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Differences in vertical and horizontal distribution of fish larvae and zooplankton, related to hydrography

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2	Differences in vertical and horizontal distribution of fish larvae
3	and zooplankton, related to hydrography
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17	Running head: Vertical distribution of fish larvae in the North Sea.
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22 Abstract

23 Planktonic fish larvae have little influence on their horizontal distribution, while they are able to control their vertical position in the water column. While prey and light are among the 24 factors with an apparent influence on the vertical distribution, the effects of other factors are 25 less clear. Notably, distributional differences between larvae of different fish species are 26 poorly understood. Information on the horizontal distribution of larvae of 27 species and the 27 vertical distribution of seven species of Gadidae, two Pleuronectidae and one Scopthalmidae, 28 29 was compiled from one survey in the northern North Sea. Horizontally, fish larvae aggregated near frontal structures, correlating with high densities of zooplankton. Increasing 30 31 length and decreasing numbers indicated an origin in the western North Sea, followed by an eastward drift. Vertically, the different species exhibited similarities but also notable 32 differences in their vertical distribution. Most gadoid species aggregated in the upper (<40 m) 33 34 or middle water column (>40 m) during the day with an increase in abundance at shallower depths during the night, while all flatfish were distributed at greater depths under all light 35 36 conditions. Hence, larvae differed in their distributional patterns, but the relative depth 37 distributions among the species in the larval community generally remained constant.

38



41 Introduction

42 Compared to current speeds the swimming ability of fish larvae is of minor importance, limiting their capability to influence their location by horizontal swimming. However, larvae 43 44 can migrate vertically in the water column and so influence their horizontal transport, as current speed and direction often changes with depth (Fortier & Leggett 1983; Sclafani et al. 45 1993). Vertical migration patterns of fish larvae can be broadly classified into three 46 categories: i) type I migrations as upward movement at the beginning of night and downward 47 movement at the beginning of day; ii) type II as the opposite (Neilson & Perry 1990) and iii) 48 a pattern of aggregation during the day and dispersal throughout the night (Gray 1998; Leis 49 50 1991; Olivar & Sabatés 1997). Exogenous factors that influence the observed patterns are, for example, light, prey and predator distribution, as well as effects of temperature and salinity. 51 Individuals of given species and congeners often exhibit similar distribution patterns, 52 regardless of the prevailing environmental conditions and form distinct assemblages in 53 54 different depth strata (Gray & Kingsford 2003; Olivar & Sabatés 1997; Röpke 1993; 55 Southward & Barrett 1983). Size and consequently swimming ability, important for determining vertical distribution, changes throughout development and many species exhibit 56 different vertical behaviours as the larvae develop (c. f. Table 1; Neilson & Perry 1990). 57 Lough and Potter (1993) observed the initiation of vertical migration in cod (Gadus morhua 58 59 Linnaeus, 1758) and haddock (Melanogrammus aeglefinus Linnaeus, 1758) at standard lengths (SL) of 6-8 mm, and a firmly established type I migration at lengths greater than 9 60 mm SL. Smaller larvae and particularly those in poor condition may be more strongly 61 62 influenced by buoyancy (Sclafani et al. 1993). However, even in their earliest stages, larvae will migrate if unfavourable conditions make it necessary (Grønkjær & Wieland 1997). The 63 influence of hydrography, in particular the position of the thermocline, is less clear. Some 64 65 studies indicate a connection between larval distributions and the thermocline for certain taxa

(Olivar & Sabatés 1997) and/or size classes (Lough et al. 1996; Lough & Potter 1993), while
others show the same distributional patterns, both of single taxa and larval assemblages,
irrespective of water column stratification (Gray 1998; Gray & Kingsford 2003). Gray and
Kingsford (2003) attributed their failure to find a relationship between distributions and the
thermocline, to a combination of the gradual and ephemeral character of thermoclines in their
study region and the lag-phase between the occurrence of hydrographic cues and the reaction
of the larvae.

73

74 The influence of prey and predator distributions was pointed out by Pearre (1973) who, based on his studies of an arrow worm Sagitta elegans (Verrill, 1873), introduced the hunger-75 76 satiation hypothesis. In this case vertical movements were related to the concurrent needs of feeding in the upper water column and hiding from visual predators at greater depths. The 77 hypothesis was later applied to other planktonic organisms, including fish larvae (Pearre 78 79 2003). Visually hunting fish larvae can follow different strategies to satisfy these needs. They 80 may rise in the water column at night, together with their zooplankton prey or may stay deeper and feed on vertically migrating prey (Lovetskaya 1953). Neilson and Perry (1990) 81 82 suggested a feeding/avoidance window at dusk and dawn, when light conditions are sufficient for feeding but predators may still be at greater depths. The influence of light differs among 83 species. Some species appear to select a specific isolume, which primarily governs their 84 vertical distribution (Woodhead 1966). This has been suggested as the cause of aggregations 85 during the day and diffuse distribution during the night when the primary cue is missing (Leis 86 87 1991). However, the effect of light is species specific as has been shown in concurrent laboratory studies (Catalán et al. 2011; Vollset et al., in press), for example some species are 88 shown to be adapted to low illumination (e.g. Downing & Litvak 2001; Huse 1994; Yoon et 89 90 al. 2010).

91 Statistical models of the vertical distribution of different taxa have high predictive power 92 with several interacting factors (Hernandez et al. 2009) and even when only using a single factor (Huebert et al. 2010). Control by a single factor is, however, rare. While prey 93 94 abundance was one controlling factor for mesopelagic larvae in the Arabian Sea (Röpke 1993) and for Sardinella aurita (Valenciennes, 1847) in the northwestern Mediterranean Sea 95 (Sabatés et al. 2008), the fish species were also limited by physical factors. The mesopelagic 96 97 species were limited by a warm mixed layer above, and S. aurita most likely by the cool (ca. 15°C) water below the pycnocline. The vertical distribution of larvae will influence 98 99 horizontal transport, as different currents at different depths might lead to retention within or a displacement out of an area (Fortier & Leggett 1982; Fortier & Leggett 1983; Govoni & 100 101 Pietrafesa 1994) and several studies have shown aggregations of fish larvae in or near fronts 102 (Kiørboe et al. 1988; Munk et al. 2002; Sabatés 1990). Likewise, food availability, the 103 relationship between illumination and prey abundance, is correlated with the distribution of Baltic cod larvae (Grønkjær & Wieland 1997). 104

105 Considering the apparent species differences in vertical distributions and migrations, a 106 comparative approach might elucidate the factors that are of prime importance. Few studies have analysed the distributional patterns of a wide range of species in a comparative way 107 (Frank et al. 1992; Gray 1996; Gray & Kingsford 2003). Such an opportunity was available 108 in the northern North Sea in 2010. The area east of the Shetland Isles is particularly species 109 rich (Economou 1987) with an assemblage primarily consisting of Gadidae, Lotidae, 110 Pleuronectidae and Scophthalmidae. In regard to abundance, the dominant species were 111 whiting, ling (Molva molva, Linnaeus, 1758), Norway redfish (Sebastes viviparus, Krøyer, 112 1845) and Norway pout. It is an important spawning ground for several fish species which 113 spawn in spring and we were able to describe both the major horizontal distributional patterns 114 from transects of stations, and the vertical patterns by vertical stratified sampling over an 18 115

hour period. In this contribution we focus on the distributional patterns of larval fish in
relation to hydrography and in relation to the distribution of zooplankton 180 - 1000 µm. We
hypothesize that relative to each other larvae of different species would retain their position
in the water column.

120

121 Materials and Methods

122 *Field sampling*

Sampling was undertaken on the *RV G.O. Sars* (IMR, Bergen, Norway), between 25^{th} April and 5^{th} of May 2010, covering transects between 59.3 and 60.75° N (Figure 1). Five additional stations were sampled over the course of 18 hours in a 5 x 5 nautical miles (NM) sized area

126 (designated 18h-station) east of the Shetland Islands.

Depth integrated samples were taken in double oblique hauls with a 76 cm diameter GULF 127 VII high speed sampler (Nash et al. 1998), down to about 100 m depth. The sampler was 128 equipped with a mechanical flow meter (General Oceanics, USA) in the mouth of the nose 129 cone. A SCANMAR depth sensor was attached to the sampler and provided both depth and 130 temperature measurements. For discrete depth sampling, a MOCNESS (Wiebe et al. 1985) 131 with a 1 m^2 opening and 4 nets (180 μ m mesh) was deployed to ca. 100 m and then hauled 132 obliquely to the surface, sampling the water column in strata with nets opening at about 100, 133 75, 40 and 20 m. Flow meters and a CTD were attached to the MOCNESS and the filtered 134 volume (m³) estimated for each stratum. Larvae were sorted on board and were preserved in 135 borax buffered 4% formaldehyde. Zooplankton was split in two fractions before preservation, 136 using a Motoda splitting device. One half was preserved for identification and enumeration 137 whilst the other half was size fractioned into $<1000 \,\mu m$, $1000-2000 \,\mu m$ and $>2000 \,\mu m$ 138

samples. Each size fraction was dried at 60°C to constant weight in order to obtain dry
weights, which were converted to milligrams dry weight per m³ (mg DW m⁻³) based on the
volume of water filtered and to g DW m⁻² based on filtered volume and sampled depth.

142

143 *Laboratory procedures*

The preserved larvae were cleaned of formalin under running water for 10-15 minutes. All
larvae were then identified to the lowest taxonomic level, using either Russell (1976),
Schmidt (1906) or Munk and Nielsen (2005). Standard length (SL; tip of the snout to the end
of the notochord) was measured to the nearest 0.1 mm with an ocular micrometer. To correct
for shrinking, live SL was calculated using the equation from Bolz and Lough (1984), after
correcting for formalin shrinkage (Theilacker 1980).

150

151 *Data treatment and analysis*

Density anomaly (σ_t) was calculated according to UN standards (Millero & Poisson 1981)
from temperature and salinity measured by CTD casts during the transects. The vertical
profiles of calculated densities were interpolated on a regular grid (0.5° x 5 m) with kriging in
Surfer 8 (Golden Software 2002), while contour plots were constructed in Sigmaplot 12
(Systat Software 2011). The vertical profiles for the five hauls at the 18h-station are given as
line graphs.

For each species in the depth integrated hauls, the catch was converted to nos. m⁻² by dividing by the filtered volume and multiplying by the maximum sampler depth. Catch of larvae in the depth discrete hauls was converted to nos. m⁻³ by dividing by the filtered volume in a given

stratum and these values were used in calculation of the depth of the centre of abundance
(Z_{cm}) from

163

$$Z_{cm} = \frac{\sum D_j \times W D_j \times A_j}{\sum W D_j \times A_j} \tag{1}$$

164

165 Where D_j is the midpoint of stratum *j*, WD_j the width of the individual stratum and A_j is the 166 abundance of the larvae. The depth of mass for zooplankton <1000 µm was calculated using 167 the same formula, but replacing abundance with dry weight in mg m⁻³. The relative 168 abundance of larvae in each stratum was plotted as a % of total abundance for day and night. 169 Z_{cm} was calculated and plotted for day, dusk, night and for single samples.

Only species for which the maximum abundance of larvae in a given stratum was above 2 per 100 m³ were used (10 out of 27 species; 37%), as was the abundance of zooplankton <1000 μ m. The station sampled at 06:20 UTC was excluded from calculations for day distributions and Z_{cm}, as it was the first sample after sunrise and considered to be biased by the night distribution. Abundances per stratum were compared visually between species and between day and night. Similarly, Z_{cm} was compared among species for day (19:14 UTC, 08:22 UTC), dusk (21:52 UTC) and night (23:56 UTC) as well as the relationship of species to the

177 hydrography in the transects.

178 The depth of the centre of abundance was tested for significant differences between species,

using one-factorial ANOVA for all species together and for Gadidae and flatfish separately.

180 Data were tested beforehand with a Shapiro-Wilks and Levene's test and were found to fulfil

- the requirements for normality and homogeneity of variance. Post hoc Tukey's HSD was
- applied to discern between which species significant differences occurred.

183 **Results**

184 *Hydrography*

Along both transects we observed a cool ($<7^{\circ}$ C), low saline (<34) surface layer over the 185 Norwegian trench, extending to ca. 50 m depth (Figure 2), representing the Norwegian 186 Coastal Current (NCC). Coldest temperatures occurred at ca. 30 m, while lowest salinities 187 188 and densities were at about 10 m depth (Figures 2a, b). Correspondingly, σ_t was increasing with depth and ranged from 25.5 kg m^{-3} to 27 kg m^{-3} . Beneath the NCC water, the 189 temperature increased down to 200-300 m, while at greater depths temperatures fell below 190 7° C and σ_{t} rose to 27.6 in the deepest parts of the Norwegian trench. On the shallow plateau, 191 between 1°W and 3°E, temperature changed markedly with depth, while salinity was almost 192 193 homogenous throughout the water column, except for the eastern margins. In the southern transect a thermocline at about 50 m was separating water of >7°C and σ_t of 27.5 kg m⁻³ from 194 cooler and denser water below. In the northern transect the warmer water reached down to a 195 196 depth of 100 m and the thermocline was less strong. On the western margins of the southern transect water temperature increased rapidly between 0.5°W and 1°W, while salinity 197 decreased from about 1.7°W westwards. Together this led to the formation of a frontal 198 199 structure. In the North, temperature increased more gradually, while salinity did not change. Overall the highest temperatures were measured at $>8^{\circ}$ C on the western margins. Throughout 200 the northern transect the surface water exhibited a σ_t of <27.5 kg m⁻³ while on the western 201 margin these lower densities reached down to a hundred metres. 202

The hydrography at the 18h-station exhibited little variability in time or depth (Figure 3).
Salinity was relatively high and stable, only changing from 35.32 to 35.33 in the sampled
water column of 120 m. The temperature likewise varied little; it was about 8°C to 50 m and

then declined continuously to 7.6°C. Fluorescence peaked at $0.12 \ \mu g \ L^{-1}$, but estimates varied during the period of investigation.

208

209 Horizontal distribution - zooplankton

At the stations closest to the Norwegian trench the total zooplankton concentration in both 210 transects ranged between 3.7 g DW m^{-2} and 5.0 g DW m^{-2} . Peak zooplankton concentrations 211 were found at the stations near 1°E, 30.5 g DW m⁻² in the South and 38.7 g DW m⁻² in the 212 North. However, at these stations the distribution between size fractions differed. While at the 213 northern station, the zooplankton biomass was nearly equally distributed between the three 214 different size fractions (Table 1), at the southern station the bulk of the zooplankton (20.1 g 215 DW m⁻²) was in the 1000 – 2000 μ m size fraction, while the zooplankton <1000 μ m was at 216 7.1 g DW m⁻² and the >2000 μ m size fraction was at 3.2 g DW m⁻². At the westernmost 217 stations zooplankton concentrations were again lower, with 19.9 g DW m^{-2} in the southern 218 and 11.7 g DW m⁻² in the northern transect for all size fractions combined. 219

220

221 *Horizontal distribution – fish larvae*

222 During the survey, a total of 2030 fish larvae of 27 species in 9 families were identified 223 (Table 2). Species richness and abundance of fish larvae increased from east to west. In the 224 area of the Norwegian trench, abundances were mostly $<30 \text{ m}^{-2}$ (Figures 2a, b). In this area 225 there were no flatfish and there were only gadoid larvae close to the western slope of the 226 trench. Over the shallow plateau abundances were mostly low ($<10 \text{ m}^{-2}$), however long 227 rough dab (*Hippoglossoides platessoides* Fabricius, 1780) and Norway pout (*Trisopterus* 228 *esmarkii* Nilsson, 1855) occurred at abundances of ca. 200 m⁻² and 300 m⁻², respectively. 229 Both the stations with these high abundances were at the boundary of salinities between 35 and 35.2, where also sharp changes in σ_t and high concentrations of zooplankton <1000 µm 230 $(7.1 \text{ g DW m}^{-2} \text{ and } 5.1 \text{ g DW m}^{-2})$ were observed. Along the northern transect larval 231 abundance and species diversity increased from the western slope of the Norwegian trench 232 westward to ca. 1°E, up to a maximum abundance of 500 m⁻² (Figure 2b), coinciding with 233 peak zooplankton densities. Along both transects the dominant species was Norway pout, 234 followed by whiting (Merlangius merlangus Linnaeus, 1758). Flatfish of the families 235 Pleuronectidae and Scophthalmidae were more abundant and species rich at the northern 236 237 transect than at the southern. Notably, Ammodytidae of 3 species were limited to the southern transect with only lesser sandeel (Ammodytes marinus Raitt, 1934) at >10 m⁻². Ling (Molva 238 *molva* Linnaeus, 1758) was found in high abundance, (33.3 m^{-2}) , at one station of the 239 northern transect, but did not occur elsewhere (Table 2a). 240 241 At the single location between the two transects whiting was almost twice as abundant as Norway pout, while other gadoids were much less abundant ($<20 \text{ m}^{-2}$) than either of these 242 243 (Table 2c). Blue ling (Molva dipterygia Pennant, 1784) and northern rockling (Ciliata septentrionalis Collett, 1875) were found in abundances over 20 m⁻². Flatfish were similarly 244 species rich and abundant as in the northern transect. Long rough dab and brill 245 (Scolphthalmus rhombus Linnaeus, 1758) were most abundant, with 25.2 m⁻² and 18.6 m⁻², 246 respectively. Clupeidae, Argentinidae and Gobiidae occurred sporadically along the transects 247 as well as at the 18h-station, in some hauls and in high numbers (Table 2). 248

249

250 *Vertical distribution – 18 hours station*

Changes in zooplankton distribution between day and night varied between the size fractions.
While the distribution of zooplankton <1000 µm varied only little (Figures 3b, c and 4) and

253 being most abundant in the two topmost strata (>30% each), coincided positively with the level of fluorescence. The larger size fractions exhibited stronger differences (Figures 3b, c), 254 particularly the $1000 - 2000 \,\mu m$ fraction which was proportionally most abundant in the 0 -255 256 20 m stratum during the day and almost homogenously distributed during the night. The trend towards a larger proportion in the deep strata during the night was common for all size 257 fractions and was reflected in the depth of the mass of the small zooplankton which was 258 259 relatively stable at around 40 m with noticeable but small deviations at night (Fig. 6) and when incorporating the station at 06:20 UTC (Fig. 7). 260

261 Seven gadoids and three flatfish species occurred in sufficient numbers to examine their

vertical distribution. Cod was absent from the sample taken at dusk, otherwise all species

263 occurred in all hauls. One group of gadoid larvae, consisting of cod (Gadus morhua),

264 haddock (Melanogrammus aeglefinus), whiting and pollock (Pollachius pollachius Linnaeus,

1758), was distributed in the upper water column (0 - 40 m) during day and night. Cod (62%) and haddock (52%) were most abundant at 0 - 20 m during the day and at 20 - 40 m at night, with 100% and 69% respectively (Figures 4a, b). For whiting (Figure 4c) and pollock (Figure 4d) the change between these strata was reversed, as their abundance increased by 32 and 38 percent at 0 - 20 m during the night. While cod was never found below 40 m depth, the other species occurred in the deeper strata and ascended to shallower depths at night, Z_{cm} decreased accordingly (Figures 6a, 7a).

272 Saithe (*Pollachius virens* Linnaeus, 1758) and the two *Trisopterus* species (Figures 4e, g)

were distributed in the strata below 40 m during the day. During the night saithe and poor cod

274 (*Trisopterus minutus* Linnaeus, 1758) were most common in the upper water column, while

275 53% of Norway pout larvae remained at 75 – 100 m depth.

In daylight all three flatfish species, witch (*Glyptocephalus cynoglossus* Linnaeus, 1758),
brill and long rough dab were most abundant at 40 – 75 m depth (Figure 5), varying between
45% for witch and 57% for brill. During the night, witch and long rough dab were most
abundant in the upper water column, peaking with 47% at 0 – 20 m and 77% at 20 – 40 m,
respectively. Brill remained most abundant at 40 – 75 m depth.

Except for brill, most larvae fell into a size range between 3 and 9 mm (Figs 8 - 10), with 281 larger larvae occurring at low numbers. Brill was much more common at standard lengths of 282 2 - 3 mm than other species, while no brill larvae were longer than 5 mm. Even such small 283 larvae exhibited substantial changes in their distribution across depth strata (Figure 10), 284 indicating that they were capable of controlling their position in the water column. With 285 increasing standard length, whiting exhibited a tendency to be proportionally more common 286 in the 0 - 20 m stratum, which was particularly noticeable at night (Figure 8). Norway pout, 287 288 the other gadoids found over a wide size range, did not exhibit such a trend and between 4 and 7 mm length exhibited a reversed trend of a larger proportion in the 20 - 40 m stratum at 289 290 a small size, while the larger larvae were in the deepest stratum at night (Figure 9). Larvae 291 above 9 mm appeared to aggregate in one or the other strata, depending on species and prevailing light conditions. 292

When testing the depth of the centre of mass for difference between species, results were only significant within a single family, *Gadidae* ($F_6=2.5$; p=0.047), but not for the group of flatfish ($F_2=0.2$; p=0.82) or in an analysis of all species together ($F_9=1.8$; p=0.1). The pattern in change of Z_{cm} , between different light conditions was similar for most species (Figures 6, 7). Z_{cm} decreased at night, except for cod, Norway pout and brill. While cod was found at greater depth during the night, Norway pout and brill had already ascended between day and dusk.

299

300 Discussion

301 Our study provides evidence for type I vertical migrations in the species examined, except for cod (Gadus morhua). However, in regard to timing, the migration patterns were not identical, 302 303 as Norway pout (Trisopterus esmarkii) and Brill (Scophthalmus rhombus) ascended earlier and pollock (Pollachius pollachius) continued to rise until the early morning. With the 304 exception of the two Trisopterus species the centre of abundance of all species was within the 305 20 - 40 m stratum either at dusk or during the night. In contrast to previous studies (Gray 306 1996; Olivar & Sabatés 1997) we observed distinct assemblages in the upper and lower parts 307 of the water column only during the day. 308 Our hydrographic observations are in accordance with findings described for the Feie-309 310 Shetland transect, reported by Hackett (1981). Hydrographic fronts were apparent at the

western and eastern margins of the transects. Larval abundances and zooplankton
concentrations were highest in the vicinity of these fronts which might imply that the frontal
processes aggregate the zoo- and ichthyoplankton (Olson et al. 1994; Olson & Backus 1985).
Larval drift and dispersion from spawning grounds around the Shetland Isles is indicated by
the general decline in larval abundance and diversity in parallel with an increase in larval
mean lengths from these areas towards the East. Similar patterns have been suggested for
Norway pout in other studies (Lambert et al. 2009; Nash et al. 2012).

In accordance with an east-west size gradient, the smallest average standard lengths were measured at the westerly positioned 18h-station. Cod and haddock (*Melanogrammus aeglefinus*) larvae were in the 6 - 8 mm size range in which Lough and Potter (1993) have observed the first appearance of vertical migrations. While our observations of cod larvae contain a high level of uncertainty, due to the low number of cod larvae in the samples, the distribution appears similar to earlier studies. The lack of cod larvae below 40 m is in

324 accordance with other observation of early cod larvae confined to the waters above the thermocline (Grønkjær et al. 1997; Grønkjær & Wieland 1997; Huwer et al. 2011; Lough & 325 Potter 1993). Our observations of Type II distributions in cod larvae were described earlier 326 327 for both the Atlantic and the Pacific cod (Gadus macrocephalus Tilesius, 1810) (Boehlert et al. 1985; Munk, in press). The depth distributions found for haddock, whiting, pollock, 328 Norway pout, witch (Glyptocephalus cynoglossus) and long rough dab (Hippoglossoides 329 330 platessoides) were similar to the findings of Economou (1987). The propensity for large whiting larvae to occur shallower at night may be explained by their greater ability to rise 331 332 quickly. This is supported by the increasing proportion of smaller larvae in the 20 - 40 m stratum. Apparently all whiting larvae were rising through the water column but the larger 333 larvae were rising more rapidly. In comparison, Norway pout showed a different trend and 334 335 generally less distinct differences between day and night. Saithe exhibited less variation in Z_{cm} in earlier studies (Munk, in press). Poor cod (Trisopterus minutus) was found shallower 336 than in the present study (Olivar & Sabatés 1997). During the day Frank et al. (1992) found a 337 338 shallower distribution of witch and long rough dab than in this study. However the bottom depth in their study was at 45 m, which may have restricted the depth distribution. The 339 340 distribution of brill appears not to be described in the literature. In many ways it resembled the distribution of Norway pout, concerning the particularly deep Z_{cm} and the timing of the 341 ascent. However the extent of the vertical migration was greater, covering 43 m. Notably, 342 brill larvae, which were on average smaller than those of other species, exhibited the largest 343 difference in Z_{cm} between day and night, suggesting that already at this small size brill larvae 344 were capable of controlling their position in the water column. The overall tendency of large 345 346 larvae to aggregate may reflect the developing patchiness in the distribution of older larvae (Hewitt 1981; Matsuura & Hewitt 1995). However, the low number of larvae above 9 mm SL 347 348 resulted in a great deal of uncertainty concerning diel shifts in distribution.

Thermoclines have been described to lead either to larval aggregation (Lough & Potter 1993; Sabatés et al. 2008) or serve as a boundary for their migrations (Olivar & Sabatés 1997; Röpke 1993). Other studies found no apparent influence of thermoclines on larval vertical distribution and migration patterns (Conway et al. 1997; Gray & Kingsford 2003). The weak stratification resulting in a weak thermocline observed at the 18-hours station is similar to conditions in the studies of Gray and Kingsford (2003) and this might be the cause of the apparent weak influence of the thermocline in both studies.

The aggregation in the 20 - 40 m stratum during the night suggests a support for the 356 hypothesis that a hungry population would ascend just far enough to find sufficient food 357 (Pearre 2003). The zooplankton that could be quantitatively sampled with the available 358 equipment was generally too large to be potential prey for all but the largest fish larvae. Even 359 though the small-sized copepods and nauplii are under-sampled by the 180 µm mesh, we 360 consider the distribution of the <1000 µm size fraction to reflect the distribution of smaller 361 zooplankton. The smallest size fraction was concentrated in the upper water column which 362 363 would be consistent with the aggregation of nauplii of most copepod species above the thermocline, which was observed in an earlier study (Krause & Trahms 1982). 364

Gadoid larvae in the observed size range primarily feed on *Calanus finmarchicus* (Gunnerus, 1770) eggs and copepod nauplii (Economou 1991) and require about 36% d⁻¹ of their own body mass (Jones 1973). For a larva of 6 mm standard length this would mean a requirement between 68 μ g for saithe and 125 μ g for cod (calculated following Economou 1987). Assuming a swimming speed and a reaction distance of one body length as well as

370 proportions between *C. finmarchicus* eggs and nauplii and between nauplii stages as in

Economou (1987) and Fransz et al. (1998) the corresponding number of food particles would

be 3260 m^{-3} and 5973 m^{-3} , respectively. In May such numbers are not unrealistic in the area

373 (Economou 1987) and would be well within the 27.6% loss of biomass due to the mesh size

used (interpolated from table III in Gallienne & Robins 2001) However, zooplankton concentration in deeper strata should still have been sufficient to fulfil food requirements, which may explain why Z_{cm} of Norway pout were not found any shallower than 51 m (equation from Economou 1987; based on: Jones 1973; Laurence 1985). The deepest Z_{cm} observed after the apparent feeding period, could be due to larvae resting in deeper, cooler water to save energy and avoid visual predators (Brett 1971) or less buoyancy due to a full stomach (Sclafani et al. 1993).

In conclusion, whilst the general observation that most of the larvae occur at depths with high 381 concentrations of zooplankton suggests a strong influence from the distribution of potential 382 383 prey, the general vertical displacement of the mean depth indicates that other environmental factors might set a species-specific 'background-depth' of distribution. Therefore the physical 384 water column structure might be the key factor determining the distribution of fish larvae, 385 386 rather than the prey distributions. As suggested by Sclafani (1993), the neutral buoyancy of fish larvae is influenced by their condition. Further developed or better fed larvae, may be 387 388 deeper in the water column, due to higher specific weight. As the species differ in the proportion of tissue types, the depth of neutral buoyancy may be different even when the 389 larvae are in the same condition. We find that the comparative approach used in the present 390 391 study has the potential for a new insight into the drivers behind vertical distribution patterns, and we suggest that further comparative community studies are undertaken. 392

393

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564	Table 1: Zooplankton densities (g DW m ⁻²), per size fraction for all sampled stations, based on GULF VII hauls. The highest abundances along
565	transects were found at the stations at ca. 1°E. Proportions differed between transects. While the biomass in the southern transect was dominated
566	by the 1000-2000 µm size fraction, proportions in the northern transect and at the 18h-station were more even between the two smaller size
567	fractions. Large zooplankton (>2000 μ m) was generally scarce with the exception of a few stations, where it contributed to a large proportion of
568	the biomass.

				Density per size fra	ize fraction (g DW m ⁻²)	
Transect	Station No.	Longitude	180-1000 μm	1000-2000 μm	>2000 µm	Total
60.75°N	423	0.47°W	7.0	4.6	0.1	11.7
	429	0.91°E	19.5	17.3	1.9	38.7
	433	2.60°E	2.1	11.6	0.2	13.9
	437	3.28°E	0.9	2.7	0.2	3.7
	444	4.45°E	1.7	2.0	0.0	3.7
59.3°N	388	4.83°E	3.1	1.3	0.6	5.0
	402	2.52°E	0.7	2.8	0.4	3.9
	406	1.32°E	7.2	20.1	3.2	30.5
	410	0.00°E	0.7	5.8	0.1	6.6
	414	1.32°W	5.1	10.1	4.7	19.9
18h-St.	418	0.61°W	5.0	4.6	0.9	10.5
	419	0.65°W	4.6	8.0	0.4	12.9
	420	0.68°W	4.8	5.8	0.2	10.8
	421	0.61°W	25.0	11.5	19.5	56.0
	422	0.68°W	16.0	10.4	3.2	29.6

570 **Table 2:** Average abundances and standard lengths (±1 SD) for all species identified in the northern transect (a), the southern transect (b) and at

- 571 the 18h-station (c). Numbers are based on depth integrated GULF VII, except for species which were only found in MOCNESS hauls. These
- 572 species are denoted with asterisks.

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	Taxon			Abundance			%	Std. Length		
Transect	Family	Species	(nos. m ⁻²)			caught	measured	(mm)		
60.75°N	Clupeidae	Clupea harengus	6.9	±	5.8	30	93.3	17.8	±	2.7
	Gadidae	Melanogrammus aeglefinus	2.7	±	4.8	4	100.0	8.9	\pm	3.1
		Merlangius merlangus	22.4	\pm	40.4	27	100.0	7.4	±	1.4
		Pollachius pollachius	2.3	\pm	2.4	7	100.0	8.5	±	1.9
		Pollachius virens	8.5	\pm	11.6	14	100.0	9.7	±	2.9
		Trisopterus esmarkii	51.3	\pm	71.9	100	100.0	9.2	±	2.5
		Trisopterus minutus	7.0	\pm	14.7	8	100.0	8.4	±	1.0
		Unidentified	3.0	±	4.3	8	62.5	6.6	±	4.1
	Gobiidae	Gobiusculus flavescens	3.3	±	7.4	3	100.0	6.9	±	1.6
	Lotidae	Ciliata septentrionalis	2.4	±	4.9	3	100.0	5.4	\pm	1.0
		Molva dipterygia	1.1	\pm	2.5	1	100.0	6.5	±	-
		Molva molva	6.7	\pm	14.9	6	100.0	5.2	±	0.6
	Pleuronectidae	Glyptocephalus cynoglossus	2.5	\pm	4.8	3	100.0	9.7	±	1.0
		Hippoglossoides platessoides	6.0	\pm	9.7	10	100.0	8.6	\pm	1.8
		Limanda limanda	4.7	\pm	6.9	6	100.0	8.0	\pm	4.0
		Pleuronectes platessa	2.2	\pm	5.0	2	100.0	6.3	\pm	0.4
		Unidentified	0.7	±	1.1	3	66.7	7.8	±	1.1
	Scophthalmidae	Lepidorhombus whiffiagonis	1.1	\pm	2.5	1	100.0	10.6	\pm	-
		Phrynorhombus norvegicus	0.4	\pm	0.6	2	100.0	9.2	±	2.0
		Scophthalmus rhombus	3.6	\pm	7.3	4	100.0	4.9	±	1.5

Taxon			Abundance			nos.	%	Std. Length			
Transect	Family	Species	(nos. m ⁻²)			caught	measured	(mm)			
59.3°N	Ammodytidae	Ammodytes marinus	3.1	±	6.9	7	100.0	16.2 ±	-		
		Hyperoplus lanceolatus	1.8	\pm	2.8	10	100.0	$18.8 \pm$	5.3		
		Unidentified	0.5	\pm	1.0	2	100.0	$17.5 \pm$	3.7		
	Argentinidae	Argentina sphyraena	0.4	\pm	1.0	1	100.0	$10.0 \pm$	-		
	Clupeidae	Clupea harengus	5.8	±	12.6	117	94.9	17.1 ±	26.3		
	Gadidae	Gadus morhua	4.4	\pm	7.7	11	90.9	$8.3 \pm$	3.5		
		Melanogrammus aeglefinus	7.6	\pm	9.8	22	100.0	9.7 \pm	5.0		
		Merlangius merlangus	5.2	\pm	8.7	12	100.0	5.9 \pm	8.2		
		Pollachius pollachius	3.1	\pm	4.8	8	100.0	9.6 ±	6.7		
		Pollachius virens	2.6	\pm	4.8	6	100.0	$10.4 \pm$	10.5		
		Trisopterus esmarkii	60.9	\pm	79.9	159	96.2	$8.8 \pm$	5.0		
		Trisopterus minutus	4.4	\pm	7.7	10	100.0	6.3 ±	7.9		
		Unidentified	2.2	\pm	3.8	5	20.0	$9.2 \pm$	-		
	Lotidae	Ciliata mustela	0.9	\pm	2.0	2	100.0	5.1 ±	0.2		
	Pleuronectidae	Hippoglossoides platessoides	4.6	±	5.6	14	100.0	11.2 ±	3.1		
		Limanda limanda	2.6	±	4.8	6	83.3	7.8 \pm	1.7		
	Scophthalmidae	Scophthalmus rhombus	0.9	±	2.0	2	100.0	3.8 ±	0.3		

b

c											
	Taxon			Abundance			% Std. J		Ler	Length	
Transect	Family	Species	(nos. m^{-2})			caught	measured	(mm)			
18h-St.	Ammodytidae	Hyperoplus immaculatus*	0.2	±	0.4	1	100.0	11.9	±	0	
		Hyperoplus lanceolatus	0.8	±	1.8	1	100.0	40	±	-	
	Argentinidae	Argentina sphyraena	20.0	±	29.2	11	90.9	10.1	±	2.4	
	Clupeidae	Clupea harengus	0.5	±	1.1	1	100.0	14.9	±	-	
	Gadidae	Gadus morhua	1.3	\pm	2.8	1	100.0	5.8	\pm	-	
		Melanogrammus aeglefinus	3.6	\pm	2.4	4	100.0	6.3	±	1.4	
		Merlangius merlangus	152.3	\pm	138.5	113	99.1	6.3	±	1.3	
		Pollachius pollachius	9.4	\pm	15.0	5	100.0	7.1	\pm	1.6	
		Pollachius virens	15.3	\pm	8.5	13	100.0	7.0	±	2.4	
		Trisopterus esmarkii	81.2	\pm	77.6	57	98.2	7.2	±	1.6	
		Trisopterus minutus	7.4	\pm	7.6	5	80.0	6.2	\pm	1.4	
		Unidentified	14.4	±	11.7	13	76.9	5.3	±	1.0	
	Gobiidae	Gobius niger	0.8	±	1.8	1	100.0	5.5	±	-	
		Gobiusculus flavescens	3.6	\pm	8.0	1	100.0	6.8	\pm	-	
		Unidentified*	0.2	±	0.4	1	100.0	2.7	\pm	-	
	Lotidae	Ciliata septentrionalis	17.8	±	22.0	14	100.0	5.7	±	0.9	
		Molva dipterygia	7.2	\pm	16.0	2	100.0	7.9	\pm	0.2	
		Molva molva	2.5	±	5.7	2	100.0	6.9	±	0.8	
	Pleuronectidae	Glyptocephalus cynoglossus	6.2	±	6.9	4	100.0	6.9	±	1.3	
		Hippoglossoides platessoides	20.2	\pm	29.2	12	83.3	8.3	±	3.3	
		Limanda limanda	2.5	\pm	5.6	5	80.0	6.8	\pm	1.6	
		Platichthys flesus*	0.4	\pm	0.9	5	100.0	3.5	±	0.5	
		Pleuronectes platessa	0.8	\pm	1.8	1	100.0	10.4	\pm	-	
		Unidentified	1.3	±	1.9	2	100.0	5.3	±	0.5	
	Scophthalmidae	Scophthalmus rhombus	18.6	±	20.2	12	100.0	4.5	±	0.7	

577 Figure captions:

Figure 1: CTD, GULF VII and MOCNESS stations sampled during the survey. The
aggregation of samples in the black rectangle represents the 18 hours station, containing 5
hauls with each gear in a 5 x 5 NM square.

581

Figure 2: Profiles of σ_t , contoured for 0.1 kg m⁻³ (thin grey lines) and 0.5 kg m⁻³ (bold grey lines) and abundance of fish larvae along the transects at 59.3°N (panel a) and 60.75°N (panel b). Only the most common species are given, while gadoids other than Norway pout and whiting, and flatfish other than long rough dab and brill are combined. Miscellaneous species comprised Clupeidae, Argentiniade, Ammodytidae, Lotidae and Gobidae which did not commonly occur.

588

Figure 3: Temperature, salinity and fluorescence at the 18h-station (panel a), averaged over all 5 hauls. The broken lines depict the boundaries between the sampled depth strata in depth discrete hauls. Most changes in hydrography and fluorescence occurred between 50 and 80 m, mainly in the stratum between 40 and 75 m. Error bars are only shown for every ten metres of depth. Panels b and c show the distribution of all size classes of zooplankton (<1000 μ m, 1000-2000 μ m and >2000 μ m) during daylight and night conditions in % of total.

596

Figure 4: Vertical distribution of gadoid fish larvae and <1000 µm zooplankton by dry
weight, during day and night as a % of total abundance or biomass. N represents the number

of larvae caught under the respective light conditions (in subscript). The y-axis depicts theboundaries between sampled strata.

601

Figure 5: Vertical distribution of flatfish larvae and <1000 µm zooplankton dry weight,
during day and night in % of total abundance or biomass. N represents the number of larvae
caught under the respective light conditions (in subscript). The y-axis depicts the boundaries
between sampled strata.

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Figure 6: Depth of the centre of abundance for gadoid (a) and flatfish larvae (b) in three different light environments. Due to the long days at this time of the year, there was only one station at dusk (21:52 UTC) and night (23:56 UTC), while three stations were in daylight (19:14 UTC, 06:20 UTC and 08:22 UTC). As it was shortly after sunrise the station at 06:20 UTC was not included into the calculation of Z_{cm} . The number of larvae caught under each light condition is given as N in the legend. The depth of mass for zooplankton (based on mg m⁻³) is depicted in both panels.

614

Figure 7: Depth of the centre of abundance for gadoid (a) and flatfish larvae (b) for
individual samples taken at the 18h-station. Daylight stations were at 19:14 UTC, 06:20 UTC
and 08:22 UTC, the station at 21:52 UTC was during dusk and the station at 23:56 UTC in
the night. The number of larvae caught at each station is given as N, with the time of
sampling given in subscript. The depth of mass for zooplankton (based on mg m⁻³) is depicted
in both panels.

Figure 8: Rounded length distribution across strata and light conditions of cod, haddock and whiting as % of total abundance. The majority of larvae ranged from 3 to 6 mm standard length. Even for the larvae at the lower end of this range, changes in distribution across strata could change substantially between the different light conditions. Empty panels indicate zero findings for the respective species in this stratum, during the entire sampling period.

627

Figure 9: Rounded length distribution across strata and light conditions of saithe, pollock, 628 Norway pout and poor cod in % of total abundance. The majority of saithe and pollock were 629 in a relatively narrow size range from 4 to 8 mm SL. Smaller larvae tended to aggregate at 630 the 20 - 40 m stratum with increasing darkness. Larger larvae were distributed throughout the 631 632 water column, but this is again based on few individuals. Norway pout covered a large size range (2 - 11 mm) and, similar to saithe larvae (4 - 6 mm), tended to aggregate in the 20 - 40 633 m stratum with increasing darkness. During day and dusk conditions poor cod of all sizes 634 635 were mostly found in the deeper strata. At night only a few large larvae in the 0 - 20 m 636 stratum were found. Empty panels indicate zero findings for the respective species in this stratum, during the entire sampling period. 637

638

Figure 10: Rounded length distribution across strata and light conditions of witch, long rough dab and brill in % of total abundance. Witch and long rough dab ranged mostly between 3 and 9 mm in standard length but with a few larvae in the extreme upper range of the size distribution which were found in the two strata between 20 and 75 m. The medium sized larvae were relatively dispersed during day and dusk and for witch appeared to aggregate in the uppermost stratum during the night. Brill was unique, as the majority of

- 645 larvae were found at the low extreme of the size range and exhibited strong fluctuations
- 646 across the depth range.







Figure 3:





Figure 5:







Figure 7:







