



A critical appraisal of phloem-mobile signals involved in tuber induction

Paula Suárez-López*

Molecular Genetics Department, Centre for Research in Agricultural Genomics, CSIC - IRTA - UAB - UB, Barcelona, Spain

Edited by:

Jurriaan Ton, University of Sheffield, UK

Reviewed by:

Chiou, Tzyy-Jen, Academia Sinica, Taiwan

Jurriaan Ton, University of Sheffield, UK

Julia Kehr, University Hamburg, Germany

David Hannapel, Iowa State University, USA

Tien-Shin Yu, Institute of Plant and Microbial Biology, Academia Sinica, Taiwan

*Correspondence:

Paula Suárez-López, Molecular Genetics Department, Centre for Research in Agricultural Genomics, CSIC - IRTA - UAB - UB, Campus UAB, Bellaterra (Cerdanyola del Vallès), 08193 Barcelona, Spain
e-mail: paula.suarez@cragenomica.es

The identification of FLOWERING LOCUS T (FT) and several FT homologs as phloem-mobile proteins that regulate flowering has sparked the search for additional homologs involved in the long-distance regulation of other developmental processes. Given that flowering and tuber induction share regulatory pathways, the quest for long-distance tuberization signals has been further stimulated. Several tuberization regulators have been proposed as mobile molecules, including the FT family protein StSP6A, the plant growth regulators gibberellins and the microRNA miR172. Although some of these hypotheses are attractive and plausible, evidence that these molecules are transmissible in potato has yet to be obtained. Two mRNAs encoding transcription factors, StBEL5 and POTATO HOMEBOX 1 (POTH1), are mobile and correlate with tuber induction. However, evidence that StBEL5 or POTH1 are required for tuberization is not available yet. Therefore, there are several good candidates for long-distance molecules in the tuberization process. Further research should test their role as systemic tuberization signals.

Keywords: tuberization, potato, long-distance signaling, FLOWERING LOCUS T, gibberellins

The induction of tuber formation is a key developmental transition for the production of potatoes, one of the most important food crops. Understanding the regulation of tuber induction is essential to devise strategies to improve tuber yield and quality. During the last two decades we have started to comprehend this regulation, with the identification of genes that control tuberization (Jackson, 1999; Abelenda et al., 2011). This has been facilitated by the tremendous progress in understanding the control of flowering, which is similar to tuberization in aspects such as the response to photoperiod and the involvement of phloem-mobile signals (Suárez-López, 2005; Abelenda et al., 2011). This Perspective paper focuses on recent findings that suggest several molecules as candidates for systemic signals controlling tuber induction.

LONG-DISTANCE SIGNALS REGULATE TUBERIZATION AND FLOWERING

Short day (SD) photoperiods promote tuberization, whereas long days (LDs), high nitrogen levels and high temperatures inhibit or delay tuberization. Within the tuberization process, it is important to distinguish between tuber induction and tuber development and growth. Induction takes place when signals are produced in leaves and transported through the phloem to underground stems (stolons), or when mobile signals that inhibit tuberization are repressed (Jackson, 1999; Suárez-López, 2005). This leads to the initiation of tuber development and growth, which determines tuber shape, number, and weight. Although tuber yield is often used to assess tuber induction,

changes in tuber yield can result from alterations in many different factors, including overall plant growth, photoassimilate partitioning, the strength of induction, tuber development, etc. (Ewing and Struik, 1992). The time of tuber initiation is therefore a much better indicator of tuber induction than tuber yield.

Grafting experiments using potato plants induced and non-induced to tuberize demonstrated the existence of transmissible substances decades ago (Gregory, 1956; Chapman, 1958), but the identification of these signals has proven difficult. Recent advances in the study of other developmental processes provide hints for finding long-distance tuberization signals. The intensive search for a phloem-mobile flowering signal, called florigen, has led to the identification of several FLOWERING LOCUS T (FT) family members as leaf-produced proteins that travel to the shoot apical meristem, where they induce flowering (Turck et al., 2008; Tsuji et al., 2013). In *Arabidopsis thaliana*, FT expression is activated by the transcriptional regulator CONSTANS (CO) in leaf phloem cells in response to floral inductive photoperiods (An et al., 2004; Ayre and Turgeon, 2004).

However, florigen is not a single molecule. Positive and negative transmissible regulators of flowering exist (Bernier, 1988; Matsoukas et al., 2012). Several FT family members can perform these functions. In rice, Heading date 3a (Hd3a) and RICE FLOWERING LOCUS T 1 (RFT1) act as florigenic signals under different photoperiods (Tamaki et al., 2007; Komiya et al., 2009). In *Arabidopsis* ATC acts as a mobile repressor or antiflorigen and TWIN SISTER OF FT (TSF) might function as a florigen

(Yamaguchi et al., 2005; Mathieu et al., 2007; Jang et al., 2009; D'Aloia et al., 2011; Huang et al., 2012). In addition, several FT-related proteins have been detected in phloem exudates of diverse species (Giavalisco et al., 2006; Lin et al., 2007; Aki et al., 2008).

Many RNAs are present in phloem exudates and a few have been reported to act in long-distance signaling (Sasaki et al., 1998; Ruiz-Medrano et al., 1999; Kim et al., 2001; Doering-Saad et al., 2002; Haywood et al., 2005). Movement of *FT* and *ATC* RNAs has been shown, but the *FT* RNA accelerates flowering less effectively than the protein and the RNA together (Li et al., 2009, 2011; Huang et al., 2012; Lu et al., 2012). Other reports indicate that translocation of the FT protein, but not the RNA, is required to promote flowering (Lifschitz et al., 2006; Mathieu et al., 2007; Notaguchi et al., 2008). These findings suggest that movement of the *FT* mRNA can help to induce flowering, but movement of the FT protein is much more crucial. In addition to FT proteins and RNA, other types of molecules, such as hormones and metabolites, have been postulated as long-distance floral signals (Turnbull, 2011; Dinant and Suárez-López, 2012).

IS FT A PHLOEM-MOBILE TUBERIZATION SIGNAL?

Transmissible signals for flowering and tuberization are interchangeable. Tobacco scions induced to flower promote tuberization when grafted onto potato stocks kept under non-tuber-inducing conditions (Chailakhyan et al., 1981). When a rice *Hd3a*-GFP fusion is expressed in potato phloem, it can move across a graft junction to stolons and induce tuber formation (Navarro et al., 2011), suggesting that a similar protein exists in potato. Indeed, several *FT*-like genes have been identified in this species. One of them encodes *StSP3D*, which mainly affects flowering, and another encodes *StSP6A*, which induces tuber formation, a role similar to that of *FT* in flowering control (Navarro et al., 2011). The effect of *StSP6A* on tuberization is transmitted through grafts (Navarro et al., 2011). Altogether, these findings strongly suggest that *StSP6A* is probably a mobile tuberization signal.

There are similarities, but also differences, in the regulation of FT genes. *StSP6A* is negatively regulated by *StCO* (Figure 1), a potato CO-like protein that represses tuberization under non-inductive LDs (Navarro et al., 2011; González-Schain et al., 2012). *StCO* does not seem to play a role under SDs (González-Schain et al., 2012). By contrast, *Arabidopsis* CO promotes *FT* transcription only under inductive photoperiods (Turck et al., 2008). In rice *Hd3a* is repressed or activated by the CO-like protein *Hd1* under non-inductive or inductive conditions, respectively, and in addition *RFT1* is up-regulated and promotes flowering much later under non-inductive conditions (Tsuji et al., 2013). These differences stress the need to test hypotheses based on flowering-time models, rather than simply extrapolating them to tuberization. Demonstrations that *StSP6A* moves are therefore eagerly awaited.

Two additional FT family members from potato, *StTFL1* and *StSP5G*, might be related to the tuberization process. *StTFL1* mRNA levels are high in stolons before induction and decrease at early stages of tuber development. Overexpression of *StTFL1* causes an increase in the number of tubers produced (Guo et al., 2010), suggesting a role in tuber induction or development. The

expression pattern of *StSP5G* suggests that this gene might play an opposite role to that of *StSP6A* in tuberization control (Navarro et al., 2011; Kloosterman et al., 2013), although a functional analysis of this gene has not been reported so far. Further analyses of *StTFL1* and *StSP5G* to determine their biological functions should be pursued, given that FT-related proteins affect other developmental processes aside from flowering and tuberization (Pin and Nilsson, 2012; Hiraoka et al., 2013). As many FT-like proteins are mobile, it would be worth testing *StTFL1* and *StSP5G* movement.

StBEL5 AND *POTH1* mRNAs AS PUTATIVE TRANSMISSIBLE SIGNALS

Two mRNAs have been proposed as long-distance signals regulating tuberization. *StBEL5* and POTATO HOMEODOMAIN 1 (*POTH1*) are homeobox transcription factors that interact with each other (Chen et al., 2003). Overexpression of *POTH1* increases the number of tubers produced relative to wild-type (WT) plants in *in vitro* tuberization assays (Rosin et al., 2003). Overexpression of *StBEL5* enhances tuber formation under SDs and promotes tuberization under non-inductive LDs. *StBEL5* mRNA moves from overexpressing scions to WT stocks and movement correlates with increased tuber yield (Chen et al., 2003; Banerjee et al., 2006). Graft transmission of *POTH1* mRNA has also been shown (Mahajan et al., 2012). Transcription of *StBEL5* and *POTH1* in vascular cells (Banerjee et al., 2006; Mahajan et al., 2012) is consistent with movement of their transcripts through the phloem. Additional experimental approaches support translocation of *StBEL5* mRNA and have been previously reviewed (Hannapel, 2010).

However, there are numerous caveats to be aware of when interpreting the movement of *StBEL5* and *POTH1* RNAs, as well as their effects on tuberization. First, *POTH1* has not been shown to affect tuber formation in soil-grown plants. Second, whether *StBEL5* and/or *POTH1* are required for tuber induction in WT plants has not been demonstrated, as only overexpression alters tuber induction or development. Third, RNA movement has been shown from overexpressing plants, but not from WT plants (Banerjee et al., 2006; Mahajan et al., 2012), and it has not been tested whether movement is required for tuberization. Fourth, *POTH1*-overexpressing plants exhibit dramatic alterations in the vasculature (Rosin et al., 2003; Mahajan et al., 2012). It is possible that the tuber phenotype of *POTH1*-overexpressing plants and graft transmission of *POTH1* mRNA are indirect consequences of these alterations. Fifth, both *POTH1* and *StBEL5* are transcribed in stolons, with an increase in *StBEL5* transcription at early stages of tuber formation (Banerjee et al., 2006; Mahajan et al., 2012), casting doubts on the need of movement from leaves. Finally, it has not been excluded that movement of *StBEL5* and/or *POTH1* proteins may occur.

Therefore, although *StBEL5* and *POTH1* RNAs are able to move, further research is needed to demonstrate whether this has any biological relevance. This can be addressed by simultaneously silencing *StBEL5* and *POTH1* or several *StBEL* paralogs, which have been proposed to act redundantly (Chen et al., 2003). Whether the *StBEL5* protein moves should also be tested.

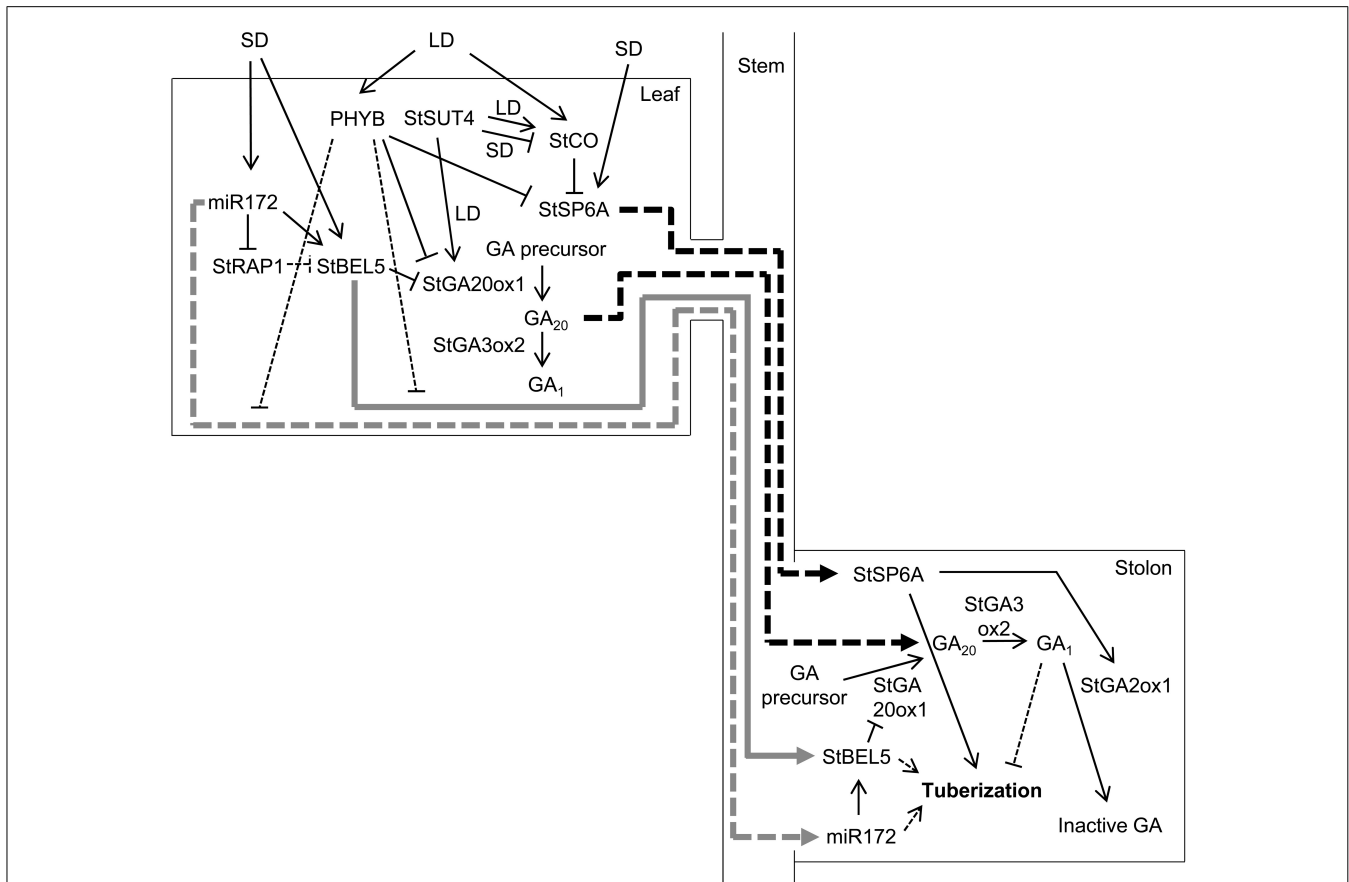


FIGURE 1 | Model for the regulation of tuber induction by phloem-mobile signals. The main candidates for mobile signals are the StSP6A protein, two RNAs – *StBEL5* and miR172 – and GAs. The production, and possibly the movement, of these four factors is regulated by a complex genetic network. PHYB, StSUT4, and StCO repress tuberization in response to LDs. GAs also seem to act as repressors, whereas StSP6A and perhaps miR172 and StBEL5 act as tuberization promoters under inductive SD conditions. Under LDs, PHYB represses the expression of *StSP6A* and *StGA20ox1*, which encodes an enzyme that catalyzes the synthesis of GA₂₀. PHYB up-regulates miR172 and StBEL5 in leaves and down-regulates them in stolons, which might result from a repression of *StBEL5* mRNA and miR172 movement from leaves to stolons. Under LDs, StSUT4 induces *StGA20ox1* and *StCO*, which represses *StSP6A*. Under SDs, StSUT4 inhibits *StCO*, relieving *StSP6A* repression. In addition to *StSP6A*, SDs up-regulate miR172 and *StBEL5*. miR172 induces *StBEL5*, probably through the repression of

miR172 target genes, such as *StRAP1*, which would act as *StBEL5* inhibitors. StBEL5 represses *StGA20ox1* in a complex with POTH1. StGA3ox2 catalyzes the conversion of GA₂₀ to GA₁, an active GA. *StSP6A*, *StBEL5* mRNA, miR172, and GAs presumably translocate to stolons through the phloem. In the stolons, *StSP6A* promotes tuber development, at least in part through up-regulation of *StGA20ox1*, which converts active GAs into inactive forms. miR172 up-regulates *StBEL5*, which together with POTH1 down-regulates *StGA20ox1*, reducing the synthesis of active GAs, which repress tuber development. Under LDs, GA₂₀ would move from leaves to stolons and would be converted to GA₁, thus repressing tuber development. Under SDs, there would be less GA₂₀ available and tuber development can occur. Thick gray arrows indicate RNA movement, and thick black arrows indicate protein or GA movement. Discontinuous lines indicate that movement or regulation has been suggested, but not demonstrated.

miR172 AFFECTS TUBERIZATION IN A GRAFT-TRANSMISSIBLE MANNER

To date, miR172, which regulates flowering in several species, is the only microRNA (miRNA) shown to affect tuber induction (Martin et al., 2009; Zhu and Helliwell, 2011). The effect of miR172 in potato has been reported in overexpressing plants, which form tubers under LDs, tuberize early under SDs and show up-regulation of *StBEL5*. Inactivation would help to confirm if miR172 is required for tuberization control.

There is growing evidence that small RNAs, including short interfering RNAs (siRNAs) and miRNAs, move cell-to-cell and systemically (Himber et al., 2003; Yoo et al., 2004; Lin et al., 2008; Pant et al., 2008; Chitwood et al., 2009; Carlsbecker et al.,

2010; Dunoyer et al., 2010; Molnar et al., 2010). The effect of miR172 overexpression is graft transmissible, suggesting that this miRNA regulates long-distance signals that control tuberization or, alternatively, that miR172 itself is a mobile signal. In grafting experiments, miR172-overexpressing scions accelerated tuberization of WT stocks, but the reciprocal graft combination did not tuberize early. The simplest interpretation is that miR172 is required in aerial organs, rather than in stolons, to promote tuberization. However, increases of miR172 levels in stolons correlate with tuber induction, while changes in leaves do not (Martin et al., 2009). At least two hypotheses can explain this apparent contradiction: (1) overexpression of miR172 in stocks might not be sufficient to counteract tuber-inhibiting signals derived from WT

scions; and (2) factors required for miR172 processing might be present or active in leaves but not in stolons. Detection of miR172 in potato phloem cells and phloem exudates of several species, as well as graft transmission in *Nicotiana benthamiana*, is consistent with the notion of this miRNA being mobile (Buhtz et al., 2008, 2010; Martin et al., 2009; Kasai et al., 2010; Varkonyi-Gasic et al., 2010). In addition to its putative role as a systemic signal, it has been proposed that miR172 might participate in cell-to-cell communication (Abelenda et al., 2011; Marín-González and Suárez-López, 2012). Given the potential of miRNAs to act as transmissible signals, it will be worth studying whether miR172 moves.

ROLE OF GIBBERELLINS IN TUBERIZATION

The plant hormones gibberellins (GAs) are present in phloem sap and seem to act as florigenic molecules in some species (Eriksson et al., 2006; King et al., 2006, 2008). The last steps in the biosynthesis of active GAs are catalyzed by GA 20-oxidase (GA20ox) and GA 3-oxidase (GA3ox). Biologically active GAs, including GA₁, GA₃, and GA₄, are inactivated by GA 2-oxidase (GA2ox) enzymes (Hedden and Thomas, 2012).

Gibberellins are involved in the control of tuber induction or development. Different observations have led to the assumption that GAs inhibit tuberization under LDs. Tuberization would take place when GA levels decrease in response to SDs (Rodríguez-Falcón et al., 2006). This decrease seems necessary to arrest longitudinal stolon growth and allow stolon swelling (Jackson, 1999). But are high GA levels really required to repress tuber induction under LDs? Silencing of a potato GA20ox (StGA20ox1) and manipulation of the levels of a GA3ox (StGA3ox2) do not induce tuberization under LDs (Carrera et al., 2000; Bou-Torrent et al., 2011). In addition, a GA2ox, StGA2ox1, affects tuberization *in vitro*, but not in soil-grown plants (Kloosterman et al., 2007), leading to the conclusion that *StGA2ox1* is a tuber-identity gene rather than a regulator of tuber induction. Local up-regulation of *StGA2ox1* in stolons by StSP6A (Navarro et al., 2011) is consistent with this interpretation.

Moreover, the expression patterns of several GA biosynthetic enzymes and the phenotypes of plants with altered levels of these enzymes do not always fit with the hypothesis of GAs repressing tuberization. For example, although *StGA3ox2* is down-regulated at the initiation of tuber development, *StGA20ox1* and *StGA20ox3* are up-regulated (Kloosterman et al., 2007). Both StGA20ox1-silenced lines and plants overexpressing StGA3ox2 tuberize earlier than WT plants under SDs, despite showing opposite changes of GA₁ levels (Carrera et al., 2000; Bou-Torrent et al., 2011). As GA biosynthesis involves feedback and feedforward regulations (Hedden and Thomas, 2012), some of these contradictions can be explained through negative feedback regulation of *StGA20ox* genes by active GAs, but this still has to be demonstrated.

To explain some of these conflicting results, it has recently been proposed that GA₂₀ – the immediate precursor of GA₁ – would be mobile, whereas GA₁ would not. In StGA3ox2-overexpressing plants, increased conversion of GA₂₀ to GA₁ in aerial parts would reduce the amount of GA₂₀ transported to stolons, resulting in low levels of GA₁ in stolons and early tuberization (Bou-Torrent et al., 2011). This interesting hypothesis fits well with some observations.

However, as *StGA20ox* genes are expressed in stolons (Carrera et al., 1999), GA₂₀ is expected to be synthesized here. StGA3ox2-overexpressing plants would then have increased conversion of GA₂₀ to GA₁ also in stolons, which should repress tuberization. Localized silencing of StGA20ox1 and StGA3ox2 in leaves and stolons and grafting experiments using plants with altered levels of these enzymes would help to elucidate the role of GA₂₀ and GA₁. It will also be necessary to test GA₂₀ movement in potato plants and whether movement is required to prevent tuberization. More work is also needed to determine whether GAs play a role in tuber induction or they regulate tuber development by preventing stolons from being competent to respond to leaf-derived inductive signals. Nowadays it cannot be excluded that GAs perform both functions.

SUCROSE AND OTHER PUTATIVE LONG-RANGE SIGNALING MOLECULES

Sucrose is a metabolite, a source of energy and a signaling molecule and it has been proposed as a transmissible substance for tuberization and flowering (Sheen et al., 1999; Suárez-López, 2005; Ruan, 2012). Transcripts of sucrose transporters are phloem mobile in several species, including potato, which suggests a possible signaling role for these RNAs (Liesche et al., 2011). A potato sucrose transporter, StSUT4, is involved in flowering and tuberization control. Inhibition of StSUT4 induces tuberization under LDs. Graft transmission of this phenotype, together with an increase in sucrose export from leaves of StSUT4-silenced plants, suggest a role for StSUT4 in long-distance signaling at least in part via source to sink carbon flux (Chincinska et al., 2008). In addition, StSUT4 regulates the production of putative long-distance signals, such as StSP6A and probably GAs (Chincinska et al., 2008, 2013).

There is additional evidence of a link between sucrose and GAs during tuberization. *In vitro* treatment with high sucrose concentrations, which induces tuber formation, reduces endogenous GA₁ levels in stolons before tuber initiation (Xu et al., 1998). Exogenous GA treatment, conversely, up-regulates StSUT4 (Chincinska et al., 2008). Altogether these observations indicate a complex interplay between GAs and sucrose during tuber induction or development. Understanding the different roles that sucrose plays in tuber formation, as a starch precursor, energy source and signal, deserves further attention.

Other molecules, such as metabolites, hormones, and peptides have the potential to act as mobile signals, but their roles in tuberization are not yet clear (Jackson, 1999; Fernie and Willmitzer, 2001; Dinant and Suárez-López, 2012). Grafting of tomato mutants onto potato stocks has been proposed as a strategy to elucidate the role of hormones in long-distance signaling, although the results so far point to effects on assimilate distribution rather than on signaling pathways (Peres et al., 2005).

THE ROLE OF PHYTOCHROME B IN REGULATING MOBILE SIGNALS

The photoreceptor phytochrome B (PHYB) plays an interesting role in the control of tuber induction, as it affects several putative systemic tuberization molecules. Grafting experiments using PHYB-silenced plants, which tuberize under LDs, led to

the proposal that PHYB induces a mobile tuberization repressor (Jackson et al., 1998). However, these plants show increased levels of *StSP6A* mRNA and reduced levels of *StBEL5* mRNA and *miR172* in leaves, as well as increased levels of these three RNAs in stolons (Martin et al., 2009; Navarro et al., 2011), indicating that PHYB inhibits the expression and/or movement of tuber-inducing molecules. Probably positive and negative regulators of tuberization respond to light signals through the action of PHYB.

Several effects of PHYB on plant development are mediated by GAs (Lau and Deng, 2010). In potato, PHYB affects GA synthesis or signaling, as *StGA20ox1* mRNA abundance is increased in PHYB-silenced plants, which show several phenotypes characteristic of alterations in GA homeostasis (Jackson et al., 2000). In addition, *StSUT4* probably mediates some effects of PHYB on plant development (Chincinska et al., 2008). How PHYB regulates all these genes is not known. PHYB controls long-distance regulation of other processes, pointing to a general role of PHYB in systemic signaling (Griebel and Zeier, 2008; Suzuki et al., 2011).

CONCLUSION AND FUTURE CHALLENGES

We have recently witnessed substantial advances in our knowledge of potato tuber induction. Although the identity of mobile

tuberization molecules is yet unknown, they are probably similar to flowering signals. Several good candidates have been proposed (Figure 1). Further research should test whether they act as genuine systemic tuberization signals.

Long-distance communication involves the production of signals, but also requires phloem loading, transport and unloading, as well as the response of target tissues to the translocated signals. Once the chemical nature of the signals is established, dissecting all these processes will be easier. The availability of the potato genome sequence (Xu et al., 2011) will facilitate these tasks. Interspecific grafting and experimental approaches used in other species, such as analyses of phloem sap composition, visualization of reporters fused to putatively mobile proteins and strategies to disrupt intercellular signaling, can be employed to address these questions.

ACKNOWLEDGMENTS

I acknowledge funding by Spanish MINECO and MICINN (Grants BFU2012-33746 and CSD2007-00036 “Centre for Research in Agrigenomics”) and the Department of Innovation, Universities and Enterprise of Catalonia (2009-SGR-703 and Catalan Biotechnology Reference Network).

REFERENCES

- Abelenda, J. A., Navarro, C., and Prat, S. (2011). From the model to the crop: genes controlling tuber formation in potato. *Curr. Opin. Biotechnol.* 22, 287–292. doi: 10.1016/j.copbio.2010.11.013
- Aki, T., Shigyo, M., Nakano, R., Yoneyama, T., and Yanagisawa, S. (2008). Nano scale proteomics revealed the presence of regulatory proteins including three FT-Like proteins in phloem and xylem saps from rice. *Plant Cell Physiol.* 49, 767–790. doi: 10.1093/pcp/pcn049
- An, H., Roussot, C., Suárez-López, P., Corbesier, L., Vincent, C., Piñeiro, M., et al. (2004). CONSTANS acts in the phloem to regulate a systemic signal that induces photoperiodic flowering of *Arabidopsis*. *Development* 131, 3615–3626. doi: 10.1242/dev.01231
- Ayre, B. G., and Turgeon, R. (2004). Graft transmission of a floral stimulant derived from *CONSTANS*. *Plant Physiol.* 135, 2271–2278. doi: 10.1104/pp.104.040592
- Banerjee, A. K., Chatterjee, M., Yu, Y., Suh, S.-G., Miller, W. A., and Hannapel, D. J. (2006). Dynamics of a mobile RNA of potato involved in a long-distance signaling pathway. *Plant Cell* 18, 3443–3457. doi: 10.1105/tpc.106.042473
- Bernier, G. (1988). The control of floral evocation and morphogenesis. *Annu. Rev. Plant Physiol. Plant Mol. Biol.* 39, 175–219. doi: 10.1146/annurev.pp.39.060188.001135
- Bou-Torrent, J., Martínez-García, J. F., García-Martínez, J. L., and Prat, S. (2011). Gibberellin A1 metabolism contributes to the control of photoperiod-mediated tuberization in potato. *PLoS ONE* 6:e24458. doi: 10.1371/journal.pone.0024458
- Buhtz, A., Pieritz, J., Springer, F., and Kehr, J. (2010). Phloem small RNAs, nutrient stress responses, and systemic mobility. *BMC Plant Biol.* 10:64. doi: 10.1186/1471-2229-10-64
- Buhtz, A., Springer, F., Chappell, L., Baulcombe, D. C., and Kehr, J. (2008). Identification and characterization of small RNAs from the phloem of *Brassica napus*. *Plant J.* 53, 739–749. doi: 10.1111/j.1365-313X.2007.03368.x
- Carlsbecker, A., Lee, J.-Y., Roberts, C. J., Dettmer, J., Lehesranta, S., Zhou, J., et al. (2010). Cell signalling by microRNA165/6 directs gene dose-dependent root cell fate. *Nature* 465, 316–321. doi: 10.1038/nature08977
- Carrera, E., Bou, J., García-Martínez, J. L., and Prat, S. (2000). Changes in GA 20-oxidase gene expression strongly affect stem length, tuber induction and tuber yield of potato plants. *Plant J.* 22, 247–256. doi: 10.1046/j.1365-313x.2000.00736.x
- Carrera, E., Jackson, S. D., and Prat, S. (1999). Feedback control and diurnal regulation of gibberellin 20-oxidase transcript levels in potato. *Plant Physiol.* 119, 765–773. doi: 10.1104/pp.119.2.765
- Chailakhyan, M. K., Yanina, L. I., Devedzhyan, A. G., and Lotova, G. N. (1981). Photoperiodism and tuber formation in grafting of tobacco onto potato. *Dokl. Akad. Nauk* 257, 1276–1280.
- Chapman, H. W. (1958). Tuberization in the potato plant. *Physiol. Plant.* 11, 215–224. doi: 10.1111/j.1399-3054.1958.tb08460.x
- Chen, H., Rosin, F. M., Prat, S., and Hannapel, D. J. (2003). Interacting transcription factors from the three-amino acid loop extension superclass regulate tuber formation. *Plant Physiol.* 132, 1391–1404. doi: 10.1104/pp.103.022434
- Chincinska, I., Gier, K., Krügel, U., Liesche, J., He, H., Grimm, B., et al. (2013). Photoperiodic regulation of the sucrose transporter *StSUT4* affects the expression of circadian-regulated genes and ethylene production. *Front. Plant Sci.* 4:26. doi: 10.3389/fpls.2013.00026
- Chincinska, I. A., Liesche, J., Krügel, U., Michalska, J., Geigenberger, P., Grimm, B., et al. (2008). Sucrose transporter *StSUT4* from potato affects flowering, tuberization, and shade avoidance response. *Plant Physiol.* 146, 515–528. doi: 10.1104/pp.107.112334
- Chitwood, D. H., Nogueira, F. T., Howell, M. D., Montgomery, T. A., Carrington, J. C., and Timmermans, M. C. (2009). Pattern formation via small RNA mobility. *Genes Dev.* 23, 549–554. doi: 10.1101/gad.1770009
- D’Aloia, M., Bonhomme, D., Bouché, F., Tamseddak, K., Ormenese, S., Torti, S., et al. (2011). Cytokinin promotes flowering of *Arabidopsis* via transcriptional activation of the *FT* paralogue *TSF*. *Plant J.* 65, 972–979. doi: 10.1111/j.1365-313X.2011.04482.x
- Dinant, S., and Suárez-López, P. (2012). “Multitude of long-distance signal molecules via phloem,” in *Biocommunication of Plants*, eds G. Witzany and F. Baluška (Heidelberg: Springer), 89–121. doi: 10.1077/978-3-642-23524-5_6
- Doering-Saad, C., Newbury, H. J., Bale, J. S., and Pritchard, J. (2002). Use of aphid stylectomy and RT-PCR for the detection of transporter mRNAs in sieve elements. *J. Exp. Bot.* 53, 631–637. doi: 10.1093/jexbot/53.369.631
- Dunoyer, P., Schott, G., Himber, C., Meyer, D., Takeda, A., Carrington, J. C., et al. (2010). Small RNA duplexes function as mobile silencing signals between plant cells. *Science* 328, 912–916. doi: 10.1126/science.1185880
- Eriksson, S., Böhlenius, H., Moritz, T., and Nilsson, O. (2006). GA₄ is the active gibberellin in the regulation of *LEAFY* transcription and *Arabidopsis* floral initiation. *Plant Cell* 18, 2172–2181. doi: 10.1105/tpc.106.042317
- Ewing, E. E., and Struik, P. C. (1992). Tuber formation in potato: induction, initiation and growth. *Hortic. Rev.* 14, 89–198. doi: 10.1002/9780470650523.ch3
- Fernie, A. R., and Willmitzer, L. (2001). Molecular and biochemical triggers of potato tuber development. *Plant Physiol.* 127, 1459–1465. doi: 10.1104/pp.010764

- Giavalisco, P., Kapitza, K., Kolasa, A., Buhtz, A., and Kehr, J. (2006). Towards the proteome of *Brassica napus* phloem sap. *Proteomics* 6, 896–909. doi: 10.1002/pmic.200500155
- González-Schain, N. D., Díaz-Mendoza, M., Żurczak, M., and Suárez-López, P. (2012). Potato *CONSTANS* is involved in photoperiodic tuberization in a graft-transmissible manner. *Plant J.* 70, 678–690. doi: 10.1111/j.1365-313X.2012.04909.x
- Gregory, L. E. (1956). Some factors for tuberization in the potato plant. *Am. J. Bot.* 43, 281–288. doi: 10.2307/2438945
- Griebel, T., and Zeier, J. (2008). Light regulation and daytime dependency of inducible plant defenses in *Arabidopsis*: phytochrome signaling controls systemic acquired resistance rather than local defense. *Plant Physiol.* 147, 790–801. doi: 10.1104/pp.108.119503
- Guo, J.-L., Yu, C.-L., Fan, C.-Y., Lu, Q.-N., Yin, J.-M., Zhang, Y.-F., et al. (2010). Cloning and characterization of a potato *TFL1* gene involved in tuberization regulation. *Plant Cell Tissue Organ Cult.* 103, 103–109. doi: 10.1007/s11240-010-9759-8
- Hannapel, D. J. (2010). A model system of development regulated by the long-distance transport of mRNA. *J. Integr. Plant Biol.* 52, 40–52. doi: 10.1111/j.1744-7909.2010.00911.x
- Haywood, V., Yu, T.-S., Huang, N.-C., and Lucas, W. J. (2005). Phloem long-distance trafficking of *GIBBERELLIC ACID-INSENSITIVE* RNA regulates leaf development. *Plant J.* 42, 49–68. doi: 10.1111/j.1365-313X.2005.02351.x
- Hedden, P., and Thomas, S. G. (2012). Gibberellin biosynthesis and its regulation. *Biochem. J.* 444, 11–25. doi: 10.1042/BJ20120245
- Himber, C., Dunoyer, P., Moissiard, G., Ritzenthaler, C., and Voinnet, O. (2003). Transitivity-dependent and -independent cell-to-cell movement of RNA silencing. *EMBO J.* 22, 4523–4533. doi: 10.1093/emboj/cdg431
- Hiraoka, K., Yamaguchi, A., Abe, M., and Araki, T. (2013). The florigen genes *FT* and *TSF* modulate lateral shoot outgrowth in *Arabidopsis thaliana*. *Plant Cell Physiol.* 54, 352–368. doi: 10.1093/pcp/pcs168
- Huang, N.-C., Jane, W.-N., Chen, J., and Yu, T.-S. (2012). *Arabidopsis thaliana* *CENTRORADIALIS* homologue (*ATC*) acts systemically to inhibit floral initiation in *Arabidopsis*. *Plant J.* 72, 175–184. doi: 10.1111/j.1365-313X.2012.05076.x
- Jackson, S. D. (1999). Multiple signaling pathways control tuber induction in potato. *Plant Physiol.* 119, 1–8. doi: 10.1104/pp.119.1.1
- Jackson, S. D., James, P., Prat, S., and Thomas, B. (1998). Phytochrome B affects the levels of a graft-transmissible signal involved in tuberization. *Plant Physiol.* 117, 29–32. doi: 10.1104/pp.117.1.29
- Jackson, S. D., James, P. E., Carrera, E., Prat, S., and Thomas, B. (2000). Regulation of transcript levels of a potato gibberellin 20-oxidase gene by light and phytochrome B. *Plant Physiol.* 124, 423–430. doi: 10.1104/pp.124.1.423
- Jang, S., Torti, S., and Coupland, G. (2009). Genetic and spatial interactions between *FT*, *TSF* and *SVP* during the early stages of floral induction in *Arabidopsis*. *Plant J.* 60, 614–625. doi: 10.1111/j.1365-313X.2009.03986.x
- Kasai, A., Kanehira, A., and Harada, T. (2010). *miR172* can move long distances in *Nicotiana benthamiana*. *Open Plant Sci. J.* 4, 1–6. doi: 10.2174/1874294701004010001
- Kim, M., Canio, W., Kessler, S., and Sinha, N. (2001). Developmental changes due to long-distance movement of a homeobox fusion transcript in tomato. *Science* 293, 287–289. doi: 10.1126/science.1059805
- King, R. W., Mander, L. N., Asp, T., MacMillan, C. P., Blundell, C. A., and Evans, L. T. (2008). Selective deactivation of gibberellins below the shoot apex is critical to flowering but not to stem elongation of *Lolium*. *Mol. Plant* 1, 295–307. doi: 10.1093/mp/ssp030
- King, R. W., Moritz, T., Evans, L. T., Martin, J., Andersen, C. H., Blundell, C., et al. (2006). Regulation of flowering in the long-day grass *Lolium temulentum* by gibberellins and the *FLOWERING LOCUS T* gene. *Plant Physiol.* 141, 498–507. doi: 10.1104/pp.106.076760
- Kloosterman, B., Abelenda, J. A., Gomez, M. d. M. C., Oortwijn, M., de Boer, J. M., Kowitzanich, K., et al. (2013). Naturally occurring allele diversity allows potato cultivation in northern latitudes. *Nature* 495, 246–250. doi: 10.1038/nature11912
- Kloosterman, B., Navarro, C., Bijsterbosch, G., Lange, T., Prat, S., Visser, R. G. F., et al. (2007). *StGA2ox1* is induced prior to stolon swelling and controls GA levels during potato tuber development. *Plant J.* 52, 362–373. doi: 10.1111/j.1365-313X.2007.03245.x
- Komiya, R., Yokoi, S., and Shimamoto, K. (2009). A gene network for long-day flowering activates *RFT1* encoding a mobile flowering signal in rice. *Development* 136, 3443–3450. doi: 10.1242/dev.040170
- Lau, O. S., and Deng, X. W. (2010). Plant hormone signaling lightens up: integrators of light and hormones. *Curr. Opin. Plant Biol.* 13, 571–577. doi: 10.1016/j.pbi.2010.07.001
- Li, C., Gu, M., Shi, N., Zhang, H., Yang, X., Osman, T., et al. (2011). Mobile *FT* mRNA contributes to the systemic florigen signalling in floral induction. *Sci. Rep.* 1, 73. doi: 10.1038/srep00073
- Li, C., Zhang, K., Zeng, X., Jackson, S., Zhou, Y., and Hong, Y. (2009). A cis element within *Flowering Locus T* mRNA determines its mobility and facilitates trafficking of heterologous viral RNA. *J. Virol.* 83, 3540–3548. doi: 10.1128/JVI.02346-08
- Liesche, J., Krügel, U., He, H., Chincinska, I., Hackel, A., and Kühn, C. (2011). Sucrose transporter regulation at the transcriptional, post-transcriptional and post-translational level. *J. Plant Physiol.* 168, 1426–1433. doi: 10.1016/j.jplph.2011.02.005
- Lifschitz, E., Eviatar, T., Rozman, A., Shalit, A., Goldshmidt, A., Amsellem, Z., et al. (2006). The tomato *FT* ortholog triggers systemic signals that regulate growth and flowering and substitute for diverse environmental stimuli. *Proc. Natl. Acad. Sci. U.S.A.* 103, 6398–6403. doi: 10.1073/pnas.0601620103
- Lin, M.-K., Belanger, H., Lee, Y. J., Varkonyi-Gasic, E., Taoka, K.-I., Miura, E., et al. (2007). *FLOWERING LOCUS T* protein may act as the long-distance florigenic signal in the cucurbits. *Plant Cell* 19, 1488–1506. doi: 10.1105/tpc.107.051920
- Lin, S.-I., Chiang, S.-F., Lin, W.-Y., Chen, J.-W., Tseng, C.-Y., Wu, P.-C., et al. (2008). Regulatory network of microRNA399 and *PHO2* by systemic signaling. *Plant Physiol.* 147, 732–746. doi: 10.1104/pp.108.116269
- Lu, K.-J., Huang, N.-C., Liu, Y.-S., Lu, C.-A., and Yu, T.-S. (2012). Long-distance movement of *Arabidopsis* *FLOWERING LOCUS T* RNA participates in systemic floral regulation. *RNA Biol.* 9, 653–662. doi: 10.4161/rna.19965
- Mahajan, A., Bhogale, S., Kang, I., Hannapel, D., and Banerjee, A. (2012). The mRNA of a Knotted1-like transcription factor of potato is phloem mobile. *Plant Mol. Biol.* 79, 595–608. doi: 10.1007/s11103-012-9931-0
- Marín-González, E., and Suárez-López, P. (2012). “And yet it moves”: cell-to-cell and long-distance signaling by plant microRNAs. *Plant Sci.* 196, 18–30. doi: 10.1016/j.plantsci.2012.07.009
- Martin, A., Adam, H., Díaz-Mendoza, M., Żurczak, M., González-Schain, N. D., and Suárez-López, P. (2009). Graft-transmissible induction of potato tuberization by the microRNA *miR172*. *Development* 136, 2873–2881. doi: 10.1242/dev.031658
- Mathieu, J., Warthmann, N., Küttner, F., and Schmid, M. (2007). Export of *FT* protein from phloem companion cells is sufficient for floral induction in *Arabidopsis*. *Curr. Biol.* 17, 1055–1060. doi: 10.1016/j.cub.2007.05.009
- Matsoukas, I. G., Massiah, A. J., and Thomas, B. (2012). Florigenic and antiflorigenic signalling in plants. *Plant Cell Physiol.* 53, 1827–1842. doi: 10.1093/pcp/pcs130
- Molnar, A., Melnyk, C. W., Bassett, A., Hardcastle, T. J., Dunn, R., and Baulcombe, D. C. (2010). Small silencing RNAs in plants are mobile and direct epigenetic modification in recipient cells. *Science* 328, 872–875. doi: 10.1126/science.1187959
- Navarro, C., Abelenda, J. A., Cruz-Oró, E., Cuéllar, C. A., Tamaki, S., Silva, J., et al. (2011). Control of flowering and storage organ formation in potato by *FLOWERING LOCUS T*. *Nature* 478, 119–122. doi: 10.1038/nature10431
- Notaguchi, M., Abe, M., Kimura, T., Daimon, Y., Kobayashi, T., Yamaguchi, A., et al. (2008). Long-distance, graft-transmissible action of *Arabidopsis* *FLOWERING LOCUS T* protein to promote flowering. *Plant Cell Physiol.* 49, 1645–1658. doi: 10.1093/pcp/pcn154
- Pant, B. D., Buhtz, A., Kehr, J., and Scheible, W.-R. (2008). MicroRNA399 is a long-distance signal for the regulation of plant phosphate homeostasis. *Plant J.* 53, 731–738. doi: 10.1111/j.1365-313X.2007.03363.x
- Peres, L. E. P., Carvalho, R. F., Zsögön, A., Bermúdez-Zambrano, O. D., Robles, W. G. R., and Tavares, S. (2005). Grafting of tomato mutants onto potato rootstocks: an approach to study leaf-derived signaling on tuberization. *Plant Sci.* 169, 680–688. doi: 10.1016/j.plantsci.2005.05.017
- Pin, P. A., and Nilsson, O. (2012). The multifaceted roles of *FLOWERING LOCUS T* in plant development. *Plant Cell Environ.* 35, 1742–1755. doi: 10.1111/j.1365-3040.2012.02558.x

- Rodríguez-Falcón, M., Bou, J., and Prat, S. (2006). Seasonal control of tuberization in potato: conserved elements with the flowering response. *Annu. Rev. Plant Biol.* 57, 151–180. doi: 10.1146/annurev.arplant.57.032905.105224
- Rosin, F. M., Hart, J. K., Horner, H. T., Davies, P. J., and Hanappel, D. J. (2003). Overexpression of a *Knotted*-like homeobox gene of potato alters vegetative development by decreasing gibberellin accumulation. *Plant Physiol.* 132, 106–117. doi: 10.1104/pp.102.015560
- Ruan, Y.-L. (2012). Signaling role of sucrose metabolism in development. *Mol. Plant* 5, 763–765. doi: 10.1093/mp/sss046
- Ruiz-Medrano, R., Xoconostle-Cázares, B., and Lucas, W. J. (1999). Phloem long-distance transport of *CmNACP* mRNA: implications for supracellular regulation in plants. *Development* 126, 4405–4419.
- Sasaki, T., Chino, M., Hayashi, H., and Fujiwara, T. (1998). Detection of several mRNA species in rice phloem sap. *Plant Cell Physiol.* 39, 895–897. doi: 10.1093/oxfordjournals.pcp.a029451
- Sheen, J., Zhou, L., and Jang, J.-C. (1999). Sugars as signaling molecules. *Curr. Opin. Plant Biol.* 2, 410–418. doi: 10.1016/S1369-5266(99)00014-X
- Suárez-López, P. (2005). Long-range signalling in plant reproductive development. *Int. J. Dev. Biol.* 49, 761–771. doi: 10.1387/ijdb.052002ps
- Suzuki, A., Suriyagoda, L., Shigeyama, T., Tominaga, A., Sasaki, M., Hiratsuka, Y., et al. (2011). *Lotus japonicus* nodulation is photomorphogenetically controlled by sensing the red/far red (R/FR) ratio through jasmonic acid (JA) signaling. *Proc. Natl. Acad. Sci. U.S.A.* 108, 16837–16842. doi: 10.1073/pnas.1105892108
- Tamaki, S., Matsuo, S., Wong, H. L., Yokoi, S., and Shimamoto, K. (2007). Hd3a protein is a mobile flowering signal in rice. *Science* 316, 1033–1036. doi: 10.1126/science.1141753
- Tsuji, H., Taoka, K.-I., and Shimamoto, K. (2013). Florigen in rice: complex gene network for florigen transcription, florigen activation complex, and multiple functions. *Curr. Opin. Plant Biol.* 16, 228–35. doi: 10.1016/j.pbi.2013.01.005
- Turck, F., Fornara, F., and Coupland, G. (2008). Regulation and identity of florigen: FLOWERING LOCUS T moves center stage. *Annu. Rev. Plant Biol.* 59, 573–594. doi: 10.1146/annurev.arplant.59.032607.092755
- Turnbull, C. (2011). Long-distance regulation of flowering time. *J. Exp. Bot.* 62, 4399–4413. doi: 10.1093/jxb/err191
- Varkonyi-Gasic, E., Gould, N., Sandanayaka, M., Sutherland, P., and MacDiarmid, R. (2010). Characterisation of microRNAs from apple (*Malus domestica* 'Royal Gala') vascular tissue and phloem sap. *BMC Plant Biol.* 10:159. doi: 10.1186/1471-2229-10-159
- Xu, X., Pan, S., Cheng, S., Zhang, B., Mu, D., Ni, P., et al. (2011). Genome sequence and analysis of the tuber crop potato. *Nature* 475, 189–195. doi: 10.1038/nature10158
- Xu, X., van Lammeren, A. A. M., Vermeer, E., and Vreugdenhil, D. (1998). The role of gibberellin, abscisic acid, and sucrose in the regulation of potato tuber formation in vitro. *Plant Physiol.* 117, 575–584. doi: 10.1104/pp.117.2.575
- Yamaguchi, A., Kobayashi, Y., Goto, K., Abe, M., and Araki, T. (2005). *TWIN SISTER OF FT (TSF)* acts as a floral pathway integrator redundantly with *FT*. *Plant Cell Physiol.* 46, 1175–1189. doi: 10.1093/pcp/pci151
- Yoo, B.-C., Kragler, F., Varkonyi-Gasic, E., Haywood, V., Archer-Evans, S., Lee, Y. M., et al. (2004). A systemic small RNA signaling system in plants. *Plant Cell* 16, 1979–2000. doi: 10.1105/tpc.104.023614
- Zhu, Q.-H., and Helliwell, C. A. (2011). Regulation of flowering time and floral patterning by miR172. *J. Exp. Bot.* 62, 487–495. doi: 10.1093/jxb/erq295

Conflict of Interest Statement: The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 02 April 2013; paper pending published: 19 April 2013; accepted: 23 June 2013; published online: 16 July 2013.

Citation: Suárez-López P (2013) A critical appraisal of phloem-mobile signals involved in tuber induction. *Front. Plant Sci.* 4:253. doi: 10.3389/fpls.2013.00253
This article was submitted to *Frontiers in Plant Physiology*, a specialty of *Frontiers in Plant Science*.

Copyright © 2013 Suárez-López. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in other forums, provided the original authors and source are credited and subject to any copyright notices concerning any third-party graphics etc.