



FACULDADE DE MEDICINA
UNIVERSIDADE DO PORTO

MESTRADO INTEGRADO EM MEDICINA

2012/2013

Diogo da Silva Miguel
Cardiac adaptations to exercise: the
role of stem cells and future
therapeutic insights.

março, 2013

FMUP



FACULDADE DE MEDICINA
UNIVERSIDADE DO PORTO

Diogo da Silva Miguel
Cardiac adaptations to exercise: the
role of stem cells and future
therapeutic insights.

Mestrado Integrado em Medicina

Área: Fisiologia

**Trabalho efetuado sob a Orientação de:
Doutor Paulo Castro Chaves**

**Trabalho organizado de acordo com as normas da revista:
Cardiovascular Research**

março, 2013

FMUP

Eu, Diogo da Silva Miguel, abaixo assinado, nº mecanográfico 070801255, estudante do 6º ano do Mestrado Integrado em Medicina, na Faculdade de Medicina da Universidade do Porto, declaro ter atuado com absoluta integridade na elaboração deste projeto de opção.

Neste sentido, confirmo que **NÃO** incorri em plágio (ato pelo qual um indivíduo, mesmo por omissão, assume a autoria de um determinado trabalho intelectual, ou partes dele). Mais declaro que todas as frases que retirei de trabalhos anteriores pertencentes a outros autores, foram referenciadas, ou redigidas com novas palavras, tendo colocado, neste caso, a citação da fonte bibliográfica.

Faculdade de Medicina da Universidade do Porto, 20/03/2013

Assinatura:



Nome: Diogo da Silva Miguel

Email: mimed07255@med.up.pt

Título da Monografia:

Cardiac adaptations to exercise: the role of stem cells and future therapeutic insights.

Orientador:

Paulo Manuel Barreiros de Castro Chaves

Ano de conclusão: 2013

Designação da área do projeto:

Fisiologia

É autorizada a reprodução integral desta Monografia para efeitos de investigação e de divulgação pedagógica, em programas e projetos coordenados pela FMUP.

Faculdade de Medicina da Universidade do Porto, 20/03/2013

Assinatura:



Cardiac adaptations to exercise: the role of stem cells and future therapeutic insights

Diogo da Silva Miguel¹

Faculty of Medicine of University of Porto, Physiology and Cardiothoracic Department

mimed07255@med.up.pt

Paulo Castro Chaves MD, PhD

Faculty of Medicine of University of Porto, Physiology and Cardiothoracic Department

pchaves@med.up.pt

Number of words: 5339

¹Corresponding author

Faculdade de Medicina da Universidade do Porto, Al. Prof. Hernâni Monteiro, 4200-319 Porto, Portugal

Telephone number: +351 22 551 3600

Fax: +351 22 551 3601

E-mail: mimed07255@med.up.pt

Diogo da Silva Miguel, Paulo Castro Chaves

Cardiac adaptations to exercise: the role of stem cells and future therapeutic insights

Abstract

The heart responds to physical exercise through changes in its morphology known as physiological hypertrophy. Besides cardiomyocyte hypertrophy, there is also an increased cellular proliferation with formation of new myocytes. Altogether, these changes lead to an improvement in cardiac function. The molecular mechanism responsible for the development of physiological hypertrophy is centred in the IGF-1R/PI3K/Akt signalling pathway, which modulates the activity of transcription factors involved in protein synthesis and cardiomyocytes proliferation. The endogenous pool of cardiac stem cells is believed to be crucial for the continuous renewal and regenerative potential of cardiac tissue, and has also been positively correlated with physiological hypertrophy. Another system involved in myocardium physiology and pathology is microRNA. The knowledge of the mechanisms underlying the beneficial effects of physical exercise over cardiac function may sustain the development of new therapeutic approaches regarding cardiac pathologies, which represent today an important cause of morbidity and mortality worldwide.

Keywords: Exercise, Heart, Hypertrophy, microRNA, Stem cells.

Introduction

The understanding of heart's physiology has always been on the top of scientific community's priorities. Indeed, there is an enormous amount of work available regarding this area of knowledge.

The heart responds to increased workload with hypertrophy. According to the nature of the stimulus, the hypertrophy can be either physiological or pathological¹. The main stimulus that leads to pathological hypertrophy is pressure overload, where the heart responds with hypertrophy of existing myocytes as an attempt to overwhelm the increased load². However, in this setting maladaptive changes, such as apoptosis, necrosis and a shift in gene expression, also occur, which leads to rapid decline in cardiac function, with heart failure as possible outcome^{1, 2}. In the opposite side there is the physiological hypertrophy, with physical exercise appearing as the best example of this type of cardiac adaptation. Unlike pressure overload, exercise training increases the blood flow through the heart (volume overload)¹, which in turn results in lengthening of myocytes and consequent eccentric hypertrophy of the whole organ². The normal structure of the myocardium is kept, the cardiac function remains unchanged or may improve³, and the loss of myocytes and increased fibrosis observed in pathological hypertrophy are actually attenuated with exercise training⁴.

As noted above, physical exercise is the main trigger of physiological hypertrophy, with all the resulting benefits. Exercise training is today extensively prescribed and is a cheap and effective way for both the prevention and management of heart diseases³. Indeed, current evidence points that an improved fitness level is a strong indicator of freedom from all-cause mortality⁵ and is enough to decrease the morbidity and mortality linked with cardiac disorders⁶. Besides, physical exercise has been shown to protect the myocardium from age induced apoptosis, fibrosis and senescence^{4, 7, 8}.

Until the past decade, it was believed that the heart was a post-mitotic organ, with no regenerative potential. However, an increasing body of evidence supports the existence of a resident population of cardiac stem cells⁹⁻¹². This endogenous pool not only reassures the continuous renewal of cardiac tissue¹³, but also supports some grade of regeneration after an injury^{9, 11}. The old paradigm of the heart as a terminally differentiated organ without any renewal potential did not give space for the exploration of regenerative medicine. However, by the light of the evidence accumulated during the past decade some work begun to be done in the field of regenerative medicine applied to the heart.

This review aims to look over the mechanisms known to take part in the control of the adaptations of myocardium to physical exercise, and open new perspectives about possible applications of this knowledge in the search of new therapeutic approaches.

Macroscopic and functional changes

In 1975 Morganroth *et al.* used echocardiography to describe for the first time the cardiac changes induced by physical exercise, concluding that endurance exercise induced an increase in left ventricle mass and diastolic cavity size, while the wall thickness remained normal (eccentric hypertrophy). On the other hand, athletes that went through a resistance training program showed increased myocardial mass with ventricular wall thickening, but with normal diastolic cavity size (concentric hypertrophy)¹⁴. This concept has been accepted for decades as the “Morganroth hypothesis”. Indeed, other studies have found an increase in both left ventricular mass and left ventricle end diastolic diameter with endurance exercise¹⁵⁻¹⁷, though some of them have failed to show any effect of resistance training in cardiac morphology^{15, 16}.

Adaptations at cellular level

Unlike the macroscopic remodelling described in the previous paragraph, due to ethical limitations there is a paucity of studies describing the changes that occur at the cellular level in humans. However, a vast amount of studies using mice have shown that an increase in individual cardiomyocyte dimensions occurs in response to several swimming training programmes¹⁸⁻²⁰. That increase comprises both the cardiomyocyte's long and short axis and mean cell area²¹, and is remarkably related with the exercise's intensity – high intensity exercise induces bigger changes than moderate intensity⁶. On the other hand, the pattern of cardiomyocyte growth is also determined by the type of physical exercise: while endurance training (swimming, running), and the associated volume overload, prompt myocyte lengthening, resistance training (wrestling, weight lifting), which is associated with an intermittent pressure overload, drives mainly to thickening of myocytes².

Until recently, cardiomyocyte hypertrophy was thought to be the only mean by which the heart increased its size, which was in line with the paradigm that heart was a post-mitotic organ without any renewal potential. However, nowadays' current evidence indicates that human myocardium is capable of, at least in part, renovate itself by new cardiomyocyte formation¹³. Actually, if it was not this self-renewing capacity, the heart would lose most of its mass in few decades¹¹. Furthermore, increased levels of division markers, along with a rise in the number of cardiomyocytes, has already been documented in mice following the application of endurance exercise protocols^{18, 22}. Interestingly, the increase in the number of cardiomyocytes was positively related with the intensity of physical exercise the rats have been exposed to²². Therefore, it seems that the regenerative capacity of myocardium is enhanced by physical activity.

Another possible mechanism by which the heart can increase its mass is through a drop in apoptotic rate. Indeed, several studies point in this direction. Endurance exercise has been found to attenuate the rise in Bax/Bcl-2 ratio seen with ageing. Consequently there is a decrease in the activation of caspase-9 and 3 and DNA fragmentation, which is reflected in the superior number of myocytes per area observed in old exercised rats compared with sedentary ones⁴. On the other hand, it was also demonstrated a rise in the levels of heat shock protein 70 (HSP70), known to inhibit apoptosis, in the ventricles of exercised mice⁷. At last but not the least, an improved telomerase activity was observed in mice exposed to endurance exercise, thereby protecting myocytes from senescence and apoptosis. This was found already after 21 days of exercise, even before physiological hypertrophy has been documented⁸. Furthermore, in studies using obese rats, endurance exercise was also shown to decrease the number of apoptotic cardiomyocytes^{23, 24} and reverse the architectural changes observed to occur in association with obesity²³.

Molecular mechanisms

IGF-1R/PI3K/Akt pathway. The IGF-1R/PI3K/Akt pathway has been extensively studied and is seen as the hallmark of physiological hypertrophy^{20, 25, 26}. In fact, the nature of myocardial adaptation to increased workload (physiological vs. pathological) begins to differentiate already at the growth factor and receptor level – while the activation of G-protein coupled receptors by adrenergic stimuli results in pathological adaptation of myocardium²⁷, in the presence of physiological stimuli such as physical exercise it is the insulin-like growth factor I (IGF-1) acting over its receptor (IGF-1R) and triggering the beneficial adaptations described above²⁵⁻²⁷.

In one study using rats, it was shown that swimming training increased the IGF-1 mRNA after 2 and 6 weeks of exercise²⁸. These findings are compatible with the work of Seneri *et al.* that assessed the concentration of this growth factor in coronary sinus and concluded that it was significantly bigger in athletes than in controls²⁹. The importance of the pair IGF-1/IGF-1R was further highlighted by knock-out studies in which the deletion of IGF-1R in mice attenuated the cardiac hypertrophy seen in controls after swimming training²⁰. On the other hand, the overexpression of IGF-1R was associated with a proportional increase in the chamber size and wall thickness of heart, without any histologic signs of cardiomyopathy, what was also reflected in the enhanced systolic function of transgenic IGF-1R animals²⁵.

IGF-1R is a tyrosine kinase receptor which once stimulated by its ligand phosphorylates and activates phosphatidylinositol 3-kinase (PI3K)³⁰. PI3Ks are a family of ubiquitous expressed lipid kinases, which are essential for a wide range of biological processes in several different types of tissues²⁷. Class-I PI3K phosphorylates phosphatidylinositol-4,5-bisphosphate to generate phosphatidylinositol-3,4,5-trisphosphate, which in turn acts as second messenger to activate Akt. Class-I_A PI3K is activated by growth factor receptors, such as IGF-1R, while class-I_B PI3K is activated by G-protein coupled receptors²⁷. The importance of class-I_A PI3K activity for the development of physiological hypertrophy was well documented by studies in which mice lacking either the catalytic or regulatory subunit of this kinase failed to show physiological hypertrophy²⁵⁻²⁷. However, the same didn't happen when in the presence of a pathological stimulus, since even in the absence of the catalytic subunit of class-I_A PI3K, pressure overload successfully induced pathological hypertrophy²⁶.

The downstream effector of the IGF-1R/PI3K/Akt pathway is the serine-threonine kinase Akt, also known as protein kinase B (PKB). There have been identified 3

isoforms of this kinase (Akt1, 2 and 3), and despite their high degree of homology, their expression varies among different tissues³¹. Data suggests that the most important isoform in cardiac physiology is Akt1^{19, 31}. As noted above, PI3K plays a key role in the activation of Akt through generation of phosphatidylinositol-3,4,5-trisphosphate. PTEN has the opposite effect, and thus decreases the activation of Akt³¹. Akt phosphorylates several different targets, which implicates it in a variety of biological processes³¹. The importance of Akt for the development of physiological hypertrophy is well supported by a study in which mice lacking Akt1 failed in achieving the degree of cardiac adaptation seen in their control littermates after a program of swimming training¹⁹. In addition, this importance is further reinforced by the evidence that overexpression of Akt causes cardiac hypertrophy both at the molecular and histological level, without any parallel increase in the collagen content of the heart.

Downstream targets. The mammalian target of rapamycin (mTOR) works as an important effector that controls protein synthesis, promoting cellular growth. This is one of the downstream targets of Akt, which phosphorylates and inactivates the product of tuberous sclerosis gene 2 (TSC2). Doing so, Akt counteracts the inhibition that TSC2 exerts over mTOR, thereby enhancing its growth promoting effect¹⁹. This effect is mediated by the phosphorylation of the ribosomal S6 kinase (S6K), that once activated phosphorylates and activates the ribosomal protein S6³². In addition, the translation initiation factor-4E binding protein-1 (4E-BP1) sees its activity increased by mTOR⁶. The net effect is an increased protein synthesis, with consequent cellular hypertrophy⁶. The facts that mice lacking Akt1 showed lower levels of phosphorylated S6K and S6 after exercise training¹⁹ and that overexpression of IGF-1R was associated with an increase in phosphorylated S6K²⁵ highlight the importance of the IGF-1R/PI3K/Akt pathway for the induction of this system. However, the absence of S6K did not

attenuate cardiac physiological hypertrophy when mice were subjected to an exercise protocol, which suggests that, while important, S6K is not essential for the development of physiological hypertrophy³².

Another mean by which Akt induces myocyte hypertrophy is by inactivation of glycogen synthase kinase-3 beta (GSK-3 β)¹⁹. GSK-3 β phosphorylates and inactivates the transcription factor GATA-4, thereby impairing protein synthesis³³. Indeed, increased levels of the inactive phosphorylated form of GSK-3 β were detected in mice overexpressing Akt, which reflected in the bigger accumulation of GATA-4 in cardiomyocytes' nuclei³⁰. Once in the nucleus, GATA-4 regulates the expression of a variety of cardiac genes related with myocardium hypertrophy³³. Moreover, GATA-4 has also been connected to myocyte proliferation in zebrafish³⁴

Besides GATA-4, several other transcription factors have been associated to cardiomyocytes hypertrophy³³. One of the identified transcription factors was C/EBP β , whose expression was found to be reduced after exercise training. Interestingly, that fall was secondary to a higher Akt1 activity, and resulted in a reduction of the repressive effect that C/EBP β has over the serum response factor (SRF). SRF is also a transcription factor that, among others, promotes the expression of GATA-4. Another downstream target negatively regulated by C/EBP β is the CBP/p300-interacting transactivator with ED-rich carboxy-terminal domain 4 (CITED4). In fact, CITED4 was markedly increased in cardiomyocytes and induced its proliferation, as reflected by an increase in the number of cells¹⁸.

The role of microRNAs

MicroRNAs (miRNA) are a class of recently discovered small molecules, each composed of a noncoding sequence of nearly 22 nucleotides, which can modulate gene

expression by destroying or repressing the translation of a target RNA. Each miRNA molecule can target more than a different mRNA, and each mRNA can be targeted by more than a different miRNA, which provides a fine control of gene expression³⁵. Some miRNAs have already been identified that are known to play important roles in cardiac development. Namely, miRNA-1 and miRNA-133 are of uttermost importance in cardiac development by regulating the proliferation and differentiation of cardiomyocytes³⁶. The role of miRNA-1 and miRNA-133 in adult heart is less certain, but both pathological and physiological hypertrophic stimuli decrease the expression of both miRNAs, which suggests an inverse correlation between miRNA-1 and miRNA-133 and myocardium hypertrophy³⁷.

In an attempt to identify miRNAs regulated by PI3K, Lin *et al.* found that mice expressing constitutively active PI3K (caPI3K) expressed lower levels of miRNA-222, miRNA-34a and miRNA-210 as opposed with controls. Moreover, although the levels of these miRNAs increased after myocardial infarction, they remained significantly lower in the group expressing caPI3K, and this was reflected in the improved cardiac function shown in this group³⁸. Another miRNA that has been shown to be related with physiological hypertrophy following a swim training programme was miRNA-29c. However, unlike the former, miRNA-29c levels were upregulated after a programme of several weeks of physical exercise. The consequence was a downregulation in the expression of COL1A1 and COL3A1, with a decrease of total left ventricle collagen content and improved ventricular compliance³⁹.

Endogenous Cardiac Stem Cells

As described above, the heart harbours a pool of endogenous cardiac stem cells (eCSC). The fact that exercised animals' hearts have shown evidence of new cardiomyocytes

formation raises a question – what is the relationship between physical exercise and eCSC?

Some studies exist that try to answer this question. In one of them, the number of c-Kit^{POS} eCSC was found to be increased in exercised rats in comparison with controls²². Also the IGF-1R/PI3K/Akt pathway has been connected with eCSC. In this field, current evidence points to a positive correlation between the activation of this pathway and the activation and proliferation of eCSC^{22, 40}. Moreover, IGF-1 overexpression increased the activity of telomerase, which resulted in a delay in eCSC senescence⁴¹. Therefore, if the activation of IGF-1R/PI3K/Akt fosters the activation and proliferation of eCSC, and if physical exercise promotes the activation of the IGF-1R/PI3K/Akt signalling pathway, so it is reasonable to hypothesize that physical exercise is an effective trigger of eCSC activation and consequent cardiac regeneration.

Clinical insights

So far, the only available therapeutic options aim to block the progression on the heart disease³⁸. However, as noted above, the last decade brought crescent scientific evidence that supports the possibility of exploring new therapeutic approaches that can improve the function of the failing heart, thereby improving the quality of life of millions of people suffering from cardiac pathology.

IGF-1R/PI3K/Akt pathway. As stated above, the IGF-1R/PI3K/Akt pathway is the central core of physiological hypertrophy. In fact, some studies have already been performed in the setting of myocardial infarction that showed a reduction in cardiac fibrosis, cardiomyocytes apoptosis and infarction size, as well as better systolic and diastolic function, following the administration of IGF-1^{42, 43}. Furthermore, in animals subjected to myocardial infarction, the treatment with an association of IGF-1 and

hepatocyte growth factor (HGF) fostered eCSC activation with consequent cardiac regeneration^{40, 44, 45}. Interestingly, this was observed along with increased expression of GATA-4⁴⁴ which, as described above, seems to have a role in physiological hypertrophy induced by physical exercise.

microRNAs. The discovery of miRNAs has brought a new biological target for future therapies. Actually, today is already possible either to increase the expression, using vectors⁴⁶, or antagonize specific miRNAs using ‘antagomirs’, which are cholesterol-associated RNAs that block the action of miRNAs over their targets⁴⁷. The expression of miRNA-34a, aforementioned to be decreased in the cardiomyocytes of mice expressing caPI3K, has been shown to rise in the ageing heart and following myocardial infarction. Interestingly, using antigomir against miRNA-34a it was possible to observe a drop in cell death and cardiac fibrosis, and consequent improvement in contractile function in mice subjected to myocardial infarction⁴⁸. Another miRNA described above as being negatively correlated with PI3K activity was miRNA-210. By those findings, it would be reasonable to think that a decrease in the expression of this miRNA would be beneficial. However, other studies found that miRNA-210, whose expression is upregulated under hypoxic conditions⁴⁹, is actually beneficial in the setting of myocardial infarction by significantly inducing angiogenesis and decreasing myocytes’ apoptotic rate, thereby contributing for a more favourable ventricular remodelling and cardiac function⁴⁶.

Cell therapy. The first attempts to regenerate myocardial tissue have been done through transplant of stem cells from other origins than the heart^{50, 51}. Although animal studies have provided exciting results, several clinical trials in humans were already performed and showed only modest, if any, improvement in cardiac function⁵⁰.

Therefore, new approaches are needed in order to make this type of therapy more effective.

One of the first questions to be answered is which type of cells suits better for cell-based therapy. The discovery of a pool of resident cardiac stem cells has opened a new window regarding the regenerative potential of this organ, which can be further explored either as a therapeutic tool or target. Indeed, a phase 1 clinical trial has documented significant improvements in both the ejection fraction and infarct size in patients that received autologous cardiac stem cells therapy compared with controls. Furthermore, the safety of the treatment was absolute, with no adverse effects registered in any of the treated patients⁵².

The pool of cardiac stem cells is complex and comprises cells with different capacity to replicate and form mature myocytes and blood vessels. In an attempt to better characterize those different cellular populations, D'Amario *et al.* concluded that cardiac stem cells expressing IGF-1R were younger and had a better replicative reserve, as opposed with cells expressing IGF-2R and AT1R. Moreover, cells expressing IGF-1R were capable of secreting both IGF-1 and IGF-2, the latter being responsible for the induction of myocyte differentiation. The superiority of IGF-1R expressing cardiac stem cells was further confirmed by the greater degree of cardiac recovery after the transplant of these cells into hearts of rats subjected to myocardial infarction⁵³.

Besides the choice of the ideal cell, other strategies can be taken to maximize the beneficial effects of cell therapy. Interestingly, the combination of cardiac stem cells administration and nanofibers containing IGF-1 resulted in a higher myocardial recovery after ischemia when compared with each therapy alone⁵⁴.

Discussion

In the last decade several steps have been taken in order to open new lines of investigation regarding new therapeutic strategies. Maybe the biggest advances have been done in the field of cell therapy, with some clinical trials already in progress. However, as stated above, there is also growing expectation about miRNAs and its manipulation.

The central role that the IGF-1R/PI3K/Akt has in the development of physiological hypertrophy has also triggered attempts to modulate this signalling system, namely through the therapy with IGF-1. In fact, the results obtained with IGF-1 administration in animal models of myocardial infarction were encouraging^{42, 43}. However, this signalling pathway is present in a wide range of cell types and because of that systemic administration of this growth factor can have diffuse effects and result in the progression of occult neoplasms³¹.

Concerning cell therapy, some obstacles have been raised regarding the type of cells that better fulfil the requirements for the success of this therapeutic approach. By one side, the application of cells from other origins than the heart has shown disappointing results. Among the causes advanced for this is the source and inadequate preparation of the cells, as well as the timing and mode of cell delivery. On the other side, it is believed that embryonic stem cells are the ones that have the greatest potential to be successful. However, to the risk of teratoma formation and immunogenic incompatibilities that the use of embryonic stem cells entails, we still have to add the ethical issues⁵⁰. Finally, it appears that autologous cardiac stem cells transplantation may be the best approach, as has been shown by a recent phase 1 clinical trial⁵². Still, more research is needed to find new strategies that can increase the feasibility and effectiveness of cell therapy.

The discovery of miRNAs has provided a new overview of cellular regulation mechanisms. In addition, it represents another process where it is possible to act to change the fate of diseased heart. Currently, it is already clear that different miRNAs are important in both the heart development and pathology⁵⁵. Furthermore, some studies performed in animals were successful in improving the pattern of cardiac remodelling after myocardial infarction^{46, 48}. Nevertheless, miRNAs represent a class of molecules that exerts a very delicate control of specific genes at a post-transcriptional level⁵⁵. Consequently, it is critical to better understand the function of each specific miRNA in the heart's physiology and pathology, and only after it will be possible to delineate the best plans to take all the potential from this promising area.

Heart diseases, namely those of ischemic cause, are a major source of morbidity and mortality worldwide⁵¹. Despite the improvement that the development of several classes of drugs have brought to the prognostic of these patients, advances are need so that it can be possible to further decrease the burden of disease of cardiac pathologies.

Conflict of interest

None declared.

Bibliography

1. Dorn GW, II. The fuzzy logic of physiological cardiac hypertrophy. *Hypertension* 2007;**49**:962-970.
2. Opie LH, Commerford PJ, Gersh BJ, Pfeffer MA. Controversies in Cardiology 4 - Controversies in ventricular remodelling. *Lancet* 2006;**367**:356-367.
3. Ellison GM, Waring CD, Vicinanza C, Torella D. Physiological cardiac remodelling in response to endurance exercise training: cellular and molecular mechanisms. *Heart (British Cardiac Society)* 2012;**98**:5-10.

4. Kwak H-B, Song W, Lawler JM. Exercise training attenuates age-induced elevation in Bax/Bcl-2 ratio, apoptosis, and remodeling in the rat heart. *Faseb Journal* 2006;**20**:791-+.
5. O'Keefe JH, Vogel R, Lavie CJ, Cordain L. Exercise like a hunter-gatherer: a prescription for organic physical fitness. *Progress in cardiovascular diseases* 2011;**53**:471-479.
6. Kemi OJ, Wisloff U. Mechanisms of exercise-induced improvements in the contractile apparatus of the mammalian myocardium. *Acta physiologica (Oxford, England)* 2010;**199**:425-439.
7. Siu PM, Bryner RW, Martyn JK, Alway SE. Apoptotic adaptations from exercise training in skeletal and cardiac muscles. *Faseb Journal* 2004;**18**:1150-+.
8. Werner C, Hanhoun M, Widmann T, Kazakov A, Semenov A, Poss J, *et al.* Effects of physical exercise on myocardial telomere-regulating proteins, survival pathways, and apoptosis. *Journal of the American College of Cardiology* 2008;**52**:470-482.
9. Beltrami AP, Barlucchi L, Torella D, Baker M, Limana F, Chimenti S, *et al.* Adult cardiac stem cells are multipotent and support myocardial regeneration. *Cell* 2003;**114**:763-776.
10. Torella D, Ellison GM, Karakikes I, Nadal-Ginard B. Resident cardiac stem cells. *Cellular and Molecular Life Sciences* 2007;**64**:661-673.
11. Nadal-Ginard B, Kajstura J, Leri A, Anversa P. Myocyte death, growth, and regeneration in cardiac hypertrophy and failure. *Circulation Research* 2003;**92**:139-150.
12. Saravanakumar M, Devaraj H. Distribution and homing pattern of c-kit(+) Sca-1(+) CXCR4(+) resident cardiac stem cells in neonatal, postnatal, and adult mouse heart. *Cardiovascular pathology : the official journal of the Society for Cardiovascular Pathology* 2012.
13. Bergmann O, Bhardwaj RD, Bernard S, Zdunek S, Barnabe-Heider F, Walsh S, *et al.* Evidence for Cardiomyocyte Renewal in Humans. *Science* 2009;**324**:98-102.

14. Lewis EJ, McKillop A, Banks L. The Morganroth hypothesis revisited: endurance exercise elicits eccentric hypertrophy of the heart. *The Journal of physiology* 2012;**590**:2833-2834.
15. Venckunas T, Raugaliene R, Mazutaitiene B, Ramoskeviciute S. Endurance rather than sprint running training increases left ventricular wall thickness in female athletes. *European journal of applied physiology* 2008;**102**:307-311.
16. Spence AL, Naylor LH, Carter HH, Buck CL, Dembo L, Murray CP, *et al.* A prospective randomised longitudinal MRI study of left ventricular adaptation to endurance and resistance exercise training in humans. *The Journal of physiology* 2011;**589**:5443-5452.
17. De Luca A, Stefani L, Pedrizzetti G, Pedri S, Galanti G. The effect of exercise training on left ventricular function in young elite athletes. *Cardiovascular ultrasound* 2011;**9**:27.
18. Bostrom P, Mann N, Wu J, Quintero PA, Plovie ER, Panakova D, *et al.* C/EBP beta Controls Exercise-Induced Cardiac Growth and Protects against Pathological Cardiac Remodeling. *Cell* 2010;**143**:1072-1083.
19. DeBosch B, Treskov I, Lupu TS, Weinheimer C, Kovacs A, Courtois M, *et al.* Akt1 is required for physiological cardiac growth. *Circulation* 2006;**113**:2097-2104.
20. Kim J, Wende AR, Sena S, Theobald HA, Soto J, Sloan C, *et al.* Insulin-Like Growth Factor I Receptor Signaling Is Required for Exercise-Induced Cardiac Hypertrophy. *Molecular Endocrinology* 2008;**22**:2531-2543.
21. McMullen JR. Role of insulin-like growth factor 1 and phosphoinositide 3-kinase in a setting of heart disease. *Clinical and Experimental Pharmacology and Physiology* 2008;**35**:349-354.
22. Waring CD, Vicinanza C, Papalamprou A, Smith AJ, Purushothaman S, Goldspink DF, *et al.* The adult heart responds to increased workload with physiologic hypertrophy, cardiac stem cell activation, and new myocyte formation. *European heart journal* 2012.

23. Lee SD, Shyu WC, Cheng IS, Kuo CH, Chan YS, Lin YM, *et al.* Effects of exercise training on cardiac apoptosis in obese rats. *Nutrition, metabolism, and cardiovascular diseases : NMCD* 2012.
24. Peterson JM, Bryner RW, Sindler A, Frisbee JC, Alway SE. Mitochondrial apoptotic signaling is elevated in cardiac but not skeletal muscle in the obese Zucker rat and is reduced with aerobic exercise. *Journal of applied physiology (Bethesda, Md : 1985)* 2008;**105**:1934-1943.
25. McMullen JR, Shioi T, Huang WY, Zhang L, Tarnavski O, Bisping E, *et al.* The insulin-like growth factor 1 receptor induces physiological heart growth via the phosphoinositide 3-kinase(p110 alpha) pathway. *Journal of Biological Chemistry* 2004;**279**:4782-4793.
26. McMullen JR, Shioi T, Zhang L, Tarnavski O, Sherwood MC, Kang PM, *et al.* Phosphoinositide 3-kinase(p110 alpha) plays a critical role for the induction of physiological, but not pathological, cardiac hypertrophy. *Proceedings of the National Academy of Sciences of the United States of America* 2003;**100**:12355-12360.
27. Luo J, McMullen JR, Sobkiw CL, Zhang L, Dorfman AL, Sherwood MC, *et al.* Class I-A phosphoinositide 3-kinase regulates heart size and physiological cardiac hypertrophy. *Molecular and Cellular Biology* 2005;**25**:9491-9502.
28. Scheinowitz M, Kessler-Icekson G, Freimann S, Zimmermann R, Schaper W, Golomb E, *et al.* Short- and long-term swimming exercise training increases myocardial insulin-like growth factor-I gene expression. *Growth hormone & IGF research : official journal of the Growth Hormone Research Society and the International IGF Research Society* 2003;**13**:19-25.
29. Neri Serneri GG, Boddi M, Modesti PA, Cecioni I, Coppo M, Padeletti L, *et al.* Increased cardiac sympathetic activity and insulin-like growth factor-I formation are associated with physiological hypertrophy in athletes. *Circ Res* 2001;**89**:977-982.
30. Condorelli G, Drusco A, Stassi G, Bellacosa A, Roncarati R, Iaccarino G, *et al.* At induces enhanced myocardial contractility and cell size in vivo in transgenic mice.

Proceedings of the National Academy of Sciences of the United States of America 2002;**99**:12333-12338.

31. Matsui T, Rosenzweig A. Convergent signal transduction pathways controlling cardiomyocyte survival and function: the role of PI 3-kinase and Akt. *Journal of Molecular and Cellular Cardiology* 2005;**38**:63-71.
32. McMullen JR, Shioi T, Zhang L, Tarnavski O, Sherwood MC, Dorfman AL, *et al.* Deletion of ribosomal S6 kinases does not attenuate pathological, physiological, or insulin-like growth factor 1 receptor-phosphoinositide 3-kinase-induced cardiac hypertrophy. *Mol Cell Biol* 2004;**24**:6231-6240.
33. Akazawa H, Komuro I. Roles of cardiac transcription factors in cardiac hypertrophy. *Circulation Research* 2003;**92**:1079-1088.
34. Kikuchi K, Holdway JE, Werdich AA, Anderson RM, Fang Y, Egnaczyk GF, *et al.* Primary contribution to zebrafish heart regeneration by gata4(+) cardiomyocytes. *Nature* 2010;**464**:601-605.
35. Cataluccia D, Latronico MVG, Condorelli G. MicroRNAs control gene expression - Importance for cardiac development and pathophysiology. In: Sideman S, Beyar R, Landesberg A, eds. *Control and Regulation of Transport Phenomena in the Cardiac System*, 2008:20-29.
36. Cordes KR, Srivastava D. MicroRNA Regulation of Cardiovascular Development. *Circulation Research* 2009;**104**:724-732.
37. Latronico MVG, Catalucci D, Condorelli G. Emerging role of MicroRNAs in cardiovascular biology. *Circulation Research* 2007;**101**:1225-1236.
38. Lin RCY, Weeks KL, Gao X-M, Williams RBH, Bernardo BC, Kiriazis H, *et al.* PI3K(p110 alpha) Protects Against Myocardial Infarction-Induced Heart Failure Identification of PI3K-Regulated miRNA and mRNA. *Arteriosclerosis Thrombosis and Vascular Biology* 2010;**30**:724-732.

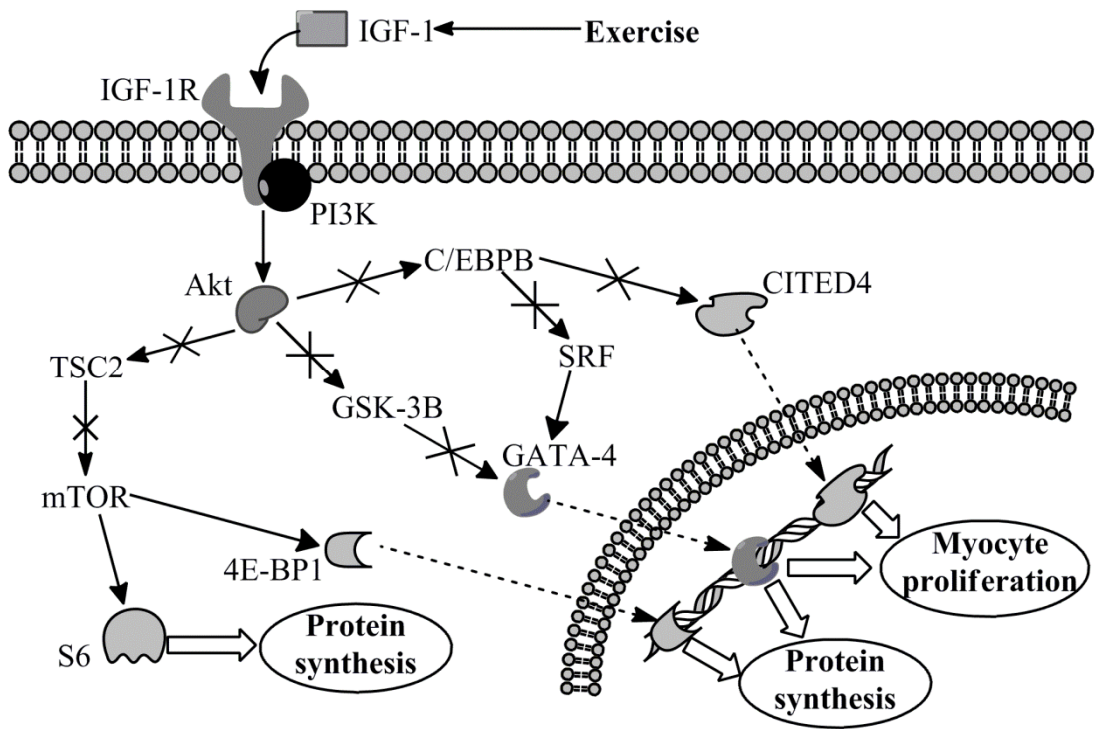
39. Soci UPR, Fernandes T, Hashimoto NY, Mota GF, Amadeu MA, Rosa KT, *et al.* MicroRNAs 29 are involved in the improvement of ventricular compliance promoted by aerobic exercise training in rats. *Physiological Genomics* 2011;**43**:665-673.
40. Ellison GM, Torella D, Dellegrottaglie S, Perez-Martinez C, Perez de Prado A, Vicinanza C, *et al.* Endogenous Cardiac Stem Cell Activation by Insulin-Like Growth Factor-1/Hepatocyte Growth Factor Intracoronary Injection Fosters Survival and Regeneration of the Infarcted Pig Heart. *Journal of the American College of Cardiology* 2011;**58**:977-986.
41. Torella D, Rota M, Nurzynska D, Musso E, Monsen A, Shiraishi I, *et al.* Cardiac stem cell and myocyte aging, heart failure, and insulin-like growth factor-1 overexpression. *Circulation Research* 2004;**94**:514-524.
42. O'Sullivan JF, Leblond AL, Kelly G, Kumar AH, Metharom P, Buneker CK, *et al.* Potent long-term cardioprotective effects of single low-dose insulin-like growth factor-1 treatment postmyocardial infarction. *Circulation Cardiovascular interventions* 2011;**4**:327-335.
43. Lai NC, Tang T, Gao MH, Saito M, Miyano-hara A, Hammond HK. Improved function of the failing rat heart by regulated expression of insulin-like growth factor I via intramuscular gene transfer. *Human gene therapy* 2012;**23**:255-261.
44. Ruvinov E, Leor J, Cohen S. The promotion of myocardial repair by the sequential delivery of IGF-1 and HGF from an injectable alginate biomaterial in a model of acute myocardial infarction. *Biomaterials* 2011;**32**:565-578.
45. Bocchi L, Savi M, Graiani G, Rossi S, Agnetti A, Stillitano F, *et al.* Growth factor-induced mobilization of cardiac progenitor cells reduces the risk of arrhythmias, in a rat model of chronic myocardial infarction. *PloS one* 2011;**6**:e17750.
46. Hu S, Huang M, Li Z, Jia F, Ghosh Z, Lijkwan MA, *et al.* MicroRNA-210 as a novel therapy for treatment of ischemic heart disease. *Circulation* 2010;**122**:S124-131.

47. Krutzfeldt J, Kuwajima S, Braich R, Rajeev KG, Pena J, Tuschl T, *et al.* Specificity, duplex degradation and subcellular localization of antagomirs. *Nucleic acids research* 2007;**35**:2885-2892.
48. Boon RA, Iekushi K, Lechner S, Seeger T, Fischer A, Heydt S, *et al.* MicroRNA-34a regulates cardiac ageing and function. *Nature* 2013.
49. Chan YC, Banerjee J, Choi SY, Sen CK. miR-210: the master hypoxamir. *Microcirculation* 2012;**19**:215-223.
50. Fraccarollo D, Galuppo P, Bauersachs J. Novel therapeutic approaches to post-infarction remodelling. *Cardiovascular research* 2012;**94**:293-303.
51. Dimmeler S, Burchfield J, Zeiher AM. Cell-based therapy of myocardial infarction. *Arteriosclerosis Thrombosis and Vascular Biology* 2008;**28**:208-216.
52. Bolli R, Chugh AR, D'Amario D, Loughran JH, Stoddard MF, Ikram S, *et al.* Cardiac stem cells in patients with ischaemic cardiomyopathy (SCIPIO): initial results of a randomised phase 1 trial. *Lancet* 2011;**378**:1847-1857.
53. D'Amario D, Cabral-Da-Silva MC, Zheng H, Fiorini C, Goichberg P, Steadman E, *et al.* Insulin-like growth factor-1 receptor identifies a pool of human cardiac stem cells with superior therapeutic potential for myocardial regeneration. *Circ Res* 2011;**108**:1467-1481.
54. Padin-Iruegas ME, Misao Y, Davis ME, Segers VF, Esposito G, Tokunou T, *et al.* Cardiac progenitor cells and biotinylated insulin-like growth factor-1 nanofibers improve endogenous and exogenous myocardial regeneration after infarction. *Circulation* 2009;**120**:876-887.
55. Small EM, Olson EN. Pervasive roles of microRNAs in cardiovascular biology. *Nature* 2011;**469**:336-342.

Figure legends

Figure 1 – IGF-1R/PI3K/Akt signalling pathway and downstream targets. Arrows mean activation; arrows with a cross mean inhibition; C/EBPB – C/EBP β ; GSK-3B – GSK-3 β . For more details consult the text.

Figure 1



ANEXO

Instruções aos Autores

Policy

Cardiovascular Research is the international basic science journal of the European Society of Cardiology. The Journal is concerned with both basic and translational research, across different disciplines and areas, enhancing insight in cardiovascular disease mechanisms and the perspective for innovation. The Journal welcomes submission of papers both at the molecular, subcellular, cellular, organ, and organism level, and of clinical proof-of-concept and translational studies.

Manuscripts may be submitted as Original Articles, Rapid Communications, or Reviews. Moreover, the Journal publishes Letters to the Editor and Editorials (the latter are usually invited).

An author should indicate whether his/her manuscript should be considered for one of the Spotlight Issues that address particular themes. Manuscripts are normally evaluated by three members from an international panel of reviewers, and an editorial decision is made on average within 22 days of receipt of a manuscript.

Preparation of manuscripts (for regular papers)

The manuscript should be typed double-spaced and pages should be numbered. Original articles should not exceed 5750 words, including the abstract, manuscript text, references, and figure legends. Abbreviations should be kept to a minimum and should not appear in the Abstract unless they may be understood by non-expert readership. Manuscripts should be submitted electronically (see below, under *Submission*). Authors presently unable to take advantage of online submission should fax the Editorial Office for further instructions (+49 641 99 47 209). Manuscripts submitted to the journal may be checked for originality using anti-plagiarism software.

(1) Title page. This is the first page of the manuscript submission file. Title length should be no longer than 120 characters, including spaces. Provide the names of all authors including first name, department where the work was performed, all authors' affiliations, name of corresponding author with address, telephone number, fax and e-mail. Also give current addresses of any authors who have moved since the work was finished. If there are more than 10 authors, a statement of the contribution of each to the study should be provided in your cover letter. The number of words should be mentioned on the title page.

(2) Abstract. The abstract should be submitted as a separate file. Repeat in normal sized, but bold font, names of the authors and the title of the manuscript at the top of the page. The abstract *should not exceed one page* of the manuscript and should be *250 words or less*. It should be structured into the subsections "Aims," "Methods and Results" and "Conclusion(s)." Give the name of the animal species, if applicable, in the subsection "Methods".

(3) Keywords. These will be published with your article. During online submission, they are typed into a window. A maximum of 5 keywords is allowed. Keywords can be selected from the linked alphabetically formatted [list](#) or can be of your own choice.

(4) Classifications. These are used for administration purposes and selection of reviewers. During online submission they are chosen by ticking boxes in a formatted list. Authors should first choose classifications concerning *Discipline, Object of Study, Level, and Expertise* from the linked [list](#) and then specific classifications, listed here alphabetically. Please tick as many keywords as you feel necessary to characterize your manuscript.

(5) Introduction. This section should position the study with regard to objective, rationale, and preceding work of other authors.

(6) Methods. This section should be divided into headed subsections. To reduce a lengthy methods section, experimental details (buffer compositions, primer sequences, etc.) may be included in a separate supplementary file for online publication. However, each method must be briefly described and thoroughly referenced in the main article.

(6a) NEW: For investigations involving procedures with animals or animal tissues, the main Methods section should provide the generic name of the anaesthetic and analgesic agent(s) used, the dose, and the route and frequency of administration. Note also that neuromuscular blocking or paralytic agents should never be used without general anaesthesia. Methods used for monitoring of the adequacy of anaesthesia must be described. Methods used for euthanasia should likewise be explicitly described. For experiments involving isolated tissues or primary cell cultures, the procedures used for their isolation should be described, including methods of anaesthesia and/or euthanasia. Finally, it should be stated whether the investigation conforms to either the [Guide for the Care and Use of Laboratory Animals](#) published by the United States National Institutes of Health.

Health or the [Directive 2010/63/EU of the European Parliament](#). Please see the [editorial statement of this journal](#) for more specific information on anesthetics.

(6b) If human subjects or tissues are used, you should state whether the investigation conforms with the principles outlined in the [Declaration of Helsinki](#).

(6c) In addition, for both animal and human research, you should declare whether approval was granted by a local or university ethics review board (approval reference number to be given, if available). All manuscripts will be sent to an ethics subeditor for approval, if applicable, before the peer-review process is initiated.

(7) Results. If pertinent, the section may be divided into headed subsections. For presentation of data, figures are preferred to tables. Also, extensive numerical data should appear in legends to the figures rather than in the main body of text. SI units should be used.

(8) Discussion. This section should not contain paragraphs dealing with topics that are beyond the scope of the study. Four manuscript pages should in general be enough to compare and interpret the data with regard to previous work by yourself and others.

(9) Funding. Details of all funding sources for the work in question should be given in a separate section entitled 'Funding'. This should appear before the 'Acknowledgements' section.

The following rules should be followed:

- The sentence should begin: 'This work was supported by ...'
- The full official funding agency name should be given, i.e. 'the National Cancer Institute at the National Institutes of Health' or simply 'National Institutes of Health' not 'NCI' (one of the 27 subinstitutions) or 'NCI at NIH' ([full RIN-approved list of UK funding agencies](#)) Grant numbers should be complete and accurate and provided in brackets as follows: '[grant number ABX CDXXXXXX]'
- Multiple grant numbers should be separated by a comma as follows: '[grant numbers ABX CDXXXXXX, EFX GHXXXXXX]'
- Agencies should be separated by a semi-colon (plus 'and' before the last funding agency)
- Where individuals need to be specified for certain sources of funding the following text should be added after the relevant agency or grant number 'to [author initials]'

An example is given here: 'This work was supported by the National Institutes of Health [AA123456 to C.S., BB765432 to M.H.]; and the Alcohol & Education Research Council [P50 CA098252 and CA118790 to R.B.S.R.]'

Oxford Journals will deposit all NIH-funded articles in PubMed Central. See http://www.oxfordjournals.org/for_authors/repositories.html for details. Authors must ensure that manuscripts are clearly indicated as NIH-funded using the guidelines above.

(10) Acknowledgements.

(11) Conflict of Interest. All authors must make a formal statement indicating any potential conflict of interest that might constitute an embarrassment to any of the authors if it were not to be declared and were to emerge after publication. Such conflicts might include, but are not limited to, shareholding in or receipt of a grant or consultancy fee from a company whose product features in the submitted manuscript or which manufactures a competing product. If none of the authors has a conflict of interest, then type: 'Conflict of Interest: none declared.'

(12) References. Note: This format has been recently changed – journal names should be in italics, volume numbers in bold, and page numbers should be fully written out. In-text citations should be numerical and superscripted.

Regular papers:

Coronel R, Opthof T, Taggart P, Tytgat J, Veldkamp M. Differential electrophysiology of repolarisation from clone to clinic. *Cardiovasc Res* 1997;**33**:503-517.

Books:

Wit AL, Janse MJ. The Ventricular Arrhythmias of Ischemia and Infarction. Electrophysiological Mechanisms. Mount Kisco, NY: Futura Publishing Company, Inc, 1992.

Chapter in book:

Weber KT. Cardiac Interstitium: Extracellular Space of the Myocardium. In: Fozzard HA, Haber E, Jennings RB, Katz AM, Morgan HE, eds. The Heart and Cardiovascular System. Scientific Foundations, 2nd ed. New York: Raven Press, 1991:1465-1480.

Thesis:

Dekker L.R.C. Role of intracellular calcium in ischemic damage and preconditioning in cardiac muscle. Amsterdam: University of Amsterdam. 1996 (Thesis).

Abstract:

Like regular paper, but add (Abstract) at end.

Please note: If the bibliography contains more than six authors, *et al.* should be added following the sixth author.

(13) Figure Legends. Figure legends should start on a new page of the manuscript, but one page may contain legends to more than one figure.

(14) Figures/Tables. A maximum of 6 figures on 6 pages is allowed. These may have multiple panels, but they must be able to withstand reduction by up to 50%. Additional figures may be uploaded as a supplement. Tables can be included in the manuscript file. Figures should be attached as a separate file(s) during the submission process and labelled (entitled "Figure 1", for example, in the box marked "Description" visible during submission). Electronically submitted figures should be of high resolution (300 dpi or greater) and in one of the following formats: tiff (.tif), bitmap (.bmp), jpeg (.jpg), portable data format (.pdf), or postscript (.ps or .eps). Any lettering in the figures should be large enough to stand photographic reduction. You should prepare your figures for either one column width (84 mm) or the entire page width (175 mm). The maximum height is 240 mm. Photomicrographs should contain a scale bar that represents a given length in the figure (e.g. 5 μm). The Publisher will determine the degree of any reduction or enlargement required and in general, line drawings will be reduced to one column width if possible.

(15) Colour Figures. For colour reproduction in print, you will receive information regarding the costs from Oxford Journals after receipt of your accepted article. Each colour page in print costs approx. £350/\$600/€525. For further information on the preparation of electronic artwork, please see <http://cpc.cadmus.com/da>.

Please note: Because of the high cost of colour, authors are advised to submit figures where the colour is not essential in black and white or greyscale. In line graphs, different lines can be indicated with dots, dashes or symbols (♦ ◇ V X + and so on) or with labels and arrows. Bars in bar charts can be black, white, and grey, or include cross hatching.

Where colour is necessary for proper interpretation, figures should be submitted in colour. Manuscripts submitted in colour will be published in colour, both online and in print. Note also that you are required to make a statement in your covering letter whether you agree to pay the cost of printing your colour figures (see below under "Submission").

Language editing

Particularly if English is not your first language, before submitting your manuscript you may wish to have it edited for language. This is not a mandatory step, but may help to ensure that the academic content of your paper is fully understood by journal editors and reviewers. Language editing does not guarantee that your manuscript will be accepted for publication. If you would like information about such services please click [here](#). There are other specialist language editing companies that offer similar services and you can also use any of these. Authors are liable for all costs associated with such services.

Supplementary Data

Supplementary material can be submitted to support and enhance your scientific research. Supplementary files supplied will be published online alongside the electronic version of your article. Authors should submit the material in electronic format together with the article online and supply a concise and descriptive caption for each file. Regarding supplementary methods, please note that a reader should be able to understand what techniques were used, with at least a simple description or adequate reference to another source in the literature. Buffer components, SDS gel composition, primer sequences, etc., may be placed in supplementary methods.

Please note: supplementary data cannot be altered or replaced after the paper has been accepted for publication. This will not undergo typesetting or copyediting.

Preparation of Review Articles

Review articles should be divided into the following sections: a short abstract (unstructured) followed by various subsections that may include an introduction and may also be further subdivided, and a summary or similar concluding section. The maximum number of words is 7500, including references.

Rapid Communications

These are high priority manuscripts that report major advances or provide important, novel insight into the field of cardiovascular medicine and basic science. They are organized like regular manuscripts (above) but are relatively short and concise (no longer than 4000 words, including references, and 5 display items, preferably no colour figures). An accompanying covering letter should justify why it belongs in this category. The decision to admit a manuscript to this track rests with the Editor.

Online copyright licence form

Upon receipt of accepted manuscripts at Oxford Journals authors will be invited to complete an online copyright licence to publish form.

Please note that by submitting an article for publication you confirm that you are the corresponding/submitting author and that Oxford University Press ("OUP") may retain your email address for the purpose of communicating with you about the article. Please notify OUP immediately if your details change. If your article is accepted for publication OUP will contact you using the email address you have used in the registration process. Please note that OUP does not retain copies of rejected articles.

Open access option for authors

Cardiovascular Research authors have the option to publish their paper under the [Oxford Open](#) initiative; whereby, for a charge, their paper will be made freely available online immediately upon publication. After your manuscript is accepted the corresponding author will be required to accept a mandatory licence to publish agreement. As part of the licensing process you will be asked to indicate whether or not you wish to pay for open access. If you do not select the open access option, your paper will be published with standard subscription-based access and you will not be charged.

You can pay Open Access charges using our Author Services site. This will enable you to pay online with a credit/debit card, or request an invoice by email or post.

Open access charges can be viewed [here](#) in detail; discounted rates are available for authors based in some developing countries (click [here](#) for a list of qualifying countries). Please note that these charges are in addition to any colour/page charges that may apply.

Orders from the UK will be subject to the current UK VAT charge. For orders from the rest of the European Union, OUP will assume that the service is provided for business purposes. Please provide a VAT number for yourself or your institution and ensure you account for your own local VAT correctly.

Self-archiving and post-print policy

Authors may deposit the post-print of their article into PubMedCentral, other subject repositories or institutional repositories, but must stipulate that public availability be delayed until 12 months after the first online publication. For further details of this policy please visit: [Author Self-archiving Policy](#)

Submission

Manuscripts should be submitted electronically by the corresponding author at the URL <http://www.editorialmanager.com/cardiovascres/default.asp>. Three files are required to be uploaded for the submission process: (1) the **abstract**; (2) the **manuscript** (with title page, *not* as a PDF file); and (3) the **covering letter** including the following declarations: (i) That "the manuscript, or part of it, has neither been published (except in form of abstract or thesis) nor is currently under consideration for publication by any other journal"; (ii) The submitting author should declare that the co-author(s) has (have) read the manuscript and approved its submission to *Cardiovascular Research*; (iii) In the case of colour figures, the authors should declare that they agree to pay for the cost of printing. A specification of costs will be sent by publisher after final acceptance of the manuscript.

Checklist

Covering letter?
Length of Title?
Addresses and affiliations?
Number of words?
Title and authors repeated at top of Abstract?
Abstract structured?
Abstract length one page?

Species mentioned in Abstract?
Ethics statement?
Figures OK?
Reference format?
Double spacing?