film is close to that of
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6 203 a five-pence coin showing that they could be easily placed in the oral cavity and get swallowed.
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8 204 Jet dispensing appeared to be capable for rapid and reproducible production of thin films
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10 205 comprising of drugs with remarkably different water solubility in a wide dose range. Compared
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12 206 to the conventional method of solvent casting where a common solvent system should be
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14 207 selected to dissolve all drugs and polymers, in jet dispensing drug and polymer solutions in
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16 208 different solvents can be prepared and dispensed separately allowing greater versatility
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18 209 especially in the case of multi-drug films. By adjusting the film dimensions the printed drug
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20 210 amounts can be easily tuned and adjusted to individual patient needs [12]. Flexible
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22 211 manufacturing processes with adjustable dose strengths like printed films are considered
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24 212 particularly innovative and interesting for applications in personalized medicine or early drug
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26 213 development [13].
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33 214 Surface morphology and solid-state characterisation

34 215 SEM images reveal the multi-layered structure of the films consisting of three layers (Fig.4a,
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36 216 c). Layer 1 is the bottom layer consisting of HCTZ and CARV embedded in a matrix of EC
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38 217 and copovidone K28, layer 2 is the middle layer consisting of AMI and layer 3 is the top layer
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40 218 that was formed by jetting few drops of CARV in copovidone K28. Despite the differences in
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42 219 the ratio of polymers used for each film formulation, the respective layers of both film
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44 220 formulations exhibit the same morphology. Specifically, both layer 1 and 3 are smooth and
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46 221 transparent indicating that the drugs are incorporated in the polymeric matrix while layer 2
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48 222 consists of fine AMI particles (Fig. 4b). This shows that jet dispensing of drug solutions
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50 223 containing polymers (layer 1 and 3) simulates conventional film casting with drug embedded
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52 224 in the film, while in the absence of polymers (layer 2) it simulates ink-jet printing and the drug
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54 225 is resided on top of the film (Fig. 4d, [14])
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226 XRD is a fast and straightforward method for determining basic information regarding the solid
227 state of a material/ formulation with a limit of crystallinity detection in amorphous drug
228 compositions around 5–10% [15]. The XRD diffractograms of the raw drugs exhibit
229 characteristic peaks indicating their crystallinity while the patterns of both EC and copovidone
230 K28 consist of an amorphous halo due to the amorphous nature of these polymers (Fig. 5a).
231 The XRD diffractograms of the film formulations exhibit an amorphous halo with some
232 characteristic peaks (2 theta: 16.7°, 19.5°, 19.9°, 21.5°, 24.8°, 28.3°) which are attributed to
233 HCTZ (Fig. 5b, [16]). These results indicate that all the components of the film are in the
234 amorphous state apart from HCTZ which is in the crystalline form. The reduced intensity of
235 HCTZ peaks for the film 50/50 EC/copovidone K28 compared to those of the film 90/10
236 EC/copovidone K28, indicates that increased incorporation of copovidone K28 suppressed the
237 crystallization of HCTZ.

238 The DSC thermographs of the raw drugs exhibit sharp endothermic peaks at 271 °C, 118 °C
239 and 293 °C due to the melting of HCTZ, CARV and AMI, respectively (Fig. 6a). Moreover, in
240 the case of AMI, the endothermic event from 120 to 150°C is related to the loss of water from
241 the hydrate form of AMI. In the DSC thermographs of the jet-dispensed films, the broad
242 endothermic peak from 40 to 100 °C can be due to the presence of copovidone K28 while the
243 small endothermic peaks around 230-250 °C can be attributed to the melting of HCTZ (Fig.
244 6b). The melting point of HCTZ in the films has shifted to lower temperatures due to the
245 presence of other compounds and the reduction of the size of the drug crystals which are now
246 embedded in the polymer matrix of the first layer forming a solid dispersion. XRD and DSC
247 are in agreement regarding the solid state of the jet-dispensed films which are partially
248 amorphous as all the components are in the amorphous state apart from HCTZ which retains
249 its crystallinity in the formulation.

250 FT-IR

251 FT-IR spectroscopy was implemented to identify interactions among the drugs and the
252 polymers. The FT-IR spectra of the raw drugs, polymers and the films are illustrated in Fig. 7.

253 To investigate the presence of interactions between copovidone K28 and CARV, films with
254 composition corresponding to layer 3 were prepared by jet dispensing (JD CARV/Copovidone
255 K28). The FT-IR spectrum of CARV raw material showed a well-defined characteristic peak
256 at 3342 cm^{-1} corresponding to the NH stretching vibration of the secondary amine. In this
257 region, copovidone K28 demonstrated a very broad peak due to water adsorption related to the
258 highly hygroscopic nature of the polymer (Figs. 7a, 8a). In the FT-IR spectrum of JD
259 CARV/Copovidone K28, the peaks assigned to NH group and water have been merged and
260 moved towards lower wavenumber indicating hydrogen bonding interaction. Also, the
261 broadness of the peak is attributed due to the amorphous nature of CARV [17]. Moreover, the
262 peaks of copovidone K28 at 1667 and 1661 cm^{-1} which correspond to the carbonyl peaks of
263 aliphatic and aromatic ring have moved towards lower wavenumbers at 1658 and 1652 cm^{-1} ,
264 constituting another indication of strong hydrogen bonding interactions between CARV and
265 copovidone K28 (Fig. 8b).

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267 The FT-IR spectra (region: $3100\text{-}3800\text{ cm}^{-1}$) of the crystalline HCTZ and the jet-dispensed
268 films are illustrated in Fig. 9a. The most important chemical groups of HCTZ molecule are the
269 four amino groups and the S=O groups, since these groups can interact either with other HCTZ
270 molecules or with other compounds by means of hydrogen bonding. Specifically, HCTZ can
271 interact with copovidone K28 via hydrogen bonding of its amino group with the carbonyl group
272 of the polymer. HCTZ has four amino groups; two are coming from the primary sulphonamide,
273 one from the secondary sulphonamide and finally there is one secondary amino group. In the
274 crystalline powder, the peaks which correspond to the NH groups are at, 3392 cm^{-1} , 3359 cm^{-1} ,

275 3262cm⁻¹, 3166 cm⁻¹ (Fig. 9a). The assignment of the NH peaks has been previously reported
276 by Tajber *et al.* [18]. Specifically, 3392cm⁻¹ corresponds to N-H secondary amines, 3362 cm⁻¹
277 3166 cm⁻¹ at asymmetric and symmetric stretch vibration of primary sulphonamide and
278 3262cm⁻¹ is the N–H stretch of the secondary sulphonamide. The symmetric stretching
279 vibration of SO groups appears at 1149cm⁻¹ and 1163cm⁻¹, whereas the antisymmetric
280 stretching vibration of SO group is detected at 1347cm⁻¹ and 1333cm⁻¹.

281 The potential hydrogen donors of HCTZ are the two amino groups of the primary
282 sulphonamides, one from the secondary sulphonamide and one from a secondary amino group.
283 In general, the best hydrogen bond donor will bond to the best hydrogen bond acceptor.
284 Moreover, hydrogen bond donating abilities correlate with the acidities, as the more highly
285 acidic group is a better donor among the similar functional groups. In the case of HCTZ,
286 according to Adsmund *et al.* [19] the amido group is more acidic than the proton in the amino
287 group, due to the strong electronegative character of the SO group which is able to polarize the
288 nitrogen atom positively which in turn facilitates the release of the proton. Consequently,
289 HCTZ will interact with the copovidone K28 via the amido group. However, the spectra did
290 not confirm any hydrogen bonding between the carbonyl group of copovidone K28 and the
291 amino group of HCTZ as the amino groups of HCTZ in the jet-dispensed films remain at the
292 same position with the pure material (Fig. 9a). However, the carbonyl peaks which appeared
293 in Fig. 9b should correspond to copovidone K28 which has moved towards lower wavenumbers
294 at 1651 and 1657 cm⁻¹, indicating that copovidone K28 participates in hydrogen bonding
295 interactions probably with CARV as it was mentioned previously. This is probably due to the
296 fact that the amino groups of CARV are more electronegative than those of HCTZ.

297 **Confocal Raman mapping**

298 The spectra of the pure compounds in the 1000 – 1650 cm⁻¹ region are illustrated in Fig.10.
299 Carvedilol has three characteristic peaks at 1577, 1593 and at 1633 cm⁻¹ and one at 1286 cm⁻¹

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300 corresponding to CC stretching vibration and at CN sym. stretch vibration. A detailed
301 description of the peaks has been reported by Marques *et al.* [20]. In HCTZ the strong peaks
302 exist at 1295 cm⁻¹ and at 1313 cm⁻¹ while the strong peak at 1596 cm⁻¹ is assigned to the NH₂
303 ending vibration. In terms of AMI, the two peaks at 1651 and at 1673 cm⁻¹ can be attributed to
304 C=N and C=O vibrations.

305 In order to investigate the distribution of the drugs on the formulation the results from confocal
306 Raman mapping technique were analysed using principal component analysis (PCA). Based
307 on the eigenvalues, the hyperspectral data can be described by two principal components (PCs,
308 Fig. 11a). This is a typical observation in multivariate data analysis of Raman spectra, which
309 leads arbitrarily to the assumption that there are only two components in the system, as the first
310 PC corresponds to the features of the strongest spectral contributors. It can be clearly observed
311 that the three first loadings contain characteristics peak pattern of HCTZ. Hence, for instance
312 the strong peaks at 1149cm⁻¹ and 1163cm⁻¹ correspond to the symmetric stretching vibration
313 of SO groups. It is interesting also to see in PC3 the latter peak has moved towards higher
314 wavenumbers at 1174cm⁻¹ from 1163cm⁻¹, probably due to interactions or due to different
315 chemical environment of these HCTZ molecules. As in the case of HCTZ, characteristics peaks
316 of CARV appear to all the PCs at 1287 cm⁻¹ and at 1633cm⁻¹, indicating that both drugs are
317 distributed across the surface (Fig.11b).

318 Also, the distribution of CARV in the 3rd layer was attempted. However, the PCA in the 3rd
319 layer showed that the predominant peaks at 1288 cm⁻¹, 1388 cm⁻¹, 1510 cm⁻¹ of the first PCs
320 correspond to AMI. This can be explained by the fact that the solution of CARV/copovidone
321 was completely absorbed by the 2nd layer.

322 **Drug content and *in-vitro* dissolution studies**

323 Drug content analysis of the films produced by jet-dispensing was close to the nominal values
324 (6% relative standard deviation or less). This can be linked to the accurate dot deposition
325 achieved by the jet dispenser which subsequently eliminates drug loss or dose variation.
326 Moreover, jetting of solutions rather than suspensions eliminates the risk of sedimentation prior
327 to jetting that can adversely impact drug content in each film.

328 The dissolution profiles of the three drugs from the two types of jet-dispensed films are
329 illustrated in Fig. 12. For both formulations studied, complete and immediate release of AMI
330 from the films is achieved as it expected, since it was jetted alone without any polymer in the
331 second layer of the films. This shows that second layer manages to act as an immediate release
332 layer following the concept of the formulation design. The release of HCTZ from the films is
333 not complete and is delayed with the increase of EC in the polymer matrix. Unlike EC, the
334 increase of copovidone K28 improved the release profile of all the drugs and this can be
335 attributed to the function of this polymer as a pore former facilitating drug release from the EC
336 matrix [21,22]. Specifically for HCTZ, when 10% of copovidone K28 was used 20% of HCTZ
337 was released, whereas in the case of 50% of copovidone, HCTZ release increased to 60%. In
338 the case of CARV, in both films there is an initial burst release where 20% of drug is released,
339 attributing to the amount of drug that was deposited on the third layer and was combined with
340 the hydrophilic copovidone K28 whereas the rest of the drug was remained trapped in the EC
341 matrix of the first layer. The entrapment of HCTZ and CARV in the EC matrix can be attributed
342 to the hydration of the EC and subsequent formation of a gel layer which prevents the
343 incorporated drug molecules to migrate to the dissolution medium.

344 **4. Conclusions**

1 345 Multi-layered films containing three antihypertensive drugs were prepared by jet dispensing.
2 346 The films were characterized regarding their morphology and solid-state properties while FTIR
3 spectroscopy and confocal Raman mapping provided information on the interactions between
4 347 drug and polymers and the drug distribution in the layers of the films, respectively.
5 348 Combinations of EC and copovidone K28 provide suitable polymeric film substrates with the
6 ability to modify drug release. Particularly, decreased EC to copovidone K28 weight ratio was
7 349 found to suppress the crystallization of hydrochlorothiazide and to increase the release rate of
8 the dispensed drugs. While future studies should focus on optimising drug release from each
9 350 layer, this preliminary study highlights the great potential of jet dispensing as a method capable
10 351 to prepare films containing drug combinations and various modes of release (e.g. immediate
11 352 and sustained drug release). Multi-layered films prepared by jet dispensing can be used as
12 353 flexible formulations allowing co-delivery of drug combinations and dose personalization.
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30 357 **Acknowledgements**

31 358 The authors are grateful to Ms Devyani Amin (University of Greenwich) for her help with the
32 359 development of the HPLC method.
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37 360 **Conflict of interest disclosure**

38 361 The authors confirm that this article has no conflict of interest.
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Figures

Fig. 1 Chemical structures of drugs and polymers used for the preparation of the multi-layered films by jet dispensing.

Fig. 2 Schematic illustration of (a) the formulation's design and (b) a jet nozzle.

Fig. 3 Image and dimensions of a 50/50 EC/PVP multi-layered film prepared by jet dispensing.

Fig. 4 SEM images of multi-layered films produced by jet dispensing.

Fig. 5 XRD patterns of (a) raw drugs and polymers and (b) films prepared by jet dispensing.

Fig. 6 DSC thermographs of (a) raw drugs and polymers and (b) films prepared by jet dispensing.

Fig. 7 FT-IR spectra of (a) raw drugs and (b) polymers and films prepared by jet dispensing.

Fig. 8 FT-IR spectra of carvedilol, copovidone K28 and jet-dispensed film of carvedilol-copovidone K28 in the region of (a) 3100-3800 cm^{-1} and (b) 1600-1800 cm^{-1} .

Fig. 9 FT-IR spectra of hydrochlorothiazide, copovidone K28 and jet-dispensed films in the region of (a) 3100-3800 cm^{-1} and (b) 1600-1800 cm^{-1} .

Fig. 10 Raman spectra of the raw drugs in the 1000-1650 cm^{-1} region.

Fig. 11 (a) Principal components (PCs) versus eigenvalues and (b) principal component analysis (PCA) of confocal Raman spectra for the JD 50/50 EC/copovidone K28 film formulation (red and brown arrows correspond to peaks of HCTZ and CARV, respectively).

Fig. 12 In-vitro dissolution profiles of the three drugs from multi-layered jet-dispensed films with different EC/copovidone K28.

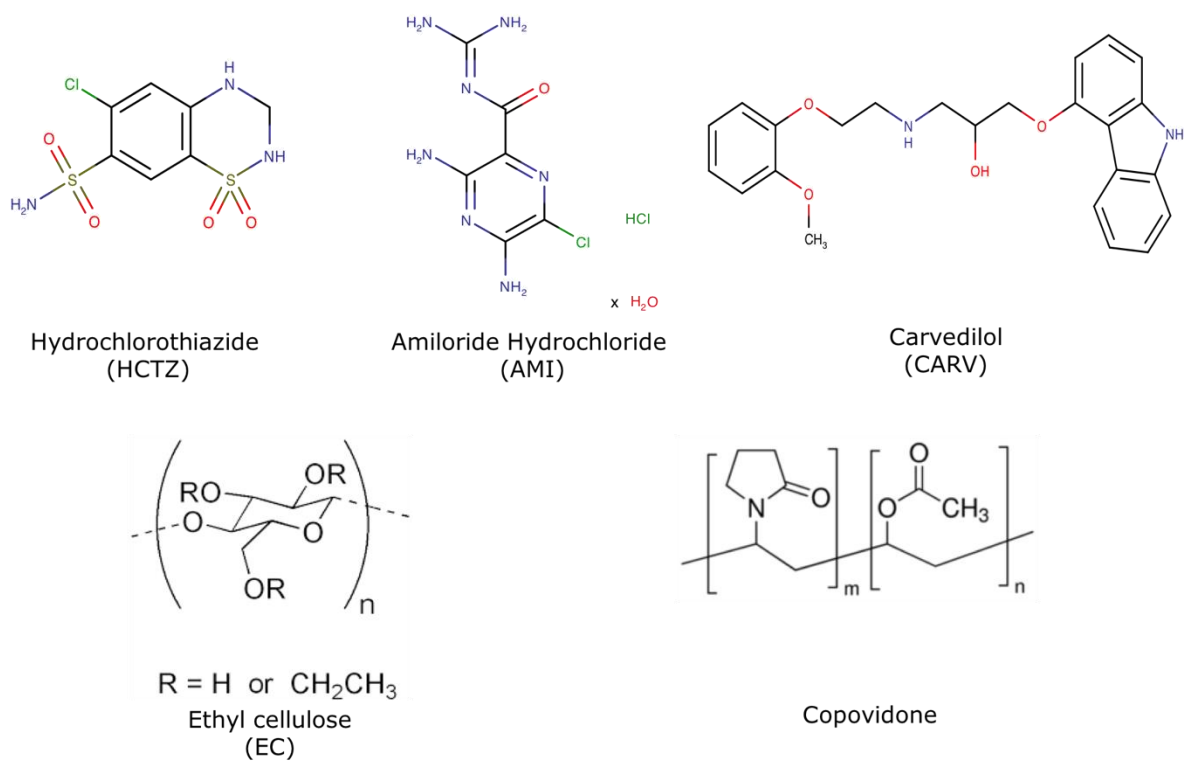


Fig. 1 Chemical structures of drugs and polymers used for the preparation of the multi-layered films by jet dispensing.

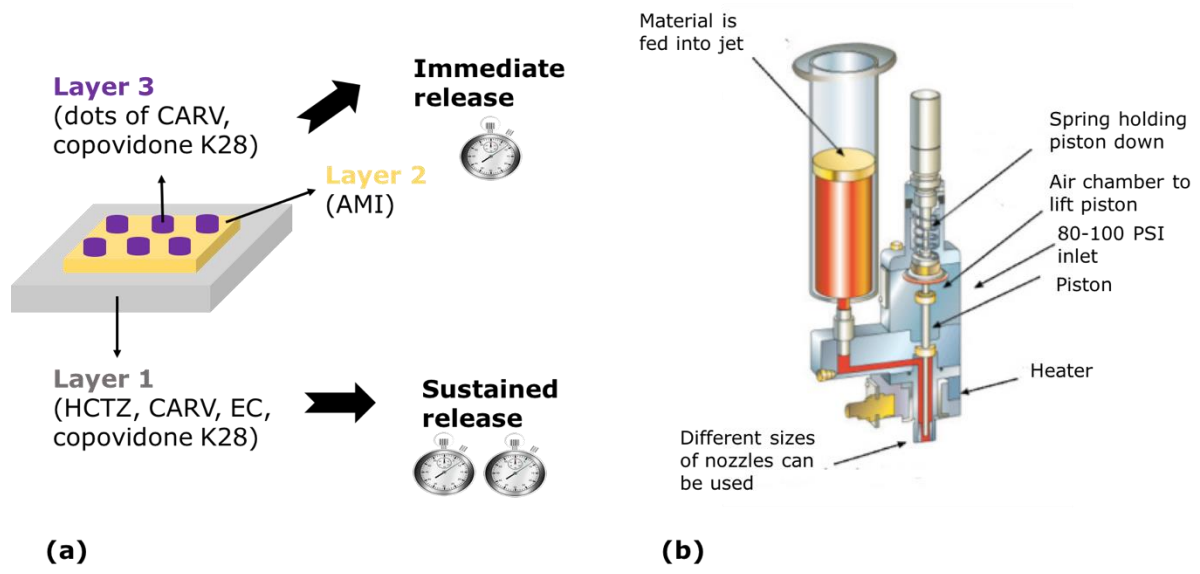


Fig. 2 Schematic illustration of (a) the formulation's design and (b) a jet nozzle.

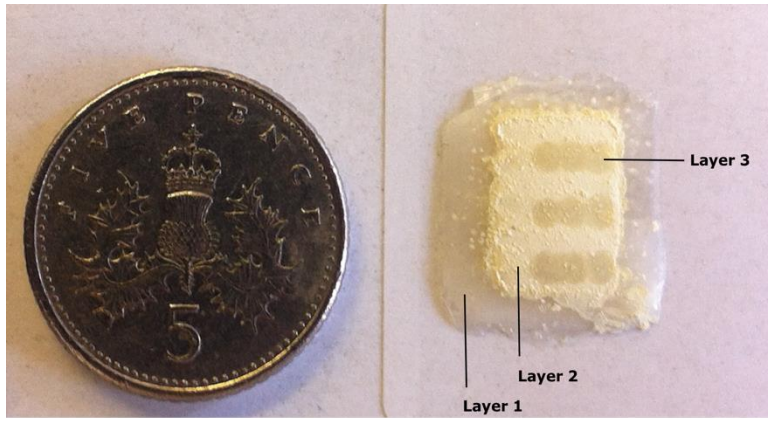


Fig. 3 Image and dimensions of a 50/50 EC/PVP multi-layered film prepared by jet dispensing.

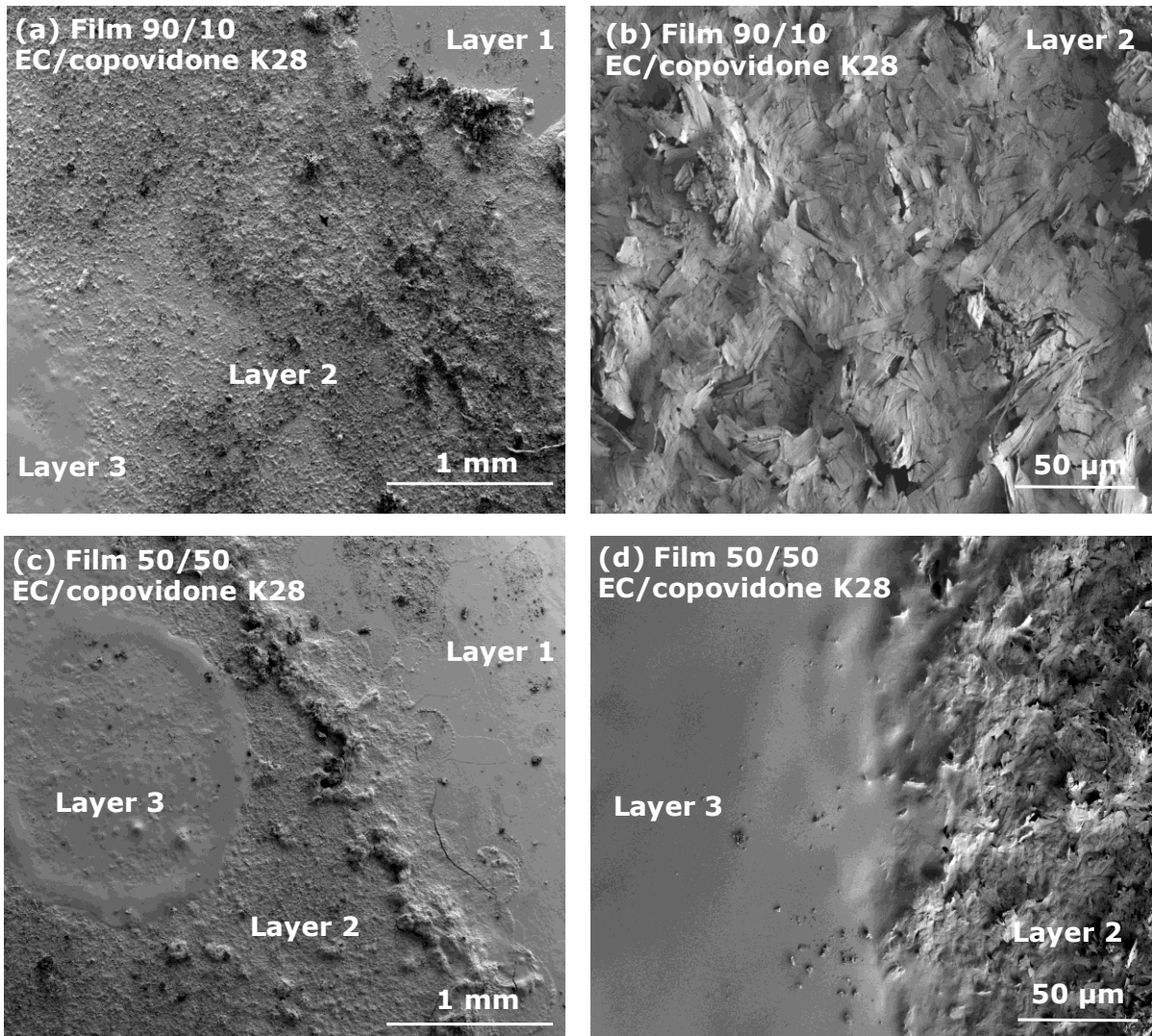


Fig. 4 SEM images of multi-layered films produced by jet dispensing.

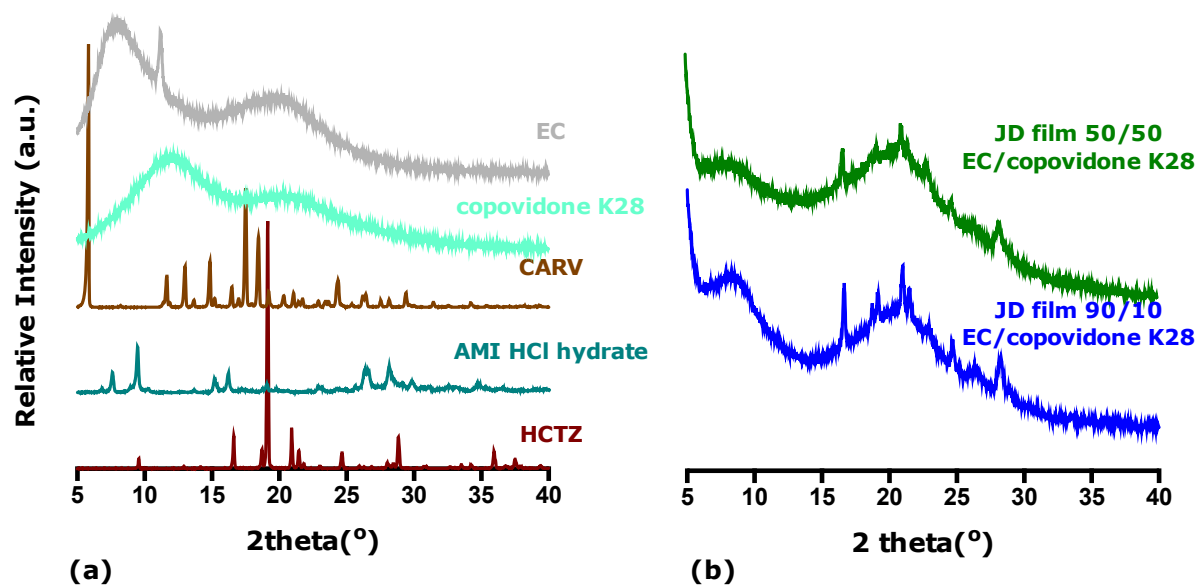


Fig. 5 XRD patterns of (a) raw drugs and polymers and (b) films prepared by jet dispensing.

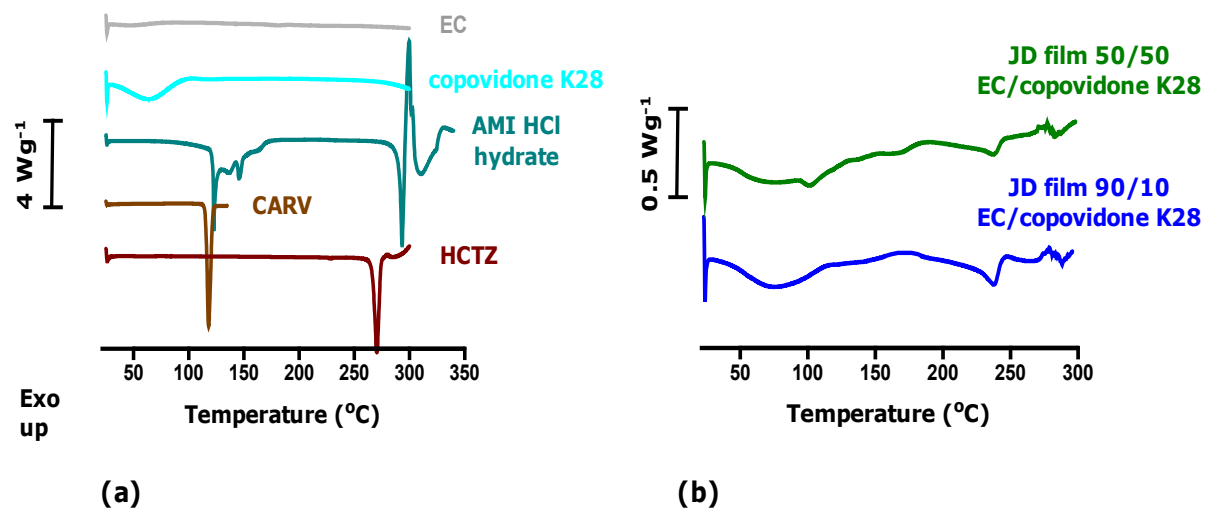


Fig. 6 DSC thermographs of (a) raw drugs and polymers and (b) films prepared by jet dispensing.

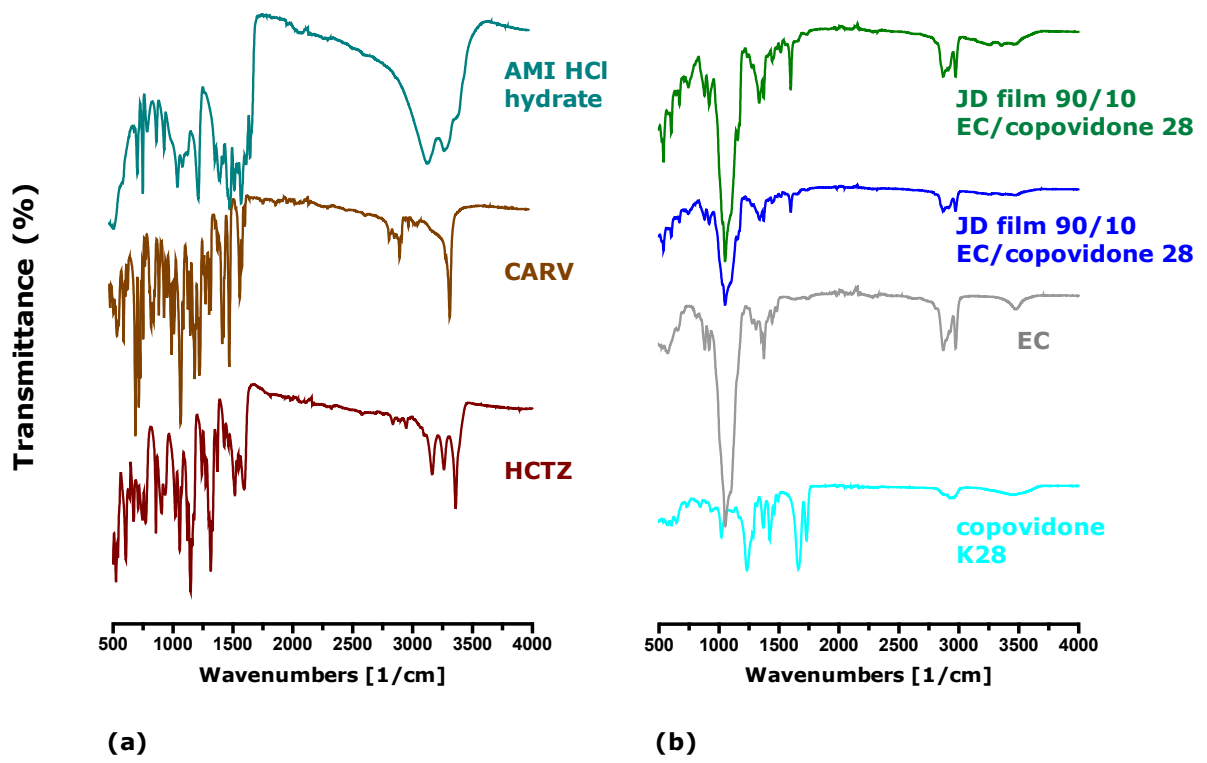


Fig. 7 FT-IR spectra of (a) raw drugs and (b) polymers and films prepared by jet dispensing.

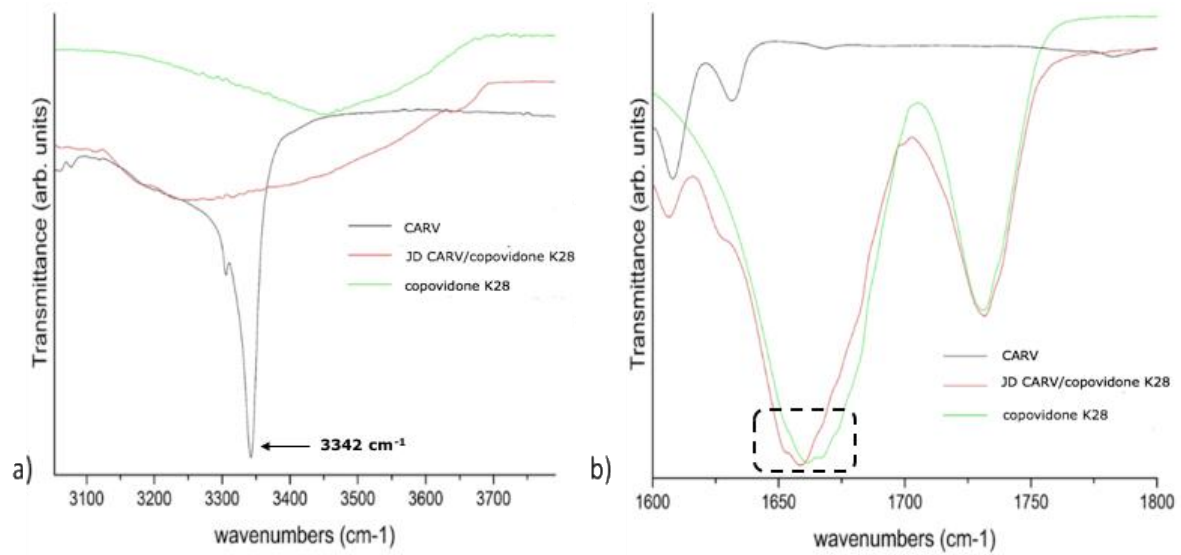


Fig. 8 FT-IR spectra of carvedilol, copovidone K28 and jet-dispersed film of carvedilol-copovidone K28 in the region of (a) 3100-3800 cm⁻¹ and (b) 1600-1800 cm⁻¹.

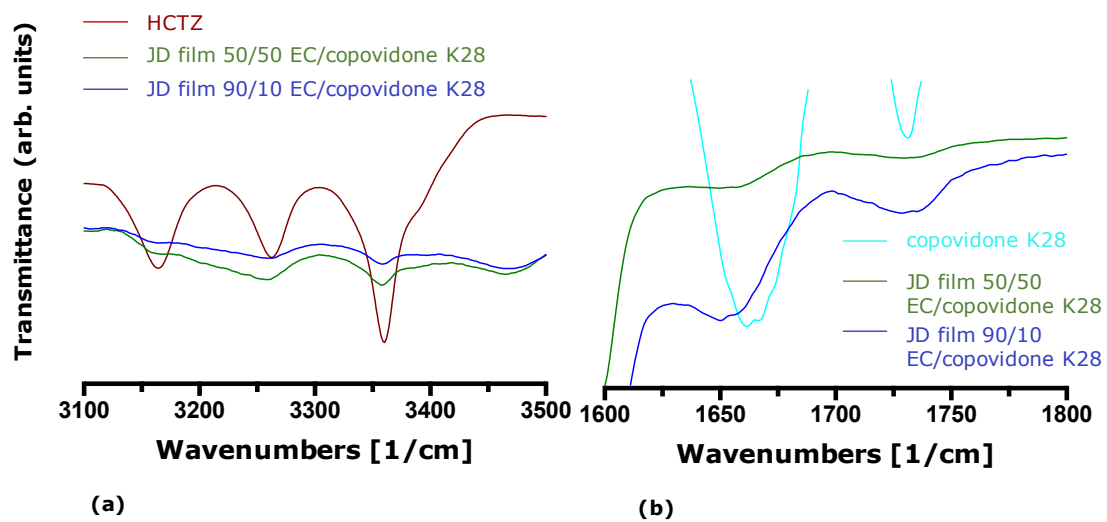


Fig. 9 FT-IR spectra of hydrochlorothiazide, copovidone K28 and jet-dispersed films in the region of (a) 3100-3800 cm^{-1} and (b) 1600-1800 cm^{-1} .

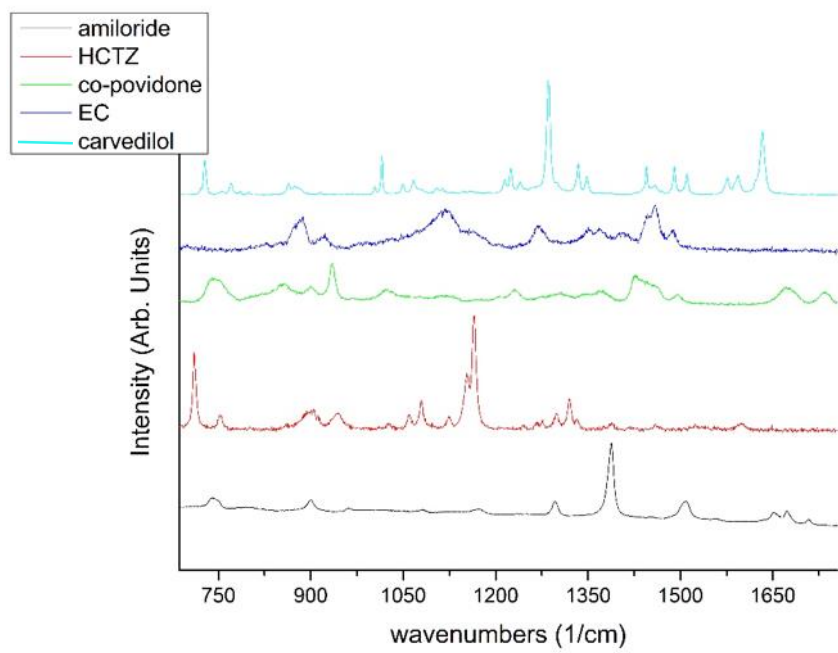
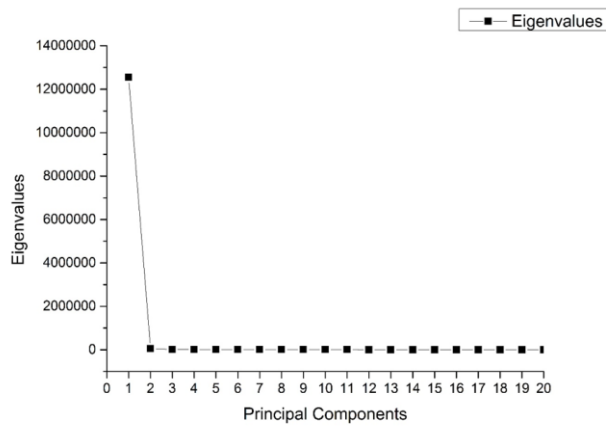
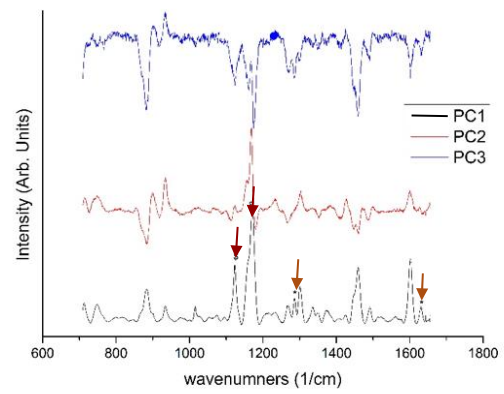


Fig. 10 Raman spectra of the raw drugs in the 1000-1650 cm^{-1} region.



(a)



(b)

Fig. 11 (a) Principal components (PCs) versus eigenvalues and (b) principal component analysis (PCA) of confocal Raman spectra for the JD 50/50 EC/copovidone K28 film formulation (red and brown arrows correspond to peaks of HCTZ and CARV, respectively).

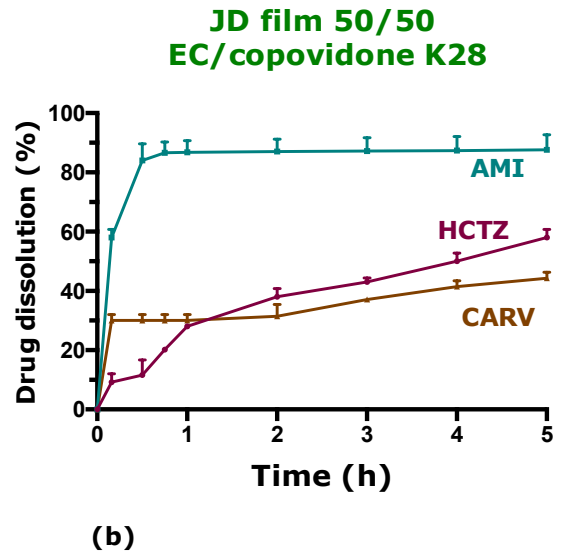
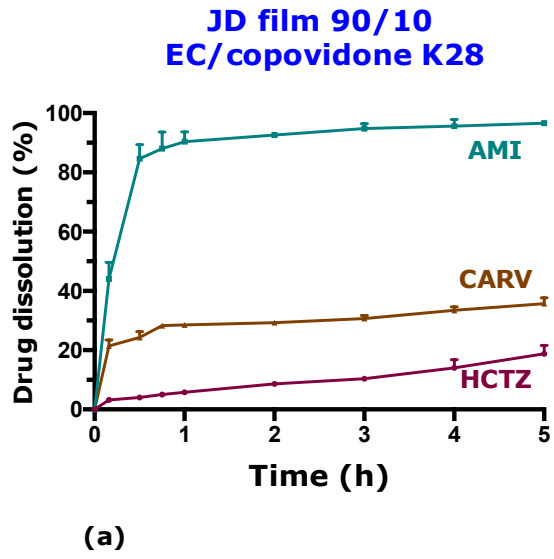


Fig. 12 *In-vitro* dissolution profiles of the three drugs from multi-layered jet-dispensed films with different EC/copovidone K28.

Table**Table 1.** Nominal composition of each layer for the jet-dispensed film formulations and volume of the precursor solutions printed per film.

	Film	HCTZ (mg)	CARV (mg)	AMI (mg)	EC (mg)	Copovidone K28 (mg)	Volume printed per film (ml)
Layer 1	90/10	25	2.5	-	49.5	5.5	3.57
	50/50	25	2.5	-	27.5	27.5	3.57
Layer 2		-	-	2.5	-	-	0.25
Layer 3		-	1	-	-	2	0.033