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Cellular Polyamine Catalogues of the Five Classes of the Phylum Proteobacteria: Distributions of Homospermidine within the Class Alphaproteobacteria, Hydroxyputrescine within the Class Betaproteobacteria, Norspermidine within the Class Gammaproteobacteria, and Spermine within the Classes Deltaproteobacteria and Epsilonproteobacteria

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Abstract : Cellular polyamines extracted from reclassified or newly validated 47 alphaproteobacteria, 46 betaproteobacteria, 96 gammaproteobacteria, 12 deltaproteobacteria and 10 epsilonproteobacteria were analyzed by high-performance liquid chromatography. Homospermidine was widely distributed within the class Alphaproteobacteria, however, homospermidine-dominant type, spermidine-dominant type and homospermidine/spermidinedominant type were found and the three triamine profiles were genus-specific. The all genera belonging to the class Betaproteobacteria, ubiquitously contained putrescine and 2hydroxyputrescine. Triamines were absent in almost betaproteobacteria. Many genera, including psychrophilic species, of the class Gammaproteobacteria, contained putrescine and spermidine as the major polyaminenes. Diaminopropane and norspermidine were selectively distributed in several genera of the class Gammaproteobacteria. Spermidine was the major polyamine in the classes Deltaproteobacteria and Epsilonproteobacteria. Spermine was found in some thermophiles within Betaproteobacteria, Deltaproteobacteria and Epsilonproteobacteria, suggesting that the occurrence of spermine correlate to their thermophily. Additional these polyamine catalogues serve for the classification of the phylum Proteobacteria, as a chemotaxonomic marker.

Key words : Homospermidine, Hydroxyputrescine, Norspermidine, Polyamine, Proteobacteria, Spermidine, Spermine

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### INTRODUCTION

The phylum Proteobacteria (formerly the class Proteobacteria) of the domain Bacteria contains the five classes (formerly subclasses) Alphaproteobacteria, Betaproteobacteria, Gammaproteobacteria, Deltaproteobacteria and Epsilonproteobacteria<sup>1, 2)</sup>. Diamines such as diaminopropane (1,3-diaminopropane), putrescine, cadaverine and hydroxyputrescine (2hydroxyputrescine), triamines such as spermidine, norspermidine, homospermidine, hydroxyspermidine (2-hydroxyspermidine) and acetylspermidine  $(N^{1}$ acetylspermidine), a triamine, spermine, and a guanidinoamine, agmatine, have been found as their cellular polyamine components. Our studies on polyamine distribution profiles within proteobacteria already provided valuable chemotaxonomic informations on their classifications<sup>3-12)</sup>. Afterward, many new members of proteobacteria have been isolated and validated in Bergey' Manual of Systematic Bacteriology<sup>1)</sup>, National Centre for Biotechnology Information (NCBI)<sup>2)</sup> and Validation Lists of International Journal of Systematic and Evolutionary Microbiology<sup>13)</sup>.

As shown in the previous studies, a novel triamine, homospermidine, was selectively distributed in the class Alphaproteobacteria and a group of the class Gammaproteobacteria within Proteobacteria<sup>6, 9, 11, 14-16)</sup>. Homospermidine distribution in the reclassified and 47 newly validated alphaproteobacteria (belonging to 39 genera), divided into five orders, were detarmined to elucidate variations in their polyamine profiles, and to evaluate the usefulness of polyamine pattern as a phenotypic marker.

A novel hydroxydiamine, hydroxyputrescine, widespread within the class Betaproteobacteria<sup>6, 8, 14,</sup> <sup>17)</sup>. Its distribution was limited in betaproteobacteria within Proteobacteria. To determine the occurrence of hydroxypolyamines, cellular polyamines of the reclassified and 46 newly validated betaproteobacteria belonging to 21 genera were analyzed.

Norspermidine was widespread within the gammaproteobacteria belonging to the order Vibrionales of the class Gammaproteobacteria<sup>6, 7, 10, 18, 19)</sup>. This uncommon triamine was found in this class and it has never been detected in other four classes

within Proteobacteria. Polyamine analyses were carried out in the reclassified and 96 newly validated gammaproteobacteria belonging to eight orders (42 genera) including various marine psychrophiles.

Within the class Deltaproteobacteria, some myxobacteria contained homospermidine, however, it was absent in the other deltaproteobacteria containing spermidine<sup>6, 12)</sup>. Spermidine was the major polyamine in the members of the class Epsilonproteobacteria, previously analyzed<sup>6, 12)</sup>. To survey spermine distribution in thermophilic proteobacteria, polyamines of 12 newly published deltaproteobacteria including a novel thermophile and 10 newly published epsilonproteobacteria including five novel thermophiles were analyzed.

#### MATERIALS AND METHODS

The proteobacteria were grown at the optimum growth temperature in the media designated by the culture collections (listed in Tables 1-4) as shown by Medium No., Marine broth (MB) (DIFCO Ltd., Detroit, USA), Nutrient broth (NB) (Nissui Pharmaceutical Co. Ltd., Tokyo, Japan), NB supplemented with 5% NaCl (NB-NaCl), Trypticase soy broth (TSB) (BBL, Bectone Dickinson and Company, Cockeysville, MD, USA), Brain heart infusion broth (BHIB) (Nissui), yeast extract-mannitol medium (YM), peptone-yeast extract medium (PY) and PY dissolved in seawater (PYSW). Polyamine-free synthetic 199 medium (Nissui Pharmaceutical Co. Ltd), 199 medium supplemented with 5% NaCl (199NaCl) and 199 medium dissolved in seawater (199SW) were also used. Organisms at the stationary phase were harvested by centrifugation. The pellets were homogenized in 0.5 M perchloric acid (HClO<sub>4</sub>). Polyamines were extracted into HClO<sub>4</sub> and analyzed by high-performance liquid chromatography (HPLC) on a Hitachi L6000 High-Speed Liquid Chromatograph<sup>20)</sup>. Estimated cellular concentrations of polyamines are listed in Tables 1, 2, 3 and 4.

### **RESULTS AND DISCUSSION**

Class Alphaproteobacteria (Table 1)

Order Sphingomonadales

The wide distribution of homospermidine in this order has been known. Homospermidine was found in

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Organism Medium.	Temp(°C)	д	olyamin	uπ) ou	ol/g we	I wr cell	ä			08-66I	·		ı	о ,	្ត	•	
	1	deC	Pul Ca	id Si	H H	Spd Bi	pm A	ця	Erythrobacter aquimaris JCM 12189 <sup>1</sup>	MB-30	•			0,74			
Order Sphingomonadales									Erythrobucter inteolus JCM 12599 <sup>T</sup>	MB-30	ŀ	,	ī	1.06		,	
Sphingomonas soli IAM 15213 <sup>1</sup> NB-3			,	0.6	.0 86	€.	о	10	Erythrobacter litoralis IAM 14332 <sup>T</sup>	MB-30°			ī	0.95	,		
(Sphingomonas composta)									Erythrobucter langus IFO 14126 <sup>7</sup>	MB-30°	,		,	0.43		•	
Splingomonas oligophenolica JCM 12082 <sup>7</sup> NB(1/10	- 02-((		•	'	Ι.(	5			Perphyrobacter sanguineus IAM 12620 <sup>T</sup>	NB-24	'	0.34		0.85		0.02	
Sphingomonas yabuuchiae JCM 11416 <sup>T</sup> 7SB-	30 .	•	ı	ö	0 0	8			(Agrobacterium sanguineum)								
Sphingemonus azotifizens IFO 15497 <sup>1</sup> PY-30	°.	•		,	1.2	' g	'		Shinella granuli JCM 13254 <sup>T</sup>	JCMB46-30	ŀ	0.25	ö '	75 0.67	,		
Sphingomonas trueperi IFO 16157 <sup>1</sup> PY-3(		'		'	1.1	- 01	'		Shinella zoogloeoides IAM 12669 <sup>1</sup>	199-30	,	0.01		0.54	8	. 20	
Sphingomonas adhaesiva IFO 15099 <sup>1</sup> PY-3	30°	•	,	ı	1.5	' 8			(CYabireella sacharophila, Zoogloea ramigera)	0							
Sphingomonas asaecharolytica IFO 15499 <sup>7</sup> PY-3	30°		•		1	30			Order Rhodobacterales								
Sphingomonas echinoides IFO 15742 <sup>7</sup> PY-31	0		•	,	1	- 91	'		Dinoroseobacker shibae NCIMB 14021 $^{\mathrm{T}}$	06-WS661	•	0.30		0.05 0	ম	'	
Sphingomonas mati IFO 15500 <sup>T</sup> PY-30		·	'		0.	- 51	•			MB-30		0.60		.70 0.0	ۍ.		
Sphingomonas parapaucimobilis IFO 15100 <sup>7</sup> PY-3	30" -		•	_	i، '	35			Paracoccus denitrificans IAM 12475 <sup>T</sup>	199-30 <sup>4</sup>	•	0.49	0,02	1.50			
Sphingomonas paucimobilis IFO 13953 <sup>T</sup> PY-3		•	:	1	2.4	' 20	•		Paracoccus thiocyanatus IAM 128167	NB-30 <sup>c</sup>	0.05	1.45	0.28 2	Ŗ	ç	8	
Sphingonunas pruni IFO 15498 <sup>T</sup> PY-2	30°		,		2.1	י גז	4		Paracuccus versums IFO 14567 <sup><math>r</math></sup>	NB-30 <sup>6</sup>	'	0.28		0.34			
Sphingomonas sanguinis IFO 13937 <sup>1</sup> PY -3.		,	'			- 16	1		Paracoccus alkenifer NCIMB 13527 <sup>n</sup>	NB-30°	•	1.45	0.02	20		•	
Sphingemenas roseifiava IAM 14823 <sup>T</sup> IAMB.	38-25		•		, ,	20	1		Paracoccus pantotrophus ATCC $35512^{T}$	NB-30 <sup>b</sup>	ı	1.60	0.49	2.05	,		
Sphingomonus aquatilis IFO 16141 <sup>T</sup> PY-	-30		1	- 13	0	'	Ċ,	R	Paracoccus alcaliphilus JCM 7364 <sup>°</sup>	199-30 <sup>4</sup>	ı	2.40	0.05	10	ð	0.0 10	÷
Sphingomonas koreensis IFO 16142 <sup>T</sup> $PY \leq$	30'			0	8	•	õ	2	Paracoccus aminophilus JCM 7686 <sup>r</sup>	199-304	,	0.52	0.04	8	0.0	0.0	
" Sphingomonus atotofornuns" IAM 15131 NB-	-30			ć ·	<b>8</b>		0	50	Purucoccus aminovorans JCM 7686 <sup>1</sup>	199-30 <sup>¢</sup>	,	0.88	0.30 1	. 19	,	,	
Sphingobium chungbukense JCM 11454 <sup>1</sup> JCM7;	5-30 -	•	•	1.4	5	'	0.1.	K	Puracoccus kocurii JCM 7684 <sup>2</sup>	199-30 <sup>4</sup>	ī	0.38 (	0.41 3.	10	0.0	N 0.52	~
Sphingobium aniense IAM 15006 <sup>T</sup> NB-3	30		,	0 -	8		0	22	Paracoccus seriviphilus NBRC 100798 <sup>1</sup>	MB-30	0.03	0.26	0.60 C	175	'	,	
691	-30			0 -	5		ő	05	Paracoccus zeazanthinifaciens MBIC 03966	<b>MB-25</b>	ľ	0.33	0.14	1.66	ļ	•	
Sphingobium japonicum NBRC 101211 PY -:	30			-	2	•	I		Paracoccus koreensis IAM 15216 <sup>T</sup>	NB-30		0.75	0.08	040		•	
Sphingopyxis taejonensis IFO 16144 <sup>1</sup> PY-3	- 00	0	. 40.0	-	37 -	,	0.0	_	Pelagibuca bernudensis JCM 13377 <sup>t</sup>	MB-25		0.70	0.49 2	.45	,	•	
(Sphingomonus taejonensis)									Catellibacteriun nectariphilum NBRC 10X)46	<sup>T</sup> NBRC388-3		1.40	,	6		'	
Novosphingobium pentaramutivorans JCM 12182 <sup>r</sup>	MB-30				1.30				Jannaschia cystaugen: NBRC 100362 <sup>v</sup>	MB-25	•	0.45	0.10	1,43			
Novospiungobtum taihuense JCM 12465 <sup>°</sup> JCM	1443-30	,	ī		1.20	,			Pseudovihrio denitrificuns JCM 12308 <sup>T</sup>	MB-30	,			0.60	,		
Novosphingobium tardaugens JCM 114347 <sup>T</sup> JC)	M12-25		0.02	1,45	0.75				Pseudovibrio ascidiaceicola IAM 15084 <sup>T</sup>	MB-30	,	0.10	,	0.89	•	0.07	
Sphingosinicella microcystinivorans 3CM 13185 <sup>T</sup> NI	B-30	,		,	,	1.15			Rosevarius tolerans IAM 14840 <sup>7</sup> IAMB	[4]SW-20	ī	0.25	0.80	. 42 -	•	•	
									Pseudorhodobacter ferrugineus IAM 12616 <sup>t</sup>	0E-WS661	•	0.33	,	08/1		•	

0.02 -

0.85

- 0.34 -

MB-24<sup>6</sup>

(Agrobacterium ferrugineum)

Roseibacterium elongatum JCM 11220 <sup>T</sup>	NB-27	,	0.15 0.	35	0.82			
Yungia pacifica JCM 12573 <sup>T</sup>	MB-30		0.10 0	0.08	0.44	,		ī
Order Rhodospirillales								
Defluvicoccus vanus NCIMB 13612 <sup>T</sup>	VCIMB481-30				0.80	,	,	ı
Saccharibacter floricola JCM 12116 <sup>T</sup>	JCM97-28	0.02	0.05		1.40		ı.	•
Azospirillum oryzae IAM 15130 <sup>7</sup>	NB-30		88	,	036 0	5		
Azospirilium irakense ATCC 51182 <sup>T</sup>	199-30	-	45	0	0.10 2.	32		
Azospirilium amazonense ATCC 35119 <sup>1</sup>	199-30*	0 ,	- 15	0	0.02	22		,
Azospirillum hatopraeferens $ATCC$ 43709 <sup>T</sup>	199-30*	,	. 15		0.05	8		
Azospiritum brasilense JCM 1224 <sup><math>T</math></sup>	NM-30°	Ċ -		0	23 0.	- 5		
Azospiritium lipoferm JCM 1247 <sup>r</sup>	NM-305	0.0	ч.	ð	08 0.6	•	•	
Tistrella mobilis ${ m IAM}~14872^{ m T}$	06- YY	Э -	06 0.5	2	96 1.1	نہ •		
Kozakia baliensis JCM 11301 <sup>1</sup>	JCM90-30	0 -	3	0	- 08'0	•		
Asaia krungthepensis NBRC 100057 <sup>1</sup>	PY G-30		. ÓE.		1.40			
Neoasaia chiangmaiensis NBRC 101099 <sup>T</sup>	PY G-30	,	383		8.0		,	·
Gluconobacter thailandicus JCM 12310 <sup>°</sup>	JCM97-30		0.46		1.50		•	•
Acetobacter cibinongensis JCM 11196 <sup>t</sup>	JCM97-30	Ĵ	101		0.98			
Acetobacter orientalis JCM 11195 <sup>r</sup>	JCM97-30		0.10		1,12			•
Acetobacter syzygii JCM 11197 <sup>r</sup>	JCM97-30	0	•		1.05			
Order Rhizobiales								
Methylocapsa acidiphila NCIMB 13765 <sup>1</sup>	NCIMB472-30		•	'	0.70	ı	•	,
Kaistia adipata IAM 15023 $^{ m T}$	NB-30			1	0.46		,	•
Rhodopseudomonas fuecalis JCM 11668 <sup>1</sup>	JCM358-30	٠	Ņ		,	1.10	,	ı
Rhodopseudortonas palustris NCIMB 8252 <sup>1</sup>	NCIMB27-30 <sup>b</sup>	•	0.02	1	0.10	130	ī	1
Rhodopseudomonas acidophila NCIMB 117	61 <sup>T</sup> NCIMB153-30 <sup>b</sup>	'	0.15	,		2.40	٠	·
Pseudaminobacter defluvii IAM 12817 <sup>r</sup>	IAMB60-30	1	6610	•	1.43	1.02	,	•
Roseomonas lacus JCM 13283 <sup>1</sup>	JCM346-30		0.10		1.50	1.10		٩
Hoeftea phototrophica NCIMB 14078 <sup>T</sup>	MB-30	۲		1	1.15		•	
Order Caulobacterales								
Asticcucaulis tuihuensis JCM 12463 <sup>T</sup> J	CM443-30			0.10	06.0	0.40		0.20
Brevundimunan nasdae JCM 11415 <sup>T</sup>	TSB-30		ı	•	0.6	•	1	1
Brevundinumas bacteroides NCIMB 9790 <sup>T</sup>	NCIMB58-30 <sup>b</sup>		'	'	0.2(	0 1.25		1
Brevundimonas subvibrioides IFO 16660 <sup>°</sup>	199-30*	3	'	٠	,	140 1	,	'

Brevundimonas bullata IAM 13153 <sup>T</sup>	199-30 <sup>b</sup>	٠	'	٢	2.20	1.80		'
Brevundimonas diminuta IFO 12697 <sup>1</sup>	199-30*	•				0.65		•
Brevundimonas vesicularis IFO 12165 <sup>T</sup>	199-30 <sup>b</sup>	'	'	,	1.17	0.20		,
Phenylobacterium koreense LAM 15119 <sup>T</sup>	NB-30	•		•	1.10			1
incertae sedis								
Pleonwrphomonas oryzae IAM 15079 <sup>7</sup>	NB-30	•	0.60	,	0.06	0.81	,	'
Terasakiella pusilla IFO 13613 <sup>1</sup>	-0E-MS661	•	0.24	,	1.19		٠	,
(Oceanospirittura pusittum)								
Kordiimonas gwangyangensis JCM 12864 <sup>1</sup>	MB-30	'	0.70		0.35	0.20		'

Dap, diaminoprupane; Put, putrescine; Cad, cadaverine; Spd, spermidine; HSpd, homospermidine; Spm, spermine;
Agen, agmaine; -, not detected (<0.005); <sup>7</sup>, Type strain; IAM, Institute of Molecular and Cellular Biosciences, The University of Tokyo, Tokyo, Japan, JCM, Japan Collection of Microorganisms, RIKEN, Wako, Sattama, Japan; IFO, Institute for Fermenation, Oaska, Japan; NERC, Biotogical Resource Center, National Institute of Technology and Evaluation, Kisurazu, Japan; MBIC, Marine Biotechnology Institute Culture Collection, Kanaishi, Iwate, Japan; IFO, Fordandion, Kisurazu, Japan; MBIC, Marine Biotechnology Institute Culture Collection, Kanaishi, Iwate, Japan;
ATCC, American Type Culture Collection, Manassas, Virginia, USA; NCIMB, National Collections of Industrial, Foods and Marine Bacteria, Scotland, UK. Fonner names are shown in parentheses. Quotation narks indicate that the scientific name has not been validly published. \* was cited from Hamana *et al.*, 1994, \* Hamana *et al.*, 2001, <sup>e</sup> Hamana & Takeuchi, 1998, <sup>a</sup> Hamana & Matsuzaki, 1992, \* Takeuchi *et al.*, 2001, <sup>f</sup> Hamana *et al.*, 2003, <sup>b</sup> Hamana & Matsuzaki, 1990, <sup>b</sup> Hamana & Matsuzaki, 1993, <sup>b</sup> Manuau

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the newly analyzed three Sphingomonas species as well as Sphingosinicella microcystinivorans<sup>21</sup>, in the present study. The major polyamine was spermidine in Sphingomonas azotoformans, Sphingobium chungbukense, Sphingobium amiense, Sphingobium japonicum, Sphingopyxis taejonensis and three Novosphingobium species. Sphingomonas soli and two Shinella species contained spermidine and homospermidine as the major polyamines, indicating a unique triamine profile within this order. Other 13 authentic Sphingomonas species contained homospermidine. It has been reported that three Sphingobium species, six Novosphingobium species and three Sphingopyxis species contained spermidine<sup>22)</sup>. New *Erythrobacter* and *Porphyrobacter* species contained spermidine alone. These results suggest that the distribution of homospermidine in this order is genus-specific, however, are chemotaxonomically conflicting with the classification of some Sphingomonas species.

### Order Rhodobacterales

Dinoroseobacter shibae contained spermidine and homospermidine. Spermidine was the major polyamine in the new alphaproteobacteria located in the genera, Paracoccus, Pelagibaca, Catallibacterium, Jannaschia, Pseudovibrio, Roseovarius, Pseudorhodobacter and Yagia species, analyzed in the present study.

### Order Rhodospirillales

Homospermidine and spermidine were found in *Azospirillum* and *Tistrella*, whereas spermidine was detected in *Defluvicoccus*, *Saccharibacter*, *Kozakia*, *Asaia*, *Neoasaia*, *Gluconobacter* and *Acetobacter* species, analyzed in the present study.

### Order Rhizobiales

Methylocapsa acidiphila, Kaistia adipata and Hoeflea phototrophica contained spermidine alone. Rhodopseudomonas faecalis contained homospermidine alone. Pseudaminobacter defluvii and Roseomonas lacus contained spermidine and homospermidine. Three types on triamine profile were observed in this order.

## Order Caulobacterales

Asticcacaulis taihuensis contained spermidine, homospermidine and agmatine. *Phenylobacterium koreense* contained spermidine alone. Distribution profiles of spermidine and homospermidine are heterogeneous in the six *Brevundimonas* species. Others

Homospermidine was distributed in the newly validated *Pleomorphomonas* and *Kordiimonas* species, however, their phylogenetic positions within the class Alphaproteobacteria are unknown. The major triamine in *Terasakiella pusilla* was spermidine.

### Class Betaproteobacteria (Table 2)

It has been demonstrated that 66 betaproteobacteria, consisting the six orders Burkholderiales, Hydrogenophilales, Methylophilales, Neisseriales, Nitrosomonadales and Rhodocyclales, ubiquitously contained hydroxyputrescine in addition to putrescine<sup>6, 8)</sup>. Hydroxyputrescine has not been detected in almost other four classes of the phylum Proteobacteria. Therefore, it has been proposed that the occurrence of this hydroxydiamine served as a chemotaxonomic marker for the class Betaproteobacteria.

Azohydromonas, Burkholderia, Caldimonas, Cupriavidus, Comamonas, Curvibacter, Delftia, Derxia , Diaphorobacter , Herbaspirillum , Hydrogenophaga, Hylemonella, Janthinobacterium, Malikia, Mitsuaria, Ottowia, Pelomonas, Ralstonia and Wautersia belonging to Burkholderiales, Zoogloea and "Pseudomonas butanovora" belonging to Rhodocyclales, Hydrogenophilus belonging to Hydrogenophilales, and Alysiella, Conchiformibium, Bergeriella, Simonsiella and Microvirgula belonging to Neisseriales, analyzed in the present study, ubiquitously contained putrescine and hydroxyputrescine as the major polyamines.

Hydroxyputrescine was found also in *Thiobacter* subterraneus, Sivamonas terrae and "Pseudomonas jianii" classified into this class. Spermidine was found in thermophiles and some mesophilic species analyzed in the present study, however, phylogenetic significance of this triamine was not clear. Hydroxyspermidine was produced by aminopropyltransfer into hydroxyputrescine, and coexisted with hydroxyputrescine in the betaproteobacteria containing spermidine as a major 
 Table 2. Cellular concentrations of polyamines of betaproteobacteria

Oreanism	Medium-Temp(°C)		olvamine	(π mol/)	t wet wi	cell):		Herbaspirilium seropedicae ATCC 35892 <sup>7</sup> ATCC8838	8-20	- 1.35	2,10			•	
3	•	Dap	H-Put F	ut Cad	H-Sp	d Spd	Som	Herbasalrillum chiwvohenolicum IAM 15024 <sup>7</sup> NB-25	5	- 0.05	0.44		,		,
Order Burkholderiales							.	Herbaspirillum huttiense IAM 14941 <sup>T</sup> NB-25	ž	- 0.35	1.40		,	,	,
Burkholderia kururiensis JCM 10599 <sup>T</sup>	NB-30	'	1.75	0.52 0.	5	0.12	,	(Pseudomdonas huttiensis)							
Burkholderiu glathei JCM 10563 <sup>T</sup>	NB-37	ı	0.45	0.04	•	0.06	ı	Herbuspiritium puteus IAM 15032 <sup>T</sup> NB-25	25	- 0.38	0.80	,	,	1	
Brukholderia sordidicola JCM 11778 <sup>1</sup>	TSB-30		0.15	- t <b>9</b> :0	ľ		ı	Herbaspirillum frisingense IAM 14974 <sup>T</sup> NB-30	30	- 0.55	66.0	1		0.01	,
Ralstonia manniolilytica JCM 11284 <sup>T</sup>	TSB-30		0.44	- 15	'	ı		Herbuspirilium rubrisubalbicans IAM 14876 $^{T}$ BHIB-	3-25	- 0.15	0.35				,
Ralstanía picketti JCM 10171 <sup>T</sup>	NB-30 t	ŀ	0.70 1.	30 -		ŀ	•	Curvibucter graciils IAM 15033 <sup>T</sup> NB-25	2	05.0	1.22		,		,
Cupriavidus eutropha IAM 13533 <sup>T</sup>	199-30		64 04 04	. 02.5		•	,	Curvibacter lanceolatus IAM 14947 <sup>T</sup> MB-25	5	- 0.14	1.03	'		,	,
(Ralstoria entropha, Wantersia entropha)	(1							(Pseudomonas lanceolata)							
Cupriavidus oxalaticus IFO 13593°	NB-30	•	0.70	0.65 0.0	2	ı	ı	Curvibacter delicatus IFO 14919 <sup>T</sup> PY-30	30°	- 0.02	0.58			1	,
(Ratrionia ozalatia, Wantersia ozulatica)								(Aquaspirillum delicatum)							
Cupriavidus gilardii JCM 11283 <sup>1</sup>	TSB-30		0,40 (	0.75 -	,	٢	ı	Comanonas budia IAM 14839 <sup>T</sup> NB-2	-25	- 0.42	0.56	0.03	,	0.19	4.
(Rakstonia gilardii, Wautersia gilardii)								Commonas terrigena JFO 132997 199-3	30	- 0.81	0.35	ī	ŀ	,	
Cuprisvidus pauculus JCM 11286 <sup>7</sup>	TSB-30		0.52	- 01.1	ı	ľ	,	Comanonas testosteroni IAM 12419 <sup>1</sup> [99-30	30*	0.32	0.95		,	0.02	
(Raistonia pauculu, Wautersia panaula)								Malikia spinosa IAM 14918 <sup>1</sup> NB-2	-25	- 0.37	1.26	0.02		0.10	
"Wautersia metadolorelans" IAM 14785	5 NB-30	'	1.12 0	- 997	,	•	ı	(Pseudomonas spinosa, Hydrogenophuga spinosa)							
( "Ratstonia metalophila")	06-661	•	0.77	0.26	ı	'	•	Pelomonas saccharophila IAM 14368 <sup>1</sup> NB	8-30	- 0,4	9 1.02		٠	,	•
"Wautersia silverii" IAM 14786	NB-30	,	0.62 0	- 48				(Pseudomonus saccharophila)							
("Ratstonia silverii")	199-30		0.50 0.	32 -	·	,	,	Janthinobacterium tividum IAM 13948 <sup>T</sup> 199-2	-25	1.74	2.50		·		
"Wautersia tsushimaenvis" IAM 14787	NB-30		0.65 (	137 -		'		Janthinobacterium agaricidamnosum [AM 14973 <sup>1</sup> NE	(B-25	- 0.15	S 0.96	,	,	•	
( "Rakstonia tsushimae" )	199-30	•	0.70	.46	•	•		Hydrogenophuga flava IAM 14931 <sup>1</sup>	18-30	- 0.65	5 1.00	,	3	,	•
Delftia acidovoruns IAM 12409 <sup>r</sup>	PY-30		0.30 1	. 55	1	ı.		Hydrogenophaga intermedia IAM 14919 <sup>T</sup> NB	B-27	- 0.1	5 0.40	·		0.03	•
Delftia tsuruhatensis NBRC 16741 <sup>T</sup>	PY-30	1	0.42 2	20	'	ı		Hydrogenapliaga palleronii IAM 14930 <sup>T</sup> NB.	B-30	1.1	5 1.90	•		,	
Diaphorobacter nitroreducens JCM $11421^{T}$	<sup>T</sup> JCM22-30	ı	0.16	- 58.1	'	ı	,	Hydrogenophaga pseudofuwa IAM 14928 <sup>1</sup> NB	B-37	- 0.4	15 0.97	'	ľ	ī	,
Caldimonus manganoxidans JCM 10698 <sup>7</sup>	JCM74-45		0.49 0	- 5	•	'	L	Hydrogenophaga taentospirutis IAM 14929 <sup>n</sup> NB	B-30	- 0.4	0 1.28			ī	,
Ottowia thiouxydaus JCM 11629 <sup>T</sup> J	JCNB46-26	•	0.65 1.	. 50.	•	•		Derxia gummora ATCC 15994 <sup>T</sup> ATCC165-	-30	1.45	1.50	0.03		,	,
Hylemonella gracilis IPO 14920 <sup>T</sup>	PY-30	•	1.40 ]	- 01.		'		IAM 13946" [AMB1-	-25	- 1.02	1.60	,		0.10	0.02
(Aquuspirillum gracile)								1AM 14990 IAMB1-2	27	06'0	1.08	0.02		,	
Herbaspirillum autotrophicum IFO 15327 <sup>1</sup>	199-30°		0.65 1	- 45		,		Mitsuaria chitosunitabida IAM 14711 <sup>4</sup> NB-3(	30	- 0.09	1.17		ı	۲	
(Aquaspirillum autotrophicum)								Azothydramonas lata IAM 12599 <sup>T</sup> 199-3(	30	0.70	0.95	0.02	,	0.10	
2								(Atcallyenes latus)							
								Azohydromenas australica IAM 12664 <sup>7</sup>	-30	0.3	5 0.45	,	•	0.18	ł

(Alcaligenes latus)								
Giesbergeria anulus IFO 14917 <sup>1</sup>	PY-30°	,	1,15	2.00			,	,
(Aquaspirillum anulus)								
Giesbergeria giesbergeri IFO 13959 <sup>1</sup>	PY-30°		0.25	0.50		•		,
(Aquaspirilium glesbergeri)								
Giesbergerla sinuosa IFO 14925 <sup>r</sup>	PY -30 <sup>h</sup>		0.10	9.0		ī	i.	ı
(Aquaspiritivan sinuosum)								
Simplicispira metamorpha IFO 13960 <sup>T</sup>	199-30°		0.77	0.49		,		
(Aquaspirillum metamorphum)								
Simplicispira psychrophila IFO 13611 <sup>T</sup>	PY-30 <sup>6</sup>		0.25	<u>'</u>			0.08	
(Aquaspiriilan psychrophilum)								
Order Rhodocyclales								
"Pseudomonus butanovora" IAM 12574	NB-30		0.65	2.20	,		0.38	0.02
Zaoglova rumigera IAM 12136'	199-304	•	0.35	0.12	,		ı.	
Zoogloea resiniphila ATCC 700687 <sup>f</sup>	ATCC2136-30 <sup>4</sup>		0,60	J.45	,		ľ	,
Zoogloea oryzae IAM 15218 <sup>r</sup>	08-30		0.17	0.25		'	•	,
Order Hydrogenophilales								
Hydrogenophilus thermaluteolus IFO 14978	y <sup>T</sup> IFO820-50 <sup>c</sup>	,	1.32 0	0 08	02 0.	3	24	02
Hydrogenophilus hirschil JCM 10831 <sup>7</sup>	JCM279-60		1.70 (	8	.15 0	5	0,24 0	0.67
Order Neissenales								
Alysiella crassa IAM 14969 <sup>7</sup>	IAMB127-37		0.50	0.35	0.07			
(Simonsiella crassa)								
Alysiella filiformis IAM 14895 <sup>1</sup>	IAMB (27-37	'	0.30	0.02				
Conchiformibium steedae IAM 14972 <sup>7</sup>	IAMB127-37		1.07	0.60	140		,	
(Simonsiella steedae)								
Conchiformibium kuhniae LAM 15037 <sup>1</sup>	199-30	٠	0.25	0.95			·	
Bargeriella denitrificans IAM 14975 <sup>1</sup>	NB-30	'	0.04	0.15	0.03		1.18	,
	119-30	ı	0.04	0.10	ı		0.35	
Microvirgula aerodenitrificans [AM 14991	1AMB104-30	•	0.55	0.10	0.16	,		
IAM 14943*	IAMB33-27	•	0.15	0.0	1.88		0.05	
(Aquaspirillum disper) IFO 15328	199-25	0.20	0.95	0.01	1.30		,	,
Incertae sedis								
Thiobacter subterraneus $JCM 12421^T$ JC	CM 419-55	'	0.10	0.20	1,40		0.10	,

Sivunonas terrae IAM $15217^{T}$	0E-BN	- 0.50	1.10	,		•
		-	:	-		:
Dap, diaminopropane; H-Put, 2-hydroxyputa	rescine; Put, putrescir	ie; Cad, cadaven	ne; H-Sj	д 2-р	NOTOX	yspermudiae
Spd, spermidine; Spm, spermine; -, not deter	cted (<0.005); <sup>T</sup> , Type	e strain; IAM, In	stitute of	Mole	cular a	nd Cellular
Biosciences, The University of Tokyo, Toky	yu, Japan; JCM, Japar	n Collection of N	licroorgs	nistra	, RIKE	EN, Wako,
Saitama, Japan; IFO, Institute for Fernentat	tion, Osaka, Japan; N	BRC, Biological	Resourc	e Cen	ter, Na	tional
Institute of Technology and Evaluation, Kis	arazu, Japan; ATCC,	American Type (	Culture (	Collect	ion, M	anassas,

,

.

J

- 0.19 0.40

NB-30

"Pseudomonas jianii" IAM 12403

Virginia, USA. Former names are shown in parentheses. Quotation marks indicate that the scientific name has not been validly published. "was cited from Hamana et al., 1992, <sup>b</sup> Hamana et al., 1994, <sup>b</sup> Hamana et al., 1991,<sup>a</sup>

Натапа *et al*., 2000.

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polyamine. Norspermidine and homospermidine have not been found in the betaproteobacteria analyzed in the previous<sup>6, 8</sup> and present studies.

Spermidine and spermine were detected in a moderately thermophilic species of *Hydrogenophilus*, *H. hirschii*, growing at 60°C as a major polyamine, suggesting that the occurrence of spermine, a tetra-amine, correlate to its thermophily upper 60°C. On the other hand, significant amount of spermine was not found in slightly thermophilic (thermotolerant) *Caldimonas manganoxidans* growing at 45°C, *Hydrogenophilus thermoluteolus* growing at 50°C and *Thiobacter subterraneus* growing at 55°C. Uptake of spermine by other mesophilic species from media is not excluded.

## Class Gammaproteobacteria (Table 3)

#### Order Alteromonadales

Norspermidine was detected in two newly analyzed species of *Psychromonas* in the present study and a *Moritella* species previously analyzed<sup>7</sup>. Four types of polyamine profiles, putrescine alone, putrescine plus cadaverine, hydroxyputrescine plus putrescine, and putrescine plus spermidine, were found in Shewanella species including newly analyzed species. Cadaverine was the major polyamine in the two new species of Moritella, however, spermidine had not been found in this genus. Two species of Colwellia contained no cellular polyamines. Polyamine disribution profiles were varied within the mesophilic/psychrophilic marine proteobacteria belonging to the genera Psychromonas, Shewanella, Colwellia and Moritella, indicating that these genera are phylogenetically and chemotaxonomically conflicting taxa. Hydroxyputrescine was detected in several Shewanella and a Colwellia species belonging to this order. The occurrence of this hydroxydiamine related to branching point of the two classes Betaproteobacteria and Gammaproteobacteria, therefore, serve as a chemotaxonomic marker for betaproteobacteria within the phylum Proteobacteria. Spermidine was the major polyamine in Alteromonas, Idiomarina, Marinobacter, Marinobacterium, Pseudoalteromonas and Ferrimonas species. Pseudidiomarina taiwanensis contained putrescine alone. The occurrence of spermine in

Pseudoalteromonas phenolica and Psychromonas marina growing at  $15^{\circ}$ C is interesting, however, spermine has been detected ordinarily in thermophilic proteobacteria as shown in the present study. Spermine was not found in the psychrophilic Psychromonas, Shewanella, Colwellia and Moritella species growing under 10°C.

## Order Vibrionales

Norspermidine was found in three new species of *Vibrio* and three new species of *Photobacterium*. This triamine had been detected in almost species of these two genera previously analyzed<sup>19)</sup>.

## Order Oceanospirillales

Former Oceanospirillum species have been devided into the three genera Oceanospirillum, Pseudospirillum and Oceanobacter belonging to the order Oceanospirillales and the genus Marinobacter belonging to the order Alteromonadales<sup>1, 2, 13)</sup>. However, the analyzed species belonging to the four genera as well as the related genera Marinomonas, Halomonas and Chromohalobacter belonging to this order contained putrescine and spermidine as the major polyamine. Microbulbifer and Agarivorans contained cadaverine and spermidine. Occurrence of cadaverine seems to be species-specific in the genera Halomonas and Marinomonas.

#### Order Pseudomonadales

Almost species of the genera Azotobacter, Azorhizophilus, Psychrobacter, Moraxella and Pseudomonas contained putrescine, cadaverine and spermidine. Two Cellvibrio species contained putrescine alone. A new species as well as previously analyzed species of Acinetobacter contained diaminopropane alone<sup>23)</sup>.

## Order Thiotrichales

Five *Thiothrix* species of the order Thiotrichales were heterogeneous in their polyamine profiles. *Thiomicrospira* and *Methylophaga* had same polyamine profile, putrescine plus spermidine. Order Xanthomonadales

Among *Pseudoxanthomonas*, *Rhodanobacter* and *Aquimonas* species analyzed in the present study, a species of *Pseudoxanthomonas*, *P. koreensis*, lacked polyamines. Others contained spermidine as the major polyamine. Chemotaxomomic significance of the absence of cellular polyamines, found in *P*.

Table 3. Cellular concentrations of polyamines of gammaproteobacteria

Organism	um-Temp(°C)	щ	Polyam	uine ( μ	mol/g	stet wt	cell):			Shewanella gaetbuli JCM 11814 <sup>T</sup>	MB-30		1.25	0.30					
		Dap	H-Put	t Put	Cad 1	VSpd S	pd Sp	m Aj	шä	Shewunella japonica IAM 14956 <sup>T</sup>	MB-25		,	0:30	0.75				
Order Alteromonadales										Shewanella marinintesänu JCM 11558 <sup>r</sup>	MB-25	,	41	1.25	1	,		,	,
Alteromonas macteodii LAM 12920 <sup>3</sup> 199	9NaCI-25		,	,	0.14	-1 -	52	0	8	Shewarella marisflavi JCM 12192 <sup>1</sup>	MB-30	,		1.06			80		
	MB-25				0.05		±5 -	0.0	80	Shewanella profunda JCM 12080T	MB-25	,	,	0.73	0.05		,		
Alteromonus marina JCM 11804 <sup>T</sup>	MB-30	,		,	9.39	.0	8	Ċ	17	Shewanella sairae $JCM 11563^{T}$	MB-25	,	٠	0.72	,	,		,	
Alteromonas litorea JCM 12188 <sup>T</sup>	MB-30			0 -	130	3.1	' 9	0.1	νi	Shewanella sediminis NCIMB 14036 <sup>T</sup>	01-EM	,	0.75	01.1			.02		
Pseudoalteromonas peptidolytica MBIC 01416 <sup>7</sup>	5 <sup>7</sup> MB-30	•				- 1	.10	Ö	315	Shewanella schlegellana JCM 11561 <sup><math>T</math></sup>	MB-25	,	4	0.25	ı	,		,	
$Pseudoalteromonus spongiae JCM 12884^{ m I}$	MB-30	,		,			- 20	Ģ	05	Shewanella hafniensis NBRC 100975 <sup>1</sup>	MB-25	,	0.87	0.02	,			,	,
Pseudoalteromonas phenolica IAM 14989 $T$	MB-15	ŀ			,		0.60	030		Shewavella halifarensis NCIMB 14093 <sup>1</sup>	MB-15	'	0.60	050				,	,
W	Bagar-15	,	ī	,	0.07		ନ୍ତ	<b>J</b> 30		Shewanella morhuze NBRC $100978^{T}$	MB-25	,	1.27	ı	,	•	'	•	ī
Pseudoalteromonus saguniensis JCM 1146 $^{ m I}$	MBagar-27	ľ			,		0.10			Shewanella pneunatophori JCM 13187 $^{ m t}$	MB-27	•	•	0.75	,	,		ī	
Psychromonas marina LAM 14899 <sup>t</sup>	MB-15		ı	10.0	0.22	0.80	0.04	0.65	ı	Colwellia piezophila JCM 11831 <sup>T</sup>	MB-10	•	,	,	,				,
Psychronionus kaikoae JCM 11054 <sup>7</sup>	Mf3-10	,	ı	0.04	1.25	0.25			,	Colwellia maris JCM 10085 <sup>1</sup>	JCM253-10		0.16 (	0.56		-	34	,	,
Psychromonus profunda JCM 11437 <sup>T</sup>	MB-4	ı	ı	0.15	1.10	•				Colwellia psychrerythrae ATCC 27364 <sup>T</sup>	MB-10 <sup>4</sup>	,		,	ŀ	4			ī
Idiomurina seosinensis JCM 12526 <sup>T</sup>	MB-30			,		ī	0.75		0.20	Moritella profunda JCM 11435 <sup>r</sup>	MB-2	'	,	,	0.40				,
Pseudidiomarina taiwanensis JCM 13360 <sup>T</sup>	PY-30	,	,	0.70	ï		,			Moritella abyssi JCM 11436 <sup>t</sup>	MB-4		•	ŗ	0.38	1			
Shewanella algae IAM 14159 <sup>T</sup> 15	1995 W-20°			1.45	0.50				ı	Movitella marina ATCC 15381 <sup><math>T</math></sup>	PYSW-25	'		۱	1.10	,		•	'
Shewanella baltica NCIMB 1733	MB-20	ï	ī	1.20	1.50					Moritella japonica JChi 10249 <sup>1</sup>	NB-10 <sup>6</sup>	'	•		0.50		,	,	
Shewavella bentica ATCC 43992 <sup>7</sup>	MB-2"			0.25		•				Moritella yayunosii JCM $10263^{T}$	MB-10'	'		0.02		1.50	,		
Shewanella frigidimarina NCIMB 400	MB-30	ī		0.80	,	,	0.02		,	Marinohacter litoralis JCM 11547 <sup>r</sup>	MB-30				ŀ		1.10		,
Shewanelta hanedai IAM 12641 <sup>T</sup>	1995W-20*	,	,	1.50	•		•			Marinobacter lutaoensis JCM 11179 <sup>r</sup>	MBagar-37	ı			0.33	•	1.16		ŀ
Shewanetta putrefaciens IAM 12079 <sup>T</sup> 1:	1995W-30*	,		3.00		,			ı	Marinobacter hydrocarbonoclasticus AT	CC 496407199SW-20	۔ ۲	•	١	,	,	0.98		'
Shewanella violacea JCM 10179 <sup>1</sup>	MB-4'	•	0.60	0.45		ı	ī	ī	ı	(Pseudomonas nauticu) IAM 12929(=A	TCC 27132) MB-25	ı	ī	0.03	0.10		1.35	,	0.03
Shewanetta woodyi NCIMB 13526 <sup>1</sup>	MB-20 <sup>f</sup>			0.50		ı	0.25			Marinubacter flavimaris JCM 12323 <sup>7</sup>	MB-30			0.05	,	,	0.69	,	0.03
Shewanella amazonensis ATCC 700329 <sup>t</sup> ATCC	C1065-20*		,	1.05	,	٢	0.08	,	,	Marinobucter daepoensis JCM 12324 <sup>F</sup>	MB-30	'	,	0.07	ı	,	1.20	ı	•
Shewanella peateana ATCC 700345 <sup>1</sup>	MB-30 <sup>6</sup>	1	·	0.67	1		•	ı	ı	Marinobacter alkaliphilus JCM 12291 <sup>T</sup>	JCM392-30	1	,	0.10	0.12		0.65	0.01	0.05
Shewanella oneidensis ATCC 700550 <sup>T</sup>	TSB-30 <sup>a</sup>	1		1.50	1.10		,		,	Marinobacter maritimus JCM 12521 <sup>T</sup>	JCM443-22	,	,	0.18	0.04		0.56	•	0.02
Shewanella gelidimarina ATCC 700752 <sup>4</sup>	MB-30 <sup>6</sup>	'		0.01	0.01	,	1.12	,	,	Marinobacterium georgiense ATCC 700	774 <sup>™</sup> MB-25 <sup>1</sup>	,	,	0.04	0.10	ı	06.1	0.03	
Shewunella aquimarina JCM 12193 <sup>T</sup>	MB-30	ı	ı.	1.08	,	•	,	,	ı	(Pseudonionus iners) LAM 1419	) NB-25	•	,	0.33	0.10	,	0.75	0.01	'
Shewanella decolorationis IAM 15094 <sup>T</sup>	MB-25	,		11.1	,		,	1		Marinobacterium januaxchii IFO 15466 <sup>1</sup>	1998W-30	'	•	0.02	,	•	0.80	,	
										(Oceanospirillum jannaschii)									
										Ferrimonus marina MBIC 06486 <sup>T</sup>	MB-30	,	ľ		'	•	1.05	,	,

Order Vibrionales									
Photobacterium ganghwense JCM 12487 <sup><math>T</math></sup>	MB-25				ı	0.60	0.40		
Photobacterium frigidiphilum JCM 12947 <sup>1</sup>	MB-10					0.24	0.48		
Photobucterium indicum IFO 14233 <sup>T</sup>	MB-30°			0.01	,	0.52	0.62	,	
( "Hyphomicrobum indicuon" )									
Vibrio metschnikovii IAM 14406 <sup>T</sup>	MB-37			0.08	,	0.50	0.48	,	
Vibrio superstes IAM 15009 <sup>T</sup>	MB-15		ī	,		0.02	0.65	0.10	
	MBagar-15	·	·	0.0	,	0.20	0.38	•	1
Vibrio ruber JCM 11486 <sup>r</sup>	TSB-30		,	,	0.55	0.62	,	0.02	۱
	fSBagar-30	'		'	0.20	0.50	,	,	,
Order Oceanospiriliales									
. Oceanobacter kriegii IFO 15467 <sup>T</sup>	199SW-30	'		0.03	,		0.84		,
(Oceanospirillum kriegii)									
Pseudospirillum japonicum IFO 15446 <sup>1</sup>	05-W-30			0.01			1.11		ı
(Oceanospirillun juponicum)									
Hulomonas alimentaria JCM 10888 <sup>-</sup>	MB-30	•	,	0.02	۱	'	0.83	L	•
Halomonas gluchei JCM 11692 <sup>T</sup>	MB-20			0.07	0.15		0.89	,	,
Halomonas marisfiavi JCM 10873 <sup>T</sup>	MB-20						1.26		
Halemonas koreensis JCM 12237 <sup>1</sup>	MB-35			0.0			1.32	,	0.10
Chromohalobacter salexigens NCIMB 1376	8 <sup>r</sup> NCIMB219-30	•	٠				0.80	•	
Chromolialobacter canadensis NCIMB 1370	57 <sup>T</sup> NCIMB473-30	•	۱	0.02		•	1.32	,	
(Halomonas canadensis)									
Chromohalobauter israelensis NCIMB 1376	6 <sup>7</sup> NCIMB473-3()			ľ	•	'	0.71		
(Hulomonas istaelensis)									
Marinomonas communis IAM 12914 <sup>r</sup>	06-W2691		ī	1.05	4		0.35	.,	
Marinomonas vaga IAM 12923 <sup>1</sup>	1998W-30		,	01.0			0.50		
Marinomonas mediterranea IAM 14944 <sup>1</sup>	MB-25			0.04	0.16		0.48		,
Marinonwnas prinwyensis IAM 15010 <sup>7</sup>	MB-30			0.35	05.0		0.64		1
Marinomonas ushuaiensis JCM 12170 <sup>T</sup>	MB-30			0.36	0.02		0.37		
Microbulbifer satipatudis $JCM 11542^{T}$	MB-37		,	,	0.20	1	0.60	,	0.15
Microbulbifer hydrolynicus ATCC 700072 <sup>T</sup>	£2-WS991			0.02	.65		0.25		0.06
(Microbulbifer hydrolynca)									
Agarivorans albus [AM 14998 <sup>T</sup>	MB-25	0.01		0.05	0.37	,	0.98	0.03	

Order Pseudomonadales

Azotobacter beijerinckii IAM 12683 <sup>1</sup>	$YM-30^{h}$	,		0.11	0.01	-	507		
Azotobacter chroacoccum IAM 12994 <sup>°</sup>	YM-30			1. <del>4</del> 4	0.22		050		
Azotobacter nigricans IAM 15005 <sup>T</sup>	NB-28	,		0.10	0.10		0.44		
Azvtobacter vinelandü 🛛 IAM 15004 <sup>r</sup>	14MB138-28	•		1.30	0.12	,	0.32		
IA	MB 138agar-28	•		0.70	0.10		0.12		
IAM 1078	YM-30°	,	,	0.94	0.27		0.28 0	8	
Azotobacter armeniacus IAM 15047 <sup>°</sup>	IAMB129-30	ı	ı	0.80	0.08		0.28		,
Azotobacter salinestris AM 15020 <sup>T</sup>	IAMB129-26			0.75	0.35	,	0:30		
Azorhizophilus paspuli IAM 12667 <sup>T</sup>	IAMB13-30			1.37	,		0.34		,
(Azotobacter paspait)	109-30*	'	,	2.16	•		0.85	,	,
Psychrobacter phenylpyravicus IAM 1228	32 <sup>1</sup> 1 <del>99</del> -25 <sup>-</sup>	ı	•	1	0.20	·	1.35		
Psychrobucter urativarans ATCC15174 <sup>7</sup>	199-254	,	,	0.61	0.14	'	0.07	,	
Psychrobacter friginola ATCC 700361 <sup>T</sup>	ATCC1849-15 <sup>4</sup>	•	•	•	,	'	1.08		
Prychrobacter glacincola ATCC 700754 <sup>1</sup>	MB-15 <sup>6</sup>	'	'	0.0	0.06	,	0.86	0.02	
Psychrobacter immobilis IAM 12280 <sup>T</sup>	NB-25*	,		0.25	,		1.30		
Psychrobacter pacificensis IFO 16270 <sup>T</sup>	MB-25 <sup>s</sup>	,		0.10	•		1.15	,	
Psychrobacter jeotgali JCM 11463 <sup>T</sup>	MB-30	4		0.04	0.20		1.60		,
Psychrobacter okhowskensis JCM 11840 <sup>T</sup>	MB-20	,	,	01,0	0.07		1.26		
Psychrobacter adimentaris JCM 12315 <sup>7</sup>	MB-30	,	•	0.15			96.0		,
Psychrobacter celer JCM 12601 <sup>T</sup>	MB-30		•	0.10	0.05		1.10		,
Morazeila oblonga IAM 14971 <sup>T</sup>	646+SE-37	•		0.02			1.02	,	
Pseudomonas atotifigens JCM 12708 <sup>7</sup>	JCM443-30			0.50	0.40		0.40		,
Pseudononas beijerinckii LAM 14940 <sup>T</sup>	NB-NaCI-25	'	'	0.01	,	•	1.95		ı
Pseudomonas pachastrellae JCM 12285 <sup>T</sup>	MB-30	'	•	1.20	,	'	0.80		,
Pseudomonus psychrophila JCM 10889 <sup>7</sup>	NB-30	•	•	1.05	,	,	0.65		
Celtvibrio vulgaris NCIMB 8633 <sup>T</sup>	NCIMB16-20	ľ		0.40	,	,	•		,
Cellvibrio fulvus NCIMB 8634 <sup>1</sup>	NCIMB16-20	,		0:30		r	•		
Acinetobacter venetianus MBIC 01332	MB-20	1.50		•					,
Activetobacter culcoaceticus IAM 12087 <sup>r</sup>	199-30	2.41		,		ı	ı	,	ı
Acinetobacter baumannii IAM 12088 <sup>7</sup>	199-30	134	,	ı	,		•		
Acinetobacter johnsonii IAM 1517	06-901	1.35			,	٠			
Aciitetobacter lwoffti GIFU 1951T	199-30	2.30			,			ŀ	,

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Actnetochacter radioresistens ATCC 43998 <sup>T</sup>	199-30	2.50	•	·	ı.	'	•		'	
Order Thiotrichales										
Thiothrix disciformis JCM 11364 <sup>t</sup>	JCM269-30			0.77	0.30	,	0.75		,	
Thiothrix flexitis JCM 11135 <sup>T</sup>	JCM 296-25	, ,	-	Ę	0.25	,		ī		
Thiothriz eikelboomii ATCC 497188 <sup>1</sup>	ATCC1820-20 <sup>6</sup>			.,	٠		1.10	4		
Thiothrix fructosivoruns ATCC 49748 <sup>T</sup>	ATCC1820-20 <sup>6</sup>		,	•		٠	1.12	٠		Ų
Thiotheix until ATCC 49747 <sup>T</sup>	ATCC1820-208					'	01.1	,		
Thiomicrospira thermophila JCM 12397 <sup>T</sup>	JCM423-30	ı	,	0.20		٠	0.40	,		
Thiomicrospira crunogena ATCC 35932 <sup>T</sup>	ATCC 1422-26		,	0.18	'	•	0.70	'	•	
Thiomicrospira pelophila ATCC 27801 <sup>T</sup>	ATCC1036-26 <sup>f</sup>		,	0.27	,	·	0.22	·	,	
Thiomicrospira thyusirae ATCC 51452 <sup>7</sup>	ATCC1891-30			0.57	ć	۲	0.03	ľ	•	
Methylophaga sulfidovorance IAM 1503 $S^{T}$	MB+methanol			0.05			0.10	'		
Methylophaga marina JCM 6886 <sup>7</sup>	199NaCl-30t			0.20			3.70			
Methykophaga thalassica JCM 6887 <sup>7</sup>	408-30k	,		0.20		•	2.00	'		
Order Kanthomonadales										
Dyella japonica IAM 15069 <sup>T</sup>	NB-25		'	0.18	06.1	'	1.25	•	0.47	
Pseudoxanthomonas mexicana JCM 11524 <sup><math>T</math></sup>	05-83T			1	·	•	1.44		ŀ	I
Pseudoxanthomonas japonensis JCM 11525	<sup>T</sup> TSB-30				,	'	1.37	•		-
Pseudoxanthomonas koreensis IAM 15116 <sup>T</sup>	TSB-30				•	•		•		ч
	199-30	,			,			·		20
Pseudoxanthomonas duejeonensis IAM 1511	LS <sup>T</sup> TBS-30					•	1.42	'		Ŭ
Rhodanobucter fulvus IAM 15025 <sup>7</sup>	NB-25		,	,		•	0.45	,	0.05	
Aquimonas varati JCM 12896 <sup>T</sup>	JLZB-30						0.80			-
Order Chromatiales										Ŭ
Thiovirga sulfuroxydans JCM 12417 <sup>1</sup> JC	CM414-30			'	ŗ		0.80		,	4,
Halothiobacillus halophilus ATCC 49870 <sup>T</sup> A	JCC1846-30			0.60	,		1.90		,	~
Halothiobacillus hydrothermatic ATCC 5145	53 <sup>1</sup> ATCC1901-3	, ¥.		0.02	'n	,	0.80			1-4
Halothiobacillus neapolitanus JCM 3861 $^{ au}$	JCM90-30°	,	ī	0.88	0.11	'	0.05		ī	H
Thioalkalivibrio jannaschii JCM 11372 <sup>1</sup>	JCNB14-30				,	'	1.00	,	ľ	
Thioalkalivibrio thiocyanexidans JCM 1136	8° JCNB14-30		,				1.45		'	
Thioulkalivibrio puradoxus JCM 11367 <sup>7</sup>	JCM310-30				'	,	1.03	•	'	
Rheinhetmera pacifica IAM 15043 <sup>T</sup>	MB-15	,	,	1.25	•	'	0.50	•	•	
Incertae sectos										

Kangiella aquimarina JCM 12318 <sup>T</sup>		MB-3(			1	0.47	0.36		0.50		
Kanyiella koreensis JCM 12317 <sup>T</sup>		MB-30	'			.65	0,42	,	1.08	0.6	0
"Pseudomonas auricularis" IAM 1	12(137	NB-3	, ,			0.52	•	•	٠		
"Mesophilobacter marinus" [AM ]	13 185	£-WS661	-			.00	1.25		1.00	1	
Salinisphaera shahunensis JCM 112	575 <sup>°</sup> J(	CM339-30	1	,	0	1.17	,	,	0.82	. 0.6	0
Order Enterobactoriales											
		Dap H-P	ut Pu	- -	I	VSpd	Spd	Spin	t Agm	Ac-Sp	
Aquumonus fontana IAM 15072 <sup>T</sup>	BHIB-30	•			0.10		1.10	'	'	,	
Erwinia aphidicola LAM 14479 <sup>T</sup>	NB-25	0.50	0	09.1	0.19		0.32	'	•	0.0	
	199-25	0.57	0.8	8	0.60	,	0.10	•	0.05	0.20	
Erwinia chrysanthemi MAFF 30176	5 199-25 <sup>4</sup>	,	1.4	~	0.15		0.57	ŀ	•	0.14	
Erwinia nigriflaens MAFF 301435	199-25	0.70	0.5	5	57		0.15	'	0.34	t0:0	
Erwinia persicinus IAM 12843 <sup>°</sup>	199-25		Ι.	0	0.32		0.57		•	0.03	
Erwinia rhapontici MAFF 301331	199-25	0.50	0.3	ģ	0.15		0.12		0.03	ı	
Рестоваснегит сагогогонит LAM	12633										
(Ervinia carotovora)	NB-25 <sup>1</sup>	0.35	-	2	0.05	•	0.75		•		
Dan. diaminomonane: H-Put 2-hvdr	0.000 Interest	ine But m	tracci	2°C •	vehen		and enter		Level Life		1
				· ·		1	vla vla			i	
homospermiente; Spin, spermine; Ag	çm, agmatlı	ne; Ac-Spd	N <sup>+</sup> -ac	ctylspe	erraidin	с; •, ло	t detecta	¥ 8	1.005); 1	, Type	
strain; IAM, Institute of Molecular ar	nd Cellular	Bioscience	S, The	Unive	rsity of	Tokyo	, Tokyo	Japa.	a; JCM,	Japan	
Collection of Microorganisms, RIKE	SN. Wako,	Sailama, Ja	pan; IF	O, Ins	titute fo	or Ferr	cutation	, Osa	ка, Јара	i i	
NBRC, Biological Resource Center,	National Ir	istitute of 7	cchnol	ogy an	id Eval	uation,	Kisamz	u, Jap	kan; MB)	ú	
Marine Biotechnology Institute Culta	ure Collecti	on, Kamai	shi, Jwa	lte, Jap	ALC: AT	CC, AI	nerican	Type	Culture		
Collection, Manassas, Virginia, USA	, NCIMB,	National C	bllectio	ns of ]	ndustri	al, Foc	[ pus sp	Marin	e Bacler	ia,	
Scotland, UK. Former names are si	eq ni nwoq	rentheses.	Quota	tion in	arks in	dicate	that the ;	scient	ific nam	e has n	ž

Hamana & Matsuzaki, 1993,° Hamana, 1997,<sup>6</sup> Hamana *et al.*, 2000,° Hamana *et al.*, 2003, <sup>h</sup> Hamana *et al.*, 1988,<sup>i</sup> been validly published. \* was cited from Hamana & Takeuchi, 1998, <sup>b</sup> Hamana *et al.*, 2001, ° Hamana, 2000, <sup>4</sup> Hamana et al., 1994, <sup>1</sup>Hamana & Matsuzaki, 1992, <sup>1</sup>Hamana & Kishimoto, 1996, <sup>1</sup>Hamana, 1996. *koreensis* as well as two *Colwellia* species within the Gammaproteobacteria, is unclear.

# Order Chromatiales

Three *Thioalkalivibrio* species and *Thiovirga* sulfuroxydans ubiquitously contained spermidine alone. *Rheinheimera pacifica* as well as *Halothiobacillus* species contained putrescine and spermidine.

### Order Enterobacteriales

A new *Erwinia* species analyzed in the present study, as well as other four spcies and a related *Pectobacterium* species, contained acetylspermidine and diaminopropane. It has been reported that these two polyamines were serectively distributed in the family Enterobacteriaceae of this order of the class Gammaproteobacteria<sup>18)</sup>. However, *Aquamonas fontana* located in this order contained putrescine and spermidine.

### Others

*Salinisphaera* species analyzed in the present study contained putrescine, cadaverine and agmatine.

#### Class Deltaproteobacteria (Table 4)

Within the class Deltaproteobacteria, many species belonging to the heterotrophic order Myxoccocales contained homospermidine, whereas previously analyzed other deltaproteobacteria belonging to other six orders Desulfobacterales, Desulfovibrionales, Desulfuromonadales, Syntrophobacterales, Desulfurellales and Bdellovibrionales contained spermidine as the major polyamine<sup>6, 12)</sup>. In the present analysis, sulfate-reducing Desulfobacterium cetonicum belonging to the order Desulfobacterales contained cadaverine and homospermidine whereas other two Desulfobacterium species contained cadaverine and spermidine. The occurrence of homospermidine in D. cetonicum is the first datum and is unique within this order.

In the order Desulfuromonadales, thermophilic *Geothermobacter ehrlichii* belonging to the Fe(II)-reducing family Geobacteraceae contained spermidine and spermine as the major polyamines. On the other hand, psychrotolerant *Geopsychrobacter electrodiphilus* belonging to Geobacteraceae as well as mesophilic *Geobacter* and *Pelobacter* species, contained spermidine alone. New species of the

sulfate-reducing genus *Desulfovibrio* of the order Desulfovibrionales, *D. brasiliensis*, *D. aerotolerans*, *D. alkalitolerans*, *D. frigidus* and *D. ferrireducens*, as well as other four *Desulfovibrio* species analyzed previously<sup>6, 12)</sup>, contained spermidine. Although thermophilic *Desulfurella acetivorans* (the order Desulfurellales) and *Desulfacinum infernum* (the order Syntrophobacterales) were poor in spermine, the major occurrence of spermine was found in a thermophile, *Geothermobacter ehrlichii*, in the present study. Distribution of cadaverine and agmatine seems to be genus- or species-specific in this class.

### Class Epsilonproteobacteria (Table 4)

Hydrogen-oxidizing Hydrogenimonas thermophila, a novel thermophile growing at 55°C, is located in the order Campylobacterales, and contained spermidine and spermine as the major polyamines. Mesophilic sulfur-oxidizing Sulfurimonas autotrophica and Sulfuricurvum kujiense located in the order Campylobacterales, contained spermidine alone. Other various heterotrophic, mesophilic epsilonproteobacteria belonging to the families Campylobacteraceae and Helicobacteraceae of the order Campylobacterales, contained also spermidine as the major polyamine<sup>6, 12</sup>.

Spermine was found as a major polyamine in two thermophilic nitrate-ammonifying Caminibacter species and a thermophilic, acidophilic hydrogenoxidizing Lebetimonas species of the new order Nautiliales. Mesophilic nitrate-reducing Nitratifractor salsuginis and mesophilic sulfuroxidizing Sulfurovum lithotrophicum and sulfurreducing Thioreductor micantisoli contained spermidine alone. Thermophilic nitrate-reducing Nitratiruptor tergarcus contained spermine as the main polyamine component. Polyamines of various chemolithoautotrophs belonging to deltaproteobacteria and epsilonproteobacteria have been analyzed. However, polyamine profiles related to their optimum growth temperature rather than their chemolithoautotrophic type. The occurrence of spermine in the chemolithotrophic, thermophilic deltaproteobacteria growing at 50-60℃ and epsilonproteobacteria growing at 50-55℃ was Table 4. Cellular concentrations of polyamines of deltaproteobacteria and epsilonproteobacteria

			:				
Organism Medium-Lemp (U)	Polya	mme (1	t mol/g	wet w	t cell):		
	Dap	Put	Cad	pds	HSpd	Spn	Agm
Deltaproteobacteria							
Order Desulfobacterales							
Desulfobacterium cetonicum JCM 12296 <sup>T</sup> JCM395-30	,		0.20		0.72	•	
Desulfobacterium vacuolatum JCM 12295 <sup>T</sup> JCM394-30	۰		0.75	0.85	•		,
Desulfobacterium niacini JCM 12294 <sup>5</sup> JCM393-30	ī	,	0.79	0,65	•	•	,
Desulfococcus multivorans NCIMB 12965 <sup>T</sup> NCIMB295-30 <sup>b</sup>	'	0.02	,	0.70	•	0.04	,
Desulfobulbus propionicus NCIMB 12907 NCIMB17-37°		0.02		0.88	ŗ	0.04	
Order Desulturomonadales							
Geothermobacter ehrlichti JCM 12418 <sup>4</sup> JCM416-50	,		•	0.40	,	050	
Geobucter suffarreducens ATCC 51573 <sup>1</sup> ATCC1957-30 <sup>b</sup>	•	,		L.10			0.03
Geobacter hydrogenophilus AFCC 515W <sup>T</sup> AFCC1957-30 <sup>h</sup>		•	•	0.92	1		0.10
Geopsychrobacter electrodiphilus JCM 12469 <sup>7</sup> JCM425-22	,		,	0.52	•		
Pelobacter acidigatilici ATCC 49970 <sup>7</sup> ATCC1852-30 <sup>7</sup>	'	ı		1.45	'		0.24
Pelobacter masseliensis ATCC 49973 <sup>T</sup> ATCC1852-30 <sup>t</sup>	·			1.20	'		0.20
Desulfuromonus svalbardensis JCM 12927 <sup>®</sup> JCM463-15				0.44	•		
Desulfuromusa ferrireducens JCM 12926 <sup>T</sup> JCM463-15	,			0.60		٠	
Order Desulfurzilales							
Desulfurella acetivorum: ATCC 51451 <sup>T</sup> ATCC 1920-55 <sup>b</sup>				0.97	'	0.0	'
Order Desuffovibrionales							
Desulfovibrio aerotolerans JCM 12613 <sup>T</sup> JCM439-30			,	040	۰	,	
Desulfovibrio alkalitol <del>a</del> rans JCM 12612 <sup>7</sup> JCM438-40			,	0.42	,	•	
Desulfovibrio brasiliensis JCM 12178 <sup>t</sup> JCM383-30	1	ι		0.52		'	
Desulfovibrio ferrireduceus JCM 12925 <sup>T</sup> JCM462-20	•	•	•	0.29	'		ī
Desulfovitrio frigidus JCM 12924 <sup>7</sup> JCM462-20		٠	•	0,44			
Desulfovitrio africanus NCIMB 8401 <sup>T</sup> NCIMB17-37 <sup>b</sup>	10.0	0.12	0.03	0.95		0.02	0.03
Desulfovitrio desulfaricans NCIMB $8307^{T}$ NCIMB17-30 <sup>b</sup>		0.24	1.98	0.40	i.	,	
Desulfovibrio salexigens NCIMB 8403 <sup>T</sup> NCIMB104-30 <sup>b</sup>	,	0.05	0.04	0,65		٠	0.20
Desulfovierio vulgaris · NCIMB 9442 <sup>T</sup> NCIMB17-30 <sup>b</sup>	,	0.02	0.01	05.1	÷	0.10	0.03
Order Syntrophobacterales							

Desulfacinum infernum NCIMB 13416 <sup>t</sup> N	CIMB109-60*	•	0.02		0.68		0.06	
Epsilonproteobacteria								
Order Campylobacterales								
Sulfurimonas autotrophica JCM 11897 <sup>r</sup>	JCM365-24	,	ı	•	0.75	,	,	ı
Sulfarimonas denitrificans ATCC 33889 <sup>1</sup>	ATCC1255-26*	'	۰.	0.4	0 2.10	'	•	
(Thiomicrospira deninificans)								
Suljuricurvum kujiense JCM 11577 <sup>n</sup>	JCM340-25		,		0.50		ī	
Hydrogenimonas thermophila JCM 11971 $^{ extsf{T}}$	JCVB69-55		0.02	•	0.70	4	0.17	
Sulfarospirillum burnesii ATCC 700032 <sup>1</sup>	ATCC2034-30 <sup>b</sup>	•	,	•	1.10		0.01	
Sulfurospirithum arsenophihum ATCC 7000	56 <sup>T</sup> ATCC2018-30 <sup>b</sup>	0.02	Ļ	0.85	0.10		0.03	
Sulfarospirillum deleyianum ATCC 51133 <sup><math>T</math></sup>	ATCC1842-30*	'		•	1.25			
Order Nautiliales								
Caminiburier profundus JCM 12957 <sup><math>T</math></sup>	JCM367-55	•	0.27		0.12	,	0.75	,
Caminibacter mediatianticus $JCM$ $12641^{T}$	JCM440-55	ī	0.04	1	0.04		0.70	0.04
Lebetimonus acidiphila JCM 12420 <sup>T</sup>	JCM417-50		,	0.10	0.40	,	0.70	
Incertae sedis								
Nitralizuptor tergarcus JCM 12459 <sup>E</sup>	JCM421-55	٠	۰.	'	0.02		1.00	
Nitralfractor satsuginis JCM 12458 <sup>T</sup>	JCM421-37	ľ	,	'	0.54	·	,	
Sulfurovun lithotrophicun JCM 12117 <sup>T</sup>	JCM374-28	,	·		0.25	,	,	
Thioreductor micantisoli JCM 12457 <sup>†</sup>	JCM420-32	,		,	0.40			,
Dap, diaminopropante; Риг, рицеscine; Саd, с	adavenne; Spd, spenn	idine; F	(Spd, he	dsound	ermidine	Spm	, speimi	le;

Dap, diaminopropante; Put, puttescine; Cad, cadavenine; Spd, spermidine; HSpd, homospermidine; Spm, spermine; Agm, agmatine; -, not detected (-0.005); <sup>T</sup>, Type strain; ATCC, American Type Culture Collection, Manassas, Virginia, USA; JCM, Japan Collection of Microorganisms, RIKEN, Wake, Saitana, Japan; NCIMB, National Collection of Industry, Marine and Food Bacteria, Scotland, UK. \* was cited from Hamana & Takeuchi, 1998, \* Hamana et al., 2004.

### observed.

Polyamine distribution profiles in the phylum Proteobacteria

Homospermidine was distributed as the major polyamine in the alphaproteobacteria belonging to the nine genera, Sphingomonas and Sphingosinicella within the order Sphingomonadales, Dinoroseobacter within the order Rhodobacterales, Azospirillum and Tistrella within the order Rhodospirillales, Rhodopseudomonas and Pseudaminobacter within the order Rhizobiales, and Asticcacaulis and Brevundimonas within the order Caulobacterales. The three triamine profiles, homospermidinedominant type, spermidine-dominant type and homospermidine/spermidine-dominant type, were genus-specific. These results suggest the varieties of their triamine profiles and the usefulness of triamine pattern as a phenotypic marker in the class Alphapoteobacteria. The occurrence of homospermidine instead of spermidine was not ubiquitous in the genera Sphingomonas and Brevundimonas, indicating that chemotaxonomic usefulness of polyamine profile is not agree with the criteria of these genera. Spermidine and homospermidine are produced by the different pathways, aminopropyl transfer or aminobutyl transfer, however, the two biosynthetic activities correlate to their phylogenetic locations at genus level within the class Alphaproteobacteria.

All betaproteobacteria analyzed ubiquitously contained hydroxyputrescine and putrescine. Spermidine was sporadicaly distributed in some members. Norspermidine and homospermidine were not found. Since hydroxyputrescine was almost selectively distributed in the class Betaproteobacteria including 66 species previously analyzed<sup>6, 8)</sup>, the occurrence of hydroxyputrescine in 46 strains also serve as a chemotaxonomic marker for betaproteobacteria within the phylum Proteobacteria. Putrescine is produced by ornithine decarboxylation and then hydroxylated. When spermidine was synthesized from putrescine, hydroxyspermidine was also produced from hydroxyputrescine. In addition to spermidine, spermine was dominant in a thermophilic Hydrogenophilus species within this class.

Norspermidine was the major polyamine in the Photobacterium and Vibrio species analyzed, belonging to the order Vibrionales, and was detected as a major polyamine in two species of Psychromonas and one species of Moritella, within the order Alteromonadales. In the order Alteromonadales, the genera Shewanella, Colwellia and Marinobacter were heterogeneous in their hydroxyputrescine, cadaverine and spermidine levels. Spermidine was the major polyamine in the other six orders Oceanospirillales, Pseudomonadales, Thiotrichales, Xanthomonadales, Chromatiales and Enterobacteriales. Homospermidine was not found in the all gammaproteobacteria analyzed. Diaminopropane was the main component in Acinetobacter of Pseudomonadales<sup>23</sup> and a major polyamine in Erwinia and Pectobacterium of Enterobacteriales<sup>18)</sup>. Diaminopropane and norspermidine, produced from diaminopropane by an aminopropylation, were selectively detected in some genera. These polyamine distribution profiles serve for the chemotaxonomy of gammaproteobacteria, however, the profiles are conflicting in some genera.

Desulfobacterium cetanicum contained homospermidine and other deltaproteobacteria, Desulfococcus, Geothermobacter, Geopsychrobacter and Desulfovibrio species, contained spermidine as the major polyamine. Cadaverine distributed in Desulfobacterium, Desulfococcus and Desulfovibrio species. Spermidine was found as a major polyamine in epsilonproteobacteria. Spermine was detected in a novel thermophile of Geothermobacter within the class Deltaproteobacteria and in the five novel thermophiles belonging to the genera Hydrogenimonas, Caminibacter, Lebetimonas and Nitratiruptor of the class Epsilonproteobacteria, suggesting that the occurrence of spermine correlate to their thermophily. The widespread distribution of moderate thermophiles containing spermine within the phylum Proteobacteria is suggested. Additional data on cellular polyamine catalogues, shown in the present study, serve as a chemotaxonomy for further classification of the phylum Proteobacteria.

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## REFERENCES

- Brenner D J, Krieg N R, Saley J T. (eds.) The Proteobacteria. *In* Bergey' Manual of Systematic Bacteriology, 2nd ed., vol. 2, part C. Springer, Berlin, 2005.
- 2)NCBI (National Center for Biotechnology Information, U.S. National Library of Medicine), Taxonomy Browser Bacteria (2006). http//ncbi.nlm.nih.gov/Taxonomy/
- 3) Hamana K. Polyamine distribution pattern and chemotaxonomy of bacteria. Microbiol Cult Coll 2002; 18: 17-43 (in Japanese).
- Hamana K, Matsuzaki S. Polyamines as a chemotaxonomic marker in bacterial systematics. Crit Rev Microbiol 1992; 18: 261-283.
- 5) Hamana K, Matsuzaki S. Polyamine distribution patterns serve as a phenotypic marker in the chemotaxonomy of the Proteobacteria. Can J Microbiol 1993; 39: 304-310.
- 6)Hamana K, Takeuchi M. Polyamine profiles as chemotaxonomic marker within alpha, beta, gamma, delta, and epsilon subclasses of the class Proteobacteria: Distribution of 2hydroxyputrescine and homospermidine. Microbiol Cult Coll 1998; 14: 1-14.
- 7) Hamana K, Okada M, Saito T, Nogi Y. Polyamine distribution profiles among some members of the gamma subclass of the class Proteobacteria. Microbiol Cult Coll 2000; 16: 51-61.
- 8) Hamana K, Saito T, Okada M. Polyamine profiles within the beta subclass of the class Proteobacteria: Distribution of 2hydroxyputrescine. Microbiol Cult Coll 2000; 16: 63-69.
- 9) Hamana K, Saito T, Okada M. Distribution profiles of spermidine and homospermidine within the alpha subclass of the class Proteobacteria. Microbiol Cult Coll 2001; 17: 3-12.
- Hamana K, Sakamoto A, Tachiyanagi S, Terauchi E. Polyamine profiles of some members of the gamma subclass of the class Proteobacteria.

Polyamine analysis of twelve recently described genera. Microbiol Cult Coll 2003; 19: 3-11.

- 11) Hamana K, Sakamoto A, Tachiyanagi S, Terauchi E, Takeuchi M. Polyamine profiles of some members of the alpha subclass of the class Proteobacteria: Polyamine analysis of twenty recently described genera. Microbiol Cult Coll 2003; 19: 13-21.
- 12) Hamana K, Saito T, Okada M, Niitsu M. Polyamine distribution profiles among some members within delta- and epsilon-subclasses of Proteobacteria. Microbiol Cult Coll 2004; 20: 3-8.
- 13) Validation List 95-109. *In* Int J Syst Evol Microbiol (ed. by The Society for General Microbiology) vol. 54-56, 2004-2006.
- 14) Hamana K, Sakane T, Yokota A. Polyamine analysis of the genera Aquaspirillum, Magnetospirillum, Oceanospirillum and Spirillum. J Gen Appl Microbiol 1994; 40: 75-82.
- 15) Hamana K, Matsuzaki S. Polyamines and their biosynthetic activities in nonphytopathogenic marine agrobacteria. Can J Microbiol 1990; 36: 567-572.
- 16) Hamana K, Matsuzaki S. Taxonomic significance of polyamine synthesis in *Paracoccus*. J Gen Appl Microbiol 1992; 38: 93-103.
- 17) Hamana K, Satake S, Iyobe S, Matsuzaki S. Polyamine distribution patterns in *Pseudomonas*, *Alcaligenes* and *Comamonas*. Ann Rep Coll Med Care Technol Gunma Univ 1992; 13: 105-109.
- 18) Hamana K. Distribution of diaminopropane and acetylspermidine in Enterobacteraceae. Can J Microbiol 1996; 42: 107-114.
- 19) Hamana K. Polyamine distribution patterns within the families Aeromonadaceae, Vibrionaceae, Pasteurellaceae, and Halomonadaceae, and related genera of the gamma subclass of the Proteobacteria. J Gen Appl Microbiol 1997; 43: 49-59.
- 20) Hamana K. Extraction and HPLC analysis of bacterial polyamines. Ann Gunma Health Sci 2002; 23: 149-158 (in Japanese).
- 21) Maruyama T, Park H D, Ozawa K, Tanaka Y, Sumino T, Hamana K, Hiraishi A, Kato K. Sphingosinicella microcystinivorans gen. nov., sp. nov., a microcystin-degrading bacterium. Int J

Syst Evol Microbiol 2006; 56: 85-89.

22) Takeuchi M, Hamana K, Hiraishi A. Proposal of the genus *Sphingomonas* sensu stricto and three new genera, *Sphingobium*, *Novosphingobium* and *Sphingopyxis*, on the basis of phylogenetic and chemotaxonomic analyses. Int J Syst Evol Microbiol 2001; 51: 1405-1417.

23) Hamana K, Matsuzaki S. Diaminopropane occurs ubiquitously in *Acinetobacter* as the major polyamine. J Gen Appl Microbiol 1992; 38: 119-114.