

Studies toward a prebiotic protometabolism

Adam J. Coggins

UCL

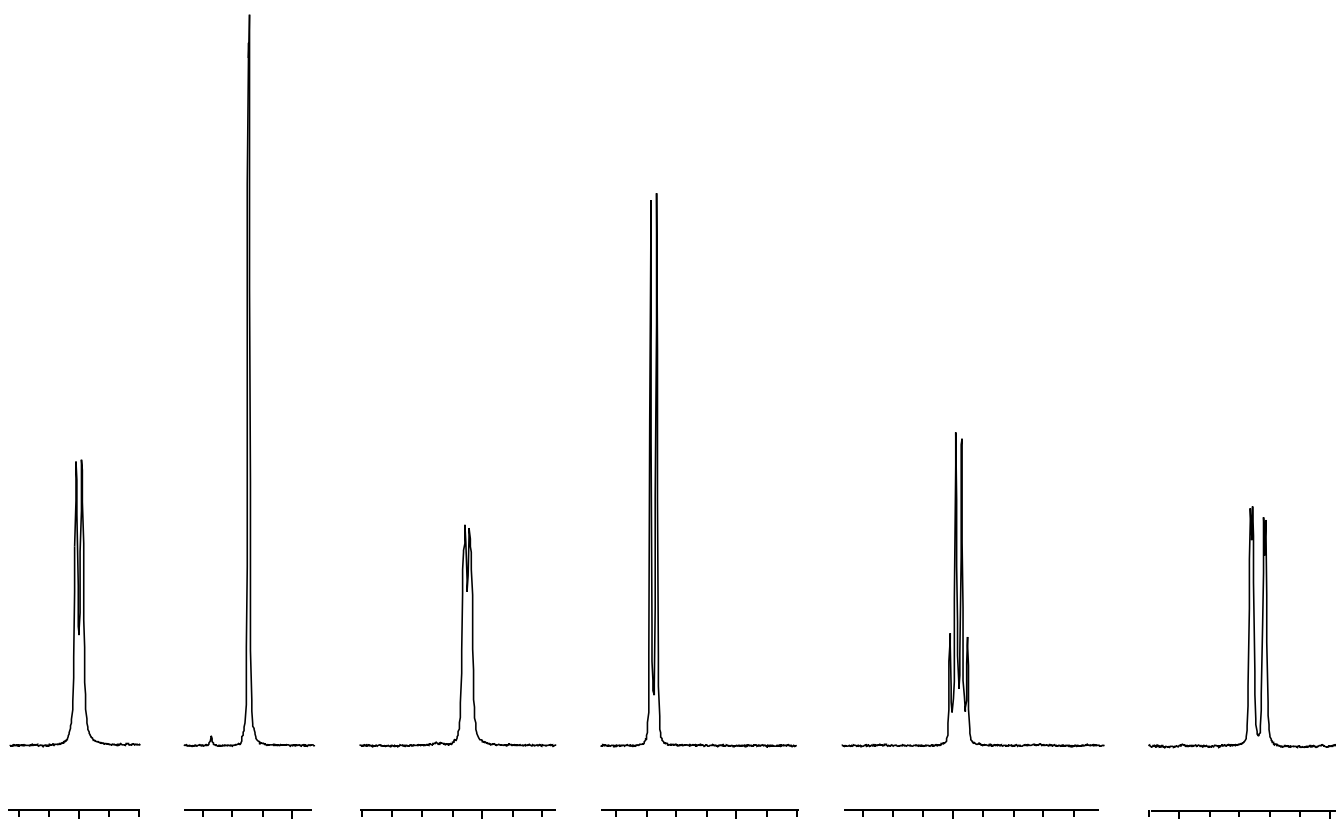
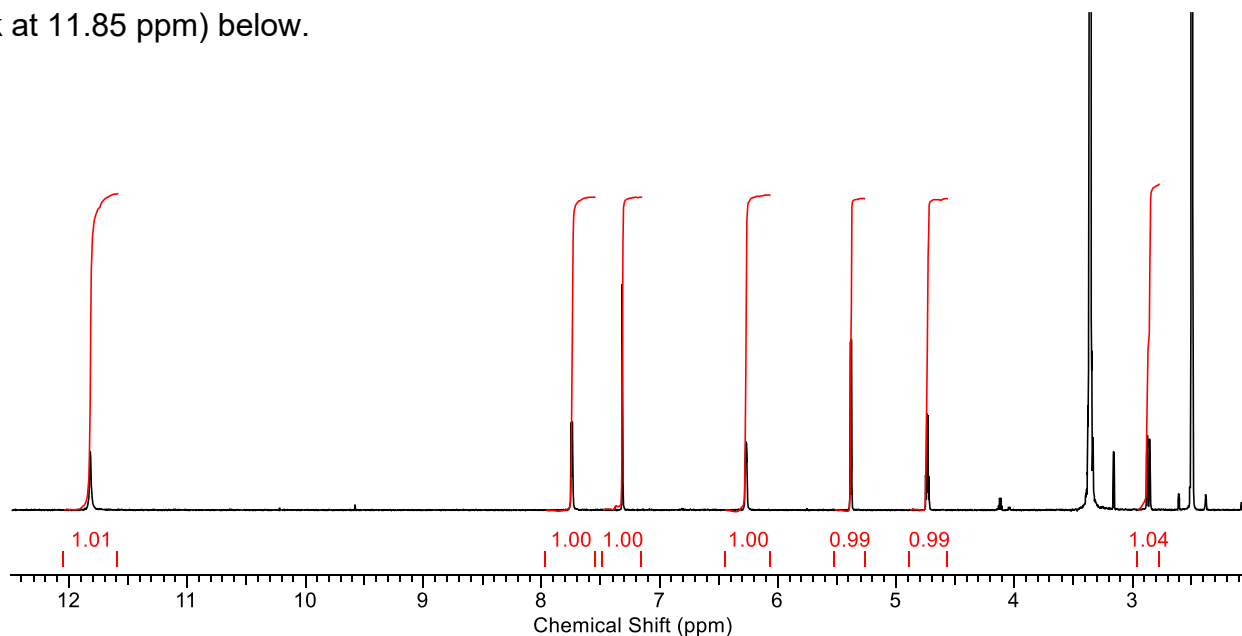
*Thesis Submitted to University College London
for the Degree of Doctor of Philosophy*

Appendix – part 4

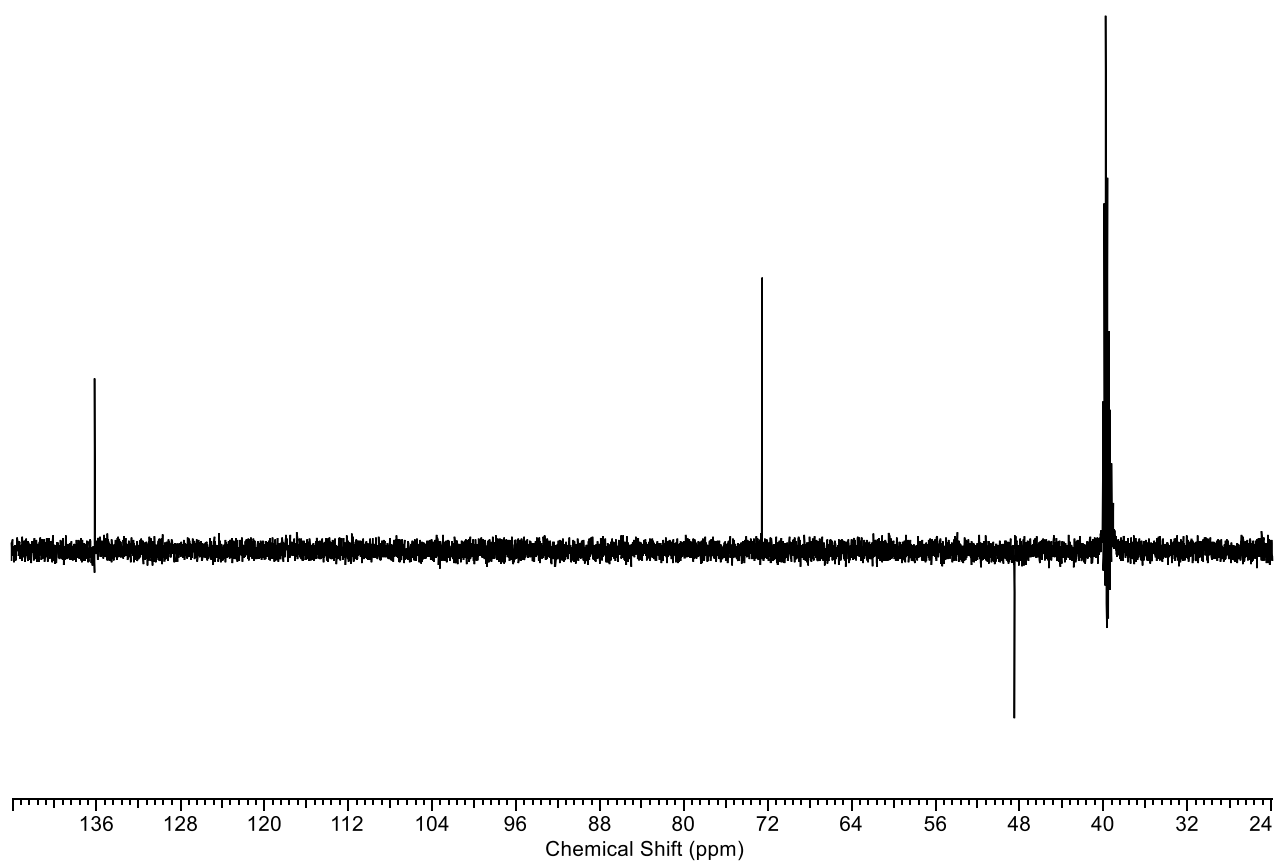
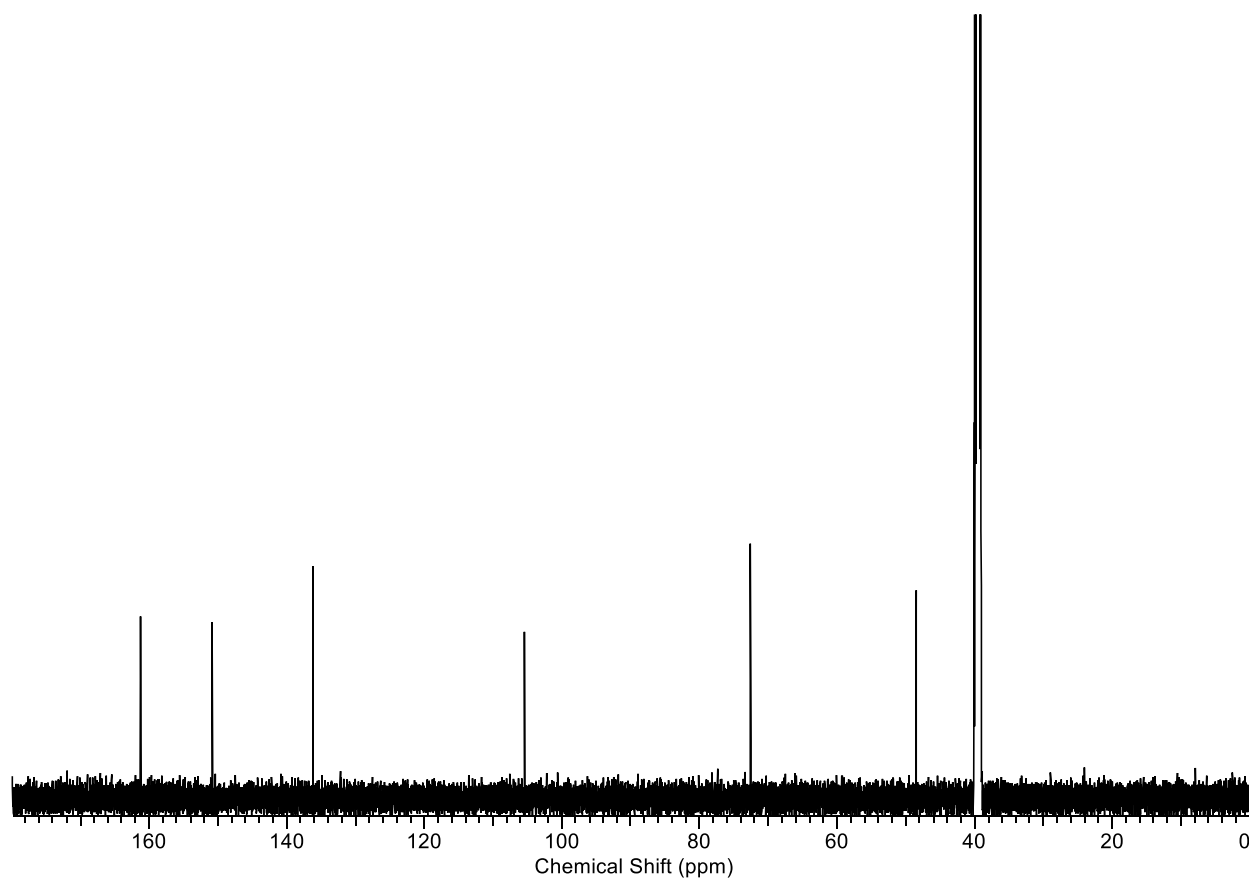
Purine-Precursor Tethering by Imine Tautomerisation

Azepinomycin (212)

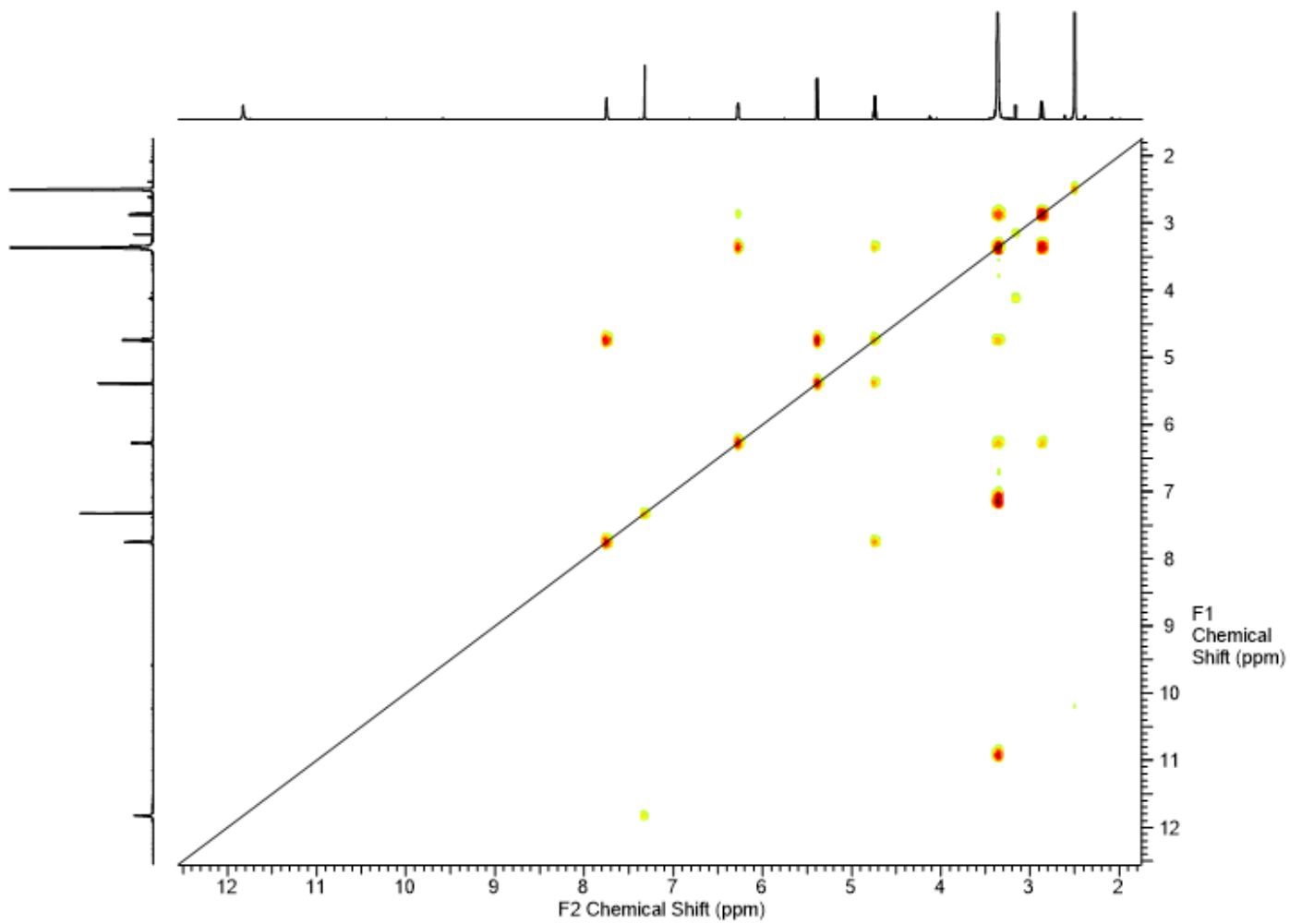
A333: ^1H NMR spectrum (600 MHz, {DMSO- d_6 }, 2.0 – 12.5 ppm) of **212**, with expansions (excluding peak at 11.85 ppm) below.



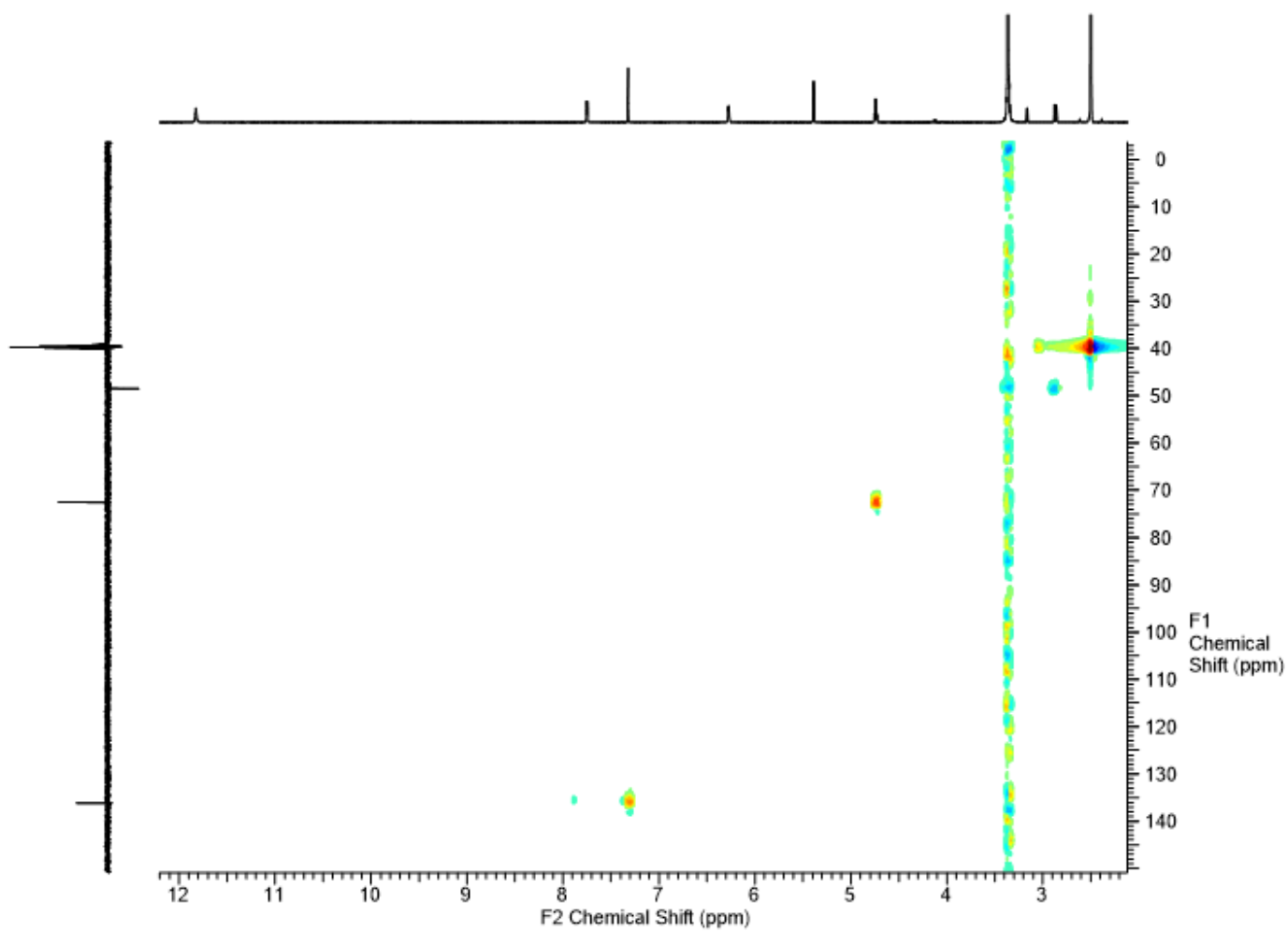
A334: ^{13}C NMR spectrum (151 MHz, {DMSO- d_6 }, 0 – 180 ppm) of **212** with DEPT135 spectrum (24 – 144 ppm) below.



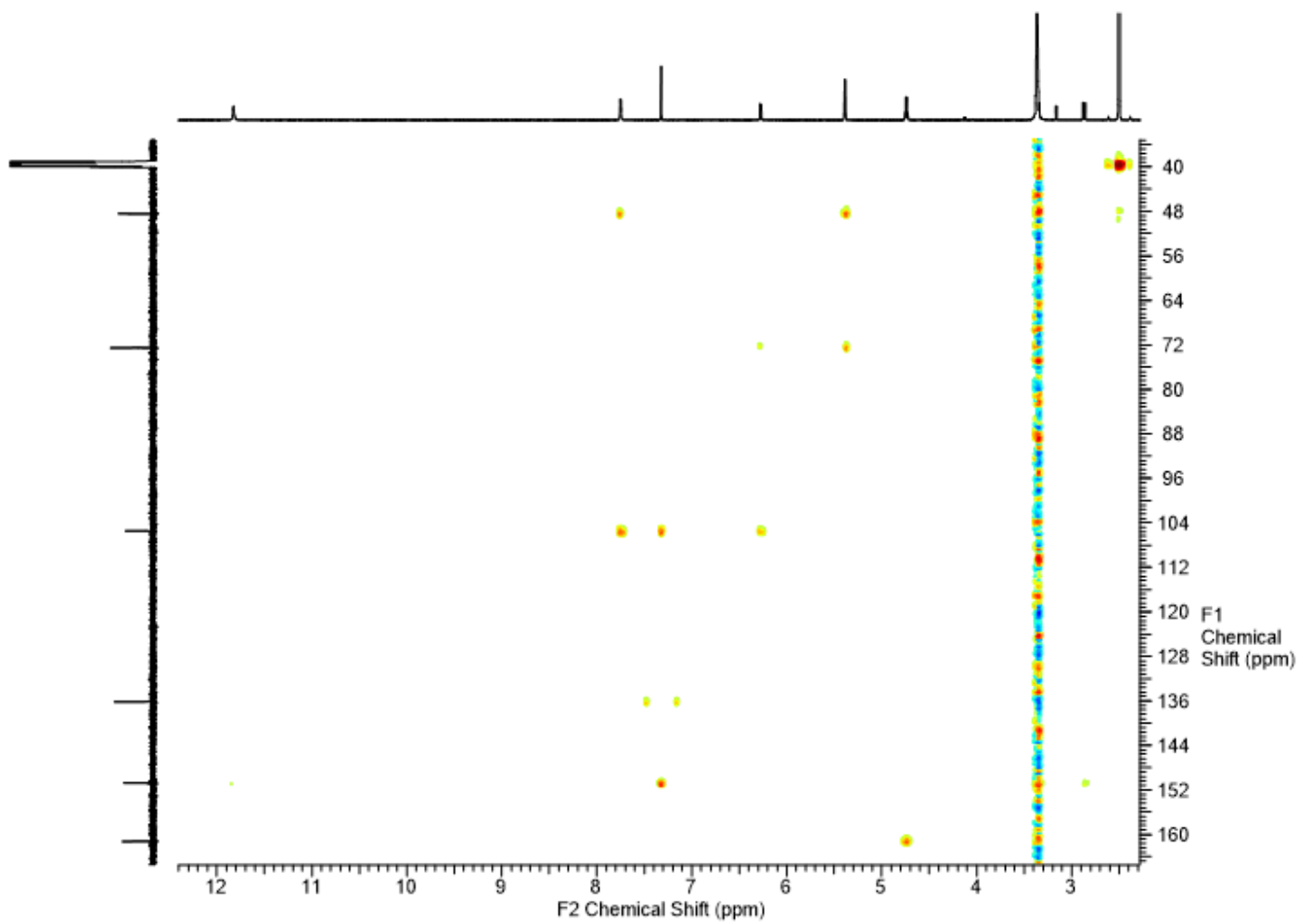
A335: ^1H - ^1H COSY NMR spectrum (600 MHz, {DMSO- d_6 }) of **212**.



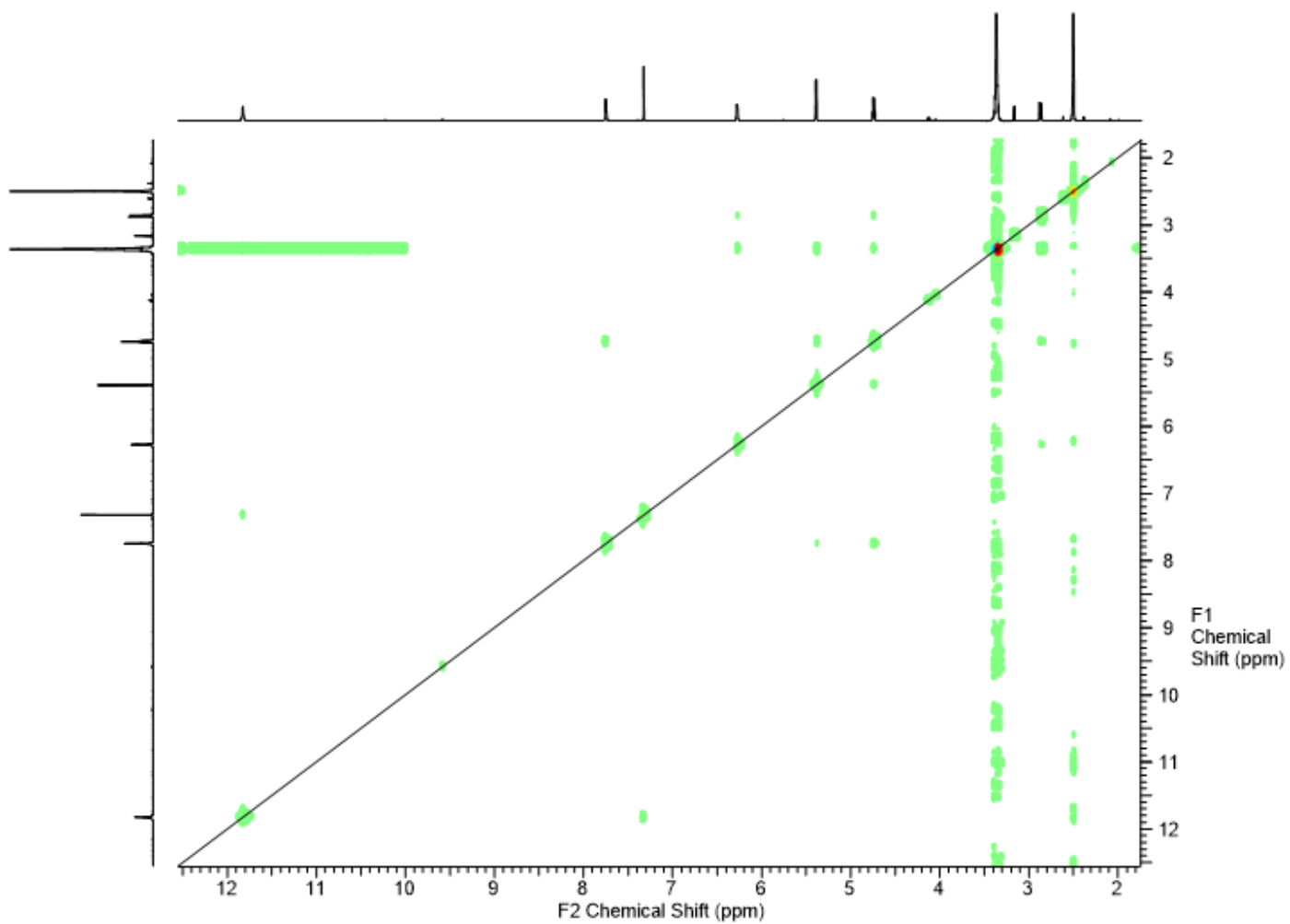
A336: ^1H - ^{13}C HSQC NMR spectrum (600 MHz, {DMSO-d₆}) of **212**.



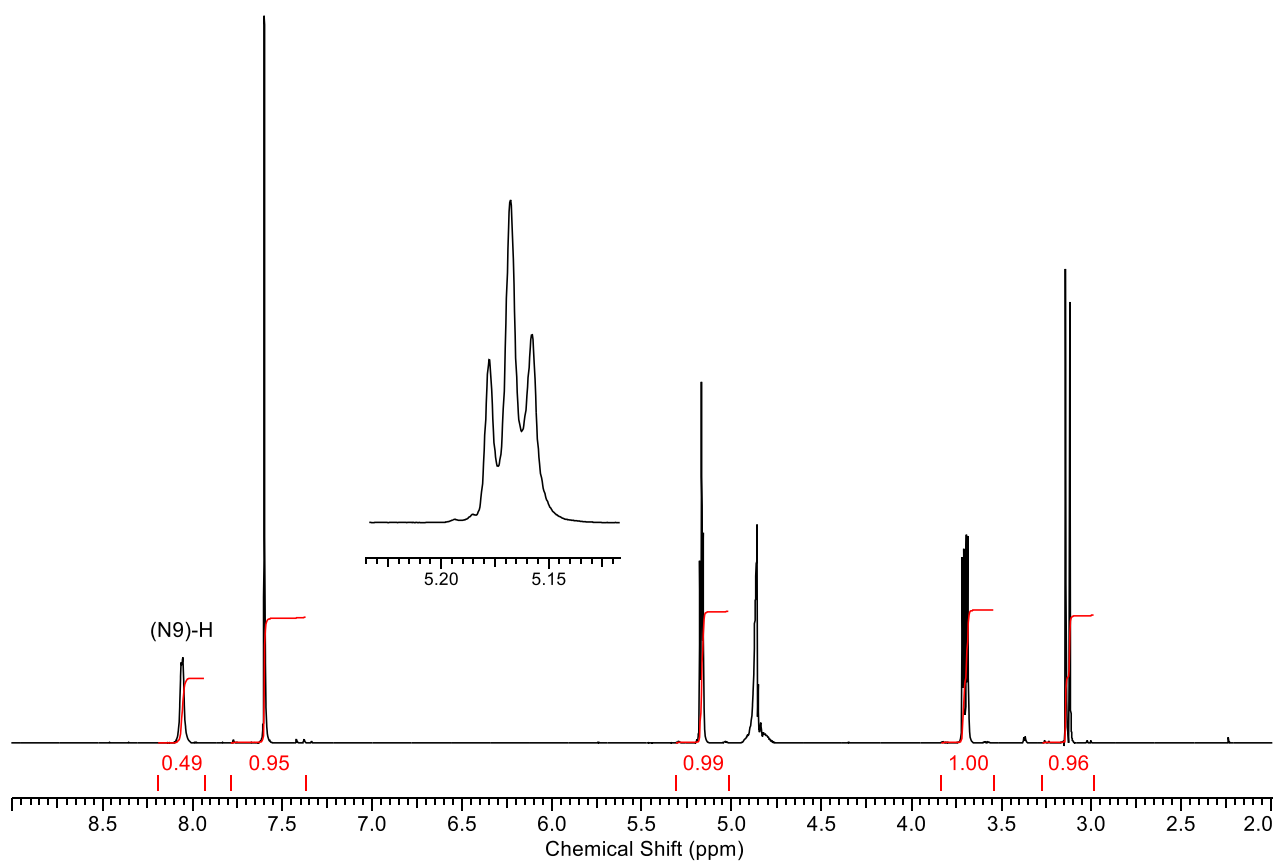
A337: ^1H - ^{13}C HMBC NMR spectrum (600 MHz, {DMSO- d_6 }) of **212**.



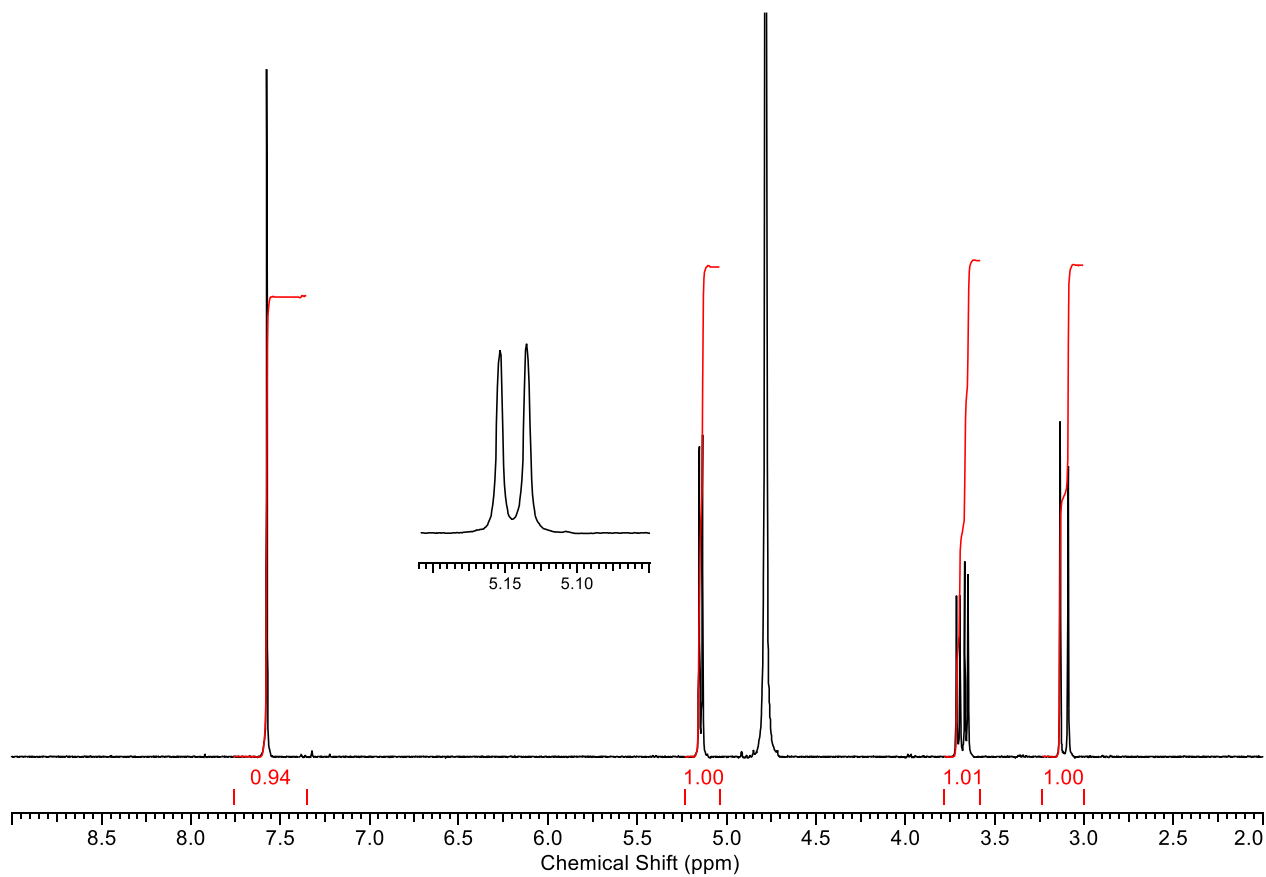
A338: ^1H - ^1H NOESY NMR spectrum (600 MHz, {DMSO- d_6 }) of **212**.



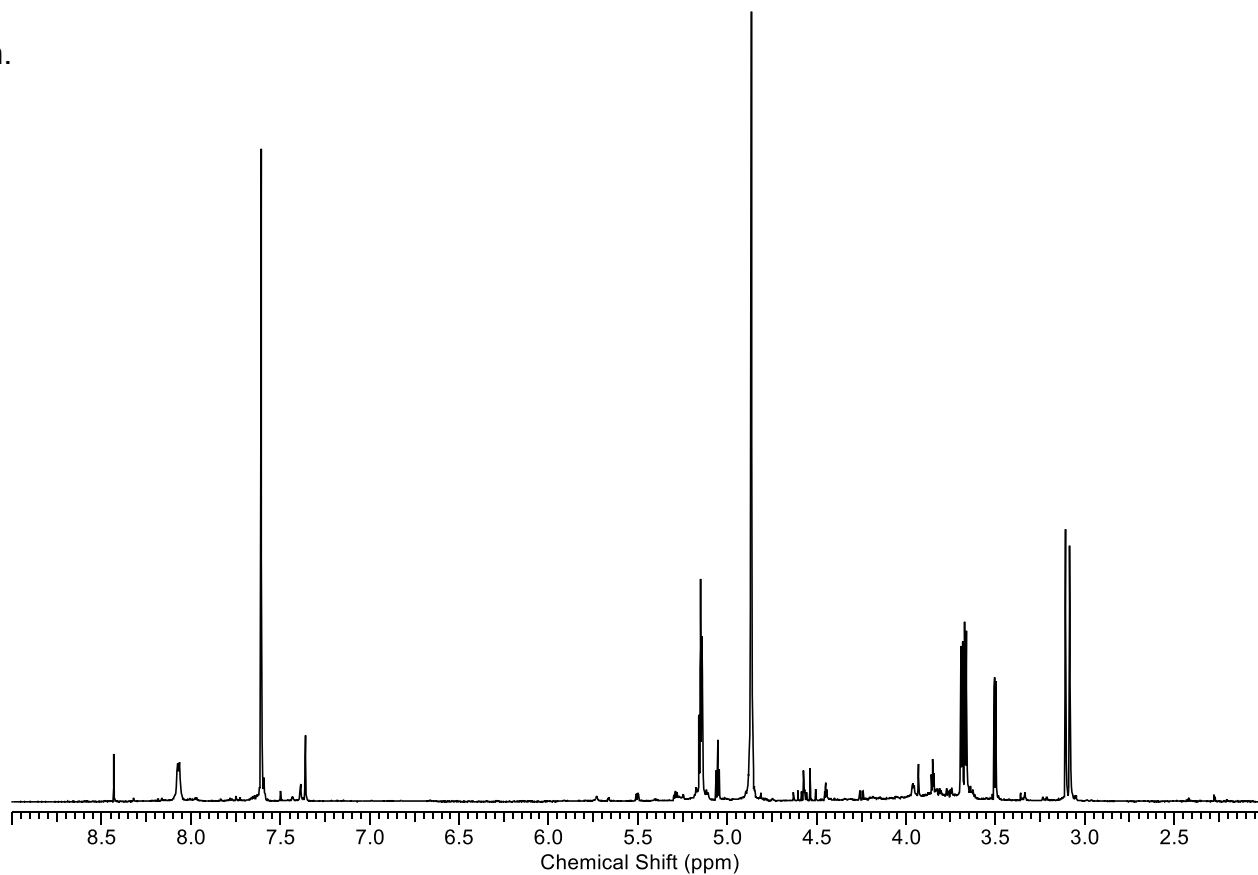
A339: ^1H NMR spectrum (600 MHz, $\{\text{H}_2\text{O}/\text{D}_2\text{O}$. 9:1}, 2.0 – 9.0 ppm) of **212** with expansion overlaid.



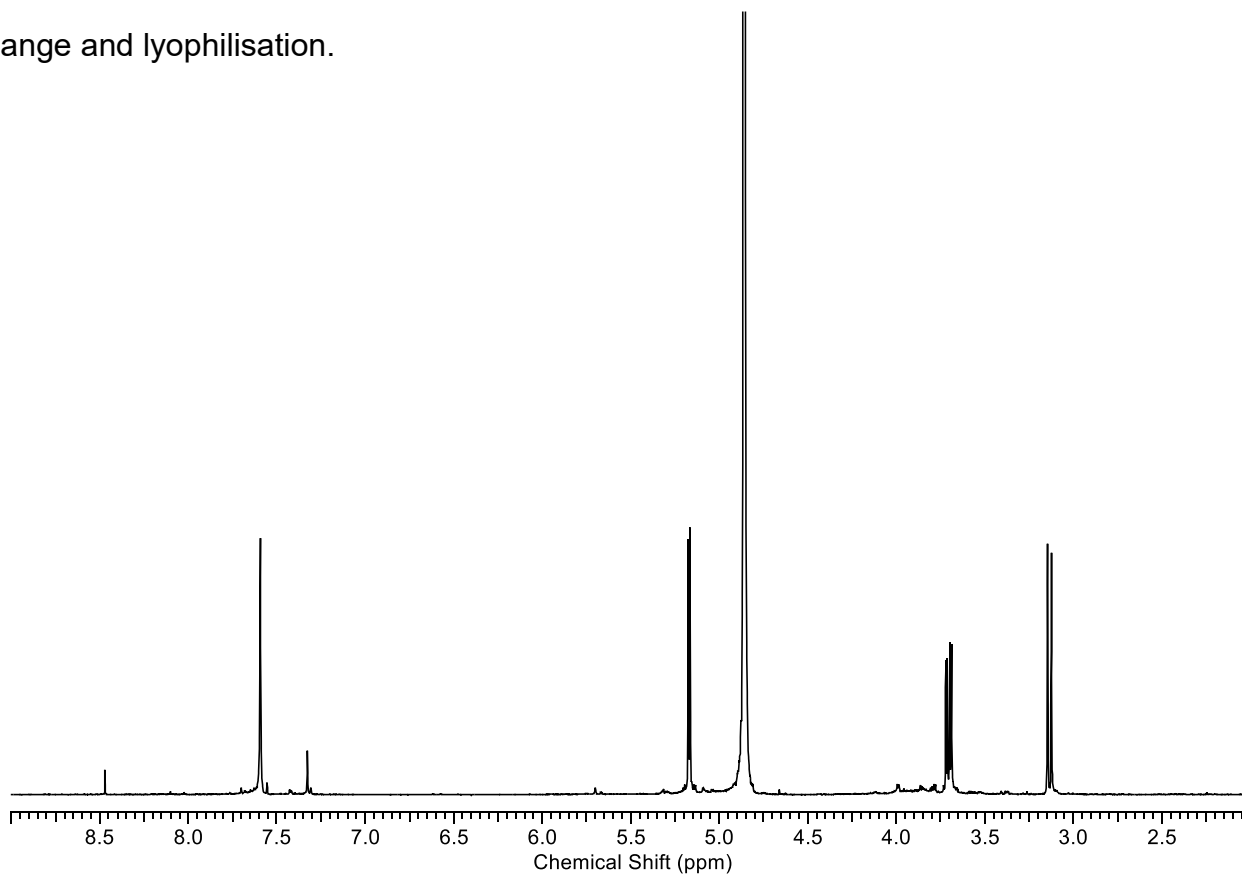
A340: ^1H NMR spectrum (600 MHz, $\{\text{D}_2\text{O}\}$, 2.0 – 9.0 ppm) of **212** with expansion overlaid.



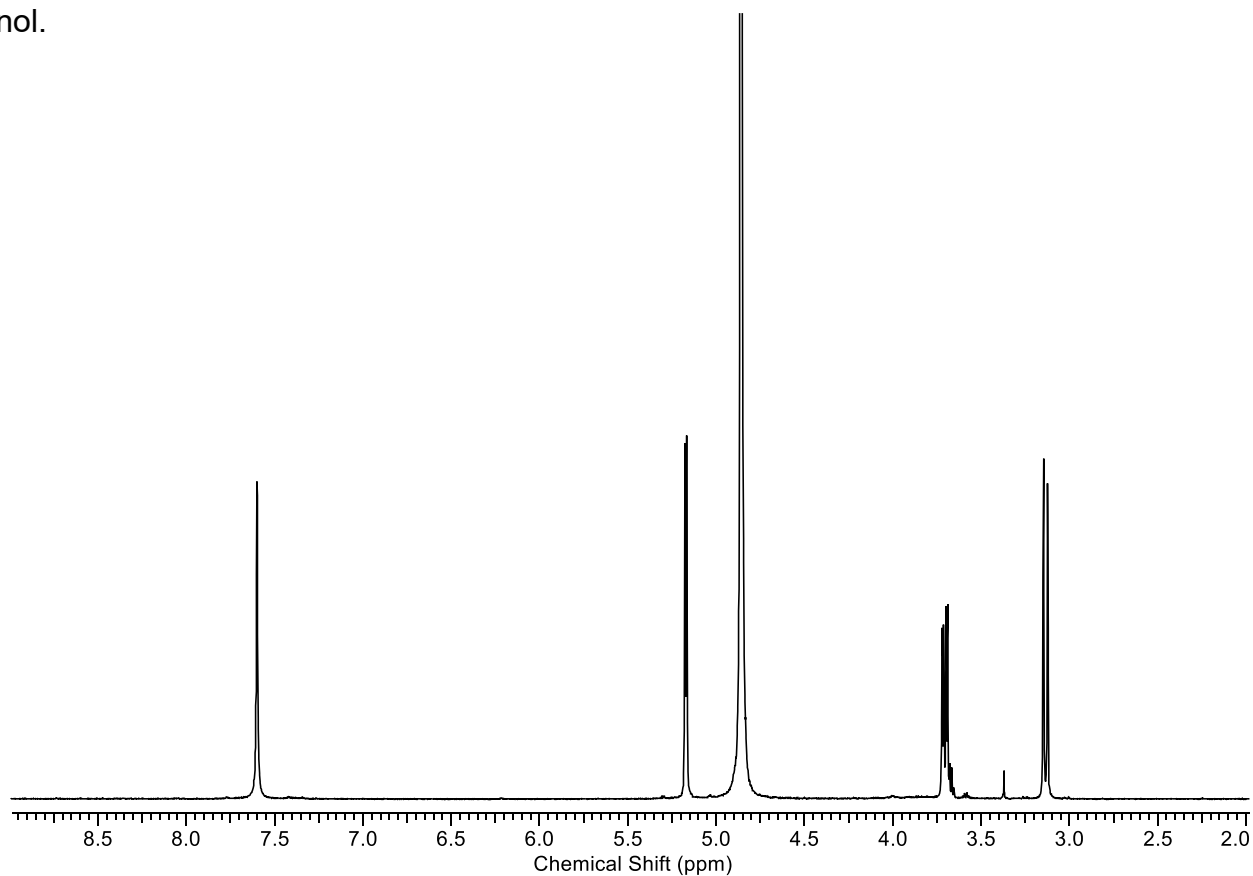
A341: ^1H NMR spectrum (400 MHz, $\{\text{H}_2\text{O}/\text{D}_2\text{O}, 9:1\}$, 2.0 – 9.0 ppm) of **212** reaction mixture after 28 h.



A342: ^1H NMR spectrum (400 MHz, $\{\text{D}_2\text{O}\}$, 2.0 – 9.0 ppm) of **212** crude product after DowexTM resin exchange and lyophilisation.

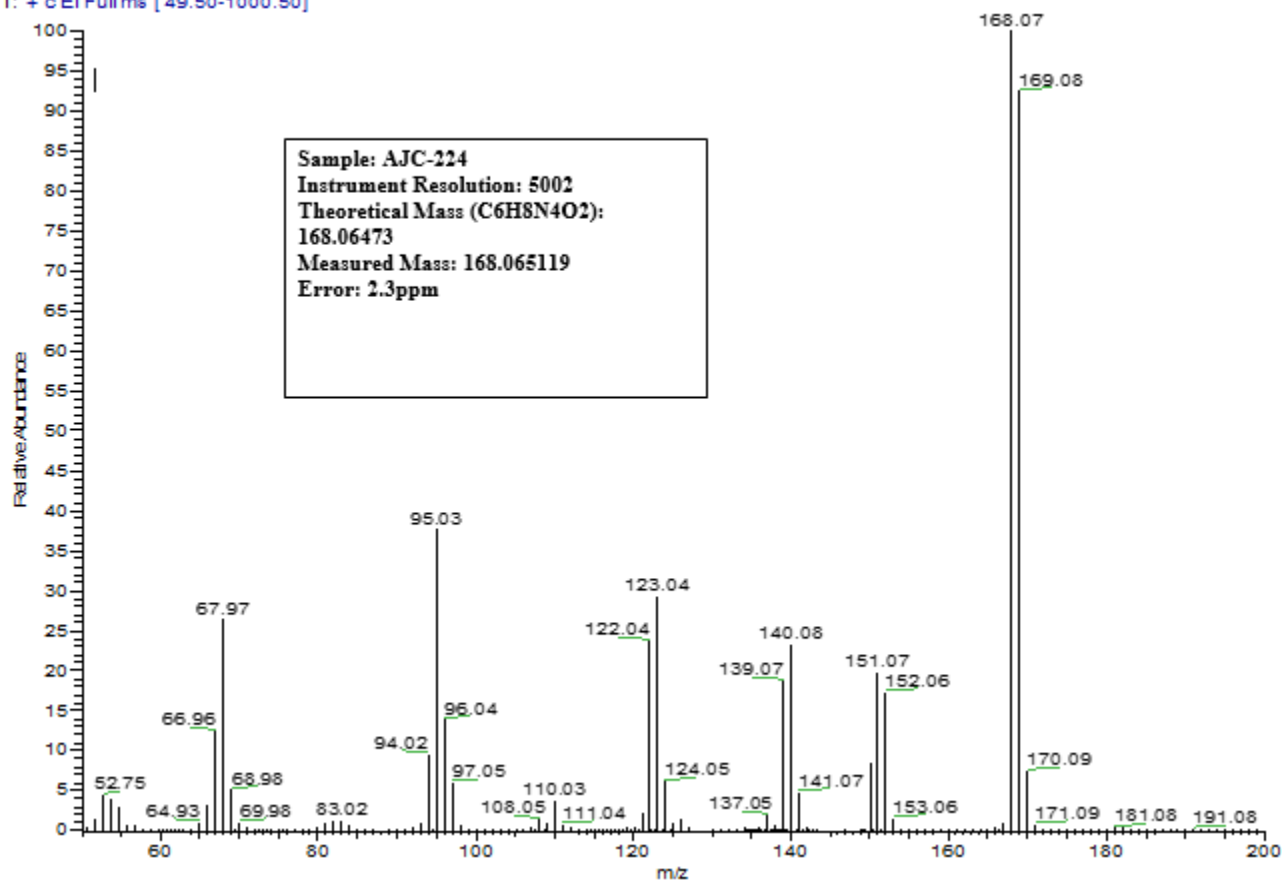


A343: ^1H NMR spectrum (400 MHz, $\{\text{D}_2\text{O}\}$, 2.0 – 9.0 ppm) of **212** after precipitation from aqueous ethanol.

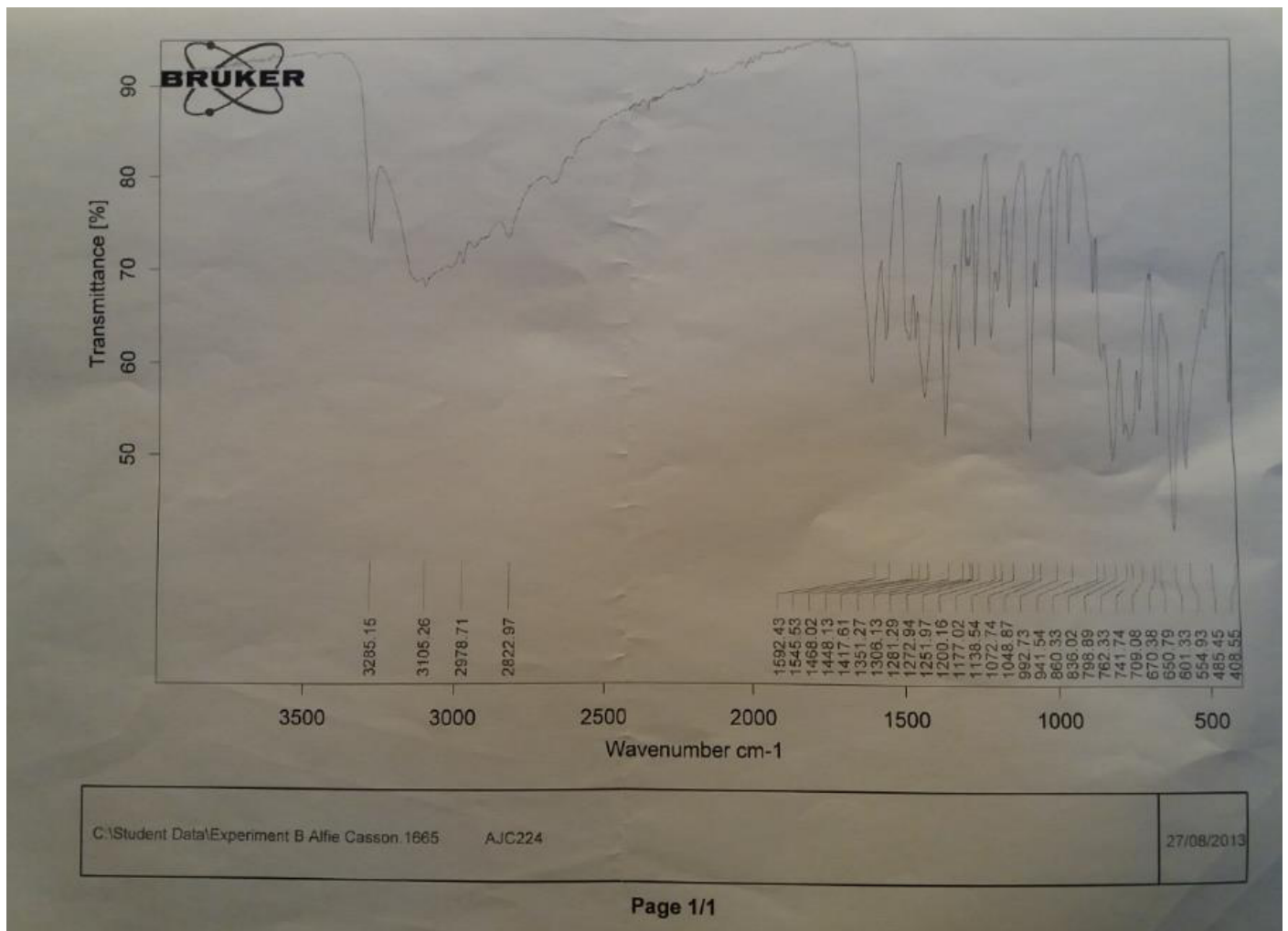


A344: EI+ mass spectrum of 212.

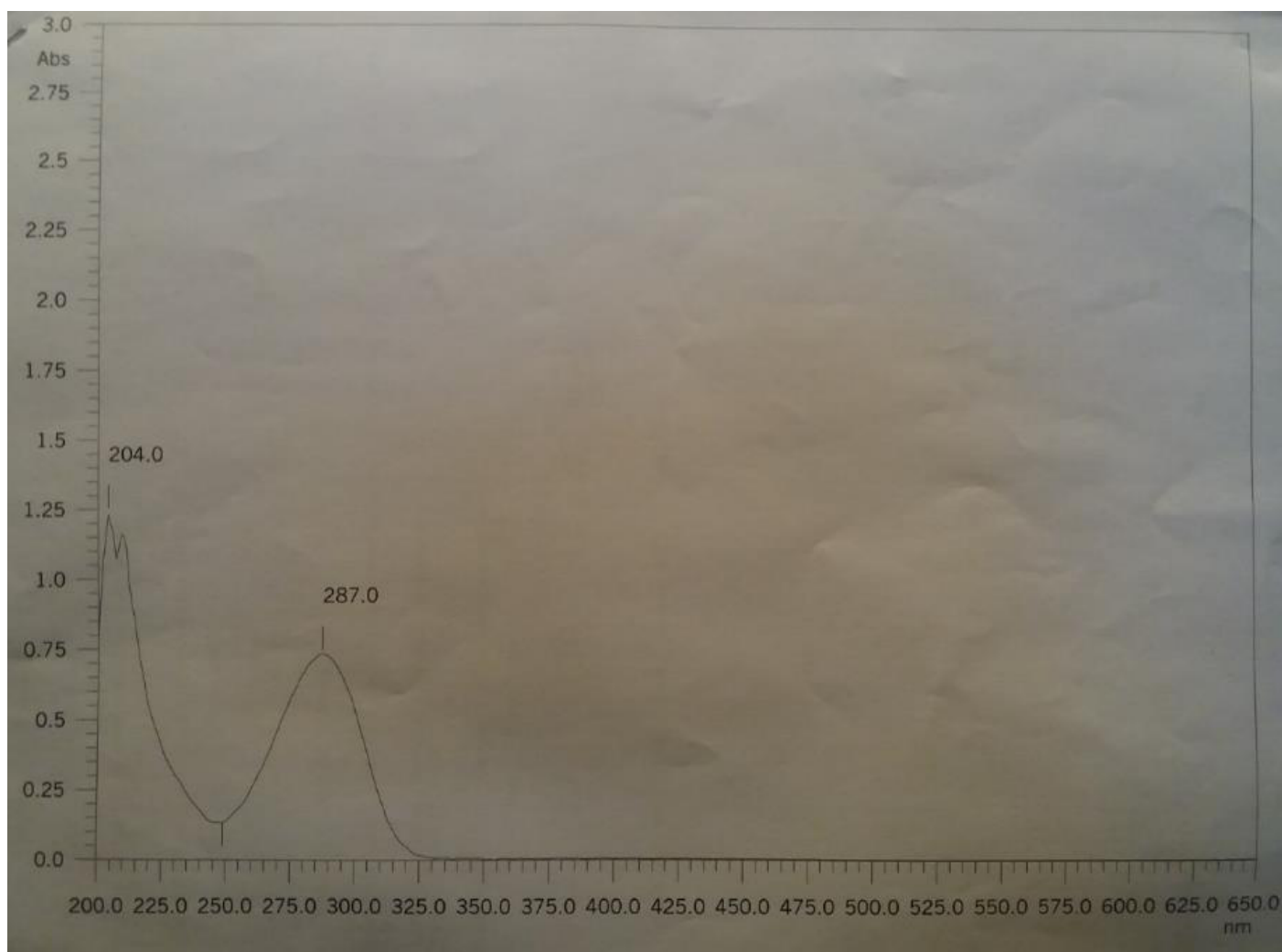
ajc224_e #7 RT: 0.80 AV: 1 NL: 2.78E7
T: + c EI Fullms [49.50-1000.50]



A345: IR spectrum of 212.

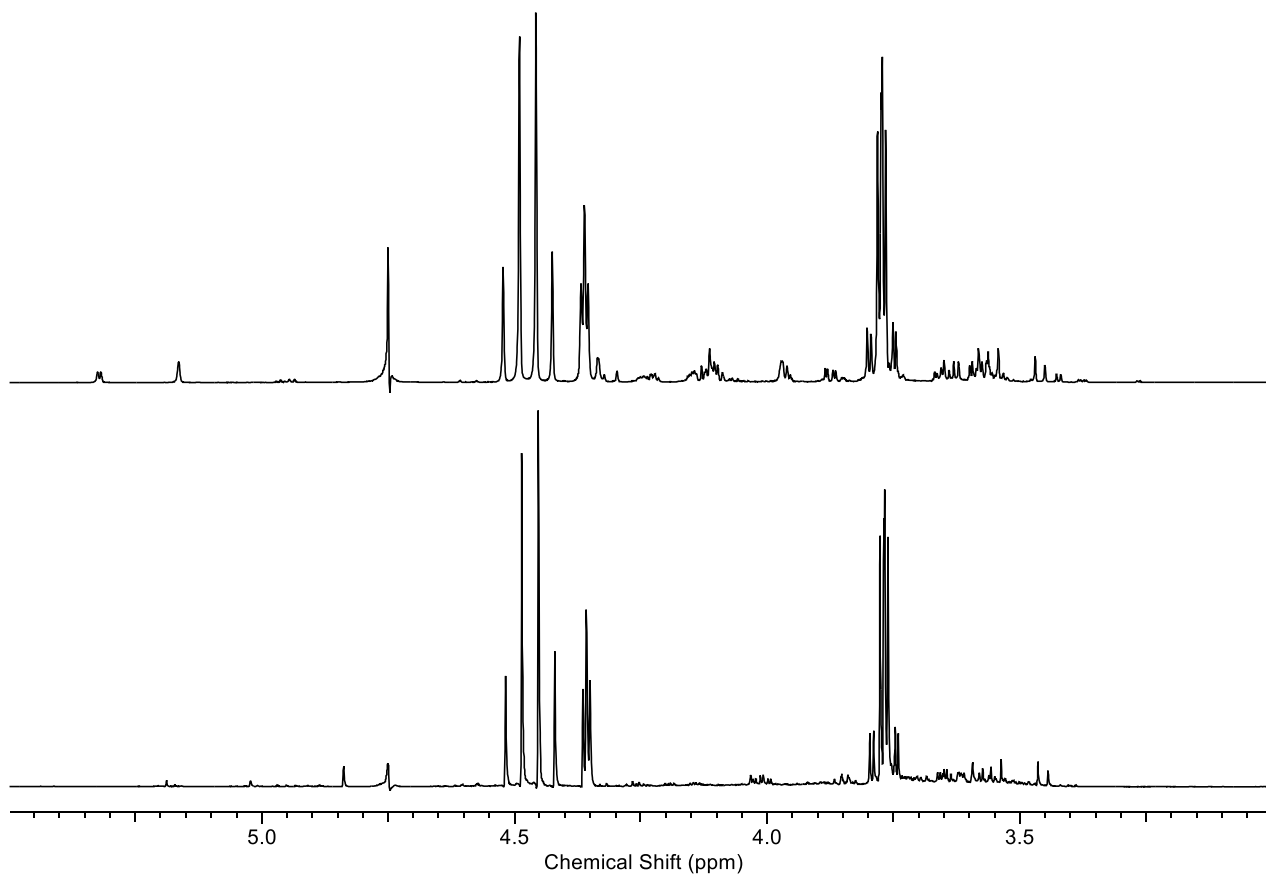


A346: UV-vis spectrum of **212**.



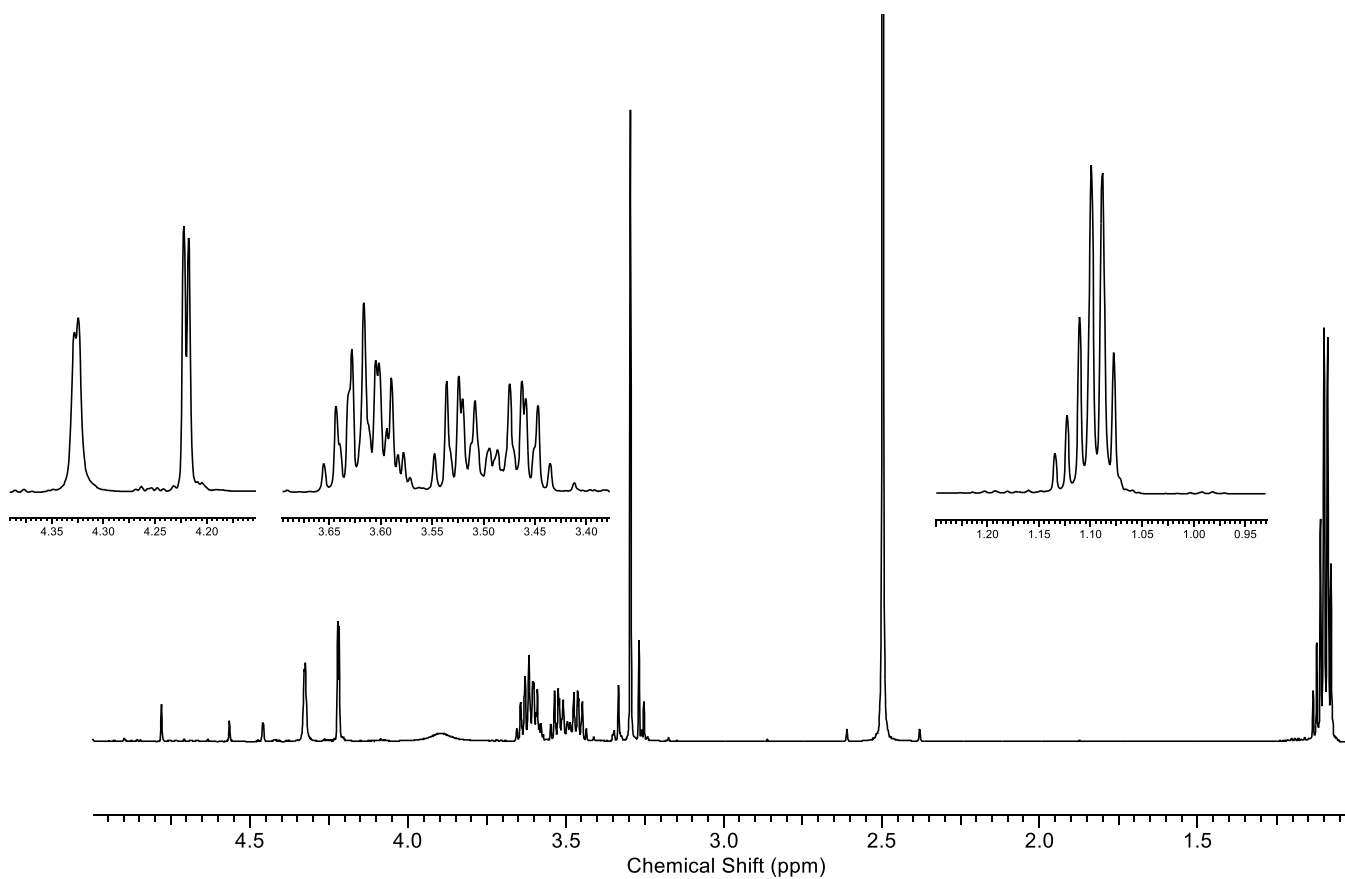
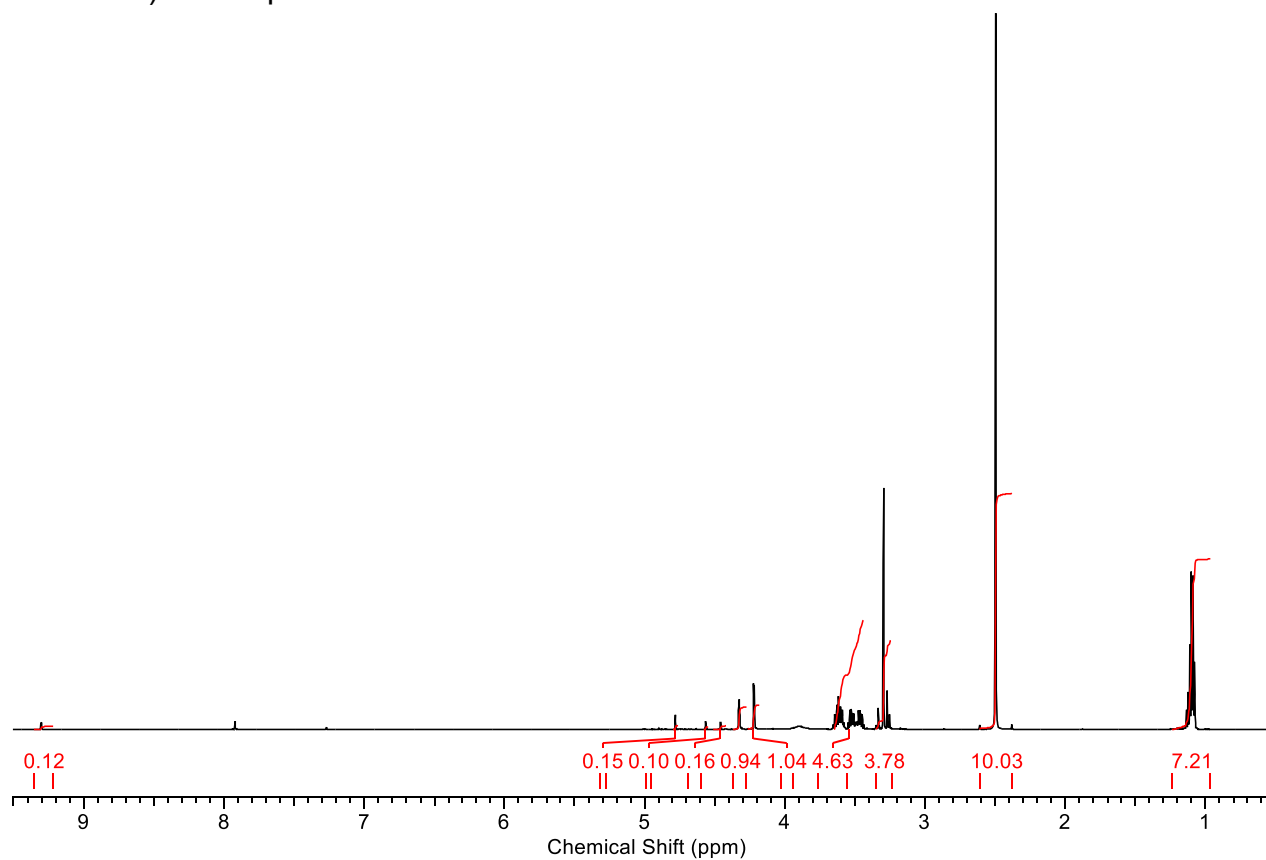
Erythrulose (215)

A347: ^1H NMR spectra (600 MHz, $\{\text{H}_2\text{O}/\text{D}_2\text{O}, 9:1\}$, 3.0 – 5.5 ppm) of **Top, 215** isolated from the reaction of glycolaldehyde (500mM) in pH 7 phosphate buffer (1M) at 60 °C for 16 h and **Bottom,** commercially obtained **215**.

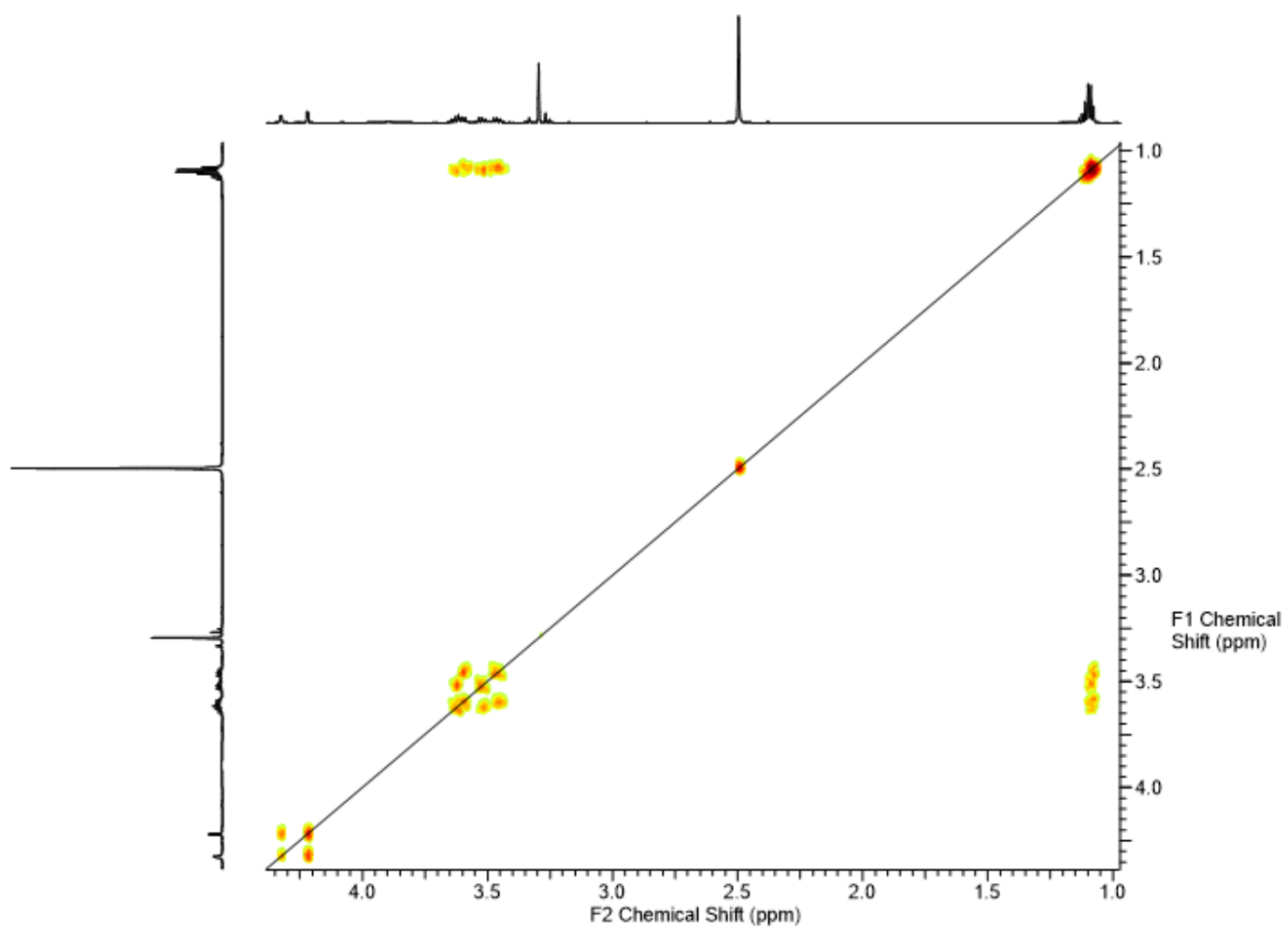


2,2-diethoxyacetaldehyde (217)

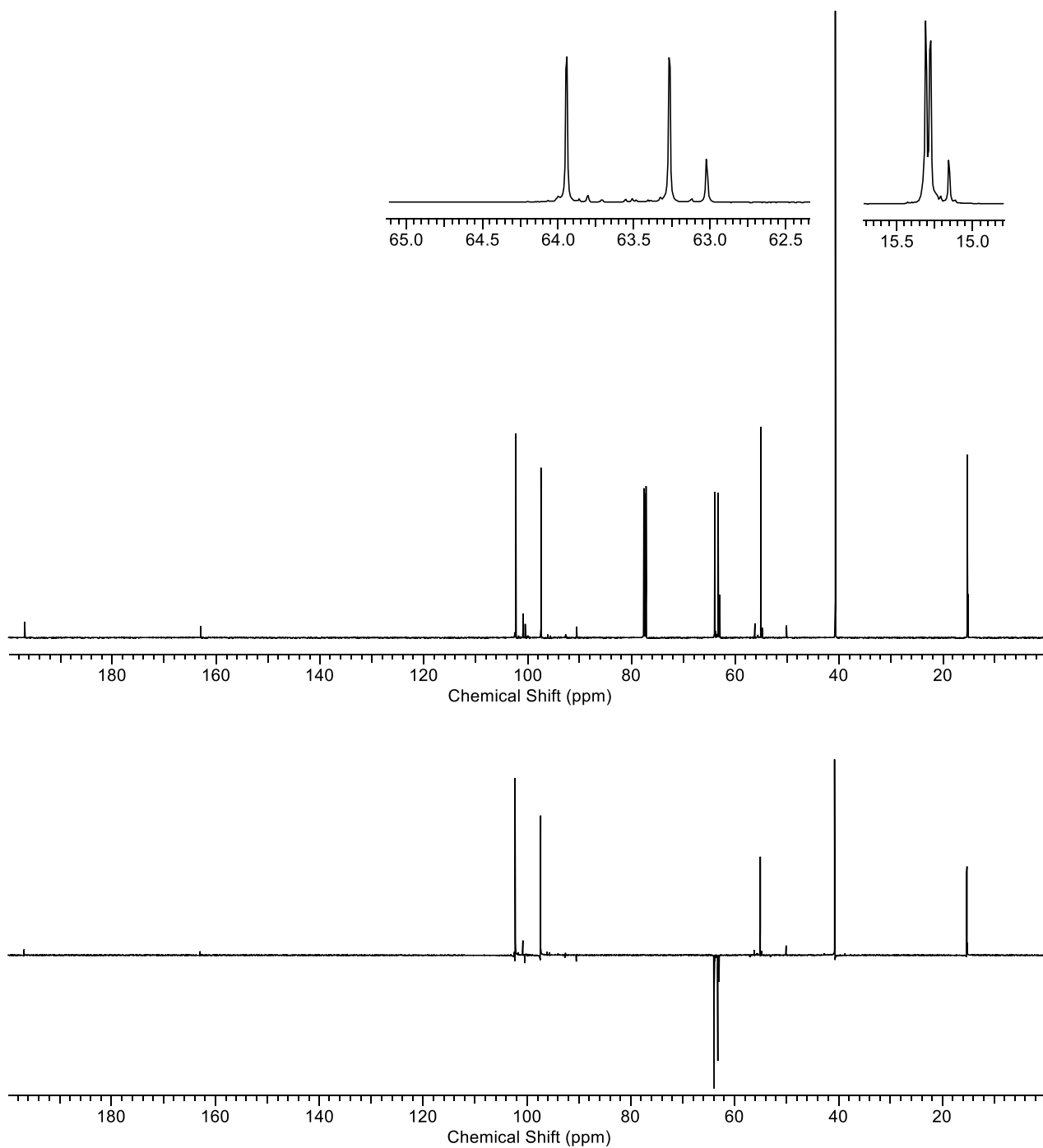
A348: ^1H NMR spectrum (600 MHz, $\{\text{CDCl}_3\}$, 0.5 – 9.5 ppm) of **217** (isolated as its 1-methyl-hemi-acetal in DMSO) with expansions below.



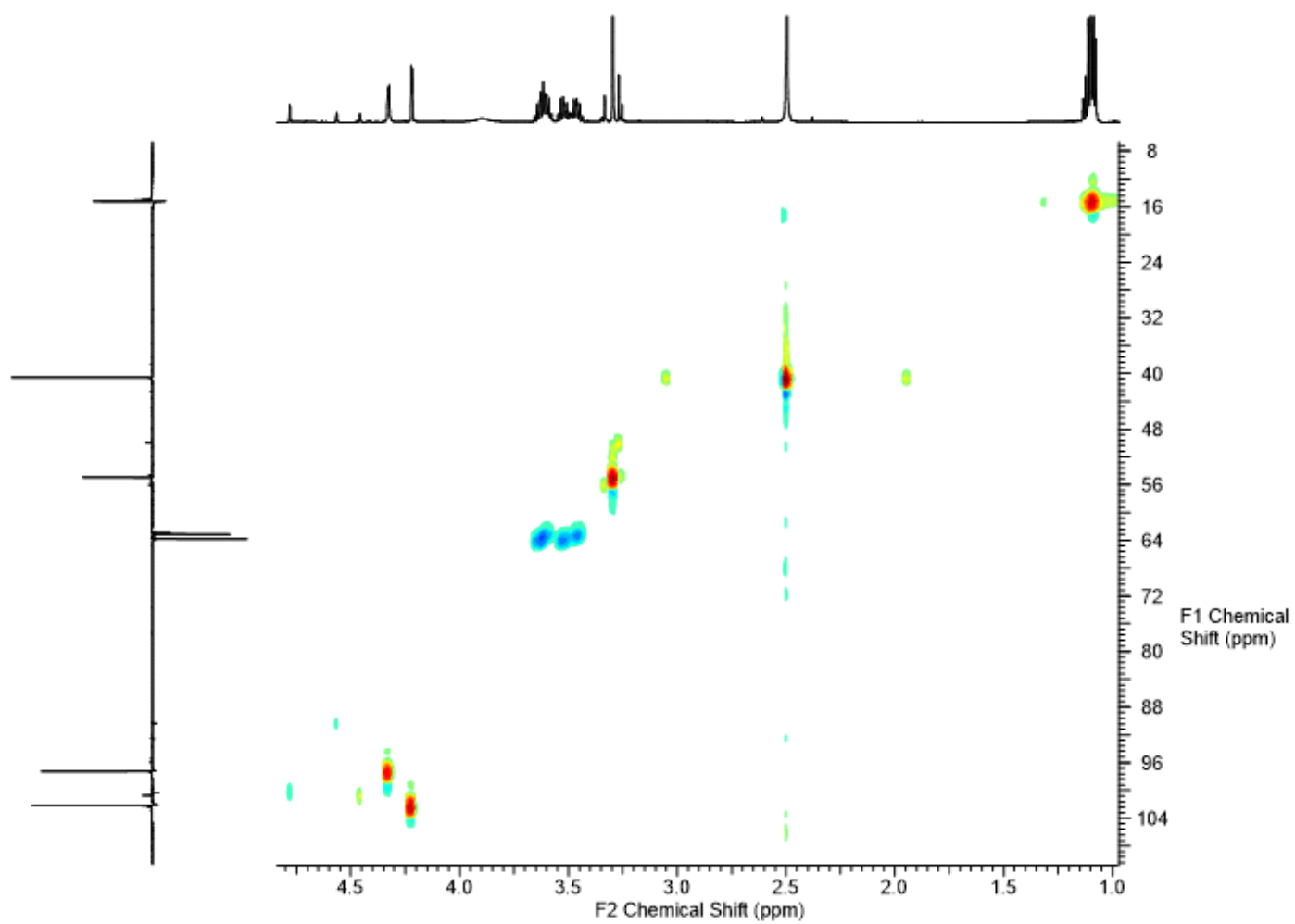
A349: ^1H - ^1H COSY NMR spectrum (600 MHz, $\{\text{CDCl}_3\}$) of **217** (isolated as its 1-methyl-hemi-acetal in DMSO).



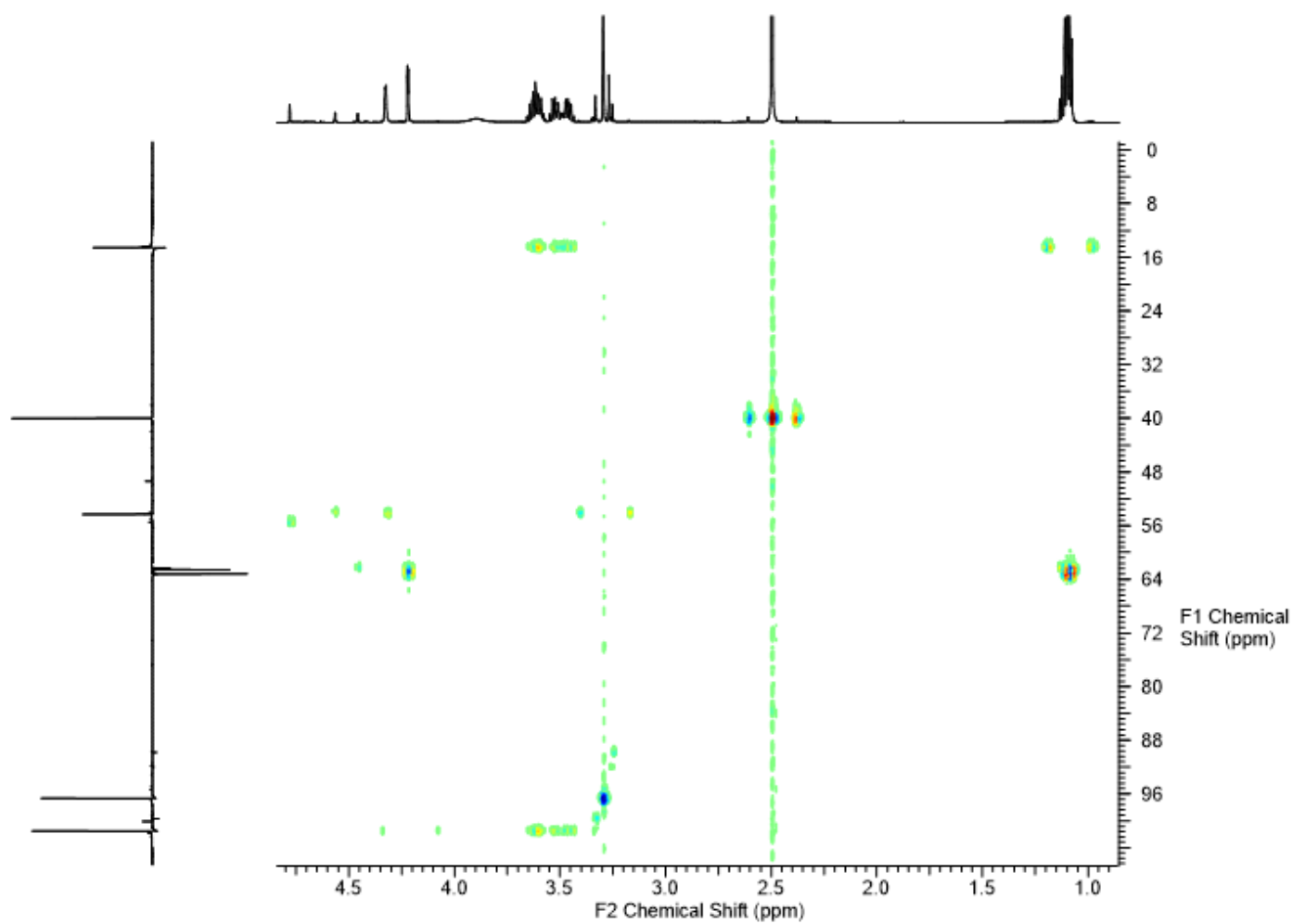
A350: ^{13}C NMR spectrum (151 MHz, $\{\text{CDCl}_3\}$, 0 – 200 ppm) of **217** (isolated as its 1-methyl-hemi-acetal in DMSO) with expansions overlaid and DEPT135 below.



A351: ^1H - ^{13}C HSQC NMR spectrum (600 MHz, $\{\text{CDCl}_3\}$) of **217** (isolated as its 1-methyl-hemi-acetal in DMSO).

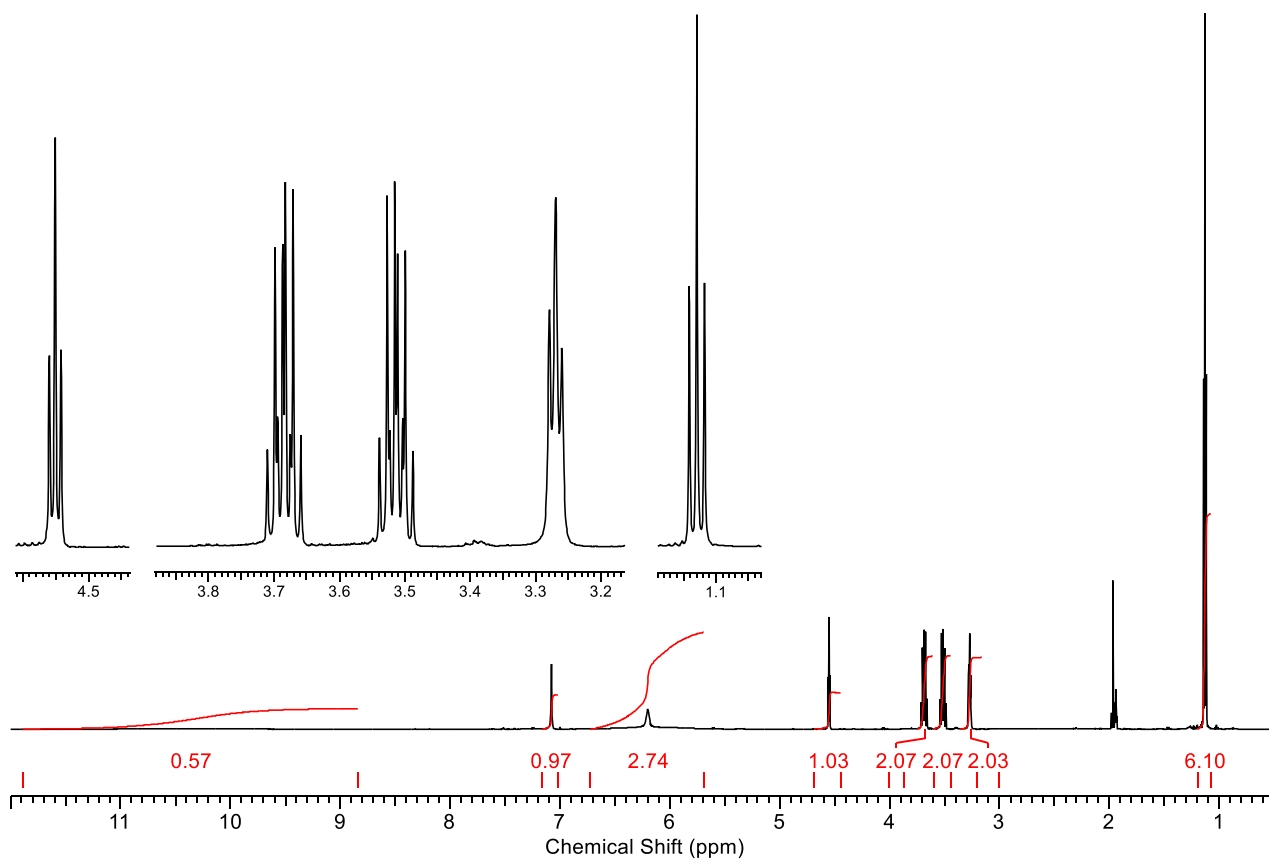


A352: ^1H - ^{13}C HMBC NMR spectrum (600 MHz, $\{\text{CDCl}_3\}$) of **217** (isolated as its 1-methyl-hemi-acetal in DMSO).

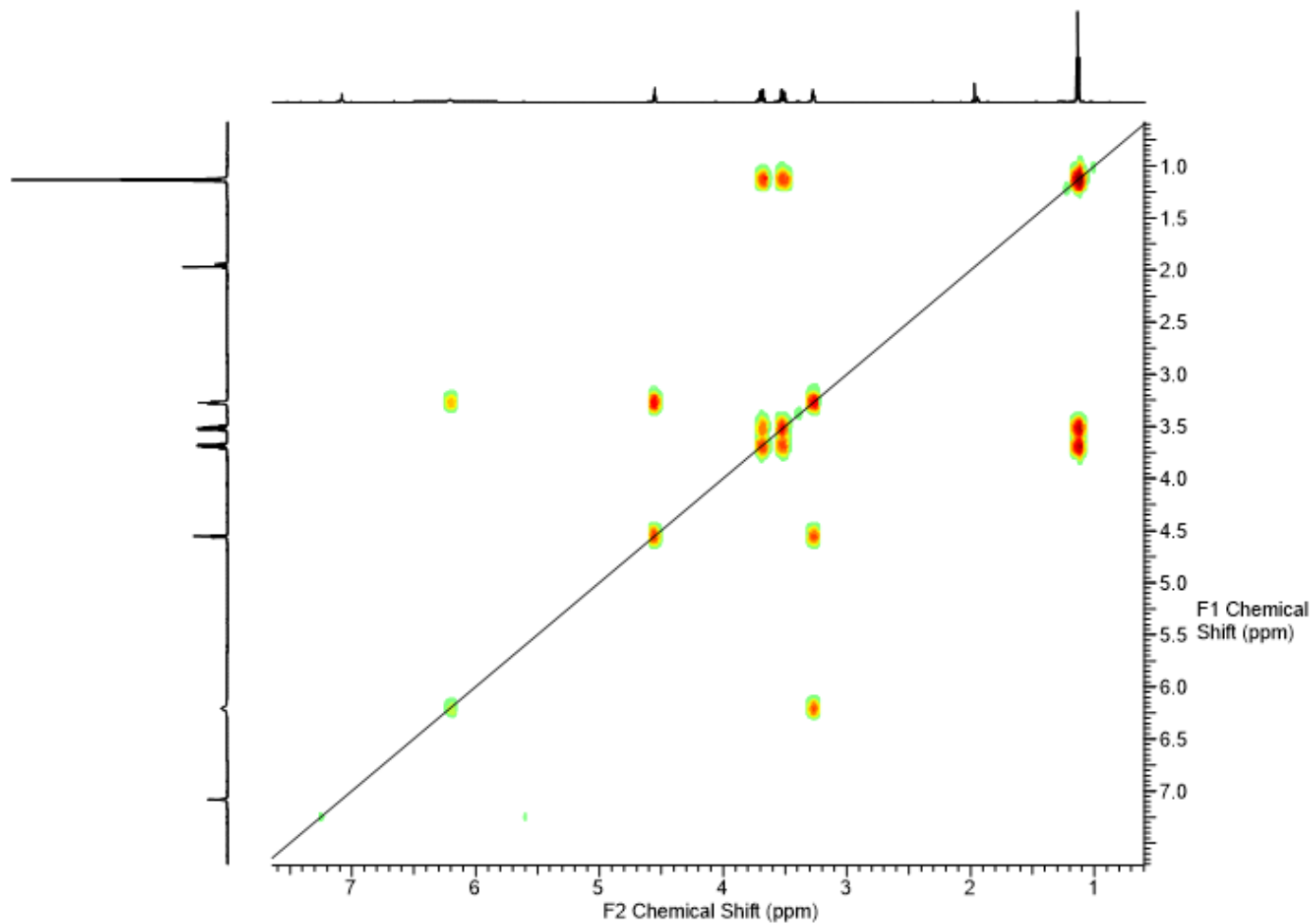


4-((2,2-diethoxyethyl)amino)-1H-imidazole-5-carboxamide (218)

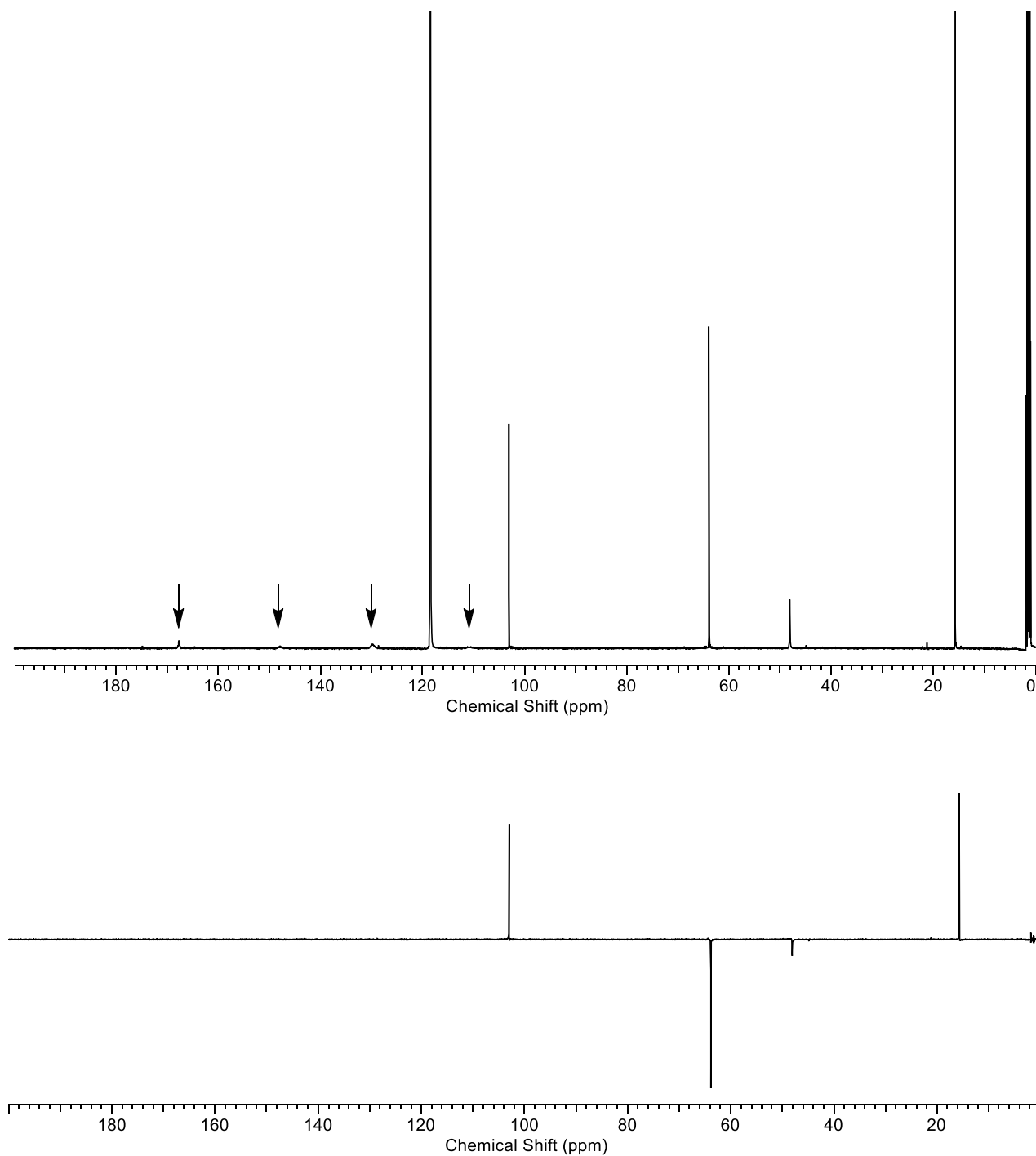
A353: ^1H NMR spectrum (600 MHz, $\{\text{MeCN-d}_3\}$, 0.5 – 12.0 ppm) of **218** with expansions overlaid.



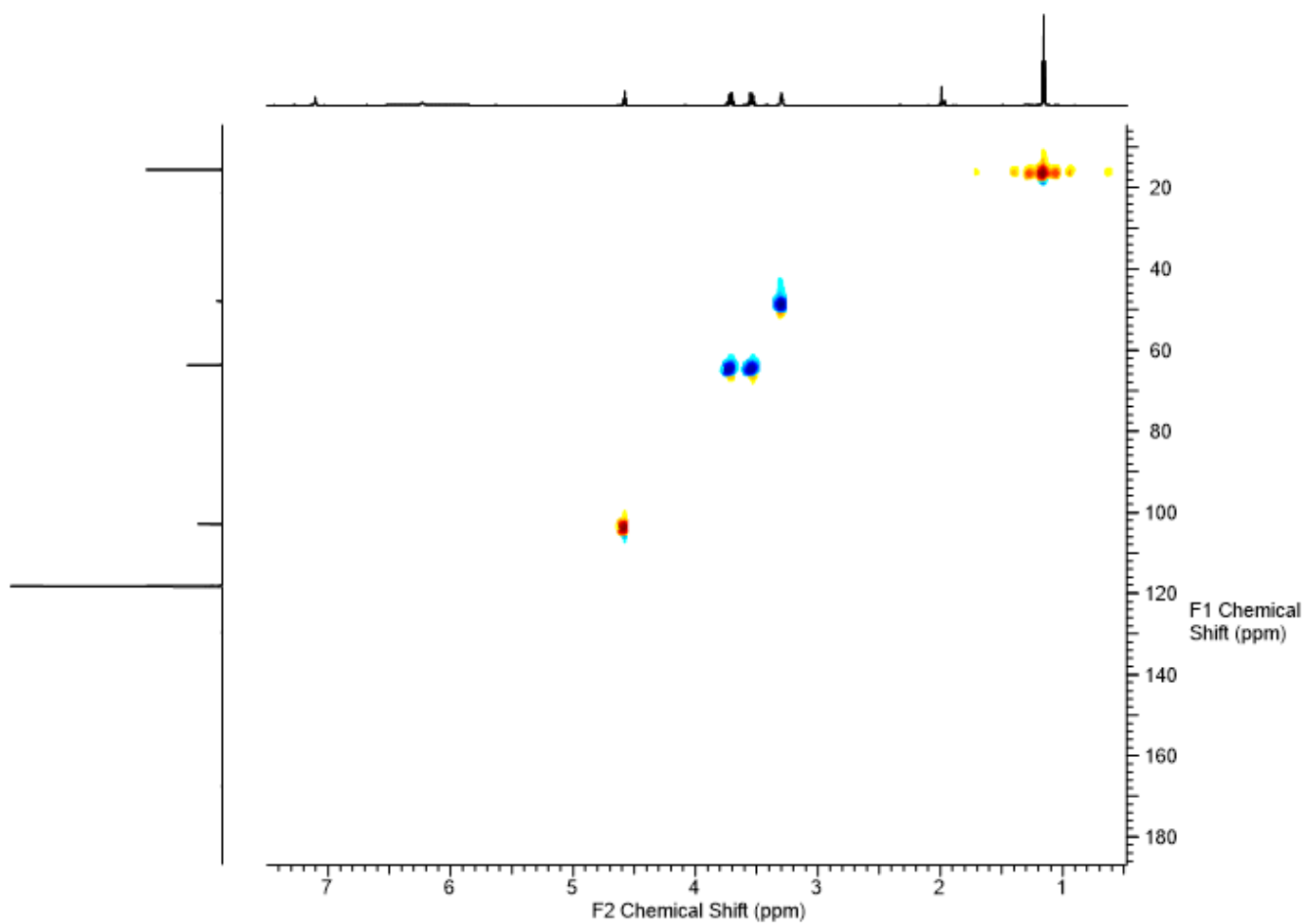
A354: ^1H - ^1H COSY NMR spectrum (600 MHz, {MeCN-d₃}) of **218**.



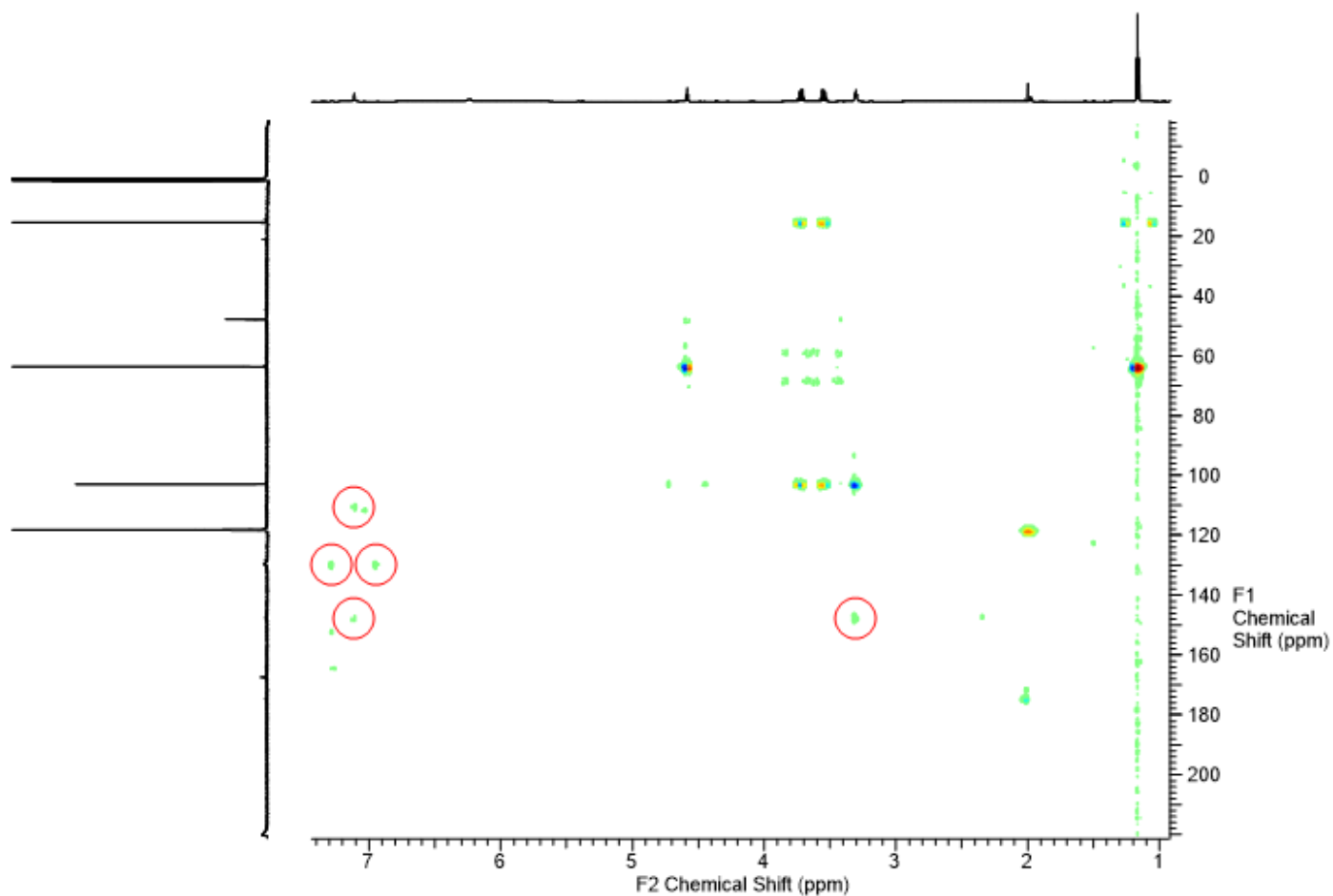
A355: ^{13}C NMR spectrum (151 MHz, $\{\text{CDCl}_3\}$, 0 – 200 ppm) of **218** with expansions overlaid and DEPT135 spectrum below. The highly broadened imidazolyl signals are indicated with arrows.



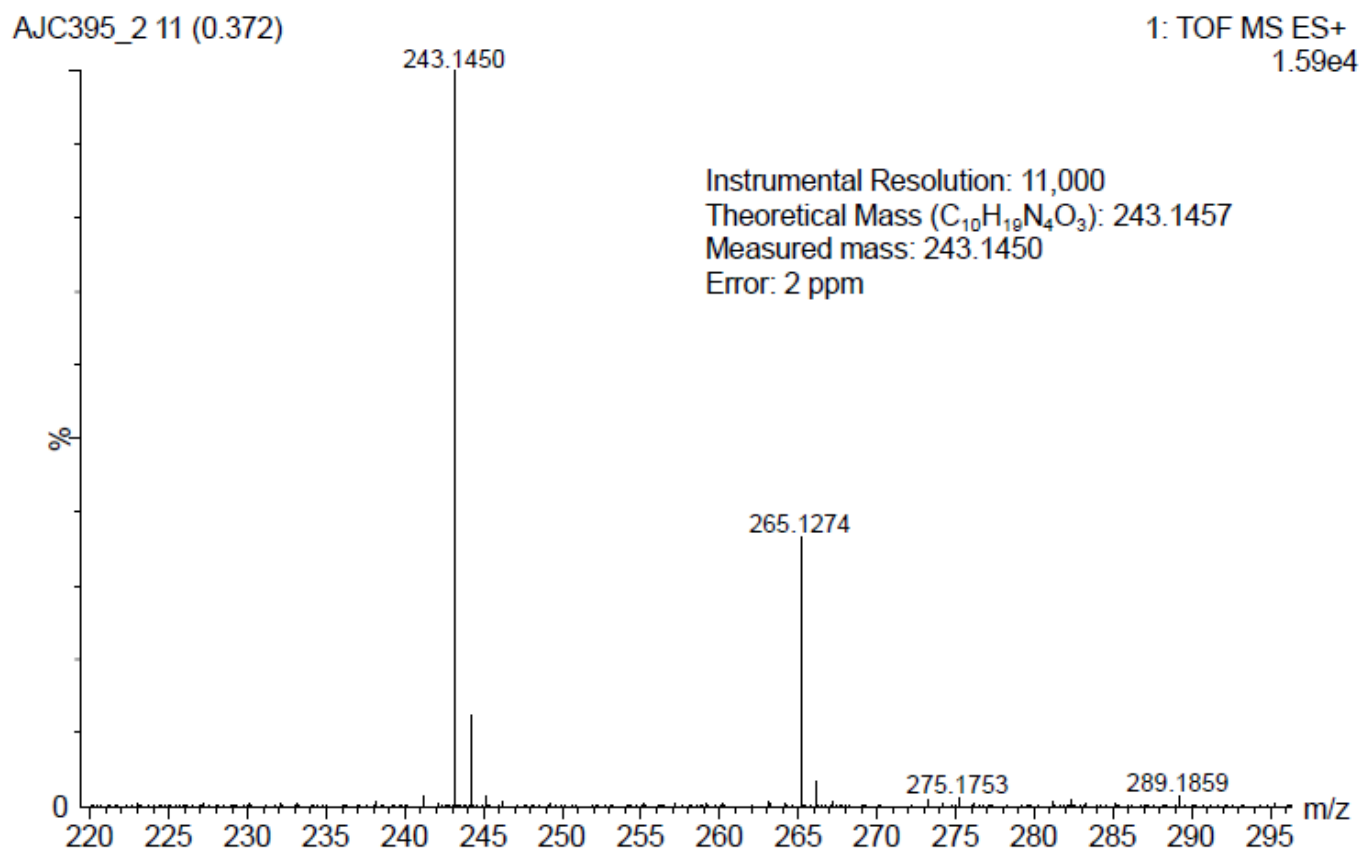
A356: ^1H - ^{13}C HSQC NMR spectrum (600 MHz, {MeCN-d₃}) of **218**.



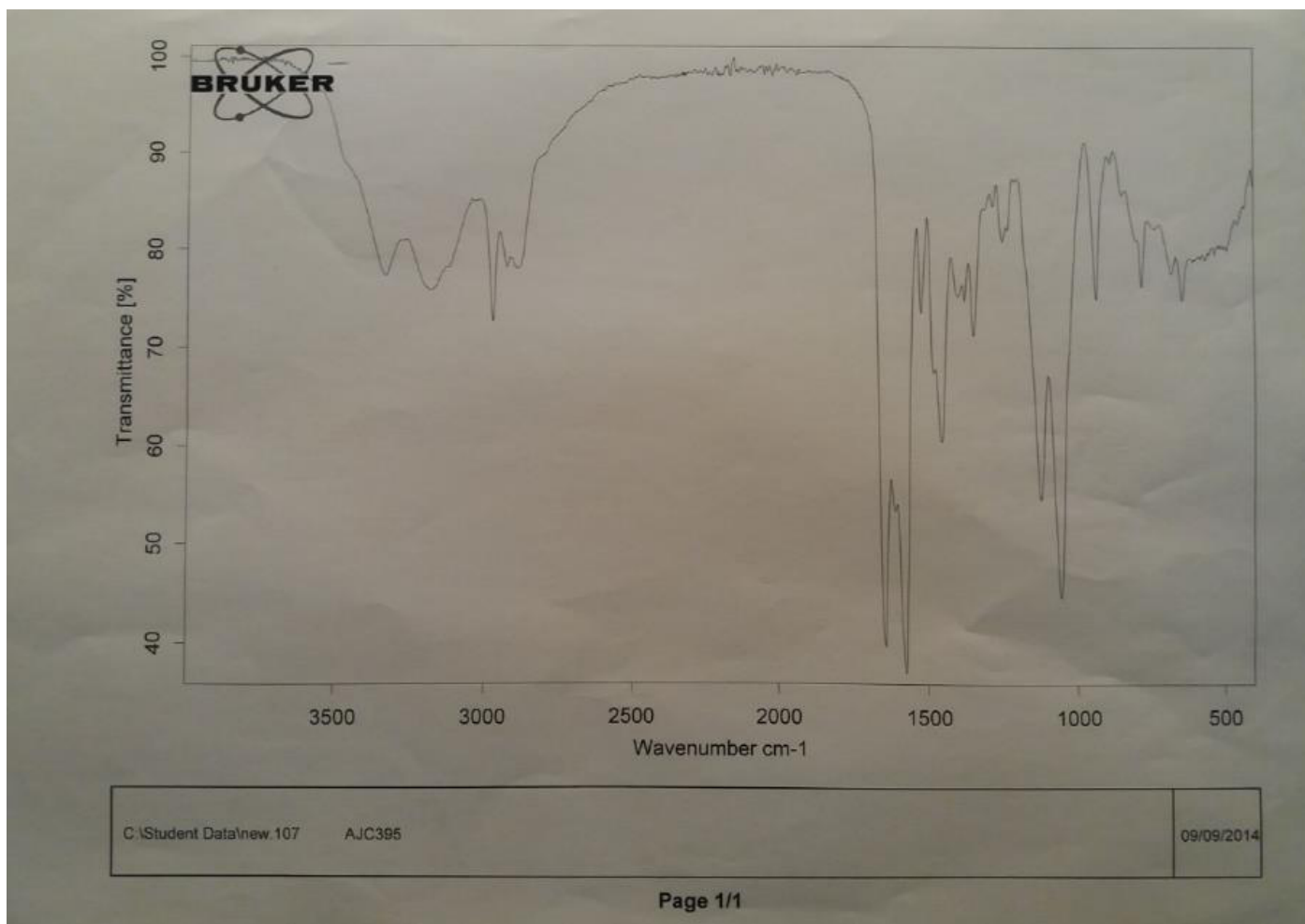
A357: ^1H - ^{13}C HMBC NMR spectrum (600 MHz, {MeCN- d_3 }) of **218**. The correlations used to confirm that the broadened peaks in the ^{13}C spectrum were due to genuine signals have been circled.



A358: ESI+ mass spectrum of **218**.



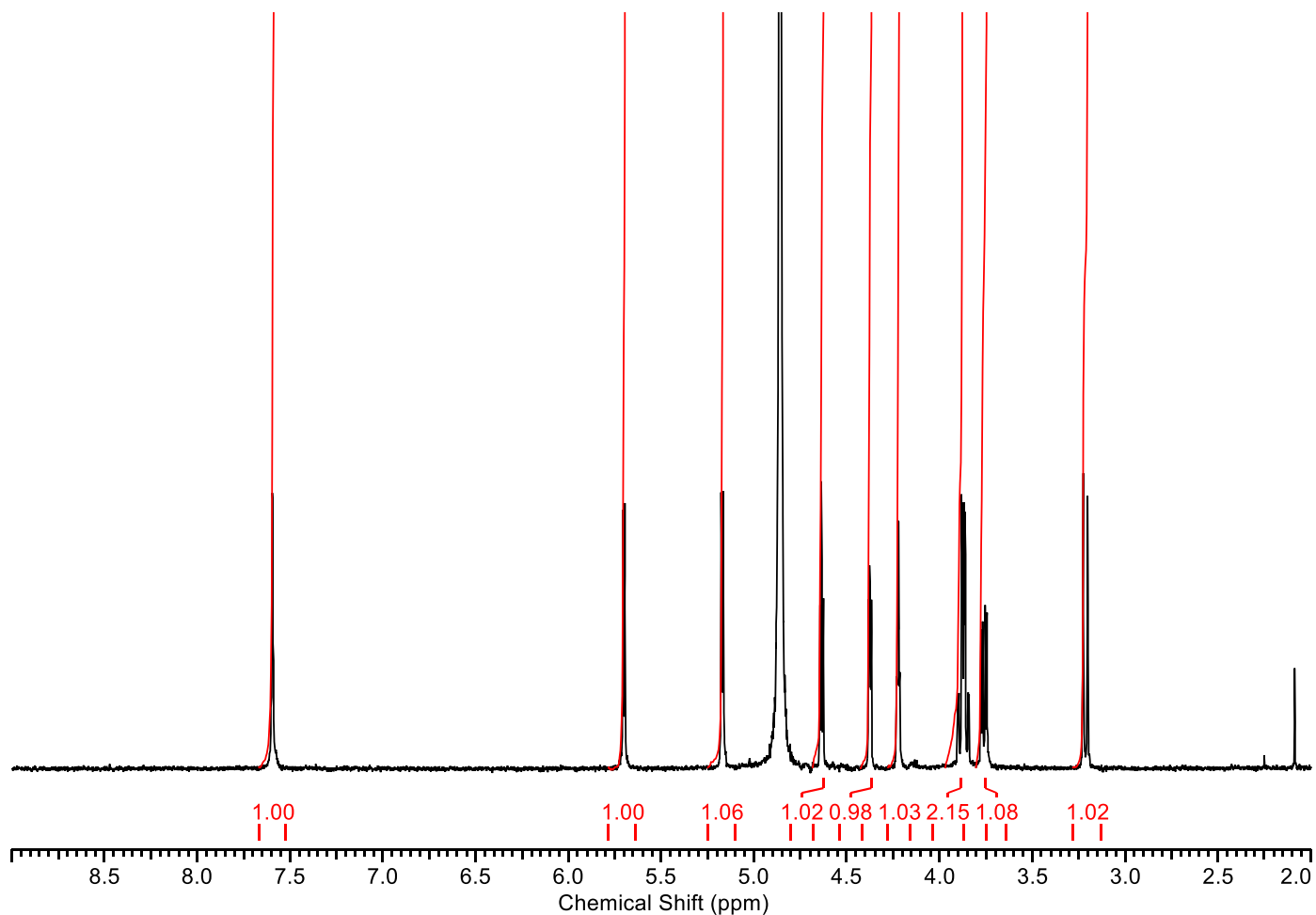
A359: IR spectrum of 218.

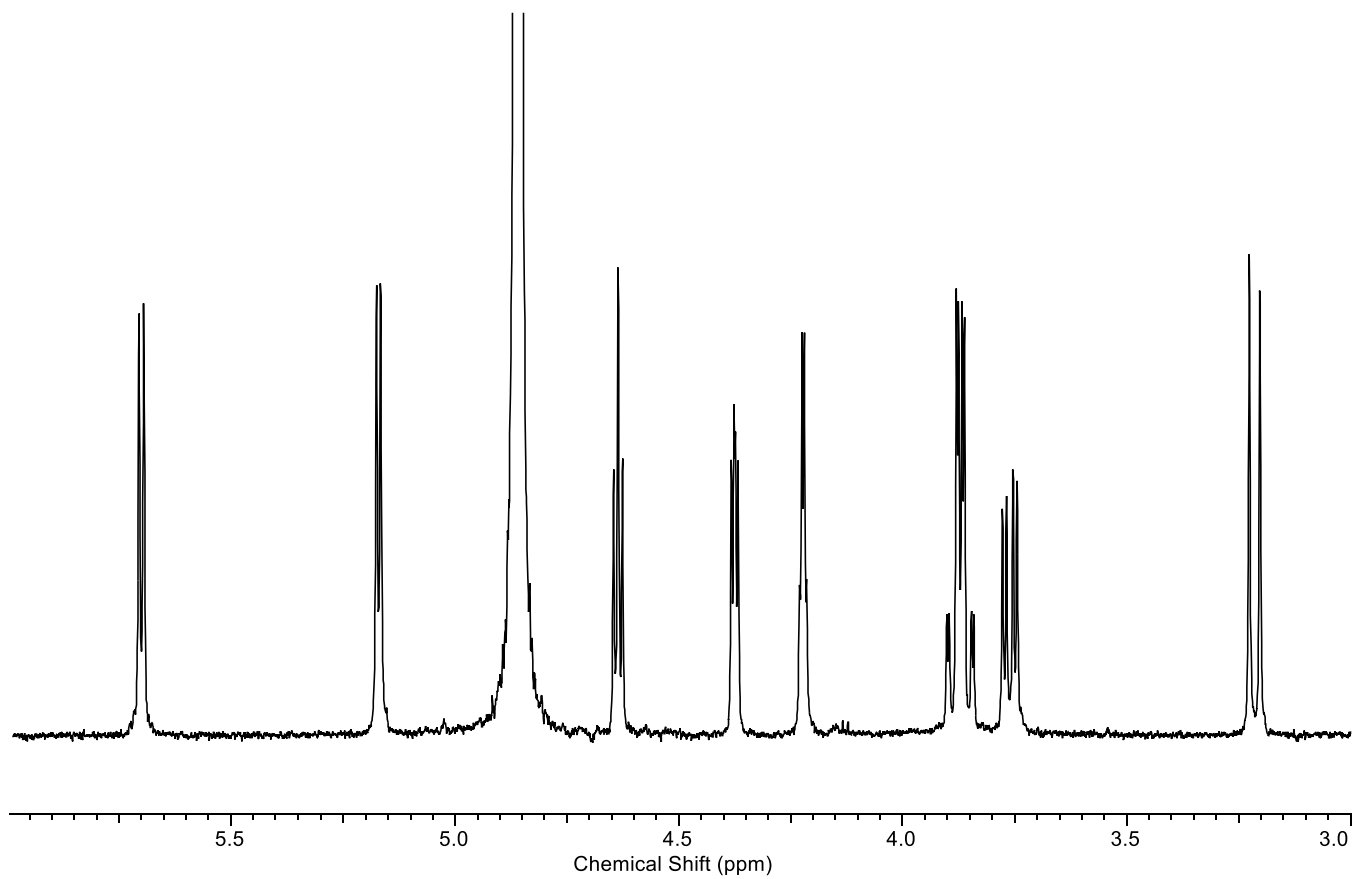


N1-(β-D-ribofuranosyl)-(8R)-azepinomyacin (214A)

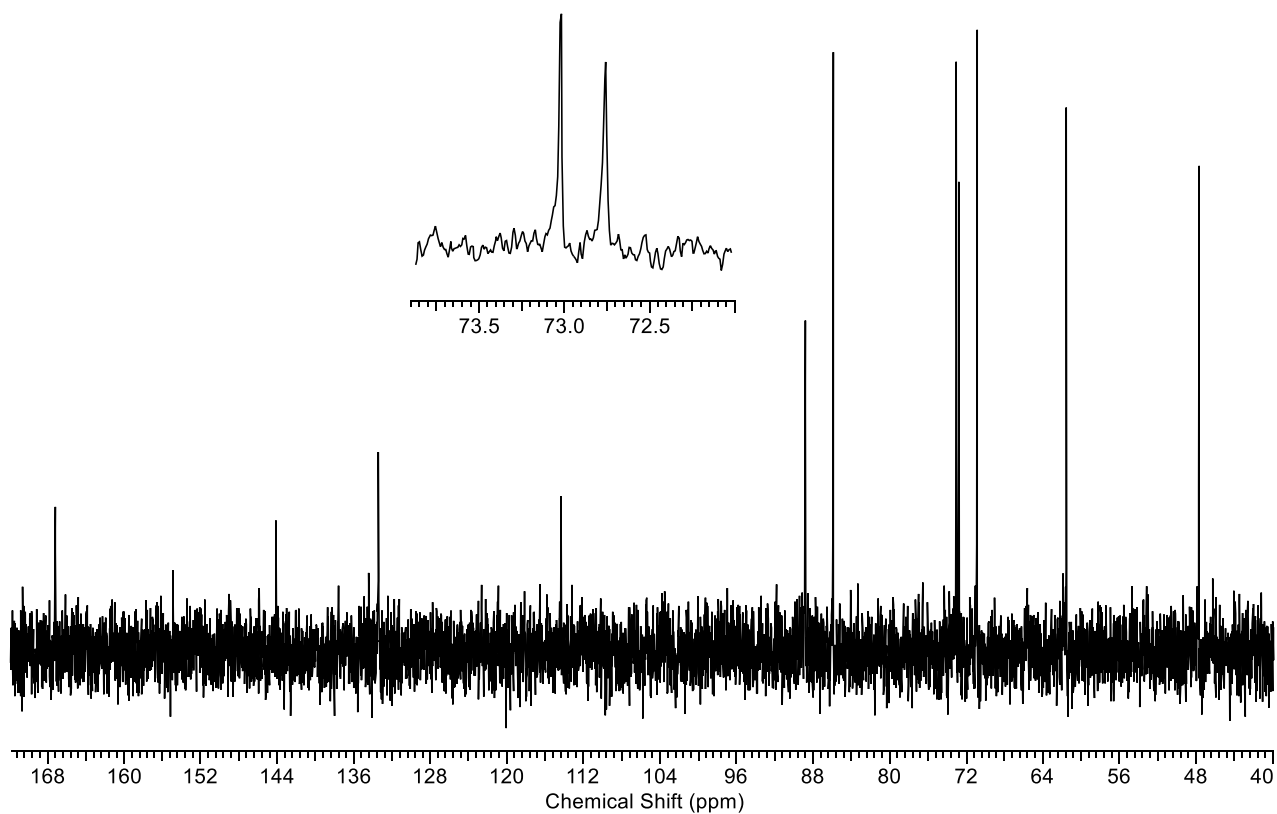
and N1-(β-D-ribofuranosyl)-(8S)-azepinomyacin (214B)

A360: ¹H NMR spectrum (600 MHz, {D₂O}, 2.0 – 9.0 ppm) of **214A** with expansion below.





A361: ^{13}C NMR spectrum (151 MHz, $\{\text{D}_2\text{O}\}$, 40 – 170 ppm) of **214A** with expansion overlaid.



A362: ESI+ mass spectrum of 214A.

Elemental Composition Report

Page 1

Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

157 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass)

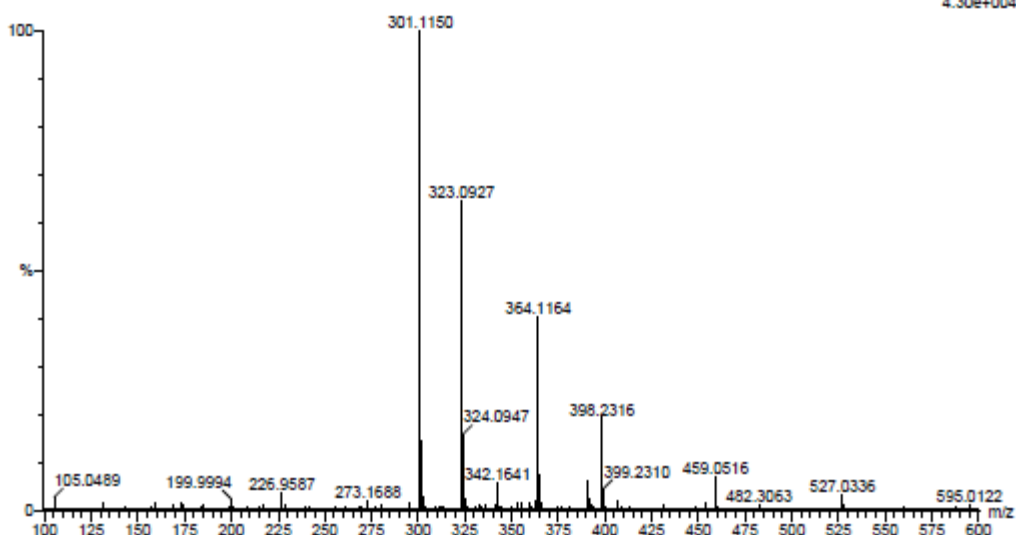
Elements Used:

C: 0-20 H: 0-30 N: 0-5 O: 0-10

AJC437A

AJC437A_2 17 (0.598)

1: TOF MS ES+
4.30e+004



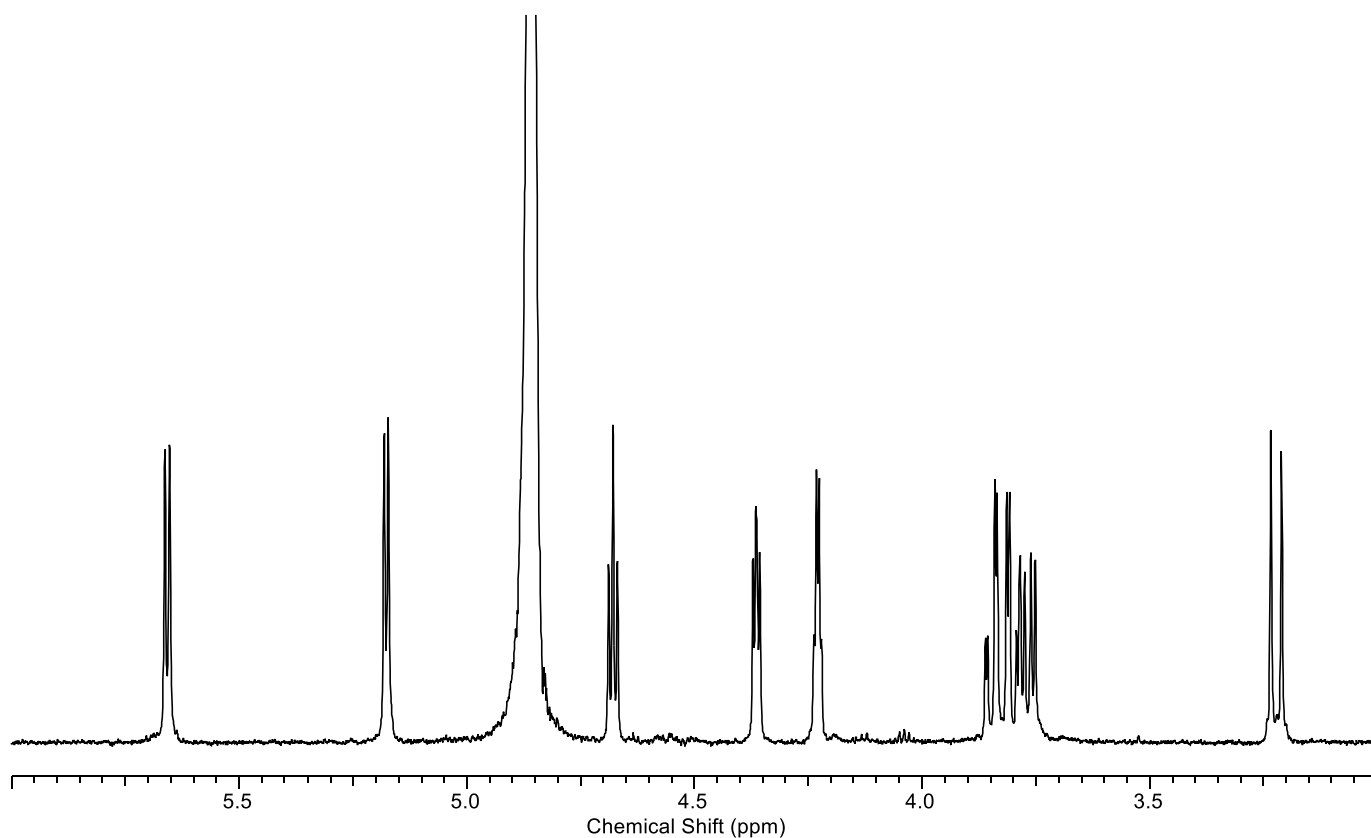
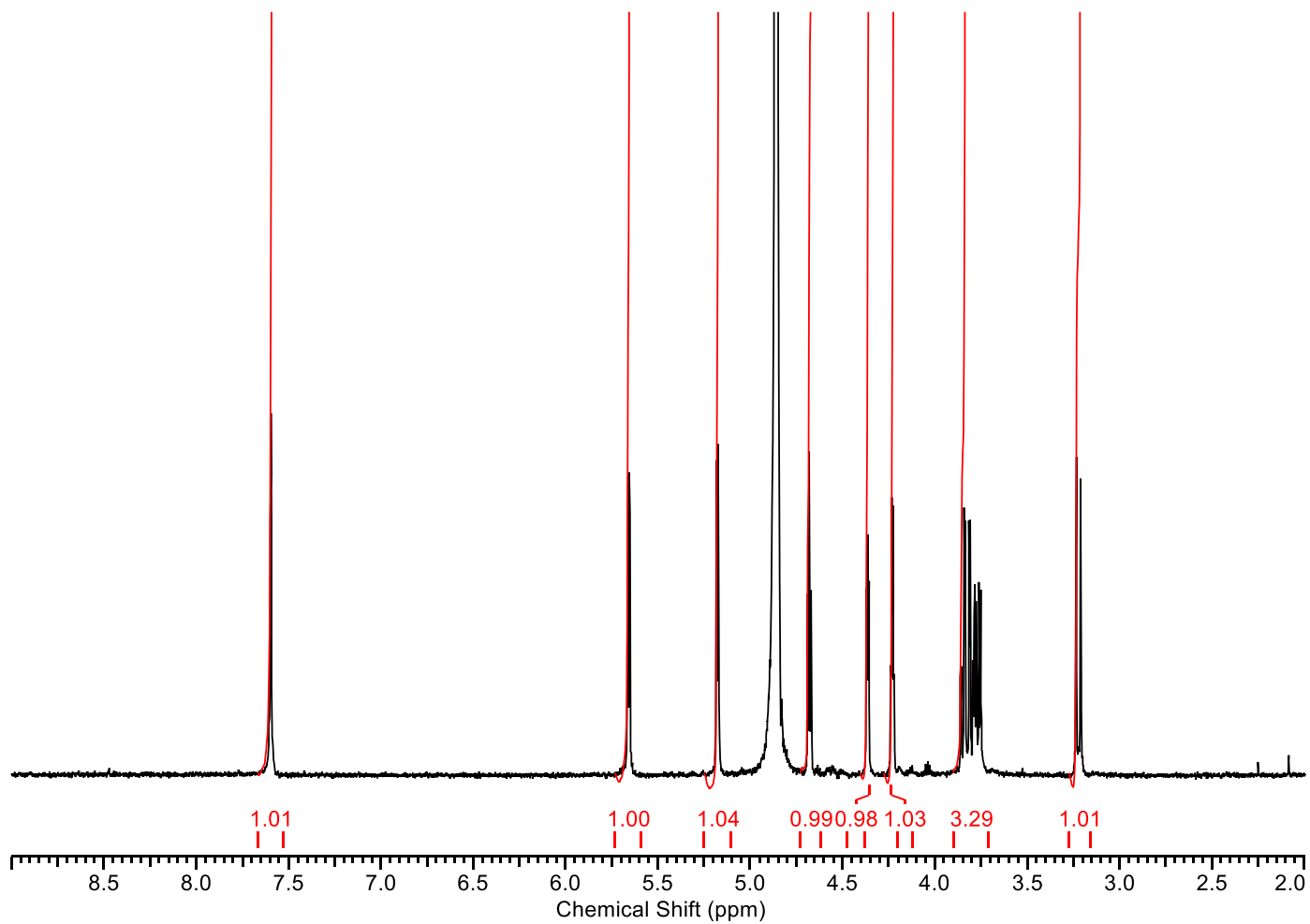
Minimum:

Maximum: 5.0 5.0 -1.5

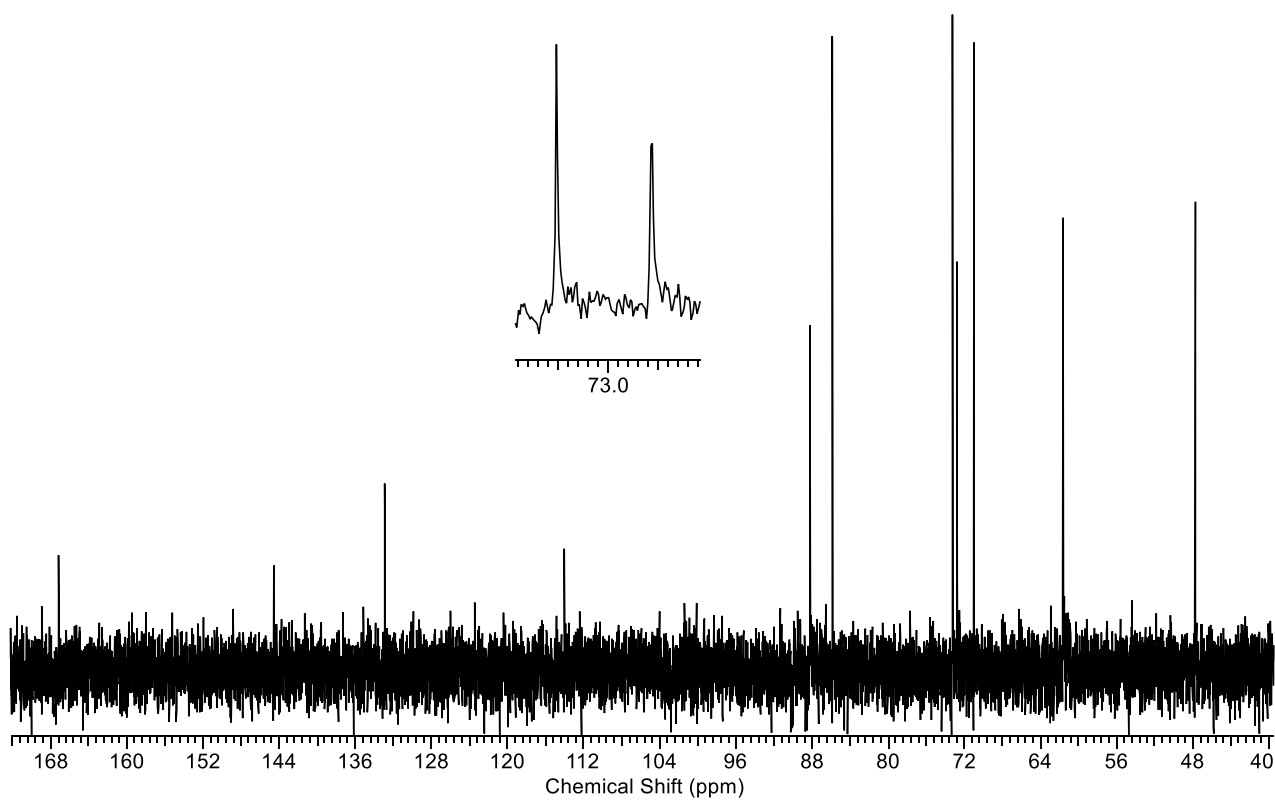
50.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	Formula
301.1150	301.1148	0.2	0.7	5.5	28.2	C11 H17 N4 O6

A363: ^1H NMR spectrum (600 MHz, $\{\text{D}_2\text{O}\}$, 2.0 – 9.0 ppm) of **214B** with expansion below.



A364: ^{13}C NMR spectrum (151 MHz, $\{\text{D}_2\text{O}\}$, 40 – 170 ppm) of **214B** with expansion overlaid.



A365: ESI+ mass spectrum of 214B.

Elemental Composition Report

Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

157 formula(e) evaluated with 2 results within limits (all results (up to 1000) for each mass)

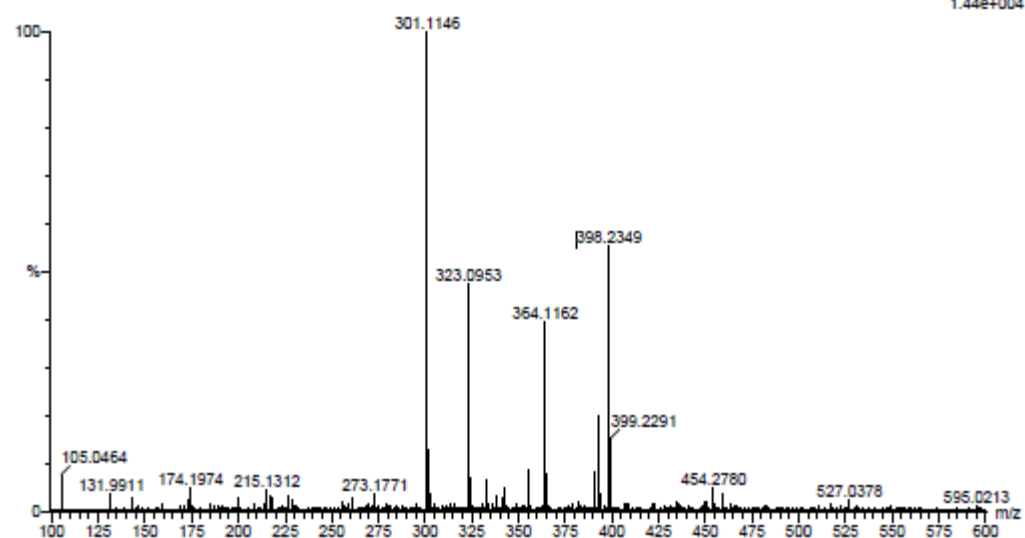
Elements Used:

C: 0-20 H: 0-30 N: 0-5 O: 0-10

AJC437B

AJC437B_2 16 (0.541)

1: TOF MS ES+
1.44e+004



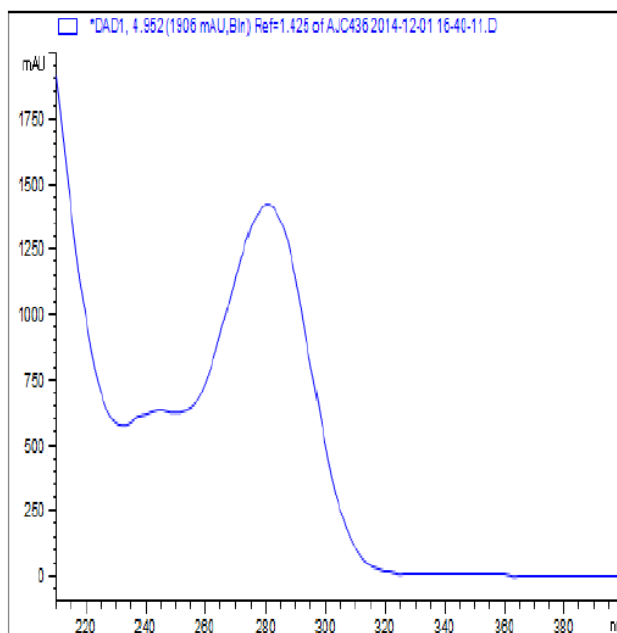
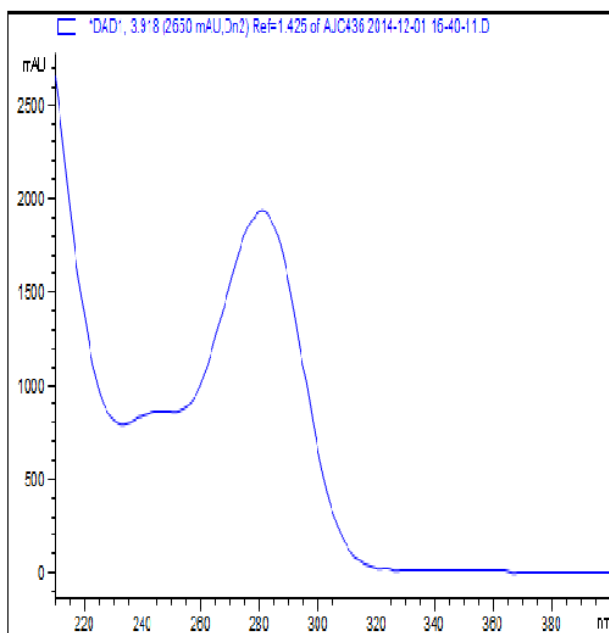
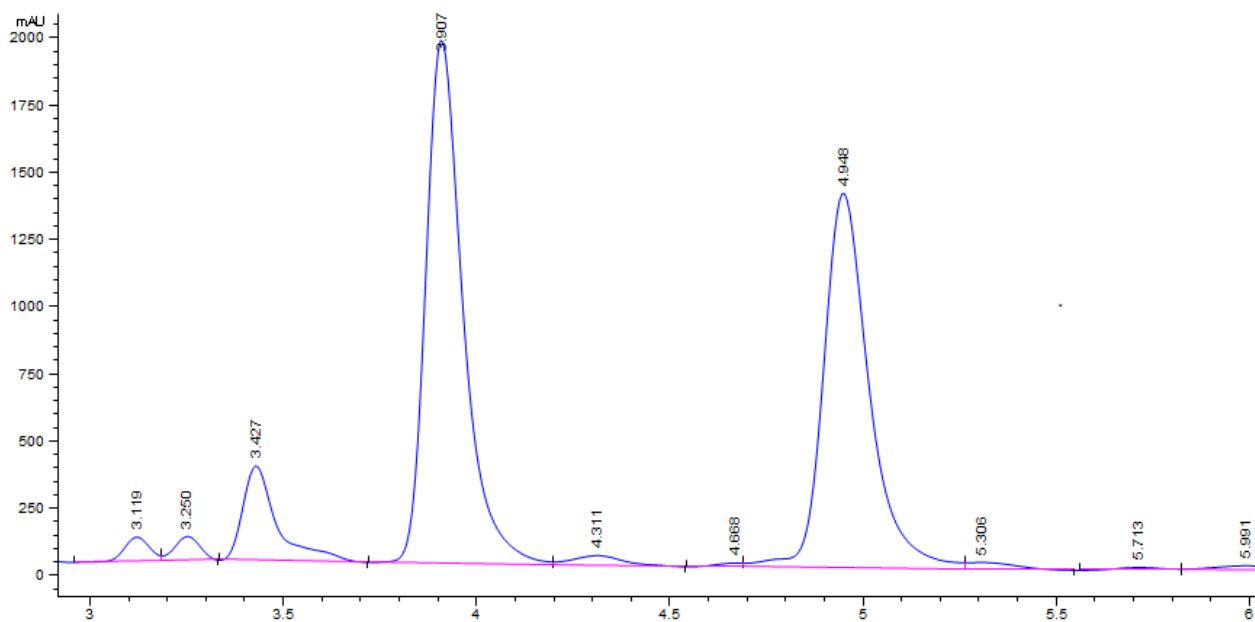
Minimum:

Maximum: 5.0 5.0 -1.5

50.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	Formula
301.1146	301.1148	-0.2	-0.7	5.5	51.5	C11 H17 N4 O6
	301.1135	1.1	3.7	0.5	18.9	C10 H21 O10

A366: HPLC trace of crude **214** (Diastereomer **A** = 3.91 min; Diastereomer **B** = 4.95 min) with corresponding UV traces of the major peaks below.



Single crystal X-ray diffraction studies

General information

Single crystal X-ray diffraction data were collected using an *Agilent SuperNova (Dual Source)* single crystal X-ray diffractometer equipped with an Atlas CCD Detector. All data sets collected at 150 K using $\text{CuK}\alpha$ radiation ($\lambda = 1.54184 \text{ \AA}$). The data were acquired and processed using the *CrysAlisPro* program and the datasets were corrected for Lorentz and polarization effects. Structure solution and refinement was accomplished using SHELXS-97 and SHELXL-97, respectively. The crystal structures were solved using direct methods. All non-hydrogen atoms were refined anisotropically. Hydrogen atoms affiliated with oxygen and nitrogen atoms were refined isotropically in positions identified in the difference Fourier map or in geometrically constrained positions, while hydrogen atoms associated with carbon atoms were refined isotropically in geometrically constrained positions. Crystallographic and refinement parameters for all crystal structures are given in the table below.

A367: Crystallographic and refinement parameters for 108, 184, 185, 198 and 212.

compound	212	185	184	108	198
chemical formula	C ₆ H ₈ N ₄ O ₂	(C ₁₁ H ₁₀ N ₆ O ₃) ₂ (H ₂ O)	C ₁₁ H ₁₂ N ₆ O ₄	C ₁₂ H ₃₈ Ca ₃ N ₁₂ O ₂₉ P ₆	(C ₁₃ H ₁₅ N ₆ O ₅) (H ₂ O) _{0.83}
<i>M_r</i> /gmol ⁻¹	168.16	566.52	292.27	1120.60	349.27
crystal system	monoclinic	monoclinic	monoclinic	monoclinic	triclinic
space group	<i>P</i> 2 ₁ / <i>c</i>	<i>P</i> 2 ₁ / <i>n</i>	<i>P</i> 2 ₁ / <i>c</i>	<i>P</i> 2 ₁ / <i>c</i>	<i>P</i> -1
<i>a</i> /Å	6.68948(7)	18.7664(4)	7.6237(2)	10.59420(10)	11.3819(2)
<i>b</i> /Å	14.04389(15)	5.30070(10)	7.24320(10)	22.20680(10)	14.1561(3)
<i>c</i> /Å	7.22458(9)	23.3702(4)	21.8767(4)	18.34810(10)	16.2079(3)
<i>α</i> /°	90	90	90	90	67.126(2)
<i>β</i> /°	93.8750(11)	93.478(2)	1199.08(4)	102.1080(10)	76.398(2)
<i>γ</i> /°	90	90	90	90	74.371(2)
<i>V</i> /Å ³	677.171(13)	2320.47(8)	1199.08(4)	4220.61(5)	2291.97(9)
<i>Z</i>	4	4	4	4	6
<i>ρ</i> _{calc} / mg mm ³	1.649	1.627	1.619	1.727	1.518
<i>F</i> (000)	352.0	1176	608	2304	1094
<i>μ</i> (CuK _α)/mm ⁻¹	1.090	1.154	1.084	6.556	1.044
<i>T</i> /K	150.0(1)	150.0(1)	150.0(1)	150.0(1)	150.0(1)
crystal size/mm	0.28 × 0.26 × 0.15	0.33 × 0.27 × 0.13	0.14 × 0.12 × 0.09	0.22 × 0.18 × 0.13	0.32 × 0.22 × 0.11
index range	-8 ≤ <i>h</i> ≤ 8 -17 ≤ <i>k</i> ≤ 17 -8 ≤ <i>l</i> ≤ 8	-22 ≤ <i>h</i> ≤ 21 -6 ≤ <i>k</i> ≤ 4 -27 ≤ <i>l</i> ≤ 27	-9 ≤ <i>h</i> ≤ 7 -7 ≤ <i>k</i> ≤ 8 -18 ≤ <i>l</i> ≤ 26	-13 ≤ <i>h</i> ≤ 13 -27 ≤ <i>k</i> ≤ 27 -22 ≤ <i>l</i> ≤ 22	-13 ≤ <i>h</i> ≤ 13 -17 ≤ <i>k</i> ≤ 17 -20 ≤ <i>l</i> ≤ 20
collected reflections	9206	8245	4244	72333	47318
unique reflections	1328	4113	2123	8486	9144
<i>R</i> _{int}	0.0198	0.0251	0.0135	0.0325	0.0328
reflections with <i>I</i> > 2σ(<i>I</i>)	1294	3582	1969	8083	8409
no. parameters	110	405	207	564	750
<i>R</i> (<i>F</i>), <i>F</i> > 2σ(<i>F</i>)	0.0383	0.0357	0.0317	0.0471	0.0371
<i>wR</i> (<i>F</i> ²), <i>F</i> > 2σ(<i>F</i>)	0.1004	0.0420	0.0339	0.0487	0.0399
<i>R</i> (<i>F</i>), all data	0.0389	0.0934	0.0826	0.1270	0.0988
<i>wR</i> (<i>F</i> ²), all data	0.1007	0.1002	0.0846	0.1287	0.1020
<i>Δ</i> _r (min., max.) eÅ ⁻³	0.69, -0.66	0.23, -0.21	0.22, -0.26	2.20, -1.34	0.59, -0.24