

# Long-term oral antibiotic treatment: why, what, when and to whom?

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Antibiotics are typically prescribed as short courses for acute infections, in order to reduce bacterial load, shift the balance in favour of host defences and thus help to overcome infection. Over the past decade, however, interest in the long-term anti-inflammatory and immunomodulatory effects of selected antibiotics has been on the increase. Since the clinical effectiveness of erythromycin was reported in diffuse panbronchiolitis in the 1980s, the use of macrolides has been adopted into many other chronic inflammatory airway diseases characterised by frequent exacerbations, including cystic fibrosis (CF), non-CF bronchiectasis, COPD, severe noneosinophilic asthma, bronchiolitis obliterans after lung transplantation and organising pneumonia. In this chapter, we discuss the indications and limitations of long-term macrolide treatment in these chronic respiratory conditions.

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# Long-term oral antibiotic treatment: why?

Airway disorders such as COPD, cystic fibrosis (CF) and bronchiectasis, in which chronic airway inflammation and (infectious) exacerbations occur, typically result in progressive lung function decline, decreased health-related quality of life and increased risk of mortality [1–5]. The pathophysiological processes typical of these chronic inflammatory lung diseases are characterised by a vicious circle hypothesis, in which exacerbations result in increased airway inflammation, more damage to airways and therefore a higher risk for new

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exacerbations, which will finally result in progression of disease (figure 1) [4, 6, 7]. To inhibit further disease progression, interventions have to be performed directed at one of these steps of the vicious circle. Among these interventions is the implementation of long-term oral antibiotic treatment. The most widely studied class of antibiotics used for oral maintenance therapy is the macrolide group.

# Long-term oral antibiotic treatment: what?

## Choice of agent

Although other antibiotics, such as the tetracycline doxycycline and the fluoroquinolone moxifloxacin, have also been assigned immunomodulatory properties [8, 9], only the use of macrolides has been well documented in RCTs. Their effects in chronic inflammatory lung diseases were first studied in patients with diffuse panbronchiolitis in the 1980s [10]. Since then macrolides have been studied extensively in CF [11–14], non-CF bronchiectasis [15–17], and recently also in patients with COPD [18–20] and severe asthma [21, 22].

## Structure and mechanisms of actions of macrolides

The family of macrolide antibiotics is structurally characterised by a lactone ring containing at least 12 members, with erythromycin, clarithromycin and roxithromycin containing a 14-membered lactone ring, and azithromycin (also called an azalide) containing a 15-membered lactone ring with a tertiary amino group [23, 24]. The neomacrolides azithromycin, clarithromycin and roxithromycin have excellent bioavailability with superior oral absorption and better tissue penetration than erythromycin, accumulating preferentially in (alveolar) macrophages, with azithromycin having the longest serum half-life, making it suitable for once-daily or even intermittent dosing [25–27]. Due to toxicity concerns and scarceness of data on immunomodulatory effects, the ketolide telithromycin is less convenient for long-term use and will therefore not be discussed further in this chapter [28–30].

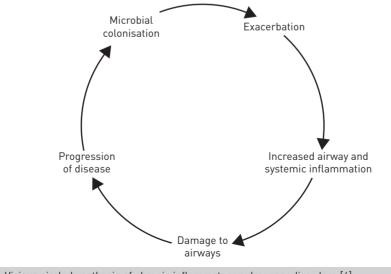


Figure 1. Vicious circle hypothesis of chronic inflammatory pulmonary disorders [6].

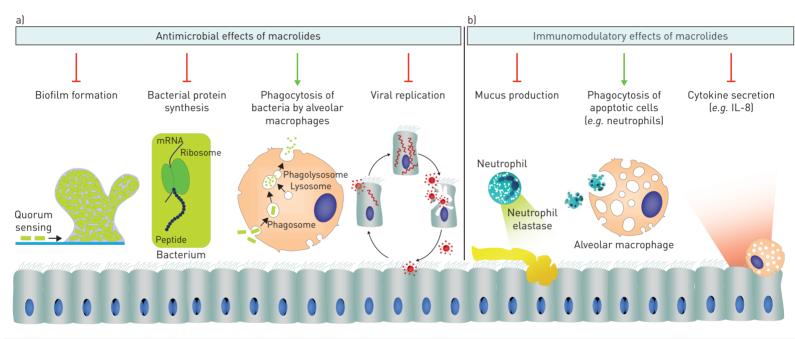


Figure 2. The antimicrobial and immunomodulatory effects of macrolides. a) Antimicrobial effects. Macrolides can directly affect bacteria by inhibiting bacterial protein synthesis, and reducing their adherence and toxin production. In addition, they are able to disrupt biofilm formation through suppression of quorum sensing proteins, in particular in the case of *Pseudomonas aeruginosa* infections. Macrolides stimulate phagocytosis of pathogenic bacteria by alveolar macrophages. In bronchial epithelial cells, they tend to induce antiviral host responses, thereby indirectly hampering viral replication. b) Immunomodulatory effects. Macrolides enhance mucociliary clearance by reducing airway secretions through inhibition of neutrophil elastase and modulation of mucin gene expression, and reduce chronic inflammation by stimulating phagocytosis of apoptotic cells and impairing the production of pro-inflammatory mediators such as IL-8 in airway epithelial cells and alveolar macrophages [53].

## Antimicrobial effects of macrolides

The direct antibiotic effect of macrolides is exerted through binding to the bacterial 50S ribosomal subunit, thereby interfering with its assembly and ultimately inhibiting bacterial protein synthesis, leading to a bacteriostatic effect [25, 31, 32]. Their antibacterial spectrum primarily includes the atypical bacteria *Chlamydophila pneumoniae*, *Mycoplasma pneumoniae* and *Legionella pneumophila*, and, although resistance is on the increase, Gram-positives such as *Streptococcus pneumoniae*, while the neomacrolides also have a better Gram-negative coverage, including susceptible strains of *Haemophilus influenzae* and *Moraxella catarrhalis* [25, 33, 34]. The neomacrolides clarithromycin and azithromycin have excellent tissue penetration, and hence form the backbone in first-line combination treatment regimens for NTM such as MAC [35, 36]. Acquired resistance to macrolides occurs through alterations in their binding site on the bacterial ribosomal RNA induced by methylases encoded by erythromycin ribosome methylase (*erm*) genes or through macrolide efflux (*mef*) genes encoding for active efflux pumps reducing intrabacterial macrolide concentrations [37, 38].

Indirect antimicrobial effects are established by stimulation of phagocytosis of bacteria by alveolar macrophages [39, 40], and *via* inhibition of quorum sensing and biofilm formation, mechanisms which have been proven particularly useful in reducing the virulence of *Pseudomonas aeruginosa* and facilitating the effects of antipseudomonal antibiotics [41–43]. At subinhibitory concentrations, azithromycin interferes with cell–cell communication through inhibition of the guanosine diphosphomannose dehydrogenase enzyme in the alginate biosynthetic pathway of mucoid *P. aeruginosa* strains, thereby mitigating biofilm production [44]. Interestingly, an increase in the expression level of type III secretion system genes, encoding for virulence factors in *P. aeruginosa*, has also been observed *in vitro*, but up to now without any clinical correlate [44, 45].

Supported by the observation that erythromycin significantly reduced the number of common colds in COPD patients [46], potential antiviral properties of macrolides have also been unravelled more recently [47–49]. *In vitro*, azithromycin decreases viral load in bronchial epithelial cells infected with rhinovirus [49]. Its antiviral effects are possibly mediated by induction of pattern recognition receptors, IFNs and IFN-stimulated genes, leading to a global amplification of the host antiviral response to human rhinoviruses [47, 48]. The antimicrobial effects of macrolides are summarised in figure 2a.

Immunomodulatory effects of macrolides

Long-term treatment courses with 14- and 15-membered-ring macrolides are known to influence chronic airway diseases mediated by neutrophilic inflammation [50–53]. In contrast to other antibiotics that have been attributed immunomodulatory effects, macrolides are known to alter a plenitude of cells within the airway, including neutrophils, alveolar macrophages, lymphocytes and epithelial cells [54, 55]. The pronounced intracellular accumulation and retention of macrolides, and in particular of azithromycin, allows prolongation of their effects within host cells [56, 57]. After an initial stimulation of neutrophil degranulation and phagocytosis-associated oxidative burst enhancing their antibacterial activity, late effects of macrolides include increased neutrophil apoptosis and attenuation of oxidative burst responses [55, 56]. Neutrophilic inflammation is further hampered by macrolide antibiotics through decreased production of chemoattractants and

decreased expression of adhesion molecules, resulting in attenuation of chemotaxis [58, 59]. Azithromycin may also promote differentiation of monocytoid cell lines into macrophages and alter the macrophage phenotype, leading to reduced secretion of pro-inflammatory cytokines and increased production of anti-inflammatory mediators [60–62]. In addition, macrolides stimulate phagocytosis of apoptotic cells by alveolar macrophages [39, 40], attenuate type 1 T-helper cell (Th1) responses following lipopolysaccharide (LPS)- or IFN- $\gamma$ -induced stimulation of macrophages, and (although conflicting observations have been reported) seem to affect the balance between Th1 and Th2 responses [63–65].

Additional modulation of host defence occurs through interaction with structural cells, such as bronchial epithelial cells, with beneficial effects on the stability of the epithelial barrier and ciliary function [54, 55]. Macrolide-induced inhibition of neutrophilic elastase and matrix metalloproteinases [66–68], and of respiratory epithelium cytokine production [56, 68, 69], is well established. Most of these effects are probably caused by modulation of mitogen-activated protein kinase and NF- $\kappa$ B signalling pathways [54]. In human airways, epithelial goblet cells and mucous cells synthesise gel-forming mucins such as MUC5AC and MUC5B [54]. Erythromycin and clarithromycin have been shown to inhibit this TNF- $\alpha$ -induced mucus secretion, resulting in improved mucociliary clearance and hence beneficial effects in various clinical conditions characterised by excessive sputum production [51, 70, 71]. The immunomodulatory effects of macrolides are summarised in figure 2b.

# Dosing of macrolides

The long half-life and interesting safety profile makes azithromycin the agent of choice for long-term use. When used for its immunomodulatory purposes, azithromycin is typically dosed as 500 mg three times weekly on Monday, Wednesday and Friday (MWF), or alternatively by a daily 250 mg dose, although the AZISAST (Azithromycin in severe asthma) study has shown efficacy in noneosinophilic asthma at lower doses of 250 mg three times weekly [21]. In case of intolerance of the usual dosage, lower doses of 250 mg MWF are worth considering as, although not widely studied, they might have the advantage of inducing less adverse events without losing efficacy. In case of obstinate gastrointestinal intolerance of azithromycin, roxithromycin can be used as an alternative, at a daily dose of 150 mg [72]. As for clarithromycin, most clinical trials have been performed with doses of 500 mg twice daily, although doses as low as 200 mg daily have also proven effective [68, 73].

# Other antibiotics

In addition to macrolides, other antibiotics have been investigated, although less intensively, for maintenance treatment in chronic inflammatory airway diseases. Doxycycline and other tetracyclines have been shown to regulate the host immune response, *e.g.* by targeting matrix metalloproteinases released from neutrophils and attenuating LPS-induced inflammation [8, 74, 75]. Immunomodulatory effects have also been attributed to fluoroquinolones, with moxifloxacin selectively inhibiting secretion of cytokines such as IL-8 and IL-6, and NF- $\kappa$ B activation in CF epithelial cell lines, and ciprofloxacin providing anti-inflammatory properties in the setting of LPS-induced lung injury in animal models [76, 77]. In *Stenotrophomonas maltophilia* infection, trimethoprim–sulfamethoxazole (cotrimoxazole) has been shown to suppress TNF- $\alpha$  production by human peripheral blood mononuclear cells [78]. Accumulating *in vitro* evidence suggests that cotrimoxazole is able

to enhance neutrophil chemotaxis, phagocytosis and intracellular killing by macrophages, to reduce lymphocytic proliferation [79]. As mentioned earlier, this chapter will elaborate mainly on the clinical applications of macrolides as only scarce data on the long-term use of other oral antibiotic treatment classes are available.

# Long-term oral antibiotic treatment: when and to whom?

## Diffuse panbronchiolitis

Diffuse panbronchiolitis is an idiopathic inflammatory disease of the respiratory bronchioles, almost exclusively occurring in Japan and other East Asian countries [80, 81]. This progressive disease is characterised by neutrophilic airway inflammation with elevated pro-inflammatory cytokines such as IL-8 and IL-1β, and often complicated by H. influenzae and/or P. aeruginosa infection [82, 83]. The prognosis of this possibly fatal disease was significantly improved by the introduction of erythromycin, leading to the suspicion of beneficial effects of erythromycin other than its antibacterial activity [10, 84]. Treatment with macrolides has been shown to decrease BAL fluid levels of neutrophils, IL-8 and pro-inflammatory  $\beta$ -defensing [83, 85, 86]. Nevertheless, most evidence came from retrospective or prospective open trials, suggesting not only efficacy of erythromycin [10, 84, 87, 88], but also of clarithromycin, azithromycin and roxithromycin [73, 89–91]. Only one small RCT was reported examining serial CT scans [92]. As the overt clinical of macrolide treatment precludes placebo-controlled trials in diffuse success panbronchiolitis, the authors of a systematic review instead recommended setting up comparative RCTs between different kinds or doses of macrolides in the future [93].

## Cystic fibrosis

CF is a multisystem, autosomal recessive genetic disease, caused by a defect in the CF transmembrane conductance regulator (CFTR) gene. Its pulmonary manifestations are characterised by recurrent airway infections, chronic inflammation and progressive lung function decline, resulting in limited life expectancy [94, 95]. However, the prognosis is gradually improving due to the introduction of medication directly targeted to the genetic cause of the disease, as well as treatments directed towards infection and inflammation [96, 97]. Frequent pulmonary exacerbations are known to be associated with increased mortality. Macrolides, which in CF will mainly have anti-inflammatory and antibacterial working mechanisms by inhibiting biofilm formation (figure 2), have been investigated for their potential to improve lung function in CF in five large RCTs (table 1) [11-14, 98]. Forced expiratory volume in 1 s (FEV1) improved in two studies and azithromycin reduced FEV1 decline in one study. Although two other trials did not show an effect of macrolides on FEV1, a reduction in the number of exacerbations was established. The 2012 Cochrane review on macrolide maintenance therapy in CF stated that 6-month azithromycin treatment courses can result in improved FEV1 and reduced exacerbation rates, but that it is not clear whether this effect will be sustained [99]. What is the effect of treatment with macrolides longer than 12 months [100]? This important issue has recently been addressed in a retrospective study showing that in the 2 years following the first year of azithromycin treatment, the exacerbation rate increased again to levels similar to the period before starting the macrolide [101].

First author [ref.]	Inclusion criteria	Intervention	Length of treatment	Primary outcome	Number of exacerbations
CF					
Saiman [12]	Age ≥6 years; weight ≥25 kg; chronic infection with <i>Pseudomonas</i> <i>aeruginosa</i>	Azithromycin 250 or 500 mg (weight <40 or ≥40 kg) three times weekly	168 days	Significantly improved FEV1	Significantly reduced (hazard ratio 0.65)
Clement [11]	Age 6-21 years	Azithromycin 250 or 500 mg (weight <40 or ≥40 kg) three times weekly	12 months	No significant difference in FEV1	Significantly reduced (rate ratio 0.5)
Εαυι [13]	Age 8–18 years	Azithromycin 250 or 500 mg (weight <40 or ≥40 kg) once daily	6 months	Significantly improved FEV1	No significant difference
Wolter [14]	Age ≥18 years	Azithromycin 250 mg once daily	3 months	Reduced rate of FEV1 decline	No significant difference
Saiman [98]	Age 6–18 years; weight ≽18 kg; no <i>Pseudomonas</i> infection	Azithromycin 250 or 500 mg (weight <36 or ≥36 kg) three times weekly	168 days	No significant difference in FEV1	50% reduction in exacerbations (hazard ratio 0.5)
Non-CF bronchiectasis		,			
Wong [15]	Age ≥18 years; ≥1 exacerbation requiring antibiotic(s) in preceding year	Azithromycin 500 mg three times weekly	6 months	Significantly reduced number of exacerbations (rate ratio 0.38)	
Serisier [17]	Age 20–85 years; ≥2 exacerbations requiring antibiotic(s) in preceding year	Erythromycin 400 mg twice daily	48 weeks	Significantly reduced number of exacerbations (rate ratio 0.57)	
Altenburg [16]	Age ≥18 years; ≥3 LRTIs requiring antibiotic(s) in preceding year; ≥1 sputum culture yielding ≥1 bacterial pathogen in preceding year	Azithromycin 250 mg once daily	52 weeks	Significantly reduced number of exacerbations (rate ratio 0.41)	

Table 1. Randomised double-blind placebo-controlled trials with long-term macrolides in cystic fibrosis (CF) and non-CF bronchiectasis

FEV1: forced expiratory volume in 1 s.

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## To whom?

As macrolides have a positive effect on lung function and exacerbation frequency, it can be suggested to start this treatment in CF patients showing lung function decline or experiencing frequent exacerbations.

## Non-CF bronchiectasis

Bronchiectasis is defined as the presence of dilated thick-walled bronchi caused by chronic bronchial inflammation and recurrent airway infections [4]. Many aetiologies have been identified, with the most common causes described in the European Bronchiectasis Registry (EMBARC) being idiopathic (39%) and post-infective (27%). Long-term oral antibiotics were used in 19.4%, consisting mainly (in 74.2%) of azithromycin [102]. One RCT evaluated amoxicillin 3 g twice daily for 32 weeks in bronchiectasis patients expectorating daily purulent sputum, which resulted in a reduction in 24-h sputum volume and purulence, and in symptom awareness [103]. Most recent trials have, however, been examining macrolides mainly based on their anti-inflammatory properties and antibacterial effects by inhibiting biofilm formation (figure 2). The three main randomised double-blind placebo-controlled studies investigating the effect of azithromycin in non-CF bronchiectasis have shown significant reductions in pulmonary exacerbations (table 1) [15–17]. In the trial conducted by WONG et al. [15], the benefits of macrolide treatment persisted for 6 months after completion of treatment. SERISIER et al. [17] showed that low-dose erythromycin also decreased sputum production, with a beneficial effect on lung function, and increased eradication of sputum pathogens.

## To whom?

Based on the three trials [15–17], expert opinion supports macrolide maintenance treatment in patients with frequent exacerbations (at least two exacerbations in the previous year) provided their airway clearance techniques have been already optimised. Patients who are stable with good self-reported quality of life and less pulmonary complaints should not be treated with long-term azithromycin [104, 105].

## COPD

COPD is defined by progressive airflow obstruction that is poorly reversible and intercurrent acute exacerbations, which have an impact on disease course and mortality risk [3, 106, 107]. Increasing evidence supports an important role for the lung microbiome in the pathogenesis of COPD [108]. Exacerbations are often caused by infectious microorganisms, accompanied by airway and systemic inflammation [109]. Prevention of exacerbations is therefore essential to inhibit further disease progression. During exacerbations, microbiome shifts have been observed, as well as differences related to the exacerbation phenotypes described as bacterial or eosinophilic, with increases in Proteobacteria (mainly *Haemophilus* and *Moraxella* spp.) or Firmicutes (mainly *Streptococcus* spp.), respectively [108, 110]. A UK retrospective cohort study demonstrated that maintenance antibiotic treatment ( $\geq 6$  months) had been prescribed only for a small proportion of COPD patients (0.61%). The antibiotics most often used were oxytetracycline, doxycycline and penicillin, with a rise in macrolide prescriptions from 2005 [111].

Many antibiotics have been assessed for long-term treatment in COPD. As early as 1956, an observational study published in *The Lancet* showed overall improvement in 60% of

patients with chronic bronchitis receiving tetracyclines during 6 months in winter. *H. influenzae* was present in sputum cultures in almost all patients [112]. In contrast, a recent RCT examining a 3-week course of doxycycline in stable COPD patients was not able to show any anti-inflammatory effects [113]. Another observational study published in 1975 showed that an improvement in sputum production and dyspnoea was seen in 50% of COPD patients with chronic sputum production receiving trimethoprim–sulfamethoxazole 960 mg twice daily during 3 months [114]. Recently, intermittent pulsed therapy with moxifloxacin 400 mg once daily for 5 days, repeated each 8 weeks during 48 weeks, reduced the exacerbation rate in COPD patients in an RCT [115].

In addition to their anti-inflammatory functions, macrolides can play a major role in prevention of exacerbations due to their antibacterial and antiviral properties (figure 2). Three major studies have investigated the role of macrolides in reducing exacerbations (table 2). While UZUN *et al.* [18] included patients with frequent exacerbations, patients with nonfrequent exacerbations were also allowed to be enrolled in the other two studies [19, 20]. Macrolides reduced the number of exacerbations in all three trials [18–20]. The beneficial effect of macrolides can be partially explained by the presence of bronchiectasis in patients with COPD. MARTÍNEZ-GARCÍA *et al.* [116] showed that bronchiectasis was present in almost 60% of the COPD patients. In the COLUMBUS (COPD: influence of macrolides on exacerbation frequency in patients) trial, CT scans were performed to exclude patients with bronchiectasis [18]. A multicentre RCT examining the effect of short-term macrolide treatment in acute COPD exacerbations is currently ongoing [117].

## To whom?

Criteria were proposed for selecting patients for long-term azithromycin treatment. In two reviews, the authors recommend to prescribe macrolides in patients with frequent exacerbations (at least two in the year before treatment) [17, 118]. The most pronounced effect of azithromycin was seen in patients of older age, in those classified in lower Global Initiative for Chronic Obstructive Lung Disease (GOLD) grade and in those not actively smoking [119]. However, this analysis was performed on data which also involved patients with nonfrequent exacerbations.

## Severe noneosinophilic asthma

Severe asthma is a heterogeneous syndrome characterised by chronic airway inflammation, either of eosinophilic nature or of noneosinophilic origin (neutrophilic or paucigranulocytic) [120]. In parallel to the chronic neutrophilic airway diseases mentioned previously, the effects of low-dose macrolides have also been examined in patients with severe noneosinophilic asthma [53]. Initially, the observed beneficial effects of macrolides in asthma were attributed to their antimicrobial properties directed towards intracellular pathogens, such as *Chlamydophila* and *M. pneumoniae* [121, 122]. As with respiratory viral infections, infection with and/or reactivation of these atypical bacteria has been associated with both asthma exacerbations and chronic severe asthma [123–125]. Increased susceptibility to infection in asthma probably leads to increased atypical bacterial infection (mainly with *C. pneumoniae*), which itself then further increases neutrophilic airway inflammation by eliciting acute antibody responses such as IgA, leading to aggravation of asthmatic symptoms [124, 125]. Although a partial contribution of their bacteriostatic effects cannot be excluded, it is widely assumed that the immunomodulatory properties of macrolides are the predominant mechanism of action [126, 127]. Whereas shorter-term trials were only able to show some

First author [ref.]	Inclusion criteria	Intervention	Length of treatment	Primary outcome
COPD				
Uzun [18]	Age ≥18 years; ≥3 exacerbations in the preceding year	Azithromycin 500 mg three times weekly	12 months	Significantly reduced number of exacerbations (rate ratio 0.60)
Albert [19]	Age ≥40 years; FEV1 <80% predicted; either use of continuous supplemental oxygen or receipt of systemic corticosteroids within the previous year	Azithromycin 250 mg once daily	12 months	Significantly reduced number of exacerbations (rate ratio 0.83)
SEEMUNGAL [20]	FEV1 30–70% predicted; no exacerbation required	Erythromycin 250 mg twice daily	12 months	Significantly reduced number of exacerbations (rate ratio 0.65)
Severe asthma		-		
Brusselle [21]	Age 18–75 years; persistent asthma (GINA step 4 or 5); ≥2 severe asthma exacerbations requiring systemic corticosteroids and/or LRTIs requiring antibiotic(s) within previous year	Azithromycin 250 mg three times weekly	6 months	No difference in rate of severe exacerbations and LRTIs requiring antibiotic(s); significantly reduced number of LRTIs and exacerbations in predefined noneosinophilic severe asthma subgroup (rate ratio 0.43)
Hahn [22]	Age ≥18 years; persistent asthma	Azithromycin 600 mg weekly	12 weeks (1 year follow-up)	No difference in symptom control, except for open-label group with more severe asthma; not powered to detect differences in exacerbation rate

Table 2. Randomised double-blind placebo-controlled trials with long-term macrolides in COPD and severe asthma

FEV1: forced expiratory volume in 1 s; GINA: Global Initiative for Asthma.

effect of macrolides on reduction of symptoms and improvement of lung function in severe asthma, insufficient to recommend their routine use for asthma control [68, 128–132], the AZISAST study was the first to examine the prevention of exacerbations as a primary end-point in adults with exacerbation-prone severe asthma receiving long-term (6 months) macrolide maintenance therapy (table 2) [21]. In contrast, clinical trials on short-term macrolide treatment in acute asthma exacerbations yielded conflicting results [133–135].

## To whom?

In the AZISAST randomised double-blind placebo-controlled trial, low-dose (250 mg three times weekly) azithromycin treatment was only associated with a significant lower exacerbation rate in the predefined subgroup of patients with noneosinophilic severe asthma [21]. The need for a careful selection of patients can also be derived from the AZMATICS (Azithromycin/asthma trial in community settings) trial, where a 12-week macrolide treatment course only showed clinical improvement in an open-label study arm encompassing individuals with severe treatment-resistant or refractory asthma [22]. However, as a recent Cochrane review points out, there is still a need for additional high-quality studies to confirm the possible benefit of macrolides in noneosinophilic severe asthma [136].

# Other indications

Azithromycin has also proven useful in bronchiolitis obliterans syndrome (BOS) after lung transplantation [137, 138]. This major complication of lung transplantation is a form of chronic rejection associated with poor survival, defined as a delayed allograft dysfunction with persistent decline in FEV1 without any other known and potentially reversible cause [139, 140]. After encouraging data showing improvements in FEV1 in observational studies [141, 142], CORRIS *et al.* [143] have recently shown efficacy of azithromycin treatment in improving FEV1 in lung allograft BOS in an RCT. In lung transplant recipients, azithromycin prophylaxis might even prevent the occurrence of BOS [144, 145]. These findings might also be extrapolated to the similar condition of BOS complicating haematopoietic stem cell transplantation; however, the scarce and rather small clinical trials available have not been able to show a significant benefit of azithromycin in this population [146–148].

Organising pneumonia, either cryptogenic, or induced by infections, drug toxicity, vasculitis or other conditions, is an inflammatory disorder characterised by buds of granulation tissue in the peripheral airways and alveoli, preferably treated with systemic corticosteroids [149]. On a case-by-case basis, chronic macrolide treatment may be adjuvant to steroid therapy or act as a steroid-sparing strategy, probably through inhibition of IL-8 release and neutrophil accumulation in the distal airways [150–154].

# Long-term oral antibiotic treatment: is it safe?

# **Drug interactions**

Drug interactions of macrolides are mainly mediated by inhibition of hepatic cytochrome CYP (P450) 3A enzymes, but are least pronounced for azithromycin compared with clarithromycin and erythromycin [155]. Through inhibition of CYP3A4, clarithromycin and erythromycin have the propensity of inducing toxic concentrations of a long list of substrates, including most statins, calcium channel blockers, amiodarone and colchicine. As an inhibitor of glycoprotein P, however, azithromycin and the other neomacrolides are able

to increase plasma concentrations of various substrates such as digoxin, everolimus, sirolimus, tacrolimus and posaconazole. All macrolides can cause altered warfarin levels to various extent. Interactions with concomitant medication should therefore always be checked before treatment initiation.

#### Adverse reactions

To date, no RCTs have thoroughly assessed the long-term safety of chronic macrolide therapy. Typical side-effects include gastrointestinal discomfort such as nausea, diarrhoea and abdominal pain, induced by dose-related effects on motilin receptors [156]. These mild-to-moderate symptoms seldom lead to drug discontinuation (reviewed in [157]) and are less frequently reported with the neomacrolides than with erythromycin [158, 159]. Within the neomacrolides, azithromycin and roxithromycin tend to induce fewer gastrointestinal complaints than clarithromycin.

While severe hepatotoxicity is rare during azithromycin treatment, transient cholestasis or other abnormal liver function tests have been described, although less frequently than with erythromycin [158–160]. Its propensity to induce severe hepatotoxicity excludes telithromycin from long-term treatment indications [28–30]. During long-term azithromycin treatment, monitoring of serum liver enzymes is advisable and treatment should be discontinued immediately if signs or symptoms of hepatitis occur.

The incidence of cardiac events has mainly been monitored in clinical trials on short-term macrolide use [161, 162]. In particular, macrolide antibiotics are known to potentially induce QT prolongation and torsades de pointes [161, 163]. Hence, azithromycin still needs to be used with caution in patients with pre-existing cardiovascular disease and risk factors [164, 165]. Before treatment initiation, measurement of the QT interval by electrocardiography is recommended, in particular in the case of polypharmacy or concomitant treatment with other QT-prolonging drugs such as fluoroquinolones.

Macrolide-induced ototoxicity typically affects the lower (speech) frequencies and is usually characterised by a bilateral sensorineural hearing loss, often reversible [157]. This rare side-effect is dose dependent and facilitated by risk factors such as renal impairment, as was demonstrated in one prospective case-control study in pneumonia patients treated with intravenous erythromycin [166]. Azithromycin-induced hearing loss has been reported in case of long-term high dosage (*e.g.* 500 mg daily) and mostly within a context of treatment of NTM infections [167, 168]. However, its incidence during low-dose long-term treatment still needs to be established [157], although one study in COPD patients observed more hearing decrements in participants receiving azithromycin than in those receiving placebo [19]. While routine audiometric screening is not considered mandatory, some authors suggest ordering audiometry before initiation of therapy in risk groups [169].

Long-term macrolide treatment will undoubtedly have an influence on the respiratory microbiome. A subanalysis of the BLESS (Bronchiectasis and low-dose erythromycin study) trial showed that in patients with non-CF bronchiectasis who were not colonised with *P. aeruginosa*, erythromycin caused a shift in the microbiome from a dominant presence of *H. influenzae* to an increase in concentration of *P. aeruginosa* [17]. Further research on this important area needs to be performed to investigate whether this change in the microbiome will also influence the clinical outcome.

## Macrolide resistance

Prescription of macrolides has been associated with emerging resistance, mainly in *Streptococcus* spp. and *Staphylococcus* spp. [170–174]. Unfortunately, few RCTs assessing long-term macrolide maintenance therapy have monitored for resistance induction. In CF patients, increased macrolide resistance has been observed during long-term macrolide treatment in *Staphylococcus aureus* and *H. influenzae*, particularly after multiple years of therapy [175–178]. As the clinical benefit of azithromycin in CF seems to be limited to the first 6–12 months of treatment [101], implementing shorter treatment courses and periods of treatment interruption might help to tackle resistance development. Similarly, a meta-analysis of RCTs in non-CF bronchiectasis documented an increased risk of macrolide resistance in *H. influenzae*, *S. aureus* and *S. pneumoniae* [179], while COPD trials did not find increased appearance of macrolide-resistant organisms after 3–12 months of therapy [20, 180].

Although macrolide resistance is increasing among pneumococcal isolates, literature on its clinical significance in severe infections such as pneumococcal pneumonia is inconsistent, but does not show a significant increase in mortality [181–184]. A recent retrospective study in a tertiary care hospital found no evidence suggesting that adult patients hospitalised with macrolide-resistant pneumococcal pneumonia experienced a more severe clinical presentation or worse clinical outcome than those with positive cultures for macrolide-susceptible *S. pneumoniae* [185]. From a population point of view, however, it remains important to restrict chronic macrolide use to those patients who will benefit the most in view of the increasing resistance data.

The occurrence of macrolide resistance is facilitated by macrolide monotherapy in the treatment of MAC lung disease and macrolide susceptibility is key to treatment success [186]. As azithromycin is able to block autophagosome clearance by preventing lysosomal acidification in primary human macrophages, leading to impairment of autophagy and, consequently, inhibition of intracellular killing of mycobacteria, some authors fear the induction of (macrolide-resistant) mycobacterial infection during long-term azithromycin use [187]. However, a nested case-control study in one large CF centre recently showed a reduced risk of NTM in CF adults receiving long-term azithromycin, making a plea for its use as primary prophylaxis against NTM [188]. Nevertheless, before initiating macrolide treatment, ideally three sputum samples should be delivered to exclude pre-existing NTM disease in predisposing conditions such as CF and bronchiectasis. Furthermore, current guidelines advise not to continue azithromycin treatment if a positive NTM culture is obtained in CF patients, unless as part of a multidrug treatment regimen in the context of NTM disease [189]. Recommendations for long-term macrolide treatment monitoring are summarised in table 3.

# Conclusions

The pleiotropic effects of macrolides such as erythromycin, clarithromycin and azithromycin encompass anti-inflammatory and immunomodulatory capacities, in addition to their antimicrobial effects. The beneficial effects of macrolides have almost unequivocally been accepted in chronic inflammatory airway diseases such as diffuse panbronchiolitis, CF, non-CF bronchiectasis, COPD and severe noneosinophilic asthma (table 4). However, when macrolides are widely used in the community, population antimicrobial resistance will possibly increase. Therefore, indications to start macrolides should be very carefully

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Adverse events	Timing	Recommendation		
Drug hypersensitivity Drug interactions Hepatotoxicity	At start Any time At start At 6 weeks	Absence of known allergy to macrolides Verify concomitant medication Aminotransferase levels <3× upper limit of normal Monitor serum liver enzymes		
	Yearly	Monitor liver enzymes (more frequently in risk groups)		
Cardiotoxicity	At start	Perform electrocardiography to assess corrected QT interval (<450 ms)		
	Any time	Repeat electrocardiography if new concomitant medication influencing QT duration		
Ototoxicity	At start	Audiometric screening for pre-existing hearing loss in risk groups ( <i>e.g.</i> elderly)		
NTM infection	At start	Perform sputum samples to exclude pre-existing NTM infection		
	6 monthly	Monitor sputum cultures for NTM (particularly in risk groups, <i>e.g.</i> cystic fibrosis)		

#### Table 3. Recommendations for long-term macrolide treatment monitoring

Table 4. Summary of considerations for chronic immunomodulatory therapy with macrolides

Airway disease	Subgroup of patients with expected benefit		
Diffuse panbronchiolitis			
CF	Patients with FEV1 decline and/or frequent (infectious) exacerbations, particularly in case of chronic <i>Pseudomonas aeruginosa</i> infection		
Non-CF bronchiectasis	Patients with frequent (infectious) exacerbations		
COPD	Patients with frequent (infectious) exacerbations		
Asthma	Noneosinophilic severe asthmatic patients with frequent exacerbations		
Bronchiolitis obliterans	Bronchiolitis obliterans syndrome in lung transplant recipients		
Organising pneumonia	Corticosteroid-dependent patients		

CF: cystic fibrosis; FEV1: forced expiratory volume in 1 s.

considered, particularly when used in common diseases such as COPD and asthma, and well monitored (table 3). Azithromycin 500 mg three times weekly (MWF) is the regimen of first choice. In general, long-term macrolides should be considered in the described diseases when patients present with at least two exacerbations per year despite optimal treatment with the ordinary medications as recommended in the relevant guidelines.

## References

- 1. Seemungal TA, Donaldson GC, Paul EA, *et al.* Effect of exacerbation on quality of life in patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 1998; 157: 1418–1422.
- Miravitlles M, Ferrer M, Pont A, *et al.* Effect of exacerbations on quality of life in patients with chronic obstructive pulmonary disease: a 2 year follow up study. *Thorax* 2004; 59: 387–395.
- 3. Donaldson GC, Seemungal TAR, Bhowmik A, *et al.* Relationship between exacerbation frequency and lung function decline in chronic obstructive pulmonary disease. *Thorax* 2002; 57: 847–852.

- Pasteur MC, Bilton D, Hill AT, et al. British Thoracic Society guideline for non-CF bronchiectasis. Thorax 2010; 65: Suppl. 1, i1–i58.
- 5. Cantin AM, Hartl D, Konstan MW, *et al.* Inflammation in cystic fibrosis lung disease: pathogenesis and therapy. *J Cyst Fibros* 2015; 14: 419–430.
- 6. Cole PJ. Inflammation: a two-edged sword the model of bronchiectasis. Eur J Respir Dis Suppl 1986; 147: 6–15.
- Sethi S, Murphy TF. Infection in the pathogenesis and course of chronic obstructive pulmonary disease. N Engl J Med 2008; 359: 2355–2365.
- 8. Rempe S, Hayden JM, Robbins RA, *et al.* Tetracyclines and pulmonary inflammation. *Endocr Metab Immune Disord Drug Targets* 2007; 7: 232–236.
- Bode C, Diedrich B, Muenster S, *et al.* Antibiotics regulate the immune response in both presence and absence of lipopolysaccharide through modulation of Toll-like receptors, cytokine production and phagocytosis *in vitro*. *Int Immunopharmacol* 2014; 18: 27–34.
- 10. Kudoh S, Azuma A, Yamamoto M, *et al.* Improvement of survival in patients with diffuse panbronchiolitis treated with low-dose erythromycin. *Am J Respir Crit Care Med* 1998; 157: 1829–1832.
- 11. Clement A, Tamalet A, Leroux E, *et al.* Long term effects of azithromycin in patients with cystic fibrosis: a double blind, placebo controlled trial. *Thorax* 2006; 61: 895–902.
- 12. Saiman L, Marshall BC, Mayer-Hamblett N, *et al.* Azithromycin in patients with cystic fibrosis chronically infected with *Pseudomonas aeruginosa*: a randomized controlled trial. *JAMA* 2003; 290: 1749–1756.
- 13. Equi A, Balfour-Lynn IM, Bush A, *et al.* Long term azithromycin in children with cystic fibrosis: a randomised, placebo-controlled crossover trial. *Lancet* 2002; 360: 978–984.
- 14. Wolter J, Seeney S, Bell S, *et al.* Effect of long term treatment with azithromycin on disease parameters in cystic fibrosis: a randomised trial. *Thorax* 2002; 57: 212–216.
- 15. Wong C, Jayaram L, Karalus N, *et al.* Azithromycin for prevention of exacerbations in non-cystic fibrosis bronchiectasis (EMBRACE): a randomised, double-blind, placebo-controlled trial. *Lancet* 2012; 380: 660–667.
- Altenburg J, de Graaff CS, Stienstra Y, *et al.* Effect of azithromycin maintenance treatment on infectious exacerbations among patients with non-cystic fibrosis bronchiectasis: the BAT randomized controlled trial. *JAMA* 2013; 309: 1251–1259.
- 17. Serisier DJ, Martin ML, McGuckin MA, *et al.* Effect of long-term, low-dose erythromycin on pulmonary exacerbations among patients with non-cystic fibrosis bronchiectasis: the BLESS randomized controlled trial. *JAMA* 2013; 309: 1260–1267.
- Uzun S, Djamin RS, Kluytmans JAJW, et al. Azithromycin maintenance treatment in patients with frequent exacerbations of chronic obstructive pulmonary disease (COLUMBUS): a randomised, double-blind, placebo-controlled trial. *Lancet Respir Med* 2014; 2: 361–368.
- 19. Albert RK, Connett J, Bailey WC, *et al.* Azithromycin for prevention of exacerbations of COPD. *N Engl J Med* 2011; 365: 689–698.
- 20. Seemungal TAR, Wilkinson TMA, Hurst JR, *et al.* Long-term erythromycin therapy is associated with decreased chronic obstructive pulmonary disease exacerbations. *Am J Respir Crit Care Med* 2008; 178: 1139–1147.
- 21. Brusselle GG, Vanderstichele C, Jordens P, *et al.* Azithromycin for prevention of exacerbations in severe asthma (AZISAST): a multicentre randomised double-blind placebo-controlled trial. *Thorax* 2013; 68: 322–329.
- 22. Hahn DL, Grasmick M, Hetzel S, *et al.* Azithromycin for bronchial asthma in adults: an effectiveness trial. *J Am Board Fam Med* 2012; 25: 442–459.
- 23. Neu HC. New macrolide antibiotics: azithromycin and clarithromycin. Ann Intern Med 1992; 116: 517-519.
- 24. Mazzei T, Mini E, Novelli A, et al. Chemistry and mode of action of macrolides. J Antimicrob Chemother 1993; 31: Suppl. C, 1–9.
- 25. Sturgill MG, Rapp RP. Clarithromycin: review of a new macrolide antibiotic with improved microbiologic spectrum and favorable pharmacokinetic and adverse effect profiles. *Ann Pharmacother* 1992; 26: 1099–1108.
- 26. Ballow CH, Amsden GW. Azithromycin: the first azalide antibiotic. Ann Pharmacother 1992; 26: 1253-1261.
- 27. Nilsen OG. Pharmacokinetics of macrolides. Comparison of plasma, tissue and free concentrations with special reference to roxithromycin. *Infection* 1995; 23: Suppl. 1, S5–S9.
- 28. File TM. Telithromycin new product overview. J Allergy Clin Immunol 2005; 115: S1-S13.
- 29. Ross DB. The FDA and the case of Ketek. N Engl J Med 2007; 356: 1601–1604.
- 30. Brinker AD, Wassel RT, Lyndly J, et al. Telithromycin-associated hepatotoxicity: clinical spectrum and causality assessment of 42 cases. *Hepatology* 2009; 49: 250–257.
- 31. Rapp RP, McCraney SA, Goodman NL, *et al.* New macrolide antibiotics: usefulness in infections caused by mycobacteria other than *Mycobacterium tuberculosis. Ann Pharmacother* 1994; 28: 1255–1263.
- 32. Champney WS, Tober CL, Burdine R. A comparison of the inhibition of translation and 50S ribosomal subunit formation in *Staphylococcus aureus* cells by nine different macrolide antibiotics. *Curr Microbiol* 1998; 37: 412–417.
- 33. Retsema J, Girard A, Schelkly W, *et al.* Spectrum and mode of action of azithromycin (CP-62,993), a new 15-membered-ring macrolide with improved potency against gram-negative organisms. *Antimicrob Agents Chemother* 1987; 31: 1939–1947.

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- 34. Ferrara A, Santos Dos C, Cimbro M, *et al.* Comparative antimicrobial activity and post-antibiotic effect of azithromycin, clarithromycin and roxithromycin against some respiratory pathogens. *Int J Antimicrob Agents* 1996; 7: 181–186.
- 35. Klemens SP, Cynamon MH. Activities of azithromycin and clarithromycin against nontuberculous mycobacteria in beige mice. *Antimicrob Agents Chemother* 1994; 38: 1455–1459.
- 36. Griffith DE, Aksamit T, Brown-Elliott BA, *et al.* An official ATS/IDSA statement: diagnosis, treatment, and prevention of nontuberculous mycobacterial diseases. *Am J Respir Crit Care Med* 2007; 175: 367–416.
- 37. Shortridge VD, Doern GV, Brueggemann AB, *et al.* Prevalence of macrolide resistance mechanisms in *Streptococcus pneumoniae* isolates from a multicenter antibiotic resistance surveillance study conducted in the United States in 1994–1995. *Clin Infect Dis* 1999; 29: 1186–1188.
- 38. Leclercq R. Mechanisms of resistance to macrolides and lincosamides: nature of the resistance elements and their clinical implications. *Clin Infect Dis* 2002; 34: 482–492.
- Hodge S, Hodge G, Brozyna S, et al. Azithromycin increases phagocytosis of apoptotic bronchial epithelial cells by alveolar macrophages. Eur Respir J 2006; 28: 486–495.
- 40. Hodge S, Hodge G, Jersmann H, *et al.* Azithromycin improves macrophage phagocytic function and expression of mannose receptor in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2008; 178: 139–148.
- 41. Tateda K, Comte R, Pechere JC, et al. Azithromycin inhibits quorum sensing in *Pseudomonas aeruginosa*. *Antimicrob Agents Chemother* 2001; 45: 1930–1933.
- 42. Hoffmann N, Lee B, Hentzer M, *et al.* Azithromycin blocks quorum sensing and alginate polymer formation and increases the sensitivity to serum and stationary-growth-phase killing of *Pseudomonas aeruginosa* and attenuates chronic *P. aeruginosa* lung infection in *Cftr<sup>-/-</sup>* mice. *Antimicrob Agents Chemother* 2007; 51: 3677–3687.
- 43. Lutz L, Pereira DC, Paiva RM, *et al.* Macrolides decrease the minimal inhibitory concentration of anti-pseudomonal agents against *Pseudomonas aeruginosa* from cystic fibrosis patients in biofilm. *BMC Microbiol* 2012; 12: 196.
- 44. Romero D, Traxler MF, López D, et al. Antibiotics as signal molecules. Chem Rev 2011; 111: 5492–5505.
- 45. Nalca Y, Jänsch L, Bredenbruch F, *et al.* Quorum-sensing antagonistic activities of azithromycin in *Pseudomonas aeruginosa* PAO1: a global approach. *Antimicrob Agents Chemother* 2006; 50: 1680–1688.
- 46. Suzuki T, Yanai M, Yamaya M, et al. Erythromycin and common cold in COPD. Chest 2001; 120: 730–733.
- 47. Gielen V, Johnston SL, Edwards MR. Azithromycin induces anti-viral responses in bronchial epithelial cells. *Eur Respir J* 2010; 36: 646–654.
- 48. Schögler A, Kopf BS, Edwards MR, *et al.* Novel antiviral properties of azithromycin in cystic fibrosis airway epithelial cells. *Eur Respir J* 2015; 45: 428–439.
- 49. Menzel M, Akbarshahi H, Bjermer L, *et al.* Azithromycin induces anti-viral effects in cultured bronchial epithelial cells from COPD patients. *Sci Rep* 2016; 6: 28698.
- 50. Scaglione F, Rossoni G. Comparative anti-inflammatory effects of roxithromycin, azithromycin and clarithromycin. J Antimicrob Chemother 1998; 41: Suppl. B, 47–50.
- 51. Rubin BK. Immunomodulatory properties of macrolides: overview and historical perspective. *Am J Med* 2004; 117: Suppl. 9A, 2S–4S.
- 52. Spagnolo P, Fabbri LM, Bush A. Long-term macrolide treatment for chronic respiratory disease. *Eur Respir J* 2013; 42: 239–251.
- 53. Brusselle GG, Joos G. Is there a role for macrolides in severe asthma? Curr Opin Pulm Med 2014; 20: 95-102.
- 54. Kanoh S, Rubin BK. Mechanisms of action and clinical application of macrolides as immunomodulatory medications. *Clin Microbiol Rev* 2010; 23: 590–615.
- 55. Parnham MJ, Erakovic Haber V, Giamarellos-Bourboulis EJ, *et al.* Azithromycin: mechanisms of action and their relevance for clinical applications. *Pharmacol Ther* 2014; 143: 225–245.
- Culić O, Eraković V, Cepelak I, *et al.* Azithromycin modulates neutrophil function and circulating inflammatory mediators in healthy human subjects. *Eur J Pharmacol* 2002; 450: 277–289.
- 57. Bosnar M, Kelnerić Z, Munić V, *et al.* Cellular uptake and efflux of azithromycin, erythromycin, clarithromycin, telithromycin, and cethromycin. *Antimicrob Agents Chemother* 2005; 49: 2372–2377.
- 58. Ianaro A, Ialenti A, Maffia P, *et al.* Anti-inflammatory activity of macrolide antibiotics. *J Pharmacol Exp Ther* 2000; 292: 156–163.
- 59. Tamaoki J. The effects of macrolides on inflammatory cells. Chest 2004; 125: 41S-50S.
- 60. Sunazuka T, Yoshida K, Oohori M, *et al.* Effect of 14-membered macrolide compounds on monocyte to macrophage differentiation. *J Antibiot* 2003; 56: 721–724.
- 61. Murphy BS, Sundareshan V, Cory TJ, *et al.* Azithromycin alters macrophage phenotype. *J Antimicrob Chemother* 2008; 61: 554–560.
- 62. Meyer M, Huaux F, Gavilanes X, *et al.* Azithromycin reduces exaggerated cytokine production by M1 alveolar macrophages in cystic fibrosis. *Am J Respir Cell Mol Biol* 2009; 41: 590–602.
- 63. Asano K, Kamakazu K, Hisamitsu T, *et al.* Modulation of Th2 type cytokine production from human peripheral blood leukocytes by a macrolide antibiotic, roxithromycin, *in vitro. Int Immunopharmacol* 2001; 1: 1913–1921.

- 64. Morikawa K, Zhang J, Nonaka M, *et al.* Modulatory effect of macrolide antibiotics on the Th1- and Th2-type cytokine production. *Int J Antimicrob Agents* 2002; 19: 53–59.
- 65. Park S-J, Lee Y-C, Rhee Y-K, *et al.* The effect of long-term treatment with erythromycin on Th1 and Th2 cytokines in diffuse panbronchiolitis. *Biochem Biophys Res Commun* 2004; 324: 114–117.
- 66. Gorrini M, Lupi A, Viglio S, *et al.* Inhibition of human neutrophil elastase by erythromycin and flurythromycin, two macrolide antibiotics. *Am J Respir Cell Mol Biol* 2001; 25: 492–499.
- 67. Kohri K, Ueki IF, Nadel JA. Neutrophil elastase induces mucin production by ligand-dependent epidermal growth factor receptor activation. *Am J Physiol Lung Cell Mol Physiol* 2002; 283: L531–L540.
- 68. Simpson JL, Powell H, Boyle MJ, *et al.* Clarithromycin targets neutrophilic airway inflammation in refractory asthma. *Am J Respir Crit Care Med* 2008; 177: 148–155.
- 69. Khair OA, Devalia JL, Abdelaziz MM, *et al.* Effect of erythromycin on *Haemophilus influenzae* endotoxin-induced release of IL-6, IL-8 and sICAM-1 by cultured human bronchial epithelial cells. *Eur Respir J* 1995; 8: 1451–1457.
- 70. Tamaoki J, Takeyama K, Tagaya E, *et al.* Effect of clarithromycin on sputum production and its rheological properties in chronic respiratory tract infections. *Antimicrob Agents Chemother* 1995; 39: 1688–1690.
- 71. Shimizu T, Shimizu S, Hattori R, *et al. In vivo* and *in vitro* effects of macrolide antibiotics on mucus secretion in airway epithelial cells. *Am J Respir Crit Care Med* 2003; 168: 581–587.
- 72. Liu J, Zhong X, He Z, *et al.* Effect of low-dose, long-term roxithromycin on airway inflammation and remodeling of stable noncystic fibrosis bronchiectasis. *Mediators Inflamm* 2014; 2014: 708608.
- 73. Kadota J, Mukae H, Ishii H, *et al.* Long-term efficacy and safety of clarithromycin treatment in patients with diffuse panbronchiolitis. *Respir Med* 2003; 97: 844–850.
- 74. Nukarinen E, Tervahartiala T, Valkonen M, *et al.* Targeting matrix metalloproteinases with intravenous doxycycline in severe sepsis a randomised placebo-controlled pilot trial. *Pharmacol Res* 2015; 99: 44–51.
- 75. Wiggins-Dohlvik K, Stagg HW, Han MS, *et al.* Doxycycline attenuates lipopolysaccharide-induced microvascular endothelial cell derangements. *Shock* 2016; 45: 626–633.
- 76. Blau H, Klein K, Shalit I, et al. Moxifloxacin but not ciprofloxacin or azithromycin selectively inhibits IL-8, IL-6, ERK1/2, JNK, and NF-kappaB activation in a cystic fibrosis epithelial cell line. Am J Physiol Lung Cell Mol Physiol 2007; 292: L343–L352.
- 77. Huang H-C, Shieh C-C, Yu W-L, *et al.* Comparing the protective effects of ciprofloxacin, moxifloxacin and levofloxacin in mice with lipopolysaccharide-induced acute lung injuries. *Respirology* 2008; 13: 47–52.
- 78. Vickers IE, Smikle MF. The immunomodulatory effect of antibiotics on the secretion of tumour necrosis factor alpha by peripheral blood mononuclear cells in response to *Stenotrophomonas maltophilia* stimulation. *West Indian Med J* 2006; 55: 138–141.
- 79. Church JA, Fitzgerald F, Walker AS, *et al.* The expanding role of co-trimoxazole in developing countries. *Lancet Infect Dis* 2015; 15: 327–339.
- 80. Poletti V, Casoni G, Chilosi M, et al. Diffuse panbronchiolitis. Eur Respir J 2006; 28: 862-871.
- 81. Kudoh S, Keicho N. Diffuse panbronchiolitis. Clin Chest Med 2012; 33: 297-305.
- 82. Ichikawa Y, Koga H, Tanaka M, *et al.* Neutrophilia in bronchoalveolar lavage fluid of diffuse panbronchiolitis. *Chest* 1990; 98: 917–923.
- 83. Sakito O, Kadota J, Kohno S, *et al.* Interleukin 1 beta, tumor necrosis factor alpha, and interleukin 8 in bronchoalveolar lavage fluid of patients with diffuse panbronchiolitis: a potential mechanism of macrolide therapy. *Respiration* 1996; 63: 42–48.
- 84. Kudoh S, Uetake T, Hagiwara K, *et al.* [Clinical effects of low-dose long-term erythromycin chemotherapy on diffuse panbronchiolitis.] *Nihon Kyobu Shikkan Gakkai Zasshi* 1987; 25: 632–642.
- 85. Fujii T, Kadota J, Kawakami K, *et al.* Long term effect of erythromycin therapy in patients with chronic *Pseudomonas aeruginosa* infection. *Thorax* 1995; 50: 1246–1252.
- 86. Hiratsuka T, Mukae H, Iiboshi H, *et al.* Increased concentrations of human beta-defensins in plasma and bronchoalveolar lavage fluid of patients with diffuse panbronchiolitis. *Thorax* 2003; 58: 425–430.
- 87. Nagai H, Shishido H, Yoneda R, *et al.* Long-term low-dose administration of erythromycin to patients with diffuse panbronchiolitis. *Respiration* 1991; 58: 145–149.
- 88. Koyama H, Geddes DM. Erythromycin and diffuse panbronchiolitis. Thorax 1997; 52: 915–918.
- 89. Kadota J-I, Mukae H, Tomono K, *et al.* Efficacy of long-term macrolide antibiotic therapy in patients with diffuse panbronchiolitis: comparison between HLA-B54-positive and -negative cases. *Int J Antimicrob Agents* 2004; 24: 550–554.
- 90. Li H, Zhou Y, Fan F, *et al.* Effect of azithromycin on patients with diffuse panbronchiolitis: retrospective study of 51 cases. *Intern Med* 2011; 50: 1663–1669.
- 91. Hui D, Yan F, Chen R-H. The effects of azithromycin on patients with diffuse panbronchiolitis: a retrospective study of 29 cases. *J Thorac Dis* 2013; 5: 613–617.
- 92. Akira M, Higashihara T, Sakatani M, *et al.* Diffuse panbronchiolitis: follow-up CT examination. *Radiology* 1993; 189: 559–562.

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- 93. Lin X, Lu J, Yang M, et al. Macrolides for diffuse panbronchiolitis. Cochrane Database Syst Rev 2015; 1: CD007716.
- 94. Elborn JS. Cystic fibrosis. Lancet 2016; 388: 2519-2531.
- 95. Dodge JA, Lewis PA, Stanton M, *et al.* Cystic fibrosis mortality and survival in the UK: 1947–2003. *Eur Respir J* 2007; 29: 522–526.
- 96. de Boer K, Vandemheen KL, Tullis E, *et al.* Exacerbation frequency and clinical outcomes in adult patients with cystic fibrosis. *Thorax* 2011; 66: 680–685.
- 97. Stephenson AL, Berthiaume Y, Singer LG, et al. A contemporary survival analysis of individuals with cystic fibrosis: a cohort study. Eur Respir J 2015; 45: 670–679.
- Saiman L, Anstead M, Mayer-Hamblett N, et al. Effect of azithromycin on pulmonary function in patients with cystic fibrosis uninfected with Pseudomonas aeruginosa: a randomized controlled trial. JAMA 2010; 303: 1707–1715.
- 99. Southern KW, Barker PM, Solis-Moya A, et al. Macrolide antibiotics for cystic fibrosis. Cochrane Database Syst Rev 2012; 11: CD002203.
- Willekens J, Eyns H, Malfroot A. How long should we maintain long-term azithromycin treatment in cystic fibrosis patients? *Pediatr Pulmonol* 2015; 50: 103–104.
- 101. Samson C, Tamalet A, Thien HV, et al. Long-term effects of azithromycin in patients with cystic fibrosis. Respir Med 2016; 117: 1–6.
- Haworth CS, Johnson C, Aliberti S. Management of bronchiectasis in Europe: data from the European bronchiectasis registry (EMBARC). Eur Respir J 2016; 48: OA273.
- Currie DC, Garbett ND, Chan KL, et al. Double-blind randomized study of prolonged higher-dose oral amoxycillin in purulent bronchiectasis. Q J Med 1990; 76: 799–816.
- 104. Wilson R, Wells AU. Azithromycin in bronchiectasis: when should it be used? Lancet 2012; 380: 627-629.
- 105. Elborn JS, Tunney MM. Macrolides and bronchiectasis: clinical benefit with a resistance price. JAMA 2013; 309: 1295–1296.
- 106. Decramer M, Janssens W, Miravitlles M. Chronic obstructive pulmonary disease. Lancet 2012; 379: 1341–1351.
- Suissa S, Dell'Aniello S, Ernst P. Long-term natural history of chronic obstructive pulmonary disease: severe exacerbations and mortality. *Thorax* 2012; 67: 957–963.
- Wang Z, Bafadhel M, Haldar K, et al. Lung microbiome dynamics in COPD exacerbations. Eur Respir J 2016; 47: 1082–1092.
- Wedzicha JA, Seemungal TAR. COPD exacerbations: defining their cause and prevention. Lancet 2007; 370: 786–796.
- 110. Gomez C, Chanez P. The lung microbiome: the perfect culprit for COPD exacerbations? *Eur Respir J* 2016; 47: 1034–1036.
- 111. James GDR, Petersen I, Nazareth I, *et al.* Use of long-term antibiotic treatment in COPD patients in the UK: a retrospective cohort study. *Prim Care Respir J* 2013; 22: 271–277.
- 112. May JR, Oswald NC. Long-term chemotherapy in chronic bronchitis. Lancet 1956; 271: 814-818.
- 113. Prins HJ, Daniels JMA, Lindeman JH, *et al.* Effects of doxycycline on local and systemic inflammation in stable COPD patients, a randomized clinical trial. *Respir Med* 2016; 110: 46–52.
- 114. Jordan GW, Krajden SF, Hoeprich PD, et al. Trimethoprim-sulfamethoxazole in chronic bronchitis. Can Med Assoc J 1975; 112: 91–95.
- 115. Sethi S, Jones PW, Theron MS, et al. Pulsed moxifloxacin for the prevention of exacerbations of chronic obstructive pulmonary disease: a randomized controlled trial. Respir Res 2010; 11: 10.
- 116. Martínez-García MÁ, la Rosa Carrillo de D, Soler-Cataluña JJ, *et al.* Prognostic value of bronchiectasis in patients with moderate-to-severe chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2013; 187: 823–831.
- 117. Vermeersch K, Gabrovska M, Deslypere G, *et al.* The Belgian trial with azithromycin for acute COPD exacerbations requiring hospitalization: an investigator-initiated study protocol for a multicenter, randomized, double-blind, placebo-controlled trial. *Int J Chron Obstruct Pulmon Dis* 2016; 11: 687–696.
- 118. Wenzel RP, Fowler AA, Edmond MB. Antibiotic prevention of acute exacerbations of COPD. *N Engl J Med* 2012; 367: 340–347.
- 119. Han MK, Tayob N, Murray S, et al. Predictors of chronic obstructive pulmonary disease exacerbation reduction in response to daily azithromycin therapy. Am J Respir Crit Care Med 2014; 189: 1503–1508.
- 120. Wenzel SE. Asthma phenotypes: the evolution from clinical to molecular approaches. Nat Med 2012; 18: 716–725.
- 121. Black PN, Blasi F, Jenkins CR, et al. Trial of roxithromycin in subjects with asthma and serological evidence of infection with *Chlamydia pneumoniae*. Am J Respir Crit Care Med 2001; 164: 536–541.
- 122. Kraft M, Cassell GH, Pak J, et al. Mycoplasma pneumoniae and Chlamydia pneumoniae in asthma: effect of clarithromycin. Chest 2002; 121: 1782–1788.
- 123. Cunningham AF, Johnston SL, Julious SA, et al. Chronic Chlamydia pneumoniae infection and asthma exacerbations in children. Eur Respir J 1998; 11: 345–349.
- 124. Wark PAB, Johnston SL, Simpson JL, et al. Chlamydia pneumoniae immunoglobulin A reactivation and airway inflammation in acute asthma. Eur Respir J 2002; 20: 834–840.

- 125. Johnston SL, Martin RJ. Chlamydophila pneumoniae and Mycoplasma pneumoniae: a role in asthma pathogenesis? Am J Respir Crit Care Med 2005; 172: 1078–1089.
- 126. Essilfie A-T, Horvat JC, Kim RY, *et al.* Macrolide therapy suppresses key features of experimental steroid-sensitive and steroid-insensitive asthma. *Thorax* 2015; 70: 458–467.
- 127. Brusselle GG. Are the antimicrobial properties of macrolides required for their therapeutic efficacy in chronic neutrophilic airway diseases? *Thorax* 2015; 70: 401–403.
- 128. Shoji T, Yoshida S, Sakamoto H, et al. Anti-inflammatory effect of roxithromycin in patients with aspirin-intolerant asthma. Clin Exp Allergy 1999; 29: 950–956.
- 129. Richeldi L, Ferrara G, Fabbri LM, et al. Macrolides for chronic asthma. Cochrane Database Syst Rev 2005; 3: CD002997.
- 130. Sutherland ER, King TS, Icitovic N, *et al.* A trial of clarithromycin for the treatment of suboptimally controlled asthma. *J Allergy Clin Immunol* 2010; 126: 747–753.
- 131. Coeman M, van Durme Y, Bauters F, *et al.* Neomacrolides in the treatment of patients with severe asthma and/or bronchiectasis: a retrospective observational study. *Ther Adv Respir Dis* 2011; 5: 377–386.
- 132. Reiter J, Demirel N, Mendy A, et al. Macrolides for the long-term management of asthma a meta-analysis of randomized clinical trials. Allergy 2013; 68: 1040–1049.
- 133. Johnston SL, Blasi F, Black PN, *et al.* The effect of telithromycin in acute exacerbations of asthma. *N Engl J Med* 2006; 354: 1589–1600.
- 134. Johnston SL, Szigeti M, Cross M, *et al.* Azithromycin for acute exacerbations of asthma: the AZALEA randomized clinical trial. *JAMA Intern Med* 2016; 176: 1630–1637.
- Brusselle GG, Van Braeckel E. AZALEA trial highlights antibiotic overuse in acute asthma attacks. JAMA Intern Med 2016; 176: 1637–1638.
- 136. Kew KM, Undela K, Kotortsi I, et al. Macrolides for chronic asthma. Cochrane Database Syst Rev 2015; 9: CD002997.
- 137. Vos R, Vanaudenaerde BM, Verleden SE, *et al.* Anti-inflammatory and immunomodulatory properties of azithromycin involved in treatment and prevention of chronic lung allograft rejection. *Transplantation* 2012; 94: 101–109.
- 138. Verleden GM, Vos R, Vanaudenaerde B, et al. Current views on chronic rejection after lung transplantation. *Transpl Int* 2015; 28: 1131–1139.
- 139. Meyer KC, Raghu G, Verleden GM, et al. An international ISHLT/ATS/ERS clinical practice guideline: diagnosis and management of bronchiolitis obliterans syndrome. Eur Respir J 2014; 44: 1479–1503.
- Verleden SE, Sacreas A, Vos R, et al. Advances in understanding bronchiolitis obliterans after lung transplantation. Chest 2016; 150: 219–225.
- 141. Verleden GM, Vanaudenaerde BM, Dupont LJ, et al. Azithromycin reduces airway neutrophilia and interleukin-8 in patients with bronchiolitis obliterans syndrome. Am J Respir Crit Care Med 2006; 174: 566–570.
- 142. Gottlieb J, Szangolies J, Koehnlein T, *et al.* Long-term azithromycin for bronchiolitis obliterans syndrome after lung transplantation. *Transplantation* 2008; 85: 36–41.
- 143. Corris PA, Ryan VA, Small T, *et al.* A randomised controlled trial of azithromycin therapy in bronchiolitis obliterans syndrome (BOS) post lung transplantation. *Thorax* 2015; 70: 442–450.
- 144. Vos R, Vanaudenaerde BM, Verleden SE, *et al.* A randomised controlled trial of azithromycin to prevent chronic rejection after lung transplantation. *Eur Respir J* 2011; 37: 164–172.
- 145. Ruttens D, Verleden SE, Vandermeulen E, *et al.* Prophylactic azithromycin therapy after lung transplantation: *post hoc* analysis of a randomized controlled trial. *Am J Transplant* 2016; 16: 254–261.
- Lam DCL, Lam B, Wong MKY, et al. Effects of azithromycin in bronchiolitis obliterans syndrome after hematopoietic SCT – a randomized double-blinded placebo-controlled study. Bone Marrow Transplant 2011; 46: 1551–1556.
- 147. Lemonnier F, Rivaud E, Neveu H, et al. Azithromycin in bronchiolitis obliterans syndrome after hematopoietic SCT. Bone Marrow Transplant 2012; 47: 1374–1374.
- 148. Yadav H, Peters SG, Keogh KA, *et al.* Azithromycin for the treatment of obliterative bronchiolitis after hematopoietic stem cell transplantation: a systematic review and meta-analysis. *Biol Blood Marrow Transplant* 2016; 22: 2264–2269.
- 149. Cordier JF. Organising pneumonia. Thorax 2000; 55: 318-328.
- 150. Ichikawa Y, Ninomiya H, Katsuki M, et al. Low-dose/long-term erythromycin for treatment of bronchiolitis obliterans organizing pneumonia (BOOP). Kurume Med J 1993; 40: 65–67.
- 151. Hotta M. Neutrophil chemotactic activity in cryptogenic organizing pneumonia and the response to erythromycin. *Kurume Med J* 1996; 43: 207–217.
- 152. Stover DE, Mangino D. Macrolides: a treatment alternative for bronchiolitis obliterans organizing pneumonia? *Chest* 2005; 128: 3611–3617.
- 153. Pathak V, Kuhn JM, Durham C, et al. Macrolide use leads to clinical and radiological improvement in patients with cryptogenic organizing pneumonia. Ann Am Thorac Soc 2014; 11: 87–91.

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- 154. Ding Q-L, Lv D, Wang B-J, *et al.* Macrolide therapy in cryptogenic organizing pneumonia: a case report and literature review. *Exp Ther Med* 2015; 9: 829–834.
- Ray WA, Murray KT, Meredith S, et al. Oral erythromycin and the risk of sudden death from cardiac causes. N Engl J Med 2004; 351: 1089–1096.
- 156. Broad J, Sanger GJ. The antibiotic azithromycin is a motilin receptor agonist in human stomach: comparison with erythromycin. *Br J Pharmacol* 2013; 168: 1859–1867.
- 157. Altenburg J, de Graaff CS, van der Werf TS, *et al.* Immunomodulatory effects of macrolide antibiotics part 2: advantages and disadvantages of long-term, low-dose macrolide therapy. *Respiration* 2011; 81: 75–87.
- 158. Periti P, Mazzei T, Mini E, et al. Adverse effects of macrolide antibacterials. Drug Saf 1993; 9: 346–364.
- 159. Principi N, Esposito S. Comparative tolerability of erythromycin and newer macrolide antibacterials in paediatric patients. *Drug Saf* 1999; 20: 25–41.
- 160. Rubinstein E. Comparative safety of the different macrolides. Int J Antimicrob Agents 2001; 18: Suppl. 1, S71–S76.
- 161. Mosholder AD, Mathew J, Alexander JJ, et al. Cardiovascular risks with azithromycin and other antibacterial drugs. N Engl J Med 2013; 368: 1665–1668.
- 162. Mortensen EM, Halm EA, Pugh MJ, et al. Association of azithromycin with mortality and cardiovascular events among older patients hospitalized with pneumonia. JAMA 2014; 311: 2199–2208.
- 163. Hancox JC, Hasnain M, Vieweg WVR, et al. Azithromycin, cardiovascular risks, QTc interval prolongation, torsade de pointes, and regulatory issues: a narrative review based on the study of case reports. Ther Adv Infect Dis 2013; 1: 155–165.
- Ray WA, Murray KT, Hall K, et al. Azithromycin and the risk of cardiovascular death. N Engl J Med 2012; 366: 1881–1890.
- 165. Svanström H, Pasternak B, Hviid A. Use of azithromycin and death from cardiovascular causes. N Engl J Med 2013; 368: 1704–1712.
- 166. Swanson DJ, Sung RJ, Fine MJ, et al. Erythromycin ototoxicity: prospective assessment with serum concentrations and audiograms in a study of patients with pneumonia. Am J Med 1992; 92: 61–68.
- 167. Wallace MR, Miller LK, Nguyen MT, et al. Ototoxicity with azithromycin. Lancet 1994; 343: 241.
- 168. Brown BA, Griffith DE, Girard W, et al. Relationship of adverse events to serum drug levels in patients receiving high-dose azithromycin for mycobacterial lung disease. Clin Infect Dis 1997; 24: 958–964.
- Hill AT. Macrolides for clinically significant bronchiectasis in adults: who should receive this treatment? Chest 2016; 150: 1187–1193.
- 170. Bergman M, Huikko S, Huovinen P, *et al.* Macrolide and azithromycin use are linked to increased macrolide resistance in *Streptococcus pneumoniae. Antimicrob Agents Chemother* 2006; 50: 3646–3650.
- 171. Karlowsky JA, Lagacé-Wiens PRS, Low DE, *et al.* Annual macrolide prescription rates and the emergence of macrolide resistance among *Streptococcus pneumoniae* in Canada from 1995 to 2005. *Int J Antimicrob Agents* 2009; 34: 375–379.
- 172. Goossens H. Antibiotic consumption and link to resistance. Clin Microbiol Infect 2009; 15: Suppl. 3, 12-15.
- 173. Coles CL, Mabula K, Seidman JC, et al. Mass distribution of azithromycin for trachoma control is associated with increased risk of azithromycin-resistant *Streptococcus pneumoniae* carriage in young children 6 months after treatment. *Clin Infect Dis* 2013; 56: 1519–1526.
- 174. Serisier DJ. Risks of population antimicrobial resistance associated with chronic macrolide use for inflammatory airway diseases. *Lancet Respir Med* 2013; 1: 262–274.
- 175. Hansen CR, Pressler T, Høiby N, *et al.* Long-term, low-dose azithromycin treatment reduces the incidence but increases macrolide resistance in *Staphylococcus aureus* in Danish CF patients. *J Cyst Fibros* 2009; 8: 58–62.
- Phaff SJ, Tiddens HAWM, Verbrugh HA, et al. Macrolide resistance of Staphylococcus aureus and Haemophilus species associated with long-term azithromycin use in cystic fibrosis. J Antimicrob Chemother 2006; 57: 741–746.
- 177. Tramper-Stranders GA, Wolfs TFW, Fleer A, *et al.* Maintenance azithromycin treatment in pediatric patients with cystic fibrosis: long-term outcomes related to macrolide resistance and pulmonary function. *Pediatr Infect Dis J* 2007; 26: 8–12.
- Hansen SG, Vieville C, Whizin N, et al. Effector memory T cell responses are associated with protection of rhesus monkeys from mucosal simian immunodeficiency virus challenge. Nat Med 2009; 15: 293–299.
- 179. Fan L-C, Lu H-W, Wei P, et al. Effects of long-term use of macrolides in patients with non-cystic fibrosis bronchiectasis: a meta-analysis of randomized controlled trials. BMC Infect Dis 2015; 15: 160.
- 180. Banerjee D, Khair OA, Honeybourne D. The effect of oral clarithromycin on health status and sputum bacteriology in stable COPD. *Respir Med* 2005; 99: 208–215.
- Moreno S, García-Leoni ME, Cercenado E, et al. Infections caused by erythromycin-resistant Streptococcus pneumoniae: incidence, risk factors, and response to therapy in a prospective study. Clin Infect Dis 1995; 20: 1195–1200.
- Aspa J, Rajas O, Rodríguez de Castro F, et al. Drug-resistant pneumococcal pneumonia: clinical relevance and related factors. Clin Infect Dis 2004; 38: 787–798.
- 183. Lujan M, Gallego M, Fontanals D, *et al.* Prospective observational study of bacteremic pneumococcal pneumonia: effect of discordant therapy on mortality. *Crit Care Med* 2004; 32: 625–631.

- 184. Song J-H, Jung S-I, Ki HK, et al. Clinical outcomes of pneumococcal pneumonia caused by antibiotic-resistant strains in Asian countries: a study by the Asian Network for Surveillance of Resistant Pathogens. Clin Infect Dis 2004; 38: 1570–1578.
- 185. Cilloniz C, Albert RK, Liapikou A, et al. The effect of macrolide resistance on the presentation and outcome of patients hospitalized for Streptococcus pneumoniae pneumonia. Am J Respir Crit Care Med 2015; 191: 1265–1272.
- Griffith DE, Brown-Elliott BA, Langsjoen B, et al. Clinical and molecular analysis of macrolide resistance in Mycobacterium avium complex lung disease. Am J Respir Crit Care Med 2006; 174: 928–934.
- 187. Renna M, Schaffner C, Brown K, *et al.* Azithromycin blocks autophagy and may predispose cystic fibrosis patients to mycobacterial infection. *J Clin Invest* 2011; 121: 3554–3563.
- 188. Coolen N, Morand P, Martin C, et al. Reduced risk of nontuberculous mycobacteria in cystic fibrosis adults receiving long-term azithromycin. J Cyst Fibros 2015; 14: 594–599.
- 189. Floto RA, Olivier KN, Saiman L, et al. US Cystic Fibrosis Foundation and European Cystic Fibrosis Society consensus recommendations for the management of non-tuberculous mycobacteria in individuals with cystic fibrosis: executive summary. *Thorax* 2016; 71: 88–90.

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