



THE AGA KHAN UNIVERSITY

eCommons@AKU

Department of Pathology and Laboratory Medicine

Medical College, Pakistan

January 2014

β -lactamase production and antimicrobial susceptibility pattern of moraxella catarrhalis isolates: report from Pakistan

Sadia Omer Sheikh
Aga Khan University

Naima Fasih
Aga Khan University

Seema Irfan
Aga Khan University

Afia Zafar
Aga Khan University, afia.zafar@aku.edu

Follow this and additional works at: http://ecommons.aku.edu/pakistan_fhs_mc_pathol_microbiol



Part of the [Pathology Commons](#)

Recommended Citation

Sheikh, S., Fasih, N., Irfan, S., Zafar, A. (2014). β -lactamase production and antimicrobial susceptibility pattern of moraxella catarrhalis isolates: report from Pakistan. *Asian Pacific Journal of Tropical Medicine*, 7(1), S228-S231.

Available at: http://ecommons.aku.edu/pakistan_fhs_mc_pathol_microbiol/601



Contents lists available at ScienceDirect

Asian Pacific Journal of Tropical Medicine

journal homepage: www.elsevier.com/locate/apjtm



Document heading doi: 10.1016/S1995-7645(14)60237-6

β -Lactamase production and antimicrobial susceptibility pattern of *Moraxella catarrhalis* isolates: report from Pakistan

Sadia Omer Sheikh, Naima Fasih, Seema Irfan, Afia Zafar*

Department of Pathology and Microbiology, Aga Khan University Hospital, Karachi, Pakistan

ARTICLE INFO

Article history:

Received 6 Dec 2013

Received in revised form 15 Mar 2014

Accepted 3 May 2014

Available online 28 Jul 2014

Keywords

Moraxella catarrhalis

Antimicrobial resistance in Pakistan

 β -lactamase enzyme*Moraxella catarrhalis* resistance pattern in Pakistan

ABSTRACT

Objective: To assess the frequency of β -lactamase production and antimicrobial resistance in *Moraxella catarrhalis* isolated from clinical specimens in Pakistan.

Methods: This cross sectional study (January to December 2010) was conducted in clinical microbiology laboratory of Aga Khan University Hospital. A total of 97 clinical respiratory specimens growing *Moraxella catarrhalis* were included. Frequency of β -lactamase production and antimicrobial resistance rates against ampicillin, erythromycin, ciprofloxacin and tetracycline were noted by performing minimum inhibitory concentration (MIC). MICs were calculated as MIC₅₀ and MIC₉₀.

Results: β -Lactamase production was detected in 84% of isolates, which correlated well with high MIC of ampicillin. Majority of isolates were susceptible to erythromycin (97%) and tetracycline (96%) with MIC₅₀=0.12 mg/L and MIC₉₀=1 mg/L respectively. All isolates were found susceptible to ciprofloxacin (MIC₅₀=0.06 mg/L).

Conclusions: Result suggests that empirical use of ampicillin should be discouraged while treating respiratory tract infections. This also emphasizes the importance of continuous surveillance in order to detect emerging resistance in *Moraxella* isolates.

1. Introduction

Over the last few decades the increasing pathogenicity and capacity to produce β -lactamase enzyme has evolved *Moraxella catarrhalis* (*M. catarrhalis*) from a benign commensal to a genuine pathogen^[1,2]. Currently, it is considered as one of the common cause of upper and lower respiratory tract infections. Though rarely reported, it can also cause severe infections including pneumonia, endocarditis, septicaemia and meningitis *etc*^[1-4]. *M. catarrhalis* also contributes in mixed respiratory tract infections with other pathogens such as *Streptococcus pneumoniae* (*S. pneumoniae*) and/or *Haemophilus influenzae* (*H. influenzae*). In such cases, β -lactamase produced by this organism may render ampicillin ineffective against susceptible *S. pneumoniae* and *H.*

influenzae^[5].

Sweden was the first country to report the presence of β -lactamase in *M. catarrhalis* in 1977^[6]. Since then β -lactamase production has been reported from various countries with increasing frequency (even above 90%)^[7-9]. Though it remains rare, reports of increasing resistance to other oral antibiotics useful to treat community acquired respiratory tract infections (CARTI) are also documented in *M. catarrhalis*^[10,11].

Information regarding frequency of β -lactamase production in *M. catarrhalis* is limited from Pakistan. A study from Pakistan reported 93.5% ampicillin susceptibility in *M. catarrhalis* but authors did not report β -lactamase activity in those isolates^[12]. Another study reported 68.7% β -lactamase production in *M. catarrhalis*. However this study had the limitations of small sample size and use of less sensitive filter paper acidometric test for β -lactamase detection^[13]. The local antimicrobial susceptibility data of other oral antibiotics used for CARTI against *M. catarrhalis* is also missing. Surveillance to monitor shifting trends in resistance is vital as it ultimately influences the selection of antimicrobial agents

*Corresponding author: Afia Zafar, Department of Pathology and Microbiology, Aga Khan University Hospital, Stadium Road, Karachi 74800, Pakistan
Tel: 009221 3486 1601, 009221 3486 1641
E-mail: afia.zafar@aku.edu

Foundation Project: Supported by the departmental seed money grant of Aga Khan University Hospital, Karachi, Pakistan (Grant No. SM2011-04).

available for use against a particular organism. Therefore, this study aimed to assess the frequency of β -lactamase production in *M. catarrhalis* and also drug resistance against ampicillin, erythromycin, ciprofloxacin and tetracycline, so as to guide empirical therapy in CARTI.

2. Material and methods

This cross sectional study was conducted from January 2010 to December 2010 in the clinical microbiology laboratory of Aga Khan University Hospital (AKUH). The AKUH laboratory has an extensive network of more than 200 collection units throughout Pakistan. Through this it caters a vast outpatient population across the country.

2.1. Respiratory specimens

Clinical respiratory specimens (sputum, tracheal aspirate, bronchoalveolar lavage, middle ear fluid, nasopharyngeal swab/aspirate and sinus aspirate) from community that were received in AKUH laboratory for culture and sensitivities during study period were included.

2.2. Isolation of *M. catarrhalis*

The identification of the *M. catarrhalis* was confirmed by colony morphology, Gram stain appearance and conventional biochemical tests such as positive oxidase, catalase, and DNAase test. Clinical isolates were saved in glycerol–phosphate buffer at -80°C . Duplicate samples from same patient were excluded. Demographic data including gender and age and seasonal distribution were also noted.

2.3. Antimicrobial susceptibility testing

Ampicillin, erythromycin, ciprofloxacin, and tetracycline powder were obtained from Sigma–Aldrich, UK. Minimum inhibitory concentrations (MICs) of the antimicrobials were performed according to agar dilution method recommended by British Society for Chemotherapy (BSAC) [14]. Agar plates were prepared with the antimicrobials to be tested incorporated in the media supplemented with 5% defibrinated blood in double dilution series. Bacterial suspension 10^4 CFU/spot was prepared and applied by the multipoint inoculator. For testing ampicillin, inoculum of 10^6 CFU was used. Plates were incubated at 37°C aerobically for 18 h. MIC breakpoints provided by the BSAC [15] were used for interpretation and isolates were categorized as being sensitive (S), intermediately susceptible (I), or resistant (R).

Interpretation of the MIC breakpoints was as follows: Ampicillin: $S \leq 1$ mg/L, $R > 1$ mg/L; Ciprofloxacin: $S \leq 0.5$ mg/L, $R > 0.5$ mg/L; Tetracycline: $S \leq 1$ mg/L, $I = 2$ mg/L, $R \geq 2$ mg/L; Erythromycin: $S \leq 0.25$ mg/L, $I = 0.5$ mg/L, $R \geq 0.5$ mg/L.

β -Lactamase production was detected by nitrocefin (Oxoid) test. The reference strains used for quality control included *S. pneumoniae* ATCC strain 49619 and *Staphylococcus aureus* ATCC strain 29213. MICs were calculated as MIC_{50} and MIC_{90} (MIC causing inhibition of 50% and 90% of isolates, respectively).

2.4. Data management and statistical analysis

Data was entered and analysed by the statistical software SPSS version 19.0. Frequencies with percentages for age and seasonal distribution, β -lactamase production and resistance for each drug were computed.

2.5. Ethical approval

This study was approved by the Ethics Review Committee of the Aga Khan University, Pakistan.

3. Results

A total of 4440 respiratory specimens were received during study period. Out of these, 30% (1333) were reported positive for respiratory pathogens. About 60% (2680) specimens were received from male patients.

A total of 97/1333 clinical respiratory specimens yielded growth of *M. catarrhalis*. Amongst studied isolates of *M. catarrhalis*, 71% (69) were from male and 65% (63) were recovered from patients aged greater than 65 years. The age distribution is shown in Table 1.

Table 1

Distribution of *M. catarrhalis* isolates ($n=97$) among different age groups.

Age range in years	Number (%)
0–15	11 (11.5)
16–30	11 (11.5)
31–45	12 (12.0)
46–65	24 (25.0)
>65	39 (40.0)

Majority of isolates (50.5%, 49) were grown in the winter months (November–February) with the peak incidence in the month of January (Figure 1). β -Lactamase production was detected in 84.5% (81) of isolates, which correlated well with the high ampicillin MIC among them. Ninety-seven percent of isolates were found susceptible to erythromycin with MIC_{90} of 0.12 mg/L and ninety-six percent susceptible to tetracycline with MIC_{90} of 1 mg/L. All isolates were susceptible to ciprofloxacin. The MIC distribution for the

four antibiotics tested is given in Table 2.

Table 2

Distribution of MICs ($\mu\text{g/mL}$) for antimicrobials tested against *M. catarrhalis* isolates ($n=97$).

MIC	Ampicillin	Ciprofloxacin	Tetracycline	Erythromycin
128	–	–	–	–
64	2	–	–	–
32	3	–	–	–
16	4	–	–	–
8	17	–	1	–
4	21	–	2	–
2	34	–	1	–
1	9	–	10	2
0.5	4	3	57	1
0.25	–	2	13	1
0.125	3	5	8	41
0.06	–	11	5	40
0.03	–	32	–	12
0.015	–	44	–	–
MIC ₉₀	8	0.06	1	0.125
MIC ₅₀	2	0.03	0.5	0.06
MIC range	0.125–64	0.015–0.5	0.06–8	0.03–1
Sensitive (%)	15.5	100	96	97

BSAC break points of MICs for ampicillin (mg/L): $S \leq 1$, $R > 1$, ciprofloxacin (mg/L): $S \leq 0.5$, $R > 0.5$, tetracycline (mg/L): $S \leq 1$, $I = 2$, $R \geq 2$ and erythromycin (mg/L): $S \leq 0.25$, $I = 0.5$, $R \geq 0.5$.

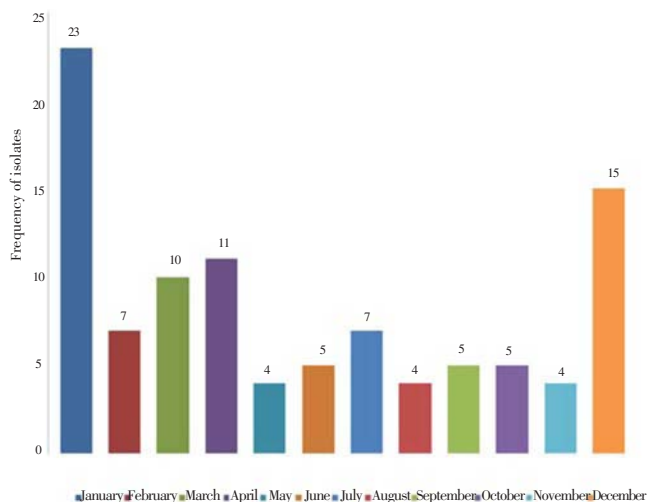


Figure 1. Monthly rate of isolation of *M. catarrhalis*.

4. Discussion

This study suggests that empirical use of ampicillin/amoxicillin in CARTI would miss β -lactamase producing *M. catarrhalis* strains and may lead to treatment failure. It may also hamper the treatment of certain concomitant organisms, such as penicillin susceptible *S. pneumoniae* and *H. influenzae* in cases of co-infection[1,5].

This study demonstrates that frequency of β -lactamase production among *M. catarrhalis* is entirely consistent with

what reported globally[7–9]. Previous study from Pakistan reported 68.7% β -lactamase production. However, in that study they used less sensitive acidometric test for enzyme detection[13]. Increasing frequency of enzyme production was reported from the neighbouring country India. Larson *et al.* reported enzyme production in 68% isolates in 1999 and Anita *et al.* demonstrated 86% of isolates with enzyme activity in 2011[16–17].

In this study, high ampicillin MIC correlates well with high β -lactamase production. Previously published work from Pakistan reported 93.5% susceptibility to this drug. However, that study had major limitation that authors did not test the β -lactamase production in their isolates by the standard method[12].

Another important and promising finding of our study is that, resistance to other commonly used oral antibiotics in CARTI such as tetracycline and erythromycin remained low. Our finding regarding the tetracycline resistance is comparable to those obtained from most Asia pacific regions (3.2%) except Taiwan where it is around 19%[9,18]. Similarly macrolide resistance is comparable to China 5.8%, however it is much lower than that reported previously from Pakistan (25%)[7,12].

Although ciprofloxacin is an accessible over the counter drug in Pakistan and easily available with or without a prescription, fortunately none of the isolate was found to be ciprofloxacin resistant. This finding is consistent with other reports published globally[7–9]. Though Tabussum *et al.* from Karachi reported 7% and a study from India reported 20% ciprofloxacin resistance[12,19].

Globally, recovery of majority of the isolates from respiratory samples from male patients was also observed in this study[20]. We also found that *M. catarrhalis* infections are more frequent among older age group. Similar finding was reported from India and Nepal[20,21]. The predilection of CARTI in the older age group has been studied and several reasons have been proposed including age related changes in systemic immunity, respiratory tract mucosal changes due to smoking and several other co-morbidities[1,2,22].

Majority of isolates were recovered in winter season, which is similar to finding of others[20–22]. A viral co-infection has been anticipated as the mechanism for the seasonal variation, it causes weakening of local immunity thereby promoting infection by these pathogens[22].

This study is limited by its inability to perform molecular detection of BRO 1 and 2 for identification of β -lactamase enzymes. But it may be anticipated that majority were BRO 1 β -lactamase enzyme as they had high MICs for ampicillin[23].

In summary, result of this study supports the recommendation of BSAC in 2011 that clinical laboratories are no more required to check the susceptibility of ampicillin against *M. catarrhalis* and should report them resistant[24]. Therefore appropriate alternative antibiotic should be

selected empirically in regard of treating CARTI. Finally we emphasize the judicious use of antibiotics in this region and continued surveillance to detect emerging resistance.

Conflict of interest statement

We declare that we have no conflict of interest.

Acknowledgement

This research project is supported by the departmental seed money grant of Aga Khan University hospital, Karachi, Pakistan (Grant No. SM2011–04).

References

- [1] Verdulin CM, Hol C, Fleer A, Dijk HV, Belkum AV. *Moraxella catarrhalis*: from emerging to established pathogen. *Clin Microbiol Rev* 2002; **15**(1): 125–144.
- [2] Murphy TF, Parameshwaran GI. *Moraxella catarrhalis*, a human respiratory tract pathogen. *Clin Infect Dis* 2009; **49**(1): 124–131.
- [3] Sano N, Matsunaga S, Akiyama T, Nakashima Y, Kusaba K, Nagasawa Z, et al. *Moraxella catarrhalis* bacteraemia associated with prosthetic vascular graft infection. *J Med Microbiol* 2010; **59**(Pt 2): 245–250.
- [4] Sy MC, Robinson JL. Community-acquired *Moraxella catarrhalis* pneumonia in previously healthy children. *Pediatr Pulmonol* 2010; **45**: 674–678.
- [5] Saafan AE. Inefficacy of β -lactams in treatment of otitis media caused by *Streptococcus pneumoniae* or *Haemophilus influenzae* associated with *Moraxella catarrhalis*: an *in vitro* study. *Egypt J Med Microbiol* 2007; **16**(3): 407–415.
- [6] Malmvall BE, Brorsson JE, Johnsson J. *In vitro* sensitivity to penicillin V and beta-lactamase production of *Branhamella catarrhalis*. *J Antimicrob Chemother* 1977; **3**(4): 374–375.
- [7] Wang H, Chen M, Xu Y, Sun H, Yang Q, Hu Y, et al. Antimicrobial susceptibility of bacterial pathogens associated with community-acquired respiratory tract infections in Asia: report from the community-acquired respiratory tract infection pathogen surveillance (CARTIPS) study, 2009–2010. *Int J Antimicrob Agents* 2011; **38**(5): 376–383.
- [8] Morrissey I, Maher K, Williams L, Shackcloth J, Felmingham D, Reynolds R. Nonsusceptibility trends among *Haemophilus influenzae* and *Moraxella catarrhalis* from community-acquired respiratory tract infections in the UK and Ireland, 1999–2007. *J Antimicrob Chemother* 2008; **62**(Suppl 2): ii97–ii103.
- [9] Beekmann SE, Heilmann KP, Richter SS, Garcia-de-Lomas J, Doern GV. Antimicrobial resistance in *Streptococcus pneumoniae*, *Haemophilus influenzae*, *Moraxella catarrhalis* and group A beta-haemolytic streptococci in 2002–2003. Results of the multinational GRASP Surveillance Program. *Int J Antimicrob Agents* 2005; **25**: 148–156.
- [10] Flamm RK, Sader HS, Farrell DJ, Jones RN. Macrolide and tetracycline resistance among *Moraxella catarrhalis* isolates from 2009 to 2011. *Diagn Microbiol Infect Dis* 2012; **74**(2): 198–200.
- [11] DiPersio JR, Jones RN, Barrett T, Doern GV, Pfaller MA. Fluoroquinolone-resistant *Moraxella catarrhalis* in a patient with pneumonia: report from the SENTRY Antimicrobial Surveillance Program (1998). *Diagn Microbiol Infect Dis* 1998; **32**: 131–135.
- [12] Tabassum N, Ahmed A. Determination of anti-microbial susceptibilities of *Haemophilus influenzae*, *Streptococcus pneumoniae* and *Moraxella catarrhalis*. *J Pak Med Assoc* 2002; **52**(2): 87–90.
- [13] Abbasi SA, Karamat KA, Hanan A. Beta-lactamase production in isolates of *Moraxella catarrhalis*. *J Pak Inst Med Sci* 1997; **7**(2): 526–529.
- [14] Reynolds R, Hope R, Williams L. Survey, laboratory and statistical methods for the BSAC Resistance Surveillance Programmes. *J Antimicrob Chemother* 2008; **62**(Suppl 2): ii15–ii28.
- [15] Andrews JM. BSAC standardized disc susceptibility testing method (version 8). *J Antimicrob Chemother* 2009; **64**(3): 454–489.
- [16] Larsson C, Kanungo R, Kahlmeter G, Rao RS, Krantz I, Norrby SR, et al. High frequency of multiresistant respiratory tract pathogens at community level in South India. *Clin Microbiol Infect* 1999; **5**: 740–747.
- [17] Anita KB, Faseela TS, Fernandez N, Malli CS, Mallya S. The prevalence of *Moraxella catarrhalis* in lower respiratory tract infections. *J Clin Diagn Res* 2011; **5**(2): 240–241.
- [18] Hsu SF, Lin YT, Chen TL, Siu LK, Hsueh PR, Huang ST, et al. Antimicrobial resistance of *Moraxella catarrhalis* isolates in Taiwan. *J Microbiol Immunol Infect* 2012; **45**(2): 134–140.
- [19] Ramana BV, Chaudhury A. Antibiotic sensitivity pattern of *Moraxella catarrhalis* at a tertiary care hospital. *Int J Pharm Life Sci* 2012; **3**(7): 1805–1806.
- [20] Tamang MD, Dey S, Makaju RK, Jha BK, Shivananda PG, Bhramadatan KN. Prevalence of *Moraxella catarrhalis* infections of the lower respiratory tract in elderly patients. *Kathmandu Univ Med J* 2005; **3**(1): 39–44.
- [21] Anita KB, Faseela TS, Rai YK, Malli CS, Mallya S. *Moraxella catarrhalis*: an often overlooked pathogen of the respiratory. *J Clin Diagn Res* 2011; **5**(3): 495–497.
- [22] Binks MJ, Cheng AC, Smith-Vaughan H, Sloots T, Nissen M, Whitley D, et al. Viral-bacterial co-infection in Australian indigenous children with acute otitis media. *BMC Infect Dis* 2011; **11**: 161.
- [23] Esel D, Ay-Altintop Y, Yagmur G, Gokahmetoglu S, Sumerkan B. Evaluation of susceptibility patterns and BRO β -lactamase types among clinical isolates of *Moraxella catarrhalis*. *Clin Microbiol Infect* 2007; **13**(10): 1023–1025.
- [24] Andrews JM, Howe RA. BSAC standardized disc susceptibility testing method (version 10). *J Antimicrob Chemother* 2011; **66**(12): 2726–2757.