



Review

Plant sterols and plant stanols in the management of dyslipidaemia and prevention of cardiovascular disease [☆][☆]



Helena Gylling ^{a,1}, Jogchum Plat ^{b,1}, Stephen Turley ^{c,1}, Henry N. Ginsberg ^{d,*,1}, Lars Ellegård ^e, Wendy Jessup ^f, Peter J. Jones ^g, Dieter Lütjohann ^h, Winfried Maerz ^{i,j,k}, Luis Masana ^l, Günther Silbernagel ^m, Bart Staels ⁿ, Jan Borén ^o, Alberico L. Catapano ^{p,q}, Guy De Backer ^r, John Deanfield ^s, Olivier S. Descamps ^t, Petri T. Kovanen ^u, Gabriele Riccardi ^v, Lale Tokgözoğlu ^w, M. John Chapman ^{x,y}, for the European Atherosclerosis Society Consensus Panel on Phytosterols²

^a Department of Medicine, Division of Internal Medicine, University of Helsinki, Biomedicum Helsinki, Finland

^b Department of Human Biology, Maastricht University, The Netherlands

^c Department of Internal Medicine, UT Southwestern Medical Center, Dallas, USA

^d Irving Institute for Clinical and Translational Research, Columbia University, New York, USA

^e Department of Internal Medicine and Clinical Nutrition, Sahlgrenska Academy at University of Gothenburg, Sweden

^f ANZAC Research Institute, Concord Hospital, Sydney, Australia

^g Richardson Centre for Functional Foods and Nutraceuticals, University of Manitoba, Winnipeg, Canada

^h Institute of Clinical Chemistry and Clinical Pharmacology, University Clinics Bonn, Germany

ⁱ Synlab Academy, Synlab LLC, Mannheim Center of Laboratory Diagnostics, Heidelberg, Germany

^j Mannheim Institute of Public Health, Social and Preventive Medicine, Medical Faculty Mannheim, University of Heidelberg, Germany

^k Clinical Institute of Medical and Chemical Laboratory Diagnostics, Medical University of Graz, Austria

^l Vascular Medicine and Metabolism Unit, University Hospital Sant Joan, IISPV, CIBERDEM, Rovira and Virgili University, Reus, Spain

^m Department of Angiology, Swiss Cardiovascular Center, Inselspital, University of Bern, Switzerland

ⁿ Faculty of Pharmacy, University of Lille 2, Lille, France

^o Strategic Research Center, Sahlgrenska Center for Cardiovascular and Metabolic Research (CMR), University of Gothenburg, Sweden

^p Department of Pharmacological and Biomolecular Sciences, University of Milan, Italy

^q Multimedita IRCSS Milano, Italy

^r Department of Public Health, University Hospital Ghent, Ghent, Belgium

^s National Centre for Cardiovascular Prevention and Outcomes, University College London, UK

^t Lipid Clinic, Hopital de Jolimont, Haine-Saint-Paul, Belgium

^u Wihuri Research Institute, Helsinki, Finland

^v Department of Clinical and Experimental Medicine, Federico II University, Naples, Italy

^w Department of Cardiology, Hacettepe University, Ankara, Turkey

^x University of Pierre and Marie Curie, Paris, France

^y Dyslipidaemia and Atherosclerosis Research Unit, INSERM U939, Pitié-Salpêtrière University Hospital, Paris, France

[☆][☆] This European Atherosclerosis Society (EAS) Consensus Panel dedicates this Position paper to the memory of Professor Tatu Miettinen, a pioneer in the study of the impact of plant sterols/stanols on cholesterol absorption and homeostasis.

* Corresponding author. Irving Professor of Medicine and Director, Irving Institute for Clinical and Translational Research Columbia University, 622 West 168 Street, PH-10, New York 10032, USA. Tel.: +1 212 305 9562; fax: +1 212 305 3213.

E-mail addresses: Helena.Gylling@hus.fi (H. Gylling), j.plat@maastrichtuniversity.nl (J. Plat), Stephen.Turley@UTSouthwestern.edu (S. Turley), hng1@columbia.edu (H.N. Ginsberg), lasse.ellegard@nutrition.gu.se (L. Ellegård), wendy.jessup@gmail.com (W. Jessup), Peter.Jones@umanitoba.ca (P.J. Jones), Dieter.Luetjohann@ukb.uni-bonn.de (D. Lütjohann), Winfried.Maerz@synlab.com (W. Maerz), luis.masana@urv.cat (L. Masana), guenther.silbernagel@yahoo.com (G. Silbernagel), Bart.Staels@pasteur-lille.fr (B. Staels), Jan.Boren@wlab.gu.se (J. Borén), alberico.catapano@unimi.it (A.L. Catapano), guy.debacker@ugent.be (G. De Backer), john.deanfield@gmail.com (J. Deanfield), olivierdescamps@hotmail.com (O.S. Descamps), Petri.Kovanen@wri.fi (P.T. Kovanen), riccardi@unina.it (G. Riccardi), lale@hacettepe.edu.tr (L. Tokgözoğlu), john.chapman@upmc.fr (M.J. Chapman).

¹ These authors contributed equally.

² Affiliations are listed here and in Author contribution.

ARTICLE INFO

Article history:

Received 11 November 2013

Accepted 11 November 2013

Available online 23 November 2013

Keywords:

Hypercholesterolaemia

Phytosterols

Intestinal cholesterol absorption

Cardiovascular risk

Safety

Functional foods with added plant sterols
and plant stanols

ABSTRACT

Objective: This EAS Consensus Panel critically appraised evidence relevant to the benefit to risk relationship of functional foods with added plant sterols and/or plant stanols, as components of a healthy lifestyle, to reduce plasma low-density lipoprotein-cholesterol (LDL-C) levels, and thereby lower cardiovascular risk.

Methods and results: Plant sterols/stanols (when taken at 2 g/day) cause significant inhibition of cholesterol absorption and lower LDL-C levels by between 8 and 10%. The relative proportions of cholesterol versus sterol/stanol levels are similar in both plasma and tissue, with levels of sterols/stanols being 500-/10,000-fold lower than those of cholesterol, suggesting they are handled similarly to cholesterol in most cells. Despite possible atherogenicity of marked elevations in circulating levels of plant sterols/stanols, protective effects have been observed in some animal models of atherosclerosis. Higher plasma levels of plant sterols/stanols associated with intakes of 2 g/day in man have not been linked to adverse effects on health in long-term human studies. Importantly, at this dose, plant sterol/stanol-mediated LDL-C lowering is additive to that of statins in dyslipidaemic subjects, equivalent to doubling the dose of statin. The reported 6–9% lowering of plasma triglyceride by 2 g/day in hypertriglyceridaemic patients warrants further evaluation.

Conclusion: Based on LDL-C lowering and the absence of adverse signals, this EAS Consensus Panel concludes that functional foods with plant sterols/stanols may be considered 1) in individuals with high cholesterol levels at intermediate or low global cardiovascular risk who do not qualify for pharmacotherapy, 2) as an adjunct to pharmacologic therapy in high and very high risk patients who fail to achieve LDL-C targets on statins or are statin-intolerant, 3) and in adults and children (>6 years) with familial hypercholesterolaemia, in line with current guidance. However, it must be acknowledged that there are no randomised, controlled clinical trial data with hard end-points to establish clinical benefit from the use of plant sterols or plant stanols.

© 2013 The Authors. Published by Elsevier Ltd. Open access under [CC BY-NC-ND license](#).

Contents

1. Introduction and rationale	348
2. Biology and mode of action of plant sterols/stanols	348
2.1. Origins	348
2.2. Transport and circulating levels	349
2.3. Tissue levels	350
2.4. Markers of intestinal cholesterol absorption and synthesis	350
2.5. Intestinal handling of cholesterol and non-cholesterol sterols: mechanism by which plant sterols/stanols added to the diet inhibit the absorption of cholesterol	350
3. Lipid modifying effects of plant sterols/stanols	351
3.1. LDL cholesterol lowering	351
3.2. Additional effects on the plasma lipid profile	352
4. Effects on atherosclerosis	352
4.1. Studies in animal and cell-based models	352
4.1.1. Animal models	352
4.1.2. Cell-based models	352
4.2. Studies in humans	353
4.2.1. Relationship of plant sterol/stanol consumption and vascular health	353
4.2.2. Relationship of circulating plant sterols/stanols and CV risk: Cohort studies	353
5. Safety	354
6. Optimising the use of plant sterols/stanols in lipid lowering	354
6.1. Combination therapy	354
6.1.1. Statins	354
6.1.2. Ezetimibe	354
6.1.3. Other combination lipid therapy	355
6.2. Postprandial lipaemia	355
7. EAS consensus panel recommendations	355
Acknowledgements	356
Author contribution	357
European Atherosclerosis Society (EAS) Consensus Panel on Phytosterols	357
Funding	357
Conflict of interest	357
Supplementary material	357
References	357

1. Introduction and rationale

The on-going pandemic of obesity, metabolic syndrome and diabetes is directly associated with an ever-increasing worldwide incidence of premature atherosclerosis and cardiovascular disease (CVD). Indeed, the European Union budget for CVD is currently estimated at €196 billion a year, with about one-half of this attributed to direct healthcare costs [1]. However, limiting the expenditure for CVD to the healthcare systems of the European Union grossly underestimates its true cost. Most public health expenses are linked to treatment, a notion which strongly argues for urgent investment in CVD prevention to improve health in European populations and to stem the socioeconomic consequences.

Recent studies of the relationship of lifetime risk of CVD to risk factor burden clearly indicate that individuals with an optimal risk factor profile, (including well-controlled blood pressure and cholesterol, non-smoking and non-diabetic), display substantially lower risk of CV events over their lifetime than those with two or more of these major risk factors [2]. Ranking of nine CV risk factors in the INTERHEART cross-sectional study in 52 countries revealed that dyslipidaemia alone accounted for most of the population-attributable risk for myocardial infarction; here, dyslipidaemia was defined as an excess of cholesterol transported in atherogenic apolipoprotein (apo)B-containing lipoproteins (among which low-density lipoproteins [LDL] predominate), relative to that in non-atherogenic apoA-I-containing, high-density lipoproteins (HDL) [3].

Robust data attest to the causal role of LDL-cholesterol (LDL-C) in coronary atherosclerosis. Reductions in LDL-C levels achieved by treatment with diet, statins or bile acid sequestrants, or by ileal bypass surgery, in prospective clinical trials of 3 or more years duration, have been demonstrated in meta-regression analyses to significantly reduce CV morbidity and mortality [4,5].

As atherosclerosis is a chronic, progressive disease typically initiated during the first three decades of life, it follows that lowering LDL-C early may substantially delay or even prevent the onset of atherosclerosis, particularly in the coronary circulation. Indeed, evidence that prolonged exposure to low plasma LDL-C levels is associated with markedly greater reduction in CV risk compared with current strategies aimed at lowering LDL-C in middle age, has been provided by a meta-analysis of Mendelian randomisation studies, involving polymorphisms in six distinct genes of cholesterol metabolism [6].

These findings prompt a key question: How can LDL-C be maintained at low levels throughout life without imposing additional burden on the healthcare system? Clearly, lifestyle, which encompasses dietary habits, must be seriously considered, particularly as extensive nutritional and behavioural changes may lower LDL-C levels by up to 20% [7].

The liver is the principal regulator of circulating LDL-C levels. Not only is it the site of formation of very low-density lipoproteins (VLDL), the precursors of most LDL particles in the circulation, but it is also the site of most receptor-mediated clearance of LDL [8]. Both the liver and intestine are central to body cholesterol homeostasis. Indeed, after lipolysis-mediated removal of dietary triglycerides from chylomicrons, the liver rapidly clears circulating chylomicron remnants, which carry cholesterol that was absorbed in the small intestine [9]. The resultant increase in hepatic cholesterol stimulates VLDL secretion and hence LDL formation, and down-regulates hepatic LDL receptor activity. Such events potentially lead to elevations in plasma LDL-C levels. Both chylomicron remnants and VLDL remnants, in addition to LDL, can deliver cholesterol to the artery wall, initiating or exacerbating atherosclerosis. When cholesterol absorption in the small intestine was inhibited by ezetimibe in apoE deficient mice, atherosclerosis was dramatically reduced [10]. In

humans, inhibition of cholesterol absorption by ezetimibe results in lowering of plasma LDL-C levels because of increased fractional removal of LDL from the circulation, consistent with less dietary cholesterol arriving at the liver via chylomicron remnants with subsequent upregulation of LDL receptors [11,12]. Inhibition of cholesterol absorption by ezetimibe was also associated with reduced chylomicron secretion into plasma as determined by reduced production of apoB48 (Fig. 1) [12].

Thus it follows that the cholesterol absorption pathway represents an attractive target in the management of dyslipidaemia, with a specific focus on both reducing the cholesterol content of chylomicron and VLDL remnants and lowering LDL-C levels. This pathway presents clinical opportunities for dietary supplementation with agents that attenuate intestinal cholesterol absorption, among which plant sterols and plant stanols are prominent.

The European Atherosclerosis Society (EAS) convened an international Consensus Panel of basic scientists and clinical investigators with expertise in cholesterol metabolism, plant sterol and plant stanol biology, and CVD. Our goals were (i) to critically appraise state-of-the-art knowledge pertaining to the potential of plant sterols and plant stanols (hereafter referred to as plant sterols/stanols) for lowering LDL-C, with a view to preventing premature atherosclerosis and CVD, and (ii) to propose recommendations for optimal integration of foods with added plant sterols/stanols, as part of a healthy lifestyle, for attenuation of CV risk. These recommendations can provide guidance and support for clinicians and health professionals in the prevention of CVD across the spectrum of CV risk.

2. Biology and mode of action of plant sterols/stanols

2.1. Origins

Plant sterols/stanols are bioactive components with similar functions as that of cholesterol in mammals. Plant sterols are steroid alkaloids which differ from cholesterol in the structure of their

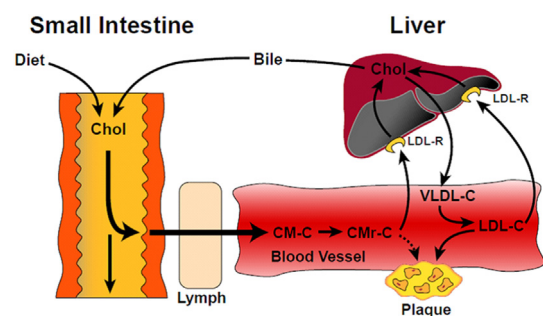


Fig. 1. Scheme illustrating the potential impact of cholesterol absorption from the small intestine on plasma levels of chylomicron remnants and LDL, with detrimental effects on the vascular wall. Cholesterol (Chol, C) entering the intestinal lumen primarily from the bile and the diet is absorbed to varying degrees and packaged in chylomicrons (CM-C) for transport via the lymph to the circulation. Therein, hydrolysis of much of the triacylglycerol present in the nascent CM results in the formation of cholesterol-rich remnant particles (CMr-C) that are ordinarily rapidly cleared from the circulation by the liver. Delivery of excess intestinal cholesterol to the liver can result in suppression of low density lipoprotein-receptor (LDL-R) activity and endogenous cholesterol synthesis, or acceleration of hepatic very low density lipoprotein-cholesterol (VLDL-C) secretion, or both. Such events will potentially raise plasma LDL-cholesterol (LDL-C) concentration. If hepatic clearance of CMr is delayed, then these particles may contribute directly to plaque formation. Together, these various pathways illustrate how agents that limit cholesterol absorption may elicit favourable changes in atherogenic plasma lipoproteins that culminate in attenuating plaque formation.

side chain, while plant stanols are 5α -saturated derivatives of plant sterols (Fig. 2).

The main food sources of plant sterols are vegetable oils, spreads and margarines, breads, cereals, and vegetables (Table 1); these contribute 50–80% of the daily plant sterol intake, with fruits adding a further 12% [13–15]. In the typical Western diet, the mean daily intake of plant sterols is about 300 mg [13,14], but can be as high as 600 mg in vegetarians [16]. The most abundant are sitosterol and campesterol, which contribute 60% and 20%, respectively, of plant sterol intake [13,14]. By comparison, the amounts of plant stanols in the diet are much lower, with only about 17–24 mg per day (predominantly sitostanol and campestanol) [14]. Cereals, especially wheat and rye, are the richest source of plant stanols.

2.2. Transport and circulating levels

These dietary components undergo low fractional absorption in the intestine, of the order of 0.5–2% for plant sterols and 0.04–0.2% for plant stanols [17]. As a result of low absorption and efficient excretion into bile after uptake by the liver, circulating levels are low, varying from 7 to 24 $\mu\text{mol/L}$ (0.3–1.0 mg/dL) for plant sterols, and from 0.05 to 0.3 $\mu\text{mol/L}$ (0.002–0.012 mg/dL) for plant stanols [18]; these levels are of the order of 500-fold and 10,000-fold lower, respectively, than those of cholesterol.

Long-term consumption of foods with added plant sterols (mean intake \pm standard deviation [SD] 1.1 ± 0.6 g/day) increases their circulating levels (from 19 to 30 $\mu\text{mol/L}$ [0.8–1.2 mg/dL] with plant sterol consumption), which overlap those within the normal range [19]. For foods with added plant stanols (mean intake 0.6 ± 0.4 g/day), increases in circulating levels of plant stanols (from 0.3 to 0.7 $\mu\text{mol/L}$ [0.012–0.028 mg/dL] with plant stanol consumption), and a decrease in circulating plant sterols (from 16 to 23%), were observed [19]. The quantitative distribution of plant sterols/stanols across the major lipoprotein classes is similar to that of cholesterol, and thus they circulate primarily in LDL particles (65–70%).

Table 1

Plant sterol and plant stanol contents in different foods. Data are given as mg/100 g (dry weight, either range or mean value).

Food item	Plant sterols	Plant stanols
<i>Vegetable oils</i>		
Corn oil	686–952	23–33
Rapeseed oil (canola oil)	250–767	2–12
Soybean oil	221–328	7
Sunflower oil	263–376	4
Olive oil	144–193	0.3–4
Palm oil	60–78	Traces
<i>Cereals</i>		
Corn	66–178	–
Rye	71–113	12–22
Wheat	45–83	17
Barley	80	2
Millet	77	–
Rice	72	3
Oats	35–61	1
<i>Nuts</i>		
Peanuts	320	–
Almond	143	–
<i>Vegetables</i>		
Broccoli	39	2
Cauliflower	18–40	Traces
Carrot	12–16	Traces
Lettuce	9–17	0.5
Potato	7	0.6
Tomato	7	1
<i>Fruits and berries</i>		
Avocado	75	0.5
Passion fruit	44	Not detected
Raspberry	27	0.2
Orange	24	Not detected
Apple	12–18	0.8
Banana	12–16	Not detected

–, Not reported.

Source: Adapted from Piironen V & Lampi AM (2004) [15].

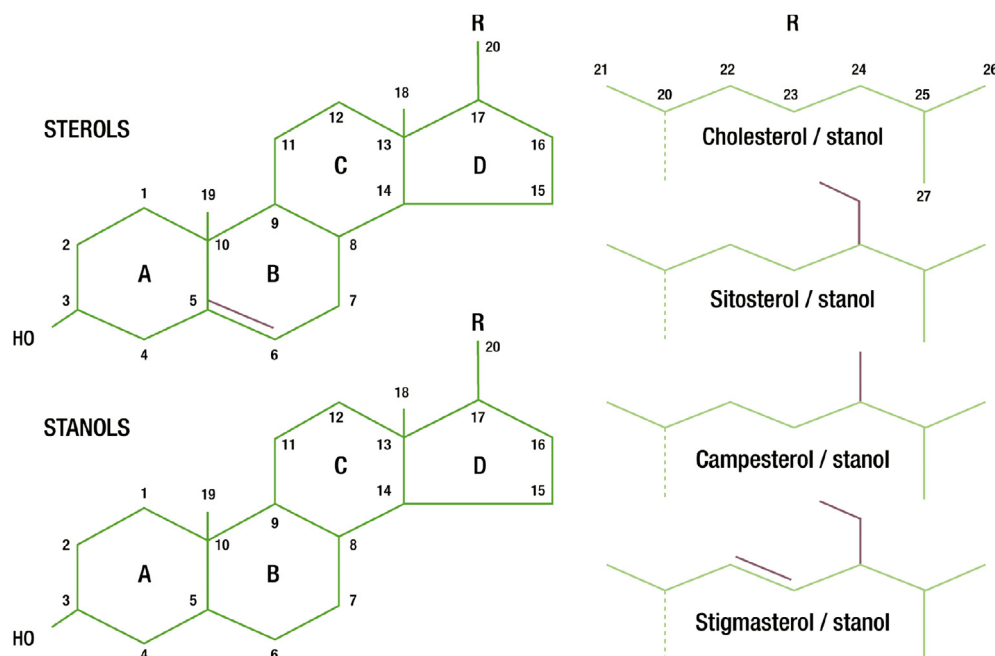


Fig. 2. Chemical structure of plant stanols and plant sterols.

The ATP-binding cassette co-transporters G5 and G8 (ABCG5/ABCG8) play a crucial role in controlling the absorption of plant sterols/stanols by secreting absorbed molecules back into the intestinal lumen [20,21]. In phytosterolaemia, severe loss of function mutations in genes coding for the ABCG5/ABCG8 transporters result in dramatic elevation in plasma plant sterol/stanol levels, which are more than 50-fold higher than those in normal individuals after consumption of plant sterols, and many of these individuals develop premature atherosclerosis (Box 1) [20–22]. Further consideration of this rare genetic disease is, however, beyond the scope of the present paper, which is focussed on plant sterol/stanol consumption as part of a healthy diet for prevention of CVD.

2.3. Tissue levels

There are limited data available for tissue levels in subjects receiving foods with added plant sterols/stanols. In healthy human subjects, plant sterols/stanols are taken up into tissues in similar proportions relative to cholesterol, and the ratio of cholesterol to plant sterols/stanols, which displays a wide range ($0.001–0.01 \times 10^{-4}$), is similar to or less than that in plasma, consistent with the absence of preferential accumulation or retention in tissues (see Supplementary Table 1). Similar findings have been reported for cerebrospinal fluid and brain tissue in healthy control subjects, and also in phytosterolaemia [23,24]. In healthy volunteers, data on tissue plant sterol/stanol levels are available for the carotid artery wall and for non-stenotic and stenotic aortic valve cusps [25–28]. In these tissues, cholesterol content is 20 µg/mg of tissue, while total plant sterol and plant stanol concentrations are ~0.04 µg/mg and ~0.001 µg/mg of tissue, respectively. Plant sterol concentrations vary widely between tissues (see Supplementary Table 1) [25–30].

Box 1 Phytosterolaemia

- Phytosterolaemia (previously referred to as sitosterolaemia) is due to rare, loss of function mutations in genes coding for the ATP-binding cassette transporters G5 and G8 (ABCG5/ABCG8), and is characterised by very high serum levels of plant sterols (up to 1.3 mmol/L or 50 mg/dL) and plant stanols (0.2 mmol/L or 8 mg/dL) [18].
- Major clinical manifestations may include premature atherosclerosis although this complication is variable; there are a number of patients with phytosterolaemia who do not have evidence of atherosclerosis (*D. Lütjohann, personal communication; E. Bruckert, privileged communication*). The presence of premature atherosclerosis appears to depend on whether there is co-existing severe hypercholesterolaemia. Other complications of phytosterolaemia include episodes of haemolysis, and xanthomas. Moreover, it is noteworthy that recent findings in 4 adult phytosterolaemic patients have not detected any significant degree of atherosclerosis (*E. Bruckert, privileged communication*).
- Plant sterols have been shown to accumulate in atherosclerotic lesions of phytosterolaemic subjects in the same ratio as present in serum. However, the phytosterol/cholesterol ratio is higher in phytosterolaemia than in normal subjects in plasma and in tissues.
- The potential relevance of markedly elevated amounts of plasma and tissue plant sterols and plant stanols to tissue deposition and the atherosclerotic process remains indeterminate.

During consumption of plant sterol-enriched foods, there was a 5-fold elevation in campesterol in stenotic aortic valve cusps [28]. However, in another study, the concentration of plant stanols in the arterial wall was not modified during consumption of foods enriched with plant stanols [27]. Clearly, there is a need for large, long-term clinical studies to exclude the possibility that consumption of dietary plant sterols/stanols might result in accumulation in arterial tissues, and evaluation of these agents as modulators of plaque formation or regression is equally of interest. Such studies present multiple methodological challenges, not least of which is access to plaque tissue for analysis of sterol composition, and for determination of the expression profile of key genes of cholesterol and plant sterol metabolism.

2.4. Markers of intestinal cholesterol absorption and synthesis

Serum cholesterol precursors and plant sterols, especially when expressed as ratios relative to cholesterol, can constitute markers of the rates of synthesis and absorption of cholesterol in non-dyslipidaemic subjects, as well as in several clinical conditions including primary and familial hypercholesterolaemia (FH), obesity and type 2 diabetes, and during some interventions including plant stanol consumption [31,32]. Overall, such markers have good validity, but there are exceptions (see Supplementary Appendix) [31–33], and full validation in multiple conditions is essential. Moreover, there are a number of methodological issues relating to standardisation of these markers. An on-going survey is investigating the degree of variability in sterol/stanol analytical data across different laboratories (see Supplementary Appendix).

2.5. Intestinal handling of cholesterol and non-cholesterol sterols: mechanism by which plant sterols/stanols added to the diet inhibit the absorption of cholesterol

The handling of sterols and stanols entering the intestinal lumen from the diet and bile is essentially a triphasic process (Fig. 3) [34–37]. The first phase is largely physicochemical in nature, occurs intraluminally, and culminates in the incorporation of cholesterol and other sterols and stanols into mixed micelles that serve as vehicles to carry these poorly water-soluble substances up to the surface of the brush border membrane on the enterocyte. This solubilisation step is essential for the subsequent entry of any type of sterol into the absorptive cell, from where it may potentially reach the circulation.

The second major phase is the uptake of cholesterol and other sterols or stanols into the enterocyte, a process that is facilitated by a plasma membrane-localised, general sterol transporter protein, Niemann-Pick C1-Like1 (NPC1L1). Within the absorptive cells, sterols undergo different fates, depending mainly on their chemical structure. Collectively, these events within the enterocyte broadly constitute the third major phase of intestinal handling. The bulk of the cholesterol is esterified by acyl CoA: cholesterol acyltransferase-2 (ACAT2) and incorporated into nascent chylomicrons, which initially enter the lymph before joining the circulation via the thoracic duct [38,39]. The absolute content of plant sterols/stanols in chylomicrons is markedly lower than that of cholesterol, with up to 50% in esterified form [39]. Importantly, the bulk of plant sterols/stanols is pumped back into the gut lumen via the ABCG5/ABCG8 transporter, resulting in minimal entry of these plant-derived molecules into the circulation [34,40,41]. Finally, foods with added gram quantities of plant sterols/stanols cause a significant inhibition of cholesterol absorption, most likely through disruption of the intraluminal solubilisation step [37], although other possible mechanistic explanations have been proposed [42].

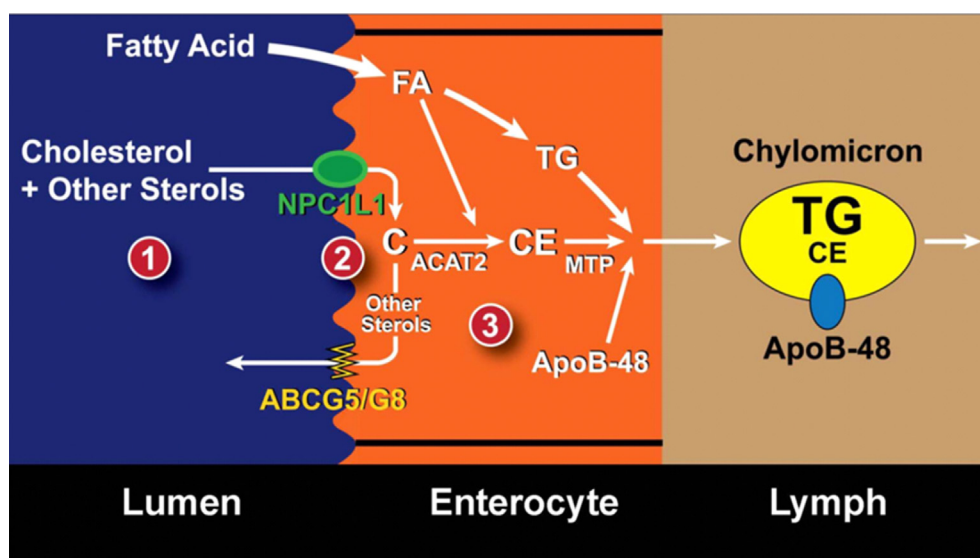


Fig. 3. Schematic summarising the three main steps (indicated by the numbers in red circles) involved in the intestinal handling of cholesterol and non-cholesterol sterols. In the first step within the intestinal lumen, sterols are incorporated into mixed micelles (1). At the brush border membrane, sterols are released from the micelles and transported into the cell via the Niemann-Pick C1-Like1 transporter (NPC1L1) (2). Once internalised, these sterols can be handled in different ways (3). For the bulk of the internalised non-cholesterol sterols, efflux back into the lumen occurs via ABCG5/ABCG8 (ATP binding cassette transporters G5 and G8). In contrast, a significant proportion of the internalised cholesterol undergoes esterification via ACAT2 (acyl CoA-cholesterol acyltransferase-2). The esterified cholesterol, along with much smaller quantities of esterified non-cholesterol sterols, are incorporated into nascent chylomicrons which enter the lymphatic system and ultimately the circulation. Dietary plant sterol or stanol supplementation is believed to inhibit the absorption of cholesterol most likely through disruption of the intraluminal solubilisation step [37]. Abbreviations: apo apolipoprotein; C cholesterol; CE cholesteryl ester; FA fatty acid, MTP microsomal triglyceride transfer protein; TG triglyceride.

As discussed above, the *ABCG5/G8* genes play a crucial role in regulating circulating levels of plant sterols [20,21]. In the KORA study, common variants of *rs41360247* and *rs4245791* in the *ABCG8* gene accounted for 8% of the variance in circulating levels of plant sterols [43]. The relationships of these and other variants in high linkage disequilibrium with circulating plant sterol levels have been confirmed in the CARLA, LURIC, and YFS cohorts [43,44].

3. Lipid modifying effects of plant sterols/stanols

3.1. LDL cholesterol lowering

Epidemiological studies in the UK ($n = 22,256$), Sweden ($n = 77,652$) and China ($n = 3940$) observed that naturally-occurring dietary plant sterol intake is inversely related to plasma total- and LDL-C levels [45–47]. However, in a well-controlled study in healthy subjects, low (126 mg plant sterols/2000 kilocalories) or high intake of plant sterols (449 mg plant sterols/2000 kilocalories) did not affect plasma LDL-C concentrations in spite of modulating cholesterol metabolism [48]. Furthermore, even at the highest levels of dietary intake, plant sterols/stanols occurring naturally in the diet have a modest hypocholesterolaemic effect. On the other hand, when natural plant sterols were omitted from the diet, serum LDL-C concentrations increased [49].

In contrast to the minimal effects of variation in consumption of naturally-occurring plant sterols/stanols in the diet, early studies such as that by Farquhar et al. (1956) demonstrated that beta-sitosterol supplementation lowered both total serum cholesterol and LDL-C (as beta-lipoprotein lipid) in young men with atherosclerotic heart disease [50]. Subsequently, Miettinen et al. (1995) demonstrated for the first time that foods (such as margarine, see Box 2) enriched with sitostanol ester lowered both total serum cholesterol and LDL-C in mildly hypercholesterolaemic subjects [51].

Subsequent data show consistent support for the LDL-C lowering effects of foods with added plant sterols/stanols [52–

54]. In the most recent meta-analysis, consumption of foods with added plant sterols/stanols (2 g/day) lowered LDL-C to a similar extent (8.2% and 9.3%, respectively) [54]. Regrettably, there is a paucity of data relating to the potential of higher plant sterol/stanol doses to further lower LDL-C levels, and thus CV risk [6,54,55]. It has been suggested that maximal LDL-C lowering may be greater with plant stanols (up to 16%) and plant stanol esters (17%), but this conclusion relies on limited studies using doses of 4–9 g/day, none of which involved direct head-to-head comparisons with plant sterols. Whether there is a position for plant sterol and/or plant stanol intakes to be raised higher than those currently recommended (2–3 g/day) in prevention strategies for the general population remains open. The fact that there is consistent, robust evidence indicating that lowering of LDL-C by different mechanisms (statins, diet, partial ileal bypass, bile acid sequestrants) results in reduction in CV risk, underlies the rationale for inclusion of plant sterols/stanols in international clinical guidelines for the management of dyslipidaemia [5,56]. In this context, it must be recognised that the use of functional foods enriched with plant sterols/stanols is currently not advised for children under 6 years. There is, however, a substantial database showing consistent LDL-C lowering efficacy in children. In controlled clinical trials in children

Box 2

Types of foods with added plant sterols or plant stanols

Low-fat spread/margarine
Yoghurt drinks
Dairy-free drinks
Low- or reduced-fat milk
Soft cheese
Orange juice
Muesli
Bread
Biscuits

and adolescents aged 4–15 years, consumption of foods supplemented with plant sterols/stanols (1.5–3 g/day) resulted in consistent LDL-C reduction (by 5–15%) in normolipaeamic children ($n = 98$), and equally in children with FH ($n = 224$; by 9–19%) (see [Supplementary Table 2](#)) [57–66]. The magnitude of such reductions is comparable to that observed in adults. On this basis, the Panel proposes that dietary supplementation with plant sterols/stanols may be considered in children (from the age of 6 years) with FH who require lipid lowering treatment as an adjunct to lifestyle advice and potential pharmacotherapy, although long term safety studies are clearly needed.

3.2. Additional effects on the plasma lipid profile

Most studies with plant sterols/stanols were conducted in individuals with isolated hypercholesterolaemia. The available data suggest that triglyceride levels are reduced by 6–20% at intakes of 1.5–2 g/day of plant sterol/stanol, with essentially no effect on HDL-C [67–69]. Pooled analyses showed a modest reduction in plasma triglycerides of 6% and 4% for recommended intakes of plant sterols (1.6–2.5 g/day) or plant stanols (2 g/day), respectively [69,70]. Indeed, evidence suggests a relationship between baseline triglyceride levels and the magnitude of this effect, with 9% reduction when baseline triglycerides were 1.9 mmol/L (170 mg/dL), but no effect at baseline levels of 1.0 mmol/L (90 mg/dL) [69]. In subjects with metabolic syndrome and moderate hypertriglyceridaemia, plant stanols lowered hepatic production of both large (>60 nm) and medium size (35–60 nm) VLDL particles [67]. Other studies documented a reduction in small, dense LDL particles in patients with type 2 diabetes or metabolic syndrome after consumption of plant stanols/sterols [68,71]. Based on one study, it appears that plant sterols/stanols do not influence lipoprotein(a) levels [72].

Finally, it is of considerable interest that LDL-C reduction, subsequent to consumption of a plant stanol ester-enriched diet in a population of metabolic syndrome subjects, was without effect on plasma levels of proprotein convertase subtilisin/kexin type 9 (PCSK9) (J Plat, unpublished data), a potential contrast to the elevation in PCSK9 levels induced upon statin-mediated LDL-C lowering [73].

Future studies of the potential effects of foods with added plant sterols/stanols in attenuating the atherogenicity of the postprandial period in well-phenotyped cohorts of subjects with metabolic syndrome or type 2 diabetes would be of special interest. Such investigations should focus on normalisation of both the qualitative and quantitative features of atherogenic triglyceride-rich lipoproteins and their remnants during this phase.

4. Effects on atherosclerosis

4.1. Studies in animal and cell-based models

Because plant sterols/stanols effectively reduce plasma LDL-C concentrations, their use could constitute a potential protective strategy against the initiation and progression of atherosclerosis. Most of the available supportive data relate to studies in animal- and cell-based models, however, these have inherent limitations. Thus, investigations in animal models have been of short duration, while very high (pharmacological) doses have been typically applied in both animal and cell models, thereby highlighting the need for cautious interpretation of the experimental findings.

4.1.1. Animal models

To date, more than 30 studies have investigated the effect of plant sterol/stanol supplementation on experimental

atherosclerosis in various animal models (see [Supplementary Table 3](#) and reviewed by Kritchevsky and Chen [2005]) [74]. In genetically-modified mouse models of atherosclerosis, protective effects were observed despite increases (up to 10-fold) in plasma plant sterol/stanol concentrations [28,75,76]. Such effects included reduction in arterial lipid accumulation, and inhibition of lesion formation and progression. Moreover, regression of existing lesions correlated with the cholesterol-lowering action of plant sterols/stanols.

4.1.2. Cell-based models

4.1.2.1. Cellular metabolism of plant sterols/stanols and potential impact on cholesterol metabolism. The proportions of plant sterols/stanols relative to cholesterol found in plasma are maintained in tissue sterols [24,26], suggesting that they are handled similarly to cholesterol in most cells. While direct studies of cellular plant sterol metabolism are relatively limited in scope, they support this view. For example, the rates of uptake and accumulation of sitosterol and cholesterol by macrophages are similar [77], and substantial esterification of beta-sitosterol (=sitosterol) and other plant sterols can be measured in many tissues and cells [26,78,79]. On the basis of limited data, efflux of sitosterol and sitostanol to HDL from human macrophages appears to be more efficient than that of cholesterol [77].

Cholesterol homeostasis is tightly controlled at the transcriptional level via Sterol Regulatory Element-Binding Protein-2 and liver X receptor (LXR)-dependent regulation, and also by post-translational control of the turnover of a number of key enzymes, receptors and transporters. Plant sterols appear to have little or no impact on these processes. Their ability to activate LXR-dependent genes is negligible or very low [80–84]. A plant sterol-enriched diet had no effect on mouse intestinal LXR target gene expression [85]. In future studies, more comprehensive evaluation of the effects of 'physiological' concentrations of plant sterols/stanols on the hepatic and intestinal gene expression profile would be of considerable interest.

4.1.2.2. Influence of plant sterols/stanols on inflammatory pathways. In macrophages, recent studies have shown that lipid accumulation and inflammatory responses are co-ordinated through LXR-mediated transrepression of inflammatory genes by desmosterol, an intermediate in cholesterol synthesis and an endogenous LXR ligand [86,87]. Interestingly, as plant sterol/stanol consumption results in a compensatory increase in endogenous cholesterol synthesis [31], the corresponding increase in intracellular desmosterol concentrations could result in plant sterol/stanol-induced anti-inflammatory effects. Indeed, a number of *in vitro* studies indicate that some plant sterols exert anti-inflammatory effects on activated macrophages. However, these studies have significant limitations, as they involved addition of plant sterols and/or stanols under conditions, which (although seldom directly measured), almost certainly significantly increase the plant sterol:cholesterol ratio to values well in excess of the normal 1:500 to 1:1000 ratio that exists *in vivo*. This is likely even when plant sterols are added at 'physiological' concentrations. Future studies must include direct measurement of the cellular levels of cholesterol and plant sterols and should aim to mimic those found *in vivo*.

There are conflicting findings with respect to the effects of sitosterol and campesterol on production of proinflammatory factors by macrophages [88–92]. Interestingly, sitosterol stimulated anti-oxidant pathways and inhibited either phorbol ester- or lipopolysaccharide-stimulated prostaglandin synthesis in mouse macrophages [93–96]. The potential relevance of these findings to the atherosclerotic process is unclear at present, but warrants further research. In addition to macrophages, substantial evidence

attests to the implication of T-lymphocytes in the immunoinflammatory dimension of atherosclerosis [97]. Interestingly, both plant sterols and plant stanols exert immune-modulating properties, to the extent that the evoked T-helper cell (Th) 1 response, as shown by enhanced production of the Th1 cytokines, interferon-gamma and interleukin-2, and activation of toll receptor 2, is an essential feature of this response [92].

Thus, in summary and based on the evidence discussed, we cannot exclude the possibility that accumulation of plant sterols/stanols might occur in vascular cells as a consequence of an increase in their circulating concentrations (see Box 3).

4.2. Studies in humans

4.2.1. Relationship of plant sterol/stanol consumption and vascular health

Human studies evaluating the effect of dietary plant sterols/stanols on measures of arterial structure or function using carotid intima media thickness, brachial artery size, flow-mediated dilatation (FMD), and arterial stiffness have not shown any major, consistent effects on vascular or endothelial function during short and mid-term plant sterol or plant stanol intake [64,66,98–103]. All studies were small and mostly of short duration, and importantly, several studies were in subjects with low CV risk exhibiting normal vascular function at baseline. Despite significant reductions in LDL-C levels, no consistent changes in measures of inflammation, oxidative stress or endothelial function were observed. In a study of apparently healthy non-smokers, daily use of plant stanol margarine for ≥ 2 years did not improve carotid artery compliance [103], and in a shorter (3 months) study, failed to impact FMD [98]. Similarly, in two studies in pre-pubertal children with FH, dietary plant sterol/stanol consumption did not improve endothelial function despite significant LDL-C lowering [64,66].

Recent research has focused on the characteristics of the microcirculation as markers of early arterial disease [104]. In one trial [105], intake of foods enriched with plant sterols/stanols (2.5 g/day) for 85 weeks by statin-treated subjects showed an association between the increase in serum campesterol concentrations and changes in both retinal arteriolar and venular diameters. Retinal venular diameters increased (by $2.3 \pm 3.1 \mu\text{m}$) and arteriolar diameter decreased (by $0.7 \pm 4.7 \mu\text{m}$) in the group receiving added plant sterols, although these effects did not reach significance. However, the change in cholesterol-standardised campesterol concentrations correlated positively with the change in venular diameter independent of changes in LDL-C concentration. Other studies with endpoints including coagulation [72], platelet

aggregation measures [106], oxidant stress [107], inflammation [108] or other biomarkers did not demonstrate convincing benefit. It is possible, however, that the reduction of LDL-C by 10%, as typically mediated by intake of foods with added plant sterols/stanols, may not attain the threshold required for impact on these parameters. Adequately powered, controlled studies of the effects of plant sterols/stanols on morphological, functional and biochemical surrogate markers of atherosclerosis are needed. Based on present evidence, there is no indication that dietary supplementation with plant sterols/stanols are associated with either benefit or harm to vascular function.

4.2.2. Relationship of circulating plant sterols/stanols and CV risk: Cohort studies

Several observational studies have investigated the association between circulating plant sterols and atherosclerosis in the general population. Some early reports found moderately elevated plant sterol levels to be positively associated with vascular disease [109,110], although others suggested an inverse or lack of relationship between circulating plant sterols and CV risk [111–113]. A recent meta-analysis including 17 studies ($n = 11,182$) [114], showed no significant associations of circulating campesterol and sitosterol with vascular disease over a range of circulating plant sterol concentrations (average concentrations for first and third tertiles, 0.17 and 0.47 mg/dL [4 and 12 $\mu\text{mol/L}$] for campesterol, and 0.13 and 0.38 mg/dL [3 and 10 $\mu\text{mol/L}$] for sitosterol). Two studies showed inverse relationships between plant sterols and CV risk, one study did not find any association, and a further study reported a positive association between plant sterols and CV risk [115–118].

Since these retrospective/case-control and prospective/cohort studies do not provide the highest level of evidence in defining causality, and as placebo-controlled trials with these endpoints are lacking, Mendelian randomisation studies may be informative. Moderate elevations of circulating plant sterols, which are associated with plant sterol-raising variants in the *ABCG8* gene, showed a positive association with prevalent coronary artery disease [43,119–122]. Such an increase in risk can, however, be entirely explained by the association of *ABCG8* variants with intestinal cholesterol absorption, as reflected by circulating cholestanol levels, and with LDL-C plasma concentration, rather than circulating levels of plant sterols [44,123–126]. The *ABO* gene has also been associated with elevated circulating sterol levels and CV risk [43]. However, the *ABO* locus exhibits even greater pleiotropy than the *ABCG5/G8* locus, as it regulates von Willebrand factor, coagulation factor VIII, intercellular adhesion molecule-1, P-selectin, and E-selectin levels [127–129]. In the largest report to date ($n = 1242$), circulating levels of plant sterols were lower in the group with coronary artery disease (CAD), and concentrations of the plant stanols, campestanol and sitostanol, which are much lower than those of plant sterols, were not different between the groups with and without CAD [130]. Thus, the available Mendelian randomisation studies do not provide a scientific basis to discourage the use of plant sterol- or plant stanol-containing functional foods.

A healthy diet is the cornerstone of CVD prevention. In this respect, it is of critical importance that indisputable evidence supports the contention that lowering of plasma LDL-C levels confers clinical benefit, irrespective of mechanism, and that such mechanisms include dietary intervention [4–7,131–133]. Indeed, robust data show that there is no qualitative difference between statin- and non-statin-mediated reduction in LDL-C when comparing their estimated effects on myocardial infarction or CHD death on the basis of the Bayes factor [5]. Thus, the regression lines for all individual diet ($n = 5$), bile acid sequestrant ($n = 3$), surgery ($n = 1$), and statin ($n = 10$) trials were similar and consistent with a one-to-one relationship between LDL-C lowering and reduction in

Box 3

Implications from studies in animal- and cell-based models of the impact of plant sterols and plant stanols on atherosclerosis

- The limited available data indicate that most cells handle (uptake, esterification, export) plant sterols/stanols in a similar manner as cholesterol.
- *In vivo*, plant sterols represent $\sim 0.1\%$ of total cellular sterol; however, most *in vitro* studies have involved the use of much higher levels of plant sterols.
- Evidence suggests that plant sterols do not affect cellular cholesterol homeostasis, although further studies are needed to confirm this.
- Some studies indicate mild anti-inflammatory effects of plant sterols/stanols, but confirmation is required at 'physiological' levels of plant sterols.
- Plant sterols are not toxic at 'physiological' concentrations.

CHD and stroke over 5 years of treatment [5]. In this context, and as discussed above, it is of immediate relevance that consumption of 2 g/day of plant sterols/stanols, as part of a healthy diet, lowers LDL-C plasma levels by approximately 10% [52–54].

The EAS Consensus Panel on Phytosterols, however, recognises that there is a lack of randomised data pertaining specifically to the impact of foods with added plant sterols/stanols on CVD prevention. Large-scale outcome trials of food products with added plant sterols/stanols for CVD prevention in the setting of low to intermediate risk are not practically feasible, given the very large number of subjects (>50,000) required for adequate power (see [Supplementary Appendix 2. Feasibility of an outcomes study for food products with added plant sterols/stanols](#)). Furthermore, the benefit of consistent, but relatively small, additional LDL-C lowering from consumption of foods with added plant sterols/stanols, as part of a healthy diet, would be difficult to demonstrate definitively in a clinical trial of optimally treated patients at high CV risk, even if 25–30,000 individuals were enrolled. The absence of randomised, controlled trial data with hard end-points to establish clinical benefit from the use of plant sterols or plant stanols must be considered when health professionals choose to advise their use as supplements to a healthy diet.

5. Safety

In the context of benefit-risk considerations, substantial interest has focused on the safety aspects of plant sterols/stanols when used as cholesterol-lowering agents. Given that the level of plant sterol/stanol intake required for cholesterol lowering ranges from 1 to 3 g/day, well above typical consumption patterns which rarely exceed 400–600 mg/day, a considerable literature has amassed exploring possible deleterious effects of prolonged plant sterol/stanol consumption. Indeed, for some individuals zealously consuming multiple foods with added plant sterols/stanols, it is feasible that intakes could rise considerably above the 3 g/day level considered to represent the ceiling at which the dose-response curve for cholesterol-lowering with plant sterols begins to plateau. However, evidence from clinical studies and post-launch monitoring indicates that overconsumption of foods with added plant sterols/stanols is not an issue [19,134].

Specific areas of concern surrounding plant sterol/stanol consumption include (i) possible negative effects on fat-soluble vitamin status, and (ii) and cancer risk. Overall, longer-term post-launch monitoring efforts have failed to answer the question whether foods with added plant sterols/stanols cause any unexpected negative health effects [135,136]. Several longer-term feeding trials have similarly failed to identify any negative action of plant sterol/stanol consumption on clinical chemistry, haematology or clinical symptomatology [52–54,137,138].

A repeated observation in some, but not all, plant sterol feeding trials is modest suppression of plasma carotenoid concentrations (by 10%), especially for the highly lipophilic hydrocarbon carotenoids (beta-carotene, alpha-carotene and lycopene) [139,140]. It has been suggested that this may arise from suppression of intestinal absorption [141]. Increasing consumption of fruits and vegetables offsets any decline in fat-soluble vitamin levels induced by plant sterol/stanol intakes in the range that lowers cholesterol [139].

Considerable controversy has occurred over the past decade regarding a possible atherogenic role of high circulatory levels of plant sterols/stanols. Reports are conflicting, although the most recent work seems to indicate that plasma levels of plant sterols associated with recommended intakes (2 g/day) do not pose a health risk. Mendelian randomisation studies have claimed to show that increased circulatory levels of plant sterols increase CVD risk, but since the *ABCG5/G8* polymorphisms examined have pleiotropic

effects, the results of these studies can readily be explained by an increased cholesterol absorption rate (see above).

Lastly, considerable evidence from animal and cell studies suggests, if anything, a protective role of sitosterol against certain cancers [142–145]. Thus, there is no increase in cancer risk with recommended daily intakes of functional foods containing 2–3 g/day of plant sterols/stanols. In fact, surveillance data associate reduced risk of certain cancers with plant sterol/stanol intakes [146,147]. Possible mechanisms based on animal and cell work suggest direct actions on apoptosis or indirect actions through the intracellular cholesterol-lowering ability of these agents [145].

Overall, evidence from longer-term monitoring trials, as well as experimental models, indicates that plant sterols/stanols present a favourable safety profile, thereby supporting their use in cholesterol lowering, either alone or adjunctive to pharmacotherapy. Based on the epidemiology, the genetics, and the wealth of current clinical trial evidence demonstrating LDL-C lowering with plant sterols/stanols and lack of significant safety concerns, the use of plant sterols/stanols in treating hypercholesterolaemia can be encouraged [56]. The question remains how best to optimise their use in lipid management.

6. Optimising the use of plant sterols/stanols in lipid lowering

6.1. Combination therapy

In addition to a role in primary prevention in the general population, foods with added plant sterols/stanols may provide additional LDL-C lowering in dyslipidaemic patients at high CV risk treated with lipid-lowering drugs. Thus, it is important to define the lipid-modifying effects of dietary plant sterols/stanols (2–3 g/day) in combination with pharmacotherapies so as to optimise their clinical use.

6.1.1. Statins

Statins are inhibitors of the rate-limiting enzyme of cholesterol biosynthesis, HMG-CoA reductase. As such, their action directly decreases intracellular levels of cholesterol and its precursors, enhances the catabolism of apoB-containing lipoproteins (mainly LDL) via upregulation of hepatic LDL receptors, and reduces *de novo* hepatic (and potentially intestinal) lipoprotein production. Since plant sterols/stanols act via a distinct mechanism, i.e. by lowering bioavailability of intestinal cholesterol for entry into the circulation, it can be speculated that plant sterols/stanols may exert an additive effect when combined with a statin. In clinical studies, dietary plant sterols/stanols induce an incremental decrease in LDL-C levels of 10–15% when added on top of statin therapy, which is superior to that (6%) obtained by doubling the statin dose [148–151]. *In vivo* studies of LDL-apoB kinetics in patients with type 2 diabetes mellitus indicated that the additive effects on LDL-C reduction when stanols were added to statins resulted from decreased production of LDL [152]. Thus, the generally accepted efficacy of plant sterol/stanol consumption (2–3 g/day) is maintained on top of statin therapy.

6.1.2. Ezetimibe

As the lipid-lowering mechanism of ezetimibe is mediated by inhibition of intestinal cholesterol absorption, ezetimibe could be considered a competitor of dietary plant sterols/stanols at the molecular level. Importantly, however, the targets differ; ezetimibe blocks the NPC1L1 transporter, while plant sterols/stanols displace cholesterol from intestinal micelles. Moreover, as NPC1L1 is also the entry gate for dietary plant sterols/stanols into the body, ezetimibe-mediated inhibition of this mechanism should both enhance their effects in the lumen of the intestine, and also reduce their plasma

Box 4

Consensus panel recommendations

- Daily consumption of foods with added plant sterols and/or plant stanols in amounts of up to 2 g/day is equally effective in lowering plasma atherogenic LDL-C levels by up to 10%, and thus may be considered as an adjunct to lifestyle in subjects at all levels of CV risk. At higher daily intakes (9 g/day), the effects of plant stanols appear more pronounced, but additional studies are needed to confirm these results and examine safety at higher doses.
- Plant sterols and plant stanols can be efficaciously combined with statins. Very limited data suggest plant sterols/stanols may also lower LDL-C levels in combination with a fibrate or ezetimibe. In this way, the potential for attainment of LDL-C goals as a function of overall CV risk can be enhanced.
- Enhanced consumption of plant sterols and plant stanols may be considered as an adjunct to lifestyle and dietary approaches for modestly reducing elevated plasma triglyceride levels, especially when levels are elevated before treatment. This needs further study in appropriate populations with elevated triglycerides.

levels. However, clinical data are limited, and in the largest study to date, there was a significant incremental reduction in intestinal cholesterol absorption during administration of ezetimibe plus plant sterols (2 g/day) associated with a significant 8% reduction in LDL-C compared to ezetimibe alone [153].

6.1.3. Other combination lipid therapy

The relevance of other combination therapies merits consideration, especially from the perspective of comprehensive lipid control in cardiometabolic disease. Clinical studies show a trend for an additive effect on LDL-C levels when foods enriched with plant sterols/stanols are consumed with a fibrate [154,155]. Given that N – 3 fatty acids have a small effect on cholesterol metabolism and mainly influence triglyceride levels, consumption of plant sterols/stanols and N – 3 fatty acids may exert a complementary beneficial effect on the lipid profile. Indeed, both are recommended as

components of a healthy diet for prevention of CVD [56,156]. Currently there are insufficient data regarding the lipid-lowering efficacy of a combination of bile acid sequestrants and dietary plant sterols/stanols. However, as bile acid sequestrants interact with lipophilic substances, it is likely that such a combination will interfere with intestinal sterol/stanol absorption. Additionally, dietary plant sterols, but not plant stanols, suppressed bile acid synthesis, probably altering bile acid sequestrant-mediated cholesterol-lowering efficacy [157].

In conclusion, the 10% reduction in serum LDL-C concentrations typical of consumption of foods with added plant sterols/stanols at doses of approximately 2 g/day, persists on top of the effect of statins. Limited data suggest an additive effect of plant sterols/stanols with ezetimibe and fibrates.

6.2. Postprandial lipaemia

Since foods with added plant sterols/stanols reduce cholesterol absorption, they might also reduce the production of intestinally-derived chylomicrons and chylomicron remnants. The few studies that have been conducted, using either a single standardised meal or day-long measures of plasma triglycerides, have not shown any effect of either acute or chronic plant sterol/stanol intake on postprandial triglycerides [150,158,159]. Where measured, levels of postprandial plant sterols/stanols were variable [158,159]. Interestingly, consumption of plant sterols esterified with fish oil for one month resulted in lower postprandial triglyceride levels than fish oils alone [160]. Overall, the limited data examining the potential lowering of postprandial triglyceride excursions do not show significant effects, which might relate to low levels of baseline triglycerides in subjects in these studies.

7. EAS consensus panel recommendations

Currently, foods with added plant sterols/stanols may be considered in individuals with high cholesterol levels but equally in those with intermediate or low global CV risk who do not qualify for pharmacotherapy [56]. On the basis of critical appraisal of the evidence base above, this EAS Consensus Panel therefore considers that there is a place for these products, in conjunction with other

Total CV risk (SCORE) %	LDL-C levels				
	<70 mg/dL <1.8 mmol/dL	70 to 100 mg/dL 1.8 to <2.5 mmol/dL	100 to <155 mg/dL 2.5 to <4.0 mmol/dL	155 to <190 mg/dL 4.0 to <4.9 mmol/dL	>190 mg/dL >4.9 mmol/dL
<1	No lipid intervention	No lipid intervention	Lifestyle intervention	Lifestyle intervention	Lifestyle intervention, consider drug if uncontrolled
≥1 to <5	Lifestyle intervention	Lifestyle intervention	Lifestyle intervention, consider drug if uncontrolled	Lifestyle intervention, consider drug if uncontrolled	Lifestyle intervention, consider drug if uncontrolled
>5 to <10 or high risk	Lifestyle intervention, consider drug*	Lifestyle intervention, consider drug*	Lifestyle intervention, and immediate drug intervention	Lifestyle intervention, and immediate drug intervention	Lifestyle intervention, and immediate drug intervention
≥10 or very high risk	Lifestyle intervention, consider drug*	Lifestyle intervention, and immediate drug intervention	Lifestyle intervention, and immediate drug intervention	Lifestyle intervention, and immediate drug intervention	Lifestyle intervention, and immediate drug intervention

* In patients with MI, statin therapy should be considered irrespective of LDL-C levels

Fig. 4. Addition of functional foods with plant sterols/stanols, as a component of lifestyle intervention, may have potential value in individuals with high LDL-C levels at intermediate or low global cardiovascular risk who do not qualify for pharmacotherapy (as indicated by pink shading), in line with the joint ESC/EAS Guidelines for Management of Dyslipidaemia [56]. Equally, foods with added plant sterols/stanols may be considered in the context of lifestyle intervention in subjects at high or very high cardiovascular risk (as indicated by blue shading). Adapted from Reiner et al. (2011) [56].

Table 2

Comparison of the cost of foods with and without added plant sterols. Data based on a strategic analysis of the European market (Frost & Sullivan Research Service, London, UK, 2005) [163].

Product type	Cost per kg (UK sterling)		Incremental cost ratio ^a
	Foods with added plant sterols	Foods without added plant sterols	
Spreads	7.46–7.98	1.80	4.14–4.43
Health drinks	4.95–8.10	2.20	2.25–3.68
Yoghurt	2.80–4.00	2.10	1.33–1.90

^a Ratio of cost of foods with added plant sterols to cost of foods without plant sterols.

lifestyle interventions, in patients receiving lipid-lowering therapy with statins or other agents who do not achieve LDL-C targets, or in those with statin intolerance (see [Box 4](#) and [Fig. 4](#)). Finally, given the increasing importance of early preventive strategies in hypercholesterolaemia [6], the potential for inclusion of plant sterol- or plant stanol -enriched foods in the diet of adults and children (>6 years) with FH, as an adjunct to lifestyle and pharmacotherapy, may be considered. We base these recommendations on the proven ability of plant sterols/stanols to lower plasma LDL-C levels in the absence or presence of concomitant statin therapy, and equally on evidence that they reduce plaque size in atherosclerosis-prone animal models. We understand that, in the absence of CVD outcome data from randomised clinical trials, the evidence for use of plant sterols/stanols is incomplete.

Healthcare professionals should take into account the level of overall CV risk of patients, and their preferences. In addition, the

Box 5

Unresolved questions

- Does inter-individual variability occur in the cholesterol-lowering efficacy of plant sterols/stanols, i.e. is it possible to identify hyper- versus hypo-responders? If so, is it also possible to overcome lack of or poor responsiveness with higher plant sterol/stanol intake?
- Do plant sterols/stanols affect lipoproteins beyond LDL-C, i.e. atherogenic triglyceride-rich lipoproteins and their remnants, LDL and HDL subfractions, and lipoprotein(a)?
- Do plant sterols/stanols reduce the potential atherogenicity of postprandial lipid and lipoproteins, especially in subjects with cardiometabolic disease such as type 2 diabetes?
- How do plant sterols/stanols affect lipoprotein metabolism in specific populations, for instance in patients with the metabolic syndrome or type 2 diabetes mellitus who exhibit an atherogenic lipoprotein phenotype?
- How do plant sterols affect cellular lipid homeostasis?
- How do plant sterols affect the function of cells involved in the development of atherosclerosis, such as endothelial cells, monocytes and macrophages under resting and 'activated' conditions *in vitro*, *ex vivo* and *in vivo*?
- Do plant sterols/stanols have significant effects on biochemical surrogate markers of atherosclerosis including coagulation, platelet aggregation, oxidant stress, and subclinical inflammation?
- Is consumption of foods with added plant sterols/stanols (2 g/day), as part of a healthy diet, associated with clinical outcomes benefits? Given that large randomised outcomes studies in low to moderate risk subjects are not practically feasible, can sufficiently powered, randomised controlled studies show significant effects of plant sterols and/or stanols on morphological and functional (e.g. endothelial function) surrogate markers of arterial phenotype and/or atherosclerosis?

Box 6

Health economic evaluation of consumption of foods with added plant sterols and/or plant stanols

Nutrition economics follows a 4-step process in order to compute the health and economic impact of penetration of the market place of foods with added plant sterols/stanols.

To adopt the 4-step cost-of-illness approach:

- Determine a success rate for adoption of foods with added plant sterols/stanols across the target population
- Evaluate the extent of LDL-C reduction due to consumption of foods with added plant sterols/stanols
- Assess the decrease in coronary heart disease (CHD) prevalence due to the estimated reduction in plasma LDL-C levels
- Estimate the healthcare savings resulting from the reduction in CHD prevalence

This multistep approach can be applied with a range of inputted values for the 4 steps above, ranging from optimistic to pessimistic.

cost of these products is relevant as there appears to be a significant relationship between socioeconomic level and the profile of consumption of food products [161,162]. Based on UK data, it is evident that the cost/kg of food products with added plant sterols can range from 1.3-fold to up to 4-fold higher than that of their conventional counterparts ([Table 2](#)) [163]. Thus, cost may potentially constitute a deterrent to the regular purchase of these products, especially among less affluent, higher-risk groups. Indeed, this is supported by recent analyses from the Predi-Med study in Spain [164], which suggest that economic difficulties in Southern Europe may have had a detrimental effect on adoption of favourable dietary behaviours, including reduced adherence to the Mediterranean diet.

Further studies are needed to address unresolved questions highlighted in this appraisal (see [Box 5](#)). Key priorities include (i) evaluation of the effects of plant sterols/stanols in patients with metabolic syndrome; (ii) Mendelian randomisation studies to investigate the effects of plant sterols/stanols on clinical outcomes; and (iii) evaluation of long-term safety and effects on clinical outcomes in registries. Finally, although this EAS Consensus Panel did not find evidence for any health risk associated with consumption of these functional foods, the Panel recognises the lack of outcomes data showing clinical benefit. Given practical constraints, and the inability to differentiate LDL-C lowering effects of concomitant pharmacotherapeutic and dietary approaches in polymedicated patients, health economic modelling (see [Box 6](#)) may offer a feasible approach to investigate whether wider use of these functional foods has the potential for healthcare savings due to reduction in CVD prevalence. On the other hand, the EAS Consensus Panel would clearly welcome, and applaud, efforts by the food industry to plan and conduct a well-designed and adequately powered study of the effects of plant sterol/stanol-supplemented foods on CVD outcomes.

Acknowledgements

We are indebted to Professor E. Bruckert for privileged communication of unpublished findings in genetically-defined phytosterolaemic patients.

Jane Stock provided outstanding editorial support to the Consensus Panel.

Author contribution

European Atherosclerosis Society (EAS) Consensus Panel on Phytosterols

Writing committee: Helena Gylling (University of Helsinki, Finland), Lars Ellegård (Sahlgrenska Academy at University of Gothenburg, Sweden), Wendy Jessup (ANZAC Research Institute, Concord Hospital, Sydney, Australia), Peter J. Jones (Richardson Centre for Functional Foods and Nutraceuticals, University of Manitoba, Winnipeg, Canada), Dieter Lütjohann (Institute of Clinical Chemistry and Pharmacology, University Clinics Bonn, Germany), Winfried Maerz (Synlab Academy, Synlab LLC, Mannheim Center of Laboratory Diagnostics; and Mannheim Institute of Public Health, Social and Preventive Medicine, Medical Faculty Mannheim, University of Heidelberg, Heidelberg, Germany; and Clinical Institute of Medical and Chemical Laboratory Diagnostics, Medical University of Graz, Austria), Luis Masana (University Hospital Sant Joan, IISPV, CIBERDEM, Rovira and Virgili University, Reus, Spain), Jogchum Plat (Maastricht University, The Netherlands), Günther Silbernagel (Swiss Cardiovascular Center, Inselspital, University of Bern, Switzerland), Bart Staels (University of Lille 2, Lille, France), Stephen Turley (UT Southwestern Medical Center, Dallas, USA).

Co-chairs: M. John Chapman (INSERM U939, Pitié-Salpêtrière University Hospital, Paris, France) and Henry N. Ginsberg (Columbia University, New York, USA)

Other Panel members: Jan Borén (Sahlgrenska Center for Cardiovascular and Metabolic Research (CMR), University of Gothenburg, Sweden), Alberico Catapano (University of Milan and Multimedica IRCSS Milano, Italy), Guy De Backer (Ghent University Hospital, Belgium), John Deanfield (National Centre for Cardiovascular Prevention and Outcomes, University College London, UK), Olivier S. Descamps (Hopital de Jolimont, Belgium), Petri T. Kovanen (Wihuri Research Institute, Helsinki, Finland), Gabriele Riccardi (Federico II University, Naples, Italy), and Lale Tokgözoğlu (Hacettepe University, Ankara, Turkey).

The EAS Consensus Panel met twice in London, and the meetings were organised and chaired by MJC and HNG. The first meeting critically reviewed the literature while the second meeting reviewed additional literature and scrutinized the first draft of the consensus paper. Each Member of the Writing Committee drafted sections of the manuscript. All Panel members agreed to conception and design, contributed to interpretation of available data, suggested revisions for this document and all members approved the final document before submission.

Funding

This work including Consensus Panel meetings were supported by unrestricted educational grants to the EAS from Unilever R & D Vlaardingen B.V., The Netherlands; Danone Research, France; Raisio Group, Finland; BASF SE, Germany and Johnson & Johnson Consumer Services EAME Ltd, UK. These companies were not present at the Consensus Panel meetings, had no role in the design or content of the Consensus Statement or in the nomination of the Panel, and had no right to approve or disapprove the final document.

ST is fully supported by US Public Health Service Grant RO1HL09610.

Conflict of interest

In addition to unrestricted educational grants received by the EAS reported under Funding, several of the Consensus Panel members have received lecture honoraria, consultancy fees and/or research funding from Abbott (HNG, LT), Aegerion (MJC,ALC), Amgen (MJC,LM), Astra

Zeneca (MJC,HNG,JB,OSD,LM,LT), Baxter (LE), Bayer (LT), Boehringer (HNG,LT), Bristol Myers Squibb (HNG), Danone (MJC,LM,OSD,JP,ST), Esteve (LM), Ferre (LM), Fresenius Kabi (LE), Genzyme (MJC,HNG), Hoffman-La Roche (MJC,LM), Kowa (ALC,MJC,HNG,LM,LT), Merck (MJC,HNG,JB, ALC,LM,OSD,ST,LT), Nestlé (LE), Novartis (HNG,LM,LT), Nutricia (LE), Pfizer (MJC,HNG,JB, OSD,LT), Raisio Nutrition Ltd (HCy,JP), Sanofi-Aventis/Regeneron (JB, OSD,HNG,LM,LT), Solvay (OSD), Unilever (DL,GS).

Appendix A. Supplementary material

Supplementary data related to this article can be found online at <http://dx.doi.org/10.1016/j.atherosclerosis.2013.11.043>.

References

- [1] Nichols M, Townsend N, Luengo-Fernandez R, et al. European cardiovascular disease statistics 2012. Brussels/Sophia Antipolis: European Heart Network, European Society of Cardiology; 2012.
- [2] Berry JD, Dyer A, Cai X, et al. Lifetime risks of cardiovascular disease. *N Engl J Med* 2012;366:321–9.
- [3] Yusuf S, Hawken S, Ounpuu S, et al. Effect of potentially modifiable risk factors associated with myocardial infarction in 52 countries (the INTERHEART study): case–control study. *Lancet* 2004;364:937–52.
- [4] Baigent C, Keech A, Kearney PM, et al., Cholesterol Treatment Trialists' (CTT) Collaborators. Efficacy and safety of cholesterol-lowering treatment: prospective meta-analysis of data from 90,056 participants in 14 randomised trials of statins. *Lancet* 2005;366:1267–78.
- [5] Robinson JG, Smith B, Maheshwari N, Schrott H. Pleiotropic effects of statins: benefit beyond cholesterol reduction? A meta-regression analysis. *J Am Coll Cardiol* 2005;46:1855–62.
- [6] Ference BA, Yoo W, Alesh I, et al. Effect of long-term exposure to lower low-density lipoprotein cholesterol beginning early in life on the risk of coronary heart disease: a Mendelian randomization analysis. *J Am Coll Cardiol* 2012;60:2631–9.
- [7] Estruch R, Martínez-González MA, Corella D, et al. Effects of a Mediterranean-style diet on cardiovascular risk factors: a randomized trial. *Ann Intern Med* 2006;145:1–11.
- [8] Dietschy JM, Turley SD, Spady DK. Role of liver in the maintenance of cholesterol and low density lipoprotein homeostasis in different animal species, including humans. *J Lipid Res* 1993;34:1637–59.
- [9] Cooper AD. Hepatic uptake of chylomicron remnants. *J Lipid Res* 1997;38:2173–92.
- [10] Davis Jr HR, Compton DS, Hoos L, Tetzloff G. Ezetimibe, a potent cholesterol absorption inhibitor, inhibits the development of atherosclerosis in ApoE knock-out mice. *Arterioscler Thromb Vasc Biol* 2001;21:2032–8.
- [11] Chan DC, Watts GF, Gan SK, Ooi EM, Barrett PH. Effect of ezetimibe on hepatic fat, inflammatory markers, and apolipoprotein B-100 kinetics in insulin-resistant obese subjects on a weight loss diet. *Diabetes Care* 2010;33:1134–9.
- [12] Tremblay AJ, Lamarche B, Cohn JS, Hogue JC, Couture P. Effect of ezetimibe on the in vivo kinetics of apoB-48 and apoB-100 in men with primary hypercholesterolemia. *Arterioscler Thromb Vasc Biol* 2006;26:1101–6.
- [13] Klingberg S, Andersson H, Mulligan A, et al. Food sources of plant sterols in the EPIC Norfolk population. *Eur J Clin Nutr* 2008;62:695–703.
- [14] Valsta LM, Lemström A, Ovaskainen ML, et al. Estimation of plant sterol and cholesterol intake in Finland: quality of new values and their effect on intake. *Br J Nutr* 2004;92:671–8.
- [15] Piironen V, Lampi AM. Occurrence and levels of phytosterols in foods. In: Dutta PC, editor. *Phytosterols as functional food components and nutraceuticals*. New York: Marcel Dekker, Inc; 2004. pp. 1–32.
- [16] Vuoristo M, Miettinen TA. Absorption, metabolism, and serum concentrations of cholesterol in vegetarians: effects of cholesterol feeding. *Am J Clin Nutr* 1994;59:1325–31.
- [17] Ostlund Jr RE, McGill JB, Zeng C-M, et al. Gastro-intestinal absorption and plasma kinetics of soy Δ5-phytosterols and phytostanols in humans. *Am J Physiol Endocrinol Metab* 2002;282:E911–6.
- [18] Björkhem I, Boberg KM, Leitersdorf E. Inborn errors in bile acid biosynthesis and storage of sterols other than cholesterol. In: Scriver CR, Beaudet AL, Sly WS, Valle D, editors. *The metabolic and molecular bases of inherited disease*. 8th ed. New York: McGraw-Hill; 2001. pp. 2961–88.
- [19] Fransen HP, de Jong N, Wolfs M, et al. Customary use of plant sterol and plant stanol enriched margarine is associated with changes in serum plant sterol and stanol concentrations in humans. *J Nutr* 2007;137:1301–6.
- [20] Berge KE, Tian H, Graf GA, et al. Accumulation of dietary cholesterol in sitosterolemia caused by mutations in adjacent ABC transporters. *Science* 2000;290:1771–5.
- [21] Lee MH, Lu K, Hazard S, et al. Identification of a gene, ABCG5, important in the regulation of dietary cholesterol absorption. *Nat Genet* 2001;27:79–83.

- [22] Salen G, von Bergmann K, Lütjohann D, et al. Multicenter Sitosterolemia Study Group. Ezetimibe effectively reduces plasma plant sterols in patients with sitosterolemia. *Circulation* 2004;109:966–71.
- [23] Vanmierlo T, Popp J, Kölsch H, et al. The plant sterol brassicasterol as additional CSF biomarker in Alzheimer's disease. *Acta Psychiatr Scand* 2011;124:184–92.
- [24] Salen G, Horak I, Rothkopf M, et al. Lethal atherosclerosis associated with abnormal plasma and tissue sterol composition in sitosterolemia with xanthomatosis. *J Lipid Res* 1985;26:1126–33.
- [25] Helske S, Miettinen T, Gylling H, et al. Accumulation of cholesterol precursors and plant sterols in human stenotic aortic valves. *J Lipid Res* 2008;49:1511–8.
- [26] Miettinen TA, Railo M, Lepäntalo M, Gylling H. Plant sterols in serum and in atherosclerotic plaques of patients undergoing carotid endarterectomy. *J Am Coll Cardiol* 2005;45:1792–801.
- [27] Miettinen TA, Nissinen M, Lepäntalo M, et al. Non-cholesterol sterols in serum and endarterectomized carotid arteries after a short-term plant stanol and sterol ester challenge. *Nutr Metab Cardiovasc Dis* 2011;21:182–8.
- [28] Weingärtner O, Lütjohann D, Ji S, et al. Vascular effects of diet supplementation with plant sterols. *J Am Coll Cardiol* 2008;51:1553–61.
- [29] Mellies MJ, Ishikawa TT, Glueck CJ, Bove K, Morrison J. Phytosterols in aortic tissue in adults and infants. *J Lab Clin Med* 1976;88:914–21.
- [30] Jessup W, Herman A, Chapman MJ. Phytosterols in cardiovascular disease: innocuous dietary components, or accelerators of atherosclerosis? *Fut Lipidol* 2008;3:301–10.
- [31] Gylling H, Radhakrishnan R, Miettinen TA. Reduction of serum cholesterol in postmenopausal women with previous myocardial infarction and cholesterol malabsorption induced by dietary sitostanol ester margarine: women and dietary sitostanol. *Circulation* 1997;96:4226–31.
- [32] Miettinen TA, Gylling H, Nissinen MJ. The role of serum non-cholesterol sterols as surrogate markers of absolute cholesterol synthesis and absorption. *Nutr Metab Cardiovasc Dis* 2011;21:765–9.
- [33] Descamps OS, De Sutter J, Guillaume M, Missault L. Where does the interplay between cholesterol absorption and synthesis in the context of statin and/or ezetimibe treatment stand today? *Atherosclerosis* 2011;217:308–21.
- [34] Rozner S, Garti N. The activity and absorption relationship of cholesterol and phytosterols. *Colloids Surf A: Physicochem Eng Aspects* 2006;282–283:435–56.
- [35] Wang DQ-H. Regulation of intestinal cholesterol absorption. *Annu Rev Physiol* 2007;69:221–48.
- [36] Turley SD. Role of Niemann-Pick C1-Like 1 (NPC1L1) in intestinal sterol absorption. *J Clin Lipidol* 2008;2:S20–8.
- [37] Nissinen M, Gylling H, Vuoristo M, Miettinen TA. Micellar distribution of cholesterol and phytosterols after duodenal plant stanol ester infusion. *Am J Physiol Gastrointest Liver Physiol* 2002;282:G1009–15.
- [38] Nguyen TM, Sawyer JK, Kelley KL, Davis MA, Rudel LL. Cholesterol esterification by ACAT2 is essential for efficient intestinal cholesterol absorption: evidence from thoracic lymph duct cannulation. *J Lipid Res* 2012;53:95–104.
- [39] Gylling HK, Hallikainen M, Vidgren H, Agren J, Miettinen TA. Ester percentages of plant sterols and cholesterol in chylomicrons and VLDL of humans with low and high sterol absorption. *Atherosclerosis* 2006;187:150–2.
- [40] Calandra S, Tarugi P, Speedy HE, Dean AF, Bertolini S, Shoulders CC. Mechanisms and genetic determinants regulating sterol absorption, circulating LDL levels, and sterol elimination: implications for classification and disease risk. *J Lipid Res* 2011;52:1885–926.
- [41] Lee S, Gershkovich P, Darlington J, Wasan K. Inhibition of cholesterol absorption: targeting the intestine. *Pharm Res* 2012;29:3235–50.
- [42] De Smet E, Mensink RP, Plat J. Effects of plant sterols and stanols on intestinal cholesterol metabolism: suggested mechanisms from past to present. *Mol Nutr Food Res* 2012;56:1058–72.
- [43] Teupser D, Baber R, Ceglarek U, et al. Genetic regulation of serum phytosterol levels and risk of coronary artery disease. *Circ Cardiovasc Genet* 2010;3:331–9.
- [44] Silbernagel G, Chapman MJ, Genser B, et al. High intestinal cholesterol absorption is associated with risk alleles in ABCG8 and ABO and with cardiovascular disease: evidence from the LURIC and YFS Cohorts and from a meta-analysis. *J Am Coll Cardiol* 2013;62:291–9.
- [45] Andersson SW, Skinner J, Ellegård L, et al. Intake of plant sterols is inversely related to serum cholesterol concentration in men and women in the EPIC Norfolk population: a cross-sectional study. *Eur J Clin Nutr* 2004;58:1378–85.
- [46] Klingberg S, Ellegård L, Johansson I, et al. Inverse relation between naturally occurring dietary plant sterols and serum cholesterol in northern Sweden. *Am J Clin Nutr* 2008;87:993–1002.
- [47] Wang P, Chen YM, He LP, et al. Association of natural intake of dietary plant sterols with carotid intima-media thickness and blood lipids in Chinese adults: a cross-section study. *PLoS ONE* 2012;7:e32736.
- [48] Lin X, Racette SB, Lefevre M, et al. The effects of phytosterols present in natural food matrices on cholesterol metabolism and LDL-cholesterol: a controlled feeding trial. *Eur J Clin Nutr* 2010;64:1481–7.
- [49] Racette SB, Lin X, Lefevre M, et al. Dose effects of dietary phytosterols on cholesterol metabolism: a controlled feeding study. *Am J Clin Nutr* 2010;91:32–8.
- [50] Farquhar JW, Smith RE, Dempsey ME. The effect of beta sitosterol on the serum lipids of young men with arteriosclerotic heart disease. *Circulation* 1956;14:77–82.
- [51] Miettinen TA, Puska P, Gylling H, Vanhanen H, Vartiainen E. Reduction of serum cholesterol with sitostanol-ester margarine in a mildly hypercholesterolemic population. *N Engl J Med* 1995;333:1308–12.
- [52] Katan MB, Grundy SM, Jones P, Law M, Miettinen T, Paoletti R, Stresa Workshop Participants. Efficacy and safety of plant stanols and sterols in the management of blood cholesterol levels. *Mayo Clin Proc* 2003;78:965–78.
- [53] Demonty I, Ras RT, van der Knaap HC, et al. Continuous dose-response relationship of the LDL-cholesterol-lowering effect of phytosterol intake. *J Nutr* 2009;139:271–84.
- [54] Musa-Veloso K, Poon TH, Elliot JA, Chung C. A comparison of the LDL-cholesterol efficacy of plant stanols and plant sterols over a continuous range: Results of a meta-analysis of randomized, placebo-controlled trials. *Prostaglandins Leukot Essent Fatty Acids* 2011;85:9–28.
- [55] Demonty I, Ras RT, Trautwein EA. Comment on: "A comparison of the LDL-cholesterol lowering efficacy of plant stanols and plant sterols over a continuous dose range: results of a meta-analysis of randomized, placebo controlled trials" by Musa-Veloso K, Poon T H, Elliot JA, Chung C. *Prostaglandins Leukot Essent Fatty Acids* 2011;85:9–28.
- [56] Reiner Z, Catapano AL, De Backer G, et al. ESC/EAS Guidelines for the management of dyslipidaemias: the Task Force for the management of dyslipidaemias of the European Society of Cardiology (ESC) and the European Atherosclerosis Society (EAS). *Eur Heart J* 2011;32:1769–818.
- [57] Williams CL, Bollella MC, Strobino BA, Boccia L, Campanaro L. Plant stanol ester and bran fiber in childhood: effects on lipids, stool weight and stool frequency in preschool children. *J Am Coll Nutr* 1999;18:572–81.
- [58] Tammi A, Rönnemaa T, Miettinen TA, et al. Effects of gender, apolipoprotein E phenotype and cholesterol-lowering by plant stanol esters in children: The STRIP study. *Acta Paediatr* 2002;91:1155–62.
- [59] Guardamagna O, Abello F, Baracco V, et al. Primary hyperlipidemias in children: effect of plant sterol supplementation on plasma lipids and markers of cholesterol synthesis and absorption. *Acta Diabetol* 2011;48:127–33.
- [60] Becker M, Staab D, von Bergmann K. Treatment of severe familial hypercholesterolemia in childhood with sitosterol and sitostanol. *J Pediatr* 1993;122:292–6.
- [61] Gylling H, Siimes MA, Miettinen TA. Sitostanol ester margarine in dietary treatment of children with familial hypercholesterolemia. *J Lipid Res* 1995;36:1807–12.
- [62] Vuorio AF, Gylling H, Turtola H, Kontula K, Ketonen P, Miettinen TA. Stanol ester margarine alone and with simvastatin lowers serum cholesterol in families with familial hypercholesterolemia caused by the FH-North Karelia mutation. *Arterioscler Thromb Vasc Biol* 2000;20:500–6.
- [63] Amundsen AL, Ose L, Nenseter MS, Ntanios FY. Plant sterol ester-enriched spread lowers plasma total and LDL cholesterol in children with familial hypercholesterolemia. *Am J Clin Nutr* 2002;76:338–44.
- [64] de Jongh S, Vissers MN, Rol P, Bakker HD, Kastelein JJ, Stroes ES. Plant sterols lower LDL cholesterol without improving endothelial function in prepubertal children with familial hypercholesterolemia. *J Inherit Metab Dis* 2003;26:343–51.
- [65] Ketomäki AM, Gylling H, Antikainen M, Siimes MA, Miettinen TA. Red cell and plasma plant sterols are related during consumption of plant stanol and sterol ester spreads in children with hypercholesterolemia. *J Pediatr* 2003;142:524–31.
- [66] Jakulj L, Vissers MN, Rodenburg J, Wiegman A, Trip MD, Kastelein JJ. Plant stanols do not restore endothelial function in prepubertal children with familial hypercholesterolemia despite reduction of low-density lipoprotein cholesterol levels. *J Pediatr* 2006;148:495–500.
- [67] Plat J, Mensink RP. Plant stanol esters lower serum triacylglycerol concentrations via a reduced hepatic VLDL-1 production. *Lipids* 2009;44:1149–53.
- [68] Sialvera TE, Pounis GD, Koutelidakis AE, et al. Phytosterols supplementation decreases plasma small and dense LDL levels in metabolic syndrome patients on a westernized type diet. *Nutr Metab Cardiovasc Dis* 2012;22:843–8.
- [69] Demonty I, Ras RT, van der Knaap HC, et al. The effect of plant sterols on serum triglyceride concentrations is dependent on baseline concentrations: a pooled analysis of 12 randomised controlled trials. *Eur J Nutr* 2013;52:153–60.
- [70] Naumann E, Plat J, Kester ADM, Mensink RP. The baseline serum lipoprotein profile is related to plant stanol induced changes in serum lipoprotein cholesterol and triacylglycerol concentrations. *J Am Coll Nutr* 2008;27:117–26.
- [71] Gylling H, Miettinen TA. Serum cholesterol and cholesterol and lipoprotein metabolism in hypercholesterolaemic NIDDM patients before and during sitostanol ester-margarine treatment. *Diabetologia* 1994;37:773–80.
- [72] Plat J, Mensink RP. Vegetable oil based versus wood based stanol ester mixtures: effects on serum lipids and hemostatic factors in non-hypercholesterolemic subjects. *Atherosclerosis* 2000;148:101–12.
- [73] Mayne J, Dewpura T, Raymond A, et al. Plasma PCSK9 levels are significantly modified by statins and fibrates in humans. *Lipids Health Dis* 2008;7:22.
- [74] Kritchevsky D, Chen SC. Phytosterols – health benefits and potential concerns: a review. *J Nutr Res* 2005;25:413–28.
- [75] Plat J, Beugels I, Gijbels MJ, de Winther MP, Mensink RP. Plant sterol or stanol esters retard lesion formation in LDL receptor-deficient mice independent of changes in serum plant sterols. *J Lipid Res* 2006;47:2762–71.
- [76] Weingärtner O, Ulrich C, Lütjohann D, et al. Differential effects on inhibition of cholesterol absorption by plant stanol and plant sterol esters in apoE^{-/-} mice. *Cardiovasc Res* 2011;90:484–92.

- [77] Hovenkamp E, Loubakos A, Duchateau GS, Tareilus EW, Trautwein EA. Preferential efflux of phytosterols over cholesterol from macrophages. *Lipids* 2007;42:1125–32.
- [78] Bao L, Li Y, Deng SX, Landry D, Tabas I. Sitosterol-containing lipoproteins trigger free sterol-induced caspase-independent death in ACAT-competent macrophages. *J Biol Chem* 2006;281:33635–49.
- [79] Tabas I, Feinmark SJ, Beatini N. The reactivity of desmosterol and other shellfish- and xanthomatosis-associated sterols in the macrophage sterol esterification reaction. *J Clin Invest* 1989;84:1713–21.
- [80] Plat J, Nichols JA, Mensink RP. Plant sterols and stanols: effects on mixed micellar composition and LXR (target gene) activation. *J Lipid Res* 2005;46:2468–76.
- [81] Brauner R, Johannes C, Ploessl F, Bracher F, Lorenz RL. Phytosterols reduce cholesterol absorption by inhibition of 27-hydroxycholesterol generation, liver X receptor α activation, and expression of the basolateral sterol exporter ATP-binding cassette A1 in Caco-2 enterocytes. *J Nutr* 2012;142:981–9.
- [82] Kaneko E, Matsuda M, Yamada Y, Tachibana Y, Shimomura I, Makishima M. Induction of intestinal ATP-binding cassette transporters by a phytosterol-derived liver X receptor agonist. *J Biol Chem* 2003;278:36091–8.
- [83] Yang C, Yu L, Li W, Xu F, Cohen JC, Hobbs HH. Disruption of cholesterol homeostasis by plant sterols. *J Clin Invest* 2004;114:813–22.
- [84] Sabeva NS, McPhaul CM, Li X, Cory TJ, Feola DJ, Graf GA. Phytosterols differentially influence ABC transporter expression, cholesterol efflux and inflammatory cytokine secretion in macrophage foam cells. *J Nutr Biochem* 2011;22:777–83.
- [85] Calpe-Berdiel L, Escolá-Gil JC, Blanco-Vaca F. Are LXR-regulated genes a major molecular target of plant sterols/stanols? *Atherosclerosis* 2007;195:210–1.
- [86] Joseph SB, Castrillo A, Laffitte BA, Mangelsdorf DJ, Tontonoz P. Reciprocal regulation of inflammation and lipid metabolism by liver X receptors. *Nat Med* 2003;9:213–9.
- [87] Spann NJ, Garmire LX, McDonald JG, et al. Regulated accumulation of desmosterol integrates macrophage lipid metabolism and inflammatory responses. *Cell* 2012;151:138–52.
- [88] Bouic PJ, Lamprecht JH. Plant sterols and sterolins: a review of their immunomodulating properties. *Altern Med Rev* 1999;4:170–7.
- [89] Ding Y, Nguyen HT, Kim SI, Kim HW, Kim YH. The regulation of inflammatory cytokine secretion in macrophage cell line by the chemical constituents of *Rhus sylvestris*. *Bioorg Med Chem Lett* 2009;19:3607–10.
- [90] Alappat L, Valerio M, Awad AB. Effect of vitamin D and β -sitosterol on immune function of macrophages. *Int Immunopharmacol* 2010;10:1390–6.
- [91] Kurano M, Iso ON, Hara M, et al. Plant sterols increased IL-6 and TNF- α secretion from macrophages, but to a lesser extent than cholesterol. *J Atheroscler Thromb* 2011;18:373–83.
- [92] Brüll F, Mensink RP, van den Hurk K, Duijvestijn A, Plat J. TLR2 activation is essential to induce a Th1 shift in human peripheral blood mononuclear cells by plant stanols and plant sterols. *J Biol Chem* 2010;285:2951–8.
- [93] Moreno JJ. Effect of olive oil minor components on oxidative stress and arachidonic acid mobilization and metabolism by macrophages RAW 264.7. *Free Radic Biol Med* 2003;35:1073–81.
- [94] Vivancos M, Moreno JJ. Beta-sitosterol modulates antioxidant enzyme response in RAW 264.7 macrophages. *Free Radic Biol Med* 2005;39:91–7.
- [95] Vivancos M, Moreno JJ. Effect of resveratrol, tyrosol and beta-sitosterol on oxidised low-density lipoprotein-stimulated oxidative stress, arachidonic acid release and prostaglandin E2 synthesis by RAW 264.7 macrophages. *Br J Nutr* 2008;99:1199–207.
- [96] Awad AB, Toczek J, Fink CS. Phytosterols decrease prostaglandin release in cultured P388D1/MAB macrophages. *Prostaglandins Leukot Essent Fatty Acids* 2004;70:511–20.
- [97] Tedgui A, Mallat Z. Cytokines in atherosclerosis: pathogenic and regulatory pathways. *Physiol Rev* 2006;86:515–81.
- [98] Raitakari OT, Salo P, Gylling H, Miettinen TA. Plant stanol ester consumption and arterial elasticity and endothelial function. *Br J Nutr* 2008;100:603–8.
- [99] Gylling H, Hallikainen M, Raitakari OT, et al. Long-term consumption of plant stanol and sterol esters, vascular function and genetic regulation. *Br J Nutr* 2009;101:1688–95.
- [100] Hallikainen M, Lyyra-Laitinen T, Laitinen T, et al. Endothelial function in hypercholesterolemic subjects: effects of plant stanol and sterol esters. *Atherosclerosis* 2006;188:425–32.
- [101] Hallikainen M, Lyyra-Laitinen T, Laitinen T, Moilanen L, Miettinen TA, Gylling H. Effects of plant stanol esters on serum cholesterol concentrations, relative markers of cholesterol metabolism and endothelial function in type 1 diabetes. *Atherosclerosis* 2008;199:432–9.
- [102] Gylling H, Halonen J, Lindholm H, et al. The effects of plant stanol ester consumption on arterial stiffness and endothelial function in adults: a randomised controlled clinical trial. *BMC Cardiovasc Disord* 2013;13:50.
- [103] Raitakari OT, Salo P, Ahotupa M. Carotid artery compliance in users of plant stanol ester margarine. *Eur J Clin Nutr* 2008;62:218–24.
- [104] Wang JJ, Liew G, Wong TY, et al. Retinal vascular calibre and the risk of coronary heart disease-related death. *Heart* 2006;92:1583–7.
- [105] Kelly ER, Plat J, Mensink RP, Berendschot TJJM. Effects of long term plant sterol and -stanol consumption on the retinal vasculature: a randomized controlled trial in statin users. *Atherosclerosis* 2011;214:225–30.
- [106] Kozłowska-Wojciechowska M, Jastrzebska M, Naruszewicz M, Foltynska A. Impact of margarine enriched with plant sterols on blood lipids, platelet function, and fibrinogen level in young men. *Metabolism* 2003;52:1373–8.
- [107] De Jong A, Plat J, Bast A, Godschalk RW, Basu S, Mensink RP. Effects of plant sterol and stanol ester consumption on lipid metabolism, antioxidant status and markers of oxidative stress, endothelial function and low-grade inflammation in patients on current statin treatment. *Eur J Clin Nutr* 2008;62:263–73.
- [108] Othman RA, Moghadasian MH. Beyond cholesterol-lowering effects of plant sterols: clinical and experimental evidence of anti-inflammatory properties. *Nutr Rev* 2011;69:371–82.
- [109] Sudhop T, Gottwald BM, von Bergmann K. Serum plant sterols as a potential risk factor for coronary heart disease. *Metabolism* 2002;51:1519–21.
- [110] Assmann G, Cullen P, Erbey J, Ramey DR, Kannenberg F, Schulte H. Plasma sitosterol elevations are associated with an increased incidence of coronary events in men: results of a nested case-control analysis of the Prospective Cardiovascular Münster (PROCAM) study. *Nutr Metab Cardiovasc Dis* 2006;16:13–21.
- [111] Pinedo S, Vissers MN, von Bergmann K, et al. Plasma levels of plant sterols and the risk of coronary artery disease: the prospective EPIC-Norfolk Population Study. *J Lipid Res* 2007;48:139–44.
- [112] Windler E, Zyriax BC, Kuipers F, Linseisen J, Boeing H. Association of plasma phytosterol concentrations with incident coronary heart disease Data from the CORA study, a case-control study of coronary artery disease in women. *Atherosclerosis* 2009;203:284–90.
- [113] Escurriol V, Cofán M, Moreno-Iribas C, et al. Phytosterol plasma concentrations and coronary heart disease in the prospective Spanish EPIC cohort. *J Lipid Res* 2010;51:618–24.
- [114] Genser B, Silbernagel G, De Backer G, et al. Plant sterols and cardiovascular disease: a systematic review and meta-analysis. *Eur Heart J* 2012;33:444–51.
- [115] Tilvis RS, Valvanne JN, Strandberg TE, Miettinen TA. Prognostic significance of serum cholesterol, lathosterol, and sitosterol in old age; a 17-year population study. *Ann Med* 2011;43:292–301.
- [116] Weingärtner O, Pinsdorf T, Rogacev KS, et al. The relationships of markers of cholesterol homeostasis with carotid intima-media thickness. *PLoS ONE* 2010;5:e13467.
- [117] Weingärtner O, Lütjohann D, Vanmierlo T, et al. Markers of enhanced cholesterol absorption are a strong predictor for cardiovascular diseases in patients without diabetes mellitus. *Chem Phys Lipids* 2011;164:451–6.
- [118] Fukushima M, Miura S, Mitsutake R, Fukushima T, Fukushima K, Saku K. Cholesterol metabolism in patients with hemodialysis in the presence or absence of coronary artery disease. *Circ J* 2012;76:1980–6.
- [119] IBC 50K CAD Consortium. Large-scale gene-centric analysis identifies novel variants for coronary artery disease. *PLoS Genet* 2011;7:e1002260.
- [120] Reilly MP, Li M, He J, et al., Myocardial Infarction Genetics Consortium, Wellcome Trust Case Control Consortium. Identification of ADAMT57 as a novel locus for coronary atherosclerosis and association of ABO with myocardial infarction in the presence of coronary atherosclerosis: two genome-wide association studies. *Lancet* 2011;377:383–92.
- [121] Schunkert H, König IR, Kathiresan S, et al., CARDIoGRAM Consortium. Large scale association analysis identifies 13 new susceptibility loci for coronary artery disease. *Nat Genet* 2011;43:333–8.
- [122] CARDIoGRAMplusC4D Consortium, Deloukas P, Kanoni S, Willenborg C, DIAGRAM Consortium, CARDIOGENICS Consortium, MuTHER Consortium, Wellcome Trust Case Control Consortium. Large-scale association analysis identifies new risk loci for coronary artery disease. *Nat Genet* 2013;45:25–33.
- [123] Strandberg TE, Tilvis RS, Pitkala KH, Miettinen TA. Cholesterol and glucose metabolism and recurrent cardiovascular events among the elderly: a prospective study. *J Am Coll Cardiol* 2006;48:708–14.
- [124] Silbernagel G, Fauler G, Renner W, et al. The relationships of cholesterol metabolism and plasma plant sterols with the severity of coronary artery disease. *J Lipid Res* 2009;50:334–41.
- [125] Matthan NR, Pencina M, LaRocque JM, et al. Alterations in cholesterol absorption/synthesis markers characterize Framingham offspring study participants with CHD. *J Lipid Res* 2009;50:1927–35.
- [126] Silbernagel G, Fauler G, Hoffmann MM, et al. The associations of cholesterol metabolism and plasma plant sterols with all-cause and cardiovascular mortality. *J Lipid Res* 2010;51:2384–93.
- [127] Smith NL, Chen MH, Dehghan A, et al., Wellcome Trust Case Control Consortium. Novel associations of multiple genetic loci with plasma levels of factor VII, factor VIII, and von Willebrand factor: the CHARGE (Cohorts for Heart and Aging Research in Genome Epidemiology) Consortium. *Circulation* 2010;121:1382–92.
- [128] Kiechl S, Paré G, Barbalić M, et al. Association of variation at the ABO locus with circulating levels of soluble intercellular adhesion molecule-1, soluble P-selectin, and soluble E-selectin: a meta-analysis. *Circ Cardiovasc Genet* 2011;4:681–6.
- [129] Karakas M, Baumert J, Kleber ME, et al. A variant in the abo gene explains the variation in soluble e-selectin levels—results from dense genotyping in two independent populations. *PLoS ONE* 2012;7:e51441.
- [130] Fassbender K, Lütjohann D, Dik MG, et al. Moderately elevated plant sterol levels are associated with reduced cardiovascular risk—the LASA study. *Atherosclerosis* 2008;196:283–8.
- [131] Buchwald H, Varco RL, Boen JR, et al. Effective lipid modification by partial ileal bypass reduced long-term coronary heart disease mortality and morbidity: five-year posttrial follow-up report from the POSCH. Program on the Surgical Control of the Hyperlipidemias. *Arch Intern Med* 1998;158:1253–61.

- [132] Brown L, Rosner B, Willett WW, Sacks FM. Cholesterol-lowering effects of dietary fiber: a meta-analysis. *Am J Clin Nutr* 1999;69:30–42.
- [133] Pereira MA, O'Reilly E, Augustsson K, et al. Dietary fiber and risk of coronary heart disease: a pooled analysis of cohort studies. *Arch Intern Med* 2004;164:370–6.
- [134] Willems JI, Blommaert MAE, Trautwein EA. Results from a post-launch monitoring survey on consumer purchases of foods with added phytosterols in five European countries. *Food Chem Toxicol* 2013;62:48–53.
- [135] Lea LJ, Hepburn PA. Safety evaluation of phytosterol-esters. Part 9: Results of a European post-launch monitoring programme. *Food Chem Toxicol* 2006;44:1213–22.
- [136] EFSA Panel on Food Additives and Nutrient Sources added to Food (ANS). Scientific opinion on the safety of stigmasterol-rich plant sterols as food additive. *EFSA J* 2012;10:2659.
- [137] Hendriks HF, Brink EJ, Meijer GW, Princen HM, Ntanos FY. Safety of long-term consumption of plant sterol esters-enriched spread. *Eur J Clin Nutr* 2003;57:681–92.
- [138] de Jong A, Plat J, Lütjohann D, Mensink RP. Effects of long-term plant sterol or stanol ester consumption on lipid and lipoprotein metabolism in subjects on statin treatment. *Br J Nutr* 2008;100:937–41.
- [139] Noakes M, Clifton P, Ntanos F, Shrapnel W, Record I, McInerney J. An increase in dietary carotenoids when consuming plant sterols or stanols is effective in maintaining plasma carotenoid concentrations. *Am J Clin Nutr* 2002;75:79–86.
- [140] Tuomilehto J, Tikkanen MJ, Högström P, et al. Safety assessment of common foods enriched with natural nonesterified plant sterols. *Eur J Clin Nutr* 2009;63:684–91.
- [141] Plat J, Mensink RP. Effects of diets enriched with two different plant stanol ester mixtures on plasma ubiquinol-10 and fat-soluble antioxidant concentrations. *Metabolism* 2001;50:520–9.
- [142] Baskar AA, Ignacimuthu S, Paulraj GM, Al Numair KS. Chemopreventive potential of beta-sitosterol in experimental colon cancer model – an in vitro and in vivo study. *BMC Compl Altern Med* 2010;10:24.
- [143] Lea LJ, Hepburn PA, Wolfreys AM, Baldrick P. Safety evaluation of phytosterol esters. Part 8. Lack of genotoxicity and subchronic toxicity with phytosterol oxides. *Food Chem Toxicol* 2004;42:771–83.
- [144] Wolfreys AM, Hepburn PA. Safety evaluation of phytosterol esters. Part 7. Assessment of mutagenic activity of phytosterols, phytosterol esters and the cholesterol derivative, 4-cholesten-3-one. *Food Chem Toxicol* 2002;40:461–70.
- [145] Woyengo TA, Ramprasath VR, Jones PJ. Anticancer effects of phytosterols. *Eur J Clin Nutr* 2009;63:813–20.
- [146] De Stefani E, Boffetta P, Ronco AL, et al. Plant sterols and risk of stomach cancer: a case-control study in Uruguay. *Nutr Cancer* 2000;37:140–4.
- [147] Mendilaharsu M, De Stefani E, Deneo-Pellegrini H, Carzoglio J, Ronco A. Phytosterols and risk of lung cancer: a case-control study in Uruguay. *Lung Cancer* 1998;21:37–45.
- [148] Blair SN, Capuzzi DM, Gottlieb SO, Nguyen T, Morgan JM, Cater NB. Incremental reduction of serum total cholesterol and low-density lipoprotein cholesterol with the addition of plant stanol ester-containing spread to statin therapy. *Am J Cardiol* 2000;86:46–52.
- [149] Neil HA, Meijer GW, Roe LS. Randomised controlled trial of use by hypercholesterolaemic patients of a vegetable oil sterol-enriched fat spread. *Atherosclerosis* 2001;156:329–37.
- [150] Castro Cabezas M, de Vries JH, Van Oostrom AJ, Iestra J, van Staveren WA. Effects of a stanol-enriched diet on plasma cholesterol and triglycerides in patients treated with statins. *J Am Diet Assoc* 2006;106:1564–9.
- [151] Simons LA. Additive effect of plant sterol-ester margarine and cerivastatin in lowering low-density lipoprotein cholesterol in primary hypercholesterolemia. *Am J Cardiol* 2002;90:737–40.
- [152] Gylling H, Miettinen TA. Effects of inhibiting cholesterol absorption and synthesis on cholesterol and lipoprotein metabolism in hypercholesterolemic non-insulin-dependent diabetic men. *J Lipid Res* 1996;37:1776–85.
- [153] Lin X, Racette SB, Lefevre M, et al. Combined effects of ezetimibe and phytosterols on cholesterol metabolism: a randomized, controlled feeding study in humans. *Circulation* 2011;124:596–601.
- [154] Becker M, Staab D, Von Bergman K. Long-term treatment of severe familial hypercholesterolemia in children: effect of sitosterol and bezafibrate. *Pediatrics* 1992;89:138–42.
- [155] Nigon F, Serfaty-Lacrosnière C, Beucler I, et al. Plant sterol-enriched margarine lowers plasma LDL in hyperlipidemic subjects with low cholesterol intake: effect of fibrate treatment. *Clin Chem Lab Med* 2001;39:634–40.
- [156] Micallef MA, Garg ML. Beyond blood lipids: phytosterols, statins and omega-3 polyunsaturated fatty acid therapy for hyperlipidemia. *J Nutr Biochem* 2009;20:927–39.
- [157] O'Neill FH, Brynes A, Mandeno R, et al. Comparison of the effects of dietary plant sterol and stanol esters on lipid metabolism. *Nutr Metab Cardiovasc Dis* 2004;14:133–42.
- [158] Relas H, Gylling H, Miettinen TA. Effect of stanol ester on postabsorptive squalene and retinyl palmitate. *Metabolism* 2000;49:473–8.
- [159] Gylling H, Hallikainen M, Simonen P, Miettinen HE, Nissinen MJ, Miettinen TA. Serum and lipoprotein sitostanol and non-cholesterol sterols after an acute dose of plant stanol ester on its long-term consumption. *Eur J Nutr* 2012;56:663–70.
- [160] Demonty I, Chan YM, Pelled D, Jones PJ. Fish-oil esters of plant sterols improve the lipid profile of dyslipidemic subjects more than do fish-oil or sunflower oil esters of plant sterols. *Am J Clin Nutr* 2006;84:1534–42.
- [161] Darmon N, Drewnowski A. Does social class predict diet quality? *Am J Clin Nutr* 2008;87:1107–17.
- [162] Bonaccio M, Bonanni AE, Di Castelnuovo A, et al. Low income is associated with poor adherence to a Mediterranean diet and a higher prevalence of obesity: cross-sectional results from the Moli-sani study. *BMJ Open* 2012;2:e001685. <http://dx.doi.org/10.1136/bmjopen-2012-001685>.
- [163] European Food Safety Authority. A report from the data collection and exposure unit in response to a request from the European Commission. *EFSA J* 2008;133:1–21. http://www.efsa.europa.eu/en/scdocs/doc/datex_report_ej133_phytosterols_en.pdf [accessed 04.07.13].
- [164] Bes-Rastrollo M. Costs of Mediterranean and Western dietary patterns and their relationship with prospective weight change. *EuroPrevent* 2013 [abstract 610].