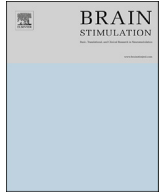




Contents lists available at ScienceDirect

Brain Stimulation

journal homepage: <http://www.journals.elsevier.com/brain-stimulation>

tDCS changes in motor excitability are specific to orientation of current flow

Vishal Rawji ^{a,*}, Matteo Ciocca ^a, André Zacharia ^{a,c}, David Soares ^a, Dennis Truong ^b, Marom Bikson ^b, John Rothwell ^a, Sven Bestmann ^a

^a Sobell Department of Motor Neuroscience and Movement Disorders, UCL Institute of Neurology, University College London, London, UK

^b Department of Biomedical Engineering, The City College of the City University of New York, New York City, USA

^c Department of Neurology, University Hospitals of Geneva, Switzerland

ARTICLE INFO

Article history:

Received 15 June 2017

Received in revised form

30 September 2017

Accepted 2 November 2017

Available online xxx

Keywords:

Transcranial magnetic stimulation

Transcranial direct current stimulation

Primary motor cortex

ABSTRACT

Background: Measurements and models of current flow in the brain during transcranial Direct Current Stimulation (tDCS) indicate stimulation of regions in-between electrodes. Moreover, the folded cortex results in local fluctuations in current flow intensity and direction, and animal studies suggest current flow direction relative to cortical columns determines response to tDCS.

Methods: Here we test this idea by using Transcranial Magnetic Stimulation Motor Evoked Potentials (TMS-MEP) to measure changes in corticospinal excitability following tDCS applied with electrodes aligned orthogonal (across) or parallel to M1 in the central sulcus.

Results: Current flow models predicted that the orthogonal electrode montage produces consistently oriented current across the hand region of M1 that flows along cortical columns, while the parallel electrode montage produces non-uniform current directions across the M1 cortical surface. We find that orthogonal, but not parallel, orientated tDCS modulates TMS-MEPs. We also show modulation is sensitive to the orientation of the TMS coil (PA or AP), which is thought to select different afferent pathways to M1.

Conclusions: Our results are consistent with tDCS producing directionally specific neuromodulation in brain regions in-between electrodes, but shows nuanced changes in excitability that are presumably current direction relative to column and axon pathway specific. We suggest that the direction of current flow through cortical target regions should be considered for targeting and dose-control of tDCS.

Crown Copyright © 2017 Published by Elsevier Inc. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

Funding

This research was funded by grants from the Medical Research Council, Department of Defense Air Force Office of Scientific Research (#FA9550-13-1-0073) and National Institutes of Health (#5R01MH092926 and #1R01NS101362)

Introduction

To date, the majority of studies in humans using transcranial direct current stimulation (tDCS) to modulate cortical function

Abbreviations: PA, postero-anterior; AP, antero-posterior; ML, medio-lateral; tDCS, transcranial direct current stimulation; MEP, motor evoked potential; M1, primary motor cortex; TMS, transcranial magnetic stimulation; AP-TMS-MEPs, motor evoked potentials elicited with anterior-posterior directed TMS; PA-TMS-MEPs, motor evoked potentials elicited with posterior-anterior directed TMS.

* Corresponding author. UCL Institute of Neurology, London, WC1N 3BG, UK.

E-mail address: vishal.rawji.11@ucl.ac.uk (V. Rawji).

employ a bipolar electrode montage: one electrode is usually placed over the target site and the other at a distance. So, for the hand area of motor cortex (M1), a large anode is conventionally centred over the anatomical location of the “hand knob” of the precentral gyrus, with a cathode over the contralateral orbit [1]. This montage, based on canonical studies by Nitsche, Paulus and colleagues on how the position of large electrodes influences population-averaged modulation of TMS-MEPs [2–4], is now widely applied for targeting diverse cortical target regions [5,6] though rarely with consideration for nuanced dose response [7–11]. Intra-cranial recordings [12] and clinical imaging [13,14], supported by current flow models [15,16] show bipolar electrode montages produce current flow in brain regions between electrodes. Though putative brain targets between electrodes have been considered [17–19], previous tDCS studies have not systematically isolated the consequences of inter-electrode current flow.

The “inter-electrode” considerations provoke a second question. Animal studies in lissencephalic animals indicate polarity specific

<https://doi.org/10.1016/j.brs.2017.11.001>

1935-861X/Crown Copyright © 2017 Published by Elsevier Inc. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

(anodal/cathodal) excitability changes for current directed normal to the cortical surface [2], which corresponds to current flow directed along the primary dendritic axis of cortical pyramidal neurons [20,21]. In the human gyrencephalic cortex, such controlled stimulation cannot easily be achieved and the directions of current flow underneath an electrode are complex [22,23]. The position of primary motor cortex in the anterior wall of the central sulcus suggests that electrode montages that direct current flow perpendicular through this gyral wall (and thus predominantly along the primary dendritic axis of cortical pyramidal neurons) may optimally modulate corticospinal excitability (CSE). The second question we address here is therefore whether there are differences in the effect of tDCS on CSE when current is oriented perpendicularly across, compared with parallel to, the cortical surface at the level of the M1 hand area. To this end, we positioned tDCS electrodes 7 cm anterior and posterior to the hand area of M1 to direct current flow across the central sulcus (Fig. 1). This means that depending on the position of the anode and cathode, current will flow through M1 in anterior-posterior (AP-tDCS) or posterior-anterior (PA-tDCS) direction, respectively. In a second condition, we positioned electrodes 7 cm medial and lateral to the M1 hand area to direct current flow in parallel along the cortical surface of central sulcus (Fig. 1). We refer to this as medio-lateral tDCS (ML-tDCS). Motor-evoked potentials (MEPs) elicited with TMS (TMS-MEPs) were used to access CSE changes after stimulation with these two orthogonal tDCS orientations.

We also addressed a third question. The effects of TMS on motor cortex are well-known to be directional [24,25]. TMS with a monophasic pulse that induces an electric current flowing from approximately posterior to anterior across the central sulcus (perpendicular to the line of the individual's central sulcus at that point) evokes MEPs (PA-TMS-MEPs) that have a shorter latency and lower threshold than stimulation with an anterior-posterior induced current (AP-TMS-MEPs). It is thought that this is because the two directions of stimulation activate different sets of presynaptic inputs to corticospinal neurones [20]. Indeed, brain slice studies of tDCS established modulation varies across afferent axonal pathways or varied orientation [23,26]. We therefore hypothesised that any effects of tDCS across M1 might also be directionally selective, and that they would interact in different ways with the direction of TMS used for eliciting MEPs. Specifically, using tDCS to direct current perpendicular to the M1 hand area, we expected that stimulation with a posterior cathode and anterior cathode (PA-tDCS) would influence MEPs evoked by PA and AP TMS in a different way to tDCS applied with an anterior anode and posterior cathode (AP-tDCS).

Methods

Participants

22 healthy volunteers (17 male, 21 right handed) aged 21–44 (mean age 28.95, SD 6.14) participated in this experiment. The study was approved by the UCL Ethics Committee and none had contraindications to TMS or tDCS as assessed by a TMS/tDCS screening questionnaire.

Current flow modelling

Finite Element Method (FEM) models of tDCS were generated to predict electric field (E-field) orientation along the motor cortex. High resolution T1 and T2 weighted MRI scans (GRE sequence, TR = 1900 ms, TE = 2.2 ms and SPACE sequence, TR = 3200 ms, TE = 402 ms respectively for subject S0; MPRAGE, TR = 6.92 ms, TE = 3.2 ms for subject S4) were previously collected and

segmented using a combination of automated and manual segmentation techniques [27]. The two individualized subjects selected had contrasting size, gender, hand knob shape, and induced electric field sensitivity [27]. Automated segmentation algorithms derived from Unified Segmentation in SPM8 [28,29] were combined with updated tissue probability maps and morphological filters (smoothing, dilation, erosion) specifically developed for current flow modelling [30]. Additional image masks (fat, electrodes, gels) and regions of interest (M1) were segmented using manual and semi-manual tools (Simpleware, Synopsys) to remove aliasing artifacts, incorporate gyri-precise detail, and position stimulation electrodes along or across the hand knob of the motor cortex. Adaptive tetrahedral meshes were generated using a voxel-based algorithm (Simpleware, Synopsys) with multiple domains corresponding to different material conductivities verified by intra-cranial recording (in S/m: Scalp 0.465, fat 0.025, skull 0.01, CSF 1.65, Grey matter 0.276, White matter 0.126, Air 1e-15, electrode 5.99e7, gel 1.4) [31–33]. Meshes were imported into a FEM solver (COMSOL Multiphysics) where the Laplace equation for electrostatics ($\nabla \cdot (\sigma \nabla V) = 0$) was solved as the field equation given a normal current density boundary condition on the anode equivalent to 1 mA, ground boundary condition on the cathode, and insulation on all other external boundaries. Results were scaled linearly to match experimental conditions when necessary. E-field orientation was visualized with surface arrows seeded evenly along M1. Arrow colors corresponded to E-field normal ($n \cdot E$) to the cortical surface. By convention, positive normal E-field represented inward “anodal” E-field (red) while negative normal E-field represented outward “cathodal” E-field (blue). Streamlines representing current flow through M1 were generated by seeding 100 points randomly along the surface of M1 and the gel-skin contact. Current density was then traced throughout the model. Line thickness was a logarithmic function of current density magnitude and colored to Voltage (red, anode; blue, cathode).

EMG recordings

Throughout the experiment, subjects were seated comfortably in a non-reclining chair, with their right hand rested on a cushion. Electromyographic (EMG) activity was recorded from the right first dorsal interosseous (FDI) muscle using Ag/AgCl cup electrodes arranged in a belly-tendon montage. The raw signals were amplified and a bandpass filter was also applied. (20 Hz to 2 kHz (Digitimer, Welwyn Garden City, UK)) Signals were digitised at 5 kHz (CED Power 1401; Cambridge Electronic Design, Cambridge, United Kingdom) and data were stored on a computer for offline analysis (Signal Version 5.10, Cambridge Electronic Design, UK was used).

Transcranial magnetic stimulation

Single pulse, monophasic TMS was employed using a Magstim 200² stimulator (The Magstim Co. Ltd) connected via a figure-of-eight coil with an internal wing diameter of 7 cm. The hotspot was identified as the area on the scalp where the largest and most stable MEPs could be obtained for the right FDI muscle, using a given suprathreshold intensity. The coil was held approximately perpendicular to the presumed central sulcus and held tangentially to the skull with the coil handle pointing backwards for postero-anterior (PA) stimulation and handle pointing forwards for antero-posterior (AP) stimulation. A coloured pencil was used to draw the boundaries around the coil so that it could be accurately positioned to the hotspot for further recordings, for PA and AP coil orientations. Hence we found the optimal orientation and location pertaining to the FDI muscle in each individual. TMS was delivered at a rate of 0.2 Hz during data collection.

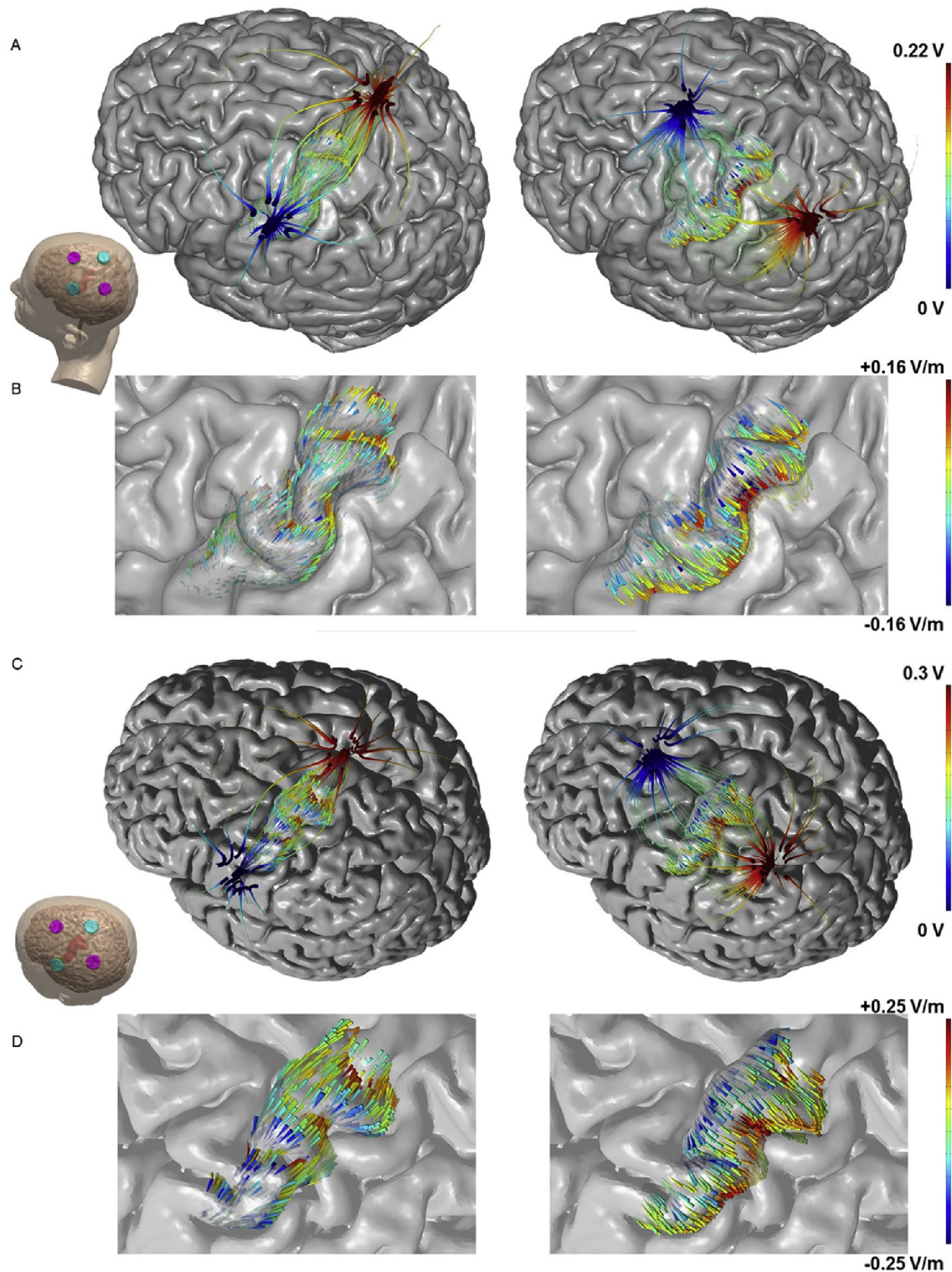


Fig. 1. Comparison of electrical field modelling for montages directing current across and along the cortical surface.

Electric field orientation on the cortex as applied by electrodes along (A) or across (B) the motor strip. Note: The streamlines and arrows have two separate colorscales. Starting outside the motor-strip, streamlines trace the direction of current density from high to low voltage (red to blue), anode to cathode. The streamlines confirm that current flows down the voltage gradient established by the electrodes and shaped by the head anatomy. On the motor strip, arrows illustrate the direction of electric field passing through the motor cortex surface. Arrow colour represents normal electric field where red is inward and blue is outward. Inward field corresponds to expected pyramidal soma depolarization and outward corresponds to expected pyramidal soma hyper-polarization (though we note that the polarization of axons and terminals must also be considered). It is important to not confuse the red/blue of voltage with the red/blue of electric field/polarization since the two are not simply related; none-the-less, this representation allows correlation of macro-scale current flow patterns set by electrode montage with gyri-scale current flow pattern which determine cellular polarization. These modelling results show how current can be directed to flow more uniformly through M1. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

Transcranial direct current stimulation

Transcranial direct current stimulation (tDCS; Starstim, Barcelona; 1 mA) was applied via 3.14 cm² Ag/AgCl gelled electrodes yielding an average electrode current density of 0.318 mA/cm². The stimulation was applied for a total of 10 min, ramped up and down for 5 s at the beginning and end of the stimulation. Participants asked to stay awake and at rest during the stimulation. Sham stimulation involved ramping up then down both at the start and end of the 10 min period, with zero stimulation for the remaining time.

TDCS electrodes were positioned 7 cm anterior and posterior to the TMS hotspot along the orientation of the TMS coil; two further electrodes were placed 7 cm medial and lateral to the hotspot perpendicular to the coil orientation (i.e. along the length of the central sulcus). We note that this convention is with regards to the approximate orientation of the central sulcus, which is generally oriented at about 45° with respect to the midline. For simplicity, we will assume that this corresponds to the anterior-posterior and medio-lateral orientation of the brain.

Stimulation was set up remotely on a computer and delivered via a Bluetooth receiver connected to the electrodes. We refer to stimulation with a posterior anode and anterior cathode as PA-tDCS, to indicate the direction of the electric field; AP-tDCS refers to stimulation with an anterior anode and posterior cathode. We refer to stimulation with a medial anode and lateral cathode (directing current flow approximately in parallel to the central sulcus) as ML-tDCS (see Fig. 2). Subjects were blinded to the type of stimulation.

Experimental parameters

Resting motor threshold (RMT) was defined as the lowest TMS stimulus intensity to evoke a response of 50 μ V in 5 out of 10 trials in the relaxed FDI using the optimal PA orientation [34].

Twenty MEPs were collected before and after tDCS, with post tDCS MEPs collected every 10 min from T0-T40. The TMS stimulus intensity was set at the intensity required to evoke a response of 1 mV peak-to-peak amplitude (SI1mV), and this intensity was kept constant throughout the entire experiment. The mean amplitude of these MEPs was calculated for each time point for each subject.

Experiment 1

In experiment 1, we investigated whether corticospinal excitability could be modulated with an electrode montage for which the region of interest (M1 hand region) was positioned between our stimulating electrodes. This was accomplished using two different stimulating montages, PA and ML tDCS (as described above), to investigate whether direction of current flow across M1 could differentially modulate responses. To this end, fifteen people participated in a crossover study, which consisted of three randomised sessions (ML-tDCS, PA-tDCS and sham), each separated by at least five days. MEPs were assessed before and after tDCS using PA TMS.

Experiment 2

Following the interesting results from experiment 1, we next asked whether the observed effect of PA-tDCS on corticospinal excitability might be explained by specific modulation of posterior-to-anterior or anterior-to-posterior inputs into M1, as probed by exploiting the known directional sensitivity of TMS over M1. Fourteen people participated in this experiment, eight of whom participated in experiment 1. PA-tDCS was applied and the effects on PA-TMS-MEPs and AP-TMS-MEPs were assessed. For each TMS stimulus direction at each time point, 20 MEPs were acquired.

Experiment 3

To complete the permutations, we finally assessed the effect of AP-tDCS on both PA and AP TMS pulses in the same group as in experiment 2.

Data analyses

The amplitude of each single MEP at each time point was measured and averaged in each individual. Data from each individual was then averaged into a grand mean and entered into a two-way repeated measures analyses of variance (rmANOVA) with main factors "STIMULATION" (in experiment 1: ML-tDCS, PA-tDCS and Sham) or "COIL DIRECTION" (PA and AP TMS coil orientations in experiments 2 and 3) and "TIME" (Baseline, T0, T10, T20, T30 and T40 for all experiments). Absolute MEP values were used in each statistical test. In cases where there was a significant "STIMULATION" x "TIME" OR "COIL DIRECTION x TIME" interaction, analysis showed no overall effect of TIME from T0-T40 (i.e. post-tDCS). Therefore we calculated the mean post-tDCS effect and expressed this as a fraction of the baseline for post hoc testing. To examine test-retest reliability between individuals having repeated PA-tDCS sessions, we used a (2,k) intraclass correlation coefficient.

Results

Modelling current flow

The electrode montages we used have not been explored in detail previously. Adapting models with parameters previously validated by intra-cranial recording [12], we calculated the expected electric field distribution in the central area of the cerebral cortex using the electrodes and stimulation sites in the present study. Fig. 1 shows the predictions for ML-tDCS (left) and PA-tDCS (right). As reported by others [16,30,35,36], the modelling shows that with bipolar montages substantial current flow (field intensities) is produced between the two electrodes. There is a notable and clear difference between the two electrode montages: whereas ML-tDCS does not produce any uniformly directed electrical fields through the main surface of motor cortex located in the anterior bank of the central sulcus, PA-tDCS leads to relatively uniform inward and outward electrical fields, which are perpendicular relative to the cortical surface of M1. For AP-tDCS the current directions reverse (not shown). Based on data from animal models [21], inward and outward electric fields would correspond to preferential pyramidal soma depolarization and hyperpolarization, respectively. For ML-tDCS, the direction of the electric field is predominantly parallel long the cortical surface in central sulcus, which would suggest no dominant pyramidal soma polarization polarity. A following prediction would be that electrode montages that lead to relatively uniform electrical fields directed perpendicular to the cortical surface in M1 should be more efficient in modulating CSE.

Physiological measurements

No significant differences were found between TMS thresholds or amplitudes of the test MEP across sessions. As expected AP TMS thresholds were higher than those for PA stimulation (see Table 1).

PA-versus ML-tDCS: effects on PA-TMS-MEPs

15 individuals in experiment 1 were given 10 min of 1 mA tDCS using each of three separate tDCS montages tested in separate sessions at least one week apart: PA-tDCS, ML-tDCS or sham-tDCS. MEPs were evaluated at rest before and up to 40 min after tDCS

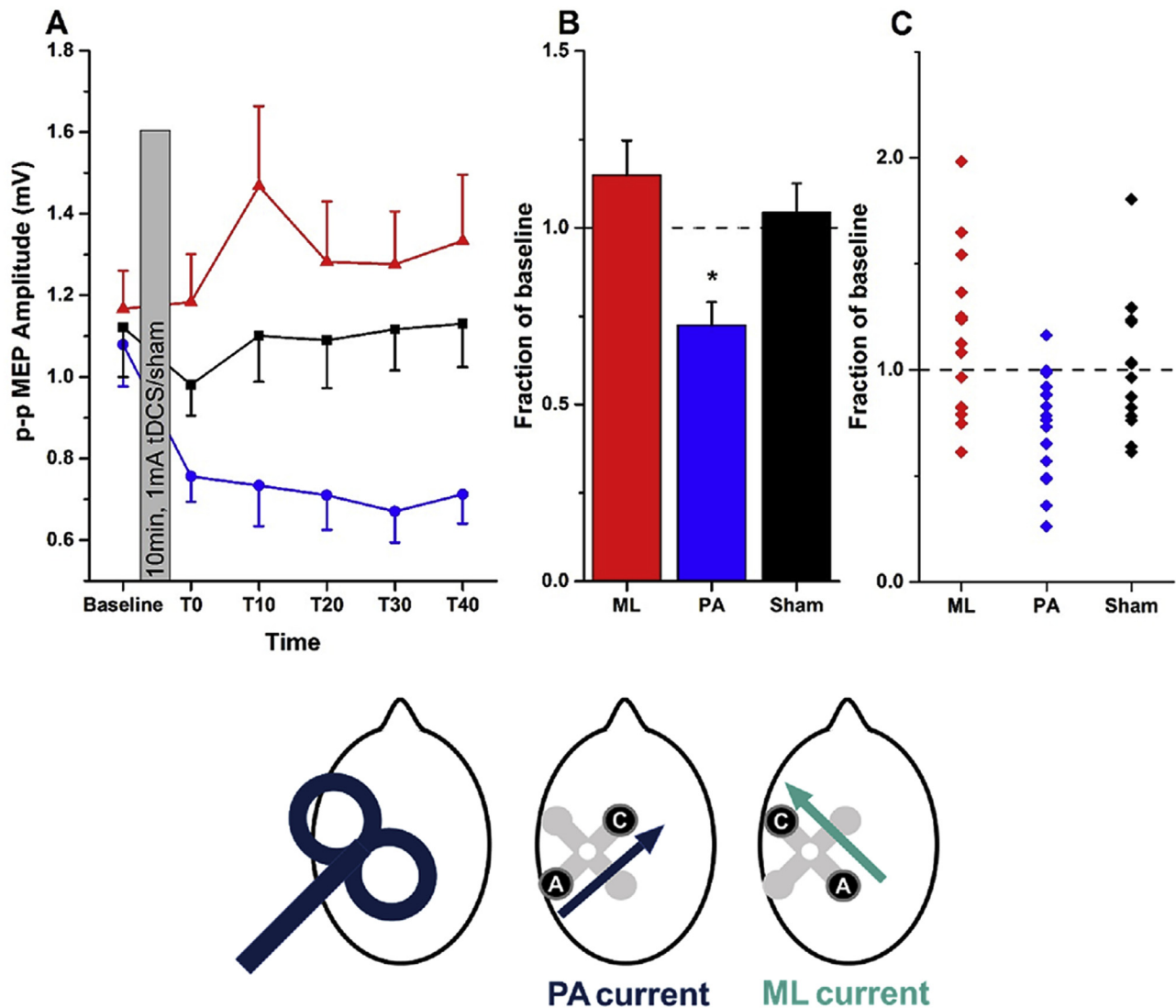


Fig. 2. Effect of PA- and ML-tDCS on the amplitude of MEPs evoked by PA-TMS-MEPs.

Fig. 2 corresponds to experiment 1. **A**, mean (\pm SEM) MEP amplitudes at baseline and every 10 min between T0 and T40 (red triangles = ML-tDCS, black squares = Sham, blue circles = PA-tDCS). **B**, mean (\pm SEM) post-stimulation effect (averaged from T0 to T40) expressed as a fraction of the baseline value in each group (ML, red; PA, blue; sham, black). Asterisks represent paired *t*-test significant differences ($p < 0.05$ with Bonferroni's multiple correction) when compared to sham. **C**, individual data points contributing to the mean values plotted in B. The head diagrams represent coil orientation and electrode configuration used: PA-TMS-MEPs, PA-tDCS and ML-tDCS. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

Table 1

Physiological parameters in each experiment.

	rMT	1 mV MEP Intensity	1 mV MEP Amplitude (mV)
Experiment 1 (n = 15)			
ML-tDCS	40.8 \pm 1.85	49.3 \pm 2.60	1.17 \pm 0.094
PA-tDCS	40.4 \pm 1.84	48.4 \pm 2.81	1.08 \pm 0.102
Sham	40.6 \pm 1.93	48.2 \pm 2.40	1.12 \pm 0.121
Experiment 2 (n = 14)			
PA-TMS		47.5 \pm 3.34*	1.04 \pm 0.057
AP-TMS		60.9 \pm 2.99*	0.98 \pm 0.054
Experiment 3 (n = 14)			
PA-TMS		48.8 \pm 3.44*	1.12 \pm 0.056
AP-TMS		62.3 \pm 3.18*	1.02 \pm 0.057

The first two columns give values for resting motor threshold (rMT) and the intensity required to evoke a 1 mV p-p MEP response (both expressed as percentage of maximum power output). The last column gives the actual MEP amplitude achieved in each condition to approximate the 1 mV target value. Errors are standard error of the mean. Asterisks represent statistically significant differences between rows in the columns ($p < 0.05$).

(Fig. 2A). PA-tDCS decreased the amplitude of MEPs whereas there was no effect of either sham or ML-tDCS. This was confirmed in a two-way repeated measures ANOVA with STIMULATION (PA, ML, sham) and TIME as main factors which showed a significant STIMULATION \times TIME interaction ($F(10, 140) = 1.991$; $p = 0.039$, $\eta^2 = 0.125$) indicating that MEPs were affected differently by each type of tDCS. In order to understand the source of the interaction, the mean post-tDCS effect has been expressed as a fraction of the baseline in Fig. 2B. A one way ANOVA showed a significant effect of type of STIMULATION ($F[2,28] = 7.134$; $p = 0.002$, $\eta^2 = 0.311$), with posthoc (Bonferroni corrected) paired *t*-tests and effect sizes showing a significant difference between the effect of PA-tDCS and sham ($t = -3.279$; $p = 0.005$; $d = -1.110$) and PA v. ML-tDCS ($t = -3.196$; $p = 0.006$) but not between sham and ML-tDCS ($t = 0.760$; $p = 0.46$; $d = 0.302$).

PA-tDCS: contrasting effects on PA- and AP-TMS-MEPs

In experiment 2, we examined whether PA-tDCS had different effects on the MEPs evoked by TMS directed in AP or PA fashion (i.e. the coil handle pointing forward or backward) in 14 participants. Baseline MEPs to each direction of TMS had the same amplitude (AP-TMS-MEPs: $0.983 \text{ mV} \pm 0.201$ vs PA-TMS-MEPs: $1.042 \text{ mV} \pm 0.226$), although the absolute intensity required for AP-TMS-MEPs ($49.1 \pm 2.6\%$) was higher than for PA-TMS-MEPs ($62.9 \pm 2.8\%$). Collapsing all the post-tDCS MEPs and expressing them as a fraction of the baseline (Fig. 3A) revealed a significant effect of PA-tDCS on PA-TMS-MEPs ($t = -2.73$; $p = 0.017$, $d = -0.980$) but no effect on AP-TMS-MEPs. There was a significant difference in the effect on PA-TMS-MEPs vs AP-TMS-MEPs ($t = -2.565$; $p = 0.024$, $d = -0.946$).

AP-tDCS: contrasting effects on PA- and AP-TMS-MEPs

Given the equivocal effects of PA-tDCS on AP-TMS-MEPs (experiment 2), we finally tested whether more consistent effects might be observed on AP-TMS-MEPs when AP-tDCS was employed (experiment 3). In the same participants as in experiment 2 we

compared the effects of AP-tDCS on PA-TMS-MEPs and AP-TMS-MEPs (Fig. 3B). Collapsing all the post-tDCS MEPs and expressing them as a proportion of the baseline, post hoc paired t-tests revealed that there was no difference in the effect of AP-tDCS on MEPs evoked by the two coil orientations ($t = -0.112$; $p = 0.913$, $d = 0.035$) nor were the MEPs to either type of stimulation changed in size after AP-tDCS (Fig. 3B).

Variability in responses to PA-tDCS

In total, 22 different individuals were examined for the effects of PA-tDCS on PA-TMS-MEPs. The results for all of them are plotted in Fig. 4A to illustrate the variability of the effect. Averaging over all post-tDCS time points gives a mean reduction of the MEP to $74.3 \pm 5.1\%$ of baseline values.

In addition, 15 of these participants were tested on two separate occasions. The mean data from session 1 and 2 are shown in Fig. 4B. A two-way ANOVA showed no effect of SESSION ($p = 0.2$, $F [1,14] = 1.808$; $p = 0.2$, $\eta^2 = 0.028$) and no SESSION*TIME interaction ($F (5, 70) = 0.555$; $p = 0.70$, $\eta^2 = 0.223$) indicating no significant differences between the two sessions. There was however, a main effect of TIME ($F (5, 70) = 7.328$; $p = 0.017$, $\eta^2 = 0.630$). The

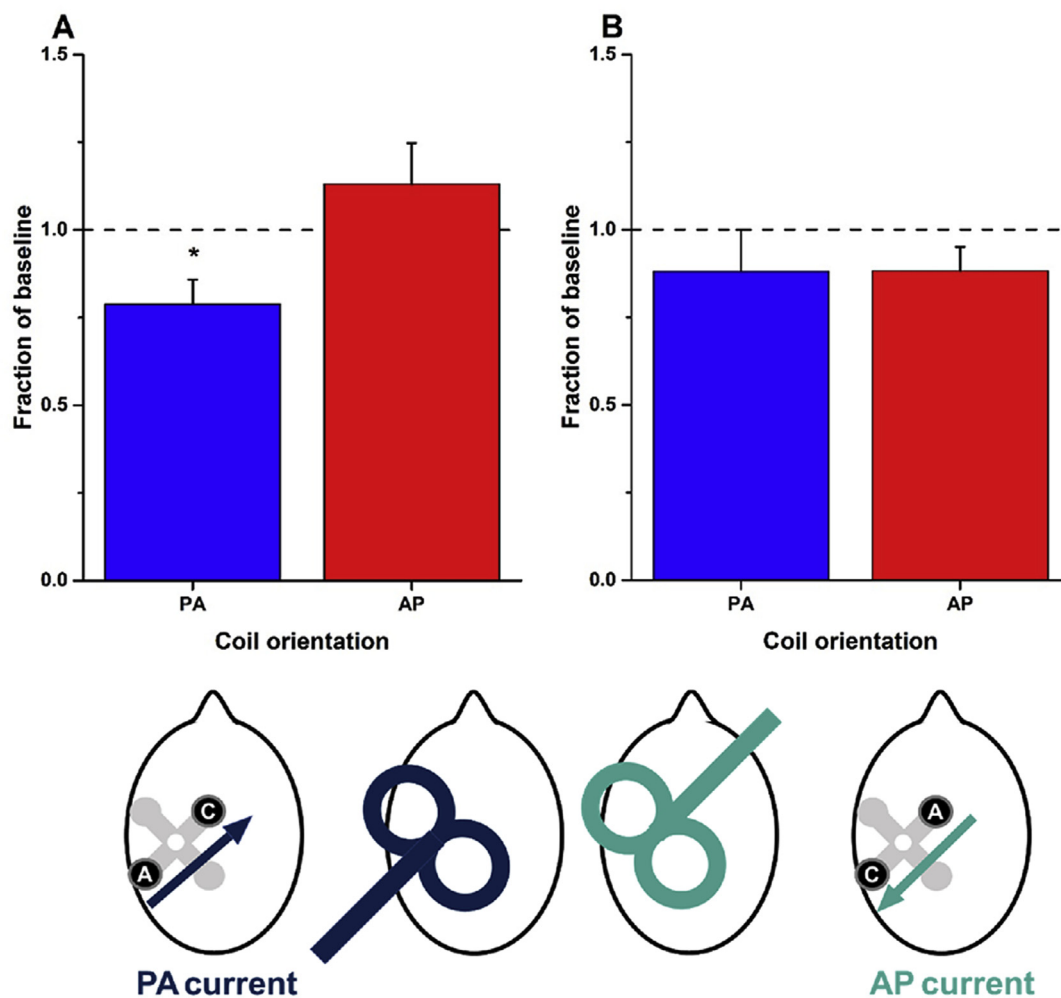


Fig. 3. Effect of PA-tDCS (A) and AP-tDCS (B) on the amplitude of MEPs evoked by PA- and AP-TMS-MEPs.

Plots A and B correspond to experiments 2 and 3 respectively. Graphs plot overall mean (\pm SEM) post-stimulation effects (averaged from T0 to T40 and expressed as a fraction of baseline values) for PA- (blue bars) and AP-TMS-MEPs (red bars). Asterisks represent significant differences between the two coil orientations ($p < 0.05$ with Bonferroni's multiple correction). The head diagrams represent coil orientation and electrode configuration used: PA- and AP-TMS-MEPs, and PA- and AP-tDCS. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

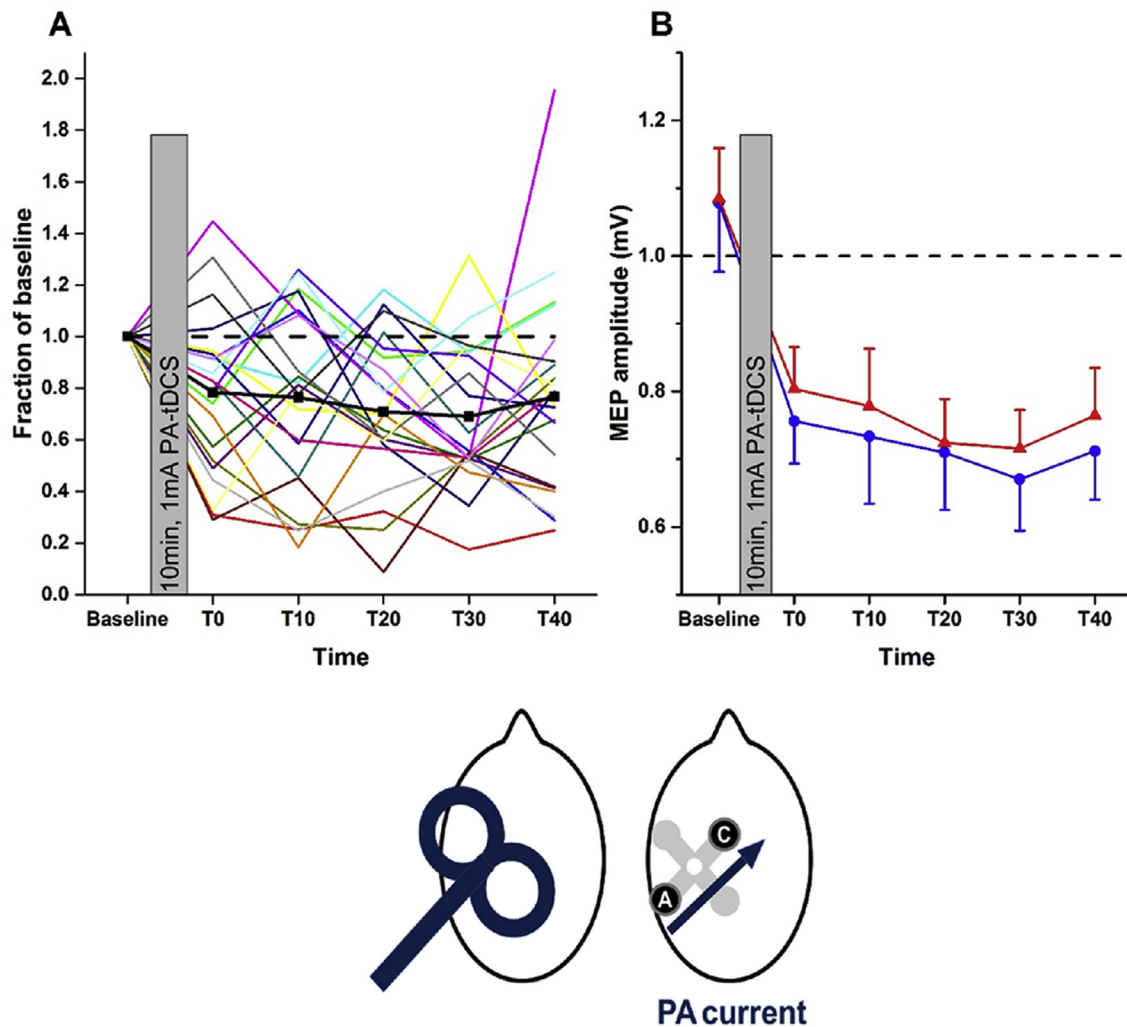


Fig. 4. A, inter-individual variation in the effect of PA-tDCS; B, repeatability of group mean response to PA-tDCS on the amplitude of MEPs evoked by PA-TMS-MEPs. A, In all 22 individuals, MEPs evoked at each post-TDCS time point have been expressed as a fraction of baseline. The solid black line (square symbols) represents the average response from all individuals. B, mean (\pm SEM) amplitudes at baseline and every 10 min, post-tDCS in a group of 15 participants who were tested on 2 separate occasions (red and blue symbols and lines). The head diagrams represent coil orientation and electrode configuration used: PA-TMS-MEPs and PA-tDCS. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

interclass correlation coefficient for the mean percentage reduction in MEP was 0.68, which is generally classified as moderate-strong reproducibility.

Discussion

Selecting the scalp position of electrodes relative to a brain target is a paramount consideration in tDCS [5] – the common bipolar montage places one electrode over the target and the second electrode at some distance. Motivated by previous studies (see below) and specific modelling predictions (Fig. 1), the present study shows that bipolar tDCS produces quantifiable changes in the excitability of primary motor cortex when it is located between the positions of the two stimulation electrodes, as opposed to placing one electrode over M1. For the hand area, the effects on corticospinal excitability depend also on the direction of the electric field: in line with the predictions from current modelling, a montage orthogonal (perpendicular) to the gyrus generated consistently directed electric fields as compared with a montage along (parallel) to the gyrus. This was reflected in the after-effects on MEPs: they were suppressed after delivering perpendicular current but there

was no effect after parallel current. Finally, the direction in which tDCS was applied across the sulcus (i.e. anode anterior or posterior) interacted differentially with the direction of TMS pulses: PA-tDCS affected PA-TMS-MEPs, but had no effect on AP-TMS-MEPs. AP-tDCS had no significant effect on either direction of TMS.

Stimulation between two electrodes

Previous modelling studies have predicted bipolar tDCS will produce cortical electric fields between the two stimulation sites which may be as large or larger than those immediately under the electrodes [16,37,38] – predictions recently validated [12,39,40]. Moreover, our own modelling presented here suggests that perpendicular current flow through the sulcus produces more uniformly directed current at the target site, compared to montages that direct current along the sulcus. This leads to the prediction that tDCS of M1 hand area will have different effects when current is passed between electrodes posterior and anterior to the axis of the central sulcus, compared with electrodes placed medial and lateral. These predictions were indeed borne out by the present results: PA-tDCS leads to aftereffects on corticospinal excitability whereas

ML-tDCS has no effect. This also indicates that not just the strength of current but also its direction with regards to the cortical surface play an important role in mediating changes in corticospinal excitability.

Although it is possible that the posterior anode changes activity in parietal cortex and this secondarily leads to changes in M1, previous work suggests that electrodes positioned more posteriorly do not effectively modulate corticospinal excitability [35]. We conclude that stimulation of sites between two scalp electrodes occurs with bipolar tDCS, as dictated by the physics of current flow, which has important implications for studies seeking to target a specific brain region. Moreover, our results indicate that controlling for the direction of current flow through a target region may help to improve the efficacy of tDCS. For example, it is conceivable that in many studies a mix of PA and ML currents will occur between subjects. If one adds to this the notion that the intensity of stimulation will vary greatly across subjects when controlling stimulator output rather than effectively applied current inside the brain [41,42], we have the situation of large variability in applied current intensity and direction at the presumed target site. Indeed, given our results, in which ML-tDCS produced no reliable effects on CSE, such a mix of current flow direction may contribute to reports of inter-subject variability in physiological and behavioural stimulation outcomes [43–46]. At least for motor cortex, control of current flow direction could be easily achieved based on the optimal orientation and position of TMS for eliciting motor-evoked potentials.

Directionality of tDCS directed across central sulcus

“Anodal stimulation” with a large electrode placed directly over M1 can increase cortical excitability. This is usually explained in the following way. Anodal stimulation produces inward current flow [17,36], though not exclusively depending on cortical folding [23]. Cortical pyramidal neurons, including those on the gyral surface of M1, are aligned perpendicular to the surface of the cortex, such that an inward current flow hyperpolarises their dendrites and depolarises the cell body [21]. Neurophysiological studies in animal indicate that the net effect of this is an increase the excitability of the neuron [47,48] including to synaptic inputs [23], favouring build-up of an LTP-like effect over the 10 min of tDCS [2,49]. This is thought to result in larger MEPs when the same inputs are activated using TMS.

The current flow modelling work presented here shows that our present placement of electrodes produces directional current which, depending on the polarity, enters and exits from the posterior and anterior banks of the precentral gyrus. For PA-tDCS, inward flow on the anterior bank of the central sulcus should polarise the pyramidal neurones in the sulcal wall (which are oriented parallel to the surface of the brain) in the same way as direct “anodal stimulation” over M1 is presumed to operate (though we note this inference stems from animal studies with well-controlled current flow). If so, then we might expect that PA-tDCS has the same effect as conventional anodal tDCS. In fact, the opposite was observed here: 10 min of 1 mA PA-tDCS suppresses MEPs, whereas “anodal stimulation” directly over M1 enhances MEPs, despite this effect also being variable [43].

The different excitation by “anodal tDCS” applied with conventional or focal 4×1 HD electrodes [50], and inhibition by PA-tDCS may be explained by difference in which neuronal elements are modulated. In contrast to the above proposed action on cortical neurons in the gyri-wall by PA-tDCS, “anodal tDCS” may modulate TMS responses by polarisation of cortical neurons specifically in gyral crowns, where inward direct current is indeed more likely

[23]. PA-tDCS will produce current at the gyri-crown parallel to the cortical surface, which is orthogonal to cortical neurons but aligned with cortico-cortical axons afferents. This leads to an alternative hypothesis where PA-tDCS modulates TMS response by polarisation of afferent axons in the gyri crown, which are sensitive to direct current. Animal neurophysiology suggests direct current orientation toward the activated axon terminal (PA in this case) will indeed decrease excitability [23,26].

A range of further alternative explanations can be proposed, that to varying extents explain the lack of the expected modulation of directional tDCS on changes in PA-TMS-MEPs or AP-TMS-MEPs. Even using HD electrodes, tDCS is not focal so net changes motor excitability may reflect actions on other cortical regions [23,51]. A non-trivial dependence on tDCS polarity was already known [8] and the non-linear properties of neurons [52] and networks can produce preferential responses to one tDCS polarity [53].

The seemingly null findings for ML-tDCS are also of interest. Using this montage, any current flowing should flow orthogonal to parts of the hand knob, thereby creating a polarized half and depolarized half. In doing so, one might predict that the heterogeneity in the field induced should cause some subjects to have inhibited responses. However, as can be seen, there is a trend towards an excitatory effect of anodal, ML-tDCS. It would be interesting to individualize tDCS based on high resolution M1 images, in light of the potentially opposing effects on intrinsic hand muscles depending on their medio-lateral location.

The heterogeneity of the induced electric field emphasises the simplistic and outdated notion of anodal-exciting/cathodal-inhibiting tDCS. We suggest that rather than a given montage yielding a particular result, directing the current in such a way gives an overall more consistent effect across the population.

Another explanation for the anisotropic effects of tDCS lies in the difference in magnitude of the electric field induced between PA- and ML-tDCS. The sensitivity of TMS coil orientation is partially explained by higher generated fields for AP directed currents than LM [54]. This has been proposed to be a result of the boundary effect at the CSF-grey matter interface [55]. A similar effect should therefore exist for tDCS generated currents. In fact, the modelling performed for our montages indicated a difference in the magnitude of the currents at the region of interest between PA- and ML-tDCS, which could account for the difference in effects seen on corticospinal excitability.

In general, the nuance in neuromodulation identified here derives from details of cortical folding and cellular morphology. Given that AP-TMS-MEPs and PA-TMS-MEPs activate different inputs to corticospinal output neurones [20] it is not unforeseen that AP-TMS-MEPs were unaffected by PA-tDCS. The dependence on idiosyncratic anatomy (and gradation in coil positioning) may lead to inter-individual variability that masks population effects for many conditions, and produce individual variability.

Variability

We examined the after-effects of PA-tDCS on PA-TMS-MEPs in 22 individuals and found that in 15 of them tDCS reduced corticospinal excitability by 10% or more. Furthermore, the ICC for repeated assessments within an individual was 0.68. Both of these figures are higher than previously reported for standard montage tDCS [56]. It would need a larger study to power this comparison adequately, but it could be that by aligning the tDCS to the individual best direction for PA-TMS-MEPs we achieved an increased uniformity of electric field normal to the surface of M1 compared with a single central anode.

Limitations

It has been pointed out that detailed mapping of the output representation of the hand knob requires that the coil orientation is rotated to maintain perpendicular current at all locations [57]. Since different muscles can be represented at different points on the curved surface of the hand knob, placing tDCS electrodes approximately PA, AP or ML will not ensure optimal stimulation in every individual. However, we hoped to account for this by first finding, in each individual, the best coil orientation for evoking FDI responses, and then using this orientation to guide placement of tDCS electrodes. In doing so, we likely stimulated the majority of FDI output and accounts for the individual differences in hand knob anatomy that undoubtedly exists between people. Therefore, we think that this might be an important feature, rather than a limitation, as it potentially could be exploited for individualized “dose-control” without requiring complex and expensive current flow modelling, which many users will not have access to.

The effects of PA- and ML-tDCS indeed outlasted the period of stimulation. However, corticospinal excitability did not return to baseline by the end of the testing period. Unfortunately, we did not track excitability changes for longer than 40 min and so cannot provide any concrete statements on the duration of offline effects. Hence, further experiments should assess this in more detail.

Conclusion

We provide strong support for the notion that bipolar tDCS has effects on cortex between the primary sites of stimulation, as predicted by models of current flow in the brain. Furthermore, these effects can be directionally-dependent. These factors may be important when interpreting (and comparing) results from conventional tDCS. More generally, our results indicate how current flow models can guide electrode placement and motivate experimental questions concerning the key factors for optimizing tES.

References

- Nitsche MA, Paulus W. Excitability changes induced in the human motor cortex by weak transcranial direct current stimulation. *J Physiol* 2000;527(3): 633–9.
- Bindman LJ, Lippold OC, Redfearn JW. The action of brief polarizing currents on the cerebral cortex of the rat (1) during current flow and (2) in the production of long-lasting after-effects. *J Physiol* 1964;172:369–82.
- Nitsche MA, Paulus W. Sustained excitability elevations induced by transcranial DC motor cortex stimulation in humans. *Neurology* 2001;57(10): 1899–901.
- Nitsche MA, Doemkes S, Karakose T, Antal A, Liebetanz D, Lang N, et al. Shaping the effects of transcranial direct current stimulation of the human motor cortex. *J Neurophysiol* 2007;97(4):3109–17.
- Brunoni AR, Nitsche MA, Bolognini N, Bikson M, Wagner T, Merabet L, et al. Clinical research with transcranial direct current stimulation (tDCS): challenges and future directions. *Brain Stimul* 2012;5(3):175–95.
- Lefaucheur JP, Antal A, Ayache SS, Benninger DH, Brunelin J, Cogiamanian F, et al. Evidence-based guidelines on the therapeutic use of transcranial direct current stimulation (tDCS). *Clin Neurophysiol* 2017;128(1):56–92.
- Nitsche MA, Fricke K, Henschke U, Schlitterlau A, Liebetanz D, Lang N, et al. Pharmacological modulation of cortical excitability shifts induced by transcranial direct current stimulation in humans. *J Physiol* 2003;553(Pt 1): 293–301.
- Batsikadze G, Moliadze V, Paulus W, Kuo MF, Nitsche MA. Partially non-linear stimulation intensity-dependent effects of direct current stimulation on motor cortex excitability in humans. *J Physiol* 2013;591(7):1987–2000.
- Thirugnanasambandam N, Grundey J, Adam K, Drees A, Skwirba AC, Lang N, et al. Nicotinic impact on focal and non-focal neuroplasticity induced by non-invasive brain stimulation in non-smoking humans. *Neuropsychopharmacology* 2011;36(4):879–86.
- Jamil A, Batsikadze G, Kuo HI, Labruna L, Hasan A, Paulus W, et al. Systematic evaluation of the impact of stimulation intensity on neuroplastic after-effects induced by transcranial direct current stimulation. *J Physiol* 2017;595(4): 1273–88.
- Ammann C, Lindquist MA, Celnik PA. Response variability of different anodal transcranial direct current stimulation intensities across multiple sessions. *Brain Stimul* 2017;10(4):757–63.
- Huang Y, Liu AA, Lafon B, Friedman D, Dayan M, Wang X, et al. Measurements and models of electric fields in the in vivo human brain during transcranial electric stimulation. *eLife* 2017;6.
- Lang N, Siebner HR, Ward NS, Lee L, Nitsche MA, Paulus W, et al. How does transcranial DC stimulation of the primary motor cortex alter regional neuronal activity in the human brain? *Eur J Neurosci* 2005;22(2):495–504.
- Pena-Gomez C, Sala-Lonch R, Junque C, Clemente IC, Vidal D, Bargallo N, et al. Modulation of large-scale brain networks by transcranial direct current stimulation evidenced by resting-state functional MRI. *Brain Stimul* 2012;5(3):252–63.
- Datta A, Bansal V, Diaz J, Patel J, Reato D, Bikson M. Gyri-precise head model of transcranial direct current stimulation: improved spatial focality using a ring electrode versus conventional rectangular pad. *Brain Stimul* 2009;2(4):201–7. e1.
- Dmochowski JP, Datta A, Bikson M, Su Y, Parra LC. Optimized multi-electrode stimulation increases focality and intensity at target. *J Neural Eng* 2011;8(4): 046011.
- Wagner T, Fregni F, Fecteau S, Grodzinsky A, Zahn M, Pascual-Leone A. Transcranial direct current stimulation: a computer-based human model study. *Neuroimage* 2007;35(3):1113–24.
- Teichmann M, Lesoil C, Godard J, Vernet M, Bertrand A, Levy R, et al. Direct current stimulation over the anterior temporal areas boosts semantic processing in primary progressive aphasia. *Ann Neurol* 2016;80(5):693–707.
- Seibt O, Brunoni AR, Huang Y, Bikson M. The pursuit of DLPCF: non-neuronavigated methods to target the left dorsolateral pre-frontal cortex with symmetric bicephalic transcranial direct current stimulation (tDCS). *Brain Stimul* 2015;8(3):590–602.
- Di Lazzaro V, Profice P, Ranieri F, Capone F, Dileone M, Oliviero A, et al. I-wave origin and modulation. *Brain Stimul* 2012;5(4):512–25.
- Radman T, Ramos RL, Brumberg JC, Bikson M. Role of cortical cell type and morphology in subthreshold and suprathreshold uniform electric field stimulation in vitro. *Brain Stimul* 2009;2(4). 215–28, 28 e1–3.
- Dmochowski JP, Bikson M, Datta A, Richardson J, Fridriksson J, Parra LC. On the role of electric field orientation in optimal design of transcranial current stimulation. *Conf Proc IEEE Eng Med Biol Soc* 2012;2012:6426–9.
- Rahman A, Reato D, Arlotti M, Gasca F, Datta A, Parra LC, et al. Cellular effects of acute direct current stimulation: somatic and synaptic terminal effects. *J Physiol* 2013;591(10):2563–78.
- Mills KR, Boniface SJ, Schubert M. Magnetic brain stimulation with a double coil: the importance of coil orientation. *Electroencephalogr Clin Neurophysiol* 1992;85(1):17–21.
- Werhahn KJ, Fong JK, Meyer BU, Priori A, Rothwell JC, Day BL, et al. The effect of magnetic coil orientation on the latency of surface EMG and single motor unit responses in the first dorsal interosseous muscle. *Electroencephalogr Clin Neurophysiol* 1994;93(2):138–46.
- Bikson M, Inoue M, Akiyama H, Deans JK, Fox JE, Miyakawa H, et al. Effects of uniform extracellular DC electric fields on excitability in rat hippocampal slices in vitro. *J Physiol* 2004;557(1):175–90.
- Truong DQ, Magerowski G, Blackburn GL, Bikson M, Alonso-Alonso M. Computational modeling of transcranial direct current stimulation (tDCS) in obesity: impact of head fat and dose guidelines. *NeuroImage Clin* 2013;2: 759–66.
- Ashburner J, Friston KJ. Unified segmentation. *NeuroImage* 2005;26(3): 839–51.
- Penny WD, Friston KJ, Ashburner JT, Kiebel SJ, Nichols TE. Statistical parametric mapping: the analysis of functional brain images. Academic Press; 2011. 2011/04/28/ 689 p.
- Huang Y, Dmochowski JP, Su Y, Datta A, Rorden C, Parra LC. Automated MRI segmentation for individualized modeling of current flow in the human head. *J Neural Eng* 2013;10(6):066004.
- Wagner T, Fregni F, Fecteau S, Grodzinsky A, Zahn M, Pascual-Leone A. Transcranial direct current stimulation: a computer-based human model study. *NeuroImage* 2007;35(3):1113–24.
- Gabriel C, Gabriel S, Corthout E. The dielectric properties of biological tissues: I. Literature survey. *Phys Med Biol* 1996;41(11):2231–49.
- Huang Y, Liu AA, Lafon B, Friedman D, Dayan M, Wang X, et al. Measurements and models of electric fields in the in vivo human brain during transcranial electric stimulation. *eLife* 2017;6:e18834.
- Rossini PM, Barker AT, Berardelli A, Caramia MD, Caruso G, Cracco RQ, et al. Non-invasive electrical and magnetic stimulation of the brain, spinal cord and roots: basic principles and procedures for routine clinical application. Report of an IFCN committee. *Electroencephalogr Clin Neurophysiol* 1994;91(2): 79–92.
- Kojima S, Onishi H, Miyaguchi S, Kotan S, Sugawara K, Kirimoto H, et al. Effects of cathodal transcranial direct current stimulation to primary somatosensory cortex on short-latency afferent inhibition. *Neuroreport* 2015;26(11):634–7.
- Datta A, Elwassif M, Battaglia F, Bikson M. Transcranial current stimulation focality using disc and ring electrode configurations: FEM analysis. *J Neural Eng* 2008;5(2):163–74.
- Miranda PC, Lomarev M, Hallett M. Modeling the current distribution during transcranial direct current stimulation. *Clin Neurophysiol* 2006;117(7): 1623–9.

- [38] Rampersad SM, Janssen AM, Lucka F, Aydin U, Lanfer B, Lew S, et al. Simulating transcranial direct current stimulation with a detailed anisotropic human head model. *IEEE Trans Neural Syst Rehabil Eng* 2014;22(3):441–52.
- [39] Jog MV, Smith RX, Jann K, Dunn W, Lafon B, Truong D, et al. In-vivo imaging of magnetic fields induced by transcranial direct current stimulation (tDCS) in human brain using MRI. *Sci Rep* 2016;6:34385.
- [40] Opitz A, Falchier A, Yan CG, Yeagle EM, Linn GS, Megevand P, et al. Spatio-temporal structure of intracranial electric fields induced by transcranial electric stimulation in humans and nonhuman primates. *Sci Rep* 2016;6:31236.
- [41] Laakso I, Tanaka S, Koyama S, De Santis V, Hirata A. Inter-subject variability in electric fields of motor cortical tDCS. *Brain Stimul* 2015;8(5):906–13.
- [42] Bestmann S, Ward N. Are current flow models for transcranial electrical stimulation fit for purpose? *Brain Stimul* 2017;10(4):865–6.
- [43] Horvath JC, Forte JD, Carter O. Evidence that transcranial direct current stimulation (tDCS) generates little-to-no reliable neurophysiologic effect beyond MEP amplitude modulation in healthy human subjects: a systematic review. *Neuropsychologia* 2015;66:213–36.
- [44] Tremblay S, Hannah R, Rawji V, Rothwell JC. Modulation of iTBS after-effects via concurrent directional TDCS: a proof of principle study. *Brain Stimul* 2017;10(4):744–7.
- [45] Wiethoff S, Hamada M, Rothwell JC. Variability in response to transcranial direct current stimulation of the motor cortex. *Brain Stimul* 2014;7(3):468–75.
- [46] Li LM, Uehara K, Hanakawa T. The contribution of interindividual factors to variability of response in transcranial direct current stimulation studies. *Front Cell Neurosci* 2015;9:181.
- [47] Gartside IB. Mechanisms of sustained increases of firing rate of neurones in the rat cerebral cortex after polarization: role of protein synthesis. *Nature* 1968;220(5165):383–4.
- [48] Radman T, Su Y, An JH, Parra LC, Bikson M. Spike timing amplifies the effect of electric fields on neurons: implications for endogenous field effects. *J Neurosci* 2007;27(11):3030–6.
- [49] Fritsch B, Reis J, Martinowich K, Schambra HM, Ji Y, Cohen LG, et al. Direct current stimulation promotes BDNF-dependent synaptic plasticity: potential implications for motor learning. *Neuron* 2010;66(2):198–204.
- [50] Kuo HI, Bikson M, Datta A, Minhas P, Paulus W, Kuo MF, et al. Comparing cortical plasticity induced by conventional and high-definition 4 x 1 ring tDCS: a neurophysiological study. *Brain Stimul* 2013;6(4):644–8.
- [51] Polania R, Nitsche MA, Paulus W. Modulating functional connectivity patterns and topological functional organization of the human brain with transcranial direct current stimulation. *Hum Brain Mapp* 2011;32(8):1236–49.
- [52] Lafon B, Rahman A, Bikson M, Parra LC. Direct current stimulation alters neuronal input/output function. *Brain Stimul* 2017;10(1):36–45.
- [53] Reato D, Rahman A, Bikson M, Parra LC. Low-intensity electrical stimulation affects network dynamics by modulating population rate and spike timing. *J Neurosci* 2010;30(45):15067–79.
- [54] Thielscher A, Opitz A, Windhoff M. Impact of the gyral geometry on the electric field induced by transcranial magnetic stimulation. *Neuroimage* 2011;54(1):234–43.
- [55] Miranda PC, Hallett M, Basser PJ. The electric field induced in the brain by magnetic stimulation: a 3-D finite-element analysis of the effect of tissue heterogeneity and anisotropy. *IEEE Trans Biomed Eng* 2003;50(9):1074–85.
- [56] Lopez-Alonso V, Fernandez-Del-Olmo M, Costantini A, Gonzalez-Henriquez JJ, Cheeran B. Intra-individual variability in the response to anodal transcranial direct current stimulation. *Clin Neurophysiol* 2015;126(12):2342–7.
- [57] Raffin E, Pellegrino G, Di Lazzaro V, Thielscher A, Siebner HR. Bringing transcranial mapping into shape: sulcus-aligned mapping captures motor somatotopy in human primary motor hand area. *Neuroimage* 2015;120:164–75.