

**Review Article** 

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# The Marine Macroalgae of the Genus *Ulva*: Chemistry, Biological Activities and Potential Applications

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#### Abstract

This review summarizes a literature survey of the marine macroalgae of the genus *Ulva* (Phylum Chlorophyta), covering the period of 1985 to 2012. The secondary metabolites isolated from members of this genus and biological activities of the organic extracts of some *Ulva* species as well as of the isolated metabolites are discussed. The emphasis on their application in food industry and their potential uses as biofilters are also addressed.

**Keywords:** Chlorophyta; *Ulva*; Macroalgae; Secondary metabolites; Biological activities; Food; Biofilters

### Introduction

The genus *Ulva* (Phylum Chlorophyta, Class Ulvophyceae, Order Ulvales, Family Ulvaceae) was first identified by Linnaeus in 1753 [1]. Since then many taxonomists and phycologists have been involved in the identification of *Ulva* species [2] which are notoriously difficult to classify due to the morphological plasticity expressed by many members as well as the few reliable characters available for differentiating taxa [2,3]. Its morphology resembles bright green sheets [4], heavily influenced by environmental conditions, age of the thallus, and life style, making difficult the delineation of species by morphological features alone [2]. There is an important relationship between morphological features of *Ulva* and salinity and nutrient concentrations in these freshwater habitats; however, the most essential physical factor positively correlated with mats development was water depth [3,5].

At present, there are 562 species names in the Algaebase and 99 of which have been flagged as currently accepted taxonomically [6]. These organisms have a potential for rapid and proliferous growth [3] with a ubiquitous distribution with species living in a wide range of habitats and environments [2,3,5,7]. Although *Ulva* species are primarily marine taxa found in saline and salty waters, they can also proliferate in freshwater habitats [6].

# Secondary Metabolites

Although the members of the phylum Chlorophyta are abundant and relatively easy to collect, their chemistry continues to be underrepresented [8]. Among all macroalgae, the green algae with less than 300 known compounds are the least producers of natural compounds when compared to the red (Rhodophyta) and brown algae (Phaeophyta) [9,10]. Anyhow, a wide range of compounds, predominantly terpenes, polyphenols and steroids, have been reported in various marine green algae [11,12]. The chemical composition of these macroalgae was found to vary depending on geographical distribution and seasons and the principal environmental factors affecting the composition being water temperature, salinity, light, nutrients and minerals availability [5].

The samples of *U. lactuca*, collected from different geographical locations, were investigated for their chemical constituents. By using gas chromatography coupled to mass spectrometry (GC-MS), Flodin

1) from the crude extract of *U. lactuca* [13], collected in Turimetta Head, North of Sydney, on the Eastern coast of Australia. Later,  $3-O-\beta$ -D-glucopyranosylstigmasta-5,25-diene (2) (Figure 1) was isolated from the methanol extract of *U. lactuca*, collected from the coast of Alexandria, Egypt [12,14,15]. Using High Performance Liquid Chromatography (HPLC), the Chinese group was able to isolate five *nor*-isoprenoids: (3*S*,5*R*,6*S*,7*E*) 3,5,6-trihydroxy-7-megastigmen-9-one (3), (3*R*,5*R*,6*R*,7*E*) 3,5,6-trihydroxy-7-megastigmen-9-one (4), (3*S*,6*R*) 3,6-dihydroxy-4,7-megastigmadien-9-one (5), grasshopper ketone (6), and isololiolide (7) (Figure 1), from the methanol extract of *U. lactuca*, collected off the coast of Bohai in China [16].

and Whitfield were able to detect 2,4,6-tribromophenol (1) (Figure

This has also happened to *U. fasciata*, a derivative of sphingosine (2-amino-4-octadecene-1,3-diol) and *N*-palmitoyl-2-amino-1,3,4,5-tetrahydroxy-octadecane (8) (Figure 2) were reported from *U. fasciata*, collected in India [15]. The extract of this alga, collected in Asbu-Qir, Egypt, was also found to contain three unsaturated fatty acids: 8-hydroxy-9-decenoic acid (9), 3,3-diol-4-decenoic acid (10), and 3-oxo-4-decenoic acid (11) [8] (Figure 2).

From the dichloromethane extract of *U. fasciata* Delile, collected in the intertidal zone in Vizhinjam harbor - Kerala at the Southwest coast of India, seven diterpenes: labda-14-ene-8-ol (12), labda-14-ene-3 $\alpha$ ,8 $\alpha$ diol (13), labda-14-ene-8 $\alpha$ -9 $\alpha$ -diol (14), labda-14-ene-8 $\alpha$ -hydroxy-3one (15), *ent*-labda-13(16)14-diene-3-one (16), *ent*-labda-13(16),14diene (17), and *ent*-labda-13(16),14-diene-3 $\alpha$ -ol (18) (Figure 3) were isolated and purified by column chromatography and Thin Layer Chromatography (TLC) [11].

Additionally, guai-2-en-10 $\alpha$ -methanol (19) and guai-2-en-10 $\alpha$ -ol (20) (Figure 4) were isolated and identified from the methanol extract

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**Figure 1:** Structures of 2,4,6-tribromophenol (1),  $3-O-\beta$ -D-glucopyranosylstigmasta-5,25-diene (2), (3S,5R,6S,7E) 3,5,6-trihydroxy-7-megastigmen-9-one (3), (3R,5R,6R,7E) 3,5,6-trihydroxy-7-megastigmen-9-one (4), (3S,6R) 3,6-dihydroxy-4,7-megastigmadien-9-one (5), grasshopper ketone (6), and isololiolide (7).



of *U. fasciata* Delile, collected in Vizhinjam and Mullur in Kerala, India [12]. From the methanol extract of the same species collected in the same local, Chakraborty and Paulraj have reported isolation of five sesquiterpenes: 2,5,5-trimethyl-4-(4'-methyl-3'-pentenyl)-2cyclohexen-1-ol (21), 4-isopentyl-3,4,5,5-tetramethyl-2-cyclohexen-1-ol (22, two diastereoisomeric compounds), 6-isopentyl-1,5,5,6tetramethyl-1-cyclohexene (23), and 3,4,5,5-tetramethyl-4-(3'oxopentyl)-2-cyclohexen-1-one (24) (Figure 4) [17].

The only report on chemical constituents of *U. pertusa* by Garson described isolation of two fatty acids: 2(*R*)-hydroxy-hexadecanoic acid (25), and 2-oxo-hexadecanoic acid (26) [18] (Figure 5).

Yildiz et al., reported isolation of gallic acid (27) from the methanol extract of *U. rigida*, collected at the Marmara Sea-Turkey [19]. Additionally, 24-*nor*-22-dehydrocholesterol (28) [20] and isofucosterol (29) [21] were also isolated from this specie (Figure 6).

Roussis et al. have investigated the production and release of

volatile metabolites from *U. rigida* during light and darkness, under natural conditions and after the biological death of the plant [22]. Using GC and GC-MS methods to analyze the chemical constituents released from live *U. rigida* to the atmosphere, they have found that oxygenated metabolites, mainly aldehydes and alcohols, were the major components and aliphatic and aromatic hydrocarbons were present in significant amounts; however vestigial quantities of the halogenated and sulfated compounds were also detected. In order to determine the chemical load of the volatile compounds that would be liberated in the atmosphere at the end of the biological cycle, the volatile oil obtained from steam distillation of *U. rigida* was analyzed by GC-MS. The results showed that dimethyl sulfide, acetaldehyde and dichloromethane were the major compounds. Furthermore, dichloromethane was found to be a major metabolite during the light period [22].

## **Biological Activities**

In the recent years, the importance of marine algae as a supply for new bioactive substances has been growing very rapidly [15,23-27] owing to their capacity to produce metabolites that exhibit various biological activities such as antibacterial, anti-inflammatory, antiproliferative,



**Figure 3:** Structures of labda-14-ene-8-ol (12), labda-14-ene- $3\alpha$ , $8\alpha$ -diol (13), labda-14-ene- $8\alpha$ - $9\alpha$ -diol (14), labda-14-ene- $8\alpha$ -hydroxy-3-one (15), ent-labda-13(16),14-diene-3-one (16), ent-labda-13(16),14-diene (17), and ent-labda-13(16),14-diene- $3\alpha$ -ol (18).



**Figure 4:** Structures of guai-2-en-10α-methanol (19) and guai-2-en-10α-ol (20), 2,5,5–trimethyl-4-(4'-methyl-3'-pentenyl)-2-cyclohexen-1-ol (21), 4-isopentyl-3,4,5,5-tetramethyl-2-cyclohexen-1-ol (22, two diastereoisomeric compounds), 6-isopentyl-1,5,5,6-tetramethyl-1-cyclohexene (23), and 3,4,5,5-tetramethyl-4-(3'-oxopentyl)-2-cyclohexen-1-one (24).



Figure 5: Structures of 2(*R*)-hydroxy-hexadecanoic acid (25) and 2-oxo-hexadecanoic acid (26).



antiviral [28,29], antifungal [27], antineoplastic [28], anticancer, anti-obesity, antidiabetic, antihypertensive, antihyperlipidemic, and antioxidant [30]. Although not many members of the genus *Ulva* have been extensively investigated, there are some interesting aspects of biological activities of the extracts and isolated metabolites. Table 1 lists biological activities of extracts and the secondary metabolites isolated from some *Ulva* species.

It is important to mention also that ulvan, a major Sulfated Polysaccharide (SP) found in the cell wall of green algae, composed mainly of rhamnose, glucuronic acid, iduronic acid, xylose, glucose, sulphate with small amounts of iduronic acid and traces of galactose and represents 8-29% of the algae dry weight [25,26,44-51]. The presence of the sulfate groups and the unusual chemical composition and structure of ulvan render it different biological properties [26,52,53]. This complex SP can also be of potential interest for food, pharmaceutical and agricultural applications [26,51-54]. In the recent years, ulvan has been extensively investigated for development of novel drugs and functional foods [26,49,53]. Because of its peculiar composition and structure [50], it represents a potential source of new functional biopolymer and it is widely used as biomaterial for tissue engineering in regenerative medicine. Furthermore, studies showed that sulfated polysaccharides can also exhibit beneficial biological activities such as anticoagulant, antiviral, antioxidant, antiinflammatory [54] and antiproliferative [27]. As attention has been focused on natural antioxidants in the past years, since synthetic antioxidants such as Butylated Hydroxyanisole (BHA) and Butylated Hydroxytoluene (BHT) were found to implicate in liver damage and carcinogenesis, several natural SPs, including ulvan, were evaluated for their antioxidant activity. Interestingly, ulvan from U. pertusa as well as acetylated and benzoylated ulvans were found to have antioxidant activity including scavenging activity against superoxide and hydroxyl radicals, reducing power and chelating ability [55]. Qi et al., in their study of the antioxidant activities of different molecular weight ulvans from *U. pertusa* (Chlorophyceae), obtained by  $H_2O_2$  degradation, have found that low molecular weight ulvans exhibited a strong antioxidant activity, and their rationale for this was that low molecular weight SPs may incorporate into the cells more efficiently and donate proton more effectively when compared to high molecular weight SPs [56]. Ulvan, from *U. pertusa*, was also found to be a potential antihyperlipidemic agent and had significantly reduced serum Triglyceride (TG), total and Low Density Lipoprotein Cholesterol (LDL-cholesterol) and elevated High Density Lipoprotein Cholesterol (HDL-cholesterol) in mice [26]. The antihyperlipidemic activity of ulvan was found to depend on the molecular weight of ulvan fractions; high molecular weight fraction was more effective on total serum and LDL-cholesterol whereas low molecular weight fractions were more effective on TG and HDLcholesterol [26,57].

Toskas et al. were able to fabricate defect free nanofibers by electrospinning ulvan and Polyvinyl Alcohol (PVA) solutions with appropriate ulvan to PVA mass ratios and polymer concentrations [58]. The spinnability of ulvan, in combination with its interesting biological and physicochemical properties, can lead to new biomedical applications such as drug release system. Recently, Toskas et al. have also succeeded in synthesizing ulvan/chitosan polyelectrolyte membrane which can promote the attachment and proliferation of 7F2 osteoblasts and maintain the cell morphology and viability. This new generation of biomaterials composed of ulvan and chitosan might have high impact in biomedical applications as potential scaffold materials [59].

Another class of secondary metabolites of green algae is sterols. Sterols from green algae have been reported to be non-toxic and capable of reducing blood cholesterol level and were found to be able to reduce the tendency to form a greasy liver and excessive fat deposition in the heart [60].

Finally, it is worth mentioning green algae as an important source of Polyunsaturated Fatty Acids (PUFAs). Due to nutritional values and beneficial effects of PUFAs, many researchers have investigated the suitability of using macroalgae as novel dietary sources of PUFAs. Pereira et al. have detected higher percentages (16%) of  $\alpha$ -Linolenic Acid (ALA), in comparison to Linoleic Acid (LA), in *Ulva* sp., collected at the Algarve coast, Portugal [61]. Moreover, PUFAs can be considered also as having an important ecological role. Alamsjah et al. have found that Hexadeca-4, 7, 10, 13-Tetraenoic Acid (HDTA), Octadeca-6,9,12,15-Tetraenoic Acid (ODTA), and  $\alpha$ -Linolenic Acid, isolated from the methanol extract of *U. fasciata* by bioassay-guided fractionation, exhibited a potent algicidal activity against the red-tide phytoplankton *Heterosigma akashiwo* (with LC<sub>50</sub> 1.35 µg/ml, 0.83 µg/ml and 1.13 µg/ml, respectively) and this result demonstrated the potential of these PUFAs for practical harmful algal bloom control [62].

# Use of Ulva spp. as Food

Although algae are known to contain a multitude of bioactive compounds, the interest in these organisms is also due to their nutritional proprieties [30,31,63]. Edible marine algae, sometimes referred as seaweeds, have attracted a special interest as good sources of nutrients [26] since they are an excellent source of vitamins, carbohydrates, dietary fiber and minerals [14,31,64,65] that could be potentially exploited as functional ingredient for both human and animal health [23,25]. Interestingly, many types of seaweeds used as traditional food in many cultures are known not only to cure various diseases but also to maintain health and were found to have immunomodulating and antitumor activities [36].

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Species	Extracts or Metabolites	Biological activities	References
U. compressa	Methanol	Antioxidant	[31]
U. conglobata	Sulfated polysaccharides	Anticoagulant	[32]
U. lactuca	Dichloromethane/methanol	Antimicrobial	[33]
	Ethanol	Antimicrobial, antiparasitic	[34,35]
	Dichloromethane/methanol	Antioxidant	[33]
	Methanol	Antitumor, antimicrobial, antiviral, and immune stimulating	[36]
	Sulfated polysaccharide	Inhibits Japanese Encephalitis Virus (JEV)	[37]
	3-O-β–D–glucopyranosylstigmasta-5,25-diene (6)	Anti-inflammatory	[14,15]
	Methanol	Anti-inflammatory	[38]
U. fasciata	Dichloromethane/methanol	Antiviral	[39]
	N-palmitoyl-2-amino-1,3,4,5-tetyrahydroxyoctadecane (8)	Antiviral	[15]
	Dichloromethane/methanol	Antibacterial	[40]
U. fasciata Delile	Ethanol	Antiproliferative	[41]
	Labda-14-ene-3α,8α-diol( <b>13</b> ) and Labda-14-ene-8α-hydroxy-3-one( <b>15</b> )	Antimicrobial	[11]
	Methanol	Antioxidant	[17]
U. rigida	Diethyl ether and methanol	Antimicrobial	[42]
	Dichloromethane and ethanol	Cytotoxicity	[43]
	Ethanol	Antimutagenic, antioxidant	[10]

Table 1: Biological activities of the genus Ulva.

The green algae belonging to the genus *Ulva* are a group of edible algae that are widely distributed along the coasts of the world's oceans [7,63], and they have an interesting chemical composition that makes their commercial exploitation attractive to produce functional or health promoting food [7,66]. The macroalgae of the order Ulvales are already used in Asia as a food condiment and as a nutritional supplement in Japan, China and other Southeast Asian countries as well as in North and South America and Oceania. For instance, they are consumed as part of the traditional Hawaiian cuisine [67], in Japan, they are included in a variety of dishes such as salads, soups, cookies, meals and condiments as well as a mixed product with other green seaweeds [7,68]. Interestingly, the interest in these algae as a novelty food is expanding in the West [7], and especially in France where they were authorized for human consumption as vegetables [69].

In the recent years, ulvan has been investigated in order to develop functional foods [26] since they cannot be digested in the human gastrointestinal tract [45,47,67,70] and therefore may be regarded as a good source of dietary fiber and a potential source of prebiotics [67]. It is interesting to note that only few species of Ulva have been studied for their application in food industry. These include U. lactuca, U. pertusa, U. compressa and U. clathrata. These macroalgae exhibited a broad spectrum of nutritional composition which makes them excellent candidates for a healthy food for human nutrition [68]. With high levels of protein (between 10 and 25% of dry mass), dietary fiber, low total lipid contents, and relatively high levels of essential amino acids, they constitute a good alternative source of amino acids and of some essential polyunsaturated fatty acids such as Oleic, Linoleic and Linolenic Acids, vitamins and minerals, especially iron [38,59,68,71]. While U. lactuca is used in salads, cookies and soups [62,68], U. pertusa, which is frequently consumed under the name of "aonori" in Japan, has a high level of protein (between 20 and 26% dry product) [63], and abundant with vitamins, trace elements and dietary fibers but with low calorie [55]. The dried U. compressa is used in cooking, particularly with eggs. It is also used as an ingredient in the preparation of high fiber snack "Pakoda", a common Indian product made from chickpea flour. The crude protein levels of this alga are found to be ranging from 21 to 32% [63]. On the other hand, as U. clathrata is a good source of soluble dietary fiber, high-quality polyunsaturated fatty acids, carotenoids and some minerals, it can be a valuable source of protein and may be efficiently used as an ingredient in human and animal foods [7].

# Application of Ulva spp. as Biofilters

Besides their use as food, the macroalgae of the genus *Ulva* can also have applications in the removal of nutrients from effluent waters of sewage, industry and mariculture. Studies showed that some *Ulva* species have been tagged as pollution indicator due to their biomass accumulation when they inhabited in highly polluted waters [2,48,72]. For instance, *U. lactuca* has proven to be a good seaweed biofilter in the treatment of fishpond effluents [73]. The opportunistic growth ability of these seaweeds makes them good candidates for water recycling in integrated invertebrates or fish aquaculture systems and of urban waters [54]. Msuya and Neori [74] have demonstrated that *U. reticulata*, as well as *Gracillaria crassa*, could be cultured and used to remove nutrients from mariculture fishpond effluents by simple facilities and using only water tidal flow [74].

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#### References

- Kong F, Mao Y, Cui F, Zhang X, Gao Z (2011) Morphology and Molecular Identification of Ulva Forming Green Tides in Qingdao China. J Ocean University of China 10: 73-79.
- Wolf MA, Sciuto K, Andreoli C, Moro I (2012) Ulva (Chlorophyta, Ulvales) Biodiversity in the North Adriatic Sea (Mediterranean Italy): Cryptic Species and New Introductions. J Phycol 48: 1510-1521.
- Heech S, Broom JES, Neill KF, Farr TJ, Dalen JL, et al. (2009) Ulva, Umbraulva and Gemina: genetic survey of New Zealand taxa reveals diversity and introduced species. Eur J Phycol 44: 143-154.
- EI-Tawil NE (2010) Effects of Green Seaweeds (Ulva sp.) as Feed Supplements in Red Tilapia (Oreochromis sp.) Diet on Growth Performance, Feed Utilization and Body Composition. J Arabian Aquac Soc 5: 179-193.
- Messyasz B, Rybak A (2010) Abiotic factors affecting the development of Ulva sp. (Ulvophyceae, Chlorophyta) in freshwater ecosystems. Aquat Ecol 45: 75-87.
- http://www.algaebase.org/search/genus/detail/?genus\_id=33&sk=0;%20 accessed%20in%2018.01.2013.%20
- 7. Rodríguez AP, Mawhinney TP, Marie DR, Cruz-Suárez LEC (2011) Chemical

composition of cultivated seaweed Ulva clathrata (Roth) C Agardh. Food Chem 129: 491-498.

- Blunt JW, Copp BR, Munro MHG, Northcote PT, Prinsep MR (2011) Marine natural products. Nat Prod Rep 28: 196-268.
- 9. Amsler CD (2008) Algal Chemical Ecology. Springer, USA.
- Khanavi M, Gheidarloo R, Sadati N, Ardekani MRS, Bagher N, et al. (2012) Cytotoxicity of fucosterol containing fraction of marine algae against breast and colon carcinoma cell line. Pharmacogn Mag 8: 69-64.
- Chakraborty K, Lipton AP, Paulraj R, Viajayan KK (2010) Antibacterial labdane diterpenoids of Ulva fasciata Delile from south western coast of the Indian Peninsula. Food Chem 119: 1399-1408.
- Chakraborty K, Lipton AP, Paulraj R, Chakraborty RD (2010) Guaiane sesquiterpenes from seaweed Ulva fasciata Delile and their antibacterial properties. Eur J Med Chem 45: 2237-2244.
- Flodin C, Whitfield FB (1999) 4-Hydroxybenzoic acid: a likely precursor of 2,4,6-tribromophenol in Ulva lactuca. Phytochem 51: 249-255.
- 14. Awad NE (2000) Biologically Active Steroid from the Green Alga Ulva lactuca. Phytother Res 14: 641-643.
- 15. Gamal AAE (2010) Biological importance of marine algae. Saudi Pharma J 18: 1-25.
- Sun Y, Zhan YC, Sha Y, Pei YH (2007) Norisoprenoids from Ulva lactuca. J Asian Nat Prod Res 9: 321-325.
- Chakraborty K, Paulraj R (2010) Sesquiterpenoids with free radical scavenging properties from marine macroalga Ulva fasciata Delile. Food Chem 122: 31-41.
- Garson MJ (1993) The Biosynthesis of Marine Natural Products. Chem Rev 93: 1699-1733.
- Yildiz G, Celikler S, Vatan O, Dere S (2012) Determination of the Anti-oxidative Capacity and Bioactive Compounds in Green Seaweeds Ulva rigida C Agardh. Int J Food Properties 15: 1182-1189.
- 20. Kerr RG, Baker BJ (1991) Marine Sterols. Nat Prod Rep 8: 465-497.
- 21. Popov SS, Marekov NL, Konaklieva MI, Panayotova MI, Dimitrova-Konaklieva S (1985) Sterols from some black sea ulvaceae. Phytochem 24: 1987-1990.
- 22. Roussis V, Rapti T, Vagias C, Harvala C, Caberi H (2000) Volatile metabolites of the green alga Ulva rigida (C.Ag). J Essent Oil Res 12: 201-204.
- 23. Smit AJ (2004) Medicinal and pharmaceutical uses of seaweed natural products: A review. J App Phycol 16: 245-262.
- 24. Athukorala Y, Jung WK, Vasanthan T, Jeon YJ (2006) An anticoagulative polysaccharide from an enzymatic hydrolysate of Ecklonia cava. Carbohy Polym 66: 184-191.
- Boopathy NS, Kathiresan K (2010) Anticancer Drugs from Marine Flora: An overview. J Oncology doi:10.1155/2010/214186.
- Wijesekara I, Pangestuti R, Kim SK (2011) Biological activities and potential health benefits of sulfated polysaccharides derived from marine algae. Carbohyd Polym 84: 14-21.
- Mohamed S, Hashim SN, Rahman AH (2012) Seaweeds: A sustainable functional food for complementary and alternative therapy. Trends Food Sci Tech 23: 83-96.
- Xu N, Fan X, Yan X, Tseng CK (2004) Screening marine algae from China for their antitumor activities. J Appl Phycol 16: 451-456.
- 29. Cabrita MT, Vale C, Rauter AP (2010) Halogenated Compounds from Marine Algae. Mar Drugs 8: 2301-2317.
- 30. Shalaby EA (2011) Algae as promising organisms for environment and health. Plant Signal Behav 6: 1338-1350.
- Paiva LS, Patarra RF, Neto AI, Lima EMC, Baptista JAB (2012) Antioxidant activity of macroalgae from the Azores. Life Mar Sci 29: 1-6.
- Mao W, Zang X, Li Y, Zhang H (2006) Sulfated polysaccharides from marine algae Ulva conglobata and their anticoagulant activity. J Appl Phycol 18: 9-14.
- El-Baky HHA, El-Baz FK, El-Baroty GS (2009) Natural preservative ingredient from marine alga Ulva lactuca L. Int J Food Sci Technol 44: 1688-1695.

- 34. Kim KM, Hwang IK, Boo JK (2011) A new Agarophyte species Gelidium eucorneum sp. nov. (Gelidiales, Rhodophyta) based on molecular and morphological data. J Phycol 47: 904-910.
- 35. Spavieri J, Kaiser M, Casey R, Wilson AH, Lalvani A, et al. (2010) Antiprotozoal Antimycobacterial and Cytotoxic Potential of Some British Green Algae. Phytother Res 24: 1095-1098.
- Lee D-G, Hyun J-W, KangK-A, Lee J-O, Lee S-H, et al. (2004) Ulva lactuca: A potential seaweed for tumor treatment and immune stimulation. Biotechnol Bioprocess Eng 9: 236-238.
- Chiu YH, Chan YL, Li TL, Wu CJ (2012) Inhibition of Japanese encephalitis virus infection by the sulfated polysaccharide extracts from Ulva lactuca. Mar Biotechnol 14: 468-478.
- Margret RJ, Kumaresan S, Ravikumar S (2009) A preliminary study on the anti-inflammatory activity of methanol extract of Ulva lactuca in rat. J Environ Biol 30: 899-902.
- Mendes GS, Soares AR, Martins FO, Albuquerque MCM, Costa SS, et al. (2010) Antiviral Activity of the Green Alga Ulva fasciata on the Replication of Human Metapneumovirus. Rev Inst Med Trop Sao Paulo 52: 3-10.
- Selvin J, Lipton AP (2004) Biopotentials of Ulva fasciata and Hypnea musciformis collected from the peninsular coast of India. J Mar Sci Technol 12: 1-6.
- 41. Ryu MJ, Kim AD, Kang KA, Chung HS, Kim H, et al. (2013) The green algae Ulva fasciata Delile extract induces apoptotic cell death in human colon cancer cells. In Vitro Cell Dev Biol Anim 49: 74-81.
- Tüney I, Çadirci BH, Unal D, Sukatar A (2006) Antimicrobial Activities of the Extracts of Marine Algae from the Coast of Urla (Izmir, Turkey). Turk J Biol 30: 171-175.
- Medeiros J, Macedo M, Constancia J, LoDuca J, Cunningham G, et al. (1999) Potential anticancer activity for plants and marine organisms collected in the Azores. Açoreana 9: 55-61.
- 44. Lahaye M, Axelos MAV (1993) Gelling properties of water-soluble polysaccharides from proliferating marine green seaweeds (Ulva spp.). Carbohyd Polym 22: 261-265.
- 45. Ray B, Lahaye M (1995) Cell-wall polysaccharides from the marine green alga Ulva "rigida" (Ulvales, Chlorophyta), chemical structure of ulvan. Carbohydr Res 274: 313-318.
- 46. Quemener B, Lahaye M, Bobin-Dubigeon C (1997) Sugar determination in ulvans by a chemical-enzymatic method coupled to high performance anion exchange chromatography. J Appl Phycol 9: 179-188.
- Lahaye M, Inizan F, Vigouroux J (1998) NMR analysis of the chemical structure of ulvan and of ulvan-boron complex formation. Carbohyd Polym. 36: 239-249.
- Lahaye M (1998) NMR spectroscopic characterisation of oligosaccharides from two Ulva rigida ulvan samples (Ulvales, Chlorophyta) degraded by a lyase. Carbohydr Res 314: 1-12.
- 49. Lahaye M, Cimadevilla EAC, Kuhlenkamp R, Quemener B, Lognoné V, et al. (1999) Chemical composition and <sup>13</sup>C NMR spectroscopic characterisation of ulvans from Ulva (Ulvales, Chlorophyta). J Appl Phycol 11: 1-7.
- Robic A, Bertrand D, Sassi J-F, Lerat Y (2009) Determination of the chemical composition of ulvan, a cell wall polysaccharide from Ulva spp. (Ulvales, Chlorophyta) by FT-IR and chemometrics. J Appl Phycol 21: 451-456.
- Stengel DB, Connan S, Popper ZA (2011) Algal chemodiversity and bioactivity: Sources of natural variability and implications for commercial application. Biotechnol Adv 29: 483-501.
- Alves A, Sousa RA, Reis RL (2012) In Vitro Cytotoxicity Assessment of Ulvan a Polysaccharide Extracted from Green Algae. Phytother Res doi: 101002/ ptr4843 [Epub ahead of print].
- Lahaye M, Robic A (2007) Structure and Functional Properties of Ulvan a Polysaccharide from Green Seaweeds. Biomacromolecules 8: 1765-1774.
- 54. Costa LS, Fidelis GP, Cordeiro SL, Oliveira RM, Sabry DA, et al. (2010) Biological activities of sulfated polysaccharides from tropical seaweeds. Biomed Pharmacother 64: 21-28.
- 55. Qi H, Zhang Q, Zhao T, Hu R, Zhang K, et al. (2006) In vitro antioxidant activity of acetylated and benzoylated derivatives of polysaccharide extracted from Ulva pertusa (Chlorophyta). Bioorg Med Chem Lett 16: 2441-2445.

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- 56. Qi H, Zhang Q, Zhao T, Chen R, Zhang H, et al. (2005) Antioxidant activity of different sulfate content derivatives of polysaccharide extracted from Ulva pertusa (Chlorophyta) in vitro. Int J Biol Macromol 37: 195-199.
- 57. Pengzhan Y, Quanbin Z, Ning L, Zuhong X, Yanmei W, et al. (2003) Polysaccharides from Ulva pertusa (Chlorophyta) and preliminary studies on their antihyperlipidemia activity. J Appl Phycol 15: 21-27.
- Toskas G, Hund R-D, Laourine E, Cherif C, Smyrniotopoulos V, et al. (2011). Nanofibers based on polysaccharides from the green seaweed Ulva rigida. Carbohyd Polym 84: 1093-1102.
- 59. Toskas G, Heinemann S, Heinemann C, Cherif C, Hund R-D, et al. (2012) Ulvan and ulvan/chitosan polyelectrolyte nanofibrous membranes as a potential substrate material for the cultivation of osteoblasts. Carbohyd Polym 89: 997-1002.
- 60. Bhakuni DS, Rawat DS (2005) Bioactive Marine Natural Products. Springer, Índia.
- Pereira H, Barreira L, Figueiredo F, Custódio L, Vizetto-Duarte C, et al. (2012) Polyunsaturated Fatty Acids of Marine Macroalgae: Potential for Nutritional and Pharmaceutical Applications. Mar Drugs 10: 1920- 1935.
- Alamsjah MA, Hirao S, Ishibashi F, Fujita Y (2005) Isolation and structure determination of algicidal compounds from Ulva fasciata. Biosci Biotechnol Biochem 69: 2186-2192.
- 63. Pereira L (2011) A review of the nutrient composition of selected edible seaweeds. Nova Science Publishers Inc, New York, USA.
- 64. Fleurence J (1999) Seaweed proteins: biochemical nutritional aspects and potential uses. Trends Food Sci Tech 10: 25-28.
- 65. Gupta S, Abu-Ghannam N (2011) Bioactive potential and possible health effects of edible brown seaweeds. Trends Food Sci Tech 22: 315-326.

- Satpati GG, Pal R (2011) Biochemical composition and lipid characterization of marine green alga Ulva rigida - a nutritional approach. J Algal Biomass Utln 2: 10-13.
- Fitzgerald C, Gallagher E, Tasdemir D, Hayes M (2011) Heart health peptides from macroalgae and their potential use in functional foods. J Agric Food Chem 59: 6829-6836.
- Tabarsa M, Rezaei M, Ramezanpour Z, Waaland JR (2012) Chemical compositions of the marine algae Gracilaria salicornia (Rhodophyta) and Ulva lactuca (Chlorophyta) as a potential food source. J Sci Food Agric 92: 2500-2506.
- Mabeau S, Fleurence J (1993) Seaweed in food products: biochemical and nutritional aspects. Trends Food Sci Tech 4: 103-107.
- Lahaye M, Brunel M, Bonnin E (1997) Fine chemical structure analysis of oligosaccharides produced by an ulvan-lyase degradation of the water-soluble cell-wall polysaccharides from Ulva sp. (Ulvales, Chlorophyta) Carbohydr Res 304: 325-333.
- Yaich H, Garna H, Besbes S, Paquot M, Blecker C, et al. (2011) Chemical composition and functional properties of Ulva lactuca seaweed collected in Tunisia. Food Chem 128: 895-901.
- Largo DB, Sembrano J, Hiraoka M, Ohno M (2004) Taxonomic and ecological profile of 'green tide' species of Ulva (Ulvales Chlorophyta) in central Philippines. Hydrobiol 512: 247-253.
- Neori A, Msuya FE, Shauli L, Schuenhoff A, Kopel F, et al. (2003) A novel three-stage seaweed (Ulva lactuca) biofilter design for integrated mariculture. J Appl Phycol 15: 543-553.
- Msuya FE, Neori A (2002) Ulva reticulata and Gracilaria crassa: Macroalgae That Can Biofilter Effluent from Tidal Fishponds in Tanzania. Western Indian Ocean J Mar Sci 1: 117-126.

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