Headspace solid-phase Microextraction of Volatile and furanic Compounds in Coated Fish Sticks: Effect of the Extraction Temperature

M. Trinidad Pérez-Palacios, Catarina Petisca, Olívia Pinho, Isabel M.P.L.V.O. Ferreira

Abstract—This work evaluated the effect of temperature on headspace solid-phase microextraction of volatile and furanic compounds in coated fish sticks. The major goal was the analysis of the samples as consumed, to reproduce volatile compounds people feel when consuming those products. Extraction at 37 °C (the human body temperature) throughout the HS-SPME analysis of volatile and furanic compounds in coated fish was compared with higher extraction temperatures, which are frequently used for this kind of determinations. The profile of volatile compounds found in deepfried (F) and non-fried (NF) coated fish at 37 and 50 °C was different from that obtained at 80 °C. Concerning furan and its derivatives, an extra formation of these compounds was observed at higher extraction temperatures. The analysis of volatile and furanic compounds in fish coated sticks simulating the cooking and eating conditions can be reliably carried out setting the headspace absorption temperature at 37 °C.

Keywords—analysis of samples as consumed, fish coated sticks, furans, headspace extraction temperature, volatiles.

I. INTRODUCTION

DURING food frying complex processes occur, mainly by lipid oxidation and Maillard reactions, different components react to produce intermediary compounds or volatiles, responsible for the characteristic flavor of fried foods [1]. However, these chemical reactions are also responsible for the formation of undesirable or harmful compounds [1], [2], such us furan and its derivatives.

Furan has been classified as possibly carcinogenic to humans (Group 2B) by the International Agency for Research on Cancer [3], and has been included by the US Department of Health and Human Service [4] in the human pathogen list. However, increased attention is being given to their derivatives, since some of them are considered toxic to animals and humans [5]-[7], thus the quantification of these compounds in cooked foods is of major relevance.

M.T.P-P. Author is with the Tecnología de los Alimentos. Facultad de Veterinaria. Universidad de Extremadura. Avda. de la Universidad s/n, 10071, Cáceres, Spain. (e-mail triny@unex.es).

C.P is with REQUIMTE, Laboratório de Bromatologia e Hidrologia, Departamento de Química, Faculdade de Farmácia, Universidade do Porto, Rua Jorge Viterbo Ferreira 228, 4050-313 Porto, Portugal. (catpetisca@hotmail.com)

O.P. is with Faculdade de Ciências da Nutrição e Alimentação da Universidade do Porto, R Dr Roberto Frias, 4200-465 Porto, Portugal (oliviapinho@fcna.up.pt)

I.M.P.L.V.O.F., is with REQUIMTE, Laboratório de Bromatologia e Hidrologia, Departamento de Química, Faculdade de Farmácia, Universidade do Porto Rua Jorge Viterbo Ferreira 228, 4050-313 Porto, Portugal (isabel.ferreira@ff.up.pt) The solid phase microextraction (SPME) technique has been successfully applied to the analyses of volatile compounds in different types of foods and beverages [8], [9] and it is also appropriate for furan analysis [10], [11]. During the analysis of headspace (HS) by SPME, the effect of some variables, such us type of fiber, sample agitation, sample pH, ionic strength, volume, time and temperature, on the extraction efficiency has been studied in order to determine the optimal conditions [12]-[15]. Several SPME methods use temperatures higher than 50 °C [12], [13], [16]. However, the use of a high extraction temperature can increase the formation of thermally generated compounds, presenting artifacts in the volatile profile [14].

The first method published for furan analysis indicated extraction at 80 °C [17], [18]. In 2006, [19] US FDA decreased the extraction temperature for furan analysis from 80°C to 60 °C, because it was reported that the furan response increased as the equilibration temperature rose [20]. Since then, different extraction temperatures have been used, namely, HS-SPME at 25 °C for furan analysis in different Spanish food [21], fiber exposure temperature at 30 °C for determination of furan in commercial and home-made babyfood [22], and extraction temperature at 50 °C for furan analysis in canned and jarred food [23]. The use of extraction temperatures close to that of human body temperature is recommended since it reproduces the volatile compounds that people feel when the products are being consumed [12]. EFSA claims that furan analysis must be performed in the samples as they are consumed [11], consequently, it is important to check that the HS-SPME extraction temperature does not influence the furan determination in cooked samples [24]. This work aimed to compare the extraction of volatile compounds with a particular detail on furan and its derivatives from HS at 37 °C and at higher extraction temperatures in fried fish coated products and simulating the usual preparation and eating process.

II. MATERIALS AND METHODS

A. Chemicals and standards

Methanol was supplied by Merck (Darmstadt, Germany) and ultrapure water (0.055 μ S/cm) was obtained by using a SeralPur Pro 90 CN system. n-Alkanes were purchased by Sigma–Aldrich (St Louis, MO, USA). The deuterated internal standard d₄-furan (98%) was purchased by Isotec (Ohio, USA). Furan (≥99%) and furfuryl alcohol (99%) were supplied by Sigma-Aldrich (Steinheim, Germany) while 2-furfural was provided by Merck (99%) (Darmstadt, Germany) and 2-pentylfuran (98%) was purchased from Alfa Aesar (Karlsrula, Germany).

Frozen coated fish, with 30 g on average, were obtained from a local store, and labeled as being made of fish (65%; *Merluccius capensis*), wheat flour, water, vegetable oil, salt, spices and natural aroma. Plain sunflower oil used for frying was also from a local recognized brand.

B. Sample preparation

Frozen coated fish samples were divided in two groups. The first group (n=18) was thawed at 4 °C during 16 h and analyzed without being deep-fried (NF). The second group (n=18) was individually deep-fried (F) in sunflower oil using a domestic deep-fryer (Kenwood DF-150; 11) at 180 °C during 4 min, according to the manufacturer recommendations. Each sample was fried alone and immediately analyzed for volatiles. The oil was replaced every 5 frying sessions. After each deep-fried, sticks were slightly drained, and placed on paper towel for removing external oil. In order to simulate the mastication process and study the volatiles release through this process, samples were grinded by using a "masticator shears straight" device (Fig. 1) (Bueno Hermanos, S.A., La Rioja, Spain, ISO 9001-2000 Quality Certified Company), usually used for people unable to masticate. Eight bites were patronized.

Six samples of each group were assayed at three different temperatures, 37 ± 1 °C, 50 ± 1 °C and 80 ± 1 °C.



Fig. 1 Manual device used in this study for grinding the coated fish stick products

C. Volatile compounds analysis

The volatile compounds were determined immediately after frying and grinding, by HS-SPME with GC-MS. Briefly, a sample portion (2 g) was transferred to a 50 ml vial containing 5 ml of water and 3 g of NaCl. The vial was sealed at once and kept at -4 °C during 10 min, followed by sonication (Fungilab, Portugal) during 15 min at room temperature.

A SPME fiber coated with carboxen–polydimethylsiloxane (CAR-PDMS) (75 μ m thickness, Supelco Co., Bellefonte, PA, USA) was used to adsorb the volatile compounds. The SPME fiber was preconditioned at 300 °C for 60 min, in a gas chromatograph injection port, inserted into the sample vial through the septum and exposed to the HS for 40 min at

different temperatures (37 \pm 1 °C, 50 \pm 1 °C and 80 \pm 1 °C) under constant agitation (600 rpm). Thereafter, the SPME fiber was inserted into the injection port and desorbed for 10 min. The injection port was at 280°C, in the split-less mode, with 1 ml min⁻¹ flow Chromatographic analysis was performed using an Agilent 6890 series gas chromatograph (Agilent, Avondale, PA, USA) coupled to a mass selective detector (Agilent 5973). Volatiles were separated using a 5% phenylmethyl silicone (HP-5) bonded phase fused-silica capillary column (Hewlett-Packard, Palo Alto, CA, USA; 33 m x 250 μm i.d., film thickness 0.25 μm), operating with helium at 80 kPa column head-pressure, resulting in a flow of 1 ml min⁻¹ at 40 °C. The oven temperature program was isothermal for 5 min at 40 °C, raised to 135 °C at a rate of 3 °C min⁻¹ and then raised to 220 °C at 20 °C min⁻¹. The transfer line to the mass spectrometer was maintained at 250 °C. Mass spectra were obtained by electronic impact at 70 eV, with a multiplier voltage of 2056 V, collecting data in the fullscan mode at a rate of 1 scan s⁻¹ over the m/z range 30–500. n-Alkanes (Sigma, St Louis, MO, USA) were run under the same chromatographic conditions to calculate the retention indices (RI). Volatile compounds were identified by comparison of their mass spectrum with reference compounds in the NIST 98 data bank (NIST/EPA/NISH Mass Spectral Library, version 1.6, U.S.A.), and by comparison of RI with those described in the literature [8], [16], [25], [26]. Individual volatile compounds were determined in area units, and results of the present study were based on the relative percentages of the chemical families of volatile compounds detected. The furanic compounds were also quantified as described below.

D.Furanic compounds quantification

External calibration curves were used for quantification of furanic compounds. A standard calibration solution containing furan, 2-furfural, furfuryl alcohol and 2-pentylfuran at 8.69, 0.52, 10.84 and 0.07 mg ml⁻¹, respectively, and a d_4 -furan solution at 1 μ g μ l⁻¹ were prepared. Five consecutive dilutions of the standard calibration solution in methanol (1:10 v/v) were made. Portions of 100 µl of each standard solution and a fixed volume (100 μ l) of d₄-furan solution were prepared as the samples, extracted by the SPME fiber and injected into the gas-chromatograph. The m/z used for the quantification of the furanic compounds were m/z 68, m/z 72, m/z 96, m/z 98 and m/z 138 ions for furan, d₄-furan, 2-furfural, furfuryl alcohol, and 2-pentylfuran, respectively. For each individual furanic compound a calibration curve (furanic compound peak area/d₄-furan peak area vs. furanic compound amount/d₄-furan amount) was constructed, obtaining R² values of 0.9999. The final results, expressed in ug g⁻¹, take into account the exact weight of the sample portion in the vial.

E. Statistic analyses

The effect of HS extraction temperature on volatile and furanic compounds of fish coated sticks was analyzed by oneway analysis of variance (ANOVA). Analyses were done by using the SPSS package (v.15.0).

III. RESULTS AND DISCUSSION

A. Profile of volatile compounds in fish coated sticks as a function of the HS extraction temperature

A total of 60 volatile compounds were detected in NF and/or F samples (Table I), being clustered in the following chemical families: aldehydes, alcohols, ketones, aliphatic and aromatic hydrocarbons, esters, furans and pyrazines. Fig. 2 shows the percentage of aldehydes, ketons, furans and pyrazines extracted at 37, 50 and 80 °C by the SPME fiber, in NF and F coated fish sticks, and Figure 3 presents the percentage of esters, alcohols, aromatic and aliphatic hydrocarbons extracted at 37, 50 and 80 °C by the SPME fiber, in NF and F coated fish sticks. Figures 2 and 3 highlight that aldehydes, ketons, furans and pyrazines increased significantly (p < 0.05) from NF to F products whereas esters and aromatic hydrocarbons decreased (p < 0.05) from NF to F products. Aliphatic hydrocarbons and alcohols did not show significant differences between NF and F samples. The changes in volatile composition can be related with i) volatiles taken from the cooking oil; ii) the compounds thermally generated or degraded in the coated fish and oil during frying; iii) the compounds formed as a results of interaction between food and oil compounds at high temperature, namely Maillard products [16].

The effect of the HS extraction temperature on the percentage of each chemical family of volatile compounds was significant, being similar in both NF and F samples (Figures 2 and 3). Values of aldehydes, alcohols, furans, esters, pyrazines and aromatic hydrocarbons showed a significant increase as the HS temperature rose, whereas the content of aliphatic hydrocarbons decreased with the increase of temperature. The percentage of ketons increased from 37 to 50 °C and decreased (in F samples) or maintained (in NF samples) from 50 to 80 °C. As consequence, different profile of volatile compounds in the coated fish sticks was found depending on the HS extraction temperature, i.e. in the F group extracted at 80 °C, aldehydes were the major chemical family, followed by aliphatic hydrocarbons and the rest of chemical groups showing minor percentages, whereas at 37 and 50 °C of HS temperature, the major chemical family was aliphatic hydrocarbons, followed by aldehydes.

TABLE I VOLATILE COMPOUNDS DETECTED IN NF AND/OR F SAMPLES GROUPED BY CHEMICAL FAMILIES

ALDEHYDES	ALCOHOLS
2-methylpropanal	2-methyl-1-butanol
3-methylbutanal	1-pentanol
2-methylbutanal	2-pentanolacetate
hexanal	1-hexanol
2-hexenal	1-butanol-3-methyl, acetate
Heptanal	1-octen-3-ol
2-heptenal	2-ethylhexanol
benzaldehyde	ALIPHATIC HYDROCARBONS
octanal	2-methylpentane
benzeneacetaldehyde	3-methylpentane
2-octenal	hexane
nonanal	heptane
nonenal	2-octene
2-decenal	decane
2,4-decadienal	undecane
2-dodecenal	dodecene
	dodecane
KETONS	ESTERS
2-pentanonone	acetic acid, ethyl ester
2,3-pentanedione	propanoic acid, methyl ester
2-heptanone	butanoic acid, methyl ester
-	· · · · · · · · · · · · · · · · · · ·
*	butanoic acid, 2-
	butanoic acid, 2- methylprotylester
FURANS	butanoic acid, 2- methylprotylester butanoic acid, butyl ester
FURANS furan	butanoic acid, 2- methylprotylester butanoic acid, butyl ester acetic acid, hexil ester
FURANS furan furfural	butanoic acid, 2- methylprotylester butanoic acid, butyl ester acetic acid, hexil ester butanoic acid, 3-methyl, butyl ester
FURANS furan furfural furfurylalcohol	butanoic acid, 2- methylprotylester butanoic acid, butyl ester acetic acid, hexil ester butanoic acid, 3-methyl, butyl ester octanoic acid, methyl ester
FURANS furan furfural furfurylalcohol 2-pentylfuran	butanoic acid, 2- methylprotylester butanoic acid, butyl ester acetic acid, hexil ester butanoic acid, 3-methyl, butyl ester octanoic acid, methyl ester butanoic acid, hexyl ester
FURANS furan furfural furfurylalcohol 2-pentylfuran	butanoic acid, 2- methylprotylester butanoic acid, butyl ester acetic acid, hexil ester butanoic acid, 3-methyl, butyl ester octanoic acid, methyl ester butanoic acid, hexyl ester octanoic acid, ethyl ester
FURANS furan furfural furfurylalcohol 2-pentylfuran	butanoic acid, 2- methylprotylester butanoic acid, butyl ester acetic acid, hexil ester butanoic acid, 3-methyl, butyl ester octanoic acid, methyl ester butanoic acid, hexyl ester octanoic acid, ethyl ester
FURANS furan furfural furfurylalcohol 2-pentylfuran	butanoic acid, 2- methylprotylester butanoic acid, butyl ester acetic acid, hexil ester butanoic acid, 3-methyl, butyl ester octanoic acid, methyl ester butanoic acid, hexyl ester octanoic acid, ethyl ester AROMATIC
FURANS furan furfural furfurylalcohol 2-pentylfuran PYRAZINES	butanoic acid, 2- methylprotylester butanoic acid, butyl ester acetic acid, hexil ester butanoic acid, 3-methyl, butyl ester octanoic acid, methyl ester butanoic acid, hexyl ester octanoic acid, ethyl ester AROMATIC HYDROCARBONS
FURANS furan furfural furfurylalcohol 2-pentylfuran PYRAZINES methylpyrazine	butanoic acid, 2- methylprotylester butanoic acid, butyl ester acetic acid, hexil ester butanoic acid, 3-methyl, butyl ester octanoic acid, methyl ester butanoic acid, hexyl ester octanoic acid, ethyl ester AROMATIC HYDROCARBONS benzene
FURANS furan furfural furfurylalcohol 2-pentylfuran PYRAZINES methylpyrazine dimethylpyrazine	butanoic acid, 2- methylprotylester butanoic acid, butyl ester acetic acid, hexil ester butanoic acid, a-methyl, butyl ester octanoic acid, methyl ester butanoic acid, hexyl ester octanoic acid, ethyl ester AROMATIC HYDROCARBONS benzene methylbenzene
FURANS furan furfural furfurylalcohol 2-pentylfuran PYRAZINES methylpyrazine dimethylpyrazine ethylpyrazine	butanoic acid, 2- methylprotylester butanoic acid, butyl ester acetic acid, hexil ester butanoic acid, 3-methyl, butyl ester octanoic acid, methyl ester butanoic acid, hexyl ester octanoic acid, ethyl ester AROMATIC HYDROCARBONS benzene methylbenzene chlorobenzene
FURANS furan furfurylalcohol 2-pentylfuran PYRAZINES methylpyrazine dimethylpyrazine ethylpyrazine	butanoic acid, 2- methylprotylester butanoic acid, butyl ester acetic acid, hexil ester butanoic acid, 3-methyl, butyl ester octanoic acid, methyl ester butanoic acid, hexyl ester octanoic acid, ethyl ester AROMATIC HYDROCARBONS benzene methylbenzene chlorobenzene 1,3-dimethylbenzene
FURANS furan furfural furfurylalcohol 2-pentylfuran PYRAZINES methylpyrazine dimethylpyrazine ethylpyrazine	butanoic acid, 2- methylprotylester butanoic acid, butyl ester acetic acid, hexil ester butanoic acid, 3-methyl, butyl ester octanoic acid, methyl ester butanoic acid, hexyl ester octanoic acid, ethyl ester acetic acid, hexyl ester butanoic acid, ethyl ester acetic acid,
FURANS furan furfural furfurylalcohol 2-pentylfuran PYRAZINES methylpyrazine dimethylpyrazine ethylpyrazine	butanoic acid, 2- methylprotylester butanoic acid, butyl ester acetic acid, hexil ester butanoic acid, 3-methyl, butyl ester octanoic acid, methyl ester butanoic acid, hexyl ester octanoic acid, ethyl ester octanoic acid, ethyl ester AROMATIC HYDROCARBONS benzene methylbenzene chlorobenzene 1,3-dimethylbenzene ethenylbenzene 1-ethyl,3,5-dimethylbenzene
FURANS furan furfural furfurylalcohol 2-pentylfuran PYRAZINES methylpyrazine dimethylpyrazine ethylpyrazine	butanoic acid, 2- methylprotylester butanoic acid, butyl ester acetic acid, hexil ester butanoic acid, 3-methyl, butyl ester octanoic acid, methyl ester butanoic acid, hexyl ester octanoic acid, hexyl ester octanoic acid, ethyl ester AROMATIC HYDROCARBONS benzene methylbenzene chlorobenzene 1,3-dimethylbenzene ethenylbenzene 1-ethyl,3,5-dimethylbenzene limonene



Fig. 3 Effect of headspace extraction temperature (37 °C; 50 °C; 80 °C) on the percentage of alcohols, aliphatic hydrocarbons, esters, and aromatic hydrocarbons from ◆ non-fried (NF) and ■ deep-fried (F) coated fish samples

80

80 °C

80 °C

80 °C

Fig. 2 Effect of headspace extraction temperature (37 °C; 50 °C; 80

°C) on the percentage of aldehydes, furans, ketons, and pyrazines

from \blacklozenge non-fried (NF) and \blacksquare deep-fried (F) coated fish samples

B. Quantification of furanic compounds in coated fish sticks as a function of the HS extraction temperature

Table II shows the quantity of furan and its derivatives (furan, 2-furfural, furfuryl alcohol and 2-pentylfuran) extracted from NF and F products at 37, 50 and 80 °C. Furan, 2-furfural, and furfuryl alcohol were not detected in NF samples sampled at 37 and 50 °C. Only 2-pentylfuran was found in NF samples at these lower temperatures. At 80 °C 2-furfural, furfuryl alcohol and 2-pentylfuran were quantified in NF. The quantity of 2-furfural, furfuryl alcohol and 2-pentylfuran in F samples increased as the extraction temperature rose, whereas the levels of furan increased from 37 to 50 °C, but it was not detected when the HS extraction temperature was set at 80 °C. These results point out i) the extra formation of furanic compounds with the increase of HS temperature, and ii) the furan decomposition and/or difficulty of extraction at high HS temperatures. Extraction temperatures between 37 and 50 °C reduce these effects. Comparison with results reported in literature is difficult, because contradictory results about influence of temperature on furan extraction are described. In canned food, an increase in the HS extraction temperature from 30 to 50 °C caused an increase in the furan peak area [5], and furan peak area increased exponentially as the equilibration temperature increased from 40 to 80 °C in green coffee, tomato juice and orange juice [20]. But other authors found a decrease in the furan response in canned and jarred food samples when increasing the extraction temperature from 25 to 40 °C [21]. Probably, the effect of the extraction temperature on furan chromatographic peak area depends on the kind of product analyzed. With respect to 2-furfural, furfuryl alcohol and 2-pentylfuran, no studies were found in the literature concerning the influence of the extraction temperature on the content of these furanic compounds.

IV. CONCLUSIONS

Headspace extraction temperature influenced the profile of volatile compounds as well as the quantity of furan and its derivatives in coated fish products analyzed before and after deep-frying. Most volatile compounds increased their response as the temperature rise. Extraction temperatures between 37-50 °C are recommended, extra furanic compounds are formed when extraction is performed at 80 °C. The analysis of volatiles including furan and its derivatives in coated fish sticks products simulating the cooking and eating conditions can be reliably carried out setting the headspace extraction temperature at 37 °C. Furfuryl alcohol was the most abundant furan derivative in fried coated fish products, hazards' associated with its presence in fried foods should be explored.

ACKNOWLEDGMENT

T.P.P. acknowledges funds provided by the Junta de Extremadura-Consejeria de Economía, Comercio e Innovación, and the Fondo Social Europeo for her post-doctoral grant. C.P. wishes to thank FCT the grant SFRH/BD/48745/2008. This work has been supported by Project PTDC/AGR-ALI/101583/2008 and PEst-C/EQB/LA0006/2011.

REFERENCES

- D. S. Mottram, "Flavour formation in meat and meat products: a review", Food Chem., vol. 62, pp. 415-424, Aug. 1998.
- [2] W.W. Nawar, "Volatile components of the frying process", Grasas y Aceites, vol. 49, pp. 271-274, May 1998.
- [3] International Agency for Research on Cancer (IARC), "Dry cleaning some chlorinated solvents and other industrial chemicals" in Monographs on the Evaluation of Carcinogenic Risks to Humans. Lyon, France, 1995.
- [4] US Department of Health and Human Service. 11th Report on Carcinogens Public Health Service, National Toxicology Program, Research triangle park, NC, 2005.
- [5] A. Becalski, S. Hayward, T. Krakalovich, L. Pelletier, V. Roscoe, and E. Vavasour, "Development of an analytical method and survey of foods for furan, 2-methylfuran and 3-methylfuran with estimated exposure", Food Addit. Cont., vol. 6, pp. 764–775, June 2010.
- [6] P.S. Sujatha, "Monitoring cytotoxic potentials of furfuryl alcohol and 2furyl methyl ketone in mice", Food Chem. Toxicol., vol. 46, pp. 286-292, Jan. 2008.
- [7] J.H.E. Arts, H. Muijser, M.J. Appel, K.C. Frieke, J.G.M. Bessems, and R.A. Woutersen, "Subacute (28-day) toxicity of furfural in Fischer 344 rats: a comparison of the oral and inhalation route", Food Chem. Toxicol., vol. 42, pp. 1389–1399, March 2004.
- [8] T. Pérez-Palacios, J. Ruiz, D. Martín, R. Grau, and T. Antequera, "Influence of pre-cure freezing on the profile of volatile compounds during the processing of Iberian hams", J. Sci. Food Agric., vol. 90, pp. 882-890, Apr. 2010.
- [9] B. Ramos, O. Pinho, and I.M.P.L.V.O. Ferreira, "Volatile profile and sensory characteristics of creams used as fillers of bakery products: home made egg creams vs industrial creams", J. Food Sci., vol. 3, pp. 255-268, 2009.
- [10] European Food Safety Authority (EFSA), "Results on the monitoring of furan levels in food", The EFSA Journal, vol. 304, pp. 1-23, 2009.
- [11] European Food Safety Authority (EFSA), "Update of results on the monitoring of furan levels in food", The EFSA Journal, vol. 8, pp. 1702-1720, 2010.
- [12] K. Kaseleht, and E. Leitner, "Optimization of the automated SPME method for the analysis of volatile compounds in traditional Estonian food (Kama and Kvass) using GC/MS", Austrian Food Chem. Conf. WKO, Eisenstadt, Austria. pp. 384-389, May 2008.
- [13] M. M. Mazida, M. M. Salleh, and H. Osman, "Analysis of volatile aroma compounds of fresh chilli (Capsicum annuum) during stages of maturity using solid phase microextraction (SPME)", J. Food Comp. Anal., vol.18, pp. 427-437, Aug. 2005.
- [14] J. Ruiz, R. Cava, J. Ventanas, and M. T. Jensen, "Headspace solid microextraction for the analysis of volatiles in a meat product: dry-cured iberian ham", J. Agric. Food Chem., vol. 46, pp. 4688-4694, Nov. 1998.
- [15] O. Pinho, C. Peres, and I.M.P.L.V.O. Ferreira, "Solid-phase microextraction of volatile compounds in "Terrincho" ewe cheese: Comparison of different fibers", J. Chromatog. A, vol. 1011, pp. 1-9, Sept. 2003.
- [16] R. Ramírez, M. Estévez, D. Morcuende, and R. Cava, "Effect of the type of frying culinary fat on volatile compounds isolated in fried pork loin chops by using SPME-GC-MS", J. Agric. Food Chem., vol. 52, pp. 7637-7643, Decemb. 2004.
- [17] US Food Drug and Administration (US FDA), "Furan in foods-Thermal treatment", Washington, DC, 2004, http://www.cfsan.fda.gov/~lrd/fr040510.html.
- [18] European Food Safety Authority (EFSA). Report of the scientific panel on contaminantes in the food chain on provisional findings on furan in food. The EFSA Journal, 137, 1-20. 2004.
- [19] US Food Drug and Administration (US FDA), "Determination of furan in foods", Washington, CD, 2006, http://www.cfsan.fda.gov/~dms/furan.html.
- [20] H. Z. Senyuva, and V. Gokmen, "Analysis of fruan in foods. Is headspace sampling a fit-for-purpose technique?" Food Addit. Cont., vol. 22, pp. 1198-1202, Aug. 2005.
- [21] M. S. Altaki, F. J. Santos, and M. T. Galceran, "Analysis of furan in foods by headspace solid-phase microextraction-gas chromatographyion trap mass spectrometry", J. Chromatog. A, vol. 1146, pp. 103–109, Jan. 2007.

- [22] F. Bianchi, M. Careri, A. Mangia, and M. Musci, "Development and validation of a solid phase micro-extraction–gas chromatography–mass spectrometry method for the determination of furan in baby-food", J. Chromatog. A, vol. 1102, pp. 268–272, Octob. 2006.
 [23] T. Kim, S. Kim, and K-G. Lee, "Analysis of furan in heat-processed
- [23] T. Kim, S. Kim, and K-G. Lee, "Analysis of furan in heat-processed foods consumed in Korea using solid phase microextraction–gas chromatography/mass spectrometry (SPME–GC/MS)", Food Chem., vol. 123, pp. 1328–1333, June 2010.
- [24] C. Crews, and L. Castle, "A review of the occurrence, formation and analysis of furan in heat-processed foods", Trends Food Sci. & Technol., vol. 18, pp. 365-372, July 2007.
- [25] E. Muriel, T. Antequera, M. J. Petrón, A.I. Andrés, and J. Ruiz, "Volatile compounds in Iberian dry-cured loin", Meat Sci., vol. 68, pp. 391-400, Nov 2004.
- [26] M. L. Timón, A.I. Carrapiso, A. Jurado, J. Lagemaat, "A study of the aroma of fried bacon and fried pork loin", J. Sci. Food Agric., vol. 84, pp. 825-831, June 2004.