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# INTERNATIONAL Clostridium difficile ANIMAL STRAIN COLLECTION

V. Zidaric, S. Janezic<sup>1</sup>, B. Pardon<sup>2</sup>, A. Indra<sup>3</sup>, B. Kokotovic<sup>4</sup>, J.L. Blanco<sup>5</sup>, C. Seyboldt<sup>6</sup>, C. Rodriguez Diaz<sup>7</sup>, I.R, Poxton<sup>8</sup>, V. Perreten<sup>9</sup>, I. Drigo<sup>10</sup>, A. Jiraskova<sup>11</sup>, M. Ocepek<sup>12</sup>, J.S. Weese<sup>13</sup>, J.G. Songer<sup>14</sup>, M. Rupnik<sup>1, 15, 16</sup>

<sup>1</sup>Institute for Public Health Maribor, Maribor, Slovenia; <sup>2</sup>Ghent University, Faculty of Veterinary Medicine, Merelbeke, Belgium; <sup>3</sup>AGES, Vienna, Austria; <sup>4</sup>Technical University of Denmark, National Veterinary Institute, Copenhagen, Denmark; <sup>5</sup>Complutense University, Madrid, Spain; <sup>6</sup>Friedrich-Loeffler Institute, Jena ,Germany; <sup>7</sup>University of Liege, Faculty of Veterinary Medicine, Liege, Belgium; <sup>8</sup>University of Edinburgh, Edinburgh, UK; <sup>9</sup>University of Bern, Institute of Veterinary Bacteriology, Bern, Switzerland; <sup>10</sup>IZSVe, Treviso, Italy; <sup>11</sup>Charles University in Prague, 1<sup>st</sup> Faculty of Medicine, Prague, Czech Republic; <sup>12</sup>University of Ljubljana, Veterinary Faculty, Ljubljana, Slovenia; <sup>13</sup>University of Guelph, Ontario Veterinary College, Ontario, Canada; <sup>14</sup>Iowa State University, Ames, U.S.A.; <sup>15</sup>University of Maribor, Medical Faculty, Maribor, Slovenia; <sup>16</sup>Centre of excellence for integrated approaches in chemistry and biology of proteins, Ljubljana, Slovenia

## Background

Animals have been recognized as an important potential reservoir of *Clostridium difficile* and according to recent studies the overlap between PCR ribotypes of human and animal isolates seems to be increasing (Bakker et al., 2010; Gould and Limbago, 2010; Janezic et al., 2012; Keel et al., 2007; Koene *et al.*, 2011).

Here we report on an International Clostridium difficile animal strain collection that was established to enhance comparative studies on animal-associated strains and contribute to

### Materials and methods

### C. difficile strains:

Altogether 100 strains from 12 different countries were contributed. Collected strains originate from 11 different animal species, including pets, horses and food animals. Approximately half (58,0%) of the strains are from cattle and pigs (Table 1).

interlaboratory exchange of the strains.

The goal of the collection is to include one PCR ribotype per species per country/laboratory.

# **Results and Discussion**

All 100 included strains were distributed into 39 different PCR ribotypes. Up to 17 different PCR ribotypes can be found within a single animal species and up to 15 different PCR ribotypes per country.

Within one standard (agarose based) PCR ribotype several PCR ribotypes could be distinguished by capillary gel based PCR ribotyping (Table 1; PCR ribotypes 014/020, 078, 002, 045, 056, 033).

Five strains are nontoxigenic while toxigenic strains account for 95.0% and belong to 10 different toxinotypes: 0, I, III, IV, V, VI, VIII, XI, XII and XIX. PCR ribotypes 078, 126, 014/020, 012 and 002 that are frequently associated with animals (Keel et al., 2007; Janezic et al., 2012) represent 40.0 % of all strains. For every strain aditional available information on animal host and strain was obtained.

### Molecular characterization of *C. difficile* strains:

All collected strains were characterized by toxinotyping (Rupnik et al., 1998; http://www.mf.uni-mb.si/tox/). In adition, binary toxin genes were detected by PCR as described in Stubbs *et al.* (2000).

Standard agarose gel-based PCR ribotyping was used as described by Bidet et al. (1999) and results analyzed by BioNumerics software 5.10 (Applied Maths).

Strains were also typed by capillary gel electrophoresis-based ribotyping using primers for standard agarose gel-based PCR ribotyping with flourescein labelled 16S primer (Indra et al., 2008). PCR ribotype patterns were analyzed and identified on a web-based database Webribo (http://webribo.ages.at).

Table 1. Distribution of C. difficile genotypes by species and country given as toxinotype/standard PCR ribotype [capillary gel electrophoresis based PCR ribotype].

Country	Austria	Belgium	Canada	Czech Republic	Denmark	Germany	Italy	Scotland	Slovenia	Spain	Switzerland	U.S.A.	Total number of strains per animal species	Number of different PCR ribotypes
Animal host														per animal species
Pig	XII/056 [new RT] 0/011/049 [049/1] 0/150 [AI-12]	V/078 [078] 0/002 [002/0] 0/081 [081]	V/078 [078] 0/015 [211]	0/150 [AI-12]	V/078 [new RT] V/045 [598] V/126 [126] VI/066 [413] 0/150 [AI-12] 0/005 [005] VI/SLO 137 [new RT]		V/078 [251] 0/150 [AI-12] XIX/012 [012] 0/014/020 [014 subtype] 0/081 [081] 0/SLO 036 [050/AI-84] XII/SLO 133 [AI-15] VI/SLO 012 [new RT]	V/078 [078]	V/045 [045]	V/078 [078]		V/078 [078] 0/002 [nd]	29	17
Cattle	V/078 [078] 0/014/020 [014/0] 0/029 [029] 0/005 [005] 0/015 [211] 0/5LO 125 [014 subtype] 0/SLO 036 [AI-84/050] 0/SLO 090 [AI-9-1] 0/SLO 143 [610] 0/SLO 164 [nd] 0/SLO 165 [nd]	V/078 [078] V/078 [078] V/126 [078 ecdc] XI/033 [033] 0/012 [012] 0/014/020 [014/0] 0/002 [002/2] 0/081 [081]	0/SLO 074 [new RT] V/078 [078]				V/126 [126] XIb/033 [033]		0/002 [209] 0/014/020 [014/0]	Xlb/283 [new RT]		V/078 [078] XIa/033 [033] III/027 [027]	29	15
Dog			0/001/072 [nd] 0/002 [002/0]			0/SLO 066 [AI-60] 0/014/020 [014/0] V/SLO 024 [new RT] Tox-/010 [010]			0/014/020 [new RT] 0/012 [012]	XII/056 [new RT] Tox-/010 [010]			10	8
Cat			0/001/072 [001 ecdc]			V/SLO 024 [new RT] 0/014/020 [449]			0/014/020 [new RT]				5	4
Poultry	0/001/072 [001]					10x-/ SLO 002 [AI-34]			0/029 [029] 0/014/020 [014/0] IV/023 [023] 0/103 [AI-82/1] 0/001/072 [001]				6	5
Horse		0/014/020 [020]	XIb/033 [new RT] V/078 [078] III/027 [027]						XI/033 [nd]		V/078 [078] V/126 [126]		7	5
Rabbit/Hare							0/012 [012] V/078 [078] 0/014/020 [014/0] Tox-/SLO 132 [new RT] 0/SLO 090 [AI-9-1] 0/002 [002/0] VIII/017 [017] Tox-/SLO 084 [205]						8	8
Partridge									XII/056 [446] XI/033 [new RT]				2	2
Goose									0/SLO 090 [AI-9-1]				1	1
Crow									0/003 [003]				1	1
Raccoon			I/SLO 166 [nd] 0/103 [AI-82/1]										2	2
Total number of strains per country	15	12	12	1	7	7	18	1	16	4	2	5	100	
Number of different PCR ribotypes per country	15	7	9	1	7	5	15	1	12	4	2	4		

nd- typing not done ; new RT- PCR ribotype has on webribo database not yet been determined; subtype- subtype has on webribo not yet been determined

Bakker D, Corver J, Harmanus C, Goorhuis A, Keessen EC, Fawley WN, Wilcox MH, Kuijper EJ. 2010. Relatedness of human and animal <i>Clostridium difficile</i> PCR ribotype 078 isolates determined on the basis of Multilocus Variable-Number Tandem-Repeat Analysis and tetracycline resistance. J. Clin. Microbiol. <b>48(10)</b> :3744–3749. Bidet P, Barbut F, Lalande V, Burghoffer B, Petit JC. 199. Development of a new PCR-ribotyping method for <i>Clostridium difficile</i> based on ribosomal RNA gene sequencing. FEMS Microbiol. Lett. <b>175</b> :261-266. Gould LH, Limbago B. 2010. <i>Clostridium difficile</i> in food and domestic animals: a new foodborne pathogen? Clin Infect Dis. <b>51(5)</b> :577-582. Indra A, Huhulescu S, Schneeweis M, Hasenberger P, Kernbichler S, Fiedler A, Wewalka G, Allerberger F, Kuijper EJ. 2008. Characterization of <i>Clostridium difficile</i> isolat correlate isolates using capillary gel electrophoresis-based PCR ribotyping. J. Med. Microbiol. <b>57</b> : 1377-1382.
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