



THE AGA KHAN UNIVERSITY

eCommons@AKU

Department of Pathology and Laboratory Medicine

Medical College, Pakistan

January 2016

Naegleria fowleri meningoencephalitis associated with public water supply, Pakistan, 2014

Najia Ghanchi Aga Khan University, najia.ghanchi@aku.edu

Erum Khan Aga Khan University, erum.khan@aku.edu

Azam Khan Karachi Water and Sewerage Board

Wali Muhammad World Health Organization

Faisal Riaz Malik Aga Khan University

See next page for additional authors

Follow this and additional works at: http://ecommons.aku.edu/ pakistan_fhs_mc_pathol_microbiol Part of the Pathology Commons

Recommended Citation

Ghanchi, N., Khan, E., Khan, A., Muhammad, W., Malik, F., Zafar, A. (2016). Naegleria fowleri meningoencephalitis associated with public water supply, Pakistan, 2014. *Emerging Infectious Diseases*, 22(10), 1835-1837. **Available at:** http://ecommons.aku.edu/pakistan_fhs_mc_pathol_microbiol/596

Authors

Najia Ghanchi, Erum Khan, Azam Khan, Wali Muhammad, Faisal Riaz Malik, and Afia Zafar

of *Burkholderia mallei*, essential for the bacterium's survival, proliferation, and evasion of host adaptive immune responses (10). Our findings suggest that an approach combining culture, 16S rDNA sequencing, and multilocus sequence typing be considered for the accurate identification of uncommon bacterial infection.

This study was supported in part by a High Impact Research– Ministry of Higher Education Grant (no. E000013-20001), the Naval Medical Research Center–Asia, and the US Department of State, Biosecurity Engagement Program (NAMRU: J-55025-75053).

References

- Coenye T, Vandamme P, Govan JRW, Lipuma JJ. Taxonomy and identification of the *Burkholderia cepacia* complex. J Clin Microbiol. 2001;39:3427–36. http://dx.doi.org/10.1128/ JCM.39.10.3427-3436.2001
- Coenye T, Laevens S, Willems A, Ohlén M, Hannant W, Govan JRW, et al. *Burkholderia fungorum* sp. nov. and *Burkholderia caledonica* sp. nov., two new species isolated from the environment, animals and human clinical samples. Int J Syst Evol Microbiol. 2001;51:1099–107. http://dx.doi.org/10.1099/00207713-51-3-1099
- Gerrits GP, Klaassen C, Coenye T, Vandamme P, Meis JF. Burkholderia fungorum septicemia. Emerg Infect Dis. 2005;11:1115–7. http://dx.doi.org/10.3201/eid1107.041290
- Zhang R, Ran Y, Dai Y, Yang H, Zhang H, Lu Y, et al. Infectious granuloma caused by *Burkholderia fungorum* confirmed by laser-capture microdissection and polymerase chain reaction. Br J Dermatol. 2014;171:1261–3. http://dx.doi.org/10.1111/bjd.13094
- Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing; twenty-fourth informational supplement (M100–S24). Wayne (PA): The Institute; 2014.
- Spilker T, Baldwin A, Bumford A, Dowson CG, Mahenthiralingam E, LiPuma JJ. Expanded multilocus sequence typing for *Burkholderia* species. J Clin Microbiol. 2009;47:2607– 10. http://dx.doi.org/10.1128/JCM.00770-09
- Sam IC, Karunakaran R, Kamarulzaman A, Ponnampalavanar S, Syed Omar SF, Ng KP, et al. A large exposure to *Brucella melitensis* in a diagnostic laboratory. J Hosp Infect. 2012;80:321–5. http://dx.doi.org/10.1016/j.jhin.2011.12.004
- Axford JS. Joint and bone infections. Medicine. 2010;38:194–201. http://dx.doi.org/10.1016/j.mpmed.2009.12.006
- Laranjo M, Alexandre A, Oliveira S. Legume growth-promoting rhizobia: an overview on the *Mesorhizobium* genus. Microbiol Res. 2014;169:2–17. http://dx.doi.org/10.1016/j.micres.2013.09.012
- Nierman WC, DeShazer D, Kim HS, Tettelin H, Nelson KE, Feldblyum T, et al. Structural flexibility in the *Burkholderia mallei* genome. Proc Natl Acad Sci U S A. 2004;101:14246–51. http://dx.doi.org/10.1073/pnas.0403306101

Address for correspondence: Sazaly AbuBakar, Tropical Infectious Diseases Research & Education Centre, Block N & O, Faculty of Medicine, University of Malaya, 50603 Kuala Lumpur, Malaysia; email: sazaly@um.edu.my

Naegleria fowleri Meningoencephalitis Associated with Public Water Supply, Pakistan, 2014

Najia K. Ghanchi, Erum Khan, Azam Khan, Wali Muhammad, Faisal Riaz Malik, Afia Zafar

Author affiliations: Aga Khan University, Karachi, Pakistan (N.K. Ghanchi, E. Khan, F.R. Malik, A. Zafar); Karachi Water and Sewerage Board, Karachi (A. Khan); World Health Organization, Karachi (W. Muhammad)

DOI: http://dx.doi.org/10.3201/eid2210.151236

To the Editor: *Naegleria fowleri*, a free-living ameba, causes acute, fulminant, fatal primary amebic meningoencephalitis (PAM) in persons with history of recreational activities in warm freshwater (1,2). During 2008–2009, thirteen case-patients with PAM and no history of recreational water activity were reported from Karachi, Pakistan (3). Since then, PAM caused by domestic water exposure, nasal cleansing by using neti pots, and ablution has been reported globally (4–6). During 2014–2015, the Aga Khan University Hospital clinical laboratory in Karachi confirmed 19 PAM case-patients without history of recreational activities in warm freshwater.

Karachi has a subtropical, arid climate and long summers (March–October). The increasing number of PAM cases might be attributable to rising environmental temperatures and a dysfunctional water supply system in Karachi (7). Data indicating direct evidence of *N. fowleri* amebae in Karachi's water supply are limited, but consistent annual reemergence of PAM in patients without history of recreational water exposure raises concerns about Karachi's water supply.

In August 2014, a previously healthy 34-year-old man living in Karachi and having no recreational water exposure was admitted to the Aga Khan University Hospital with multiple episodes of vomiting, severe headache, and fever. Cerebrospinal fluid culture showed a low glucose level (46 mg/dL [reference 45–80 mg/dL]) and high levels of protein (216 mg/dL [reference 20–40 mg/dL]), erythrocytes (30 cells/mm³ [reference 0–10 cells/mm³]), and leukocytes (1,440 cells/mm³ [reference 0–5 cells/mm³]; 65% lymphocytes and 35% neutrophils). PCR confirmed presence of *N. fowleri*. The patient died 4 days after admission, and cerebrospinal fluid and blood cultures were negative for bacterial and fungal growth.

We investigated for presence of N. fowleri amebae in domestic water and for the patient's possible exposure. In September 2014, we collected 23 samples from 2 water treatment plants (plants A and B), their pumping

LETTERS

stations, and catchment areas (online Technical Appendix Figure, http://wwwnc.cdc.gov/EID/article/22/10/15-1236-Techapp1.pdf). Plant A supplied 13 of the samples from its water distribution system, which provided water to the patient's residence and neighborhood mosque (Table). The other 10 samples were from Plant B and its water distribution system (online Technical Appendix Table), for which the local government had initiated additional chlorine enhancement because of previously occurring PAM cases. Plant A had no chlorine enhancement. Real-time PCR, as described (8), confirmed presence of N. fowleri amebae in the plant's water supply distribution. Both plants are monitored for quality control by using World Health Organization water treatment procedures guidelines (http://apps.who.int/iris/ bitstream/10665/44584/1/9789241548151 eng.pdf). The plants' water distribution exit points had the highest residual chlorine levels (0.5 ppm) (9), and levels gradually

decreased beyond the plants. Residual chlorine was undetectable in plant A's water distribution to the patient's residential area (12 km from the plant); however, with plant B's additional chlorine enhancement stations, chlorine levels were detectable in all households tested.

Residual chlorine and specimen positivity for freeliving amebae were inversely correlated. Differences were most noticeable in samples collected from plant A's distributed water, compared with plant B's water distribution samples. PCR confirmed *N. fowleri* amebae in 2 water samples collected from the patient's household overhead storage tanks and neighborhood mosque. The samples, taken from plant A's distributed water, showed no residual chlorine and a temperature >30°C. Lack of detectable chlorine and water temperature >25°C might have provided favorable conditions for *N. fowleri* amebae to thrive in domestic water (5); water temperatures 25° -40°C are favorable for *N. fowleri* growth. Absence of

Table. Characteristics of 13 water samples collected from water treatment plant A and its distribution system, including water supplied
to the apartment and neighborhood mosque of a patient with primary amebic meningitis, Karachi, Pakistan, 2014*

			Total		Culture	PCR results for	Distance
	Sample		chlorine,	Temperature,	positivity	Naegleria fowleri	relationships of water
Water supply	location	Sample type	mg/L	°C†	for FLAs	amebae‡	samples
Reservoir§	Water from	Untreated	ND	30	++	-	From reservoir to
	Kinjhar						plant A, ≈100 km
Water treatment plant A	Filtration unit¶	Treatment underway	<0.5	29.5	++	-	
	Plant A exit point	Filtered and chlorinated	0.5	30	++	-	
Pumping station#	Pumping station, site 1	Filtered and chlorinated	ND	30	++	_	From plant A to pumping station, ≈11 km
	Pumping station, site 2	Filtered and chlorinated	ND	30	++	-	
Catchment areas: patient's	Mainline	Filtered and chlorinated	ND	30	-	_	From pumping station to patient's house, 1km
*ELAs free-living an	Underground boring well water**	Untreated	ND	28.5	+	-	Within patient's house, ≈10 m
	Underground tank	Mixed ⁺⁺	ND	28	+	-	Within patient's house, ≈10 m
	Overhead tank	Mixed	ND	31	++	+	Within patient's house, ≈10 m
	In-house storage tank‡‡	Mixed	ND	30	++	-	Within patient's house, ≈5 m
	Bathroom	Mixed	ND	29	+	-	Within patient's house, ≈5 m
	Neighborhood mosque	Filtered and chlorinated	ND	31	++	+	From patient's house to mosque, ≈100 m

*FLAs, free-living amebae; ND, not detected; ++, >3 amebae seen with 40x magnification; +, 1–3 amebae seen with 40 x magnification; –, no amebae detected.

†Water temperatures of 25°C-40°C are conductive for flourishing of Naegleria fowleri amebae.

‡PCR was negative for other pathogenic FLAs such as *Balamuthia* or *Acanthamoeba* spp.

Skinjhar Lake, located in Sindh province, Pakistan, is the main reservoir that supplies water to Karachi.

The filtration unit was the only site for which 2 samples were taken; only 1 sample was taken from all other specific sample locations.

#Samples were taken from a single pumping station at 2 separate sites.

**Underground wells provide additional water supplies locally known as boring wells.

††Water from these wells is not treated and is mixed with treated water from the main water supply in storage tanks.

##Underground and overhead tanks are shared by >100 households, so for continuous water supply, residents keep small tanks in their homes.

the amebae in plant B's water suggests the importance of enhanced chlorine pumping at distribution points beyond water treatment plants for maintain residual chlorine in Karachi's domestic water supply.

Because water supply can be intermittent, underground and overhead storage tanks are essential for Karachi homes. To ensure continuous domestic supply, water is stored in overhead tanks and pumped from tanks into homes as needed. Water storage in tanks perhaps facilitated propagation of *N. fowleri* amebae in domestic and mosque water. During the summer, ambient temperatures reach 44°C, leading to increased water temperatures in overhead tanks. We found water temperatures up to 34°C, which may facilitate excystation of *N. fowleri* amebae to infective forms. Slime, dirt, and high ambient temperatures likely explain *N. fowleri* multiplication in storage tanks, the possible source of infection for this patient in Karachi.

Presence of *N. fowleri* amebae in mosque water is alarming. Ablution (Wudhu) is a ritual performed by Muslims before offering prayers and involves thorough cleaning of mouth, ears, face, arms, feet, and nasal passages, the latter by inhaling water forcefully up the nostrils. Performing this activity with contaminated water could be a communal source for potential outbreaks.

Karachi water supply authorities have initiated chlorine enhancement at various sites beyond plant B, and our findings support the need for this enhancement. We recommend that the government implement measures to maintain appropriate chlorine levels in the domestic water supply and at recreational sites and to develop effective amebaemonitoring programs. The public should use boiled or filtered water for nasal cleansing, regularly clean storage tanks, and add supplemental chlorine to water in homes, especially during the summer.

Acknowledgments

We thank the Karachi Water and Sewerage Board and residents of Karachi for providing water samples.

References

- Capewell LG, Harris AM, Yoder JS, Cope JR, Eddy BA, Roy SL, et al. Diagnosis, clinical course, and treatment of primary amoebic meningoencephalitis in the United States, 1937–2013. J Pediatric Infect Dis Soc. 2015;4:e68–75. http://dx.doi.org/10.1093/jpids/ piu103
- Yoder JS, Eddy BA, Visvesvara GS, Capewell L, Beach MJ. The epidemiology of primary amoebic meningoencephalitis in the USA, 1962–2008. Epidemiol Infect. 2010;138:968–75. http://dx.doi.org/10.1017/S0950268809991014
- Shakoor S, Beg MA, Mahmood SF, Bandea R, Sriram R, Noman F, et al. Primary amebic meningoencephalitis caused by *Naegleria fowleri*, Karachi, Pakistan. Emerg Infect Dis. 2011;17:258–61. http://dx.doi.org/10.3201/ eid1702.100442

- Yoder JS, Straif-Bourgeois S, Roy SL, Moore TA, Visvesvara GS, Ratard RC, et al. Primary amebic meningoencephalitis deaths associated with sinus irrigation using contaminated tap water. Clin Infect Dis. 2012;55:e79–85. http://dx.doi.org/10.1093/cid/cis626
- Cope JR, Ratard RC, Hill VR, Sokol T, Causey JJ, Yoder JS, et al. The first association of a primary amebic meningoencephalitis death with culturable *Naegleria fowleri* in tap water from a US treated public drinking water system. Clin Infect Dis. 2015; 60:e36–42. http://dx.doi.org/10.1093/cid/civ017
- Primary amebic meningoencephalitis associated with ritual nasal rinsing—St Thomas, US Virgin Islands, 2012. Clin Infect Dis. 2014;58:ii.
- Sajjad SH, Hussain B, Khan MA, Raza A, Zaman B, Ahmed I. On rising temperature trends of Karachi in Pakistan. Clim Change. 2009;96:539–47. http://dx.doi.org/10.1007/s10584-009-9598-y
- Qvarnstrom Y, Visvesvara GS, Sriram R, da Silva AJ. Multiplex real-time PCR assay for simultaneous detection of *Acanthamoeba* spp., *Balamuthia mandrillaris*, and *Naegleria fowleri*. J Clin Microbiol. 2006;44:3589–95. http://dx.doi.org/10.1128/ JCM.00875-06
- Reed B, Shaw R, Chatterton K. Technical notes on drinking-water, sanitation and hygiene in emergencies. Loughborough (UK): World Health Organization, Water, Engineering and Development Centre; 2013.

Address for correspondence: Afia Zafar, Section of Microbiology, Department of Pathology and Laboratory Medicine, Aga Khan University, Stadium Rd, PO Box 3500, Karachi 74800, Pakistan; email: afia.zafar@aku.edu

Unmet Needs for a Rapid Diagnosis of Chikungunya Virus Infection

Elisa Burdino, Guido Calleri, Pietro Caramello, Valeria Ghisetti

Author affiliation: Amedeo di Savoia Hospital, Torino, Italy

DOI: http://dx.doi.org/10.3201/eid2210.151784

To the Editor: Chikungunya virus (CHIKV) has become a global health problem. Clinical manifestations are not specific and are difficult to differentiate from those of similar viral diseases (e.g., dengue and Zika virus disease). Diagnostic laboratories must be prepared to meet the changing epidemiology of viral diseases. CHIKV infection is currently identified by viral genome detection, using reverse transcription PCR (RT-PCR), viral culture, and serologic testing for IgG and IgM by indirect immunofluorescence (IFA) or ELISA. RT-PCR is most sensitive during the early phase of CHIKV infection (within 5–7 days of symptom onset), but its use is limited by the short viremic phase of the disease. After the acute phase, serologic testing for IgG and IgM is a more accurate indicator of disease.