### Rice Root physiology work at CIAT:

### Identification of ideal root system to improve water and Nitrogen uptake under stress conditions

# Satoshi Ogawa JSPS Post-doctoral fellow



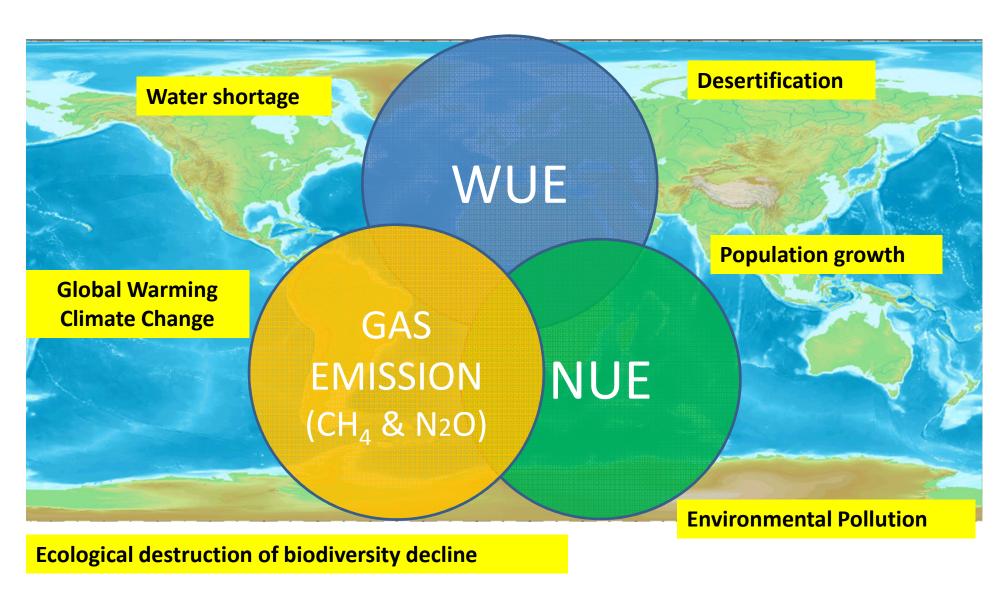








### Our challenging



These problems are further exacerbating

# **Underground Revolution**

#### **AN UNDERGROUND** REVOLUTION Plant breeders are turning their attention to roots to increase yields without causing environmental damage. Virginia Gewin unearths some promising subterranean strategies. angled, dirty and buried underfoot, to increase yields is because the treme roots are a mess to study. Digging them dous genetic variation trapped in roots has been neglected," says Lynch. Here, up is a time-consuming and sometimes back-breaking process. The shovel must Nature reports on four of the most be wielded with care to preserve the roots' delipromising leads for boosting food cate branching patterns, the root hairs and the production through roots. microbes that cling to them. All of this explains why roots have been largely out of mind, as well as out of sight, for agricultural researchers — Roots are most efficient when their architecture is tailored to their environ until now.

central to their efforts to produce crops with a better yield - efforts that go beyond the Green Revolution. That intensive period of research and development, starting in the 1940s, dramatically boosted food production through the breeding of high-yield crop varieties and the use of pesticides, fertilizers and more water. But the increases were accompanied by a depletion of groundwater and, by 1998, an eightfold increase in nitrogen-based fertilizer usage<sup>1</sup>, bringing environmental problems such as polluted waterways. The leaps in yield have still left many hungry. And the revolution missed many developing nations, some of which have poor soils and limited access to irrigation and expensive fertilizers. "Those strategies of the past aren't working now to meet growing food needs," says Jonathan Lynch, a plant nutritionist at Pennsyl-

**NEWS FEATURE FOOD** 

vania State University in University Park.

"Roots are the key to a second green revolution—one that doesn't rely on expensive inputs,'
says Lynch. Roots deliver water and nutrients,
two of the most essential, often-limiting, factors
that a plant needs. Why keep putting in more
water and fertilizers, he and others reason, when
they might instead improve roots' ability to use
what's already there and, in the process, help to
convert 'marginal' lands into productive ones.

There is room for improvement. Although plant breeders have already made huge gains by manipulating 'above ground' trusts'—for example, by breeding dwarf plant varieties, which put more energy into producing grain rather than the stalk — the same is not true for root traits. 'One reason we now have any potential

Roots are most efficient when their architecture is tailored to their environment. Deep roots can tap water beneath parched soils, whereas fine, shallow roots can exploit soils in which limiting nutrients are trapped at the surface.

Michelle Watt, a plant biologist at the Commonwealth Scientific and Industrial Research Organisation (CSIRO) in Camberra, is working to produce varieties of wheat that are better suited to drought-prone areas. In a recent study of wheat lines, Watts team found that the roots of some lines penetrate 25% deeper than others. The team crossed lines that had deeper, faster-growing roots with widely used cultivars to develop 400 new wheat lines, which are now being fidel-deared in Tolk; and Australia:

being field-tested in India and Australia. Wat is also taking advantage of new genetic tools. Rather than wade through the 17 billion base pairs of the breaching for genetic markers that are associated with deep roots in the much smaller (271 million base pair) genome of Bruchypodium distachyon, a temperate grass in the same subfamily as wheat whose genome was sequenced earlier this year. The team hopes that the markers will make it possible to identify, from seeds, which wheat varieties are likely to have deep roots, without going through the laborious process of growing the seedlings, digging them up and measuring their roots.

At Penn State, Lynch has found that, when water is limited, maize lines that incorporate a large amount of intercellular air space in root tissue have an eightfold higher yield than plants without this ability. When stressed, it may be that plants reduce the metabolic costs of

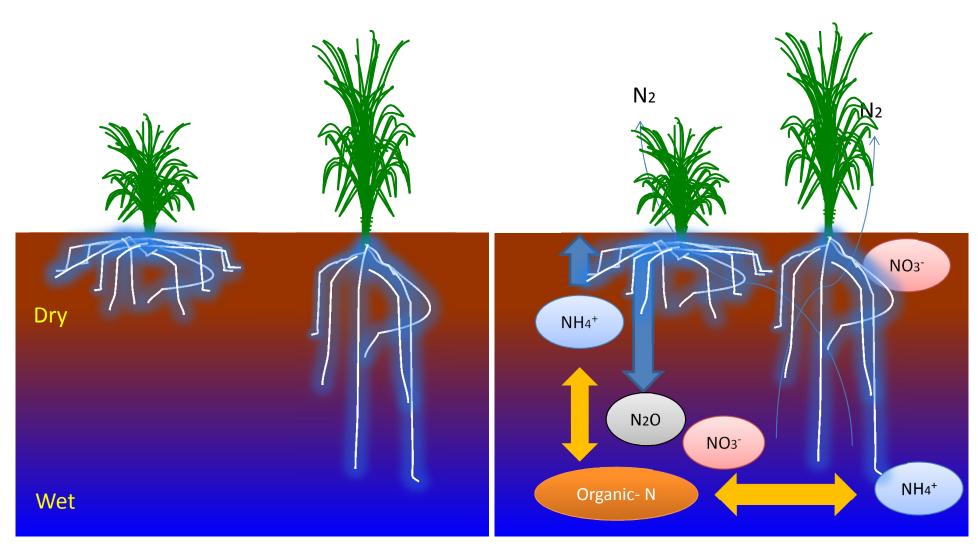
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# Increased productivity without environmental damages

- ✓ Water
- ✓ Plant nutrition
- ✓ Soil-plant-microbe interactions

552

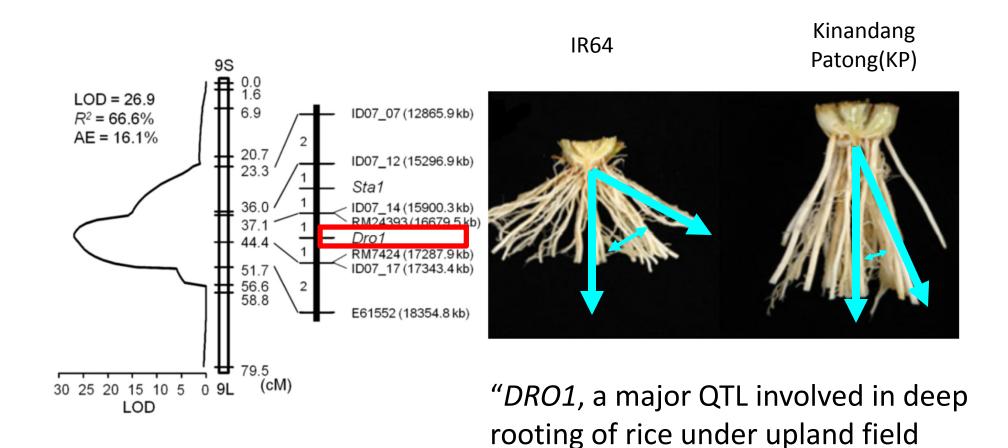
# What is an Ideal Root Type in Rice?



Water Use Efficiency (WUE)

Nitrogen Use Efficiency (NUE)

# Identification of Deeper Root Gene: Root Angle Concept

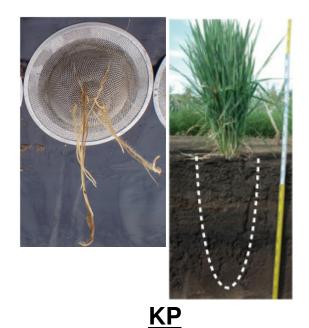


conditions" (Uga, 2013)

# Effect of *DRO1* Gene on Root Angle

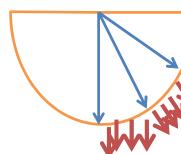






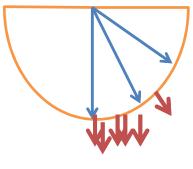
<u>IR64</u> RDR:

 $10.2 \pm 4.8$ 



**DRO1 NIL** 

RDR:  $47.1 \pm 10.2$ 



RDR:

 $77.8 \pm 9.6$ 

50%

10%

80%

### **DRO1** QTL Gene Improved Grain Yield under Water Limited Conditions

30





Rainout Shelter drought experiments



Santa Rosa rainfed experiment

Single plant yield 25 20 15 10 5 0 **IR64 IR64** IR64 Dro1 Dro1 **IR64** Dro1 Dro1 Dro1 **IR64** Rain-fed 2011 Rain-fed 2012 Drought Intermediate Wet

Rainout shelter Rainfed Ref: Uga et al. (2013)

# N Field Experiments at CIAT



#### **Measured traits**

#### At flowering period

N content of flag leaf (Kjeldahl), leaf chlorophyll content (SPAD value), Plant height

#### At harvesting period

Number of reproductive tiller

#### At post-harvesting (After drying)

Individual grain yield, individual plant biomass, panicle length and 1000 grain weight

#### **NDT traits**

NDT traits (Relative value) = measured value <sub>native</sub> / measured value <sub>FP</sub>

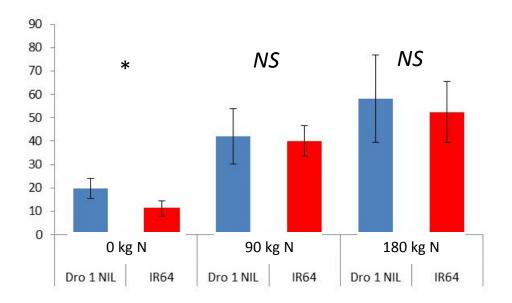
E.g.) RGY (Relative grain yield) = Individual grain yield  $_{native}$  / Individual grain yield  $_{FP}$ 

(Wei et al. 2012)

# DRO1 QTL Gene Improved Grain Yield under N Deficient Conditions



#### Single plant yield (g)



N experiments



Table 2 | Mean dry matter accumulation, grain yield, and harvest index in IR64 and Dro1-NIL lines as affected by line and fertilizer treatment

	Fertilizer treatment (F)	DWH <sup>a</sup>	DWM°	Yield <sup>b</sup>	Hle
Line (L)		(t ha <sup>-1</sup> )	(t ha <sup>-1</sup> )	(t ha <sup>-1</sup> )	
IR64		10.92	15.47	6.15	0.43
Dro1-NIL		11.24	16.27	6.80	0.45
	No treatment	8.86	12.21	4.71	0.43
	Treatment	13.30	19.52	8.24	0.45
ANOVA					
L		ns	*	**	**
F		**	**	**	ns
L×F		ns	ns	ns	ns

DWH, dry weight at heading; DWM, dry weight at maturity.

**N** experiments

Ref. Arai-Sanoh et al. (2014)

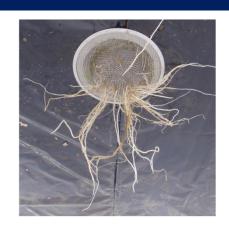
bYield is given for unhulled rice with water content of 15%.

<sup>\*</sup>Harvest index (HI) was calculated by dividing the dry weight of panicle by the dry weight of the above-ground parts.

<sup>\*</sup>Significant at P < 0.05; \*\* significant at P < 0.01; ns, not significant at P < 0.05. P values are based on ANOVA.

# Will Stronger Deep Root Contribute plant Performance?

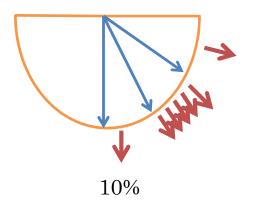


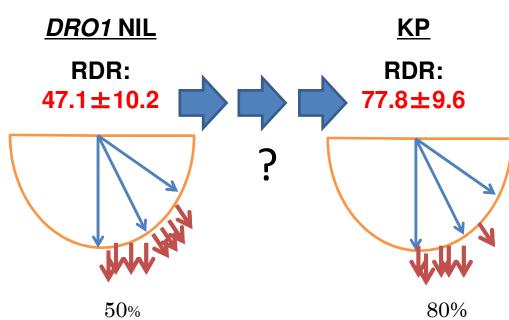




**IR64** 

RDR: 10.2±4.8



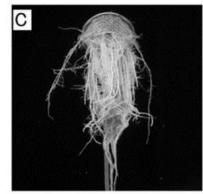


# Multi-Environmental Testing of Different Root Angle Variation

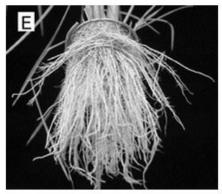
Fuente: Patent of Dr. Uga WO2011078308 A1



IR64
2; Vector control line



DRO1 NIL
2; DRO1
transgenic lines
WUE (single copy)



3; Multi-copy DRO1 transgenic lines



Transgenic rain-out shelter





Santa Rosa rainfed station

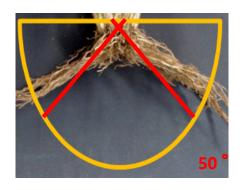


NUE

Nitrogen deficient field

### **Does Other Rooting Patterns Contribute?**

### **Diversity in Rice Rooting pattern**

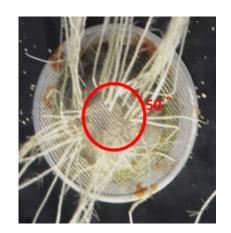


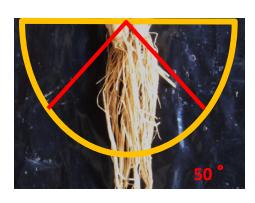
Monomorphic-Shallow (IR64) RPV =20.67



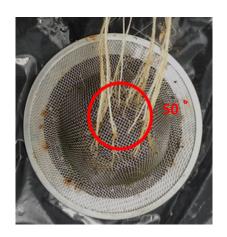


Dimorphic (*O. rufipogon*) RPV = 2.75





Monomorphic-Deep (Curinga) RPV = 12.33



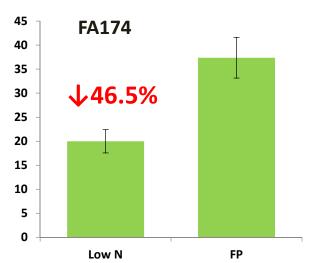
### Dimorphic is an Ideal Rooting Pattern for NUE

### **Monomorphic-Shallow**

#### 

# N experiments under lowland conditions

### **Dimorphic**

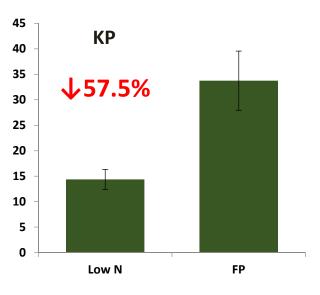


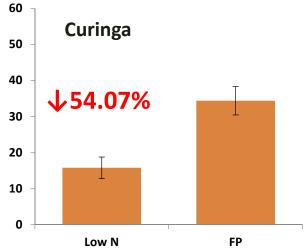
# O. rufipogon O. rufipogon O. rufipogon O. rufipogon O. rufipogon

Low N

FΡ

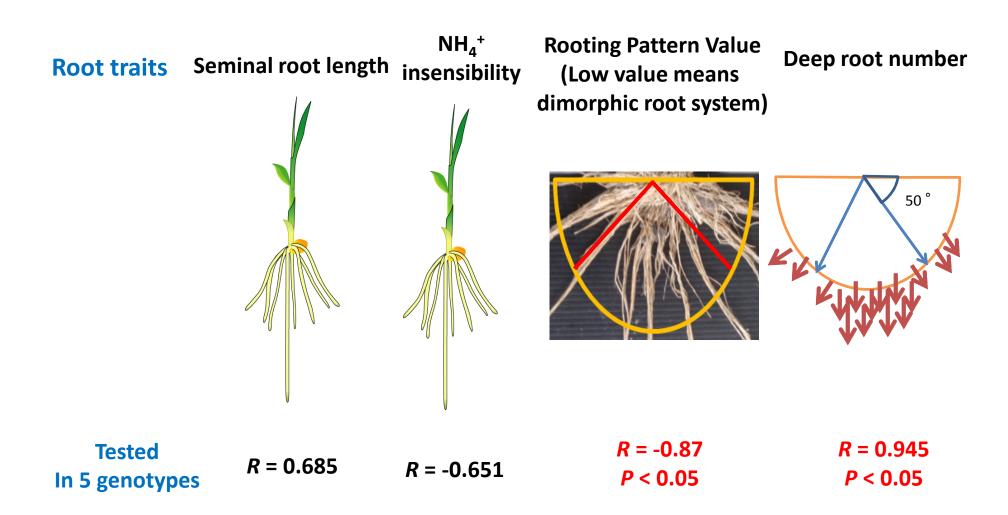
### **Monomorphic-deep**





# Dimorphic Root Contributed to Grain Yield under N Deficient Conditions

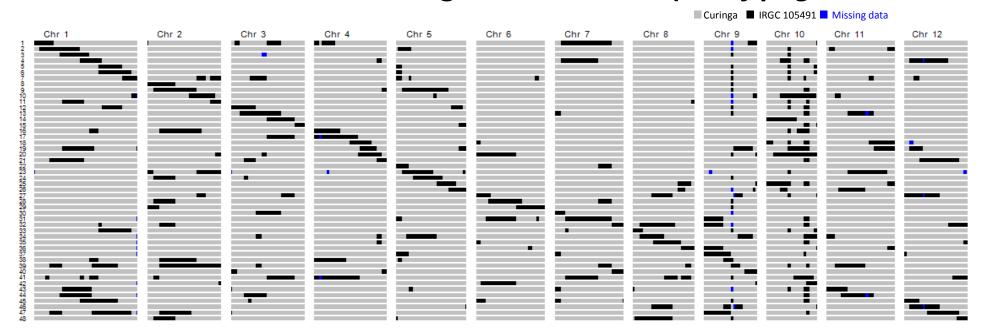
### Correlation between root traits and relative grain yield



# Identification of QTLs Regulating Root System Architecture for NUE

### **Materials**

### 48 CSSLs between Curinga x IRGA105491 (O. rufipogon)



QTLs analysis: CSSL finder (Lorieux M, 2005)

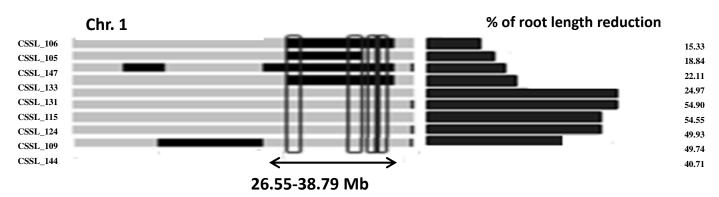




# QTLs Identified from *O. rufipogon* Regulating Root Traits

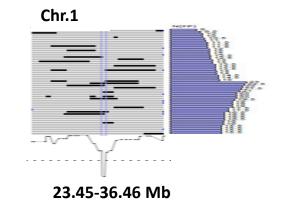
### **Seminal root elongation**

Seminal root length under 500  $\mu$ M NH<sub>4</sub><sup>+</sup> Relative seminal root length (500  $\mu$ M NH<sub>4</sub><sup>+</sup> / 5  $\mu$ M NH<sub>4</sub><sup>+</sup>)

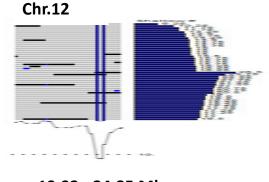


### Root growth angle

#### **Deeper root number**



#### **Shallow root number**



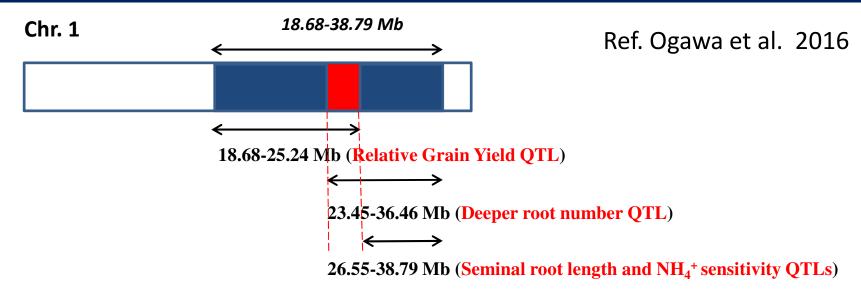
19.93 - 24.85 Mb

# QTLs Identified from *O. rufipogon* Regulating Agronomic NUE

### 8 agronomic QTLs and 3 Nitrogen Deficiency tolerance QTLs

Trait	Condition	Chr.	Marker	Position(Mb)	Positive allele	Season
Relative grain yield	NDT trait between Native and FP	1	id1010490-id103568	18.68 -25.24	O.rufipogon	Feb Jun.
Single plant yield	FP	3	id3002476-id3004123	4.32-7.68	O.rufipogon	Both trials
biomass	Native	4	id4005120-id4007907	17.68-24.36	O.rufipogon	Feb Jun.
1000 grain weight	Native and FP	5	Id5006603-id5012179	16.45-25.79	O.rufipogon	Both trials
Low SPAD value	FP	7	id7000142-id7000609	0.74-4.66	O.rufipogon	Both trials
Low Nitrogen content	FP	7	id7000142-id7000609	0.74-4.66	O.rufipogon	Feb Jun.
Relative N content	NDT trait between Native and FP	7	id7000142-id7000609	0.74-4.66	O.rufipogon	Feb Jun.
Relative SPAD value	NDT trait between Native and FP	8	id8000171	0.53	O.rufipogon	Feb Jun.
Higher tiller number	Native	9	id9000233-id9000580	0.88-10.75	O.rufipogon	Feb Jun.
Higher tiller number	FP	10	id1005370-id1006910	18.66-22.34	O.rufipogon	Aug Dec
Early flowering	Native and FP	12	id12003803-id12005677	9.54-16.74	O.rufipogon	Aug Dec

# Colocation of Three Agronomic NUE and Root Trait QTLs Identified in This Study



#### Reported QTLs in the same regions from other studies

Candidate gene name	Gene function	The Rice Annotation Project Database gene position	Reference	
PIN	Auxin efflux carrier component	Os01g0455500	Carraro et al. 2012	
IAA	Amino acid hydrolase homolog precursor (involved in auxin homeostasis)	Os01g0510600	Ding et al. 2008	
Tat protein	Twin-arginine translocation pathway signal domain containing protein	Os01g0456400	Fukao et al. 2011	
ARFs	Auxin responsive factor 3	Os01g0480600	Wang et al. 2009	
IAA8	Auxin-responsive protein	Os01g0484500	Groover et al. 2003	
IAA8	Auxin-responsive protein	Os01g0488500	Groover et al. 2003	
XPL1	Phosphoethanolamine N-methyltransferase	Os01g0500300	Luo et al. 2012	
OsGLT1	NADH-glutamate synthase	Os01g0681900	Goto et al. 1998	
OsAAT2 1 D14673	Aspartate aminotransferase	Os01g0760600	Song et al. 1996	

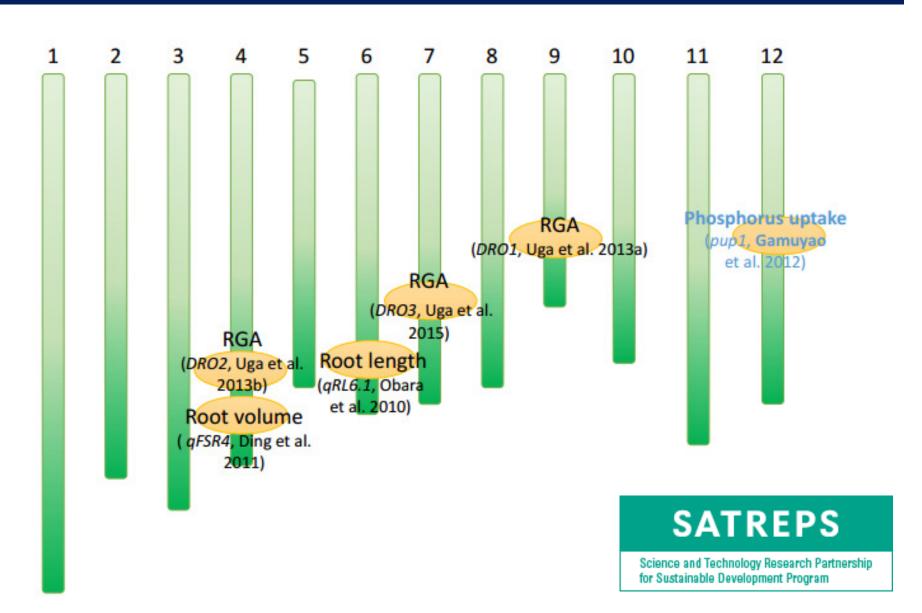
### Conclusions

 DRO1 gene can be ideal candidate for maker assisted breeding to improve water and Nitrogen use efficiency.

 Dimorphic root system showed better adaptation under water and N deficient conditions compare to monomorphic root systems

### **SATREPS Rice Project:**

### Ideal Root development by QTL pyramiding approach



### **Current Status of Root QTL Pyramiding**

	KP
Tiller number	4.0±0
Height	73.2±3.0
Deep	15.7±2.6
Shallow	13.5±2.8
RDR	53.8±3.3
MRL	26.8±2.3
Total	29.2±5.1
Shoot weight	1,150.0±151.0
Root weight	363.0±112.0
S/R	3.39±0.80
TRL	427.2±53.7



FA60
5.7±0.8
72.3±0.7
15.0±1.5
45.0±4.8
25.1±2.8
23.2±2.0
60.0±5.1
1555.3±100.2
501.2±122.0
3.30±0.82
762.2±99.8

Cross	F1	BC1F1	BC2F1	BC3F1	BC3F2	BC3F3
F 60	10	278 → 4	162 → 7	285	285	5
CT21375	10	239 → 3	162 → 4	258	→ ?	?
F 174	10	199 → 4	162 → 5	360	→?	?
F 473	10	267 → 4	162 → 7			

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Dr. Juan David Arbelaez



Natalia Espiñoza MSc



Dr. Mathias Lorieux









