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Effects of ambient versus reduced UV-B radiation on high arctic *Salix arctica* assessed by measurements and calculations of chlorophyll *a* fluorescence parameters from fluorescence transients

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A UV-B exclusion-experiment was conducted in the high Arctic Zackenberg, NE Greenland, in which *Salix arctica* leaves during most of the growing season were fixed perpendicular to the solar zenith angle, thereby receiving maximal solar radiation. Covered with Teflon and Mylar foil, the leaves received approximately 90 and 40% of the ambient UV-B irradiance, respectively. The effects were examined through recordings of chlorophyll *a* fluorescence transients, determination of biomass and analysis of total carbon and nitrogen content and amount of soluble flavonoids in the leaves. The processing of light was analysed by means of the chlorophyll *a* fluorescence transient, using the so-called JIP test, as evolved by Reto J. Strasser and his coworkers. Reduction of the UV-B irradiance caused a rise in many of the fluorescence parameters during July, but not in August (late season). Thus increases in the efficiency that an absorbed photon will be trapped by the PSII reaction centre with the resultant reduction of Q_A to Q_A^- ($ET_0/ABS = F_V/F_M$) and the efficiency that an electron residing on Q_A^- will enter the intersystem electron transport chain (ET_0/TR_0) were observed in reduced UV-B. Moreover, estimated per cross-section of leaf sample, the number of active PSII reaction centres (RC/CS_M) and electron transport rate (ET_M/CS_M) and all performance indexes (PI_{ABS} , PI_{CS_0} and PI_{CS_m}) were increased in reduced UV-B. The total soluble flavonoid content was highest in ambient UV-B. The treatment effects on fluorescence parameters that were directly measured (e.g. F_0 and F_M) and those that were derived (e.g. quantum efficiencies, parameters per PSII reaction centres and per cross-section of leaf sample) are discussed in relation to one another, in relation to daily and seasonal variation, and from the perspective of evaluating the relative importance of UV-B of donor and acceptor side capacity in Photosystem II. In conclusion, the experimental set-up and non-invasive measurements proved to be a sensitive method to screen for effects of UV-B stress.

Abbreviations – D1, core subunit of PSII; JIP test, analysis of parameters derived from the fluorescence transient (see Table 1); Mn cluster, four ion cluster which is a part of OEC going through five redox states during water splitting; OEC, oxygen evolving complex extracting electrons from water; PSI, Photosystem I; PSII, Photosystem II; Q_A , Plastoquinone A carrier of electrons from P680; Q_B , Plastoquinone B carrier of electrons from Q_A ; Tyr, secondary electron donor residue on D1 between OEC and P680; P680, the specialized chlorophyll reaction centre in PSII.

Introduction

High Arctic plants live at the limit of their distribution in an extreme environment with a short growing season, low temperature, and often nutrient limitation. Additional stress may therefore be crucial (Björn et al. 1999). Stratospheric ozone depletion increases the amount of UV-B (280–320 nm) reaching the surface of the earth (Farman et al. 1985, Webb 1997, Madronich et al. 1998), the relative increase in UV-B being largest at the higher latitudes (Björn et al. 1999). Plants in high latitude ecosystems are adapted to low UV-B levels and it is therefore expected that high latitude ecosystems are susceptible to UV-B (Caldwell et al. 1980, Robberecht et al. 1980). After many indoor studies conducted in unrealistically low background UV-A (315–400 nm) and PAR (400–700 nm) that could exaggerate UV-B response (Caldwell and Flint 1994, 1997, Caldwell et al. 1994), field experiments, in which UV-B has been increased with lamps or reduced with filters, have been published (Caldwell and Flint 1997, Caldwell et al. 1998, Paul 2001, Flint et al. 2003, Paul and Gwynn-Jones 2003). In this broad context, a growing number of studies have addressed effects on Arctic (Gehrke et al. 1996, Gwynn-Jones and Johanson 1996, Gwynn-Jones et al. 1997, Björn et al. 1999, Bredahl et al. 2004) and Antarctic ecosystems (Robson et al. 2003, Day et al. 2001, Rousseaux et al. 2001, Xiong and Day 2001). However, no one has evaluated these effects on PSII in the photosynthetic apparatus by screening leaves with chlorophyll-a fluorescence measurements, and analysing them with the so-called JIP test (Strasser et al. 2000, 2004).

Excess light can result in imbalance between the energy absorbed through the light harvesting complex and the amount of energy that can be dissipated or transduced by PSII and PSI. Such imbalance may be generated by excess light alone or may be enhanced by biotic and abiotic stress factors resulting in excess excitation energy (Bilger and Björkman 1991, Strid et al. 1996, Karpinski et al. 2001). Dissipation of excess energy is an immediate response which occurs mainly through heat dissipation (Demig-Adams and Adams 1994, Krause 1994) and detectable with the JIP test. In case of prolonged exposure to conditions causing excess light excitation, reactive oxygen species are potential photodamaging agents if the accumulation of excess light energy exceeds the capacity of antioxidant systems (Karpinski et al. 1997, 1999, 2001, Asada 1999, Mullineaux et al. 2000) and precede down-regulation of photosynthesis (Mackerness et al. 1998, Krause 1994b).

Many studies have identified PSII as the most labile component of the photosynthetic apparatus to elevated

UV-B radiation (Iwanzik et al. 1983, Strid et al. 1990, Melis et al. 1992, Vass et al. 1996, Mackerness et al. 1997, Jansen et al. 1998), but the underlying mechanisms are still a controversial subject (Andersson and Aro 2001). Both donor side mechanisms at the level of the water splitting system and/or Tyr_d and Tyr_z (Renger et al. 1989, Vass et al. 1996, Larkum et al. 2001) and acceptor side mechanisms have been reported (Jansen et al. 1996, 1998). From simultaneous monitoring of both donor and acceptor mechanisms, it could be concluded that UV-B light impairs both electron transfer from the Mn water oxidation cluster to Tyr_z⁺ and P680⁺ as well as acceptor side modifications, particularly in the Q_B pocket of the D1 protein (Vass et al. 1999). Since the full kinetics from O-step to P-step include contributions of several reaction mechanisms (Strasser et al. 2004), then the analysis of derived parameters, such as the overall grouping probability or energetic co-operativity (Stirbet et al. 1998), approximated fraction of non-Q_B reducing units which relates to heterogeneity concerning the Q_A–Q_B binding (see discussion by Strasser et al. 2004), but also the step at 300 μs (K-band) related to the oxygen-evolving complex (OEC) (see discussion by Strasser et al. 2004), which are related to donor and acceptor side of PSII, it is possible that extensive analysis and/or application of the JIP test may contribute to this subject.

This study focuses on PAR and UV-B effects on the light processing through PSII in *Salix arctica* leaves in the high arctic. The experimental approach chosen was to reduce the UV-B irradiance by means of filters. In an earlier study (Bredahl et al. 2004), a large variation in F_v/F_m measurements was observed, which was partly supposed to be associated with the large variation in leaf orientation towards the sun. Thus, we conducted an experiment where the leaves were fixed in a position exposing them to maximum solar irradiance. The processing of light through PSII was analysed by means of recording transients of chlorophyll a fluorescence, and analysing it using the JIP test and associated parameters (Krause and Weis 1991, Strasser and Strasser 1995, Strasser et al. 1995, Krüger et al. 1997, Strasser et al. 2000, 2004), a methodology discussed by Lazár (1999) and recently by Force et al. (2003).

Materials and methods

Experimental site

The fieldwork was carried out in a high arctic heathland at Zackenberg Research Station, North-east Greenland (74°eN, 21°E) in July and August of 2002.

The plant species investigated was the abundantly occurring *Salix arctica* Pall. Lge. (Böcher et al. 1978), having a robust stem and broad leaves well suited for the manipulations and measurements.

Experimental set-up and treatments

Experimental plots were established where parts of the UV-irradiance in natural daylight were reduced with ambient UV-B irradiance as reference. Reduction was achieved by filtering the solar radiation through a filter, Mylar® (type D; DuPont Teijin Films, Wilmington, DE). Mylar filter transmits $\lambda > 320$ nm (Cybulski and Peterjohn 1999). As control, a Teflon® filter (Fluoretek AB, Knivsta, Sweden.) was used. None of these filters have phytotoxic effects, as reported for cellulose diacetate (Krizek and Mirecki 2004). Teflon transmits $\lambda > 280$ nm (Cybulski and Peterjohn 1999). Measurements in the experimental area with a broad-band cosine corrected UV-B sensor (UV-S-310-T, Scintec; Atmosphärenmesstechnik GmbH, Tübingen, Germany – now manufactured as UV-S-B-T from Kipp & Zonen B.V., Delft, The Netherlands) showed that the plant canopy under Teflon and Mylar was exposed to approximately 91 and 39% of the clear sky UV-B irradiance, respectively, slightly depending on the exposure angle to the sun (Bredahl et al. 2004). In the following, the Teflon cover is referred to as 'ambient UV-B' and Mylar cover as 'reduced UV-B'.

Nylon strings were used to fix plant-shoots and leaves, to rectangular aluminium frames. Each frame was forced into the soil just in front of a *Salix* plant base. Then one long (10–20 cm) shoot was carefully fixed in a way so that all leaf surfaces were exposed in an angle of 45° facing south. Then the filter sheets were fastened to the frames with clamps. This set-up was used to expose the plants to maximum PAR and UV-B and allowed easy access to measurements.

The experiment was designed as a randomized design where each treatment consisted of 20 shoots from different plant individuals, in total 40 shoots.

Light regime

Climatic data were recorded at two nearby experimental sites and at the permanent climate station on Zackenberg throughout the season. PAR was measured with one sensor placed horizontally and maximum PAR by another sensor inclined at 45° to the south (GaAsP photodiode, Ponttailler 1990). UV-B was measured by means of a horizontally mounted broad-band cosine-corrected sensor (UV-S-B-T/C; Kipp & Zonen, Delft, The Netherlands). The sensors were connected to

dataloggers (CR 10X; Campbell Scientific, Ltd, Loughborough, UK) and hourly means of UV-B and PAR were recorded as the average of per minute recordings.

Chlorophyll a fluorescence

During July 6 to August 20, 2002, transients of chlorophyll *a* fluorescence were measured approximately every third day at 1430 h on *Salix arctica*. On 1, 6 and 13 August measurements were conducted at 0900, 1400 and 1800 h to investigate daily variations. After removal of the filter cover, a dark adaptation clip was mounted for 35–40 min on one of the exposed leaves to ensure that all PSII were in the dark-adapted state with open RC. The mounting of clips on all plants took less than 10 min. Then chlorophyll *a* fluorescence transients were recorded and digitized with a portable Handy PEA (Hansatech Instruments, Ltd. King's Lynn Norfolk, UK) with a 12-bit resolution from 10 μ s to 1 s and a time resolution of 10 μ s for the first 200 data points (Strasser et al. 1995). The excitation light intensity was 600 W m⁻² from an array of six red light-emitting diodes with a peak wavelength at 650 nm focused on the leaf surface in the clips on a spot of 4 mm in diameter. The chlorophyll *a* fluorescence signals were detected by the Handy Pea using a PIN photocell after having passed through a long pass filter (50% transmission at 720 nm) (Srivastava et al. 1997). The measurements were always done in the same order, alternating between different treatments. Initial measurements were done on 6 July 2002, on the plants after their fixation, but before attachment of filters. Afterwards, the filters were permanently attached and removed only when the measurements were conducted.

Analysis of fluorescence: the JIP test

From the fluorescence transient, the measured parameters (F_0 , F_M , tF_M , Area, $F_{50\mu s}$, $F_{100\mu s}$, $F_{300\mu s}$, F_J , F_I) led to calculation and derivation of a range of new parameters according to Strasser et al. (2000, 2004) (see Table 1). Careful translation of the measured parameters into JIP test parameters provides information of the stepwise flow of energy through PSII at different levels: (1) specific fluxes on the level per reaction centre (RC) and these are for absorption (ABS/RC), trapping (TR_0/RC), dissipation (DI_0/RC) and electron transport (ET_0/RC); and (2) phenomenological fluxes on the level of the excited sample cross-section (CS) and these are for absorption (ABS/CS), trapping (TR_0/CS), dissipation (DI_0/CS) and electron transport (ET_0/CS) (Table 1). By assuming that F_0 and F_M are reasonable measures of

Table 1. Summary of parameters, formulae and their description using data extracted from the fast fluorescence transient O-J-I-P. ABS, absorption energy flux; CS, excited cross-section of leaf sample; DI, dissipation energy flux at the level of the antenna chlorophylls; ET, flux of electrons from Q_A^- into the intersystem electron transport chain; ϕ_{PO} maximal quantum yield of primary photochemistry; ψ_o , efficiency by which a trapped exciton, having triggered the reduction of Q_A to Q_A^- , can move an electron further than Q_A^- into the electron transport chain; RC, PSII reaction centre; RC/CS, concentration of reaction centres per excited cross-section of leaf sample; TR, excitation energy flux trapped by a RC and utilized for the reduction of Q_A to Q_A^- ; PI, performance index. Note the distinction between measured and derived parameters. Adapted and reorganized from Strasser et al. (2000) and Strasser et al. (2004).

Extracted and technical fluorescence parameters

Measured parameters

F_0	Initial fluorescence intensity
F_M	Maximal fluorescence intensity
$F_{50\mu s}$	Fluorescence intensity at 50 μs
$F_{100\mu s}$	Fluorescence intensity at 100 μs
$F_{300\mu s}$	Fluorescence intensity at 300 μs
F_J	Fluorescence intensity at the J-step (at 2 ms)
F_I	Fluorescence intensity at the I-step (at 30 ms)
Area	Area between fluorescence curve and F_M
tFm	Time to reach F_M in ms

Derived parameters

Selected J-I-P-parameters

F_V	$(F_M - F_0)$
F_V/F_M	$(F_M - F_0)/F_M$
V_J	$(F_J - F_0)/(F_M - F_0)$
V_I	$(F_I - F_0)/(F_M - F_0)$
S_m	Area/($F_M - F_0$)
N	$S_m * (M_0/V_J)$
$(dV/dt)_0 = M_0$	$4 * (F_{300\mu s} - F_0)/(F_M - F_0)$
Yield P_j	$1 - (F_J/F_M)$
Yield P_i	$1 - (F_I/F_M)$
Yield P_o	F_V/F_M
F_J/F_0	F_J/F_0
F_I/F_0	F_I/F_0
F_M/F_0	F_M/F_0
Quantum efficiencies	
$\phi_{PO} = TR_0/ABS$	F_V/F_M
$\phi_{EO} = ET_0/ABS$	$F_V/F_M * (1 - V_J)$
$\phi_o = ET_0/TR_0$	$(1 - V_J)$
Specific fluxes	
ABS/RC	$M_0 * (1/V_J) * (1/\phi_{PO})$
TR_0/RC	$M_0 * (1/V_J)$
ET_0/RC	$M_0 * (1/V_J) * \phi_o$
DI_0/RC	$(ABS/RC) - (TR_0/RC)$
Phenomenological fluxes	
ABS/CS	$ABS/CS_{chl} = chl/CS$ or $ABS/CS_0 = F_0$ or $ABS/CS_M = F_M$
TR_0/CS_0	$ABS/CS_0 * \phi_{PO}$
ET_0/CS_0	$ABS/CS_0 * \phi_{PO} * \phi_o$
DI_0/CS_0	$(ABS/CS_0) - (TR_0/CS_0)$
TR_M/CS_M	$ABS/CS_M * \phi_{PO}$
ET_M/CS_M	$ABS/CS_M * \phi_{PO} * \phi_o$
DI_M/CS_M	$(ABS/CS_M) - (TR_0/CS_0)$
Density of PSII reaction centre	
RC/CS_0	$\phi_{PO} * (V_J/M_0) * ABS/CS_0$
RC/CS_M	$\phi_{PO} * (V_J/M_0) * ABS/CS_M$
Performance indexes	
PI_{ABS}	$(RC/ABS) * [\phi_{PO}/(1 - \phi_{PO})] * [\phi_o/(1 - \phi_o)]$
PI_{CS_0}	$(ABS/CS_0) * [\phi_{PO}/(1 - \phi_{PO})] * [\phi_o/(1 - \phi_o)]$
PI_{CS_M}	$(ABS/CS_M) * [\phi_{PO}/(1 - \phi_{PO})] * [\phi_o/(1 - \phi_o)]$

the absorption energy flux per excited cross-section of leaf sample (ABS/CS) in arbitrary units of a particular leaf sample in the dark-adapted state, phenomenological fluxes were estimated from both F_0 and F_M , respectively (Table 1). The specific and phenomenological fluxes are interrelated by the quantum efficiencies, which are (1) the maximum quantum yield of primary photochemistry (F_V/F_M) which in this terminology is equal to the efficiency by which an absorbed photon will be trapped by the PSII RC with the resultant reduction of Q_A to Q_A^- (TR_0/ABS); and (2) the efficiency by which a trapped exciton, having triggered the reduction of Q_A to Q_A^- can move an electron further than Q_A^- into the intersystem electron transport chain (ET_0/TR_0) (Table 1). Integrative parameters, so called performance indexes reflecting performance of the overall energy flow processing are (1) based on quantum efficiencies and hereby related to the situation of assuming equal absorption (PI_{ABS}); and (2) based on the phenomenological fluxes related to the per cross-section level (PI_{CS_0} and PI_{CS_M}) (Table 1). Estimation of the density of active PSII reaction centres per cross-section (RS/CS_0 And RC/CS_M) is also possible. See Table 1 for a full summary of all parameters, formulae, definition of symbols and Fig. 8 for pipeline models depicting the stepwise energy flow through PSII. Brief parameter descriptions are given in the results section the first time a parameter occur (see below). Although rich in information, the interpretation of JIP test parameters describing the quantitative behaviour of the PSII must be done carefully, due to both the biophysical and derived nature of these parameters.

Leaves

After the last measurements all leaves were harvested in order to determine biomass, leaf area, and the content of flavonoids, C, and N. Soil samples were analysed for C, N, and water content. Immediately after the end harvest of the season on 20 August 2002, digital photographs were taken of all the filter-covered leaves beside a brick of known area. The photographs were loaded in a pixel counting program (Bitmap, S. Danbæk, Biological Institute, University of Copenhagen) and the leaf areas were determined. The dry weight was determined after oven drying for 48–62 h at 80°C.

Total soluble flavonoids were extracted by a three step procedure. First, the pulverized leaves were heated (60°C) in 5 ml methanol for 3 min; second, after adding 5 ml HCl–H₂O–methanol (1 : 20 : 79), leaves were shaken for 10 min; third, the supernatant was diluted 10 fold in HCl–H₂O–methanol (1 : 20 : 179) after

centrifugation at approximately 3 g for 10 min (Caldwell 1968). After additional 10 min shaking, the UV-absorption was measured with a spectrophotometer (U-2010; Hitachi, Tokyo, Japan) from 280 to 315 nm at 0.2 nm intervals. The area below the absorption curve was calculated as the sum of all absorptions in the scanned range. Further, total C and N were determined on a Leco truespecTM. CN elemental determinator (Leco Corporation, St. Joseph).

Soil

At the end of the season three soil samples were taken at two depths (5 and 10 cm). Fresh weight was noted before the samples were oven dried at 80°C for 62–86 h, and then at the end dry weights were recorded. Total carbon (using Eltra CS-500, Neuss, Germany) and total nitrogen (using Leco FP-428 nitrogen determinator, Leco Corporation) were measured.

Statistical analysis

Statistical analyses were conducted using the General Linear Model (GLM) procedure (Statistical Analysis Systems Institute 1999). Levene's test was used to test for homogeneity of variance. Where necessary, parameters were transformed in order to meet the assumptions of ANOVA. Fluorescence variables were tested with a repeated measurement ANOVA, with treatment as factor and day repeated within each plot. This was to take into account that the same experimental unit (the leaves on the same shoot) was measured several times during the season. The treatment effects were tested separately on each day of measurement. Seasonal effect of treatment was tested across all measurements during the season. Furthermore, the daily variation in fluorescence parameters was tested for treatment and time effects with GLM Anova repeated measurements contrast option. Contrast variables were set to the three time levels (0900, 1400 and 1800 h) and the contrast level was set to 2 (at 1400 noon). Differences were considered at the $P < 0.05$ level. Leaf biomass, specific leaf area (SLA), flavonoid content, C, N, and C/N-ratio were tested for effects of treatment by one-way ANOVA. In cases of significant treatment effects, these analyses were followed by tests of treatment differences using Tukey test. All values presented here are non-transformed. The fluctuations in photosynthetic active radiation (PAR), Max PAR, UV-B, and Max UV-B during the whole season were tested for time effects by one-way ANOVA followed by Tukey test. Pearson correlations between PAR, Max PAR, UV-B, and Max UV-B were calculated.

Results

Climate

Irradiance data from the measurement days are included for analysis. Fluctuations in PAR, Max PAR and UV-B during the season did show some periods with different covariance between PAR and UV-B irradiance, but these were not significant (data not shown). After a period with high PAR and UV-B irradiance until 12 July, a gradual decrease was seen in PAR, Max PAR and UV-B throughout the rest of the season; there were also days with very low irradiance: 24 July, 26 July and 9 August (Fig. 1). In general, coinciding high PAR and high UV-B irradiance were observed, each being at its peak around 1400 h (data not shown). The days (24 and 26 July and 9 August) with very low irradiances were not significantly different, but deviated with respect to Tukey grouping (data not shown). Clear sky conditions with no significant difference between light variables were observed on 1, 6 and 13 August, enabling analysis of daily variation in fluorescence parameters (see below). The averages of irradiance of PAR for these three days were, at 0900 h, 255; at 1400 h, 597; and at 1800 h, 399 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$. The similar averages for UV-B were 0.167, 0.543 and 0.353 Watt m^{-2} , respectively. The Pearson correlations between PAR and UV-B ($n = 408$) were highly significant. In July, the mean air temperature was 6.7°C and, in August it was 5.6°C.

Leaves

A summary of the statistics on leaf variables shown in Table 2. The leaf biomass and leaf area were lower in ambient UV-B, but not significantly ($P < 0.2387$ and $P < 0.4866$). Furthermore, the higher specific leaf area

(SLA) in ambient UV-B was not significant ($P < 0.2238$). The flavonoid content was significantly higher in ambient UV-B ($P < 0.0001$), while no significant differences were found for C, N, and C/N ratio.

Soil

Samples taken from a depth of 5 cm had a 24% mean water content, a mean C and N content of 5.4 and 0.25%, and a mean C/N ratio of 21.5. Similarly, the averages in 10 cm depth had a 23% mean water content, C and N content 3.5 and 0.2%, and a C/N ratio of 17.7.

Chlorophyll a fluorescence

Seasonal patterns of selected fluorescence variables are shown in Figs 2–7. Significant effects of treatments were seen in July, but these effects disappeared in August. Summary statistics of July means of all fluorescence variables can be seen in Table 3, and the summary statistics for the daily variations are shown in Table 4.

After the initial measurement on 6 July, the derived parameters (Strasser et al. 2000, 2004), TR_0/ABS (efficiency by which an absorbed photon will be trapped by the PSII reaction centre, RC, with the resultant reduction of Q_A), ET_0/TR_0 (efficiency by which an electron residing on Q_A^- will enter the intersystem electron transport chain), and ET_0/ABS (quantum yield for electron transport), were significantly reduced throughout July in ambient UV-B ($P < 0.0041$, $P < 0.0038$, $P < 0.0016$), but from mid-August, the treatment effects disappeared (Table 3, Fig. 2). Quantum efficiencies increased until the season peak on 24 July, and then decreased (Table 3, Fig. 2). The derived specific fluxes (Strasser et al. 2000, 2004) per active PSII reaction

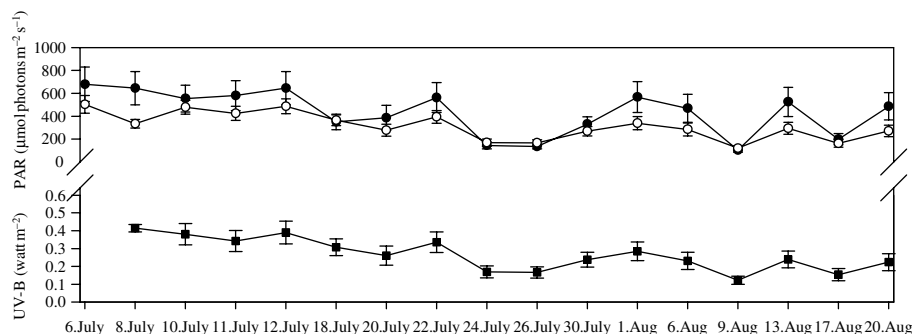


Fig. 1. PAR and maximum PAR (upper) and UV-B (lower). The curves are daily means from the days of measurements, calculated as the hours within the day, and logged by continuous measurements of PAR ($\mu\text{mol photons m}^{-2} \text{s}^{-1}$) with one horizontal placed sensor (PAR, depicted as open circles) and a second sloping 45° facing south (maximum PAR, depicted as closed circles) while UV-B (Watt m^{-2}) was continuously logged from a horizontally placed sensor (UV-B, depicted as squares). UV-B data are not available before 8 July 2002.

Table 2. Measured leaf variables. The numbers are end season means ± 1 SE tested with General linear Model (GLM) one-way ANOVA followed by Tukey test. Significant effects of treatment are shown by different letters. The resulting values of individual *F*-test and probability of correct null hypothesis are given.

Variable		Reduced UV-B	Ambient UV-B	<i>F</i> value	Prob <i>F</i>
Average leaf biomass	g	0.032 ^A \pm 0.003	0.026 ^A \pm 0.003	1.43	0.2387
SLA	cm ² g ⁻¹	8.04 ^A \pm 0.62	10.55 ^A \pm 1.93	1.53	0.2238
Flavonoid	arbitrary	176.85 ^B \pm 0.58	184.84 ^A \pm 0.57	97.29	0.0001
C	%	45.14 ^A \pm 0.23	44.93 ^A \pm 0.24	0.41	0.5273
N	%	1.25 ^A \pm 0.08	1.35 ^A \pm 0.08	0.87	0.3568
C/N ratio	ratio	40.54 ^A \pm 3.98	36.60 ^A \pm 3.46	0.55	0.4517

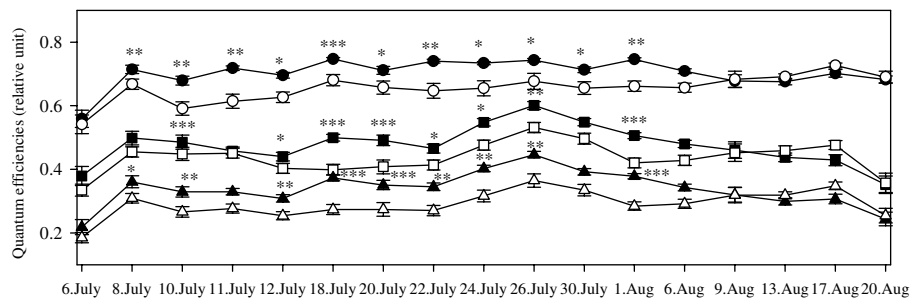


Fig. 2. Quantum efficiencies (ratios between 0 and 1.00). Closed symbols represent reduced UV-B and open symbols ambient UV-B. Curves are daily means \pm SE during the whole season tested with General linear Model (GLM) repeated measurements ANOVA ($\alpha = 0.05$). Levels of significance ****P* < 0.0010, ***P* < 0.0100, and **P* < 0.10. The efficiency by which an absorbed photon will be trapped by the PSII RC with the resultant reduction of Q_A (TR₀ ABS⁻¹) is depicted by circles, the efficiency by which an electron residing on Q_A⁻ will enter the intermediate electron transport chain between PSII and PSI (ET₀ TR₀⁻¹) is depicted by squares and the quantum yield for electron transport (ET₀ ABS⁻¹) is depicted as triangles. Across all measurements during season treatment difference were tested and revealed that TR₀ ABS⁻¹ (*P* < 0.0329), ET₀ TR₀⁻¹ (*P* < 0.1890), and ET₀ ABS⁻¹ (*P* < 0.0746). For meaning of symbols, see Table 1.

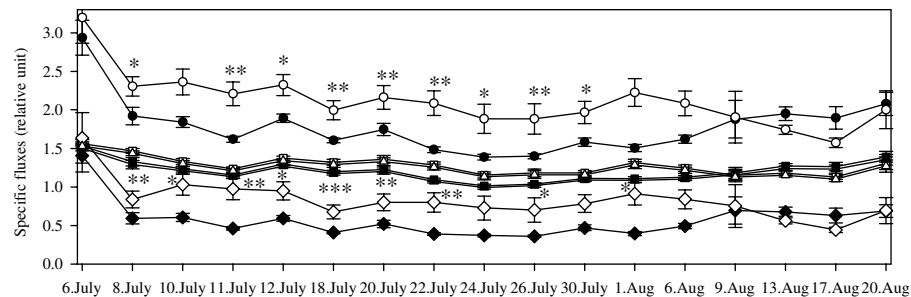


Fig. 3. Specific fluxes per PSII reaction centre (relative unit). Closed symbols represent reduced UV-B and open symbols ambient UV-B. Curves are daily means \pm SE during the whole season tested with GLM repeated measurements ANOVA ($\alpha = 0.05$). Levels of significance ****P* < 0.0010, ***P* < 0.0100, and **P* < 0.10. Variables are per active PSII RC. Absorption flux of photons (ABS RC⁻¹) is depicted by circles, the maximal trapping (TR₀ RC⁻¹) is depicted by squares, the electron transport (ET₀ RC⁻¹) is depicted as triangles, and the dissipation of untrapped excitons (DI₀ RC⁻¹) is depicted as diamonds. Across all measurements during season treatment difference were tested and revealed that ABS RC⁻¹ (*P* < 0.0934), TR₀ RC⁻¹ (*P* < 0.1367), ET₀ RC⁻¹ (*P* < 0.1350), and DI₀ RC⁻¹ (*P* < 0.0978). For meaning of symbols, see Table 1.

centre, RC, ABS/RC (absorption flux of photons per active RC), TR₀/RC (maximum trapping rate by which an exciton is trapped by the PSII RC resulting in the reduction of Q_A), DI₀/RC (effective dissipated flux of untrapped excitons per active PSII RC), and ET₀/RC (electron transport to intersystem chain per active RC)

were all significantly higher in July in ambient UV-B (*P* < 0.0091, *P* < 0.0024, *P* < 0.0153, *P* < 0.0023) (Table 3, Fig. 3).

The derived phenomenological fluxes per excited cross-section of leaf sample, estimated from *F*₀, were to some degree affected by treatment. ABS/CS₀ (photons

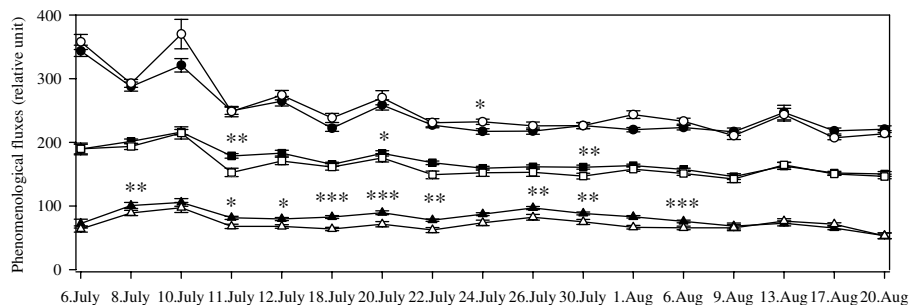


Fig. 4. Phenomenological fluxes per leaf cross-section estimated from F_0 (relative unit). Closed symbols represent reduced UV-B and open symbols ambient UV-B. Curves are daily means \pm SE error during the whole season tested with GLM repeated measurements ANOVA ($\alpha = 0.05$). Levels of significance $***P < 0.0010$, $**P < 0.0100$, and $*P < 0.10$. Variables are per cross-section of leaf sample that contains active and inactive PSII RCs. Photons absorbed, at F_0 , by antenna molecules associated with all PSII RCs per cross-section of the sample ($ABS\ CS_0^{-1}$) is depicted by circles, the trapping rate of an exciton that will lead to Q_A reduction ($TR_0\ CS_0^{-1}$) is depicted by squares, the electron transport as re-oxidation of reduced Q_A via electron transport ($ET_0\ CS_0^{-1}$) is depicted as triangles. Across all measurements during season treatment difference were tested and revealed that $ABS\ CS_0^{-1}$ ($P < 0.5153$), $TR_0\ CS_0^{-1}$ ($P < 0.2121$), and $ET_0\ CS_0^{-1}$ ($P < 0.0544$). For meaning of symbols, see Table 1.

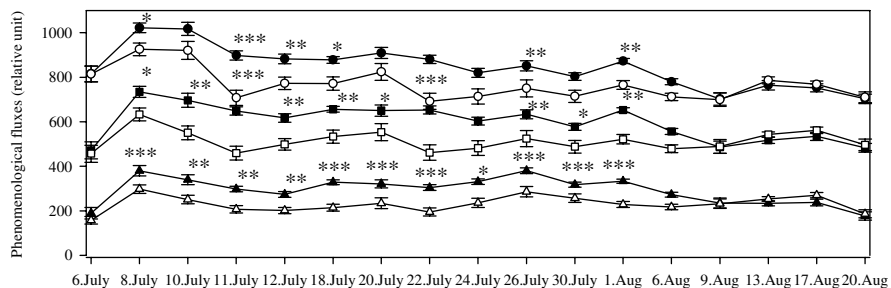


Fig. 5. Phenomenological fluxes per leaf cross-section estimated from F_M (relative unit). Closed symbols represent reduced UV-B and open symbols ambient UV-B. Curves are daily means \pm SE during the whole season tested with GLM repeated measurements ANOVA ($\alpha = 0.05$). Levels of significance $***P < 0.0010$, $**P < 0.0100$, and $*P < 0.10$. Variables are per cross-section of leaf sample that contains active and inactive PSII RCs. Photons absorbed, at F_M , by antenna molecules associated with all PSII RCs per cross-section of the sample ($ABS\ CS_M^{-1}$) is depicted by circles, the trapping rate of an exciton that will lead to Q_A reduction ($TR_M\ CS_M^{-1}$) is depicted by squares, the electron transport as re-oxidation of reduced Q_A via electron transport ($ET_M\ CS_M^{-1}$) is depicted as triangles. Across all measurements during season treatment difference were tested and revealed that $ABS\ CS_M^{-1}$ ($P < 0.0124$), $TR_M\ CS_M^{-1}$ ($P < 0.0082$), and $ET_M\ CS_M^{-1}$ ($P < 0.0185$). For meaning of symbols, see Table 1.

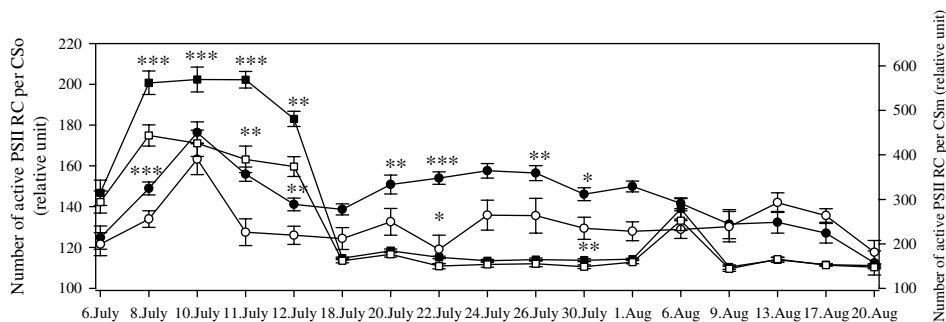


Fig. 6. Number of active PSII reaction centre per cross-section (relative unit). Closed symbols represent reduced UV-B and open symbols ambient UV-B. Curves are daily means \pm SE during the whole season tested with GLM repeated measurements ANOVA ($\alpha = 0.05$). Levels of significance $***P < 0.0010$, $**P < 0.0100$, and $*P < 0.10$. Number of active PSII RCs estimated from F_0 ($RC\ CS_0^{-1}$) depicted as circles and number of active PSII RCs estimated from F_M ($RC\ CS_M^{-1}$) depicted as squares. Across all measurements during season treatment difference were tested and revealed that $RC\ CS_0^{-1}$ ($P < 0.0780$) and $RC\ CS_M^{-1}$ ($P < 0.0191$). For meaning of symbols, see Table 1.

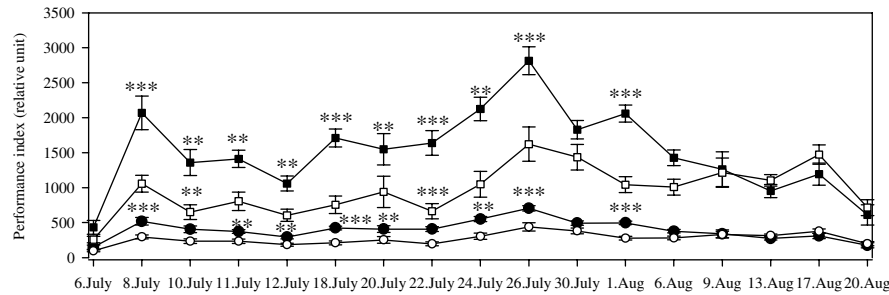


Fig. 7. Performance indexes based per leaf cross-section (relative unit). Closed symbols represent reduced UV-B and open symbols ambient UV-B. Curves are daily means \pm SE during the whole season tested with GLM repeated measurements ANOVA ($\alpha = 0.05$). Levels of significance *** $P < 0.0010$, ** $P < 0.0100$, and * $P < 0.10$. Performance index estimated from M (Pl_{CS_0}) depicted as circles and performance index estimated from M (Pl_{CS_M}) depicted as squares. Across all measurements during season treatment difference were tested and revealed that Pl_{CS_0} ($P < 0.0273$) and Pl_{CS_M} ($P < 0.0192$). For meaning of symbols, see Table 1.

absorbed, at F_0 , by antenna molecules associated with all PSII RCs per cross-section of the leaf) tended to increase in ambient UV-B ($P < 0.0773$), but TR_0/CS_0 (maximal trapping rate of excitons that will lead to Q_A reduction per cross-section of the sample) tended to decrease ($P < 0.0708$) (Table 3, Fig. 4). The ET_0/CS_0 (maximal electron transport rate per cross-section of sample) was significantly lower in July in ambient UV-B ($P < 0.0015$), but in mid-August, the effect of treatment disappeared (Table 3, Fig. 4). Phenomenological fluxes per cross-section of the sample, estimated from F_M , ABS/CS_M (photons absorbed, at F_M , by antenna molecules associated with all PSII RCs per cross-section of the sample), TR_M/CS_M (trapping rate of excitons that will lead to Q_A reduction per cross-section of the sample), and ET_M/CS_M (electron transport rate per cross-section of the sample) significantly decreased in July in ambient UV-B ($P < 0.0019$, $P < 0.0008$ and $P < 0.0003$) (Table 3, Fig. 5). Both treatments showed the same seasonal pattern with an increase from the same initial level on 6 July 2002, to two significantly different levels on 8 July (Fig. 5). Hereafter, there was a continuous effect of treatment until 9 August, when the treatment effect disappeared and the phenomenological fluxes decreased to the same low level (Fig. 5).

The estimated number of active PSII reaction centres per sample cross-section using F_0 and F_M , RC/CS_0 and RC/CS_M , were significantly decreased in July in ambient UV-B ($P < 0.0034$ and $P < 0.0024$) (Table 3 and Fig. 6). For both treatments, RC/CS_0 increased from the same initial level to a maximum on 10 July 2002. Then, the RC/CS_0 significantly decreased in ambient UV-B until 9 August, but then increased to a higher level (Fig. 6). RC/CS_M showed more or less the same pattern as RC/CS_0 during the season, but the treatment difference was most pronounced in the beginning of the

season, until 18 July (Fig. 6). After 18 July, the RC/CS_M was reduced to approximately 75% of the level on 8, 10 and 11 July. Hereafter, the RC/CS_M was marginally higher in ambient UV-B, but after 1 August, there was no effect of treatment (Fig. 6).

The parameters labelled as performance indexes (see Table 1) Pl_{CS_0} , Pl_{CS_M} , and Pl_{ABS} were all significantly decreased in ambient UV-B in July ($P < 0.0003$, $P < 0.0002$ and $P < 0.0008$) (Table 3). They all showed the same seasonal variation with an increase from the same initial level to a season peak on 26 July 2002 (Fig. 7; curve for Pl_{ABS} is not shown). Then, there was a simultaneous decrease until 9 August, where the effect of treatment disappears and performance indexes are higher for plants in ambient UV-B (Fig. 7).

The means in July for the yields at the J-I-P steps of the fluorescence transient (Yield P_j , Yield P_i and Yield P_0) significantly decreased in ambient UV-B ($P < 0.0044$, $P < 0.0016$ and $P < 0.0041$) (Table 3). They all showed the same seasonal pattern with an increase from the initial level, where no treatment difference appeared, to a period with effects of treatment on almost every day of measurements, to a season peak around 26 July 2002. Then the yields decreased and the effects of treatment disappeared after 9 August (data not shown). The ratios of fluorescence values at the J-I-P steps to those of F_0 (F_j/F_0 , F_i/F_0 and F_M/F_0) decreased in ambient UV-B and during July a significant effect of treatment was seen ($P < 0.0118$, $P < 0.0012$, and $P < 0.0007$). During the season they showed the same fluctuations as the yields at the J-I-P steps (data not shown). There was no effect of treatment in July on V_i (variable fluorescence at step I) and N (number of Q_A reduction turnover from time zero to tF_M) ($P < 0.1730$ and $P < 0.1174$), but significant effects were seen on V_j (variable fluorescence at J step), Area (area over fluorescence induction curve), tF_M (time to reach F_M in ms), M_0 (net rate of PSII closure,

Table 3. Measured fluorescence variables. The numbers are July means ± 1 SE tested with GLM ANOVA repeated measurements ($n = 20$ in each treatment and $\alpha = 0.05$). Values of F-test and probability of correct null hypothesis are given.

	Ambient UV-B	Reduced UV-B	Fvalue	ProbF
<i>Measured parameters</i>				
F ₀	263.5 \pm 3.3	251.3 \pm 2.7	3.33	0.0773
F _M	779.5 \pm 9.8	888.8 \pm 7.0	11.44	0.0019
F _{50μs}	279.6 \pm 3.5	267.5 \pm 2.8	3.20	0.0831
F _{100μs}	295.0 \pm 3.7	283.1 \pm 3.0	2.77	0.1055
F _{300μs}	355.8 \pm 4.4	347.1 \pm 3.8	1.17	0.2873
F _J	547.1 \pm 6.5	570.8 \pm 5.3	1.00	0.3243
F _I	647.4 \pm 7.5	711.0 \pm 5.7	6.86	0.0134
Area	20937.3 \pm 594.0	28425.8 \pm 647.7	11.36	0.0020
tFm	618.7 \pm 16.1	699.0 \pm 17.3	7.34	0.0107
<i>Derived parameters</i>				
Selected J-I-P parameters				
F _V	516.0 \pm 9.2	637.4 \pm 6.6	13.19	0.001
F _V /F _M	0.6423 \pm 0.0062	0.7129 \pm 0.0037	9.55	0.0041
V _J	0.5621 \pm 0.0056	0.5071 \pm 0.0052	9.75	0.0038
V _I	0.7504 \pm 0.0031	0.7227 \pm 0.0032	6.31	0.1730
S _m	39.9 \pm 0.8	44.6 \pm 1.1	4.56	0.0404
N	50.9 \pm 0.9	51.9 \pm 1.2	2.59	0.1174
M ₀	0.7539 \pm 0.0138	0.6140 \pm 0.0109	11.26	0.0021
Yield P _J	0.1623 \pm 0.0029	0.1985 \pm 0.0027	9.37	0.0044
Yield P _I	0.2864 \pm 0.0052	0.3545 \pm 0.0046	11.96	0.0016
Yield P ₀	0.6424 \pm 0.0062	0.7124 \pm 0.0037	9.55	0.0041
F _J /F ₀	2.106 \pm 0.020	2.299 \pm 0.016	7.14	0.0118
F _V /F ₀	2.515 \pm 0.030	2.887 \pm 0.024	12.57	0.0012
F _M /F ₀	3.042 \pm 0.042	3.625 \pm 0.034	13.98	0.0007
Quantum efficiencies				
$\phi_{P_0} = TR_0/ABS$	0.6423 \pm 0.0062	0.7129 \pm 0.0037	9.55	0.0041
$\phi_{E_0} = ET_0/ABS$	0.2864 \pm 0.0052	0.3545 \pm 0.0046	11.96	0.0016
$\phi_0 = ET_0/TR_0$	0.4379 \pm 0.0056	0.4929 \pm 0.0052	9.75	0.0038
Specific fluxes				
ABS/RC	2.215 \pm 0.054	1.719 \pm 0.029	7.70	0.0091
TR ₀ /RC	1.315 \pm 0.013	1.192 \pm 0.011	10.92	0.0024
ET ₀ /RC	1.287 \pm 0.013	1.166 \pm 0.011	10.93	0.0023
DI ₀ /RC	0.901 \pm 0.045	0.528 \pm 0.020	6.57	0.0153
Phenomenological fluxes				
ABS/CS ₀	263.5 \pm 3.3	251.3 \pm 2.7	3.33	0.0773
TR ₀ /CS ₀	166.6 \pm 2.0	176.7 \pm 1.4	3.49	0.0708
ET ₀ /CS ₀	72.3 \pm 1.3	86.6 \pm 1.1	12.12	0.0015
DI ₀ /CS ₀	96.9 \pm 2.6	74.6 \pm 1.8	9.03	0.0051
ABS/CS _M	779.5 \pm 9.8	888.8 \pm 7.0	11.44	0.0019
TR _M /CS _M	516.0 \pm 9.2	637.4 \pm 6.6	13.76	0.0008
ET _M /CS _M	232.4 \pm 5.6	318.0 \pm 5.1	16.95	0.0003
DI _M /CS _M	263.5 \pm 3.3	251.3 \pm 2.7	3.78	0.0608
Density of PSII reaction centres				
RC/CS ₀	130.6 \pm 1.8	149.6 \pm 1.2	10.00	0.0034
RC/CS _M	261.7 \pm 8.4	321.2 \pm 10.6	10.89	0.0024
Performance indexes				
PI _{ABS}	1.11 \pm 0.05	1.88 \pm 0.06	13.70	0.0008
PI _{CS₀}	264.9 \pm 10.7	438.4 \pm 12.2	16.94	0.0003
PI _{CS_M}	938.1 \pm 45.7	1693.8 \pm 54.8	18.35	0.0002

Table 4. Measured fluorescence variables. Numbers are daily means \pm 1 SE tested with GLM ANOVA repeated measurements contrast option ($n = 20$ in each treatment and $\alpha = 0.05$). The contrast P statistics is given for each variable. Only measurements from 1 August and 6 August at 0900, 1400 and 1800 h are included.

	0900	1400	1800	Contrast 0900–1400	Contrast 1400–1800
<i>Measured parameters</i>					
F_0	237.0 \pm 3.3	223.1 \pm 3.0	244.5 \pm 7.6	0.0002	0.0147
F_M	802.1 \pm 11.2	742.0 \pm 12.0	796.3 \pm 13.7	0.00001	0.00001
$F_{50\mu s}$	252.5 \pm 3.5	237.8 \pm 3.2	258.2 \pm 7.7	0.0002	0.0210
$F_{100\mu s}$	267.0 \pm 3.9	251.8 \pm 3.4	270.9 \pm 7.8	0.0004	0.0340
$F_{300\mu s}$	327.3 \pm 5.0	309.5 \pm 4.4	326.0 \pm 8.2	0.0009	0.0751
F_J	529.5 \pm 6.6	503.6 \pm 7.2	537.6 \pm 8.9	0.0011	0.0007
F_I	653.9 \pm 7.4	615.8 \pm 8.4	679.1 \pm 11.2	0.00001	0.00001
Area	21584.6 \pm 793.1	19715.5 \pm 704.5	25644.8 \pm 1038.2	0.0022	0.00001
tFm	537.2 \pm 18.3	528.2 \pm 14.6	896.6 \pm 20.1	0.4873	0.00001
<i>Derived parameters</i>					
Selected J-I-P parameters					
F_V	565.1 \pm 11.8	518.9 \pm 11.7	551.8 \pm 11.9	0.00001	0.0008
F_V/F_M	0.6942 \pm 0.0083	0.6880 \pm 0.0078	0.6880 \pm 0.0089	0.2611	0.9613
V_J	0.5323 \pm 0.0101	0.5568 \pm 0.0096	0.5455 \pm 0.0100	0.0030	0.2126
V_I	0.7458 \pm 0.0062	0.7671 \pm 0.0059	0.7945 \pm 0.0064	0.00001	0.00001
S_m	37.3 \pm 1.0	36.9 \pm 0.09	44.9 \pm 1.2	0.508	0.00001
N	44.6 \pm 0.9	44.5 \pm 0.8	49.1 \pm 1.3	0.7619	0.0002
M_0	0.6848 \pm 0.0275	0.7097 \pm 0.0236	0.6388 \pm 0.0280	0.128	0.0044
Yield P_J	0.1788 \pm 0.0053	0.1634 \pm 0.0051	0.1436 \pm 0.0050	0.005	0.4643
Yield P_I	0.3304 \pm 0.0092	0.3111 \pm 0.0086	0.3168 \pm 0.0084	0.0001	0.0001
Yield P_0	0.6942 \pm 0.0083	0.6880 \pm 0.0078	0.6880 \pm 0.0089	0.2631	0.9599
F_J/F_0	2.254 \pm 0.025	2.270 \pm 0.026	2.295 \pm 0.037	0.7525	0.4836
F_I/F_0	2.805 \pm 0.041	2.791 \pm 0.041	2.916 \pm 0.055	0.5587	0.0292
F_M/F_0	3.451 \pm 0.062	3.370 \pm 0.061	3.439 \pm 0.075	0.1063	0.3635
Quantum efficiencies					
$\phi_{P_0} = TR_0/ABS$	0.6942 \pm 0.0083	0.6880 \pm 0.0078	0.6880 \pm 0.0089	0.2611	0.9613
$\phi_{E_0} = ET_0/ABS$	0.3304 \pm 0.0092	0.3111 \pm 0.0086	0.3168 \pm 0.0084	0.005	0.4643
$\phi_0 = ET_0/TR_0$	0.4677 \pm 0.0101	0.4432 \pm 0.0096	0.4545 \pm 0.0100	0.003	0.2126
Specific fluxes					
ABS/RC	1.905 \pm 0.084	1.898 \pm 0.073	1.762 \pm 0.088	0.8419	0.093
TR ₀ /RC	1.245 \pm 0.025	1.245 \pm 0.021	1.135 \pm 0.026	0.7245	0.00001
ET ₀ /RC	1.215 \pm 0.024	1.216 \pm 0.020	1.110 \pm 0.025	0.7376	0.00001
DI ₀ /RC	0.661 \pm 0.065	0.653 \pm 0.057	0.628 \pm 0.065	0.9054	0.6287
Phenomenological fluxes					
ABS/CS ₀	237.0 \pm 3.3	223.1 \pm 3.0	244.5 \pm 7.6	0.0002	0.0147
TR ₀ /CS ₀	162.7 \pm 1.9	152.6 \pm 2.1	163.0 \pm 3.4	0.00001	0.0059
ET ₀ /CS ₀	74.3 \pm 2.9	70.6 \pm 2.3	75.3 \pm 2.7	0.00001	0.0054
DI ₀ /CS ₀	76.3 \pm 1.9	68.2 \pm 1.7	81.4 \pm 5.0	0.184	0.0512
ABS/CS _M	802.1 \pm 11.2	742.0 \pm 12.0	796.3 \pm 13.7	0.00001	0.00001
TR _M /CS _M	565.1 \pm 11.8	518.9 \pm 11.7	551.8 \pm 11.9	0.00001	0.0008
ET _M /CS _M	237.0 \pm 3.3	223.1 \pm 3.0	258.7 \pm 8.7	0.00001	0.0039
DI _M /CS _M	272.6 \pm 9.3	238.4 \pm 8.3	244.5 \pm 7.6	0.0002	0.0147
Density of PSII reaction centres					
RC/CS ₀	135.3 \pm 2.5	126.4 \pm 2.5	149.6 \pm 3.6	0.00001	0.00001
RC/CS _M	164.3 \pm 1.9	153.9 \pm 2.1	275.4 \pm 15.8	0.00001	0.00001
Performance indexes					
PI _{ABS}	1.62 \pm 0.10	1.38 \pm 0.08	1.64 \pm 0.11	0.0034	0.0102
PI _{CS₀}	362.3 \pm 19.9	292.6 \pm 16.3	359.9 \pm 19.7	0.00001	0.00001
PI _{CS_M}	1369.2 \pm 85.4	1080.0 \pm 70.1	1361.7 \pm 93.2	0.0002	0.0008

calculated from the initial slope of the fluorescence induction curve), and S_m (normalized complementary area, normalized for quantitative comparison of different samples) ($P < 0.0038$, $P < 0.0020$, $P < 0.0107$, $P < 0.0021$ and $P < 0.0404$) (Table 3). In late August, the fluctuation between the parameters was shifted and the effects of treatment disappeared (data not shown). There was a close positive relation between Area and tF_M (data not shown) and a close negative relation between S_m and M_0 (data not shown) during the season.

Data from 1, 6 and 13 August 2002, were pooled and the daily variations in fluorescence variables were tested for time and treatment effects (Table 4). There were no significant effects due to treatment (data not shown). The time effects are shown as the difference between 0900 and 1400 h (midday) and the difference between 1400 and 1800 h (afternoon). The quantum efficiencies ET_0/ABS and ET_0/TR_0 decreased at midday, but TR_0/ABS was unaffected. In the afternoon, the quantum efficiencies were unaffected. The estimated number of PSII RCs (RC/CS_0 and RC/CS_M) was significantly lowered at midday and increased in the afternoon. The specific fluxes per PSII RC were not significantly affected at midday, but significant decreases were observed in the afternoon for ABS/RC , TR_0/RC , and ET_0/RC . The daily variation in all phenomenological fluxes shows significant midday depression, and a tendency to increased fluxes is seen in the afternoon for the phenomenological fluxes estimated from F_M , while the opposite is seen for the fluxes estimated from F_0 (Table 4). The performance index based on equal absorption (PI_{Abs}) showed a decrease around midday and an increase during the afternoon. This was also found for the performance indexes per cross-section of the samples, using F_0 and F_M , PI_{CS_0} and PI_{CS_M} . The fluorescence intensities at 0.05 ms ($F_{50\mu s}$), at 0.1 ms ($F_{100\mu s}$), at 0.3 ms ($F_{300\mu s}$), at 2 ms (F_2), and at 30 ms (F_{30}) all showed decreased values at midday followed by an increase in the afternoon. Finally, the Area (over the fluorescence curve) was significantly decreased at midday and increased in the afternoon. An afternoon increase was also seen for tF_M , S_m , and N .

Discussion

In our experiments plants were exposed to simultaneous high ambient PAR and UV-B irradiance. There was a high correlation of PAR and UV-B ($r_s = 0.79$) and the measurements were conducted at the time of maximum irradiance of PAR and UV-B around 1400 h, the time of the maximal light stress.

Daily variation in chlorophyll a fluorescence parameters

The daily variations in most fluorescence parameters had a pattern with midday depression at noon. The marked reduction in performance indexes (PI_{CS_0} and PI_{CS_M}) sums up the effects of the high midday light values on the phenomenological fluxes (at the level of per cross-section of the sample) (Table 4). The coinciding pattern on phenomenological fluxes derived from F_0 and F_M , shows midday decreases in the total number of photons absorbed by the antenna molecule associated with all PSII RCs per cross-section of the sample (ABS/CS_M), the trapping rate of excitons, which leads to reduction of Q_A (TR_M/CS_M), and the decreased dissipation of photons (DI_M/CS_M) (Table 4). This leads to decreased electron transport, estimated as reoxidation of Q_A^- , over the leaf cross-section of the sample (ET_M/CS_M), at midday (Table 4). Moving on to the specific fluxes (at the level per PSII RC) the impact of high midday PAR and UV-B appears to be low (see below). On the other hand the performance index related to the specific fluxes (PI_{Abs}) is decreased, indicating midday depression at noon (Table 4). These findings indicate that interpretation of performance indexes (PI_{CS_0} , PI_{CS_M} and PI_{Abs}) must be done with care, but also that important differences may appear when integrating the overall response at the levels of phenomenological and specific fluxes.

Interpretation of the relation between phenomenological and specific fluxes may be done in two ways by (1) combining the fact of the decreased estimated density of active PSII RCs (RC/CS_0 and RC/CS_M) with the fact that specific fluxes (ABS/RC , TR_0/RC , DI_0/RC and ET_0/RC) are only mildly affected at midday, or (2) combining the fact of decreased quantum efficiencies (ϕ_{P_0} and ψ_0) with the fact that initial and maximal fluorescence intensities (F_0 and F_M) were also decreased (Table 4). Either way, this resulted in decreased phenomenological fluxes.

Both interpretations are reasonable from a theoretical point of view. Thus, the first interpretation (1) deduces phenomenological fluxes from the calculation of specific fluxes per RC (Strasser and Strasser 1995, Strasser et al. 2004) and the integration of the concentration of active RCs per excited cross-section of the sample (RC/CS) (Strasser et al. 2004) by assuming the absorption of photons per sample cross-section (ABS/CS) can be approximated from the fluorescence intensities F_0 and F_M (Strasser and Strasser 1995, Strasser et al. 2004). On the other hand, in the second interpretation (2) phenomenological fluxes rely on purely the mathematical derived quantum efficiencies and the approximation of ABS/CS (see above and Table 1).

Now, if it is assumed that the absorption of photons per cross-section of the sample (ABS/CS) does not change during day, then the fluctuations in either specific fluxes or quantum efficiencies will determine the fluctuations in phenomenological fluxes. In this sense, a midday depression in the phenomenological fluxes can be expected when decreases in, for example, TR_0/RC (maximal trapping rate of excitons per active PSII RC) or maximum quantum yield of primary photochemistry (ϕ_{P_0}) are found. Midday depression in phenomenological fluxes may on the other hand also be observed when decreases in F_0 or F_M are measured. In consistence with these assumptions, the only slightly decreased specific fluxes (ABS/RC, TR_0/RC , DI_0/RC and ET_0/RC), decreased quantum efficiencies (ϕ_{P_0} and ψ_0) and largely decreased F_0 and F_M (Table 4) led to a pronounced midday depression in phenomenological fluxes.

The behaviour of F_M and F_0 during the day, with a simultaneous decrease seen at midday and in the afternoon, led to a marginally decreased trapping probability (TR_0/ABS) at midday and a minor increase in the afternoon. This indicates that the plant optimizes processes related to the trapping. The decreased F_0 is therefore part of a response that actually leads to a potentially higher or equal trapping amount of photons. This does not appear to be directly reflected in F_0 , when F_0 is interpreted as a measure of absorption per cross-section (ABS/CS). These points to precaution before one interpret the ABS/CS₀ per se.

Concerning the quantum efficiencies, they may by indexing to the treatment control be used to relate the degree of stress impact on PSII donor and acceptor side. The PSII donor side related probability, TR_0/ABS (probability that an absorbed photon will be trapped by the PSII RC with the resultant reduction of Q_A to Q_A^-), was more stable than the PSII acceptor side related probability, ET_0/TR_0 (probability an electron residing on Q_A^- will enter the intersystem electron transport chain), during the day. This may be interpreted as an indication that the observed midday depression is more related to stress on the acceptor side of PSII than donor side of PSII. In relation to this, then the combination of decreased ϕ_{P_0} measured as $(TR_0/ABS = F_V/F_M)$ and the stability of the maximum trapping flux per PSII RC (TR_0/RC) at midday clearly indicates down-regulation by forming-non- Q_A -reducing RCs, so called heat sink or silent RCs (Strasser and Tsimilli-Micheal 2001, Strasser et al. 2004).

In summary, since the fluctuations of quantum efficiencies, specific fluxes, phenomenological fluxes, density of active PSII RCs, and performance indexes appear to be related to the depression, a controlled plant response to high PAR and UV-B seems to be operating. Further, the clear indication of midday depression as

down-regulation by forming-non- Q_A -reducing RCs, so-called heat sink or silent RCs (Strasser and Tsimilli-Michael 2001), also identify this time of the day to be the time when the plant is potentially most light stressed.

Relation of parameters based on means in July

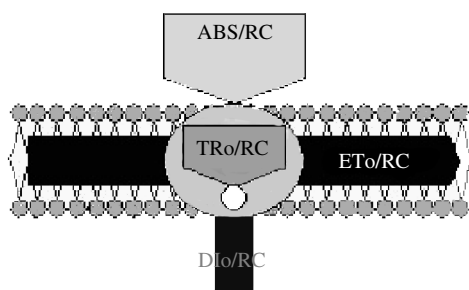
The quantum efficiencies (TR_0/ABS , ET_0/ABS , and ET_0/TR_0) were all clearly decreased in ambient UV-B (Table 3). From Table 3, it can also be seen that the fluxes per PSII RC (ABS/RC, TR_0/RC , DI_0/RC and ET_0/RC) were higher in ambient UV-B. The absorption flux of photons per active reaction centre (ABS/RC) was higher in ambient UV-B, but led only to a partly higher maximal trapping rate of excitons per PSII RC (TR_0/RC) and electron transport per active PSII RC (ET_0/RC). This is probably due to the much higher dissipation flux of untrapped excitons in the active PSII RC (DI_0/RC) in ambient UV-B. These relations are depicted in the membrane model in Fig. 8. Integration of these differences per active PSII RC leads to a decrease in the derived parameter PI_{Abs} (performance index based on equal absorption) in ambient UV-B (Table 3), indicating a less effective processing per PSII RC in ambient UV-B. Furthermore, the estimated density of active PSII RCs per cross-section of the sample (RC/CS_0 and RC/CS_M) was decreased in ambient UV-B compared to reduced UV-B (Table 3, Fig. 8). Thus, since the specific fluxes, at the level per RC (ABS/RC, TR_0/RC , DI_0/RC and ET_0/RC) were increased in ambient UV-B, but the density of PSII RCs per excited cross-section of leaf sample (RC/CS_0 and RC/CS_M) was decreased to a larger degree in ambient UV-B, decreases at the per leaf cross-section, phenomenological fluxes (TR_0/CS_0 , DI_0/CS_0 , ET_0/CS_0 , TR_M/CS_M , DI_M/CS_M , and ET_M/CS_M) were to be expected. In combination, the initial fluorescence intensity (F_0) increased, maximal fluorescence intensity (F_M) decreased plus quantum efficiencies decreased in ambient UV-B which led to decreased phenomenological fluxes (TR_0/CS_0 , DI_0/CS_0 , ET_0/CS_0 , TR_M/CS_M , DI_M/CS_M , and ET_M/CS_M) in ambient UV-B (Table 3 and pipeline model in Fig. 8). This interpretation on the relations between of specific fluxes, estimated density of PSII RC per cross-section of leaf sample (RC/CS_0 and RC/CS_M), fluorescence intensities (F_0 and F_M) and quantum efficiencies to phenomenological fluxes is consistent with the one on daily variation (see above).

The behaviour of phenomenological fluxes is summed up in the performance indexes PI_{CS_0} and PI_{CS_M} , both being decreased in ambient UV-B (Table 3).

In summary, the concerted decreases found on JIP test parameters, namely phenomenological fluxes, density

AMBIENT UV-B

Membrane model



Leaf model

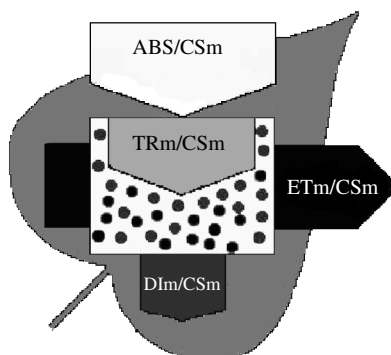
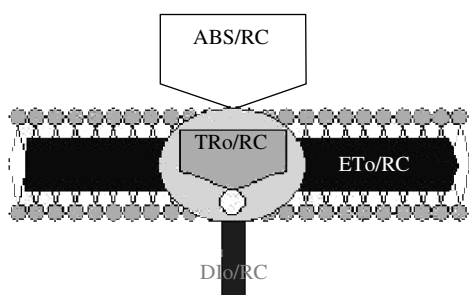


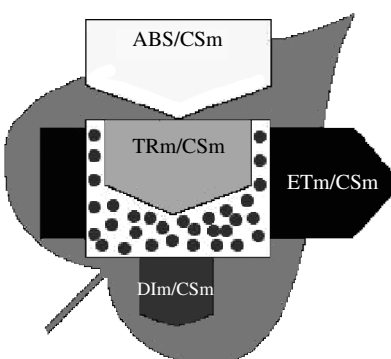
Fig. 8. Pipeline models showing the stepwise flow of energy fluxes in PSII, derived from the fluorescence transient (Table 1), at the level of individual PSII reaction centres (membrane model, displaying the specific fluxes) and at the level of excited leaf sample cross-section (leaf model, displaying phenomenological fluxes). Density of active PSII RCs (depicted as grey circles) and inactive PSII RCs (depicted as black circles) in the leaf model. The models are based on means from July (Table 3). For meaning of symbols, see Table 1. See text for discussions on the relation of fluxes.

REDUCED UV-B

Membrane model



Leaf model



of active PSII RC, quantum efficiencies and performance indexes in ambient UV-B clearly indicate that ambient UV-B is an important stressor on the photosynthetic apparatus.

Seasonal variation and climate

The seasonal variation in the fluorescence variables is related to the variation in PAR and UV-B. At the beginning of the season, from 6 to 12 July 2002, an indication of a general build-up of performance of the photosynthetic machinery was seen as a coincident increase in the normalized area over the fluorescence transient (S_m), the area between fluorescence curve and F_M (Area), and the time to reach F_M (t_{F_M}) (data not shown). Furthermore, as a consequence of this, a decreased net rate of PSII closure (M_0) (data not shown), a high level of quantum efficiencies (TR_0/ABS , ET_0/TR_0 and ET_0/ABS) (Fig. 2), and estimated number of active PSII RCs (RC/CS_0 and RC/CS_M) (Fig. 6) were seen

during the same period. After this period of clear sky conditions with high PAR and UV-B (6 to 12 July), the number of PSII RCs (RC/CS_M) was decreased to one-fifth, but RC/CS_0 only by 25% (Fig. 6). Despite this, the general increase in quantum efficiencies (TR_0/ABS , ET_0/ABS , and ET_0/ABS), phenomenological fluxes (TR_0/CS_0 , DI_0/CS_0 , ET_0/CS_0 , TR_M/CS_M , DI_M/CS_M , and ET_M/CS_M), and performance indexes (PI_{Abs} , PI_{CS_M} , and PI_{CS_0}) continued to increase. All performance indexes and quantum efficiencies had peaks on 24 July and now the opposite response, in comparison to the beginning of the season, is seen in the normalized area over the fluorescence transient (S_m), the area between fluorescence curve and F_M (Area), and the time to reach F_M (t_{F_M}), which all decreased after July 24. Furthermore, the PSII net closure rate (M_0) was increased (data not shown). This coincides with the time when the climatic variables (PAR, UV-B, and air temperature) (Fig. 1, temperature data not shown) were simultaneously decreasing. This decrease continued throughout the season. The

disappearance of treatment effects after August 9 is in contrast to the phenomena of UV-B promoted senescence (on F_V/F_M) found by Zeuthen et al. (2000). In summary, the clear seasonality of the behaviour of fluorescence parameters, derived from the fluorescence transient, clearly indicate the functionality of the JIP test as a sensitive measure of monitoring impacts on the photosynthetic machinery.

UV-B avoidance and biomass

To avoid the deleterious effects of UV-B, it was expected that the leaves exposed to UV-B would respond by growing thicker and accumulate a higher content of flavonoids as found in other studies (Tevini and Teramura 1989, Bornmann and Vogelmann 1991, Cen and Bornmann 1993, Adamse et al. 1994, Krizek et al. 1994). In this study, *Salix* leaves did have a higher content of soluble UV-B absorbing compounds in ambient UV-B, indicating accumulation of flavonoids. When no differences in biomass, but higher specific leaf area (SLA) are found it may indicate that leaf thickness is changed. Since this study found no significant difference in leaf biomass and the numeric, but insignificant, higher specific leaf area in ambient UV-B it can not be ruled out that leaf thickness has changed. This response has also been found in some UV-B supplementation experiments on other deciduous plants (Gehrke et al. 1996, Johanson et al. 1995).

Concerning UV-B screening by flavonoids, an unaffected F_V/F_M ratio (= TR_0/ABS , Table 1) has been suggested as a criterion for successful avoidance of damaging UV-B radiation (Sharma et al. 1998). The significantly reduced TR_0/ABS in ambient UV-B ($P < 0.0329$, Table 3) indicates that the deleterious effect of high UV-B could indeed not be avoided, despite the high content of soluble UV-B absorbing compounds. Therefore direct and indirect effects of ambient UV-B are to be expected further downstream (see below).

Decreased biomass in response to supplemental UV-B has been reported (Tevini and Teramura 1989, Johanson et al. 1995, Searles et al. 1995, Tosserams and Rozema 1995) and Krizek et al. (1997) found a higher biomass in reduced UV-B compared to ambient. In the present study, the leaf biomass was also lowest in ambient UV-B, although not significantly. We speculate that some of the 23% decrease in leaf biomass may potentially be explained by derived chlorophyll *a* fluorescence parameters (e.g. ET_0/ABS , ET_0/CS_0 and ET_M/CS_M). Assuming that quantum yield (ET_0/ABS) and estimated electron fluxes per cross-section of samples (ET_0/CS_0 and ET_M/CS_M) reflect potential proportions of the

flux of electrons which lead to CO_2 fixation in the Calvin–Benson cycle resulting in biomass allocation for leaf growth. In this context, the decreases in quantum yield (ET_0/ABS , 15% decrease in ambient UV-B) and electron transport per cross-section of samples (ET_0/CS_0 and ET_M/CS_M , 13 and 24% decrease in ambient UV-B, respectively) may relate to the decreased leaf biomass. In theory, higher respiration during the season or senescence phenomena at the time of harvest could also be a possible explanation for the reduced biomass, but no data in this study can evaluate these possibilities.

Other speculative possibilities may be related to the decreased leaf biomass observed, although not investigated in this study: UV-B induced decreases in photosynthesis by gene regulations (Strid et al. 1990, Jordan et al. 1991, Mackerness et al. 1996) under ecologically relevant conditions have been suggested to be most important (Björn et al. 1999). Furthermore, plants exposed to high levels of PAR and UV-B have been reported to be especially susceptible to formation of active oxygen species (Mazza et al. 1999, Andersson and Aro 2001, Karpinski et al. 2001) and precede down-regulation of photosynthesis (Mackerness et al. 1998).

Conducting further parallel measures of the relative quantum efficiency of PSII electron transport (Φ_{PSII}) and by a modulated fluorescence technique (Genty et al. 1989) and parameters derived from the fluorescence transients (e.g. ET_0/ABS , ET_0/CS and ET_M/CS) may contribute to the further understanding the derived electron flux parameters per cross-section of sample. Moreover, simultaneous measures of leaf gas exchange and modulated fluorescence may potentially identify alternative electron flux (not contributing Calvin–Benson cycle) linked to O_2 -metabolism as speculated.

Concluding remarks

The JIP test and associated parameters have been used to study stress effects related to high temperature (Srivastava et al. 1997), low temperature (Van Heerden et al. 2003), high light (Krüger et al. 1997), wind (Clark et al. 2000), nodulation status (Schmitz et al. 2001), but never used in a UV-B exclusion study. The effects of UV-B in exclusion experiments on chlorophyll fluorescence parameters have been studied once by Xiong and Day (2001) in the Antarctic, reporting that UV-B decreased the quantum yield of PSII, measured with modulated chlorophyll *a* fluorescence.

During the field season 2002, almost all measured and calculated fluorescence parameters indicated that the *Salix arctica* leaves were less stressed when screened from much of the ambient UV-B by means of filters, compared to leaves exposed to both high PAR

and high UV-B. Seasonal variation in fluorescence parameters (e.g. ET_0/ABS , ET_0/CS_0 , ET_M/CS_M , RC/CS_0 , RC/CS_M , PI_{CS_0} , PI_{CS_M}) is clearly indicated, but no UV-B promoted senescence on F_v/F_m ($= ET_0/ABS$) as reported by Zeuthen et al. (2000) was observed.

These findings clearly indicates that ambient UV-B is a stress factor lowering the potential flux of electron per leaf cross-section by decreases in quantum efficiencies and initial and maximal fluorescence. Therefore we conclude that ambient UV-B had negative effects on the photosynthetic apparatus during most of the growth season for the studied plants, with respect to the experimental set-up homogenizing and maximizing the natural light exposure.

Effective dissipated flux of untrapped excitons per active PSII per RC and per cross-section of sample (DI_0/RC , DI_0/CS_0 and DI_M/CS_M) was higher in ambient UV-B during July. Furthermore, the daily fluctuations of these parameters clearly indicated a controlled plant response to excess light and UV-B. Therefore, processes related to non-photochemical quenching are probably important for understanding the stress response, but further studies including measurements of modulated fluorescence in combination with pigment content and activity are necessary to evaluate the quenching phenomena (see below). Moreover, for establishment of a causal relationship between the relative quantum efficiency of PSII electron transport (Φ_{PSII}) and consumption of electrons for CO_2 -fixation in the Calvin-Benson cycle, but also identification of alternative electron sinks (which here may be related to O_2 -metabolism), simultaneous gas exchange and modulated chlorophyll fluorescence measurements should be conducted. In detail, seen in the light of the debate on general acceptance of derived parameters from the fluorescence transient, focus on studies comparing the general response on fluorescence parameters obtained from fluorescence transients (see Table 1) and from modulated fluorescence is needed. Basic comparison of F_0 and F_M is relevant, but also the so-called electron transport parameters estimated by means of the JIP test (ET_0/RC , ET_0/CS_0 and ET_M/CS_M) could be compared to the relative quantum efficiency of PSII electron transport (Φ_{PSII}) derived from modulated fluorescence. Moreover, the estimated energy dissipation parameters (DI_0/RC , DI_0/CS_0 and DI_M/CS_M) could be compared with the non-photochemical quenching NPQ and its components (q_E , q_I and q_T). Relation of the results may contribute to such a debate per se, but bearing in mind that only direct measurement on electron transport may be satisfactory.

Concerning the relation of effects on PSII donor and/or acceptor side capacity, the evaluation of the K-step directly addresses damage on the donor side (Strasser

1997). Further, the relative importance of donor- and acceptor side capacity can also be evaluated by deriving an index of TR_0/ABS and ET_0/ABS to treatment as done here.

With respect to the present study, the experimental set-up and the non-invasive measurements proved to be a sensitive method to screen for effects of UV-B stress.

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