

Polyphenols and their role in obesity management: A systematic review of randomized clinical trials

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List of abbreviations: *AMPK: AMP-activated protein kinase; CVD: Cardiovascular diseases; ECP: Ecklonia cava polyphenols; HSE: Hibiscus sabdariffa extract; Licorice flavonoid oil; RCTs: Randomized clinical trials; WC: Waist circumference*

Abstract

Polyphenols have been suggested to reduce body weight and modify body composition through different mechanisms. These effects have been extensively studied in animals and in vitro and to a lesser extent in humans. The aim of this review is to consider the association between polyphenols and body weight status by focusing on human intervention studies. We conducted a systematic literature search in MEDLINE (via EBSCOhost), ProQuest CENTRAL and Cochrane CENTRAL without time restrictions. Randomized controlled trials (RCTs) assessing the effect of polyphenols on weight and/or body composition in the overweight and/or obese population were included. Nineteen studies met our inclusion criteria. Results suggest that further research is required before supporting a potential role of polyphenols in reducing weight in overweight and obese individuals (9 studies showed a significant decrease in weight by a mean of 1.47 ± 0.58 Kg). Nevertheless, several studies indicated that polyphenols might be effective in preventing small increases in weight during periods of overfeeding rather than reducing weight as such. The outcomes noted do not yet support polyphenol supplementation as a complementary approach in weight-loss diets. Further larger trials with duration of 12 months or more are needed to elucidate the effect of polyphenols on body weight status.

Keywords: Polyphenols; flavonoids; catechins; obesity; Body weight; BMI; waist circumference; body fat

Introduction

Polyphenols are vastly diverse phytochemicals with complex chemical structures. They are found in a variety of commonly consumed foods such as chocolate, tea, coffee, wine, olive, dry legumes, and some vegetables (like lettuce and cabbage) and fruits (like apples and berries) (Manach et al., 2004; D'archivio et al., 2007). Around 8000 structures of polyphenols have been already identified (Martin et al., 2010). Polyphenols are classified into four groups: flavonoids, lignans, stilbenes and phenolic acids (Manach et al., 2004). **Figure 1** summarizes the most common polyphenols and their subtypes.

Polyphenols have been extensively studied over the past decade because of their potential antioxidant and anti-inflammatory roles and their possible role in the prevention and management of several diseases such as cardiovascular diseases (CVD), hypertension, diabetes, cancer, and neurodegenerative diseases (Manach et al., 2004; D'archivio et al., 2007; Michalska et al., 2010; Serban et al., 2015). Most recently, polyphenols have attracted media interest and research community because of their potential role in reducing obesity, an increasingly serious health issue in different population age ranges (Zaki et al., 2015; WHO 2016). Polyphenols such as catechins, anthocyanins, curcumin and resveratrol were suggested to exert beneficial effects on lipid and energy metabolism (Meydani et al., 2010; Min et al., 2013; Gu et al., 2014; Kunnumakkara et al., 2016; Smeriglio et al., 2016) and

potentially on weight status. Multiple mechanisms of action have been proposed mostly as a result of animal and cell studies, such as inhibition of the differentiation of adipocytes (Min et al., 2013), increased fatty acid oxidation (Dulloo et al., 1999; Shimoda et al., 2009), decreased fatty acid synthesis (Matsui et al., 2005), increased thermogenesis and energy expenditure (Nagao et al., 2007; Osakabe et al., 2013; Stohs and Badmaev, 2016) and inhibition of digestive enzymes (McDougall et al., 2005; Gu et al., 2011). Despite the popularity of this topic, the biggest contribution to this research remains through animal and in vitro studies. Few cohort studies have looked at the association between polyphenol consumption and body weight in humans (Wu et al., 2003; Hughes et al., 2008; Golomb et al., 2012), and human intervention studies are outnumbered by animal and cell studies and remain relatively limited. Results from animal and in vitro studies have mostly indicated an effect of polyphenols on reducing obesity, and have considerably contributed to the obesity-lowering effects of polyphenols discussed in the media. However, many factors could affect the replication of these outcomes to humans. Firstly, there are many differences in the metabolism and mechanism of actions of polyphenols between animals and humans (Natsume et al., 2003). For example, there is lack of clear evidence of the presence of biologically active brown adipose tissue in adults, the latter being demonstrated to be primarily triggered by cold exposure in humans (Chen et al., 2013). This has explained a part of the weight lowering effect of cocoa and tea polyphenols in animals (Matsui et al., 2005; Nomura et al., 2008). Secondly, there is a difference in body weight control between animals and free living humans. The latter exhibit large differences in daily levels of physical activity and energy balance (Leibel, 2008). Thirdly, the complexity of substituting in vitro studies for humans studies relies in the limitations in assessing polyphenol bioavailability in cell

studies (Etcheverry et al., 2012). Also, the doses of polyphenols that exert a significant effect on obesity in cells (such as the inhibition of adipogenesis (Min et al., 2013; Ejaz et al., 2009)) can be higher than the physiological levels. In view of these potential differences, and for the aim of helping the public make informed choices and directing future research, we attempt to consider the association between polyphenols and body weight status through conducting a systematic review of human intervention studies.

Methods

The review preparation was completed in 7 steps: Identification of research question, definition of inclusion and exclusion criteria, literature search, selection of eligible studies, data extraction, evaluation of the risk of bias (through *the Cochrane RoB Tool*) and presentation of results (Cornell 2016). The review was prepared according to the Prisma guidelines (Moher et al., 2009).

Search strategy

We (GF, EAD) independently conducted a systematic literature search in MEDLINE (via EBSCOhost), ProQuest CENTRAL and Cochrane CENTRAL (from 11 November 2016 to 10 December 2016) without any time restrictions. We used the following keywords: Polyphenol AND (Body weight OR Waist circumference OR Fat). We also searched the reference list of selected studies for more relevant research. Disagreements in the assessment of data were resolved by discussion and consensus was reached in all cases.

Eligibility criteria

Studies included were human RCTs investigating as a main outcome the link between polyphenols and obesity (through measurement of body weight and/or waist circumference (WC), waist/hip ratio, or body fat at least twice during the study).

There were no restrictions on the type of studies (parallel or crossover studies).

Eligible studies included overweight and/or obese adults with no history of known diseases (diabetes, CVD or hypertension) and no intake of medications that could influence lipid, carbohydrate or energy metabolism. Studies involving exercise regimens and/or hypocaloric diets in addition to the polyphenol intervention were excluded. In addition, eligibility of studies was restricted to duration no less than 4 weeks and to English language. To avoid confounding effects, studies investigating the combined effects of polyphenols and other components (such as tyrosine, fibre etc...) on anthropometric measures were excluded. Missing data and further information were requested from the authors.

Study selection and data extraction

References were checked for eligibility by two independent authors. Full texts were then verified against the inclusion criteria. Risk of bias was assessed and data about study design, participant information (BMI, age, gender) and outcome measures (Body weight, BMI, WC, waist-to-hip ratio and/or body fat percentage or mass) were extracted. Baseline and post intervention values and mean difference from baseline data were collected.

Results

The search identified 19 eligible studies. The study selection process is summarised in **Figure 2**.

Risk of bias

The allocation concealment was considered to be suitable in 2 studies and unclear in 17. Eighteen studies reported blinding participants and outcome assessors. Three studies described measures used to blind participants and outcome assessors. Data analysis of the main outcomes was reported by 15 studies. Incomplete outcome data was considered to be adequately handled in 10 studies. The two authors (GF, EAD) have reached a consensus on not to conduct a meta-analysis in this review for the following reasons 1) heterogeneity of the studies and the different types of polyphenols administered 2) missing outcome data that we failed to obtain from authors 3) missing information regarding allocation concealment and handling outcome data which makes it difficult to evaluate the risk of bias in studies. This might likely produce inappropriate summary and biased conclusions. Therefore, this review mainly aimed to clarify the effects of polyphenols on anthropometric measures and to provide directions for future research.

Study characteristics

Studies were conducted in Japan (N=5), USA (N=4), Germany (N=2), Spain (N=1), Taiwan (N=2), France (N=1), China (N=1), Denmark (N=1), Poland (N=1) and UK

(N=1). Studies investigated the effect of green tea (N=6), oolong tea (N=1), Licorice flavonoid oil (LFO) (N=3), apple juice (N=2), citrus extract (N=1), Hibiscus sabderiffa extract (N=1), grape juice (N=1), quercetin (N=1), Ecklonia cava (N=1), resveratrol (N=1) and orange juice (N=1). Samples sizes ranged from 22 to 240 participants. Sixteen studies were parallel, while 3 were crossover. All studies assessed body weight, 14 assessed waist circumference and 12 assessed body fat (percentage or mass). Four studies used a polyphenol-rich drink that provided energy, while 15 studies used capsules or drinks that provided a negligible amount of calories. Four studies assessed the effect of different doses of polyphenols on anthropometric measures. Ten studies disclosed funding from industry, and 3 studies gave no indication of funding. **Table 1** provides an overall summary of study details.

Weight loss and BMI

Out of the 19 selected studies, 10 reported a significant decrease in body weight from baseline in the experimental group (**Table 1**). While one of the studies noted a significant decrease in weight only during the first intervention period of the crossover study (Brown et al., 2011), the 9 remaining studies (8 studies lasted for 12 weeks and one study lasted for 8 weeks) showed a significant decrease in body weight by a mean of 1.47 ± 0.58 Kg. Three of these studies used a phenolic compound that provided energy (between 85-350 Kcal) (Hollis et al., 2009; Shin et al., 2012; Rangel-Huerta et al., 2015). Seventeen studies assessed BMI, of which 7 demonstrated a decrease by a mean of 0.65 ± 0.55 Kg/m² from baseline in the experimental group. On the other hand, three studies reported a significant increase in weight only in the placebo (control) group (mean increase: 1.04 ± 0.54 Kg).

Changes in waist circumference

Fourteen studies assessed the effect of polyphenols on WC. Nine studies found a significant lowering effect on WC (mean decrease: 2.58 ± 1.33 cm). Eight studies lasted for 12 weeks while one study lasted for 8 weeks (noted a significant decrease in WC by 0.63 cm) (**Table 1**).

Changes in body fat percentage and body composition

Twelve studies assessed fat composition as either body fat percentage (N=10) or body fat mass (N=2). While 4 studies showed no significant effects on body fat percentage, 6 studies showed a decrease in fat percentage by a mean of $1.86 \pm 0.98\%$. One study reported a significant decrease in fat mass (by 1.03 ± 0.24 Kg depending on the polyphenol dose). Three studies assessed fat visceral area and noted a significant decrease by 9.18 ± 1.2 cm (**Table 1**).

Discussion

Main findings

The aim of this review was to assess existing human research on polyphenols and body weight status. Eligible studies involved the incorporation of polyphenols (in different forms) in the context of a normocaloric diet and not as a part of a reduced-calorie diet or in addition to exercise. This eliminated the effect of weight loss

strategies which would mask the effect of polyphenols on weight and body composition. Demonstrating a potential effect of polyphenols on obesity would be helpful towards integrating polyphenols in weight loss regimens or coupled to exercise.

The similarities in the mechanisms of action of polyphenols on obesity provided a justification for including different types of polyphenols in the same review. For example, phlorotannins, quercetin and phenolic acids have been shown to be involved in the inhibition of adipocyte differentiation (Hsu et al., 2007; Jung et al., 2014; Seo et al., 2015). Anthocyanins, quercetin and phenolic acids have been reported to increase AMPK (adenosine monophosphate-activated protein kinase), which is involved in fatty acid oxidation (Pfeuffer et al., 2013; Chang et al., 2014). Some polyphenols have been implicated in increasing satiety, reducing energy intake (Panickar et al., 2013) and increasing thermogenesis (Meydani et al., 2010; Stohs and Badmaev, 2016). The latter mechanisms are involved in body weight control. The limited understanding of the mechanisms of actions requires further research.

The results of the selected studies do not mostly favour a weight-lowering effect of polyphenols in overweight and obese individuals. The weight loss reported by 9 out of 19 selected studies ranged between 1.47 ± 0.58 Kg. The duration of the studies (4-12 weeks) might provide an explanation for the results obtained. It has been indicated that larger longer-term trials with duration of 12 months or more are needed to understand the effect of an intervention on weight loss and weight management (Headland et al., 2016). The nine studies showed a small body weight

reduction, less than previously considered clinically relevant weight loss at 5% (Stevens et al., 2006). However, one study showed that a loss of 1 Kg of weight could potentially reduce diabetes risk by 16% (Hamman et al., 2006). Therefore, the outcomes may not be insignificant in relation to many health outcomes. It is worth noting that all these studies were double-blinded which eliminated the expectation bias. Nevertheless, there was no sufficient detail on allocation concealment in most of the studies.

A clinically relevant decrease in WC constitutes more than 3 % over the long term (Verweij et al., 2013). The mean decrease in WC in selected studies was 2.58 ± 1.33 cm. As all selected studies included overweight and obese participants, and assuming that they have an elevated WC (at least 80 cm), changes reported might not be clinically relevant in all studies.

Among studies that analysed both body weight and body fat composition, 7 studies showed a significant decrease in both body weight and body fat, while two studies reported a significant decrease only in body weight and one study reported a significant decrease in body fat percentage (by 1% , $p < 0.05$), but not body weight. Further studies are needed to explain the association between body weight and body fat in relation to polyphenol consumption, particularly because the majority of mechanisms by which polyphenols are suggested to reduce obesity is through their effects on lipid metabolism, such as inhibition of fatty acid oxidation (Dulloo et al., 1999; Shimoda et al., 2009), and decrease of fatty acid synthesis (Matsui et al., 2005).

This review suggests that further research is needed before considering polyphenols a complementary approach to aid in weight loss. This hypothesis might be helpful in avoiding misleading advertisement currently describing polyphenols as anti-obesity

agents. Nevertheless, the effects of polyphenols on cardiovascular risk seem to be more established (Dudzińska et al., 2015). Therefore, it could be suggested that polyphenols can lower CVD risk by targeting components of the metabolic syndrome such as blood pressure and HDL cholesterol (Serban et al., 2015; Santini and Novellino, 2016; Ganjali et al., 2017; Patti et al., 2017), while their implication in weight loss is less plausible. Meanwhile, a diet rich in polyphenols can be recommended as having protective effects on risk factors for CVD but their effects on weight loss in humans require further research.

Among polyphenols, flavonoids and phenolic acids appear to be the most involved in obesity and weight control. Subtypes of flavanoids that have been implicated in weight loss or change in body composition in the selected studies are catechins, phlorotannins, glabridin, anthocyanins, procyanidins and quercetin. Although the study by Rangel and Huerta (2015) showed a significant decrease in weight following the consumption of orange juice (at two different concentrations of flavanones), it is suggested that flavanone is not involved in weight loss. The decrease in weight was explained by the decrease in energy intake during the study (Rangel-Huerta et al., 2015). However, the latter study lacked a control group which could have provided more clarification on the effects of flavanones on body weight. Further studies investigating the effect of flavanones on body weight status are needed.

Other polyphenols such as curcumin, flavanols and ellagic acids have also been involved in attenuating obesity (Meydani et al., 2010) and are known to exist in several commonly consumed foods such as turmeric, grapes, nuts, pomegranate, berries, cocoa and chocolate (Daniel et al., 1989; Meydani et al., 2010; Akaberi and Hosseinzadeh, 2016; Kunnumakara et al., 2016). As far as we know, limited or no

human studies primarily investigating the potential implication of the latter polyphenols on obesity and body weight. Cocoa and dark chocolate have been the subject of extensive animal and in vitro studies due to their high flavanol content (epicatechin, procyanidins and quercetin) (Hurst et al., 2008). Although a human study demonstrated that 7 days of cocoa supplementation with 2000 mg of polyphenols reduced WC (by 1.24 ± 1.45 cm, $p \leq 0.05$) without affecting weight (Di Renzo et al., 2013), this significance remains under question due to the short study duration and the absence of a control group. Hence, this study was not included in the systematic review.

Catechins in green tea have been the most popular polyphenols studied. Research is mostly in favour of their potential effects on lowering weight and adiposity (Nagao et al., 2005; Nagao et al., 2007; Wang et al., 2010; Hsu et al., 2008). Nonetheless, a meta-analysis suggested that the effects of green tea catechins on body weight are more plausible in the Asian population due to a genetic polymorphism that leads to differences in the thermogenic effect of green tea (Hursel et al., 2009). This explains the consistent results obtained in the Asian populations (Nagao et al., 2005; Nagao et al., 2007; Wang et al., 2010; Hsu et al., 2008), while the effects are less consistent in western studies (Basu et al., 2010; Brown et al., 2011; Sulliburska et al., 2012). Therefore, it is important to find out whether findings about tea and polyphenols can be applicable to different regions. However, It seems that when the amount of caffeine is controlled, the treatment effect of green tea catechins on obesity was no longer apparent (Basu et al., 2010; Brown et al., 2011). This has been reinforced by a Dutch study which showed that the effect of green tea was not significant in participants with a habitual high caffeine intake (Westerterp-Plantenga et al., 2005).

The potential effect of caffeine or a synergistic effect between polyphenols and caffeine deserve further investigation.

Nonetheless, there is evidence to suggest that the role of polyphenols might be limited to the prevention of weight gain rather than reducing weight per se. This is in line with the majority of animal studies which demonstrated that polyphenols reduce weight gain caused by diet-induced obesity. For instance, extracts of epicatechin and quercetin and polyphenols in green tea, cocoa, resveratrol, pomegranate and curcumin have prevented an increase in weight in animals administered a hypercaloric diet, compared to those given a placebo control (no polyphenols) (Lei et al., 2007; Ahn et al., 2008; Shao et al., 2012; Gulvady et al., 2013; Tian et al., 2013; Dorenkott et al., 2014). This hypothesis provides an explanation for the results obtained in the study by Hollis et al. (2009) who showed a significant increase in weight only in the group administered grape juice with a negligible amount of polyphenols (by 1.6 ± 0.3 Kg, $p > 0.05$). Participants who consumed grape juice with polyphenols (934 mg) did not display a significant increase in weight (mean difference from baseline: 0.8 ± 0.6 Kg, $p < 0.05$). In their study on licorice flavonoid oil (LFO), Bell et al. (2011) noted a significant increase in weight only in the control group and suggested that LFO can be helpful in reducing weight gain in periods of overfeeding. Additionally, a study that did not meet the inclusion criteria for this systematic review showed that one month of pomegranate juice supplementation prevented an increase in weight, BMI and body fat mass in obese participants (mean difference from baseline: -0.5 ± 2.3 Kg, -0.2 ± 0.5 Kg/m², $-1.4 \pm 3\%$, respectively, $p > 0.05$) compared to the group administered the control juice (weight, BMI and body fat mass increased by 1.1 ± 1.3 Kg, 0.4 ± 0.5 Kg/m², $1.1 \pm 1.1\%$, respectively, $p < 0.05$) (González-Ortiz et al., 2011). Moreover, the results noted by Brown et al. (2011) led

to conclude that tea catechins may be involved in preventing weight gain during periods of positive energy imbalance such as increase in weight during seasonal changes, for instance during winter. The current authors have also shown that 400 mg of flavanols in dark chocolate avoided the increase in weight at the end of a 4-week study (Mean difference from baseline: -0.01 ± 0.9 Kg, $p=0.98$) compared to the control group (0.44 ± 0.86 Kg, $p= 0.009$) administered dark chocolate with low amount of flavanols (<60 mg) (Farhat et al., 2015). Similar results were noted by another study which included overweight and obese individuals and suggested that polyphenols can counter the effect of fat and energy contents of the diet (Al Moosawi et al., 2012), although there was no data to support this. In view of the outcomes noted, further investigation is needed before suggesting a significant effect of polyphenols on reducing weight gain in the overweight and obese population. This effect could be of importance as it was shown that an increase of 1 Kg of body weight can rise coronary mortality risk by 1 - 1.5 % (Jousilhati et al., 1996). With the increasing obesity prevalence, mainly due to a positive energy balance (Krzysztozek et al., 2015), polyphenols could then be helpful in preventing small increases in weight such as during winter or periods of overfeeding.

Further larger long-term studies (at least 12 months) controlled for energy intake and physical activity are required to determine the role of polyphenols in weight management. Studies comparing the effect of different polyphenols in periods of overfeeding and in periods of steady weight are needed in the overweight and obese population. In addition, further research investigating the effects of different doses of polyphenols will be helpful to determine the optimal dose of polyphenols for maximum benefit. Also, because of the serious effects of certain herbs and the herb-

drug interactions (Izzo et al., 2016), it is important to consider these side effects while recommending some herbal products because of their high polyphenol content. It would be important to determine whether polyphenols administered in different forms can have different effects. For example, a placebo-controlled study with three arms analyzing the effect of polyphenol-rich juice or food (introduced in the context of an energy-balanced diet) versus polyphenol extract capsules on body weight and composition in the overweight population would be valuable. Based on the results of the selected studies in this review, we can hypothesize that the administration of polyphenols in various forms does not differently affect their weight-lowering properties.

Furthermore, studies with complete data, including information about concealment and data analysis are needed. Breakdown of the amount of polyphenols in both experimental and control group is required. It is also worth noting that polyphenols can be affected by several factors such as the timing of intake, as one of the mechanisms by which polyphenols reduce obesity is through their potential effects on digestive enzymes such as inhibition of protease and lipase (McDougall et al., 2005). Therefore, it is important to test match for these differences in future studies. Lastly, it would be important to determine whether there is a combination effect of different polyphenols or polyphenol-rich foods on body weight status, especially since many of these polyphenols are available combined in nature, and whether an adaptation to the effect of polyphenols occurs over time. This would be helpful to establish conclusions regarding different foods and beverages such as apple juice, green tea as well as polyphenol extracts.

Strength and limitations

One of the review's potential weaknesses is that the comparison of significant effects between studies was based on different statistical comparisons used. In fact, 9 studies considered between-group differences, while 10 studies considered within-group differences. This has possibly affected the generation of hypotheses and conclusions. For instance, in the study by Chang et al. (2014), paired-t-tests showed a significant difference in body weight, BMI and waist-to-hip ratio ($p < 0.05$), while between-group analysis showed significant difference only in WC, waist-to-hip ratio and body fat percentage ($p < 0.05$). In addition, Tominaga et al. (2009) did not analyse whether there is a significant interaction between different doses of polyphenols and time on the effects noted. This is an important point that needs to be emphasized in both future systematic reviews and trials. Another limitation is the duration of the studies (4-12 weeks) which might not have been sufficient to understand the effect of polyphenol on body weight. Furthermore, most of the studies measured the outcomes on limited time points (baseline and week 12 were considered in 8 studies). Five other studies have taken measurements at multiple time points but did not include these data in the analysis. This an important point to consider in future studies. A suggestion would be to analyse the study parameters at 2-week interval in long term studies. Also, diet advice to participants was not properly described in the majority of selected studies. A further limitation is the restriction of the review to the English language, as the studies on polyphenols are popular in the Asian population.

Conclusion

This review of selected studies suggests that further research is needed before supporting the implication of polyphenols in weight loss and recommending polyphenols as a potential complementary approach in weight-loss diets. There is an indication to suggest that the benefits of polyphenols might only be restricted to the counteraction of the small increases in body weight and fat resulting from seasonal changes (such as during winter) or periods of overfeeding. Long-term studies involving a large cohort and controlling for diet and exercise are needed in order to confirm the potential effect of polyphenols on obesity in humans. The genetic differences between populations also need to be considered.

Acknowledgments

GF conceived the review, designed the protocol and wrote the paper. GF, E.A.D performed data extraction. SD provided detailed feedback for the review. All authors reviewed and approved the manuscript.

References

- Ahn J, Lee H, Kim S, Park J, Ha T. 2008. The anti-obesity effect of quercetin is mediated by the AMPK and MAPK signaling pathways. *Biochem Bioph Res C* **373**: 545-9.
- Akaberi M, Hosseinzadeh H. 2016. Grapes (*Vitis vinifera*) as a Potential Candidate for the Therapy of the Metabolic Syndrome. *Phytother Res* **30**: 540–556.
- Akazome Y, Kametani N, Kanda T, Shimasaki H, Kobayashi S. 2010. Evaluation of safety of excessive intake and efficacy of long-term intake of beverages containing apple polyphenols. *J Oleo Sci* **59**: 321-38.
- Almoosawi S, Tsang C, Ostertag LM, Fyfe L, Al-Dujaili EA. 2012. Differential effect of polyphenol-rich dark chocolate on biomarkers of glucose metabolism and cardiovascular risk factors in healthy, overweight and obese subjects: a randomized clinical trial. *Food Funct* **3**: 1035-43.
- Barth SW, Koch TC, Watzl B, Dietrich H, Will F, Bub A. 2012. Moderate effects of apple juice consumption on obesity-related markers in obese men: impact of diet–gene interaction on body fat content. *EJCN* **51**: 841-50.

- Basu A, Sanchez K, Leyva MJ, Wu M, Betts NM, Aston CE, Lyons TJ. 2010. Green tea supplementation affects body weight, lipids, and lipid peroxidation in obese subjects with metabolic syndrome. *J Am Coll Nutr* **29**: 31-40.
- Bell ZW, Canale RE, Bloomer RJ. 2011. A dual investigation of the effect of dietary supplementation with licorice flavonoid oil on anthropometric and biochemical markers of health and adiposity. *Lipids Health Dis* **10**: 1 (Epub ahead of print; doi: 10.1186/1476-511X-10-29.).
- Brown AL, Lane J, Holyoak C, Nicol B, Mayes AE, Dadd T. 2011. Health effects of green tea catechins in overweight and obese men: a randomised controlled cross-over trial. *BJN* **106**: 1880-9.
- Chang HC, Peng CH, Yeh DM, Kao ES, Wang CJ. 2014. Hibiscus sabdariffa extract inhibits obesity and fat accumulation, and improves liver steatosis in humans. *Food Funct* **5**: 734-9.
- Chen YC, Cypess AM, Chen YC, Palmer M, Kolodny G, Kahn CR, Kwong KK. 2013. Measurement of human brown adipose tissue volume and activity using anatomic MR imaging and functional MR imaging. *J Nucl Med* **54**: 1584-7.
- Cornell University library. A Guide to Conducting Systematic Reviews: Steps in a Systematic Review. Version current 18 August 2016. Internet: <http://guides.library.cornell.edu/c.php?g=459012&p=3142201> (accessed 17 December 2016).
- D'Archivio M, Filesì C, Di Benedetto R, Gargiulo R, Giovannini C, Masella R. 2007. Polyphenols, dietary sources and bioavailability. *Annali-Istituto Superiore di Sanita*. **43**: 38-61.
- Dallas C, Gerbi A, Elbez Y, Caillard P, Zamaria N, Cloarec M. 2014. Clinical study to assess the efficacy and safety of a citrus polyphenolic extract of red orange, grapefruit, and orange (Sinetrol-XPur) on weight management and metabolic parameters in healthy overweight individuals. *Phytother Res* **28**:212-8.
- Daniel EM, Krupnick AS, Heur YH, Blinzler JA, Nims RW, Stoner GD. 1989. Extraction, stability, and quantitation of ellagic acid in various fruits and nuts. *Journal of food composition and Analysis* **2**: 338-49.
- Di Renzo L, Rizzo M, Sarlo F, Colica C, Iacopino L, Domino E, Sergi D, De Lorenzo A. 2013. Effects of dark chocolate in a population of Normal Weight Obese women: a pilot study. *Eur Rev Med Pharmacol Sci*. **17**: 2257-66.
- Dorenkott MR, Griffin LE, Goodrich KM, Thompson-Witrick KA, Fundaro G, Ye L, Stevens JR, Ali M, O'Keefe SF, Hulver MW, Neilson AP. 2014. Oligomeric cocoa procyanidins possess enhanced bioactivity compared to monomeric and polymeric cocoa procyanidins for preventing the development of obesity, insulin resistance, and impaired glucose tolerance during high-fat feeding. *J Agr Food Chem* **62**: 2216-27.
- Dudzińska D, Boncler M, Watala C. 2015. The cardioprotective power of leaves. *AMS*, **11**: 819–839.
- Dulloo AG, Duret C, Rohrer D, Girardier L, Mensi N, Fathi M, Chantre P, Vandermander J. 1999. Efficacy of green tea extracts rich in catechin polyphenols and caffeine in increasing 24-h energy expenditure and fat oxidation in humans. *AJCN*. **70**:1040-5.
- Ejaz A, Wu D, Kwan P, Meydani M. 2009. Curcumin inhibits adipogenesis in 3T3-L1 adipocytes and angiogenesis and obesity in C57/BL mice. *J Nutr* **139**: 919-25.
- Etcheverry P, Grusak MA, Fleige LE. 2012. Application of in vitro bioaccessibility and bioavailability methods for calcium, carotenoids, folate, iron, magnesium, polyphenols, zinc, and vitamins B6, B12, D, and E. *Frontiers in physiology* **3**:317 (Epub ahead of print; DOI doi: 10.3389/fphys.2012.00317).
- Farhat G, Drummond S, Fyfe L, McDougall G, Al-Dujaili EA. 2015. Comparison of the Effects of High versus Low-Polyphenol Dark Chocolate on Body Weight and Biochemical Markers: A Randomized Trial. *EC Nutrition* **2**: 354-64.

- Ganjali S., Blesso C.N., Banach M., Pirro M, Majeed, M, Sahebkar A. 2017. Effects of curcumin on HDL functionality. *Pharmacological Research* **119**: 208–218.
- Golomb BA, Koperski S, White HL. 2012. Association between more frequent chocolate consumption and lower body mass index. *Arch Intern Med* **172**: 519-21.
- González-Ortiz M, Martínez-Abundis E, Espinel-Bermúdez MC, Pérez-Rubio KG. 2011. Effect of pomegranate juice on insulin secretion and sensitivity in patients with obesity. *Ann Nutr Metab.* **58**: 220-3.
- Gu Y, Hurst WJ, Stuart DA, Lambert JD. 2011. Inhibition of key digestive enzymes by cocoa extracts and procyanidins. *J Agr Food Chem* **59**:5305-11.
- Gu Y, Yu S, Lambert JD. Dietary cocoa ameliorates obesity-related inflammation in high fat-fed mice. 2014. *EJCN* **53**:149-58.
- Gulvady AA, Ciolino HP, Cabrera RM, Jolly CA. 2013. Resveratrol inhibits the deleterious effects of diet-induced obesity on thymic function. *J Nutr Biochem* **24**: 1625-33.
- Hamman RF, Wing RR, Edelstein SL, Lachin JM, Bray GA, Delahanty L, Hoskin M, Kriska AM, Mayer-Davis EJ, Pi-Sunyer X, Regensteiner J. 2006. Effect of weight loss with lifestyle intervention on risk of diabetes. *Diabetes care* **29**:2102-7.
- Headland M, Clifton PM, Carter S, Keogh JB. 2016. Weight-Loss Outcomes: A Systematic Review and Meta-Analysis of Intermittent Energy Restriction Trials Lasting a Minimum of 6 Months. *Nutrients*, **8**: E354. (Epub ahead of print; doi: 10.3390/nu8060354).
- Hollis JH, Houchins JA, Blumberg JB, Mattes RD. 2009. Effects of concord grape juice on appetite, diet, body weight, lipid profile, and antioxidant status of adults. *J Am Coll Nutr* **28**: 574-82.
- Hsu CH, Tsai TH, Kao YH, Hwang KC, Tseng TY, Chou P. 2008. Effect of green tea extract on obese women: a randomized, double-blind, placebo-controlled clinical trial. *Clin Nutr* **27**: 363-70.
- Hsu CL, Yen GC. 2007. Effects of flavonoids and phenolic acids on the inhibition of adipogenesis in 3T3-L1 adipocytes. *J Agr Food Chem* **55**: 8404-10.
- Hughes LA, Arts IC, Ambergen T, Brants HA, Dagnelie PC, Goldbohm RA, van den Brandt PA, Weijnenberg MP. 2008. Higher dietary flavone, flavanol, and catechin intakes are associated with less of an increase in BMI over time in women: a longitudinal analysis from the Netherlands Cohort Study. *Am J Clin Nutr* **88**: 1341–52.
- Hursel R, Viechtbauer W, Westerterp-Plantenga MS. 2009. The effects of green tea on weight loss and weight maintenance: a meta-analysis. *Int J Obesity* **33**: 956-61.
- Hurst WJ, Glinski JA, Miller KB, Apgar J, Davey MH, Stuart DA. 2008. Survey of the trans-resveratrol and trans-piceid content of cocoa-containing and chocolate products. *J Agr Food Chem* **56**: 8374-8.
- Izzo AA, Hoon-Kim S, Radhakrishnan R, Williamson E.M. 2016. A critical approach to evaluating clinical efficacy, adverse events and drug interactions of herbal remedies. *Phytother Res.***30**: 691-700.
- Jousilahti P, Tuomilehto J, Vartiainen E, Pekkanen J, Puska P. 1996. Body weight, cardiovascular risk factors, and coronary mortality 15-year follow-up of middle-aged men and women in eastern Finland. *Circulation* **93**: 1372-9
- Jung HA, Jung HJ, Jeong HY, Kwon HJ, Ali MY, Choi JS. 2014. Phlorotannins isolated from the edible brown alga *Ecklonia stolonifera* exert anti-adipogenic activity on 3T3-L1 adipocytes by downregulating C/EBP α and PPAR γ . *Fitoterapia* **92**: 260-9.
- Krzyszczoszek J, Wierzejska E, Zielinska A. 2015. Obesity. An analysis of epidemiological and prognostic research. *AMS* **11**: 24-33.

- Kunnumakkara AB, Bordoloi D, Padmavathi G, Monisha J, Roy NK, Prasad S, Aggarwal BB. 2016. Curcumin, the golden nutraceutical: multitargeting for multiple chronic diseases. *Br J Pharmacol*. (Epub ahead of print; doi: 10.1111/bph.13621)
- Lei F, Zhang XN, Wang W, Xing DM, Xie WD, Su H, Du LJ. 2007. Evidence of anti-obesity effects of the pomegranate leaf extract in high-fat diet induced obese mice. *Int J Obesity* **31**: 1023-9.
- Leibel RL. 2008. Molecular physiology of weight regulation in mice and humans. *Int J Obesity* **32**: S98-108.
- Manach C, Scalbert A, Morand C, Rémésy C, Jiménez L. Polyphenols: food sources and bioavailability. 2004. *AJCN*. **79**: 727-47.
- Martin KR, Appel CL. Polyphenols as dietary supplements: a double-edged sword. 2010. *Nutr Diet Suppl*. **2**: 1-2.
- Matsui N, Ito R, Nishimura E, Yoshikawa M, Kato M, Kamei M, Shibata H, Matsumoto I, Abe K, Hashizume S. 2005. Ingested cocoa can prevent high-fat diet-induced obesity by regulating the expression of genes for fatty acid metabolism. *Nutrition* **21**: 594-601.
- McDougall GJ, Stewart D. 2005. The inhibitory effects of berry polyphenols on digestive enzymes. *Biofactors* **23**: 189-95.
- Meydani M, Hasan ST. 2010. Dietary polyphenols and obesity. *Nutrients*. **2**:737-51.
- Michalska M, Gluba A, Mikhailidis D.P., Nowak P, Bielecka-Dabrowa, A., Rysz, J. and Banach, M., 2010. The role of polyphenols in cardiovascular disease. *Medical Science Monitor* **16**: RA110-9.
- Min SY, Yang H, Seo SG, Shin SH, Chung MY, Kim J, Lee SJ, Lee HJ, Lee KW. 2013. Cocoa polyphenols suppress adipogenesis in vitro and obesity in vivo by targeting insulin receptor. *Int J Obesity* **37**: 584-92.
- Moher D, Liberati A, Tetzlaff J, Altman DG. 2009. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *Ann Intern Med* **151**:264-9.
- Nagao T, Hase T, Tokimitsu I. 2007. A green tea extract high in catechins reduces body fat and cardiovascular risks in humans. *Obesity* **15**: 1473-83.
- Nagao T, Komine Y, Soga S, Meguro S, Hase T, Tanaka Y, Tokimitsu I. 2005. Ingestion of a tea rich in catechins leads to a reduction in body fat and malondialdehyde-modified LDL in men. *AJCN* **81**:122-9.
- Natsume M, Osakabe N, Oyama M, Sasaki M, Baba S, Nakamura Y, Osawa T, Terao J. 2003. Structures of (–)-epicatechin glucuronide identified from plasma and urine after oral ingestion of (–)-epicatechin: differences between human and rat. *Free Radical Bio Med* **34**: 840-9.
- Nomura S, Ichinose T, Jinde M, Kawashima Y, Tachiyashiki K, Imaizumi K. 2008. Tea catechins enhance the mRNA expression of uncoupling protein 1 in rat brown adipose tissue. *J Nutr Biochem* **19**: 840-7.
- Osakabe N. 2013. Flavan 3-ols improve metabolic syndrome risk factors: evidence and mechanisms.2013. *J Clin Biochem Nutr*. **52**: 186-92.
- Panickar KS. 2013. Effects of dietary polyphenols on neuroregulatory factors and pathways that mediate food intake and energy regulation in obesity. *Mol Nutr Food Res* **57**: 34-47.
- Patti AM, Toth PP, Giglio RV, Banach M, Noto M, Nikolic D, Montalto G, Rizzo M. 2017. Nutraceuticals as an Important Part of Combination Therapy in Dyslipidaemia. *Curr Pharm Des* (Epub ahead of print; doi: 10.2174/1381612823666170317145851).

Pfeuffer M, Auinger A, Bley U, Kraus-Stojanowic I, Laue C, Winkler P, Rüfer CE, Frank J, Bösch-Saadatmandi C, Rimbach G, Schrezenmeir J. Effect of quercetin on traits of the metabolic syndrome, endothelial function and inflammation in men with different APOE isoforms. *Nutr Metab Cardiovas* **23**: 403-9.

Poulsen MM, Vestergaard PF, Clasen BF, Radko Y, Christensen LP, Stødkilde-Jørgensen H, Møller N, Jessen N, Pedersen SB, Jørgensen JO. 2013. High-dose resveratrol supplementation in obese men an investigator-initiated, randomized, placebo-controlled clinical trial of substrate metabolism, insulin sensitivity, and body composition. *Diabetes* **62**:1186-95.

Rangel-Huerta OD, Aguilera CM, Martin MV, Soto MJ, Rico MC, Vallejo F, Tomas-Barberan F, Perez-de-la-Cruz AJ, Gil A, Mesa MD. 2015. Normal or high polyphenol concentration in orange juice affects antioxidant activity, blood pressure, and body weight in obese or overweight adults. *J Nutr* **145**: 1808-16.

Santini A, Novellino E. 2016. Nutraceuticals in hypercholesterolaemia: an overview. *British journal of pharmacology*. Br J Pharmacol (Epub ahead of print : doi: 10.1111/bph.13636).

Seo MJ, Lee YJ, Hwang JH, Kim KJ, Lee BY. 2015. The inhibitory effects of quercetin on obesity and obesity-induced inflammation by regulation of MAPK signaling. *J Nutr Biochem* **26**: 1308-16.

Serban C, Sahebkar A, Ursoniu S, Andrica F, Banach, M. 2015. Effect of sour tea (*Hibiscus sabdariffa* L.) on arterial hypertension: a systematic review and meta-analysis of randomized controlled trials. *Journal of hypertension* **33**: 1119-1127.

Shao W, Yu Z, Chiang Y, Yang Y, Chai T, Foltz W, Lu H, Fantus IG, Jin T. 2012. Curcumin prevents high fat diet induced insulin resistance and obesity via attenuating lipogenesis in liver and inflammatory pathway in adipocytes. *PLoS one* [serial online] **7**:e28784. Internet: <http://journals.plos.org/plosone/article/file?id=10.1371/journal.pone.0028784&type=printable> (accessed 20 December 2016).

Shimoda H, Tanaka J, Kikuchi M, Fukuda T, Ito H, Hatano T, Yoshida T. Effect of polyphenol-rich extract from walnut on diet-induced hypertriglyceridemia in mice via enhancement of fatty acid oxidation in the liver. 2009. *J Agr Food Chem*. **57**: 1786-92.

Shin HC, Kim SH, Park Y, Lee BH, Hwang HJ. 2012. Effects of 12-week Oral Supplementation of *Ecklonia cava* Polyphenols on Anthropometric and Blood Lipid Parameters in Overweight Korean Individuals: A Double-blind Randomized Clinical Trial. *Phytother Res* **26**: 363-8.

Smeriglio A, Barreca D, Bellocco E, Trombetta, D. 2016. Chemistry, pharmacology and health benefits of anthocyanins. *Phytother Res* **30**: 1265-1286.

Stevens J, Truesdale KP, McClain JE, Cai J. 2006. The definition of weight maintenance. *Int J Obes* **30**: 391-9.

Stohs SJ, Badmaev V. 2016. A Review of Natural Stimulant and Non-stimulant Thermogenic Agents. *Phytother Res* **30**: 732–740

Suliburska J, Bogdanski P, Szulinska M, Stepien M, Pupek-Musialik D, Jablecka A. 2012. Effects of green tea supplementation on elements, total antioxidants, lipids, and glucose values in the serum of obese patients. *Biol Trace Elem Res* **149**: 315-22.

Tian C, Ye X, Zhang R, Long J, Ren W, Ding S, Liao D, Jin X, Wu H, Xu S, Ying C. 2013. Green tea polyphenols reduced fat deposits in high fat-fed rats via erk1/2-PPAR γ -adiponectin pathway. *PLoS one* [serial online] **8**:e53796. Internet: <http://europepmc.org/backend/ptpmcrender.fcgi?accid=PMC3546082&blobtype=pdf> (accessed 20 December 2016).

Tominaga Y, Mae T, Kitano M, Sakamoto Y, Ikematsu H, Nakagawa K. 2006. Licorice flavonoid oil effects body weight loss by reduction of body fat mass in overweight subjects. *J Health Sci*. **52**: 672-83.

Tominaga Y, Nakagawa K, Mae T, Kitano M, Yokota S, Arai T, Ikematsu H, Inoue S. 2009. Licorice flavonoid oil reduces total body fat and visceral fat in overweight subjects: A randomized, double-blind, placebo-controlled study. *Obes Res Clin Pract* **3**:169-78.

Verweij LM, Terwee CB, Proper KI, Hulshof CT, van Mechelen W. 2013. Measurement error of waist circumference: gaps in knowledge. *Public Health Nutr*. **16**: 281-8.

Wang H, Wen Y, Du Y, Yan X, Guo H, Rycroft JA, Boon N, Kovacs EM, Mela DJ. 2010. Effects of catechin enriched green tea on body composition. *Obesity* **18**:773-9.

Westerterp-Plantenga MS, Lejeune MP, Kovacs EM. 2005. Body weight loss and weight maintenance in relation to habitual caffeine intake and green tea supplementation. *Obes Res* **13**: 1195-204.

WHO. 2016. Obesity and overweight. Internet:
<http://www.who.int/mediacentre/factsheets/fs311/en/> (accessed 20 February 2017).

Wu CH, Lu FH, Chang CS, Chang TC, Wang RH, Chang CJ. 2003. Relationship among habitual tea consumption, percent body fat, and body fat distribution. *Obes Res* **11**: 1088-95.

Zaki ME, El-Bassyouni HT, El-Gammal M, Kamal S. 2015. Indicators of the metabolic syndrome in obese adolescents. *Arch Med Sci* **11**: 92-98.

Table 1: Overall details of included studies

Study/participants characteristics	Length (weeks)	Intervention	Energy content /day	Polyphenol content per day//type (s)	Anthropometric measurements	Baseline	Post-intervention (Mean change from Baseline)
Dallas et al., 2014 (26) <i>France</i> double-blinded controlled parallel study N=95 (42% men) Age: 22-45 years BMI: 26 - 29.9 kg/m ²	12	Two capsules of a) citrus polyphenolic extract with red orange, grapefruit, sweet orange and guarana (Sinetrol-Xpur) or b) maltodextrin placebo)	NEGL	a)900 mg / Catechin (~810 mg), Flavanones (at least 20% (naringin) b) NEGL	Body weight BMI WC Waist-to-hip ratio BF %	EG: 78.14 ±1.35 CG: 77.39 ± 1.23 EG: 27.58 ± 0.16 CG: 27.27 ± 0.14 EG: 88.68 ± 1.05 CG: 88.44 ± 1.09 EG: 0.813 ± 0.113 CG: 0.809 ± 0.113 EG: 37.97 ± 1.59 CG: 36.87 ± 1.48	EG: 75.52 ± 1.25* (-2.62) CG: 75.78 ± 1.23 (-1.58) EG: 26.39 ± 0.33 (-1.19) CG: 26.12 ± 0.35 (-1.15) EG: 83.53 ± 0.87* (-5.15) CG: 87.02 ± 1.02 (-1.42) EG:0.784 ± 0.155 (-0.03) CG:0.808 ± 0.101 (-0.001) EG: 34.36 ± 1.49* (-3.01) CG: 35.85 ± 1.51 (-1.02)
Chang et al., 2014 (27) <i>Taiwan</i> double-blinded controlled parallel study N= 36 (58% men) Age: 18-65 years BMI ≥ 27 kg/m ²	12	6 capsules of a) Hibiscus sabdariffa extract (HSE) or b) starch (placebo)	NEGL	a)152 mg / Flavonoids (~39mg) Anthocyanins (68 mg) Phenolic acids (46 mg) b) NEGL	Body weight BMI WC Waist-to-hip Ratio BF %	EG: 88.52 ± 15.96 CG:84.93 ± 12.79 EG:31.51 ± 4.01 CG:30.91 ± 3.71 EG:98 ± 11.75 CG:95.32 ± 10.43 EG:0.91 ± 0.07 CG:0.9 ± 0.06 EG:37.37 ± 6.22 CG: 38.44 ± 9.8	EG: 87.28 ± 16.02* (-1.24) CG: 84.27 ± 13.14 (-0.66) EG: 31.09 ± 4.23* (-0.42) CG: 30.65 ± 3.76 (-0.26) EG: 97.16 ± 10.94 (-0.84) CG: 95.94 ± 10.25 (0.62) EG: 0.9 ± 0.06* (-0.01) CG: 0.9 ± 0.06 (0) EG:36.67 ± 6.61 (-0.7) CG: 39.08 ± 9.82 (0.64)
Hollis et al., 2009 (28) <i>USA</i> double-blinded controlled parallel study N= 76 (61% men) Age: 18-50 years BMI 25-29.9 kg/m ²	12	a) 480 ml of Concord Grape Juice (CGJ) or b) 480 ml of substitute (polyphenol-free) grape flavored drink (SGD; control) or c) no beverage (NTG)	CGJ: 350 Kcal; SGJ: 350 Kcal	a)934 mg / anthocyanins (191 mg), procyanidins (307 mg) catechin, quercetin, & myricetin. b) NEGL c) NEGL	Body weight BMI WC	EG: 79 ± 8.4 CG: 79 ± 10.7 EG: 27 ± 1.6 CG: 27 ± 1.5 EG: 83.31 ± 6.6 CG: 84.07 ± 7.62	EG : 79.7 ± 9.5 (0.7) CG: 80.6 ± 11.2* (1.6) EG: :27.1 ± 2.0 (0.1) CG: 27.2 ± 1.9 (0.2) EG: 82.04 ± 7.1* (-1.57) CG: 83.31 ± 8.64 (-0.76)

Pfeuffer et al, 2013 (29) Germany double-blinded crossover study N= 49 (100 % males) Age: 48 ± 68 years BMI: 26.3 ± 0.3 kg/m ²	Two 8-week treatment (3 week washout period)	Six capsules of a) quercetin dihydrate (150 mg/day) or b) placebo	NEGL	a)150mg / quercetin b) NEGL	Body weight	+	NS†
					BMI	+	NS†
					WC	+	† (-0.63)*
Shin et al., 2012 (30) USA double-blinded parallel controlled study N= 97 (34% males) Age: 40.5 ± 9.2 years BMI: 26.5 ± 1.6 kg/m ²	12	246 ml of a)Ecklonia cava Polyphenol extract (ECP) drink low dose b) ECP high dose or c) control drink (sugars, sodium chloride, citric acid, vit C and lemon flavour)	~ 85 Kcal (a, b,c)	a)ECP low dose: 72 mg/ Phlorotannins (unique polyphenols) b) ECP high dose: 144 mg) / Phlorotannins c) NEGL	Body weight	ECP HD: 71.2 ± 9.4 ECP LD: 70.5 ± 10 CG: 71.3 ± 9.4	ECP HD: 69.9 ± 9.6* (-1.3) ECP LD: 69.6 ± 9.6* (-0.9) CG: 70.9 ± 9.7 (-0.4)
					BMI	ECP HD: 31.4 ± 5.4 ECP LD: 26.4 ± 1.7 CG:26.5 ± 1.5	ECP HD: 29.3 ± 5.4* (-2.1) ECP LD: 26.2 ± 1.7* (-0.2) CG: 26.4 ± 1.8 (-0.1)
					Body fat %	ECP HD: 31.4 ± 5.4 ECP LD: 30.8 ± 5.3 CG: 31.7 ± 5.2	ECP HD: 29.3 ± 5.4* (-2.1) ECP LD: 29.9 ± 5.4* (-0.9) CG: 31.3 ± 5.3* (-0.4)
					WC	ECP HD: 90.7 ± 7.1 ECP LD: 90.4 ± 6.8 CG: 90.8 ± 6.3	ECP HD: 89.3 ± 6.3* (-1.4) ECP LD: 89.6 ± 5.9* (-0.8) CG: 90.3 ± 6* (-0.5)
					Waist-to hip ratio	ECP LD:90.2 ± 5.9 ECP LD: 89.9 ± 5.6 CG: 90.3 ± 4.9	ECP LD: 88.8 ± 5.5* (-1.4) ECP LD: 88.9 ± 5.8* (-1) CG: 89.7 ± 5 (-0.6)
Rangel-Huerta et al., 2015 (31), Spain Double-blinded crossover study N= 100 (8 % males) Age: 50 ± 3 years 25 ≤ BMI < 40 kg/m ²	Two 12-week period (7 week washout period)	500 ml of orange juice a)normal polyphenol concentration (NPJ) b) high polyphenol concentration (HPJ)	NPJ: 245 Kcal ; HPJ : 303 Kcal	a)NPJ : 299.5 mg/ Flavanones (237 mg of hesperidin 45 mg) of naringin, and 17 mg of didymin) b)HPJ: 742.5 mg/ Flavanones (582.5 mg mg of hesperidin, 125 mg of naringin, and 34 mg didymin)	Body weight	NPJ: 90.4 ± 1.5 HPJ: 90.6 ± 1.5	NPJ: 89.1 ± 1.5 * (-1.3) HPJ: 88.88 ± 1.5 * (-1.8)
					BMI	NPJ: 32.5 ± 0.4 HPJ: 32.6 ± 0.4	NPJ: 32.0 ± 0.4* (-0.5) HPJ: 31.9 ± 0.4* (-0.7)
					WC	NPJ: 99.1 ± 1.3 HPJ: 99.4 ± 1.1	NPJ: 95.1 ± 1.2 * (-4.0) HPJ: 95.6 ± 1.1* (-3.8)

Poulsen et al., 2013 (32) <i>Denmark</i> double-blinded parallel controlled study N= 24 (100 %males) Age: 38.4 ± 2.6 years BMI: 34.2 ± 0.7 kg/m ²	4	Four capsules of a) resveratrol or b) placebo	NEGL	a) 500 mg of resveratrol b) NEGL	Body weight BMI BF % Visceral fat volume	EG: 107.1 ± 2.7 CG: 115.9 ± 3.6 EG: 32.5 ± 0.6 CG: 35.9 ± 1.2 EG: 30.2 ± 0.7 CG: 32.3 ± 1.4 EG: 4996 ± 876 CG: 4285 ± 585	NS † NS † NS † NS †
Nagao et al., 2005 (33) <i>Japan</i> Double-blinded parallel controlled study N= 35 (100 % males) Age: 24-46 years BMI: 24.9 ± 0.4 (EG) BMI: 25 ± 0.4 (CG) (Kg/m ²)	12	One bottle (340 ml) of a) Oolong tea with amount of catechins or b) Oolong tea with low amount of catechins	NEGL	a) 690 mg of catechins b) 22 mg of catechins / CT, CG, GC, GCG, EC, ECG, EGC, EGCG	Body weight BMI WC Body Fat mass	EG: 73.9 ± 1.8 CG: 73.8 ± 1.3 EG: 24.9 ± 0.4 CG: 25.0 ± 0.4 EG: 87.9 ± 1.4 CG: 87.8 ± 1.1 EG: 19.7 ± 0.8 CG: 19.5 ± 1	EG: 71.5 ± 1.7* (-2.4) CG: 72.5 ± 1.4 (-1.3) EG: 24.1 ± 0.4* (-0.8) CG: 24.6 ± 0.4 (-0.4) EG: 84.5 ± 1.3* (-3.4) CG: 86.2 ± 1.2 (-1.6) EG: 18.3 ± 0.9* (-1.4) CG: 18.8 ± 1.1 (-0.6)
Nagao et al., 2007 (11) <i>Japan</i> Double-blinded controlled study N= 240 (58% males) Age: 41.7 ± 9.9 years BMI: 26.8 ± 2.0 kg/m ²	12	One can of a) Green tea (340 ml) high in catechins or b) Green tea low in catechins	NEGL	a) 583 mg catechins b) 96 mg of catechins/ CT, CG, GC, GCG, EC, ECG, EGC, EGCG	Body weight BMI WC BF % Visceral fat area	EG: 73.3 ± 9.7 CG: 72.1 ± 10 EG: 26.9 ± 1.9 CG: 26.7 ± 2.1 EG: 87.2 ± 5.2 CG: 86.5 ± 6.1 EG: 30.7 ± 6.4 CG: 30.7 ± 5.4 EG: 109.2 ± 42.3 CG: 107.7 ± 44	EG: 71.6 ± 9.8* (-1.7) CG: 72.1 ± 10.3 (-0.1) EG: 26.2 ± 1.9* (-0.6) CG: 26.6 ± 2.2 (-0.0) EG: 84.7 ± 5.5* (-2.5) CG: 86.5 ± 6.7 (0) EG: 28.3 ± 6.1* (-2.5) CG: 30 ± 5.6* (-0.7) EG: 98.9 ± 38.6* (-10.3) CG: 103.8 ± 38.9 (-3.9)
Wang et al., 2010 (34) <i>China</i> Double-blinded parallel controlled study N= 182 (27%males) Age: 18-55 years BMI: 24-35 kg/m ²	~12	Tea bags infusion : a) Green tea 1 (GT1); b) GT2 c) GT3 or c) control	NEGL	GT1: 458 mg; GT2: 468 mg ; GT3: 886 mg; Control: 30mg/ CT, CG, GC, GCG, EC, ECG, EGC, EGCG	Body weight WC BF % Intra-abdominal fat	GT3: 71.1 ± 11.9 GT2: 71.5 ± 11.8 GT1: 71.4 ± 9.8 CG: 69.7 ± 8.9 GT3: 95.5 ± 6.9 GT2: 95.9 ± 7 GT1: 96.1 ± 5.8 CG: 94.5 ± 6 GT3: 32.2 ± 5.3 GT2: 33.1 ± 5 GT1: 32.7 ± 5.1 CG: 33.2 ± 4.7 GT3: 83.6 ± 3.2 GT2: 84.7 ± 3.1 GT1: 85 ± 2.8	GT3: 69.9 ± 12.1* (-1.2) GT2: 70.7 ± 11.7 (-0.8) GT1: 70.7 ± 10.1 (-0.7) CG: 69.8 ± 9.1 (0.1) GT3: 93.6 ± 7* (-1.9) GT2: 94.6 ± 7 (-1.3) GT1: 95 ± 6.2 (-1.1) CG: 94.3 ± 5.8 (-0.2) GT3: 32.2 ± 5.2 (-0.5) GT2: 32.2 ± 4.9 (-0.9) GT1: 32 ± 5.1* (-0.7) CG: 33 ± 4.7 (-0.2) GT3: 78 ± 3.4* (-5.6) GT2: 81.1 ± 3.1 (-3.6) GT1: 81 ± 3 (4.1)

						CG: 80.4 ± 3.1	CG: 79.3 ± 2.9 (-1.1)
Brown et al., 2011 (35) <i>UK</i> Double-blinded cross-over controlled study N= 69 (100% males) Age: 40-69 years 28 ≤ BMI ≤ 38 kg/m ²	Two 6-week period (2 week washout period)	Capsules of a) decaffeinated green tea extract (530mg) b) lactose (placebo)	NEGL	530 mg catechins/ CT (6.15 mg), CG (0.16 mg), GC (10.7 mg), GCG (6.7 mg), EC (46 mg), ECG (32 mg), EGC (86 mg), EGCG (216 mg) b) NEGL	Body weight	EG: 101.3 ± 11.4 CG: 100.2 ± 11.1	EG: † (-0.64)* ‡ CG: † (0.53)* ‡
Basu et al., 2010 (36) <i>USA</i> Single-blinded controlled parallel study N=35 (23% men) Age: 42.5 ± 1.7 years BMI 36.1 ± 1.3 kg/m ²	8	a) Four cups of decaffeinated green tea beverage or b) 2 capsules of green tea extract capsules or c) water (4 cups)	NEGL	a) 928mg of catechins b) 870 mg/ CT, CG, GC, GCG, EC, ECG, EGC, EGCG c) NEGL	Body weight BMI WC BF %	GT: 96.4 ± 4.7 GTE: 106.2 ± 7.5 Control: 102.7 ± 6.6 GT: 34.6 ± 1.5 GTE: 38 ± 2.3 Control: 36.4 ± 2.8 GT: 105 ± 2.8 GTE: 115 ± 6.4 Control: 108 ± 5.08 GT: 42 ± 2.8 GTE: 42 ± 2.8 Control: 44 ± 3.4	† #
Hsu et al., 2008 (37) <i>Taiwan</i> Double-blinded controlled parallel study N=100 (0 % men) Age: 43 ± 11.8 years BMI: 30.8 ± 4.1 kg/m ²	12	Three capsules of a) Green tea extract (GTE) b) cellulose (placebo)	NEGL	a) 205 mg of catechins/ CT (2.76 mg), CG (2.05 mg), GC (20.5 mg), GCG (9.16 mg), EC (23 mg), ECG (10.6 mg), EGC (12.3 mg), EGCG (125 mg) b) NEGL	Body weight BMI WC	EG: 78.5 ± 10.3 CG: 76.3 ± 14.5 EG: 31.2 ± 3.5 CG: 30.5 ± 4.6 EG: 94.7 ± 7.7 CG: 93 ± 12.6	EG: 78.3 ± 10.6 (-0.15) CG: 76.2 ± 14.4 (-0.03) EG: 31.1 ± 3.7 (-0.06) CG: 30.5 ± 4.6 (-0.006) EG: 93 ± 8.5 *(-1.7) CG: 91.7 ± 11.5 (1.3)
Sulliburska et al., 2012 (38) <i>Poland</i> Double-blinded controlled parallel study N=46 (50 % men) Age: 30-60 years	12	One capsule of a) Green tea extract (GTE) or b) placebo capsules	NEGL	a) 379 mg/ CT, CG, GC, GCG, EC, ECG, EGC, EGCG (208 mg) b) NEGL	BMI WC	EG: 32.7 ± 2.4 CG: 33.45 ± 2.7 EG: 101.8 ± 6.4 CG: 105 ± 6.5	EG: 31.7 ± 2.29 (-1) CG: 33.36 ± 2.66 (-0.09) EG: 101.15 ± 6.42 (-0.65) CG: 105.02 ± 6.1 (-0.02)

BMI \geq 30 kg/m ²							
Tominaga et al., 2006 (39)	12	One capsule of a) licorice flavonoid oil (LFO) or b) MCT+ beeswax (placebo)	NEGL	24 mg/ flavonoids (Glabridin)	Body weight	EG: 72.69 \pm 1.21 CG: 72.71 \pm 1.13	EG: \dagger NS CG: (1)*
Japan Double-blinded controlled parallel study N=103 (61% men) Age: 46.1 \pm 1.2 years BMI 26.5 \pm 0.2 kg/m ²					BMI	EG: 26.37 \pm 0.21 CG: 26.5 \pm 0.2	EG: \dagger NS CG: \dagger (0.5*) :
Tominaga et al., 2009 (40)	8	a) 300 mg of licorice flavonoid oil (LFO) b) 600mg of LFO or c) 900 mg LFO d) placebo (MCT+ beeswax)	NEGL	1 % Glabridin per LFO solution	Body weight	CG:71.13 \pm 2.2 LFO: 300mg: 71.09 \pm 1.98 LFO 600 mg: 71.88 \pm 1.71 LFO 900: 72.79 \pm 1.82	CG: 73.39 \pm 2.22 (2.26) LFO 300 mg: 70.81 \pm 1.97 (-0.28) LFO 600 mg: 71.68 \pm 1.68 (-0.2) LFO 900: 72.01 \pm 1.81* (-0.78)
Japan Double-blinded controlled parallel study N=84 (67 % men) Age: 40-60 years BMI 24 - 30 kg/m ²					BMI	CG::26.51 \pm 0.34 LFO 300mg: 26.28 \pm 0.31 LFO 600 mg: 26.33 \pm 0.29 LFO 900mg: 26.22 \pm 0.3	CG: 26.59 \pm 0.33 (0.08) LFO 300 mg: 26.17 \pm 0.32 (-0.11) LFO 600 mg: 26.27 \pm 0.3 (-0.06) LFO 900 mg: 25.97 \pm 0.3* (-0.25)
					Body fat mass	CG::22.65 \pm 1.12 LFO 300mg: 22.56 \pm 1.29 LFO 600 mg: 21.46 \pm 0.92 LFO 900 mg: 22.59 \pm 0.9	CG: 22.3 \pm 1.24 (-0.35) LFO 300mg: 21.64 \pm 1.31* (-0.92) LFO 600 mg: 20.53 \pm 0.98* (-0.93) LFO 900 mg: 21.7 \pm 0.91* (-0.89)
					Visceral fat area	CG: 115.26 \pm 10.07 LFO 300mg: 120.13 \pm 10.6 LFO 600 mg: 117.16 \pm 6.3 LFO 900 mg: 122.37 \pm 8.2	CG: 110.58 \pm 9.67 (-4.68) LFO 300mg:116.03 \pm 10.57(-4.1) LFO 600 mg: 117.49 \pm 7.1(0.33) LFO 900 mg:113.02 \pm 7.72* (9.35)
Bell et al., 2011 (41)	8	Three capsules of a) Licorice Flavonoid oil (LFO) (300 mg) or b) placebo (MCT + beeswax)	NEGL	30% polyphenols / flavonoids (3% glabridin)	Body weight	EG:88.7 \pm 4 CG:92.3 \pm 2.9	EG: 88.6 \pm 3.7 (-0.1) CG: 92 \pm 3 (-0.3)
USA Double-blinded parallel controlled study N= 22 Age: 20-53 years BMI: 25-36 kg/m ²					BMI	EG:29.4 \pm 1.3 CG:30.2 \pm 1.2	EG: 29.4 \pm 1.3 (0) CG: 30.1 \pm 1.3 (-0.1)
					WC	EG:88.5 \pm 3.2 CG: 90.7 \pm 2	EG:88.9 \pm 3 (0.4) CG: 90.9 \pm 2.1 (0.2)
					BF %	EG:33 \pm 2.9 CG:32.1 \pm 2.1	EG: 32.7 \pm 3.2 (-0.3) CG: 32.2 \pm 2.1 (0.1)
Barth et al., 2012 (42)	4	a)Polyphenol-rich cloudy apply juice (CLOA) (750 ml) or	a) : 373 Kcal/ b) CB:353 Kcal	a) 802.5 mg/ phenolic acids, quercetin b) NEGL	Body weight	EG: 99 \pm 14.2 CG: 97.6 \pm 13	EG: 99.3 \pm 14.3 (0.3) CG: 97.8 \pm 12.9 (0.2)
Germany Blinded parallel controlled study					BMI	EG:31.1 \pm 3.6 CG:30.5 \pm 3	EG:: 31.1 \pm 3.6 (0) CG 30.6 \pm 3.2 (0.1)

N= 68 (100% males)		b)control beverage (CB)		WC	EG:107.3 ± 9.9 CG:106.9 ± 7.9	EG: 107.4 ± 9.7 (0.1) CG: 107.3 ± 7.9 (0.1)	
Age: 23-69 years				Body fat %	EG:29.3 ± 3.6 CG:29 ± 2.7	EG: 28.3 ± 3.9 (-1)* CG: 28.8 ± 2.7 (-0.2)	
BMI > 27 kg/m ²							
Akazome et al., 2010	12	Beverage (340 g)	NEGL	600 mg/	Body weight	EG: 74.1 ± 8.7 CG: 75.2 ± 8.5	EG: 73.2 ± 8.7* (-0.9) CG: 75.2 ± 8.2 (0)
(43)		containing a)apple			BMI	EG:27 ± 1.7 CG:26.9 ± 1.6	EG:26.7 ± 1.6* (- 0.3) CG:26.9 ± 1.5 (0)
Japan		polyphenols or b)			WC	EG: 94.3 ± 5 CG:94.3 ± 4.5	EG: 92.1 ± 4.7* (-2.2) CG:94.2 ± 4.2 (-0.1)
double-blinded		no polyphenols			Waist/hip ratio	EG: 0.951 ± 0.041 CG: 0.951 ± 0.041	EG: 0.942 ± 0.042* (-0.009) CG: 0.952 ± 0.42 (0.001)
controlled parallel		(placebo)			Body fat %	EG: 28.3 ± 5.8 CG: 28.8 ± 6.2	EG: 28.9 ± 5.6 (0.5) CG: 29.8 ± 6.1* (1)
study					Visceral fat area	EG: 106.1 ± 34.3 CG: 99.2 ± 31.5	EG: 98.2 ± 28.2* (-7.9) CG: 101.3± 35.2 (2.1)
N=94 (60 % men)							
Age: 45.8 ± 8.2 years							
BMI: 27.2± 1.6 kg/m ²							

BF: Body Fat; CG: Control group; CT: Catechins; CG: Catechin gallate ;CloA: Polyphenol-rich cloudy apply juice; GC: Galliccatechin; GCG: Galliccatechin gallate; EG: Experimental group; EC: Epicatechin; ECG: Epicatechin gallate; ECP: Ecklonia cava polyphenols; EGG: Epigallocatechin; EGCG: Epigallocatechin gallate; GT: Green tea; GTE: Green tea extract; High polyphenol concentration (HPJ); HSE: Hibiscus sabdariffa extract; LFO: Licorice flavonoid oil; ; MCT: Medium-chain Triglycerides; NEGL: Negligible; Normal polyphenol concentration (NPJ); NS: Non significant; WC: Waist circumference

Body weight (Kg), BMI (kg/m²), Waist circumference (WC), Body fat mass (Kg), visceral/abdominal fat area (cm²), Visceral fat volume area (cm³)

*significant difference p<0.05

† missing data

‡ Significant effects (p< 0.05) only in intervention period 1

No significant differences in all parameters from baseline between experimental group and control group.

