

*Linking life and technology
to shape the future*



Effect of NaCl on the aggregation behavior of rhamnolipids and implications in their biological activity

Ana I. Rodrigues, Eduardo J. Gudiña, José A. Teixeira, Lígia R. Rodrigues
e-mail: isarodrigues_4@hotmail.com

VII Iberian Meeting on Colloids and Interfaces (RICI7)
Madrid
2017

CEB - Centre of Biological Engineering
University of Minho



1

INTRODUCTION

- Surfactants;
- Biosurfactants (Rhamnolipids);

2

RESULTS

- Antifungal activity;
- Dynamic Light Scattering;
- Microscopy (Confocal);

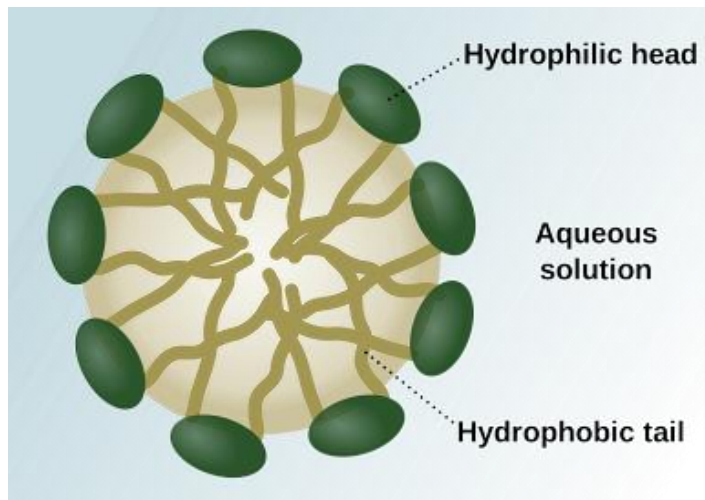
3

CONCLUSIONS

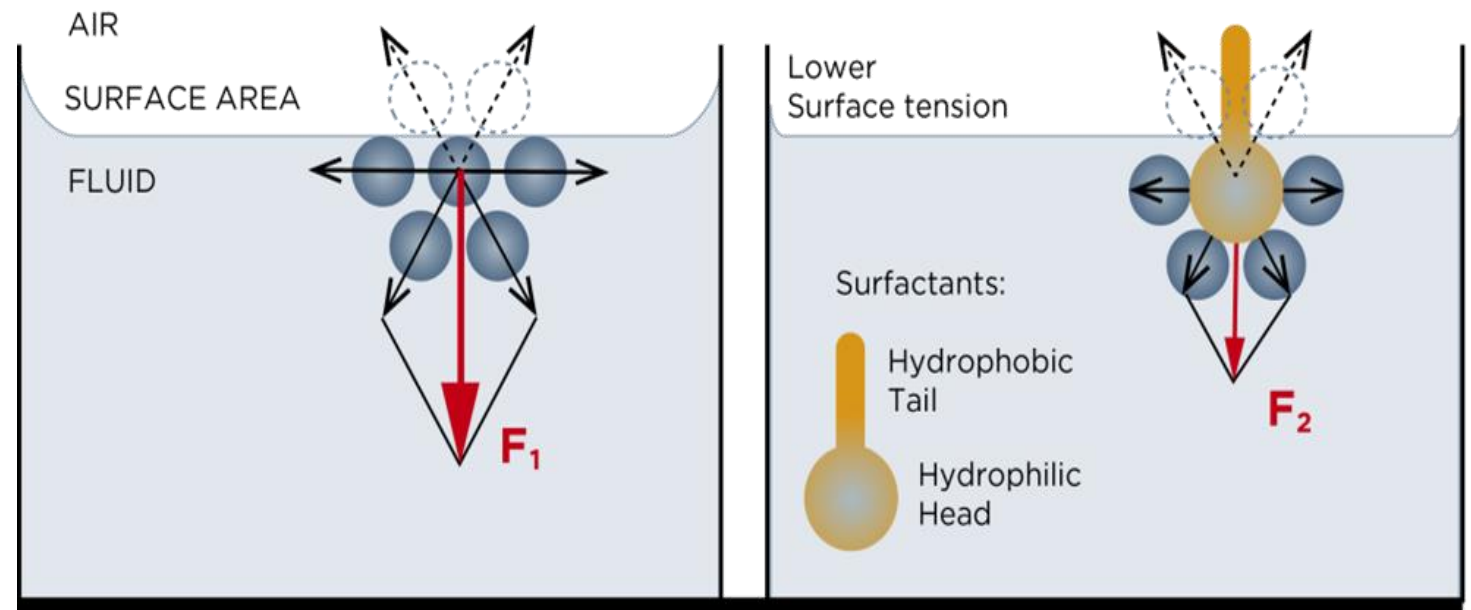
SURFACTANTS

✓ Surfactants are amphiphilic compounds

✓ Contain hydrophilic and hydrophobic groups



✓ Reduce the surface or interfacial tension between two phases with different polarities



- Synthesized by different microorganisms : bacteria, yeasts and filamentous fungi
- Properties: low toxicity
high biodegradability
high selectivity
specific activity at extreme temperatures, pH and salinities
- Can be synthesized from renewable feed-stocks and agro-industrial wastes



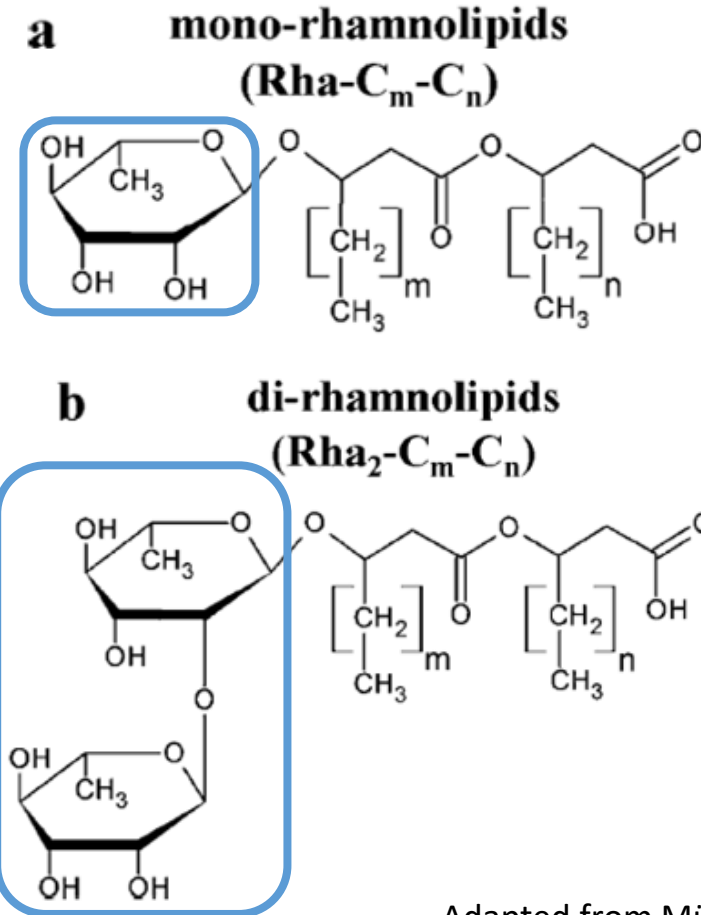
BIOSURFACTANTS

Table 1. Structural classification of biosurfactants. Adapted from Müller *et al.*, 2012

Biosurfactant size	Structural class	Examples
Low-molecular- weight	Glycolipids	Mannosylerythritol-lipids Sophorolipids Rhamnolipids ← Trehalose lipids
	Lipopeptides/ lipoamino acids	Surfactin Lysin lipids Ornithine lipids
High-molecular- weight	Polymers	Proteins Lipoproteins Polysaccharides Lipopolysaccharides
	Oil/membranes	Glycerolipids Phospholipids Fatty acids

RHAMNOLIPIDS

- **Rhamnolipids:** one (a) or two (b) rhamnose molecules linked to one or two fatty acid tails of variable length
- They are mainly produced by the Gram-negative bacterium *Pseudomonas aeruginosa*



Adapted from Müller *et al.*, 2012

Production and recovery of rhamnolipids synthesized by *P. aeruginosa* #112

RHAMNOLIPIDS PRODUCTION



Conditions

Culture medium : Corn Steep Liquor
+ Sugarcane
molasses (CSLM)

Temperature: 37 °C

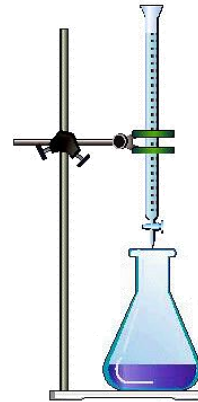
Agitation: 180 rpm

Fermentation Time : 144 h

Yield = 3194 ± 245 mg/L (cmc = 50 mg/L)

Rodrigues, A. I. *et al.* Bioresource Technology 212 (2016) 144

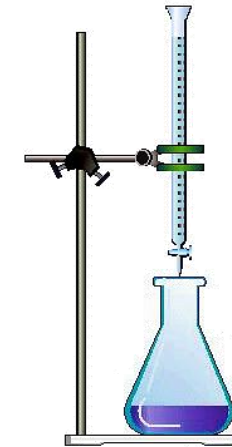
RHAMNOLIPIDS RECOVERY



Adsorption chromatography:

- polystyrene resin Amberlite XAD-2
- Elution (methanol)

RHAMNOLIPIDS CONGENERS PURIFICATION



Column chromatography:

- silica gel 60
- Elution (Chloroform:
Methanol mixtures with
increasing polarity)

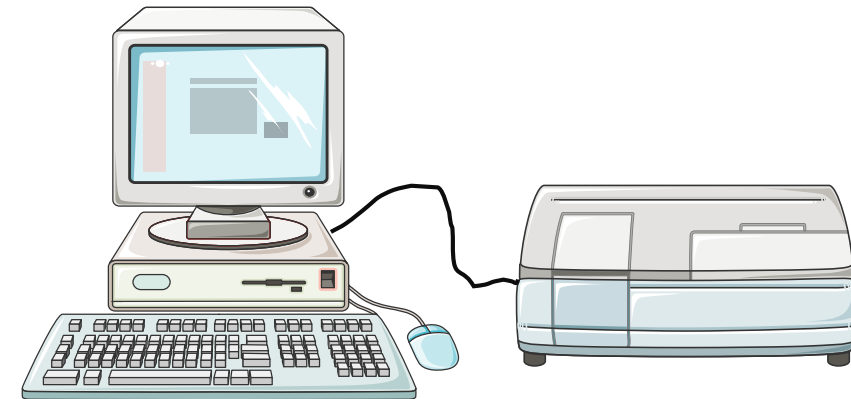
Production and recovery of rhamnolipids synthesized by *P. aeruginosa* #112

✓ Characterization

Table 2. Rhamnolipid congeners produced by *P. aeruginosa* #112 in the culture medium CSLM identified by mass spectrometry.

Rhamnolipid congeners	m/z [M+Na] ⁺	Relative abundance
Mono-Rhamnolipids		
Rha-(C ₁₀ -C ₈)	499.3	11.8 %
Rha-(C ₁₀ -C ₁₀)	527.3	100 % *
Rha-(C ₁₀ -C _{12:1})	553.3	10.4 %
Rha-(C ₁₀ -C ₁₂)	555.4	13.8 %
Di-Rhamnolipids		
Rha-Rha-(C ₈ -C ₁₀)	645.3	3.8 %
Rha-Rha-(C ₁₀ -C ₁₀)	673.3	57.8 %
Rha-Rha-(C ₁₀ -C _{12:1})	699.3	8.5 %
Rha-Rha-(C ₁₀ -C ₁₂)	701.4	14.7 %

* Most abundant ion.



Rodrigues, A. I. *et al.* Bioresource Technology 212 (2016) 144

Study of the biological activity of rhamnolipids against several fungi

Table 3. Growth inhibition percentages obtained with the cell-free supernatant and the crude rhamnolipid (RL) mixture produced by *P. aeruginosa* #112. The assays were performed at 25°C for 5 days.

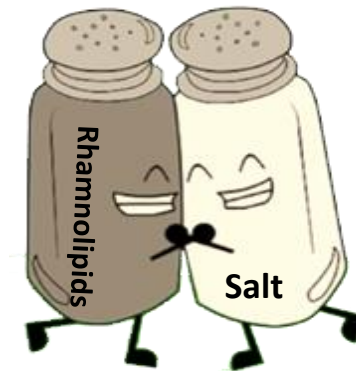
Strain	[Rhamnolipids] (g/L)	Growth Inhibition (%)	
		Cell-free supernatant	Crude RL
<i>Aspergillus niger</i> MUM 92.13	3.0	100.0 ± 0.0	20.3 ± 14.7
	1.5	31.0 ± 2.4	13.0 ± 3.7
	0.75	5.6 ± 8.5	8.1 ± 3.7
	0.375	3.5 ± 3.2	17.1 ± 6.5
<i>Aspergillus carbonarius</i> MUM 05.18	3.0	100.0 ± 0.0	22.6 ± 1.2
	1.5	100.0 ± 0.0	29.5 ± 8.3
	0.75	20.4 ± 9.1	26.7 ± 10.3
	0.375	21.7 ± 4.6	24.0 ± 2.1



Study of the biological activity of rhamnolipids against several fungi

Table 4. Growth inhibition percentages obtained with the cell-free supernatant and the crude rhamnolipid (RL) mixture produced by *P. aeruginosa* #112 at the optimized NaCl concentration. The assays were performed at 25°C for 5 days.

Strain	[Rhamnolipids] (g/L)	NaCl	Growth Inhibition (%)	
			Crude RL + NaCl	Cell-free supernatant
<i>A. niger</i> MUM 92.13	3.0	0.875 M	100.0 ± 0.0	= 100.0 ± 0.0
	1.5		51.8 ± 1.2	31.0 ± 2.4
	0.75		53.2 ± 0.0	5.6 ± 8.5
	0.375		43.3 ± 1.2	3.5 ± 3.2
<i>A. carbonarius</i> MUM 05.18	3.0	0.375 M	100.0 ± 0.0	= 100.0 ± 0.0
	1.5		58.5 ± 3.8	100.0 ± 0.0
	0.75		47.2 ± 3.8	20.4 ± 9.1
	0.375		50.9 ± 6.5	21.7 ± 4.6



Study of the biological activity of rhamnolipids against several fungi

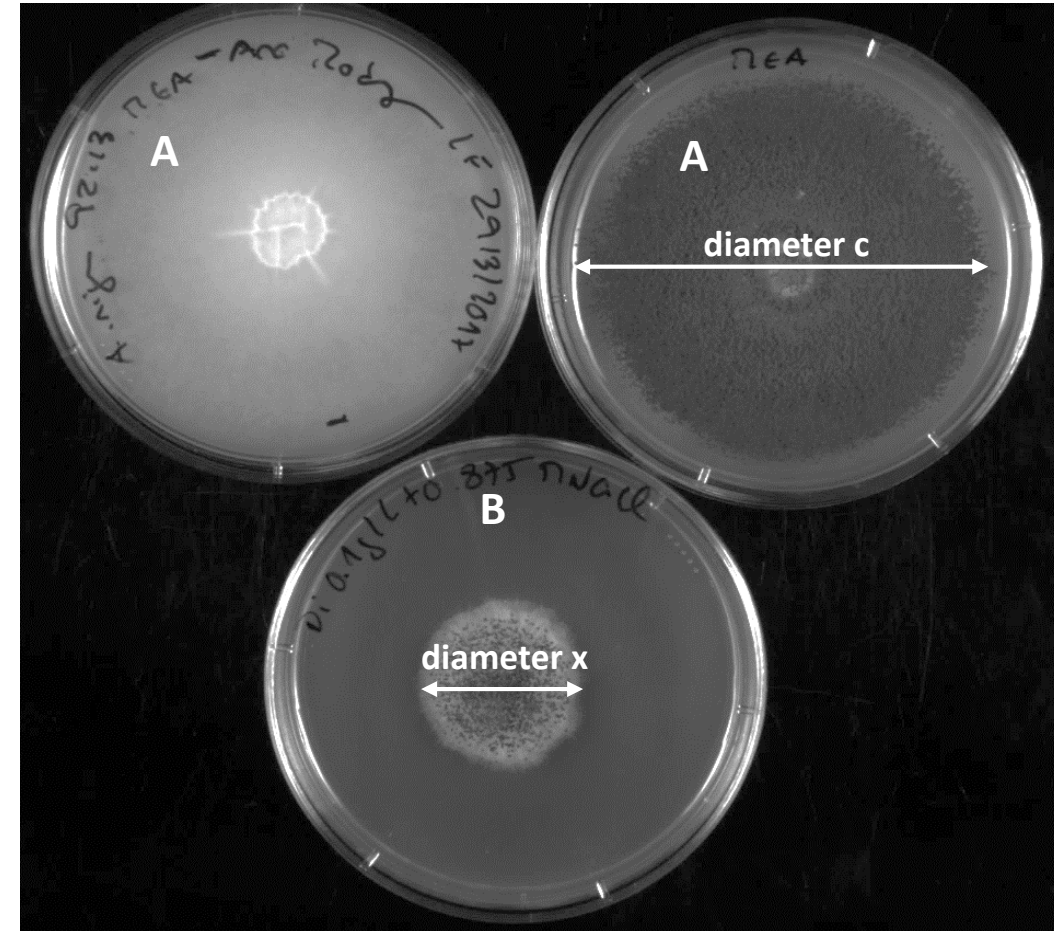
Table 5. Growth inhibition percentages obtained with the purified mono-rhamnolipid and di-rhamnolipid congeners.

A. niger MUM 92.13

NaCl	Di-Rhamnolipid (g/L)	Inhibition (%)	Mono-Rhamnolipid (g/L)	Inhibition (%)
0.875 M	0.75	100.0 ± 0.0	1.5	41.8 ± 1.4
	0.375	100.0 ± 0.0	0.75	21.2 ± 2.4
	0.2	100.0 ± 0.0	-	-
	0.1	61.9 ± 1.2	-	-
	0.05	52.4 ± 1.2	-	-

*The assays were performed at 25°C for 5 days

$$\text{Growth inhibition } x \text{ (\%)} = \left(1 - \frac{\text{diameter } x}{\text{diameter } c} \right) \times 100$$



A: Control

B: 0.1 g Di-Rhamnolipid/L +0.875M NaCl

Study of the biological activity of rhamnolipids against several fungi

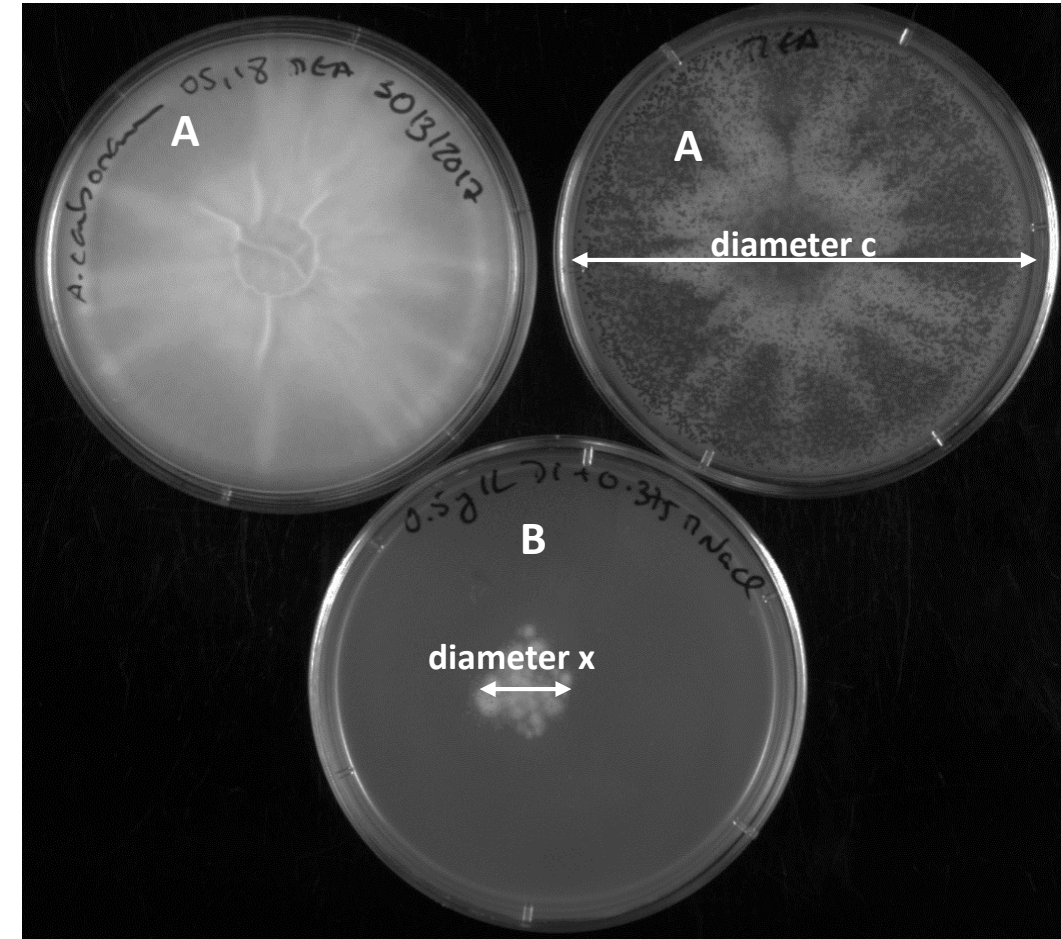
Table 6. Growth inhibition percentages obtained with the purified mono-rhamnolipid and di-rhamnolipid congeners.

A. carbonarius MUM 05.18

NaCl	Di-Rhamnolipid (g/L)	Inhibition (%)	Mono-Rhamnolipid (g/L)	Inhibition (%)
0.375 M	0.75	100.0 ± 0.0	1.5	26.4 ± 2.7
	0.6	80.7 ± 1.2	0.75	25.2 ± 4.4
	0.5	73.8 ± 1.2	-	-
	0.375	72.6 ± 1.3	-	-
	0.05	52.4 ± 1.2	-	-

*The assays were performed at 25°C for 5 days.

$$\text{Growth inhibition } x \text{ (\%)} = \left(1 - \frac{\text{diameter } x}{\text{diameter } c} \right) \times 100$$



A: Control

B: 0.5 g Di-Rhamnolipid/L + 0.375M NaCl

Study of the biological activity of rhamnolipids against several fungi

Dynamic light scattering (DLS)

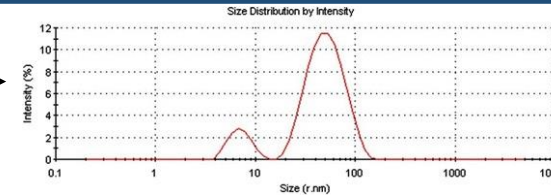
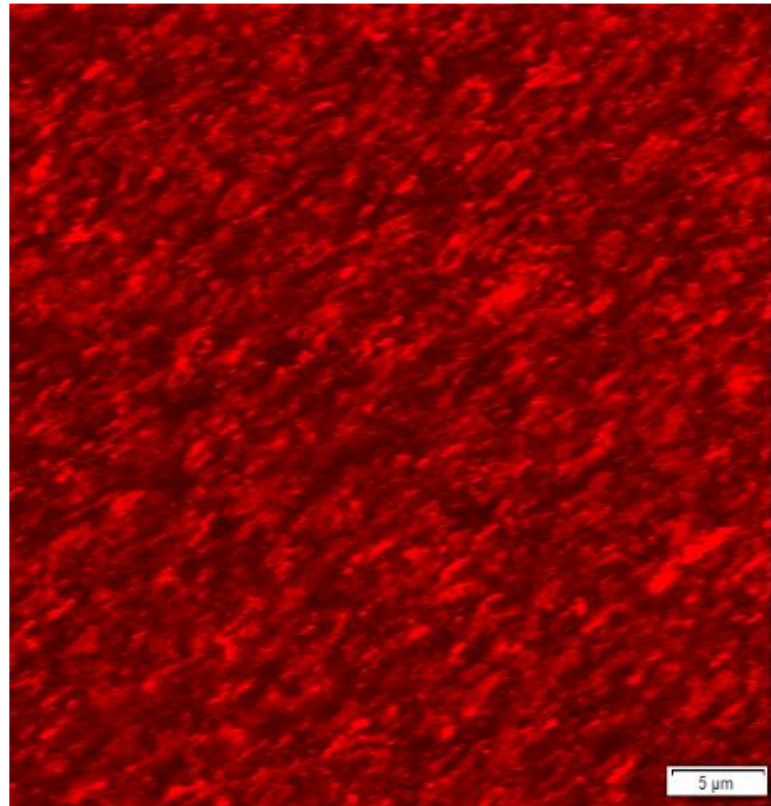


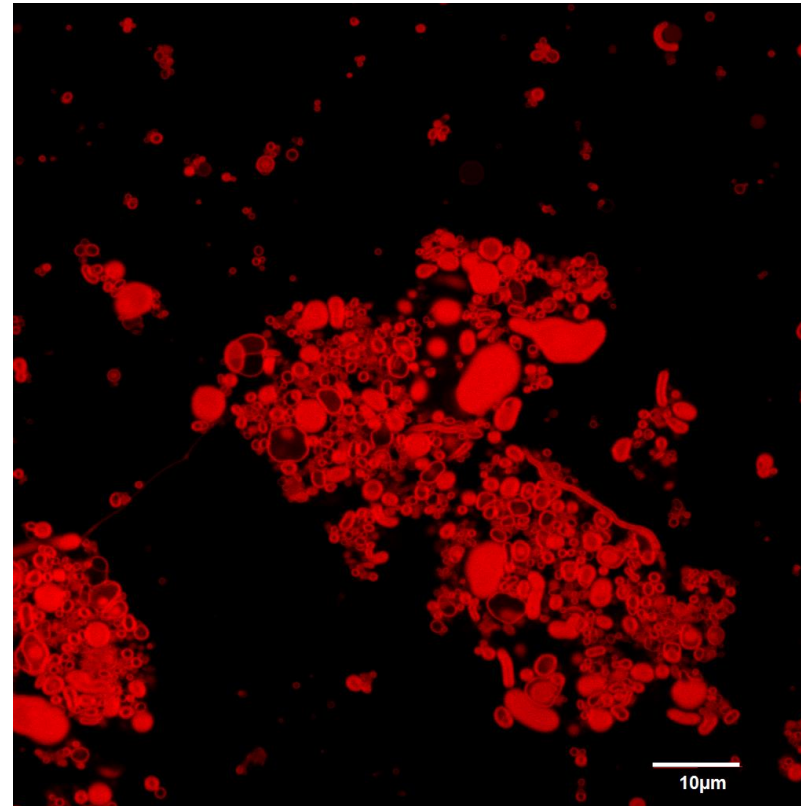
Table 7. Effect of NaCl on the micellar size distribution of the crude rhamnolipid mixture and the mono-rhamnolipid and di-rhamnolipid congeners determined by DLS analysis.

Rhamnolipid	[Rhamnolipids] (g/L)	NaCl (M)	Size (nm)	PDI
Crude (Mixture)	1.5	0.0	302.8 ± 7.4	0.549 ± 0.009
		0.375	456.6 ± 42.2	0.596 ± 0.106
		0.875	2343 ± 154.1	0.753 ± 0.190
Mono-Rhamnolipid	1.0	0.0	140.3 ± 2.0	0.263 ± 0.006
		0.375	2212 ± 444.1	0.890 ± 0.107
		0.875	4674 ± 359.8	1.000 ± 0.000
Di-Rhamnolipid	0.5	0.0	133.1 ± 4.9	0.373 ± 0.042
		0.375	> 10 000	-
		0.875	> 10 000	-

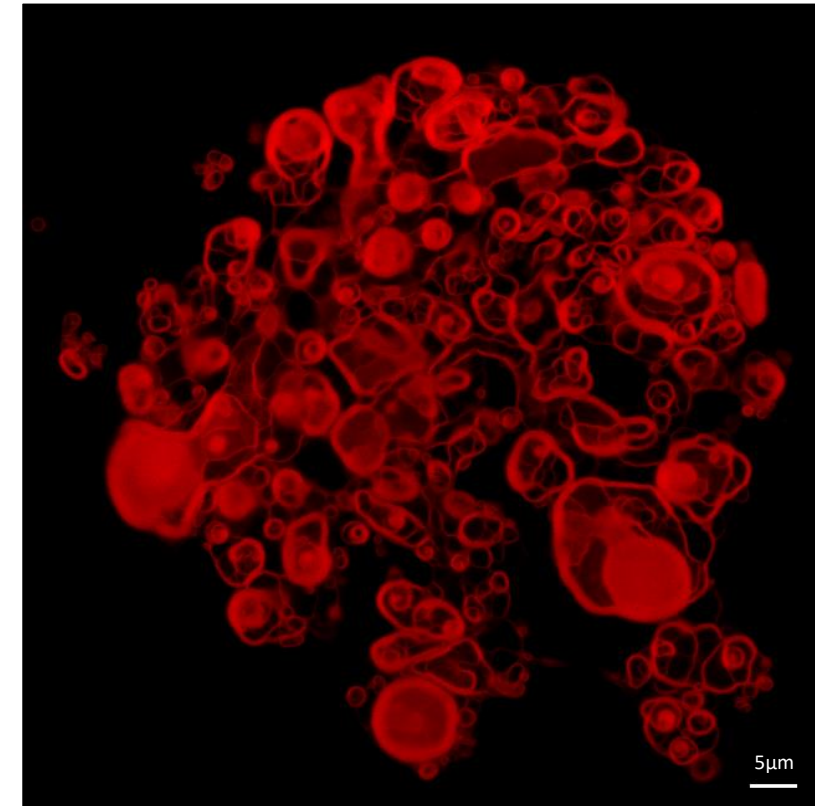
Study of the biological activity of rhamnolipids against several fungi



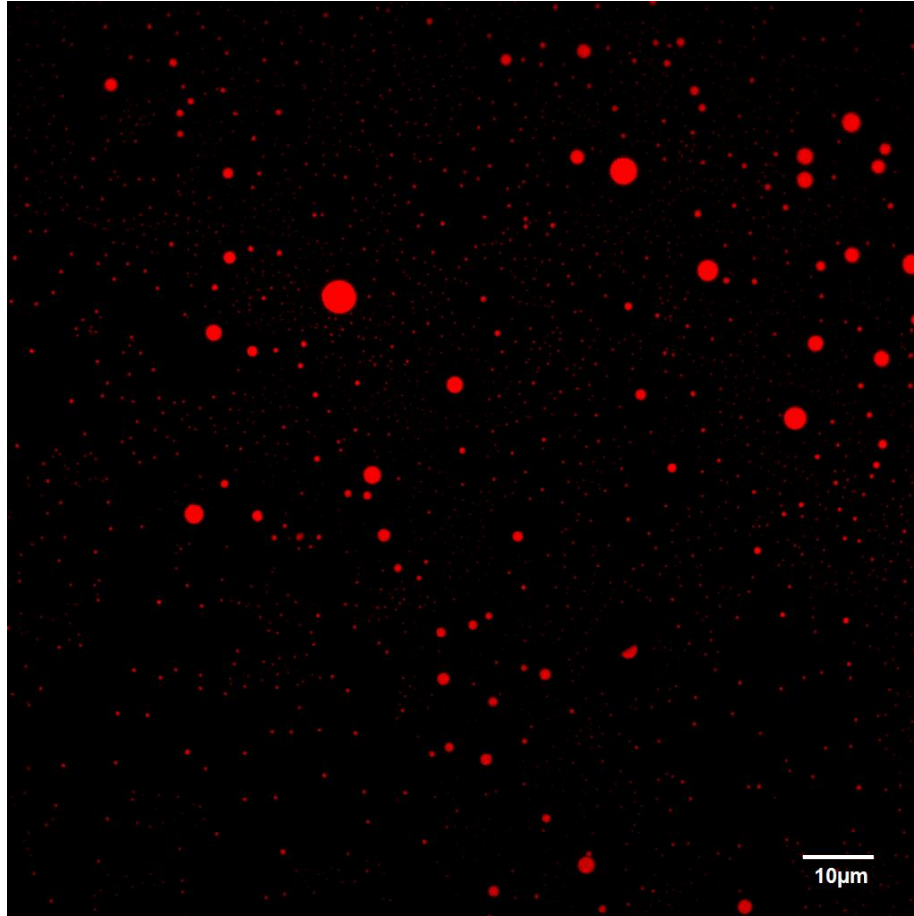
Crude Rhamnolipid (Mixture)



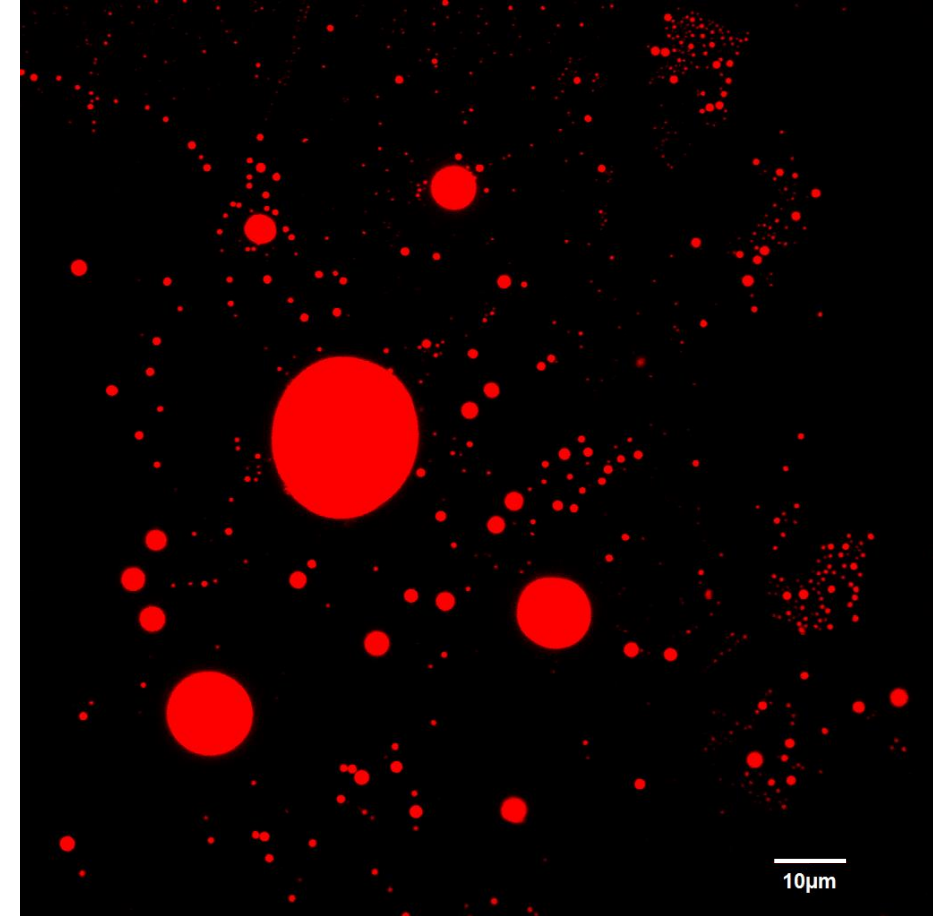
Crude Rhamnolipid (Mixture) + 0.875 M NaCl



Study of the biological activity of rhamnolipids against several fungi



Mono-Rhamnolipid + 0.875 M NaCl



Di-Rhamnolipid + 0.875 M NaCl

CONCLUSIONS

- ✓ The rhamnolipids produced in the culture medium CSLM exhibit antifungal activity against *A. niger* MUM 92.13 and *A. carbonarius* MUM 05.18
- ✓ The antifungal activity is lost during the process of rhamnolipids recovery
- ✓ The addition of NaCl alters the aggregation behavior of the crude rhamnolipids mixture restoring their antifungal activity
- ✓ The di-rhamnolipid congeners are responsible for the antifungal activity
- ✓ The rhamnolipids produced by *P. aeruginosa* #112 are a promising alternative to the chemical fungicides



ACKNOWLEDGEMENTS

together everyone
TEAM
achieves more



Prof. Lígia Rodrigues



Prof. José Teixeira



Dr. Eduardo Gudiña

This study were supported by:





*Linking life and technology
to shape the future*



University of Minho
School of Engineering

Thank you for your attention



Centre of Biological Engineering
University of Minho
Campus de Gualtar
4710-057 Braga



Email: ceb@ceb.uminho.pt
Website: www.ceb.uminho.pt

