



DTU Library

Comparison of vegetable shortening and cocoa butter as vehicles for cortisol manipulation in Salmo trutta

Birnie-Gauvin, Kim; Peiman, K. S.; Larsen, M. H.; Aarestrup, Kim; Gilmour, K. M.; Cooke, S. J.

Published in: Journal of Fish Biology

Link to article, DOI: 10.1111/jfb.13513

Publication date: 2018

Document Version Peer reviewed version

Link back to DTU Orbit

Citation (APA):

Birnie-Gauvin, K., Peiman, K. S., Larsen, M. H., Aarestrup, K., Gilmour, K. M., & Cooke, S. J. (2018). Comparison of vegetable shortening and cocoa butter as vehicles for cortisol manipulation in Salmo trutta. Journal of Fish Biology, 92(1), 229-236. DOI: 10.1111/jfb.13513

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

• Users may download and print one copy of any publication from the public portal for the purpose of private study or research.

- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

1	Comparison of vegetable shortening and cocoa butter as vehicles for cortisol manipulation
2	in Salmo trutta
3	
4	In press in Journal of Fish Biology
5	
6	K. Birnie-Gauvin ^{1†§} , K. S. Peiman ¹ , M. H. Larsen ³ , K. Aarestrup ⁴ , K. M. Gilmour ⁵ , S. J. Cooke ¹
7	
8	¹ Fish Ecology and Conservation Physiology Laboratory, Department of Biology and Institute of
9	Environmental Science, Carleton University, 1125 Colonel By Drive, Ottawa, ON, Canada K1S
10	5B6
11	³ Danish Centre for Wild Salmon, Brusgårdsvej 15, 8960 Randers, Denmark.
12	⁴ DTU AQUA, National Institute of Aquatic Resources, Section for Freshwater Fisheries and
13	Ecology, Technical University of Denmark, Vejlsøvej 39, 8600 Silkeborg, Denmark
14	⁵ Department of Biology, University of Ottawa, 30 Marie-Curie, Ottawa, ON, Canada K1N 6N5
15	
16	
17	[†] Author to whom correspondence should be addressed:
18	Email: <u>kbir@aqua.dtu.dk</u> , +45 35 88 31 14
19	§ Change of address: K. Birnie-Gauvin, DTU Aqua, National Institute of Aquatic Resources,
20	Section for Freshwater Fisheries and Ecology, Technical University of Denmark, Vejlsøvej 39,
21	8600 Silkeborg, Denmark
22	
23	Running headline: cortisol manipulation in fishes

24 ABSTRACT

25 Vegetable shortening and cocoa butter have been used as vehicles for cortisol implants in 26 a wide range of organisms, though no studies have compared the effects of these vehicles on 27 plasma cortisol and glucose, or change in mass. This study demonstrates that vegetable 28 shortening and cocoa butter are two effective vehicles for intraperitoneal cortisol implants in 29 juvenile teleost fish (brown trout, Salmo trutta) residing in north temperate freshwater 30 environments. Each vehicle showed a different pattern of cortisol elevation. Vegetable 31 shortening was found to be a more suitable vehicle for long-term cortisol elevation (elevated at 32 days 3, 6 and 9 post-treatment), while cocoa butter may be better suited for short-term cortisol 33 elevation (only elevated at 3 days post-treatment). Additionally, plasma cortisol levels were 34 higher with cortisol-vegetable shortening than with cortisol-cocoa butter implants. Plasma 35 glucose levels were elevated 6 and 9 days post-treatment for fishes injected with cortisol-36 vegetable shortening, but did not change relative to controls and shams in cortisol-cocoa butter 37 fishes. In conclusion, vegetable shortening and cocoa butter are both viable techniques for 38 cortisol manipulation in fishes in temperate climates, providing researchers with different options 39 depending on study objectives.

40

41 **KEY WORDS:** cocoa butter, cortisol implants, teleost fish, vegetable shortening

- 42
- 43
- 44
- 45
- 46

47 INTRODUCTION

Cortisol is the primary glucocorticoid stress hormone in fish (Mommsen et al., 1989; 48 49 Wendelaar Bonga, 1997; Barton, 2002). Not surprisingly, there are hundreds of papers that have 50 measured cortisol in fishes to understand the consequences of different stressors (reviewed in 51 Mommsen *et al.*, 1999). Beyond using cortisol as a biomarker of exposure to a stressor, 52 physiologists started manipulating cortisol in fishes in the 1960s to explore the mechanistic role 53 of cortisol (e.g., Slusher, 1966). This allowed researchers to move past simply observing 54 variation in cortisol levels among individuals to performing cause-and-effect studies. However, 55 despite its potential ecological relevance (Sopinka et al., 2015; Crossin et al., 2016), this 56 technique has been mainly used in the lab (reviewed in Gamperl et al., 1994). Additionally, the 57 best vehicle in which to suspend the cortisol for manipulation remains unclear. Past studies have 58 used saline oil (e.g. coconut oil), cocoa butter, and vegetable shortening to manipulate hormone 59 levels (Pottinger & Pickering, 1985; Gamperl et al., 1994; Eriksen et al., 2006; Doyon et al., 60 2006). Studies have also used mini osmotic pumps going back several decades (Theeuwes & 61 Yum, 1976). However, these are less suitable for field studies owing to expense, as fish may not 62 be recovered to retrieve the pumps, and their higher invasiveness compared to injections. The 63 main advantage of cocoa butter and vegetable shortening is that they allow for prolonged, 64 continuous release of cortisol. They are injected as liquids, and solidify once inside the fish. 65 However, cocoa butter requires high temperatures to remain in liquid form (approx. 40°C), 66 potentially resulting in the scalding of organs when injected into a fish, and becomes very hard at 67 ambient temperatures in the north temperate regions which may lead to damage to the gonads 68 (personal observation; McConnachie et al., 2012). In contrast, vegetable shortening remains in 69 liquid form at a lower temperature (approx. 30° C), and remains soft, even in cold water (5°C,

personal observation). Gamperl *et al.* (1994) originally suggested that vegetable shortening was
better than cocoa butter at lower temperatures as the hardness of cocoa butter may reduce the
absorption of cortisol.

73 This study is the first comparative study of vegetable shortening and cocoa butter as 74 vehicles for cortisol manipulation in the wild. Both vehicles are particularly suitable for field 75 studies (see Sopinka et al., 2015) owing to their low cost and ease of administration. A wild 76 population of juvenile brown trout Salmo trutta L. 1758 was used to compare the temporal 77 patterns of circulating cortisol and glucose concentrations resulting from implants of cortisol 78 suspended in vehicles of cocoa butter versus vegetable shortening. Treatment effects were 79 compared to their corresponding sham (vehicle alone) and control (no implant) groups. 80 Additionally, effects on body mass were measured treatment. It was predicted that vegetable 81 shortening implants would result in cortisol being released over a longer period of time and in 82 higher levels, resulting in higher levels of glucose and more mass loss than cocoa butter 83 implants. It was also predicted that sham treatments would not elevate cortisol or glucose concentrations or cause a change in mass compared to control fishes. 84

85

86 MATERIALS AND METHODS

The Villestrup stream is located in north-central Jutland, Denmark. The stream runs for several
kilometers across agricultural land, where a number of tributaries join in before reaching the
Mariager Fjord. The stream is home to a large population of semi-anadromous *S. trutta* (del
Villar-Guerra *et al.*, 2014). Three different sites (1 to 2km apart) within the same stream were
used. It is unlikely that there are genetic differences among populations so close (Hansen *et al.*,
2002), but even if there are, they are unlikely to have any biological significance especially when

93	comparing responses to treatments within a site. Fishes were captured via backpack
94	electrofishing (ELT 60 II GI; 300 volts; Scubla, Remanzacco, Italy) on three separate days in
95	2016: 125 fishes at Site 1 on March 3 rd (25 fishes per group), 125 fishes at Site 2 on March 4 th
96	(25 fishes per group) and 150 fishes at Site 3 on March 5 th (30 fishes per group). During this
97	period, the temperature of the water in Villestrup was between 6 and 7°C.
98	Captured fishes were held in a 60 l bin filled with oxygenated fresh stream water. Fishes
99	were anesthetized in a solution of benzocaine (0.03 g l ⁻¹ ethyl- <i>p</i> -aminobenzoate; Sigma,
100	www.sigmaaldrich.com) in stream water, then weighed (± 0.1 g), measured for total length
101	$(\pm 0.1 \text{cm})$, and tagged using a 23mm PIT tag (Passive Integrated Transponder tag, Texas
102	Instruments, RI-TRP-RRHP, 134Hz, 0.6g mass in air, Plano, Texas, USA). Tags were inserted
103	through a 5mm incision in the left side of the body, posterior to the pelvic fin. Only trout that
104	were 12-21cm in length (large enough for the PIT tag, but likely still juveniles; Larsen et al.
105	2013) were used in this study. Fishes were randomly assigned to one of the following treatment
106	groups: (1) control, (2) sham-vegetable shortening (sham-veg), (3) cortisol-vegetable shortening
107	(cort-veg), (4) sham-cocoa butter (sham-cocoa), (5) cortisol-cocoa butter (cort-cocoa). Cortisol-
108	treated fishes received an intracoelomic injection (1.5inch 18-gauge needle) of a suspension of
109	vegetable shortening (100% vegetable shortening, Crisco, OH, USA) or cocoa butter (100% pure
110	cocoa butter, NOW Foods, IL, USA) mixed with hydrocortisone 21-hemisuccinate (Sigma-
111	Aldrich, St. Louis, MO, USA, Product #H2882-1G), using a dosage of 0.01 ml vehicle (with a
112	concentration of 0.01g cortisol per ml) per 1 g of fish (equivalent to a cortisol dosage of 100mg
113	kg ⁻¹). Sham fishes were injected with only 0.01 ml g ⁻¹ fish vegetable shortening or cocoa butter.
114	The vegetable shortening and cocoa butter were heated using hot water to a temperature of 37°C
115	and 40 °C, respectively. All fishes were recovered (i.e., until full equilibrium was reached) in a

60 l tank of benzocaine-free fresh stream water following tagging. Cortisol-treated fishes were
recovered separately from sham and control fishes to prevent any cross-treatment contamination
of cortisol, and all fishes were then released at the site of capture. The tagging, weighing,
measuring and injecting process took less than one minute per fish. Overall, fishes were held in
tanks for approximately 60 minutes.

121 Fishes were recaptured via backpack electrofishing after 3, 6 and 9 days post-treatment, 122 at Site 3, Site 2 and Site 1, respectively. Immediately after shocking, we collected a blood sample 123 (<0.3 ml) from the caudal vasculature using a heparinized 1.5-inch 25-gauge needle and a 1 ml 124 syringe. All samples were collected within 3 minutes of capture. Fishes were then weighed. 125 Following recovery, fishes were returned to the river, and not recaptured. Blood samples were 126 held in a water-ice slurry until centrifuged at 2000 g for 2 minutes to separate plasma from red 127 blood cells. Plasma samples were kept at -80°C until analyzed. Environmental conditions should 128 not be a confounding factor here, as the 3 day sampling was within the 6 day sampling, and both 129 were within the 9 day sampling period. Hence, all fishes were exposed to the same conditions, 130 with day 9 fishes potentially experiencing a greater variation. However, this does not affect 131 treatment effects within a single time point, which is the focus of this study.

Plasma cortisol concentration was determined using a commercial radioimmunoassay kit
(ImmunoChem Cortisol ¹²⁵I RIA kit; MP Biomedicals, www.mpbio.com). This assay was
previously validated for use with teleost fish plasma samples (Gamperl *et al.*, 1994). All plasma
samples were measured in a single assay. Intra-assay variability (%CV) was 7.9%. Plasma
glucose levels were determined using an AccuCheck Compact Plus meter system (Roche, Basel,
Switzerland), a point-of-care device previously validated for use in teleost fishes (Stoot *et al.*,
2014).

139 Statistical analyses were conducted using JMP v12.0.1 (SAS Institute Inc., 140 Buckinghamshire, UK). Cortisol and glucose values were log-transformed to achieve normality 141 of residuals. Two-way ANOVAs were used to evaluate differences in cortisol, glucose and 142 change in mass among treatment groups over the three sampling times. A Tukey-Kramer post-143 *hoc* test was used to determine which groups differed, which is conservative with unequal 144 sample sizes as is the case here. Spearman correlations (to reduce the effect of outliers) were 145 used to determine whether cortisol levels were related to glucose levels among individuals using 146 within each category of treatment and day.

147

148 **RESULTS**

149 Between 9 and 17 fishes were recaptured per treatment group. Fishes treated with cortisol 150 suspended in vegetable shortening showed significantly higher plasma cortisol concentrations 151 after 3, 6 and 9 days post-treatment than both sham and the control treatments, with values at day 152 3 significantly higher than at day 9 (Fig. 1A; treatment × time, $F_{8,172} = 3.07$, P = 0.0029). Cort-153 cocoa fishes at day 3 had significantly higher cortisol levels than both sham and the control 154 treatments, but values for fishes sampled at days 6 and 9 did not differ from those for sham or 155 control fishes. At day 3, cort-veg fishes exhibited significantly higher plasma cortisol levels than 156 cort-cocoa fishes. Cortisol concentrations for fishes in the sham treatment were similar to fishes 157 in the control group across all time points. Glucose concentrations in cort-veg fishes were 158 significantly higher than those for sham and control treatments at days 6 and 9 (Fig. 1B; treatment \times time, F_{8,170} = 2.30, P = 0.023), whereas plasma glucose concentrations in cort-cocoa 159 160 fishes did not differ from the sham or control groups on any day. On days 6 and 9, cort-veg 161 fishes had significantly higher glucose concentrations than cort-cocoa fishes.

Initially, mass for cortisol-treated fishes did not differ from their sham or the control

163 group (all P > 0.50). Sham-veg fishes sampled on day 9 gained mass while all other groups lost

164 mass (Fig 1C, treatment × time, $F_{8,170} = 2.94$, P = 0.0042).

Plasma cortisol and glucose concentrations were positively related in day 9 cort-veg treatment ($R^2 = 0.60$, n=15, P = 0.037) No other correlation was significant (all P > 0.093).

167

168 **DISCUSSION**

169 Cortisol implants (100mg kg⁻¹) generated a significant elevation in plasma cortisol 170 concentration using either vegetable shortening or cocoa butter as a vehicle. However, the use of 171 vegetable shortening as a vehicle caused a greater elevation of cortisol concentration than cocoa 172 butter after 3 days, and this elevation lasted longer. Moreover, plasma cortisol concentration likely remained high for more than 9 days in fishes that received cortisol-vegetable shortening 173 174 implants, as found by Pickering & Duston (1983). In contrast, cocoa butter implants had short-175 lasting effects on plasma cortisol levels, with circulating concentrations returning to control 176 levels by 6 days post-treatment. The soft texture of vegetable shortening (Fig. 2), even at low 177 temperatures (solidifies at 20°C, but remains soft at lower temperatures – e.g., it was 6-7°C 178 during this study) likely allows for more effective (i.e., faster) release of the cortisol. Cocoa 179 butter, however, becomes very hard even at fairly high temperatures (solidifies at 20°C), which 180 may prevent long-lasting release of cortisol in north temperate fish species, as indicated by the 181 peak cortisol levels 3 days post-treatment. The outer cortisol likely gets released quickly, but the 182 hardness of the cocoa butter prevents the release of the inner cortisol. Alternatively, it is possible 183 that cocoa butter releases cortisol more readily than vegetable shortening, leading to the implant 184 being depleted of cortisol more rapidly and the cortisol values in cocoa butter-treated fishes

peaking earlier than the first sampling time (3 days). Unfortunately, there is no way to
distinguish between the two possibilities with our data. The conclusion however, remains the
same: vegetable shortening appears to be a more appropriate vehicle for studies seeking longterm cortisol elevation, while cocoa butter may be better suited for short-term cortisol elevation,
at least in north temperate regions.

190 Cortisol increases the rate of gluconeogenesis (reviewed by Mommsen et al., 1999). An 191 increase in plasma glucose following treatment with cortisol implants therefore would be 192 consistent with the known physiological effects of cortisol. Plasma glucose concentrations were 193 found to be higher than those of sham and control treatments at both day 6 and 9 in cort-veg 194 fishes. In contrast, plasma glucose was never elevated above sham or control treatment fishes in 195 cort-cocoa fishes, in agreement with the shorter-lasting physiological effect of cocoa butter than 196 vegetable shortening on cortisol levels. Additionally, cortisol caused an increase in glucose 197 levels earlier in the cort-cocoa treatment (day 3) than in the cort-veg treatment (day 9), further 198 supporting the hypothesis that the cocoa butter vehicle generates a shorter and faster response 199 than vegetable shortening.

200 Increased conversion of stored energy reserves to glucose during gluconeogenesis may 201 also lead to a loss in mass. Additionally, cortisol tends to suppress appetite leading to a reduction 202 in food intake, and this would also be expected to result in mass loss (Madison *et al.*, 2015). The 203 9 days of the cortisol treatment examined in the present study did not have a significant effect on 204 change in mass relative to that observed in control or sham-treated fishes, suggesting that the 205 physiological effects of elevated cortisol take more time to manifest as changes in mass. 206 Previous studies in similar systems have reported decreased growth rates of cortisol-treated 207 fishes over two weeks and longer (Madison et al., 2015; Midwood et al., 2015; Midwood et al.,

208 2016; Birnie-Gauvin *et al.*, 2017; Peiman *et al.*, 2017). Sham-veg fishes at day 9 showed a
209 significant increase in mass, which may have resulted from the vegetable shortening itself
210 starting to be absorbed internally, while in the cort-veg fishes this effect may have been offset by
211 glucose metabolized by cortisol. Indeed, it was only in this latter group that cortisol and glucose
212 were positively related. The mechanism by which this occurred is unknown and its biological
213 significance remains evasive.

214 The present study showed that vegetable shortening and cocoa butter are two effective 215 vehicles for cortisol implants in north temperate regions, and that sham treatments with the 216 vehicle alone do not result in growth impairments compared to controls over the short-term, as 217 previously observed in reproductive female S. trutta following cocoa butter sham implants 218 (Hoogenboom *et al.*, 2011). However, it was noticed that cocoa butter implants had sharp edges, 219 which could result in internal organ damage, a potentially deleterious effect which has not 220 previously been noted. Cortisol levels peaked 3 days post-treatment for both vegetable 221 shortening and cocoa butter implants, and cortisol levels remained elevated for 9 days with the 222 vegetable shortening implant. Maximum cortisol levels achieved in this experiment are beyond 223 the physiological range for salmonids (Donaldson, 1981; Gamperl et al., 1994). If the goal of the 224 study requires cortisol levels within the normal physiological range, a lower dosage of cortisol 225 may be appropriate. Glucose levels were affected by cortisol in fishes that received vegetable 226 shortening but not cocoa butter implants. Thus, in north temperate regions, vegetable shortening 227 is a more appropriate vehicle for studies seeking longer-term cortisol elevation, while cocoa 228 butter may be better suited for studies looking for short-term cortisol elevation, providing 229 researchers with different options depending on study objectives.

230

231	S. J. Cooke is supported by the Canada Research Chairs Program, the NSERC E.W.R. Steacie
232	Memorial Fellowship and the NSERC Discovery Grant (DG) program. This study was also
233	partly funded by the Danish Rod and Net Fish License Funds, and by NSERC DG funding to K.
234	M. Gilmour. We thank J. S. Mikkelsen, M. Holm, HJ. Christensen, A. Garcia Laborde and F.
235	Valenzuela Aguayo for assisting us in the field. We also thank ME. Bélair Bambrick and C.
236	Best for their help in the lab.
237	
238	REFERENCES
239	
240	Barton, B. A. (2002). Stress in fishes: a diversity of responses with particular reference to
241	changes in circulating corticosteroids. Integrative and Comparative Biology 42, 517-525. doi:
242	10.1093/icb/42.3.517
243	
244	Birnie-Gauvin, K., Peiman, K. S., Larsen, M. H., Aarestrup, K., Willmore, W. G., Cooke, S. J.
245	(2017). Short-term and long-term effects of transient exogenous cortisol manipulation on
246	oxidative stress in juvenile brown trout. Journal of Experimental Biology 220, 1693-1700. doi:
247	10.1242/jeb.155465
248	
249	Crossin, G. T., Love, O. P., Cooke, S. J., & Williams, T. D. (2016). Glucocorticoid
250	manipulations in free-living animals: considerations of dose delivery, life-history context and
251	reproductive state. Functional Ecology 30 , 116-125. doi: 10.1111/1365-2435.12482
252	

- del Villar-Guerra, D., Aarestrup, K., Skov, C., Koed, A. (2014). Marine migrations in
 anadromous brown trout (*Salmo trutta*). *Ecology of Freshwater Fish* 23, 594-603. doi:
 10.1111/eff.12110
- 256
- 257 Donaldson, E. M. (1981). Pituitary-interrenal axis as an indicator of stress in fish. In Stress and

258 Fish (Pickering, A. D., eds), pp. 11-47. London and New York: Academic Press.

259

- 260 Doyon, C., Leclair, J., Trudeau, V. L., Moon, T. (2006). Corticotropin-releasing factor and
- 261 neuropeptide Y mRNA levels are modified by glucocorticoids in rainbow trout, Oncorhynchus
- 262 *mykiss. General and Comparative Endocrinology* **146**, 126-135. doi:
- 263 10.1016/j.ygcen.2005.10.003
- 264
- 265 Eriksen, M. S., Bakken, M., Espmark, Å., Braastad, B. O., Salte, R. (2006). Prespawning stress
- 266 in farmed Atlantic salmon *Salmo salar*: maternal cortisol exposure and hyperthermia during
- 267 embryonic development affect offspring survival, growth and incidence of malformations.

268 Journal of Fish Biology 69, 114-129. doi: 10.1111/j.1095-8649.2006.01071.x

269

- 270 Gamperl, A. K., Vijayan, M. M., Boutilier, R. G. (1994). Experimental control of stress hormone
- 271 levels in fishes: techniques and applications. *Reviews of Fish Biology and Fisheries* **4**, 215-255.
- doi: 10.1007/BF00044129

- Hansen, M. M., Ruzzante, D. E., Nielsen, E. E., Bekkevold, D., Mensberg, K. L. D. (2002).
- 275 Long-term effective population sizes, temporal stability of genetic composition and potential for

- local adaptation in anadramous brown trout (*Salmo trutta*) populations. *Molecular Ecology* 11,
 2523-2535. doi: 10.1046/j.1365-294X.2002.01634.x
- 278
- 279 Hoogenboom, M. O., Armstrong, J. D., Miles, M. S., Burton, T., Groothuis, T. G., Metcalfe, N.
- 280 B. (2011). Implantation of cocoa butter reduces egg and hatchling size in Salmo trutta. Journal

281 *of Fish Biology* **79**, 587-596. doi: 10.1111/j.1095-8649.2011.03039.x

282

- 283 Larsen, M. H., Thorn, A. N., Skov, C., Aarestrup, K. (2013). Effects of passive integrated
- transponder tags on survival and growth of juvenile Atlantic salmon, Salmo salar. Animal

285 *Biotelemetry* **1**, 19-25. doi: 10.1186/2050-3385-1-19

286

- 287 Madison, B. N., Tavakoli, S., Kramer, S., Bernier, N. J. (2015). Chronic cortisol and the
- regulation of food intake and the endocrine growth axis in rainbow trout. *Journal of*

289 *Endocrinology* **226**, 103-119. doi: 10.1530/JOE-15-0186

290

- 291 McConnachie, S. H., Cook, K. V., Patterson, D. A., Gilmour, K. M., Hinch, S. G., Farrell, A. P.,
- 292 Cooke, S. J. (2012). Consequences of acute stress and cortisol manipulation on the physiology,
- 293 behavior and reproductive outcome of female Pacific salmon on spawning grounds. *Hormones*
- and Behaviour 62, 67-76. doi: 10.1016/j.yhbeh.2012.05.001

- 296 Midwood, J. D., Larsen, M. H., Boel, M., Aarestrup, K., Cooke, S. J. (2015). An experimental
- 297 field evaluation of winter carryover effects in semi-anadromous brown trout (*Salmo trutta*).
- 298 Journal of Experimental Zoology 323, 645-654. doi: 10.1002/jez.1955

300	Midwood, J. D., Larsen, M. H., Aarestrup, K., Cooke, S. J. (2016). Stress and food deprivation:
301	linking physiological state to migration success in a teleost fish. Journal of Experimental Biology
302	219 , 3712-3718. doi: 10.1242/jeb.140665
303	
304	Mommsen, T. P., Vijayan, M. M., Moon, T. W. (1999). Cortisol in teleosts: dynamics,
305	mechanisms of action, and metabolic regulation. Reviews in Fish Biology and Fisheries 9, 211-
306	268. doi: 10.1023/A:1008924418720
307	
308	Peiman, K. S., Birnie-Gauvin, K., Midwood, J. D., Larsen, M. H., Wilson, A. D., Aarestrup, K.,
309	Cooke, S. J. (2017). If and when: intrinsic differences and environmental stressors influence
310	migration in brown trout (Salmo trutta). Oecologia 184, 374-384. doi: 10.1007/s00442-017-
311	3873-9
312	
313	Pickering, A. D., Duston, J. (1983). Administration of cortisol to brown trout, Salmo trutta L.,
314	and its effects on the susceptibility to Saprolegnia infection and furunculosis. Journal of Fish
315	<i>Biology</i> 23 , 163-175. doi: 10.1111/j.1095-8649.1983.tb02891.x
316	
317	Pottinger, T. G., Pickering, A. D. (1985). The effects of 11-ketotestosterone and testosterone on
318	the skin structure of brown trout, Salmo trutta L. General and Comparative Endocrinology 59,
319	335-342. doi: 10.1016/0016-6480(85)90389-2

- 321 Slusher, M. A. (1966). Effects of cortisol implants in the brainstem and ventral hippocampus on
 322 diurnal corticosteroid levels. *Experimental Brain Research* 1, 184-194. doi:
 323 10.1007/BF00236870
- 324
- 325 Sopinka, N. M., Patterson, L. D., Redfern, J. C., Pleizier, N. K., Belanger, C. B., Midwood, J. D.,
- 326 Crossin, G. T., Cooke, S. J. (2015). Manipulating glucocorticoids in wild animals: basic and
- applied perspectives. *Conservation Physiology* 3, cov031. doi:10.1093/conphys/cov031
 328
- 329 Stoot, L. R., Cairns, N. A., Cull, F., Taylor, J. J., Jeffrey, J. D., Morin, F., Mandelman, J. W.,
- 330 Clark, T. D., Cooke, S. J. (2014). Use of portable blood physiology point-of-care devices for
- basic and applied research on vertebrates: a review. *Conservation Physiology* **2**, cou011.
- doi: 10.1093/conphys/cou011
- 333
- 334 Theeuwes, F., Yum, S. I. (1976). Principles of the design and operation of generic osmotic
- 335 pumps for the delivery of semisolid and liquid drug formulations. Annals of Biomedical

336 *Engineering* **4**, 343-353. doi: 10.1007/BF02584524

- Wendelaar Bonga, S. E. W. (1997). The stress response in fish. *Physiological Reviews* 77, 591625. doi: 10.0031-9333/97
- 340
- 341
- 342
- 343

344 FIGURE CAPTIONS

346	Fig 1. PIT-tagged brown trout (Salmo trutta) were subjected to one of 5 treatments; control (no
347	implant), sham-veg (given a vegetable shortening implant), sham-cocoa (given a cocoa butter
348	implant), cort-veg (given 100mg kg ⁻¹ of cortisol suspended in a vegetable shortening implant)
349	and cort-cocoa (given 100mg kg ⁻¹ of cortisol suspended in a cocoa butter implant), and were re-
350	captured at 3 (black bars), 6 (grey bars) or 9 (white bars) days post-treatment. (A) Plasma
351	cortisol concentration, (B) plasma glucose concentration, and (C) change in mass are presented
352	as a function of treatment group and sampling day. Values are means + SEM, $N = 9$ to 17.
353	Groups that share a letter are not significantly different from one another (see text for details).
354	
355	Fig 2. Representative images of the dissection of brown trout (Salmo trutta) post-treatment to
356	illustrate the different implant vehicles; (A) control, (B) vegetable shortening implant, and (C)
357	cocoa butter implant. Arrows point to the implants. Vegetable shortening remained soft at 3, 6
358	and 9 days post-treatment. Cocoa butter implants were hard to the touch at 3, 6 and 9 days post-
359	treatment, with some implants showing sharp edges.
360	
361	
362	
363	
364	
365	
366	

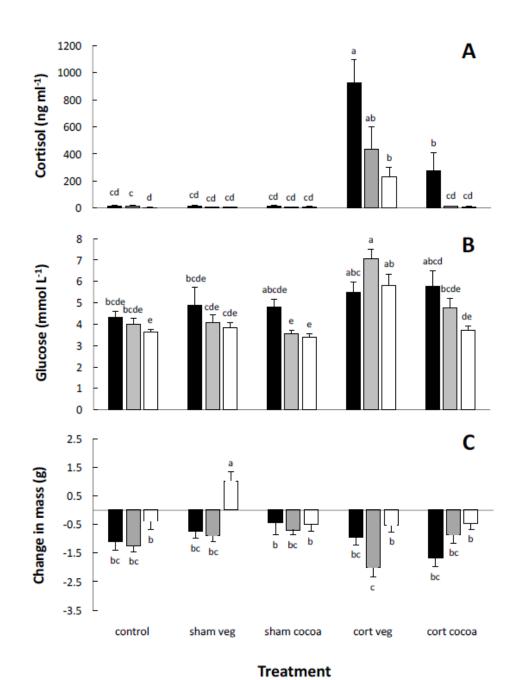


Figure 2.

