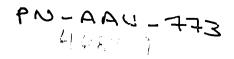
# OAT DESCRIPTORS







November 1985

# INTERNATIONAL BOARD FOR PLANT GENETIC RESOURCES

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OAT DESCRIPTORS

IBPGR Secretariat Kome 1985

The International Board for Plant Genetic Resources (IBPGR) is an autonomous international scientific organization under the aegis of the Consultative Group on International Agricultural Research (CGIAR). The IBPGR was established by the CGIAR in 1974 and its Executive Secretariat is provided by the Food and Agriculture Organization of the United Nations. The basic function of the IBPGR is to promote and of genetic coordinate international network an resources centres to further the collection. conservation, documentation, evaluation and use of plant germplasm and thereby contribute to raising the standard of living and welfare of people throughout the world. The Consultative Group mobilizes financial members to meet support from its the budgetary requirements of the Board.

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## PREFACE

1

This descriptor list for oat (<u>Avena</u> spp.) is based upon a list of descriptors selected by the Oat Working Group of the European Cooperative Programme for Conservation and Exchange of Crop Genetic Resources (ECP/GR) during its first meeting at Aegean Regional Agricultural Research Institute (ARARI), Izmir, Turkey, 25-27 September 1984. List of members of the Working Group is provided in Appendix I. Subsequently it was revised by members of the Working Group in the light of comments and criticisms received from oat scientists (listed in Appendix II). The IBPGR wishes to record its thanks to them for their contributions, which the Working Group has attempted to incorporate in the final text.

This descriptor list for <u>Avena</u> is sufficiently comprehensive and detailed to meet the needs of curators and breeders and to accommodate cultivated, weedy and wild forms and the IBPGR wishes to remind users that descriptors not relevant to their work may be disregarded.

The IBPGR encourages the collection of data on the first four categories of this list: 1. Accession; 2. Collection; 3. and 4. Characterization. The IBPGR endorses the information in categories 1-4 as the minimum that ideally should be available for any one accession. Other descriptors are given in categories 5 onwards that will enable the simple encoding of further characterization and evaluation data and which can serve as examples for the creation of additional descriptors in the IBPGR form for any user.

Although the suggested coding should not be regarded as the definite scheme, this format has the full backing of the IBPGR and is promoted world-wide. The descriptor list given here provides an international format and thereby produces a universally understood 'language' for all plant genetic resources data. The adoption of this scheme for all data encoding, or at least the production of a transformation method to convert other schemes to the IBPGR format, will produce a rapid, reliable and efficient means for information storage, retrieval and communication. This will greatly assist the utilization of germplasm throughout the international plant genetic resources network. It is recommended, therefore, that information should be produced by closely following this descriptor list with to: ordering and numbering the descriptors; using regard the descriptors specified; and using the descriptor states recommended.

Any suggestions for modifications will be welcomed by the IBPGR Secretariat, Rome.

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#### DESCRIPTOR LIST FOR OAT

The IBPGR now uses the following definitions in genetic resources documentation:

- (i) <u>passport</u> (accession identifiers and information recorded by collectors);
- (ii) <u>characterization</u> (consists of recording those characters which are highly heritable, can be easily seen by the eye and are expressed in all environments);
- (iii) <u>preliminary evcluation</u> (consists of recording a limited number of additional traits thought desirable by a consensus of users of the particular crop).

Characterization and preliminary evaluation will be the responsibility of the curators, while further characterization and evaluation should be carried out by the plant breeder. The data from further evaluation should be fed back to the curator who will maintain a data file.

The following internationally accepted norms for the scoring or coding of descriptor states should be followed as indicated below:

- (a) measurements are made according to the SI system. The units to be applied are given in brackets following the descriptor;
- **(b)** many descriptors which are continuously variable are recorded on a 1-9 scale. The authors of this list have sometimes described only a selection of the states, e.g. 3. 5 ad 7 for such descriptors. Where this has occurred the full range of codes is available for use by extension of the codes given or by interpolation between them e.g. --in Section 8 (Pest and disease susceptibility) 1 = extremely low susceptibility and 8 = high to extremely high susceptibility;
- (c) presence/absence of characters are scored as + (present) and 0 (absent);
- (d) for descriptors which are not generally uniform throughout the accession (e.g. mixed collection, genetic segregation) mean and standard deviation could be reported where the descriptor is continuous or mean and 'x' where the descriptor is discontinuous;

(e) when the descriptor is inapplicable, '0' is used as the descriptor value, e.g. if an accession does not form flowers, 0 would be scored for the following descriptor

Flower colour

1	White
2	Yellow
3	Red
4	Purple

- (f) blanks are used for information not yet available;
- (g) standard colour charts, e.g. Royal Horticultural Society Colour Chart, Methuen Handbook of Colour, Munsell Color Charts for Plant Tissues are strongly recommended for all ungraded colour characters (the precise chart used should be specified in the NOTES descriptor, 11);
- (h) dates should be expressed numerically in the format DDMMYYYY, where

DD - 2 digits to represent the day MM - 2 digits to represent the month YYYY - 4 digits to represent the year

In many characters it is necessary that their expression should be recorded at a particular stage in the development of the plant. Appropriate stages are suggested in sections 4 and 6 using the decimal code of Zadoks, Chang and Konzak  $\frac{1}{4}$  and the key to the code is reproduced on pages 3 and 4.

<sup>1/</sup> Zadoks, J.C., Chang T.T., and Konzak, C.F. 1974. <u>A Decimal Code for the Growth</u> <u>Stages of Cereals</u>. European Association for Research on Plant Breeding (EUCARPIA), Wageningen, Netherlands.

2-d cod	igit General duscription e	2d cod	igit General description e
	Germination		Stem elongation
00	Dry seed	30	Pseudo stem erection
10	Start of imbibition	31	lst node detectable
02	-	32	2nd node detectable
03	Imbibition complete	33	3rd node detectable
04	-	34	4th node detectable
05	Radicle emerged from caryopsis	35	5th node detectable
06	-	36	6th node detectable
07	Coleoptile emerged from caryopsis	37	Flag leaf just visible
08	-	38	<u> </u>
09	Leaf just at coleoptile tip	39	Flag leaf ligule/collar just visible
	Seedling growth		Booting
10	First leaf through coleoptile	40	-
11	First leaf unfolded 1/	41	Flag leaf sheath extending
12	2 leaves unfolded	42	-
13	3 leaves unfolded	43	Boots just visibly swollen
14	4 leaves unfolded	44	-
15	5 leaves unfolded	45	Boots swollen
16	6 leaves unfolded	46	-
17	7 leaves unfolded	47	Flag leaf sheath opening
18	8 leaves unfolded	48	-
19	9 or more leaves unfolded	49	First awns visible
	Tillering		Inflorescence emergence
20	Main shoot only	50 <sub>}</sub>	First spikelet of K
21	Main shoot and I tiller	51	inflorescence just visible <sup>1</sup> S
22	Main shoot and 2 tillers	52 լ	1/4 of inflorescence N
23	Main shoot and 3 tillers	53 <sup>}</sup>	emerged S
24	Main shoot and 4 tillers	54 კ	1/2 of inflorescence N
25	Main shoot and 5 tillers	55 <sup>(</sup>	emerged <sup>i</sup> S
26	Main shoot and 6 tillers	56 լ	3/4 of inflorescence N
27	Main shoot and 7 tillers	57 <sup>7</sup>	emerged <sup>1</sup> S
28	Main shoot and 8 tillers	58 t	Emergence of inflorescence IN
29	Main shoot and 9 or more tillers	59 <sup>}</sup>	completed S

Table 1. A decimal code for the growth stages of cereals

 $\underline{I}$  see next page

2-d code	igit General description a	2-digit General description code
	Antithesis	Ripening
60 61 62 63 64 65 66 67 68 69 70 71 72 73	Beginning of anthesis {N Beginning of anthesis S - - Anthesis half-way {N Anthesis half-way S - - Anthesis complete {N	<ul> <li>90 -</li> <li>91 Caryopsis hard (difficult to divide by thumb-nail) 2/</li> <li>92 Caryopsis hard (can no longer be dented by thumb- nail) 3/</li> <li>93 Caryopsis loosening in daytime</li> <li>94 Over-ripe, straw dead and collapsing</li> <li>95 Seed dormant</li> <li>96 Viable seed giving 50% germination</li> <li>97 Seed not dormant</li> <li>98 Secondary dormancy induced</li> <li>99 Secondary dormancy lost</li> </ul>
74 75 76 77 78 79	- Medium milk - Late milk - - <u>Dough development</u>	
80 81 82 83 84 85 86 87 88 89	- - Early dough - Soft dough - Hard dough -	

Table I. A decimal code for the growth stages of cereals (Continued)

N = non-synchronous crops

S = synchronous

- 1/ Stage of seedling inoculation with rust in the glasshouse
- 2/ Ripeness for binder (ca 16% water content); chlorophyll of inflorescence largely lost
- 3/ Ripaness for combine harvester (16% water content)

#### PASSPORT

# 1. ACCESSION DATA

#### 1.1 ACCESSION NUMBER

This number serves as a unique identifier for accessions and is assigned by the curator when an accession is entered into his collection. Once issigned this number should never be reassigned to another accession in the collection. Even if an accession is lost, its assigned number is still not available for re-use. Letters should occur before the number to identify the genebank or national system (e.g. MG indicates an accession comes from the genebank at Bari, Italy; PI indicates an accession within the USA system)

#### 1.2 DONOR NAME

Name of institution or individual responsible for donating the germplasm

1.3 DONOR IDENTIFICATION NUMBER

Number assigned to accession by the donor

1.4 OTHER NUMBERS ASSOCIATED WITH THE ACCESSION (other numbers can be added as 1.4.3, etc.)

Any other identification number known to exist in other collections for this accession, e.g. USDA Plant Inventory number (<u>not</u> collection number, see 2.1)

1.4.1 Other number 1

1.4.2 Other number 2

- 1.5 SCIENTIFIC NAME
  - 1.5.1 <u>Genus</u>
  - 1.5.2 Species
  - 1.5.3 <u>Subspecies</u>
  - 1.5.4 Botanical variety
  - 1.5.5 <u>Cultivar group</u>
- 1.6 PEDIGREE/CULTIVAR NAME

Nomenclature and designations assigned to breeder's material

#### 1.7 ACQUISITION DATE

The date in which the accession entered the collection

- 1.8 DATE OF LAST REGENERATION OR MULTIPLICATION
- 1.9 ACCESSION SIZE

Approximate number of seeds of accession in collection

1.10 NUMBER OF TIMES ACCESSION REGENERATED

Number of regenerations or multiplications since original collection

1.11 NUMBER OF PLANTS GROWN DURING LAST REGENERATION

#### 2. COLLECTION DATA

2.1 COLLECTOR'S NUMBER

Original number assigned by collector of the sample normally composed of the name or initials of the collector(s) followed by a number. This item is essential for identifying duplicates held in different collections and should always accompany sub-samples wherever they are sent

#### 2.2 COLLECTING INSTITUTE

Institute or person collecting/sponsoring the original sample

- 2.3 DATE OF COLLECTION OF ORIGINAL SAMPLE
- 2.4 COUNTRY OF COLLECTION OR COUNTRY WHERE CULTIVAR/VARIETY BRED

Use the 3 letter abbreviations supported by the Statistical Office of the United Nations. Copies of these abbreviations are available from the IBPGR Secretariat and have been published in the FAO/IBPGR Plant Genetic Resources Newsletter number 49

#### 2.5 PROVINCE/STATE

Name of the administrative subdivision of the country in which the sample was collected

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2.6 LOCATION OF COLLECTION SITE

Number of kilometres and direction from nearest town, village or map grid reference (e.g. TIMBUKTU /S means 7 km south of Timbuktu)

2.7 LATITUDE OF COLLECTION SITE

Degrees and minutes followed by N (north) or  $\mathcal{S}$  (south), e.g. 1030S

2.8 LONGITUDE OF COLLECTION SITE

Degrees and minutes followed by E (east) or W (west), e.g. 7625 W

2.9 ALTITUDE OF COLLECTION SITE (m)

Elevation above sea level

- 2.10 COLLECTION SOURCE
  - 1 Wild
  - 2 Farm land
  - 3 Farm store
  - 4 Backyard
  - 5 Village market
  - 6 Commercial market
  - 7 Institute
  - 8 Other (specify in the NOTES descriptor, 11)

## 2.11 STATUS OF SAMPLE

- 1 Wild
- 2 Weedy
- 3 Breeder's line
- 4 Primitive cultivar (landrace)
- 5 Inbred line
- 6 Hybrid
- 7 Synthetic
- 8 Other (specify in the NOTES descriptor, 11)

### 2.12 LOCAL/VERNACULAR NAME

Name given by farmer to cultivar/landrace/weed

2.13 NUMBER OF PLANTS SAMPLED

Approximate number of plants collected in the field to produce this accession

#### 2.14 PHOTOGRAPH

Any identification of a photograph of the accession or environment taken at collection. If no photograph was taken, record '0'

#### 2.15 HERBARIUM SPECIMEN

Was a herbarium specimen collected?

- 0 No
- + Yes

#### 2.16 TOPOGRAPHY OF COLLECTION SITE

- 1 Swamp
- 2 Flood plain
- 3 Plain level
- 4 Undulating
- 5 Hilly
- 6 Mountainous
- 7 Other (specify in the NOTES descriptor, 11)

#### 2.17 SOIL TEXTURE AT COLLECTION SITE

- 1 Organic
- 2 Clay
- 3 Loam
- 4 Sand
- 5 Rocky

#### 2.18 PESTS AND DISEASES AT COLLECTION TIME

Specify using item numbers of pests and diseases (Section 8) and severity of infection on 1-9 scale

2.19 OTHER NOTES FROM COLLECTOR

Collectors may record ecological information. For cultivated crops, cultivation practices such as irrigation, season of sowing, etc. may be recorded

# CHARACTERIZATION

- 3. <u>SITE DATA</u>
  - 3.1 COUNTRY OF CHARACTERIZATION
  - 3.2 SITE (RESEARCH INSTITUTE)
  - 3.3 NAME OF PERSON IN CHARGE OF CHARACTERIZATION
  - 3.4 SOWING DATE
  - 3.5 HARVEST DATE
  - 3.6 CULTIVATION METHOD

Record row spacing and other management practices

## 4. PLANT DATA

### Stage

- 4.1 VEGETATIVE
  - 4.1.1 Chromosome number
  - 4.1.2 Growth class (seasonality)
    - 1 Autumn sown
    - 2 Facultative
    - 3 Spring sown

# 4.1.3 Vernalization requirement

- 0 Vernalization not required
- + Vernalization required

# 4.1.4 Growth habit

At juvenile stage, angle of the tillers from the vertical

- 3 Erect
- 5 Semiprostrate
- 7 Prostrate

# 4.1.5 Plant height

80-92

25-29

Relative to specified reference varieties

- 3 Short
- 5 Medium
- 7 Tall

		<u>Stage</u>
4.1.6	Stem thickness	80-92
	Relative to specified reference varieties	
	3 Thin	
	5 Intermediate 7 Thick	
4.1.7	Nodes hairiness	60-69
	0 Glabrous	
	3 Slightly pubescent	
	5 Moderately pubescent 7 Highly pubescent	
	/ Highly pubescenc	
4.1.8	Angle of flag leaf to culm	50-53
	3 Acute < 90°	
	5 Intermediate about 90°	
	7 Obtuse > 90°	
4.1.9	Rigidity of flag leaf	50-53
	3 Bent	
	5 Slightly bent	
	7 Stiff	
4.1.10	Angle to culm of leaves (other	50-53
	than flag leaf)	
	3 Acute < 90°	
	5 Intermediate about 90°	
	7 Obtuse > 90°	
4.1.11	Rigidity of leaves (other than flag leaf)	50-53
	3 Bent	
	5 Slightly bent	
	7 Stiff	
4.1.12	2 <u>Hairiness of leaf sheath</u> (lower leaves)	25-29
	0 Glabrous	
	3 Slightly pubescent	
	5 Moderately pubescent	
	7 Highly pubescent	

			<u>Stage</u>
	4.1.1	3 <u>Hairiness of leaf margin</u> (leaf below flag leaf)	40-4
		<ul> <li>Glabrous</li> <li>Slightly pubescent</li> <li>Moderately pubescent</li> <li>Highly pubescent</li> </ul>	
4.2	INFLO	RESCENCE AND FRUIT	
	4.2.1	Shape of panicle	65
		1 Urilateral 2 Equilateral	
	4.2.2	Erectness of panicle	70-79
		3 Drooping 5 Semi-erect 7 Erect	
	4.2.3	Angle of panicle branches to the main axis	70-79
		<ol> <li>Extremely low (panicle branches erect)</li> <li>Acute</li> <li>Intermediate</li> <li>Obtuse</li> </ol>	
	4.2.4	Erectness of spikelets	60-65
		3 Drooping 5 Semi-erect 7 Erect	
	4.2.5	Waxiness of the panicle	64-69
		0 Absent + Present	
4.3	SEED		
	4.3.1	Lemma colour	92
		<ol> <li>White</li> <li>Yellow</li> <li>Grey</li> <li>Red</li> <li>Black</li> <li>Other (specify in the NOTES descriptor, 1</li> </ol>	1)

	<u>S1</u>	age
4.3.2	<u>Hairiness of lemma</u>	92
	<ul> <li>Glabrous</li> <li>Slightly pubescent</li> <li>Moderately pubescent</li> <li>Highly pubescent</li> </ul>	
4.3.3	Kernel covering (cultivated forms)	92
	0 Grains naked + Grains covered	
4.3.4	Awnedness	92
	Recorded on basal floret	
	0 No awns 3 Weak awns 7 Strong awns	
4.3.5	<u>Awn type</u> (for wild and weedy species)	92
	Recorded on basal floret	
	<ol> <li>Straight</li> <li>Geniculate</li> <li>Other (specify in the NOTES descriptor, 11)</li> </ol>	
4.3.6	<u>Position of awn insertion</u> (for wild and weedy species)	92
	Recorded on basal floret	
	<pre>1 1/4 from base 2 1/3 from base 3 1/2 from base 4 &gt; 1/2 from base</pre>	
4.3.7	<u>Dispersal unit</u> (for wild and weedy species)	<del>9</del> 2
	<ol> <li>Spikelet, minus glumes (abcission layer base of the lowest floret only)</li> <li>Floret (abcission layer at base of each flored)</li> </ol>	
4.3.8	<u>Hairiness at basal part of the primary grain</u>	92
	0 Glabrous 3 Slightly pubescent	

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- 5 Moderately pubescent 7 Highly pubescent

#### FURTHER CHARACTERIZATION AND EVALUATION

- 5. <u>SITE DATA</u>
  - 5.1 COUNTRY OF FURTHER CHARACTERIZATION AND EVALUATION
  - 5.2 SITE (RESEARCH INSTITUTE)
  - 5.3 NAME OF PERSON IN CHARGE OF EVALUATION
  - 5.4 SOWING DATE
  - 5.5 HARVEST DATE
  - 5.6 CULTIVATION METHOD

Record row spacing and other management practices

# 6. PLANT DATA

6.1	VEGETA	EGETATIVE S		
	6.1.1	Length of second leaf from top	50-69	
		Relative to specified reference varieties		
		3 Short 5 Medium 7 Long		
	6.1.2	Width of second leaf from top	50-69	
•**		At widest point. Relative to specified refe varieties	rence	
		3 Narrow 5 Medium 7 Broad		
	6.1.3	Number of tillers		
		Relative to specified referer ce varieties		
		3 Few 5 Intermediate 7 Many		
	6.1.4	Number of fertile tillers	92	

An average of 10 plants

			Stage	
	6.1.5	Lodging at immature stage	60-79	
		0 Upright (all plants) 3 Minor lodging 5 Intermediate 7 Lodged 9 Extremely lodged (all plants)		
	6.1.6	Lodging at mature stage	92	
		<ul> <li>Upright (all plants)</li> <li>Minor lodging</li> <li>Intermediate</li> <li>Lodged</li> <li>Extremely lodged (all plants)</li> </ul>		
6.2	INFLOR	ESCENCE AND FRUIT		
	6.2.1	Days to heading	54-55	
		Counted as days from sowing to 50% of panicl fully emerged	es	
	6.2.2	Days to harvest	92	
		Counted as days from sowing to harvest ripen	ess	
	6.2.3	<u>Relation between maturity of grains and straw</u>	80-89	
		<ul> <li>Grain ripe before straw</li> <li>Simultaneous</li> <li>Straw ripe before grain</li> </ul>		
	6.2.4	Number of seeds in panicle	92	
		An average of 10 panicles		
	6.2.5	Number of grains in spikelet	92	
		An average of 5 spikelets		
6.3	SEED			
	6.3.1	Grain shedding at maturity (cultivated forms)	92	
		3 Low 5 Intermediate 7 High		

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#### Stage

6.3.2	<u>1000 grain weight</u> (g)	92
6.3.3	Test weight (kg/hl)	92
6.3.4	Percentage of husk (%)	92
6.3.5	Percentage protein content of caryopsis (%)	92
	Record details of analytical method used in the NOTES descriptor, 11	
6.3.6	Percentage oil content of caryopsis (%)	92
	Record details of analytical method used in the NOTES descriptor, 11	

#### 6.3.7 Sprouting

Tendency of grains to sprout in the ear before harvest as a result of late rainfall

- 0 No sprouting
- 9 All sprouting

# 7. <u>STRESS SUSCEPTIBILITY</u>

Scored on a 1-9 scale, where

- 3 Low susceptibility
- 5 Medium susceptibility
- 7 High susceptibility

#### 7.1 LOW TEMPERATURE DAMAGE

Damage caused by cold to aerial parts of plants (distinct from winter kill of whole plants, see descriptor 7.5)

- 0 No damage
- 3 Slightly damaged
- 5 Moderately damaged
- 7 Highly damaged
- 9 All aerial parts killed
- 7.2 HIGH TEMPERATURE
- 7.3 DROUGHT

- 7.4 HICH SOIL MOISTURE
- 7.5 WINTER KILL

Susceptibility to winter stress measured as a loss of plants in a sowing

- 0 No loss
- 9 All plants dead
- 7.6 SALINITY
- 7.7 LOW PH
- 8. <u>PEST AND DISEASE SUSCEPTIBILITY</u>

These reactions are coded on a 1-9 scale with 9 representing maximum susceptibility. In each case, it is important to state the origin of the infection or infestation, i.e. natural, field inoculation, laboratory test, and pathotype or physiologic race used. Record such information in the NOTES descriptor, 11

8.1 PESTS

	8.1.1	<u>Ditylenchus dipsaci</u>	Stem and bulb eelworm
	8.1.2	<u>Heterodera avenae</u>	Root eelworm
	8.1.3	<u>Lama melanopa</u>	Leaf beetle
	8.1.4	<u>Oscinella frit</u>	Fruit fly
8.2	FUNGI		
	8.2.1	<u>Erysiphe graminis avenae</u>	Powdery mildew
	8.2.2	Drechslera spp.	
	8.2.3	<u>Puccinia coronata avenae</u>	Crown rust
	8.2.4	<u>Puccinia graminis avenae</u>	Stem rust
	8.2.5	<u>Septoria avenae</u>	
	8.2.6	<u>Ustilago avenae</u>	Loose smut
	8.2.7	<u>Ustilago kolleri</u>	Covered smut
.0.2			

'8.3 BACTERIA

- 8.4 VIRUS
  - 8.4.1 Barley yellow dwarf virus (BYDV)

8.4.2 Oat sterile dwarf virus

# 9. <u>ALLOENZYME COMPOSITION</u>

This may prove to be a useful tool for identifying duplicate accessions

# 10. CYTOLOGICAL CHARACTERS AND IDENTIFIED GENES

11. NOTES

Give additional information where descriptor is noted as "Other" as, for example, in descriptor 4.3.1. Also include here any further relevant information

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