

45
1967-2012

IITA

Research to Nourish Africa

R4D Review

Edition 8
March 2012



IITA R4D Review

Issue 8, March 2012

Editors

Lava Kumar and Katherine Lopez

Production team

Copy editor: Rose Umelo

Creative production: Godson Bright

Adegboyega Juba, and

Clement Ono-Raphael

Editorial board

Robert Asiedu, David Chikoye, and

Victor Manyong

The IITA R4D Review is published by IITA, Africa's leading research partner in finding solutions for hunger and poverty.

Copyright

IITA holds the copyright to its publications but encourages duplication of these materials for noncommercial purposes.

Proper citation is requested and modification of these materials is prohibited. Permission to make digital or hard copies of part or all of this work for personal or classroom use is hereby granted without fee and without a formal request provided that copies are not made or distributed for profit or commercial advantage and that copies bear this notice and full citation on the first page. Copyright for components not owned by IITA must be honored and permission pursued with the owner of the information. Prior specific permission is required to copy otherwise, to republish, to post on servers, or to redistribute to lists.

Disclaimer

We do not endorse, approve, or give any warranty on the content of this publication. All liability for loss and damage arising from your use of our content is excluded (except where death or personal injury arises from our negligence or loss or damage arises from any fraud on our part).

ISSN 2071-3681

Cover: Research worker in IITA's maize field collecting tassels for hand pollination.

Photo by C. Ono-Raphael.

contents

2 EDITOR'S NOTE

Mind the gap...

4 NEWS

Boosting yam productivity in Ghana and Nigeria

Pro-vitamin A cassava released

Multi-CGIAR center initiative launched

Plant Virology Symposium slated in 2013

6 FEATURES

Maize genetic improvement for enhanced productivity gains 6

A success tale on improving two legume crops in Africa 11

Breeding superior banana hybrids 16

Cassava improvement in the era of "agrigenomics" 21

Yam breeding at IITA 27

Genomics for yam breeding 31

A 'Green Revolution' in the West African cocoa belt 35

42 BEST PRACTICE

Partnerships as relationships for agricultural development

45 TOOL BOX

Afla-ELISA: a simple and low-cost quantitative test for the estimation of aflatoxins

48 WHO'S WHO

Nteranya Sanginga:

"Science can solve agriculture's problems"

52 LOOKING IN

Valerie Bemo:

"Collaboration required for major breakthroughs in African agriculture"

55 FRONTIERS

"Agrigenomics" for crop improvement 55

Transgenics in crop improvement research at IITA 58

Molecular diagnostic tools for plant health protection 61

Molecular diagnostic tools for plant health protection

Lava Kumar (L.kumar@cgiar.org)

Molecular tools in disease diagnosis

Rapid advancements in biotechnologies have led to the development of a myriad of molecular diagnostic tools in the past decade¹. These tools, either based on the properties of nucleic acid (DNA or RNA) or proteins of the target agents, have improved the efficacy, accuracy, and speed of detection and identification of disease-causing agents and characterization of the diversity of pathogens and pests.

Most popular protein detection methods depend on antigen-antibody interactions. Polyclonal or monoclonal antibodies produced against the proteins of interest are used as probes to detect the target proteins by techniques such as enzyme-linked immunosorbent assay (ELISA), Western immunoblotting, dot immunobinding assay, and several variants of these techniques. Meanwhile, nucleic



Researcher observing mouse hybridoma cell lines under microscope in the Virology and Molecular Diagnostics Unit, IITA, Ibadan, Nigeria. Photo by IITA.

acid-based diagnostic tools are based on the hybridization of homologous nucleotides, size of the DNA fragments generated by restriction enzyme treatment, order of nucleotide arrangement, or a combination of more than one of these

approaches. Polymerase chain reaction (PCR), developed in the mid-1980s, has led to the development of several new and simplified techniques, fast established as a mainstay of applied molecular biology and molecular diagnostics.

L. Kumar, Head of Germplasm Health Unit and Virologist, IITA, Ibadan, Nigeria.

Platform for developing molecular diagnostics

The objective of the molecular diagnostics research in IITA is to develop tools and technologies for better understanding, diagnosis, and monitoring of biological systems. This program emphasizes the development of simple and accurate tools and procedures for rapid identification of pathogens and pests affecting the food and horticultural crops in sub-Saharan Africa (SSA). Both protein and nucleic-acid based diagnostic tools have been developed against target agents (viruses, fungi, bacteria, phytoplasma, insect pests, and mycotoxins). These tools are critical to several programs on crop improvement and crop protection, including evaluation of germplasm for host resistance, breeding for pest and disease resistance, surveillance surveys, and monitoring programs.

ELISA-based diagnostics are preferred for the identification of plant viruses. It is simple, reliable, cost-effective, and easy to adopt in minimally-equipped labs. Backed with facilities for purifying proteins, and production of polyclonal

and monoclonal antibodies, ELISA-based diagnostics were established for about 20 economically important viruses affecting IITA's mandate crops in SSA (e.g., Maize streak virus, cassava mosaic begomoviruses, Cowpea mottle virus, Southern bean mosaic virus, and more). Antibodies were also produced against nonviral targets such as mycotoxins. Polyclonal antibodies produced against aflatoxin B1 were used to develop the 'Afla-ELISA' test for quantitative estimation of aflatoxins in maize and other commodities (see article on page 45). Monoclonal antibodies are usually produced for discriminating closely related virus species or strains (e.g., African cassava mosaic virus and East African cassava mosaic virus). The production of monoclonal antibodies is expensive and tedious, but it offers the advantage of perpetual production of antibodies from mouse hybridoma cell lines. Because of this, IITA has placed increasing emphasis on producing monoclonals for all important pathogens.

PCR-based diagnostics are developed as an alternative tool or to overcome the

limitations of ELISA in detecting viroids, viral satellites, and to discriminate strains and closely related species. Oligonucleotide primers have been developed based on the genomic data generated from our research programs and those available in the public database for the specific detection of targets in PCR assays. Procedures were also established to simplify PCR application. For instance, a procedure established for direct detection of viruses in leaf sap bypasses the need for nucleic extraction². Emphasis is placed on the development of multiplex PCR assays for the simultaneous detection of more than one virus in a single reaction. A multiplex PCR method has been developed for the simultaneous detection of African cassava mosaic virus and East African cassava mosaic like-viruses responsible for cassava mosaic disease in SSA². This test was further improved to detect cassava brown streak viruses that have emerged as a major threat to cassava in East Africa, thereby making it a one-stop test for detecting all the major viruses infecting cassava in SSA.

Similar efforts are being devised to detect all viruses infecting yam. Real-time PCR using Taqman™ probes are being developed to quantify virus concentrations within the plants to characterize host response to virus inoculation. Presently, specific and generic diagnostic tools for the detection of almost all the pathogens that affect major food staples in SSA have been established at IITA.

Pathogen diversity and DNA barcodes

Detailed knowledge of pathogen diversity is a prerequisite to developing unambiguous

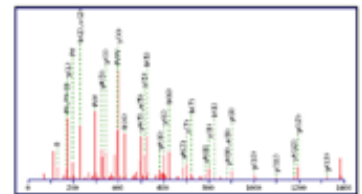
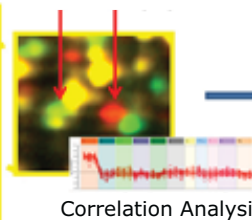
diagnostic tools. Pathogen populations are characterized by sequencing the specific genes and the data generated is used to interpret origin and spread of the pathogen, taxonomy, and phylogeny. For diversity assessment, gene targets are selected based on the pathogen that comprise, ribosomal Internal Transcribed Sequence (ITS), mitochondrial cytochrome oxidase-I (COI), histone, virus coat protein, etc. This approach has been used for assessing the diversity of *Colletotrichum*

gloeosporioides responsible for anthracnose of yam, *Cercospora* spp. causing gray leaf spot of maize, cassava brown streak virus, banana bunchy top virus, and several other agents. Information generated from these studies have provided valuable clues to understand the origin and drivers of spread, identification of previously uncharacterized pathogens^{3,4} and identification of unique markers known as "DNA barcodes" for use as genetic markers for identifying pathogens and pests⁵.

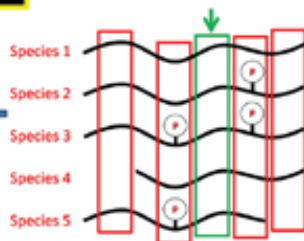
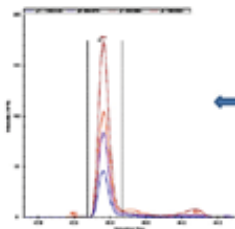
1. DIGE quantification

2. Statistics to find isoforms linked to transmission and all other isoforms in populations

3. Protein identification



4. DNA sequencing to identify polymorphisms linked to transmission phenotype



'INTSHVEYD**TASRHY**
'INTSHVEYDT RHY
'INTSHVEYD**TEL**RHY

6. Targeted proteomics assays to quantify peptides using a triple quadrupole mass spectrometer and validate using field-collected insect samples

5. Import sequences into Skyline to identify best peptides in vector populations to develop biomarker assay

Biomarkers for insect vectors

Recently a new initiative was started in collaboration with Cornell University to identify protein biomarkers to rapidly identify variation in vectoring potential of aphid and whitefly vector populations. Diagnostic tools developed in this program will aid in better understanding the virus-vector interactions, disease epidemiology, and improved management of insect vector-borne virus diseases.

Training in application of molecular diagnostics

In addition to technology development, efforts are made to transfer technology, products, and skills to stakeholders in national research and extension services. This is done through collaborative activities and organization of training courses at regular intervals in collaboration with national organizations such as the Nigerian Institute of Science Laboratory Technology. During the training courses, specific emphasis is placed on the application of diagnostics in monitoring and surveillance programs. Standard diagnostic protocols are compiled into a cook-

book style laboratory manual⁶ and distributed during the training courses.

End note

Molecular diagnostics development programs in IITA consider the latest knowledge and state-of-the-art technologies in establishing simple and robust tools that are relevant to end-users, are low-cost, and conducive for adoption in minimally equipped labs. We are adding new tools, such as, loop-mediated isothermal amplification reaction (LAMP) assay and deep sequencing approaches to broaden the knowledge on pathogens occurring in our mandate crops to increase the repertoire of available tools.

Molecular diagnostic tools are routinely used in germplasm indexing, phenotypic evaluation of germplasm, disease surveillance, and monitoring programs in SSA. They are also used in collecting baseline information and monitoring shifts in pathogen and pest dynamics due to changes in agricultural systems and climate change effect. These tools are already proving useful in rapid detection and identification of new and emerging pathogens

and pests [e.g., *Paracoccus marginatus* (papaya mealybug) in Nigeria; *Phytophthora colocasiae* causing taro leaf blight in Nigeria and Ghana; 16srII group phytoplasma responsible for witches' broom disease of soybean in Southern Africa; and Banana bunchy top virus in Benin].

References

- ¹Benali, S. et al. 2011. Advances of molecular markers application in plant pathology research. European Journal of Scientific Research. 50:110-123.
- ²Alabi, O.J. et al. 2008. Multiplex PCR method for the detection of *African cassava mosaic virus* and *East African cassava mosaic Cameroon virus* in cassava. Journal of Virology Methods. 154:111-120.
- ³Alabi, O.J. et al. 2010. Two new 'legumoviruses' (genus *Begomovirus*) naturally infecting soybean in Nigeria. Archives of Virology. 155:643-656.
- ⁴Sharma, K. et al. 2010. Genetically distinct *Cercospora* species cause grey leaf spot of maize (*Zea mays* L.) in Nigeria. Phytopathology 100 (6): S117.
- ⁵Kumar, P.L. and K. Sharma. 2010. DNA barcodes for pathogens of African food crops. R4D Review 4: 51-53. www.R4DReview.org.
- ⁶Kumar, P.L. (ed.). 2009. Methods for diagnosis of plant virus diseases: a laboratory manual. IITA, Ibadan, Nigeria. 90 pp.

A photograph of a man wearing a brown bucket hat and a checkered shirt, looking down at a corn cob he is holding in his hands. He is standing in a field of corn plants. The image is overlaid with a semi-transparent white filter.

Join the discussions online at
www.r4dreview.org

IITA

Research to Nourish Africa

ISSN: 2071-3681